

## SUPPLEMENTARY MATERIAL

### **Age-related alterations in fertilization-induced Ca<sup>2+</sup> oscillations depend on the genetic background of mouse oocytes**

by

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## **Supplemental Figure legends**

### **Supplemental Figure S1**

#### **Characteristics of the Oregon Green 488 BAPTA 1-AM staining**

(A) Initial fluorescence intensity of the Oregon Green 488 BAPTA staining recorded for oocytes from young (black squares) and old (white squares) F1, SWISS, and C57BL/6 females. Each small square reflects a mean fluorescence intensity calculated for a single experiment (either in vitro fertilization or TG/A23187 treatment). Big squares reflect average intensities for all experiments conducted for a given type of oocytes. Error bars indicate standard deviations. (B) Representative image sequences showing Oregon Green 488 BAPTA fluorescence in oocytes from young and old F1, SWISS, and C57BL/6 females at the beginning of the recording and just before and during the 1<sup>st</sup> and the 3<sup>rd</sup> Ca<sup>2+</sup> transients. Time is measured from the beginning of the recording and is shown in minutes and seconds. There is no visible compartmentalization of the dye. Scale bar 10 μm.

### **Supplemental Figure S2**

#### **Representative patterns of Ca<sup>2+</sup> oscillations in fertilized oocytes from young and old F1, SWISS and C57BL/6 mice**

(A-B) The representative patterns of Ca<sup>2+</sup> oscillations in fertilized oocytes obtained from (A) young and (B) old F1 females. (C-D) The representative patterns of Ca<sup>2+</sup> oscillations in fertilized oocytes obtained from (C) young and (D) old SWISS females. (E-F) The representative patterns of Ca<sup>2+</sup> oscillations in fertilized oocytes obtained from (E) young and (F) old C57BL/6 females.

### **Supplemental Figure S3**

#### **Characteristics of the 3<sup>rd</sup> Ca<sup>2+</sup> transient in fertilized oocytes from young and old F1, SWISS and C57BL/6 mice**

(A) Duration and (B) amplitude of the 3<sup>rd</sup> Ca<sup>2+</sup> transient in young (n = 56 F1, 92 SWISS, 36 C57BL/6) and old (n = 29 F1, 73 SWISS, 16 C57BL/6) oocytes. (C) Rate of Ca<sup>2+</sup> increase during the 3<sup>rd</sup> Ca<sup>2+</sup> transient in young (n = 56 F1, 92 SWISS, 36 C57BL/6) and old (n = 29 F1, 74 SWISS, 16 C57BL/6) oocytes. (D) Rate of Ca<sup>2+</sup> decrease during the 3<sup>rd</sup> Ca<sup>2+</sup> transient in young (n = 56 F1, 92 SWISS, 36 C57BL/6) and old (n = 29 F1, 74 SWISS, 16 C57BL/6) oocytes. (A-D) Graphs present medians and the 1<sup>st</sup> and the 3<sup>rd</sup> quartile values. The ends of the whiskers are set at 1.5\*IQR above the third quartile and 1.5\*IQR below the first quartile. Dots show the minimum and maximum values if they are outside the range (outliers).

### **Supplemental Figure S4**

#### **ITPR and ATP2A2 in oocytes from young and old F1 females**

(A) A representative immunoblot showing the amount of ITPR protein in oocytes from young and old F1 females. The dashed line shows the area presented in Fig. 3 F (upper panel). The contrast and brightness have not been adjusted. (B) Quantification of the ITPR immunostaining in young (n=49) and old (n=13) F1 oocytes. (C) Quantification of the ATP2A2 immunostaining in young (n=29) and old (n=28) F1 oocytes. All intensity values in (B-C) were normalized with the mean fluorescence intensity calculated for the control, young oocytes imaged with the same settings. Graphs present medians and the 1<sup>st</sup> and the 3<sup>rd</sup> quartile values. The ends of the whiskers are set at 1.5\*IQR above the third quartile and 1.5\*IQR below the first quartile. Dots show the minimum and maximum values if they are outside the range (outliers).

