

1 **Identification of Tse8 as a Type VI secretion system toxin from *Pseudomonas aeruginosa***
2 **that targets the bacterial transamidosome to inhibit protein synthesis in prey cells**

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4 **Laura M. Nolan^{1*}, Amy K. Cain^{2&†}, Thomas Clamens^{1†}, R. Christopher D. Furniss^{1§},**
5 **Eleni Manoli¹, Maria A. Sainz-Polo³, Gordon Dougan², David Albesa-Jové^{3,4‡}, Julian**
6 **Parkhill^{2^}, Despoina A.I. Mavridou^{1,5#} and Alain Filloux^{1#}**

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8 ¹MRC Centre for Molecular Bacteriology and Infection (CMBI), Department of Life Sciences,
9 Imperial College London, London, SW7 2AZ, United Kingdom.

10 ²Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, Cambridge,
11 CB10 1SA, United Kingdom.

12 ³Structural Biology Unit, CIC bioGUNE, Bizkaia Technology Park, 48160 Derio, Spain.

13 ⁴IKERBASQUE, Basque Foundation for Science, Bilbao, Spain.

14 ⁵Department of Molecular Biosciences, University of Texas at Austin, Austin, 78712, Texas,
15 USA

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17 ^{*}Current address: National Heart and Lung Institute, Imperial College London, London, SW3
18 6LR, United Kingdom.

19 [&]Current address: Department of Molecular Sciences, Macquarie University, NSW 2109,
20 Australia.

21 [§]Current address: Science for Life Laboratory, Department of Molecular Biosciences, The
22 Wenner-Gren Institute, Stockholm University, Stockholm 10691, Sweden

23 [^]Current address: Department of Veterinary Medicine, University of Cambridge, Cambridge
24 CB3 0ES, United Kingdom.

25 [‡]Current address: Instituto Biofisika (UPV/EHU, CSIC), Fundación Biofísica
26 Bizkaia/Biofisika Bizkaia Fundazioa (FBB) and Departamento de Bioquímica y Biología

27 Molecular, University of the Basque Country, Leioa 48940, Spain.

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29 †These authors have contributed equally to this work

30 #Correspondence to Alain Filloux: a.filloux@imperial.ac.uk; Despoina Mavridou:

31 despoina.mavridou@austin.utexas.edu

32 **Supplementary Tables**

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34 **Legend for Supplementary Table 1**

35 **Supplementary Table 1.** Contains the full TraDIS data set (Tab 1), data above our threshold
36 cut-off ($-\log_{2}FC > 2$ $q < 0.05$) (Tab 2) and data from Tab 2 limited to genes < 600 bp (Tab 3). See
37 ‘Generation of TraDIS sequencing libraries, sequencing and downstream analysis’ in Methods
38 for details on analyses. P values were corrected for multiple testing using the Benjamini-
39 Hochberg method, and genes with a corrected P value (Q value) of < 0.05 (5% false discovery
40 rate) and an absolute \log_{2} fold change ($\log_{2}FC$) of > 2 were considered significant.

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42 **Supplementary Table 2.** Strains and plasmids used in the current study.

