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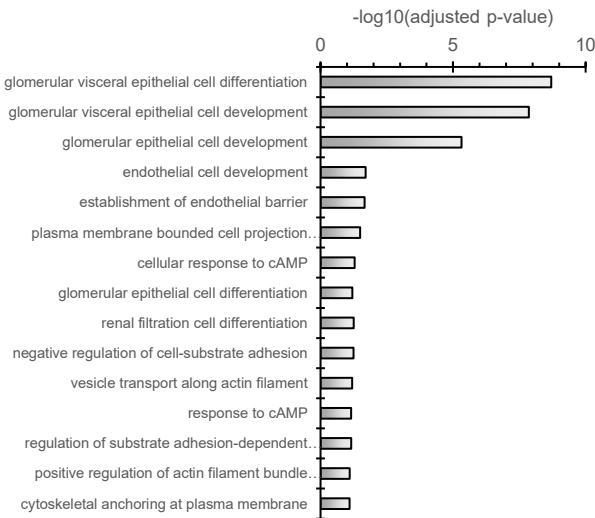
**Supplemental Video 1.** Calcium uptake by *Neb1<sup>-/-</sup>* podocytes

**Supplemental Table 1.** Rat primers used for qRT-PCR

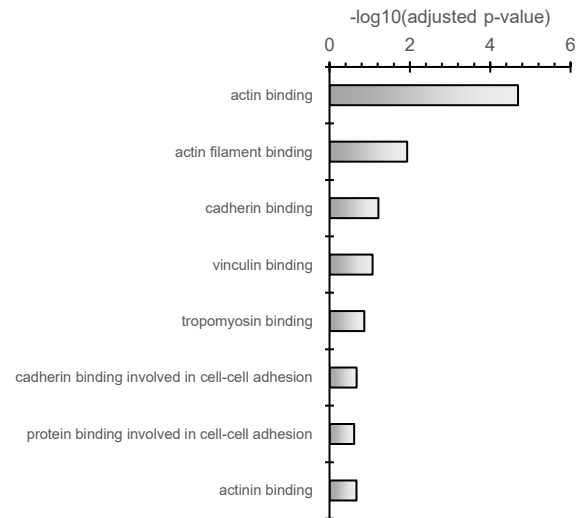
<b>Gene Name</b>	<b>Forward</b>	<b>Reverse</b>
<i>Gapdh</i>	TGCACCACCAACTGCTTAGC	GGCATGGACTGTGGTCATGA
<i>Actn4</i>	ACCATGCCTTTTCAGGAGCG	CTTCTGAGGCACACGGTCTT
<i>Nphs1</i>	CCACAGCGAGGCACTCCGTG	AGGATACGGTGCCGGGGACC
<i>Nphs1</i>	CCACAGCGAGGCACTCCGTG	AGGATACGGTGCCGGGGACC
<i>Nbl</i>	AGGAGCACCCGTCCTTCC	TAGGTCCTTAGATTTGCTGAATGCT

**Supplemental Figure 1.** Enrichment analyses for the top 52 consistently downregulated glomerular proteins in the transient rat puromycin aminonucleoside-induced nephropathy model. Proteomic hits were used to determine representation via the *EnrichR* suite within **(A)** gene ontology (GO) biological processes, **(B)** GO molecular function, **(C)** *ChEA* transcription factors, and **(D)** *MGI* mouse phenotype databases.