**Supplemental Table S1.** The pattern of Ca<sup>2+</sup> oscillations and time of pronuclei formation in fertilized oocytes obtained from young and old females of different genetic background

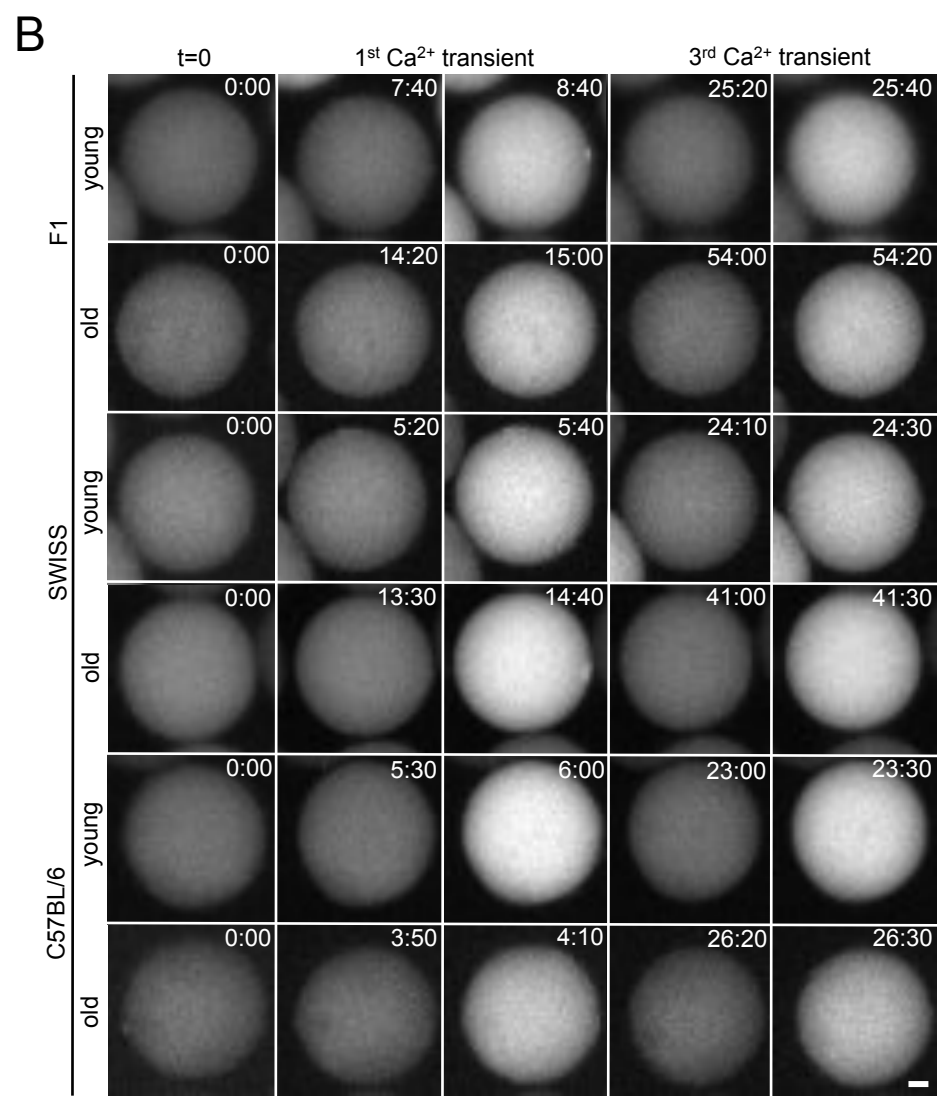
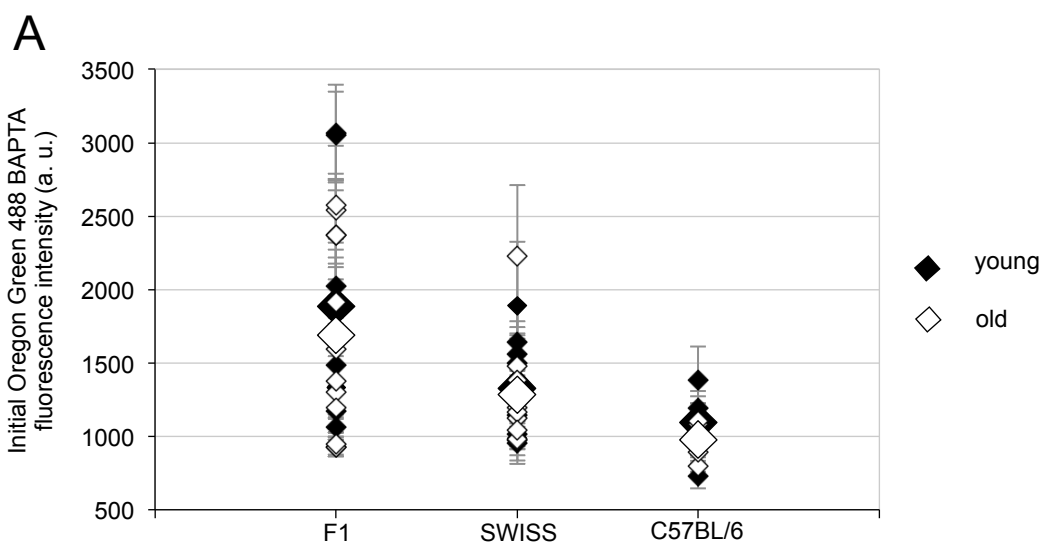
Parameter	Median (Q1; Q3)					
	Number of analyzed embryos					
	F1		SWISS		C57BL/6	
	young (3 exp)#	old (8 exp)	young (4 exp)	old (4 exp)	young (2 exp)	old (2 exp)
No. of Ca <sup>2+</sup> transients	<b>10.0</b> (8.0; 11.0) n=57	<b>7.0<sup>a</sup></b> (7.0; 8.0) n=30	<b>9.0</b> (7.0; 12.0) n=93	<b>9.0</b> (7.0; 11.0) n=74	<b>8.5</b> (8.0; 9.3) n=36	<b>7.0<sup>c</sup></b> (5.0; 8.0) n=17
Total duration of Ca <sup>2+</sup> oscillations (min)	<b>153.2</b> (130.8; 172.5) n=57	<b>162.7</b> (122.7; 176.3) n=29	<b>162.1</b> (142.7; 194.3) n=92	<b>152.4<sup>d</sup></b> (132.9; 177.6) n=74	<b>153.0</b> (133.3; 171.6) n=36	<b>111.8<sup>b</sup></b> (66.0; 143.0) n=17
Mean interval between Ca <sup>2+</sup> transients* (min)	<b>15.5</b> (13.3; 17.8) n=57	<b>21.9<sup>a</sup></b> (18.9; 23.4) n=29	<b>16.9</b> (13.7; 20.8) n=92	<b>17.8</b> (15.1; 20.6) n=74	<b>17.6</b> (15.2; 20.6) n=36	<b>15.3</b> (12.3; 19.7) n=17