Stain or plasmid	Relevant characteristics	Reference/source
Strain		
<i>Escherichia coli</i>		
Dh5α	F - <i>endA1 glnV44 thi-1 recA1 relA1 gyrA96 deoR nupG purB20 φ80dlacZ ΔM15 Δ(lacZYA-argF)</i> U169, <i>hsdR17</i> (r K-mK+), λ-	Invitrogen
CC118λpir	Host strain for pKNG101 replication; Δ(<i>ara leu</i>) <i>araD ΔlacX 74 galE galK -phoA 20 thi-1 rpsE rpoB argE (Am) recA 1 Rfr</i> λpir	Laboratory collection
SM10	Host strain for pBT20 and replication; <i>thi-1 thr leu tonA lacY supE recA ::RP4-2-Tc::Mu</i> λpir , KmR	⁵
BL21 (DE3)	B F ⁻ <i>ompT gal dcm lon hsdSB(rB⁻mB⁻)</i> λ(DE3 [<i>lacI lacUV5-T7p07 ind1 sam7 nin5</i>]) [<i>malB⁺</i>] _{K-12} (λ ^S)	Invitrogen
B834	F- <i>ompT hsdSB(rB- mB-)</i> <i>gal dcm met</i> (DE3)	Invitrogen
K12	<i>supE44 hsdS20 recA13 ara-14 proA2 lacY1 galK2 rpsL20 xyl-5 mtl-1</i>	Laboratory collection
DHM1	<i>cya-854 recA1 gyrA96 (Nal) thi1 hsdR17 spoT1 rfbD1 glnV44(AS)</i>	⁶
<i>Agrobacterium tumefaciens</i>		
C58	Wild type virulent strain containing nopaline-type Ti plasmid pTiC58	Eugene Nester
<i>Pseudomonas aeruginosa</i>		
PAK	Wild type	Filloux laboratory strain
PAKΔ <i>retS</i>	<i>retS</i> deletion mutant	⁷
PAKΔ <i>retS</i> Δ <i>vgrG1a</i>	<i>retS</i> and <i>vgrG1a</i> deletion mutant	⁷
PAKΔ <i>retS</i> Δ <i>vgrG1c</i>	<i>retS</i> and <i>vgrG1c</i> deletion mutant	⁷
PAKΔ <i>retS</i> Δ <i>vgrG1a</i> Δ <i>vgrG1c</i>	<i>retS</i> , <i>vgrG1a</i> and <i>vgrG1c</i> deletion mutant	⁷

PAK Δ <i>retSΔH1</i>	<i>retS</i> and H1-T6SS cluster (encompassing the PAK genes corresponding to PA0070-PA0095) are deleted	7
PAK Δ <i>retSΔH1-H3</i>	<i>retS</i> , H1-T6SS cluster (encompassing the PAK genes corresponding to PA0070-PA0095), H2-T6SS cluster (encompassing the PAK genes corresponding to mid PA1657 to PA1662) and H3-T6SS cluster (encompassing the PAK genes corresponding to PA2357 to PA2377) are deleted	Filloux laboratory strain
PAK Δ <i>retSΔH2ΔH3</i>	<i>retS</i> , H2-T6SS cluster (encompassing the PAK genes corresponding to mid PA1657 to PA1662) and H3-T6SS cluster (encompassing the PAK genes corresponding to PA2357 to PA2377) are deleted	This study
PAK Δ <i>retSΔ<i>tseI8</i></i>	<i>retS</i> as well as PA4163 to PA4164 are deleted	This study
PAK Δ <i>retSΔ<i>tse8</i></i>	<i>retS</i> as well as PA4163 (<i>tse8</i>) are deleted	This study
PAK Δ <i>retSΔ<i>tseI8::lacZ</i></i>	<i>retS</i> as well PA4163 to PA4164 are deleted with the <i>lacZ</i> gene from miniCTX-lac inserted at the vacant <i>att</i> site on the chromosome	This study
PAK Δ <i>retS::tse8S186A</i>	<i>retS</i> deletion mutant with <i>tse8</i> (native locus) having S186A substitution	This study
PAK Δ <i>retSΔH1::<i>tse8S186A</i></i>	<i>retS</i> and H1-T6SS deletion mutant with <i>tse8</i> (native locus) having S186A substitution	This study
Plasmid		
pCR-BluntII-TOPO Blunt	Cloning vector, Zeo ^R /Km ^R	Invitrogen
pKNG101	Suicide vector, <i>sacB</i> , Str ^R	8
pKNG101: <i>tseI8</i> mutator	Mutator construct for deletion of <i>tseI8</i> by allelic exchange	This study
pKNG101: <i>tseS186A</i> mutator	Mutator construct for generating <i>tse8S186A</i> at native site on chromosome by allelic exchange	This study
pKNG101:H2-T6SS	Mutator construct for deletion of H2-T6SS by allelic exchange	Filloux laboratory collection