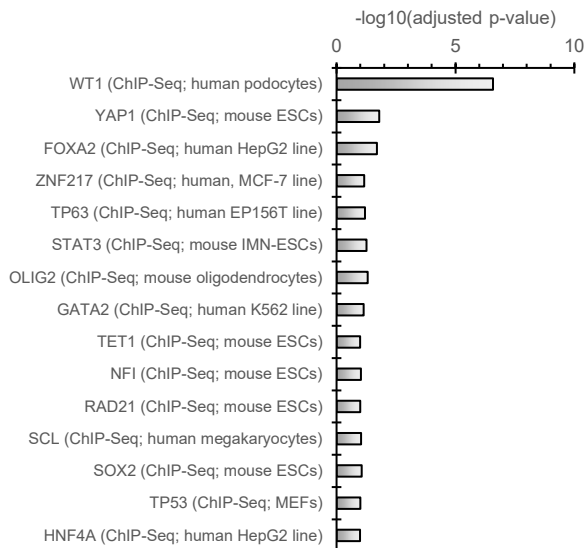
**A**



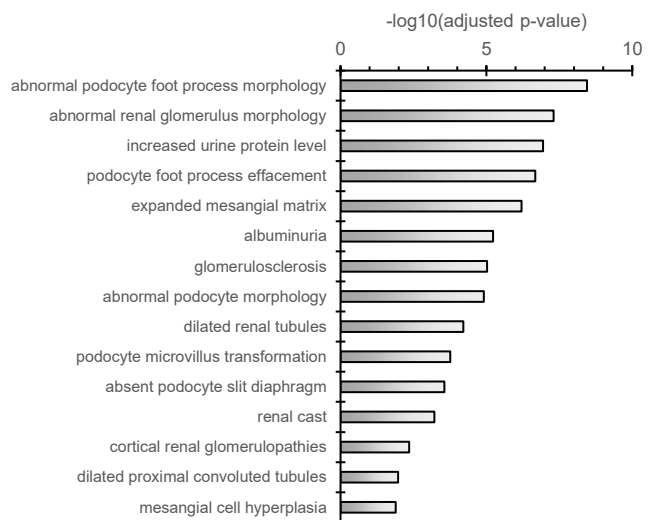
**B**



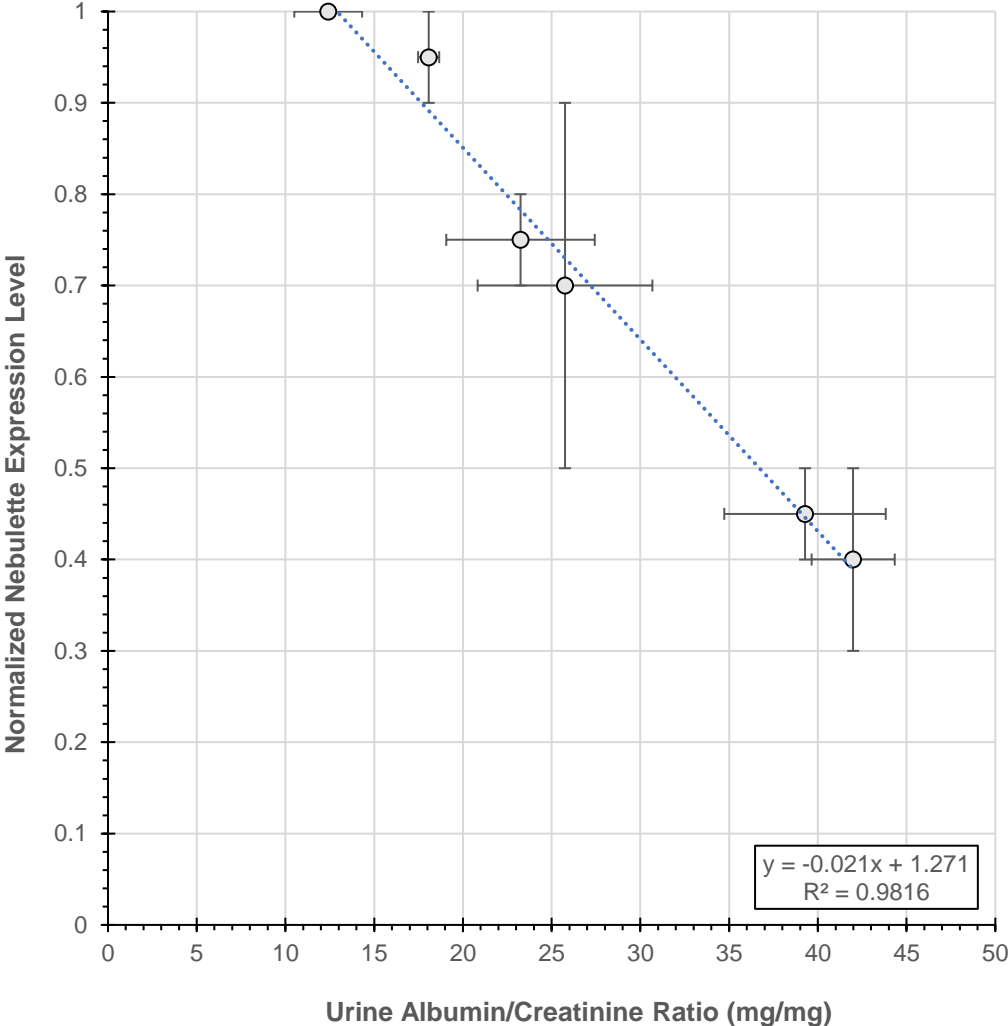
**C**



**D**



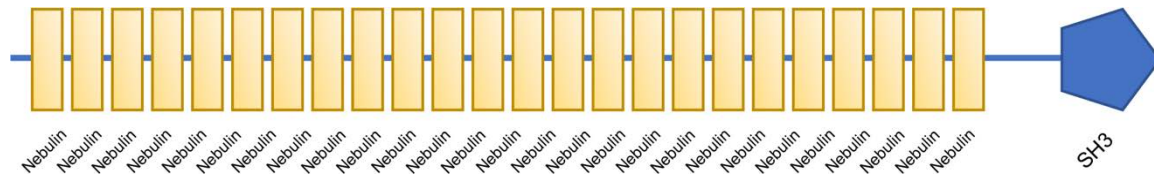
**Supplemental Figure 2.** Correlation between normalized nebullette expression levels as quantified by isobaric tagged proteomics and glomerular function as quantified by urine albumin/creatinine ratio.





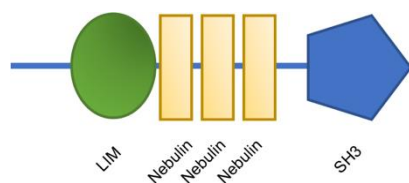
**Supplemental Figure 3.** Domains and the amino acid sequences of the two nebulette isoforms in the rat, and the mass spectrometry-detected peptides highlighted in yellow and cyan. We note that all the detected peptide spectra stem from the C-terminus region that is common between the two isoforms as shown in bold. None of the peptides unique to the first isoform are detected even though it is five times larger in size.