Duration of the 1 <sup>st</sup> Ca <sup>2+</sup> transient (min)	<b>2.3</b> (2.0; 2.4) n=55	<b>2.8<sup>a</sup></b> (2.5; 3.1) n=27	<b>3.2</b> (2.7; 3.7) n=92	<b>2.5<sup>a</sup></b> (2.0; 2.8) n=73	<b>2.2</b> (2.0; 2.5) n=36	<b>2.6<sup>c</sup></b> (2.2; 2.7) n=18
Amplitude of the 1 <sup>st</sup> Ca <sup>2+</sup> transient ( $\Delta F/F_0$ )	<b>0.8</b> (0.6; 0.9) n=55	<b>0.9</b> (0.8; 1.0) n=27	<b>0.8</b> (0.8; 0.9) n=92	<b>0.8</b> (0.8; 0.9) n=73	<b>1.0</b> (1.0; 1.1) n=36	<b>1.0<sup>d</sup></b> (0.9; 1.0) n=18
Duration of the 3 <sup>rd</sup> Ca <sup>2+</sup> transient (min)	<b>0.7</b> (0.7; 0.8) n=56	<b>0.8<sup>a</sup></b> (0.8; 1.0) n=29	<b>1.0</b> (0.8; 1.2) n=92	<b>0.8<sup>c</sup></b> (0.8; 1.0) n=73	<b>0.8</b> (0.7; 0.8) n=36	<b>0.7</b> (0.5; 0.7) n=16
Amplitude of the 3 <sup>rd</sup> Ca <sup>2+</sup> transient ( $\Delta F/F_0$ )	<b>0.7</b> (0.6; 0.9) n=56	<b>0.8<sup>c</sup></b> (0.8; 0.8) n=29	<b>0.8</b> (0.7; 0.8) n=92	<b>0.8</b> (0.7; 0.8) n=73	<b>0.8</b> (0.7; 0.9) n=36	<b>0.9</b> (0.8; 0.9) n=16
Rate of the 1 <sup>st</sup> Ca <sup>2+</sup> increase (min <sup>-1</sup> )	<b>3.0</b> (2.4; 3.9) n=54	<b>3.0</b> (2.5; 3.8) n=27	<b>3.2</b> (2.8; 3.9) n=92	<b>3.1</b> (2.6; 3.8) n=74	<b>5.0</b> (3.7; 5.9) n=36	<b>3.7<sup>c</sup></b> (2.9; 4.8) n=18