pKNG101:H3-T6SS	Mutator construct for deletion of H3-T6SS by allelic exchange	Filloux laboratory collection
pRK2013	Tra+, Mob+, Km ^R	9
pUX-BF13	OriR6K helper plasmid, mob/oriT, provides Tn7 transposition function in trans, Amp ^R	10
pBT20	For mariner transposon mutagenesis, Gm ^R /Amp ^R	11
pET-41a-3CD	Vector derived from pET-41a by Novagen produces fusion protein with N-terminal GST-His-S tag, Km ^R	12
pE221	Coding region of leaderless <i>E. coli</i> CcmE (S32-S163) with a C-terminal polyhistidine tag, pET22b, ApR	13
pET28a	Expression vector, Km ^R	Novagen
pET28a: <i>tse8</i>	Coding region of <i>tse8</i> in frame with N-terminal 6xHis)	This study
pET28a: <i>gatA</i> -V5	Coding region of <i>gatA</i> with C-terminal V5 tag (out of frame with 6xHis)	This study
pET41a: <i>gatA</i> -V5	Coding region of <i>gatA</i> with C-terminal V5 tag (out of frame with 6xHis)	This study
pET41a: <i>tse8</i> -HA	Coding region of <i>tse8</i> with C-terminal HA tag (out of frame with 6xHis)	This study
pET41a: <i>tse8</i> -HA-Strep	Coding region of <i>tse8</i> with C-terminal HA-Strep tag (out of frame with 6xHis)	This study
pET28a: <i>gatC</i> -HA	Coding region of <i>gatC</i> with C-terminal HA tag (out of frame with 6xHis)	This study
pET41a-3CD-TEV	pET41a as above but with a TEV site introduced to allow cleavage of N-terminal tags	This study
pET41a-3CD-TEV: <i>tse8</i>	pET41a with coding region of <i>tse8</i> in frame for (cleavable) N-terminal GST-His-S tag	This study
pACYCduet1	Dual expression vector, CmR	Novagen

pACYCduet1-His- <i>gatB</i>	Coding region of <i>gatB</i> with N-terminal His tag into MCS1	This study
pMMB67HE	Expression vector, Ap ^R	Filloux laboratory collection
pMMB67: <i>tse8</i>	Coding region of <i>tse8</i> with no tag	This study
pMMB67: <i>tse8</i> -HA	Coding region of <i>tse8</i> with C-terminal HA tag	This study
pMMB67: <i>vgrG1a</i> -V5	Coding region of <i>vgrG1a</i> (PA0091) with C-terminal V5 tag	This study
pMMB67: <i>vgrG1b</i> -V5	Coding region of <i>vgrG1b</i> (PA0095) with C-terminal V5 tag	This study
pMMB67: <i>vgrG1c</i> -V5	Coding region of <i>vgrG1c</i> (PA2685) with C-terminal V5 tag	This study
pBBR1MCS5	Expression vector, Gm ^R	14
pBBR1: <i>tsei8</i>	Coding region of <i>tsei8</i> with 500 bp upstream region to include native promoter	This study
pBBR1MCS4	Expression vector, Amp ^R	14
pBBR1: <i>tse8</i>	Coding region of <i>tse8</i> (untagged) in constitutively expressed plasmid	This study
miniCTX: <i>sfGfp</i>	sfGfp under control of constitutive <i>pX2</i> promoter with integration at vacant Tn7 site on the chromosome	Knut Drescher lab collection
pJN105	Expression vector, Gm ^R	15
pJN: <i>tsi8</i> -V5	Coding region of <i>tsi8</i> with C-terminal V5 tag	This study
pJN: <i>asnS</i> -His	Coding region of <i>asnS</i> from <i>E. coli</i> with C-terminal His tag	This study
pTrc200	Sm ^R , Sp ^R , pVS1 origin <i>lacI^q</i> , <i>trc</i> promoter expression vector	16
pTrC: <i>tse8</i> -HA	Coding region of <i>tse8</i> with C-terminal HA tag	This study
pUT18c	Vector for Bacterial Two-Hybrid assay	6