**Nebulette (Isoform-1), Rattus Norvegicus (Uniprot ID : D4A164\_RAT, 1015 AA)**



MKVPVSGDVKEETEEENVEEEEKPEDEVFLKPVVEDLSMELARKCTELISDIHYKEDYRKS KDKCTS VT DTP  
 TLNHVKNISAFISETKYKGTIKADLSNCLYKDMPATIDSVFAREVSQLQSEVAYKQKHEAEKGLSDYAHMKEP  
 PEVRHAMEVNRHQSNISYRKDVQGTHTYTAEMDRPDIKKATQISKIISNAEYKKGQGIVNKEPSVIGRPDFEH  
 AVEASKLSSQVKYKEKFDNEMKGGHYNPLGSASFRQHQLATVLASDVKYKQDVQTMHEPVS DLPNLLFL  
 DHALKASRMLSGWEYKKNFEENKGSYHFDAEAPHLHHRGNATLQSQVKYREEYEKNKGKTM LDFVETPS  
 YQSSKEAQMQSEKVKYKEDFEKEIKGRSSLDLDTKPAFLHVKHISNLMREKEYK KDLENEIKGKGMELSSSEV  
 LDIQRAKRASEMASEKEYK KDLLELEIKGKGMQIDADTLEIQRVKRAAKIASEKDYKRDLETEIKGKGMQVSTD  
 TLDVQRAKRASEMASQKQRKDLENEIKGKGMQVNDIPDMLRAKRASEIYSQKKYKDEAEKMLS NYSTVA  
 VTPEIQRIKTTQQNISNVSYKEEV RAGTAVRNTPEIERVKKQNHNIS SVKFEEGIKHATAISDPLELKRVTENQ  
 KDVINRFQYKEPTYKATPVTMTPEIERVRRNQEQLSAVKHKGELKQATSILDLPGLKRVRENQKTISNVYYK  
 GQLGRATALSVTPEMERVKKNQENISSVKYTQDHKQMKGRPCVILDTPALRHVKEAQN HVS MVK**YHEDFE**  
**KTKGRGFTPVVDDP****PTERVRKNTQVVSDAAYKGVQPHVEMDR** RPIIVDLKVVRTD PGSIFDIDPLEDNI  
 QSRSLHMLSEKASQYQSRLSRSHSGSTFGTGLGDDKSEISELYPSFSCCSEVTRPSDEGAPVLP GAYQQS  
 HSQGYGYMHQTSVSSVRSMQHSANLR TYR**AMYDYSAQDEDEVSFRDGDYIVNVQPIDDGW****MYGTVQR****TG**  
**RTGMLPANYIEFVN**

**LIM-nebulette (Isoform-2), Rattus Norvegicus (Uniprot ID: A0A146J2K6\_RAT, 270 AA)**

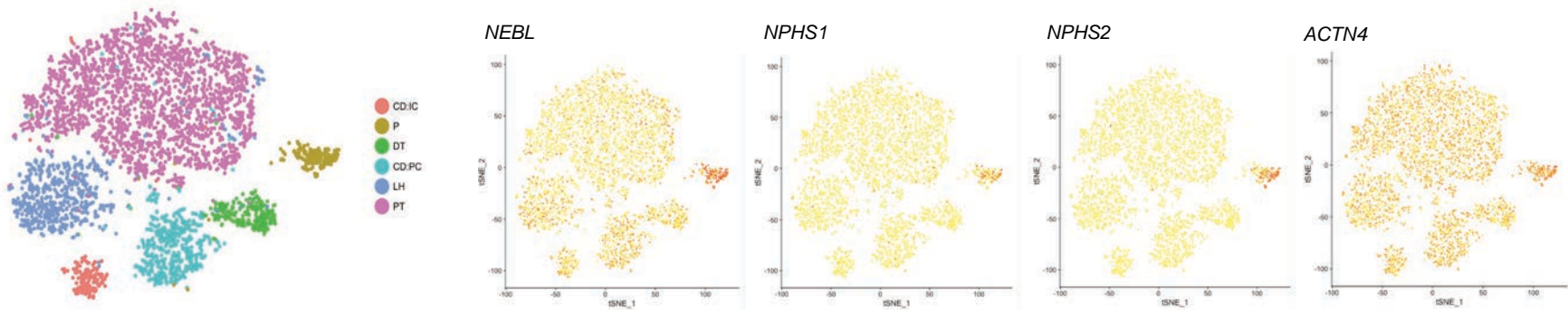


MNPQCARCGKVVPTEKVNCLDKYWHKGFHCEVCKMALNMNNYKGYEKKPYCNAHYPKQSFTTVADTP  
 ENLRLKQQSELQSQVKYKRD FEESRGRGFSIVTDTPELQRLKRTQE QISNVK**YHEDFEKTKGRGFTPVVDD**  
**PVTERVRKNTQVVSDAAYKGVQPHVEMDR** RPIIVAPVLP GAYQQSHS QGYGYMHQTSVSSVRSMQHS  
 ANLR TYR**AMYDYSAQDEDEVSFRDGDYIVNVQPIDDGW****MYGTVQR****TGRTGMLPANYIEFVN**