Rate of the 1 <sup>st</sup> Ca <sup>2+</sup> decrease (min <sup>-1</sup> )	<b>-3.9</b> (-4.5; -2.9) n=56	<b>-3.9</b> (-4.3; -3.5) n=30	<b>-3.5</b> (-3.8; -3.2) n=92	<b>-3.5</b> (-3.9; -3.0) n=74	<b>-4.5</b> (-4.9; -3.9) n=36	<b>-4.0<sup>c</sup></b> (-4.3; -3.8) n=18
Rate of the 3 <sup>rd</sup> Ca <sup>2+</sup> increase (min <sup>-1</sup> )	<b>4.0</b> (3.1; 5.0) n=56	<b>4.5<sup>c</sup></b> (4.2; 4.8) n=29	<b>3.7</b> (3.3; 4.1) n=92	<b>3.8</b> (3.4; 4.2) n=74	<b>4.8</b> (4.2; 5.4) n=36	<b>5.1</b> (4.6; 5.3) n=16
Rate of the 3 <sup>rd</sup> Ca <sup>2+</sup> decrease (min <sup>-1</sup> )	<b>-4.1</b> (-4.8; -3.4) n=56	<b>-4.5<sup>d</sup></b> (-4.7; -4.1) n=29	<b>-3.7</b> (-4.2; -3.5) n=92	<b>-3.7</b> (-4.0; -3.3) n=74	<b>-4.4</b> (-5.4; -4.1) n=36	<b>-4.9</b> (-5.3; -4.5) n=16
Time of pronuclei formation (min)	<b>189.4</b> (170.4; 229.6) n=26	<b>242.7<sup>b</sup></b> (214.0; 261.7) n=13	<b>206.5</b> (179.5; 253.2) n=42	<b>247.4<sup>b</sup></b> (229.4; 269.3) n=22	<b>225.3</b> (216.5; 251.6) n=28	<b>201.5<sup>b</sup></b> (188.3; 211.1) n=12

