

Supporting information

The genetic Ca²⁺ sensor GCaMP3 reveals multiple Ca²⁺ stores differentially coupled to Ca²⁺ entry in the human malaria parasite *Plasmodium falciparum*

Lucas Borges-Pereira^{1,2#}, Samantha J. Thomas^{2#}, Amanda Laizy dos Anjos e Silva¹, Paula J. Bartlett²; Andrew P. Thomas² * and Célia R. S. Garcia^{1,3*}

¹Departamento de Fisiologia, Instituto de Biociências, Universidade de São Paulo, Brasil.

²Department of Pharmacology, Physiology and Neuroscience, Rutgers University, New Jersey Medical School, Newark, New Jersey, USA. ³Departamento de Análises Clínicas e Toxicológicas, Faculdade de Ciências Farmacêuticas, Universidade de São Paulo, Brasil

#These authors contributed equally to this work.

*Corresponding authors:

Célia R. S. Garcia: cgarcia@usp.br

Andrew Thomas : thomasap@njms.rutgers.edu

Running title: GCaMP3 studies of Ca²⁺ pools in *P. falciparum*

Keywords: Malaria, Calcium, *Plasmodium*, Thapsigargin, SOCE, GCaMP3, Signaling

List of the material included:

Figure S.1 Thg causes an addition artefact when added in the spectrofluorometer cuvette

Figure S.2 Cell population average of single cell [Ca²⁺]_c responses to Thg.

Figure S.3 Removal of extracellular Ca²⁺ reduces [Ca²⁺]_c to a new steady state in isolated *P. falciparum*.

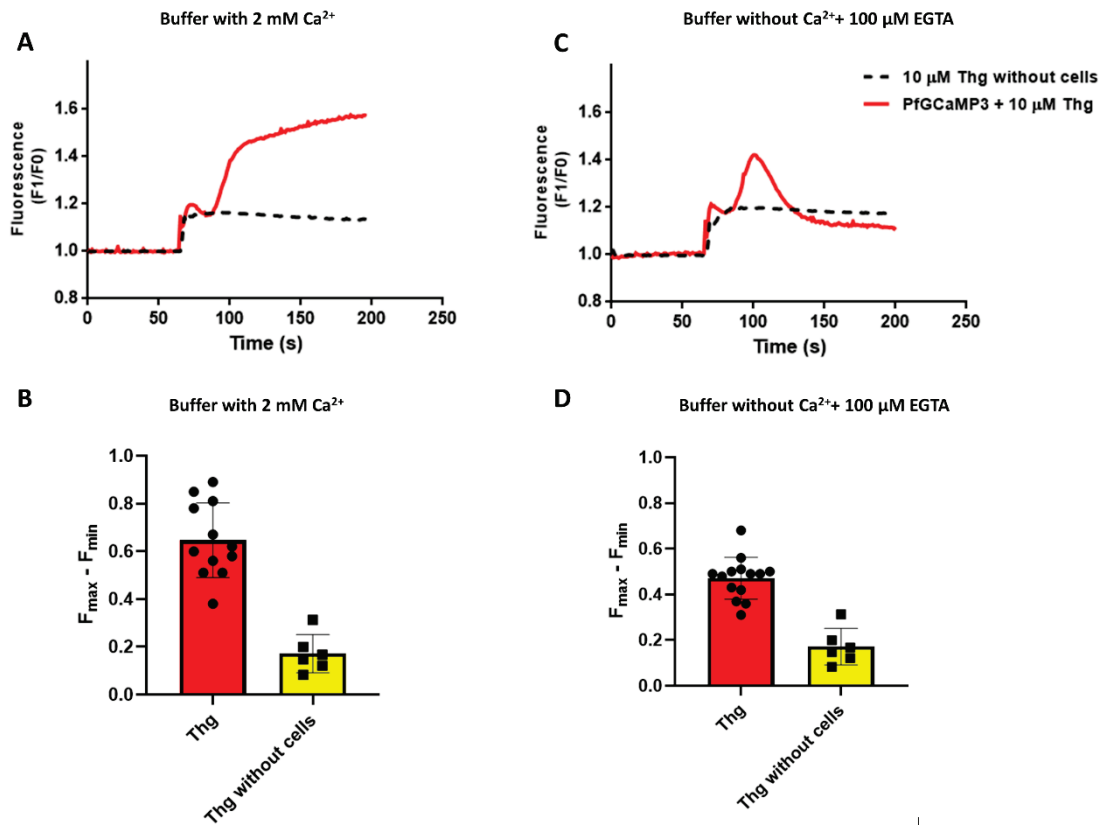


Fig. S1 Thg causes an addition artefact when added in the spectrofluorometer cuvette. Ten μM Thg was added in the spectrofluorometer cuvette in the absence of PfGCaMP3 parasites (buffer only). Thg *per se* showed a fluorescence artefact (dotted lines). This artefact was compared with the Ca²⁺ response elicited by Thg in PfGCaMP3 parasites (red lines) in the presence (A-B) and absence (C-D) of extracellular Ca²⁺.