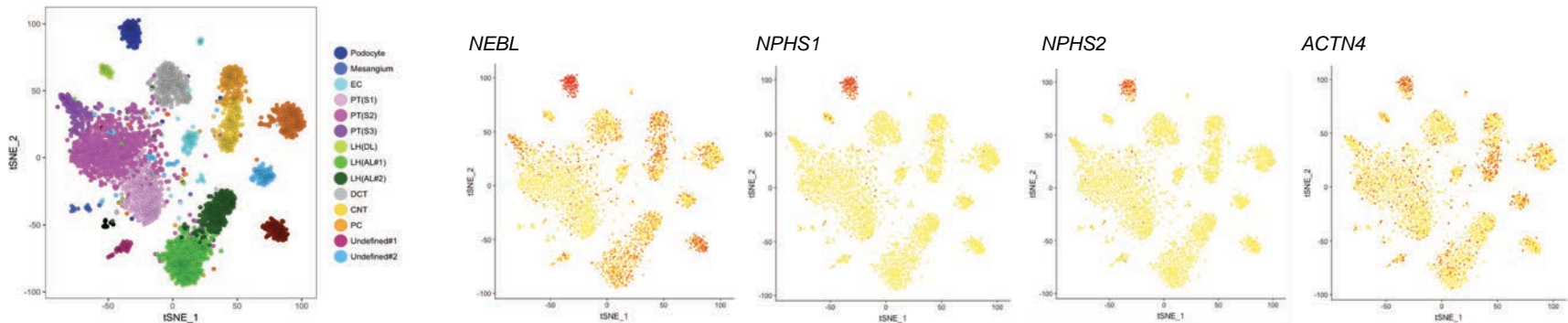
**Supplemental Figure 5.** Analysis of *NEBL* expression in public databases. **(A)** Unsupervised clustering of 4,259 single nuclei that has identified six distinct cell types in human adult kidney. These types include three tubular cell types of proximal tubule (PT), loop of Henle (LH), and distal tubule (DT); two collecting duct (CD) cell types of principal cells (PC) and intercalated cells (IC); and one podocyte population (P). **(B)** Unsupervised clustering of 4,524 single-nuclear RNA sequencing of adult human kidney that has identified 17 distinct cell types in human adult kidney including 11 tubular cell types, podocytes, mesangium, endothelial cells, and macrophages. Superimposed over the tSNE plots are the expression levels of *NEBL*, *NPHS1*, *NPHS2*, and *ACTN4* transcripts.

**A**



from Wu and Malone *et al.* J Am Soc Neph 2018 (Human Adult Kidney, Epithelial)

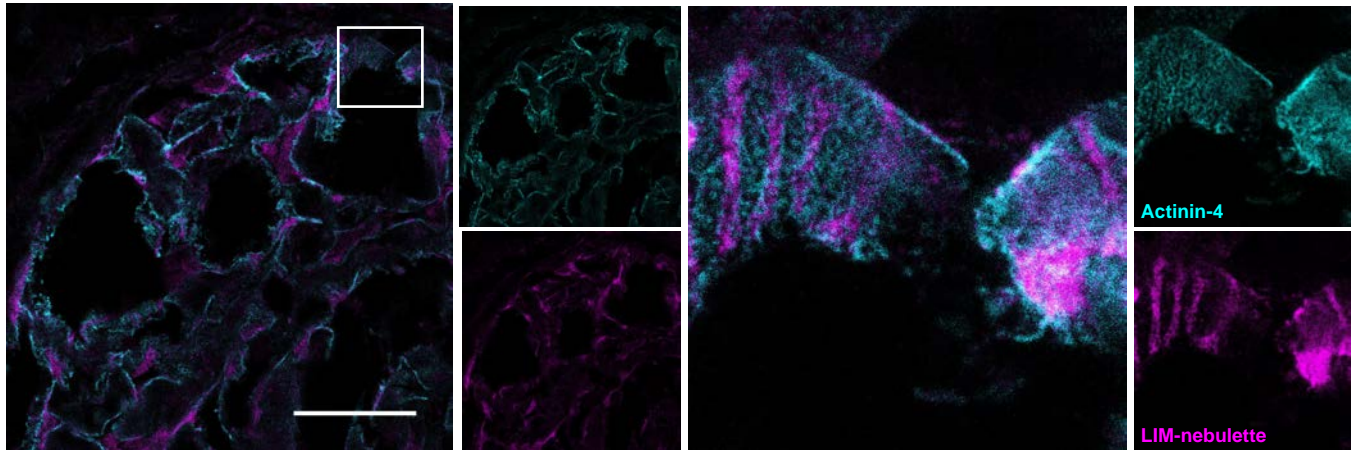
**B**



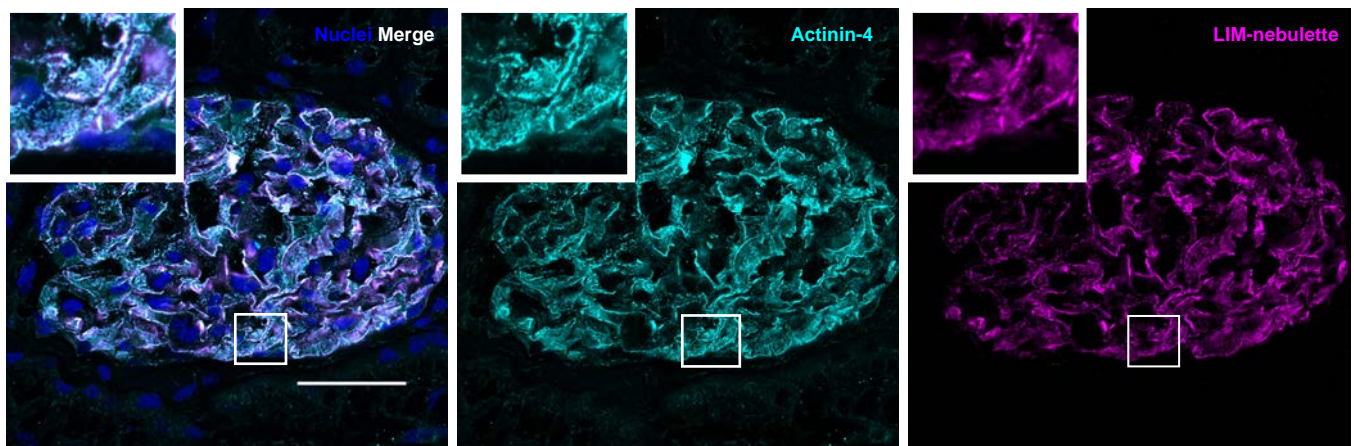
from Wu and Uchimura *et al.* Cell Stem Cell 2018 (Human Adult Kidney, Total)

**Supplemental Figure 6.** Super resolution imaging of human glomeruli. **(A)** Stimulated emission-depletion (STED) imaging at 100X and **(B)** Zeiss Airyscan laser scanning confocal imaging at 63X showing expression of LIM-nebulette (magenta) and actinin-4 (cyan) localizing to visceral epithelial processes.