\*calculated for the first 2 hrs of Ca<sup>2+</sup> oscillations

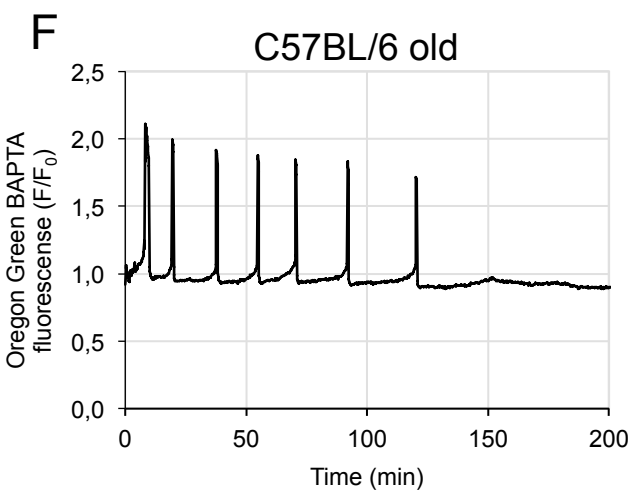
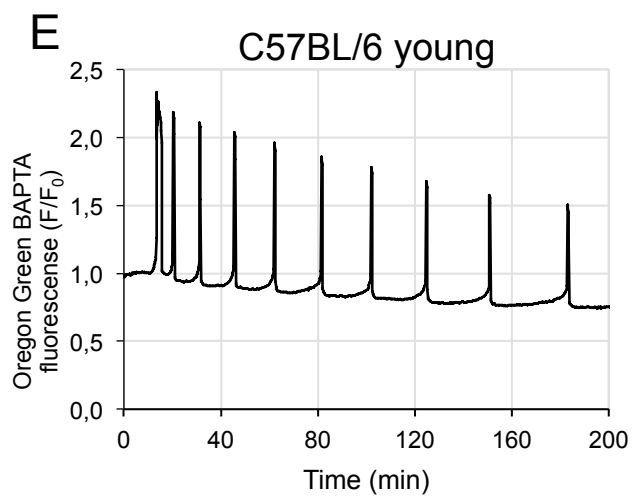
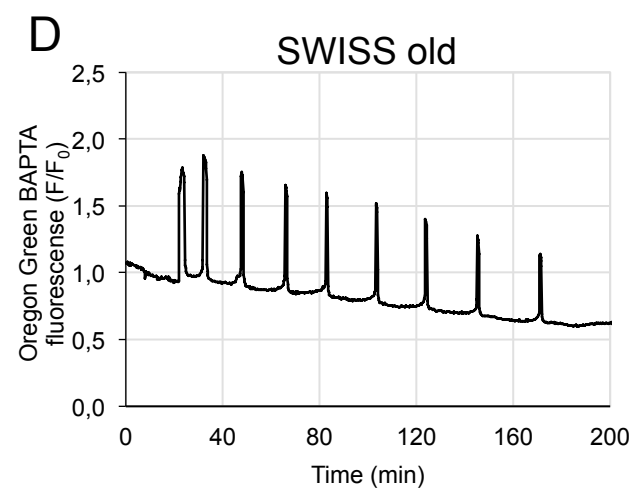
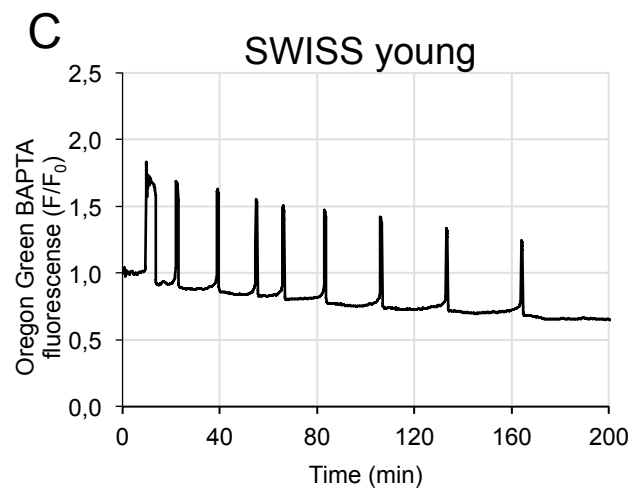
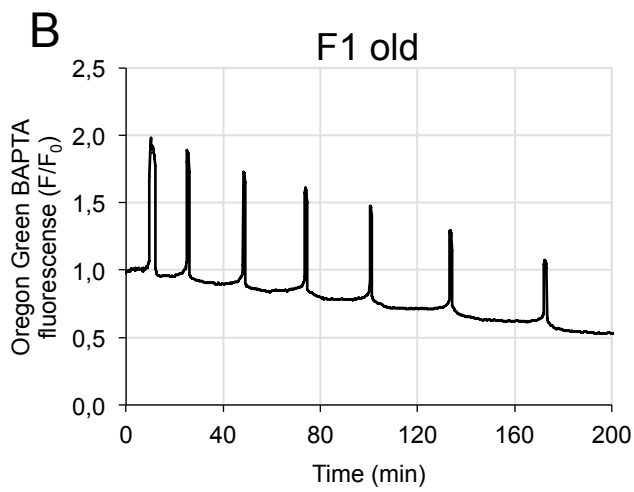
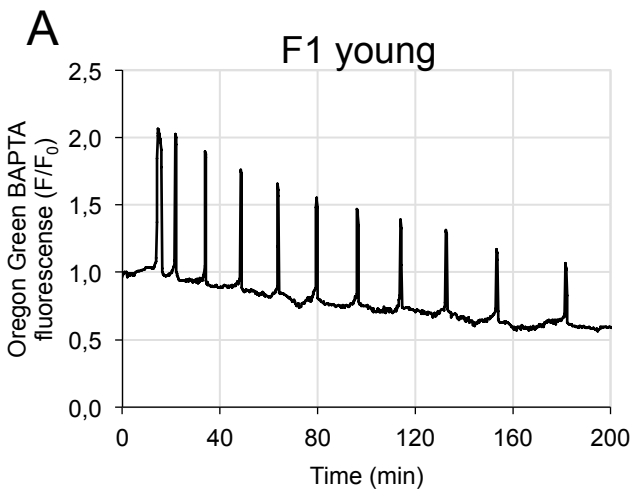
# no. of experiments conducted for the respective group of oocytes