pKT25	Vector for Bacterial Two-Hybrid assay	6
pUT18c-Zip	N-terminal T18 fusion on leucine zipper of GCN4	6
pKT25-Zip	N-terminal T25 fusion on leucine zipper of GCN4	6
pUT18c- <i>tse8</i>	N-terminal T18 fusion to coding region of <i>tse8</i>	This study
pKT25- <i>tse8</i>	N-terminal T25 fusion to coding region of <i>tse8</i>	This study
pUT18c- <i>tsi8</i> -V5	N-terminal T18 fusion to coding region of <i>tsi8</i> with C-terminal V5 tag	This study
pKT25- <i>tsi8</i> -V5	N-terminal T25 fusion to coding region of <i>tsi8</i> with C-terminal V5 tag	This study
pUT18c- <i>vgrG1a</i>	N-terminal T18 fusion to coding region of <i>vgrG1a</i>	This study
pKT25c- <i>vgrG1a</i>	N-terminal T25 fusion to coding region of <i>vgrG1a</i>	This study
pUT18c- <i>vgrG1b</i>	N-terminal T18 fusion to coding region of <i>vgrG1b</i>	This study
pKT25c- <i>vgrG1b</i>	N-terminal T25 fusion to coding region of <i>vgrG1b</i>	This study
pUT18c- <i>vgrG1c</i>	N-terminal T18 fusion to coding region of <i>vgrG1c</i>	This study
pKT25c- <i>vgrG1c</i>	N-terminal T25 fusion to coding region of <i>vgrG1c</i>	This study

44 **Supplementary Table 3.** Primers used in the current study.

Primer list	Name	Oligonucleotide sequence (5'-3')	Description
OAL2939	Primer 1 <i>tsei8</i> left F	GGCATCCACGGC GCTTTCCG	Generate deletion mutant of <i>tsei8</i> primer 1
OAL2940	Primer 2 <i>tsei8</i> left R	TCAGTCGCGCTC GATCATGCTGTC ACC	Generate deletion mutant of <i>tsei8</i> primer 2
OAL2941	Primer 3 <i>tsei8</i> right F	ATGATCGAGCGC GACTGAGCGCTT	Generate deletion mutant of <i>tsei8</i> primer 3
OAL2942	Primer 4 <i>tsei8</i> right R	GCTCTACATCGG CACGTTACCC	Generate deletion mutant of <i>tsei8</i> primer 4
OAL2943	Primer 5 upstream <i>tsei8</i>	GCGTTGACCGTG ATGCCAG	Generate deletion mutant of <i>tsei8</i> primer 5
OAL2944	Primer 6 downstream <i>tsei8</i>	GAGGACCCGGCC TACTACGG	Generate deletion mutant of <i>tsei8</i> primer 6
OAL2939	Primer 1 <i>tse8</i> left F	GGCATCCACGGC GCTTTCCG	Generate deletion mutant of <i>tse8</i> primer 1
OAL4770	Primer 2 <i>tse8</i> left R	TCACTTGCTCTC GATCATGCTGTC ACC	Generate deletion mutant of <i>tse8</i> primer 2
OAL4771	Primer 3 <i>tse8</i> right F	ATGATCGAGAGC AAGTGAAAGGC GCGG	Generate deletion mutant of <i>tse8</i> primer 3
OAL4772	Primer 4 <i>tse8</i> right R	GCTTCGCCGCCT ACACCACGG	Generate deletion mutant of <i>tse8</i> primer 4
OAL2943	Primer 5 upstream <i>tse8</i>	GCGTTGACCGTG ATGCCAG	Generate deletion mutant of <i>tse8</i> primer 5

OAL4773	Primer 6 downstream <i>tse8</i>	GCGAGCGCCTGG GATTCC	Generate deletion mutant of <i>tse8</i> primer 6
OAL996	Primer 1 H2-T6SS left F	GACTGGTTGAAA ATCCTGGAAAAC	Generate deletion mutant in H2-T6SS primer 1
OAL997	Primer 2 H2-T6SS left F	TCAGGCGAACGG CCTCCTGCTGGG CGC	Generate deletion mutant in H2-T6SS primer 2
OAL998	Primer 3 H2-T6SS left F	AGGAGGCCGTTC GCCTGAGGTGGG TGC	Generate deletion mutant in H2-T6SS primer 3
OAL999	Primer 4 H2-T6SS left F	CAACACGGTATA GGGGTTGTG	Generate deletion mutant in H2-T6SS primer 4
OAL1000	Primer 5 H2-T6SS left F	GAATTGTTAAGA TATTCATTGGCG CAC	Generate deletion mutant in H2-T6SS primer 5
OAL1001	Primer 6 H2-T6SS left F	TCGAGCAGCAGG GTTCCGCCATCC GCG	Generate deletion mutant in H2-T6SS primer 6
OAL1002	Primer 1 H3-T6SS left F	ATTTCCGACATA TGGTGAAACATC	Generate deletion mutant in H3-T6SS primer 1
OAL1003	Primer 2 H3-T6SS left F	TGCTGATCAGAA GCGCAGCTCGAC GTT	Generate deletion mutant in H3-T6SS primer 2
OAL1004	Primer 3 H3-T6SS left F	CTGCGCTTCTGA TCAGCATCAACC TCT	Generate deletion mutant in H3-T6SS primer 3