**A**

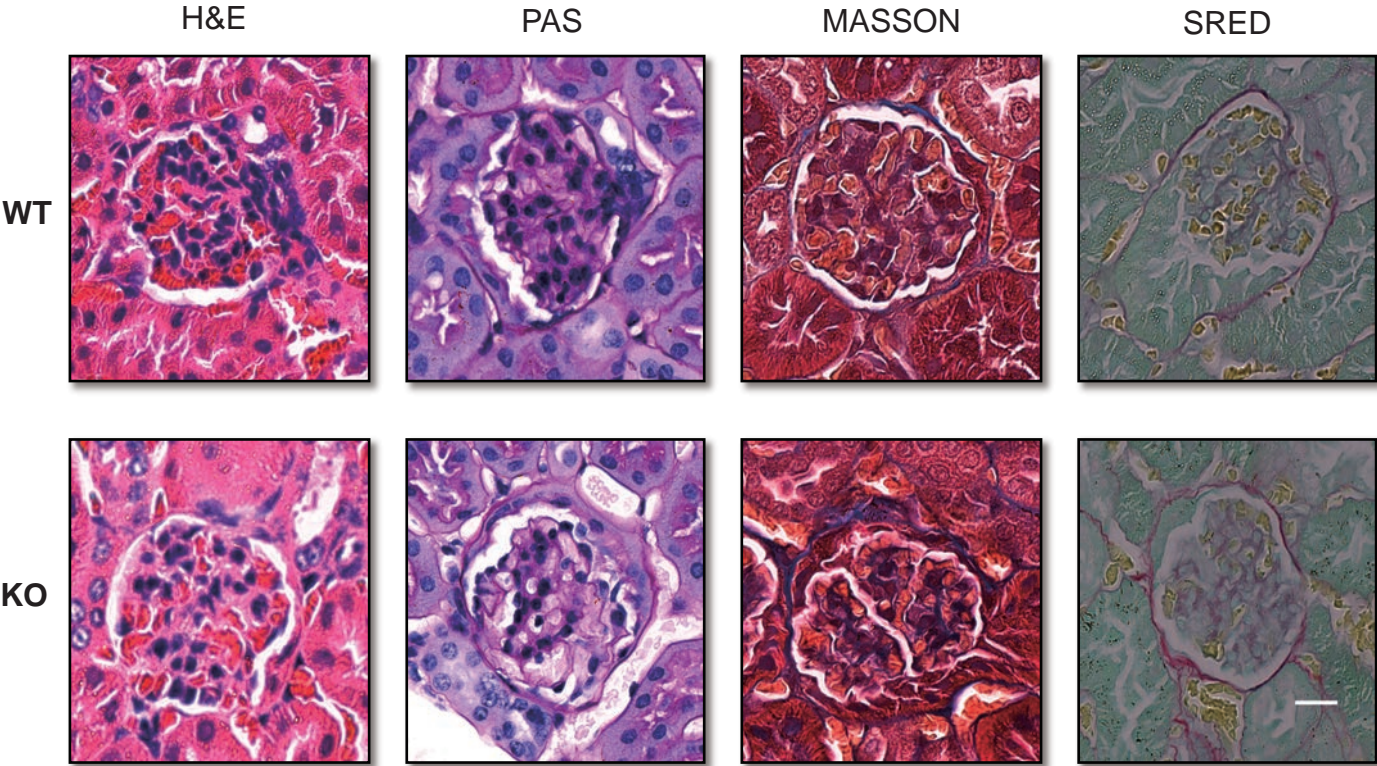


**B**



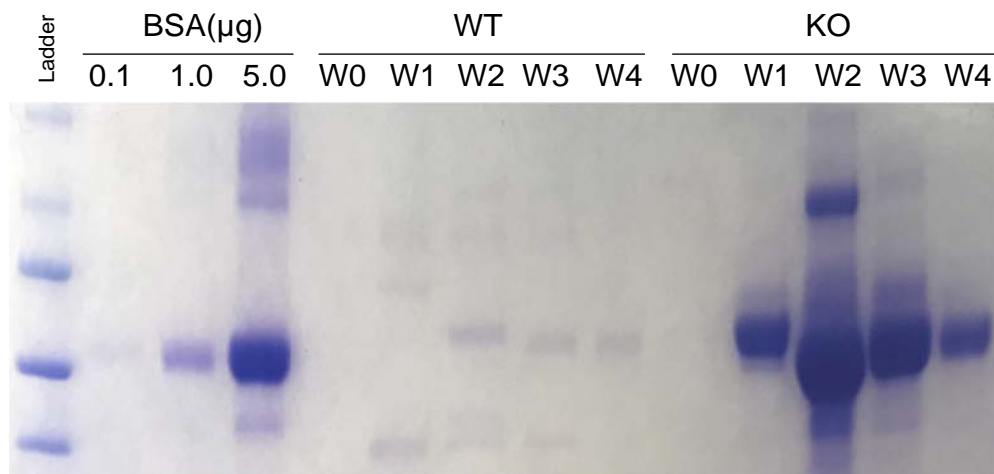


**Supplemental Figure 7.** Histopathological assessment of wild type (WT) and nebulin knockout (KO) mice using hematoxylin and eosin (H&E), Periodic acid-Schaff (PAS), Masson's trichrome (MASSON) and Picrosirius red (SRED) staining.