<sup>a</sup>p<0.001, <sup>b</sup>p<0.01, <sup>c</sup>p<0.05, <sup>d</sup>p=0.05 vs. oocytes from young females of the respective background

# Supplemental Figure S1

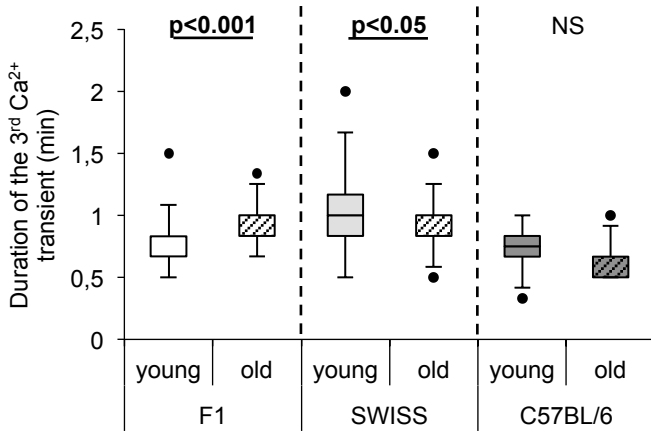


# Supplemental Figure S2

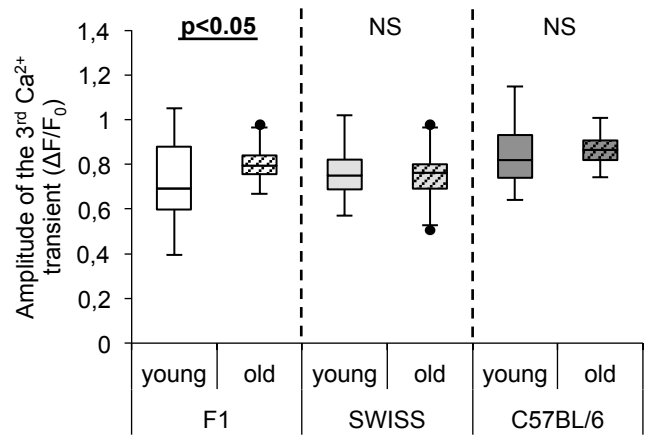


# Supplemental Figure S3

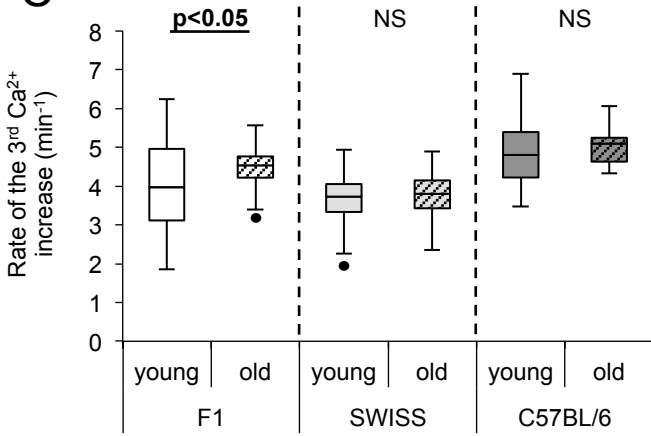
## A



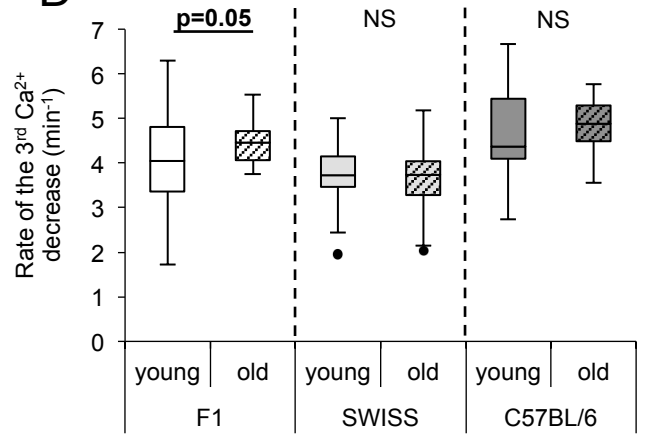
## B



## C



## D



# Supplemental Figure S4