OAL1005	Primer 4 H3-T6SS left F	GTGAATGGCACG AATAAATAGTTC ATA	Generate deletion mutant in H3-T6SS primer 4
OAL1006	Primer 5 H3-T6SS left F	AACTGCTGCCGG TAGTCGCGGCGG TAC	Generate deletion mutant in H3-T6SS primer 5
OAL1007	Primer 6 H3-T6SS left F	CACCCTTTCAG TAGTCGCACATC AGC	Generate deletion mutant in H3-T6SS primer 6
OAL3095	<i>tsei8</i> comp_F	GGATCCCAGCGT CTCGGCGGTTTG	Generate complementation construct of <i>tsei8</i> with native promoter (500 bp upstream of <i>tsei8</i>) (BamHI site)
OAL3096	<i>tsei8</i> comp_R	AAGCTTTCAGTC GCGCAGGGCGTA	Generate complementation construct of <i>tsei8</i> (HindIII site)
OAL3099	<i>tse8</i> comp_F	CCGGCGGATCCT AACAGGAGGAA TTAACCATGATC GAGGTCACCGAG GTT	Generate complementation of <i>tse8</i> with C-terminal V5 tag (BamHI site)
OAL3100	<i>tse8</i> comp_R	CCGGGCTCGAGT CACGTAGAATCG AGACCGAGGAG AGGGTTAGGGAT AGGCTTACCCTT GCTTGCCAGCGG TGG	Generate complementation of <i>tse8</i> with C-terminal V5 tag (XhoI site)
OAL3792	<i>tse8</i> _BTH_F	GAGCTCTCACTT GCTTGCCAGCGG	Generate <i>tse8</i> with N-terminal T18/T25 fusion in BTH vector (BamHI site)

OAL3792	<i>tse8</i> _BTH_R	GAGCTCTCACTT GCTTGCCAGCGG	Generate <i>tse8</i> with N-terminal T18/T25 fusion in BTH vector (SacI site)
OAL4774	<i>tse8</i> no tag_F	TCTAGAATGATC GAGGTCACCGAG GTT	Generate <i>tse8</i> with no tag (XbaI site)
OAL4775	<i>tse8</i> no tag_R	GAGCTCTCACTT GCTTGCCAGCGG T	Generate <i>tse8</i> with no tag (SacI site)
OAL3560	<i>tsi8</i> BTH_F	GCGCTCTAGAAT GCAGCGACTCTT CGTCTACGGCAG C	Generate <i>tse8</i> with N-terminal T18/T25 fusion and C-terminal V5 tag in BTH vector (XbaI site)
OAL3561	<i>tsi8</i> BTH_R	GGATCCTCACGT AGAATCGAGACC GAGGAGAGGGT TAGGGATAGGCT TACCGTCGCGCA GGGCGTAGAC	Generate <i>tse8</i> with N-terminal T18/T25 fusion and C-terminal V5 tag in BTH vector (BamHI site)
OAL4061	His- <i>tsi8</i> _F	GGATCCATGCAG CGACTCTTCGTC TA	Generate <i>tsi8</i> with N-terminal His tag in pET28a (BamHI site)
OAL4062	His- <i>tsi8</i> _R	GAGCTCTCAGTC GCGCAGGGCGT	Generate <i>tsi8</i> with N-terminal His tag in pET28a (SacI site)
OAL1511	Tn7- <i>glmS</i>	AATCTGGCCAAG TCGGTGAC	Combined with Tn7-RR109 produces a fragment of approx. 150bp that can be used to check any Tn7 insertion