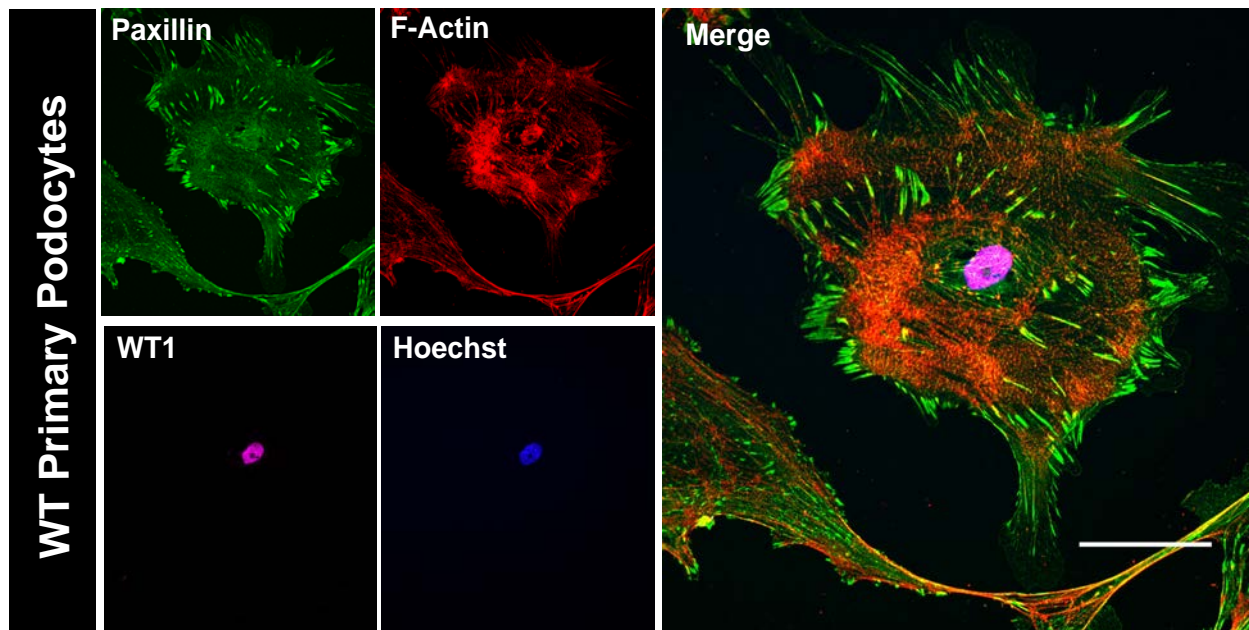
Scale bar= 50µm

**Supplemental Figure 8.** Visual comparison of albuminuria between wild type (WT) and nebulin knockout (KO) mice injected with single dose of Adriamycin (ADR). Urine was collected at baseline (W0), and weekly for four weeks (W1, W2, W3, W4). SDS-PAGE Coomassie brilliant blue staining of bovine serum albumin (BSA) standards and urine samples from WT and KO mice.

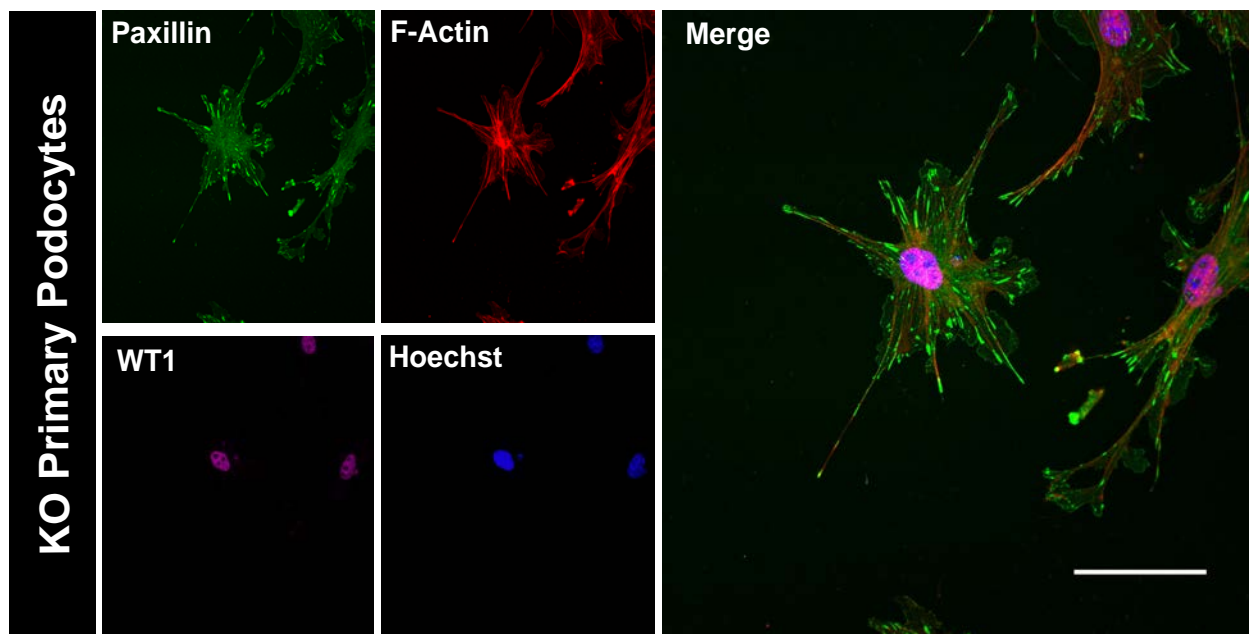


**Supplemental Figure 9.** Freshly isolated mouse primary podocytes from **(A)** wild type (WT) and **(B)** nebullette knockout (KO) animals showing expression of paxillin, F-actin, Wilm's tumor 1 (WT1), and nuclei (Hoechst). Scale bars = 50  $\mu$ m.