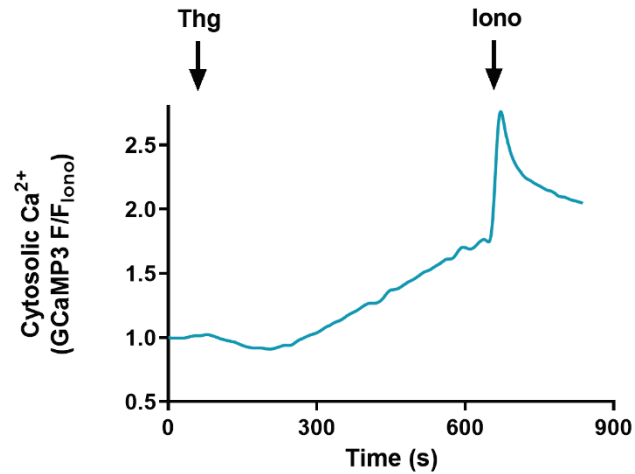


Fig. S2 Cell population average of single cell $[Ca^{2+}]_c$ responses to Thg.

Averaged $[Ca^{2+}]_c$ trace from 43 single trophozoite-stage *P. falciparum* parasites expressing GCaMP3 (PFGCaMP3). Cells were challenged with 5 μ M Thg in the presence of extracellular Ca^{2+} (2 mM), followed by 10 μ M Ionomycin (Iono). Responses in individual cells comprise complex oscillatory changes in $[Ca^{2+}]_c$ (see Fig. 4 C and F), whereas the population average is similar to fluorimeter cell population experiments (Figs. 1-3), albeit with a longer delay before the response, presumably because there is no mixing of drugs in the imaging experiments.

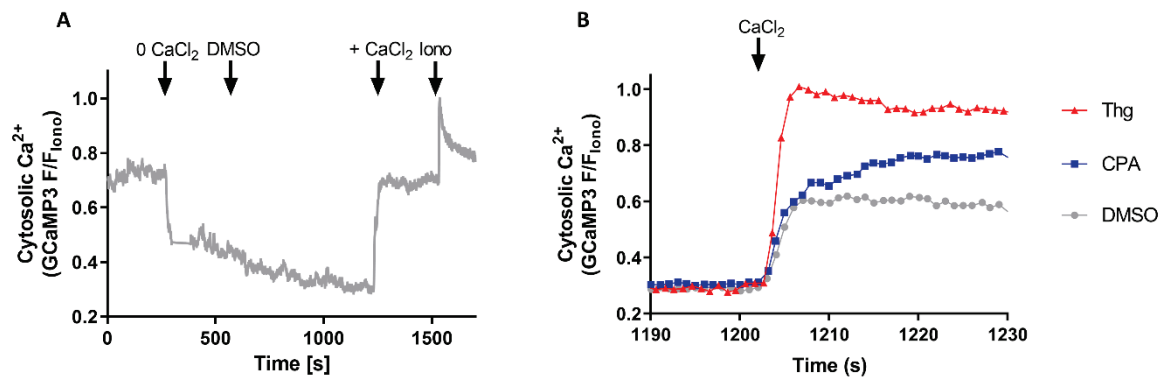


Fig. S3 Removal of extracellular Ca^{2+} reduces $[Ca^{2+}]_c$ to a new steady state in isolated *P. falciparum*. (A) Representative DMSO vehicle control trace to show the change in $[Ca^{2+}]_c$ upon removal and add back of extracellular Ca^{2+} in trophozoite-stage PfGCaMP3 parasites. Ionomycin ($10 \mu M$, Iono) was added at the end of each experiment. (B) Representative traces of the rise in $[Ca^{2+}]_c$ following addback of $2 \text{ mM } Ca^{2+}$ after removal of extracellular Ca^{2+} and 10 min treatment with DMSO vehicle, $10 \mu M$ CPA or $5 \mu M$ Thg). The extracellular Ca^{2+} washout period shown in the first 400 s of panel A was omitted for the traces in Figure 4. Following addback of $2 \text{ mM } CaCl_2$, $[Ca^{2+}]_c$ returned to basal levels in DMSO- and CPA-treated parasites. By contrast, Thg-treated parasites showed a larger amplitude and faster rate of $[Ca^{2+}]_c$ rise, consistent with SOCE. All data are representative of 3 independent experiments and responses were normalized to the peak ionomycin ($10 \mu M$, Iono) response (F/F_{Iono}).