OAL1512	Tn7-RR109	CAGCATAACTGG ACTGATTTTCAG	Combined with Tn7-glmS produces a fragment of approx. 150bp that can be used to check any Tn7 insertion
OAL3231	<i>vgrGla</i> _F	ATGCAACTGACC CGCCTGGTCCAG GTGGA	Generation of <i>vgrGla</i> (PA0091) with C-terminal V5 tag
OAL3232	<i>vgrGla</i> _R	TCAGCACGCGTA GTCCGGCACGTC GTACGGGTAGCC CTTCGCCGGCGG CGGAA	Generation of <i>vgrGla</i> (PA0091) with C-terminal V5 tag
OAL3233	<i>vgrGlb</i> _F	ATGGCACTTGCG CAACAGACCCGC CTGGT	Generation of <i>vgrGlb</i> (PA0095) with C-terminal V5 tag
OAL3234	<i>vgrGlb</i> _R	TCAGCACGCGTA GTCCGGCACGTC GTACGGGTAGTT CTGGAGGATCTT GCGT	Generation of <i>vgrGlb</i> (PA0095) with C-terminal V5 tag
OAL3235	<i>vgrGlc</i> _F	GTGGCTATTGGC CAGCCTTTCGCG ACGGC	Generation of <i>vgrGlc</i> (PA2685) with C-terminal V5 tag
OAL3236	<i>vgrGlc</i> _R	TCAGCACGCGTA GTCCGGCACGTC GTACGGGTAACA GTTGATATCGAC ATTGG	Generation of <i>vgrGlc</i> (PA2685) with C-terminal V5 tag

OAL1740	<i>vgrG1a</i> BTH_F	GCGCGGGATCCC ATGCAACTGACC CGCCTG	Generate <i>vgrG1a</i> with N-terminal T18/T25 fusion in BTH vector (BamHI site)
OAL1741	<i>vgrG1a</i> BTH_R	GCGCGGAATTCT CAGCCCTTCGCC GGCGG	Generate <i>vgrG1a</i> with N-terminal T18/T25 fusion in BTH vector (EcoRI site)
OAL1860	<i>vgrG1b</i> BTH_F	GCGCGTCTAGAG ATGGCACTTGCG CAACAGACC	Generate <i>vgrG1b</i> with N-terminal T18/T25 fusion in BTH vector (XbaI site)
OAL1861	<i>vgrG1b</i> BTH_R	GCGCGGAATTCT CAGTTCTGGAGG ATCTTGCG	Generate <i>vgrG1b</i> with N-terminal T18/T25 fusion in BTH vector (EcoRI site)
OAL2390	<i>vgrG1c</i> BTH_F	GCGCGTCTAGAA TGCAACACACCC GCCTGGTACACG	Generate <i>vgrG1c</i> with N-terminal T18/T25 fusion in BTH vector (XbaI site)
OAL2391	<i>vgrG1c</i> BTH_R	GCGCGGAATTCT CAACAGTTGATA TCGACATTGGGC	Generate <i>vgrG1c</i> with N-terminal T18/T25 fusion in BTH vector (EcoRI site)
OAL2458	<i>vgrG6</i> BTH_F	GCGCGTCTAGAA TGTCGCCCCCG CCAACCAGACGC	Generate <i>vgrG6</i> with N-terminal T18/T25 fusion in BTH vector (XbaI site)
OAL2459	<i>vgrG6</i> BTH_R	GCGCGGAATTCT CATGGCGTGGGC TCATCCTTGTCG	Generate <i>vgrG6</i> with N-terminal T18/T25 fusion in BTH vector (EcoRI site)
OAL3099	<i>tse8</i> -HA_F	CCGGCGGATCCT AACAGGAGGAA TTAACCATGATC	Generate <i>tse8</i> -HA (BamHI site and Shine-Dalgarno)