**A**

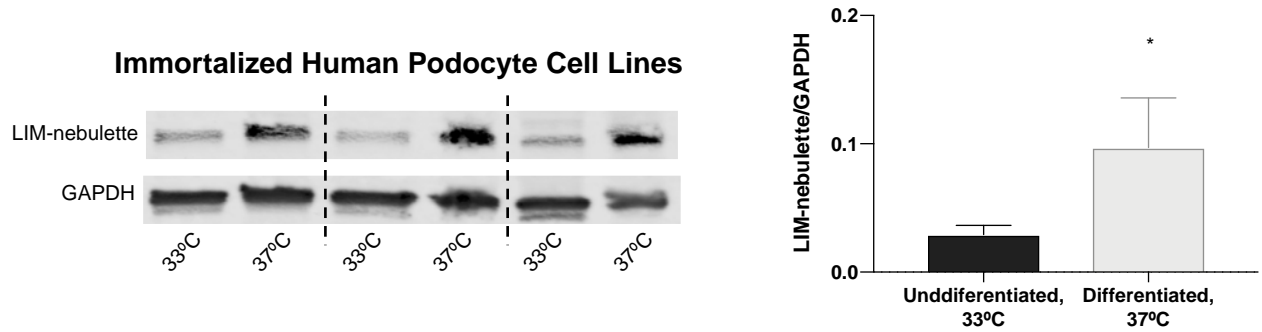


**B**

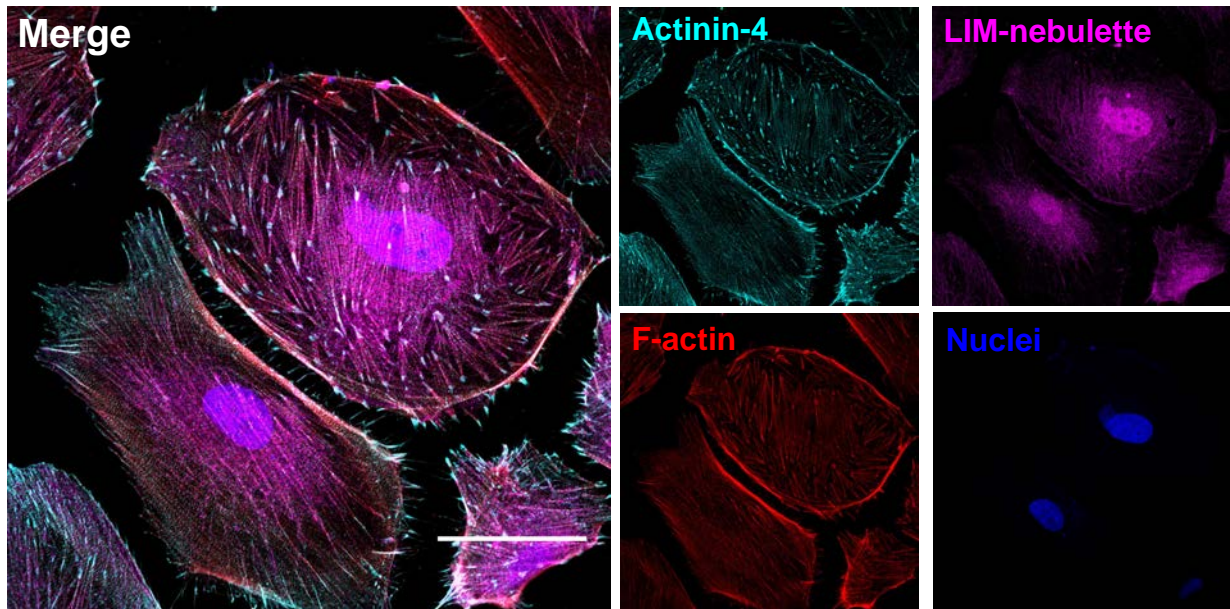


**Supplemental Figure 10.** Expression of LIM-nebulette in immortalized human podocyte cell line **(A)** as assessed by western blotting before (33°C) and after (37°C) differentiation for 10 days and quantified. **(B)** Spatial distribution of LIM-nebulette (magenta), actinin-4 (cyan), F-actin (red) and nuclei (blue) in the immortalized human podocyte cell line differentiated for 10 days at 37°C. Scale bar = 50 µm.

**A**

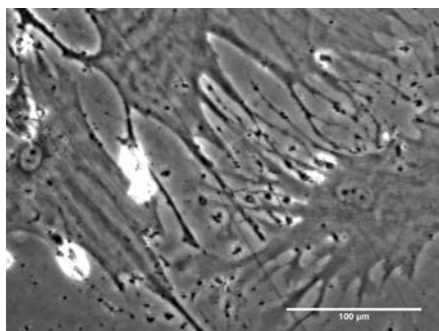


**B**

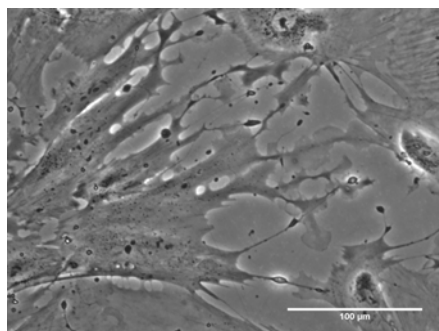




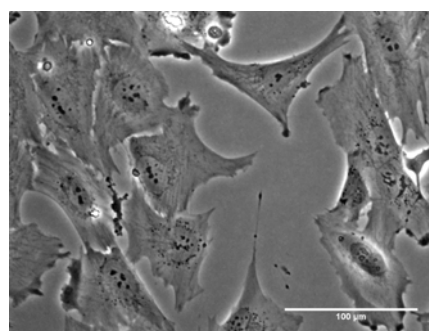
**Supplemental Figure 11.** Morphological comparison of arborization in freshly isolated mouse primary podocytes (left), human induced pluripotent stem cell (hiPSC)-derived podocytes (middle), and immortalized human podocyte cell line (right) under phase contrast bright field at 40X magnification. Scale bars = 100  $\mu$ m.



**Mouse primary podocytes**

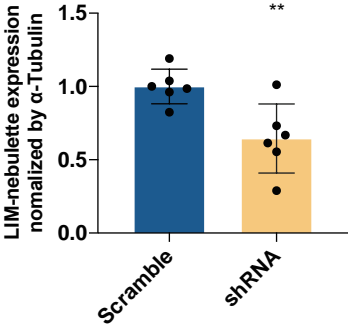


**hiPSC-derived podocytes**

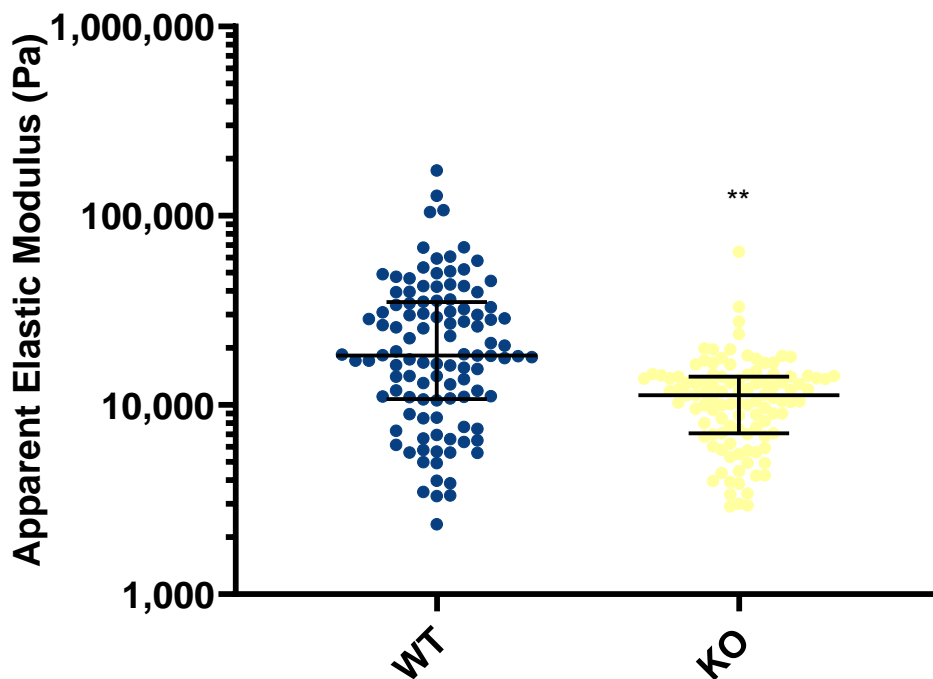


**Immortalized human podocytes cell line**

**Supplemental Figure 12.** Expression of nebulette in immortalized human podocyte cell line expressing scrambled or PAN-nebulette shRNA was assessed using Western blotting and quantified.



**Supplemental Figure 13.** Spatial distribution of subcellular elasticity of primary podocytes isolated from *Neb1<sup>+/+</sup>* (WT) or *Neb1<sup>-/-</sup>* (KO) mice. Cells were cultured on 50-mm type-I collagen coated dishes for 48 hours and probed using a standard pyramidal tip over a 6 x 6 perinuclear indentation array on an Asylum MFP-3D atomic force microscope (AFM) at 37°C. Podocytes from KO animals had a significantly lower elastic modulus (\*\* $p = 0.0013$ ; repeated measures ANOVA,  $n = 108$ ). Median  $\pm$  interquartile range of apparent elastic modulus for WT cells was  $18.3 \pm 11.9$  kPa versus  $11.3 \pm 3.5$  kPa in KO cells. AFM indentation rate was 10  $\mu\text{m}/\text{sec}$ . Probed cellular volumes were  $20 \times 20 \times 5$   $\mu\text{m}$ .



**Supplemental Figure 14.** Changes in spatial distribution of LIM-nebulette (magenta) in differentiated immortalized human podocytes upon exposure to F-actin (Cytochalasin D and Latrunculin B) or vimentin (Arylquin-1 and Withaferin A) disruptors. Representative immunofluorescence images were taken at 63X using Zeiss Airyscan super-resolution confocal microscope. Scale bars = 25  $\mu$ m.