		GAGGTCACCGAG GTT	
OAL3310	<i>tse8</i> -HA_R	GCCGGCTGCAGT CAGCACGCGTAG TCCGGCACGTCG TACGGGTACTTG CTTGCCAGCGGT GGAG	Generate <i>tse8</i> -HA (PstI site)
OAL3669	<i>tse8</i> exp_F	GCGCGGATCCAT CGAGGTCACCGA GGTTTC	Generate <i>tse8</i> with N-terminal GST-His-S tag for purification (BamHI site)
OAL3670	<i>tse8</i> exp_R	GCGGCCGCTCAC TTGCTTGCCAGC GG	Generate <i>tse8</i> with N-terminal GST-His-S tag for purification (NotI site)
OAL5140	<i>tse8</i> -HA-Strep_F	CATATGATCGAG GTCACCGAGGTT TCCATCGCCGAG CTGCGTG	Generate <i>tse8</i> with C-terminal HA and Strep tags (NdeI site) with pET41a: <i>tse8</i> -HA as template; use with primer <i>gatC</i> -HA-Strep ^R
OAL3301	<i>tsi8</i> -V5_F	CCGCCTCTAGAT AACAGGAGGAA TTAACCATGCAG CGACTCTTCGTC TAC	Generate <i>tsi8</i> with C-terminal V5 tag (XbaI site and Shine-Dalgarno)
OAL3302	<i>tsi8</i> -V5_F	CTGCAGTCACGT AGAATCGAGACC GAGGAGAGGGT TAGGGATAGGCT TACCCGTAGAAT	Generate <i>tsi8</i> with C-terminal V5 tag (PstI site)

		CGAGACCGAGG AGAGGGTTAGG GATAGGCTTACC GTCGCGCAGGGC GTAGA	
OAL3273	<i>gatA</i> -V5_F	CCGGCCTCGAGT AACAGGAGGAA TTAACCATGCTG CATCAATTGACC CT	Generate <i>gatA</i> with C-terminal V5 tag (XhoI site and Shine-Dalgarno)
OAL3274	<i>gatA</i> -V5_R	CCGGGATGCATT TACGTAGAATCG AGACCGAGGAG AGGGTTAGGGAT AGGCTTACCGAA GCCGGCCGGGGT GCG	Generate <i>gatA</i> with C-terminal V5 tag (NsiI site)
OAL3275	<i>gatB</i> _F	GCGCGGGATCCG CAATGGGAAACC GTGATC	Generate <i>gatB</i> with N-terminal His tag (BamHI site)
OAL3276	<i>gatB</i> _R	AAGCTTTCACGC TTCGAGCTTTTT	Generate <i>gatB</i> with N-terminal His tag (HindIII site)
OAL3277	<i>gatC</i> -HA_F	CATATGGCGCTT GAACGCTCCGAC	Generate <i>gatA</i> with C-terminal HA tag (NdeI site)
OAL3278	<i>gatC</i> -HA_R	AATTACTCGAGT CAGCACGCGTAG TCCGGCACGTCG	Generate <i>gatA</i> with C-terminal HA tag (XhoI site)

		TACGGGTATGAC TCGATGACTTTC GG	
OAL3913	<i>asnS</i> -His_F	GCGCGCTGCAGT AACAGGAGGAA TTAACCATGAGC GTTGTGCCTGTA GCCGA	Generate <i>asnS</i> (asparagine tRNA synthase from <i>E. coli</i> MG1655) with C-terminal His tag (SpeI site and Shine-Dalgarno)
OAL3914	<i>asnS</i> -His_R	GGCGCACTAGTT TAGTGATGGTGA TGGTGATGGAAG CTGGCGTTACGC GG	Generate <i>asnS</i> (asparagine tRNA synthase from <i>E. coli</i> MG1655) with C-terminal His tag (NotI site)

46 **Supplementary Table 4.** Output of bioinformatic analyses of relevant bacteria possessing
 47 GatCAB, AsnS and GlnS.

Transamidosome (GatCAB Components)		
GatA (pBLAST)	GatB (pBLAST)	GatC (pBLAST)
<i>Agrobacterium tumefaciens</i>	<i>Agrobacterium tumefaciens</i>	<i>Agrobacterium tumefaciens</i>
<i>Pseudomonas aeruginosa</i>	<i>Pseudomonas aeruginosa</i>	<i>Pseudomonas aeruginosa</i>
Operons containing gatCAB (nBLAST)		
<i>Agrobacterium tumefaciens</i>		
<i>Pseudomonas aeruginosa</i>		
Cognate tRNA synthases (AsnS and/or GlnS)		
AsnS (pBLAST)	GlnS (pBLAST)	
<i>Escherichia coli</i>	<i>Escherichia coli</i>	
	<i>Pseudomonas aeruginosa</i>	

48

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