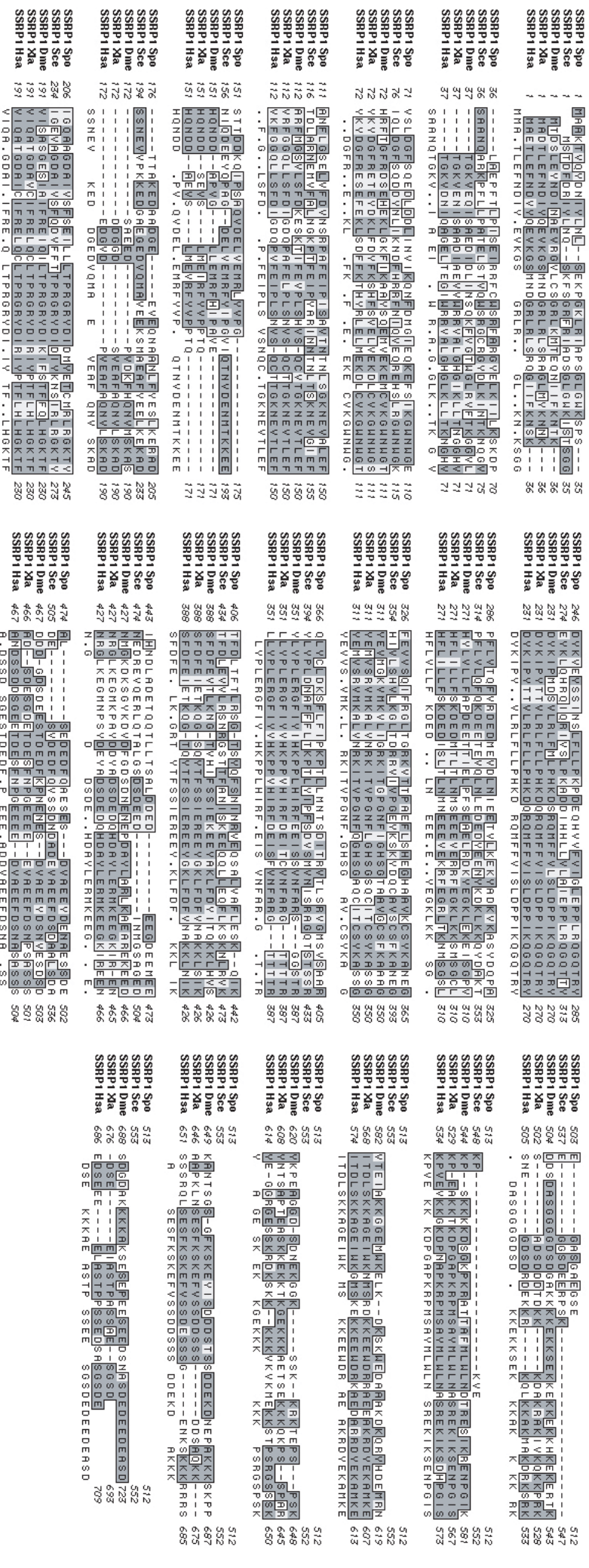


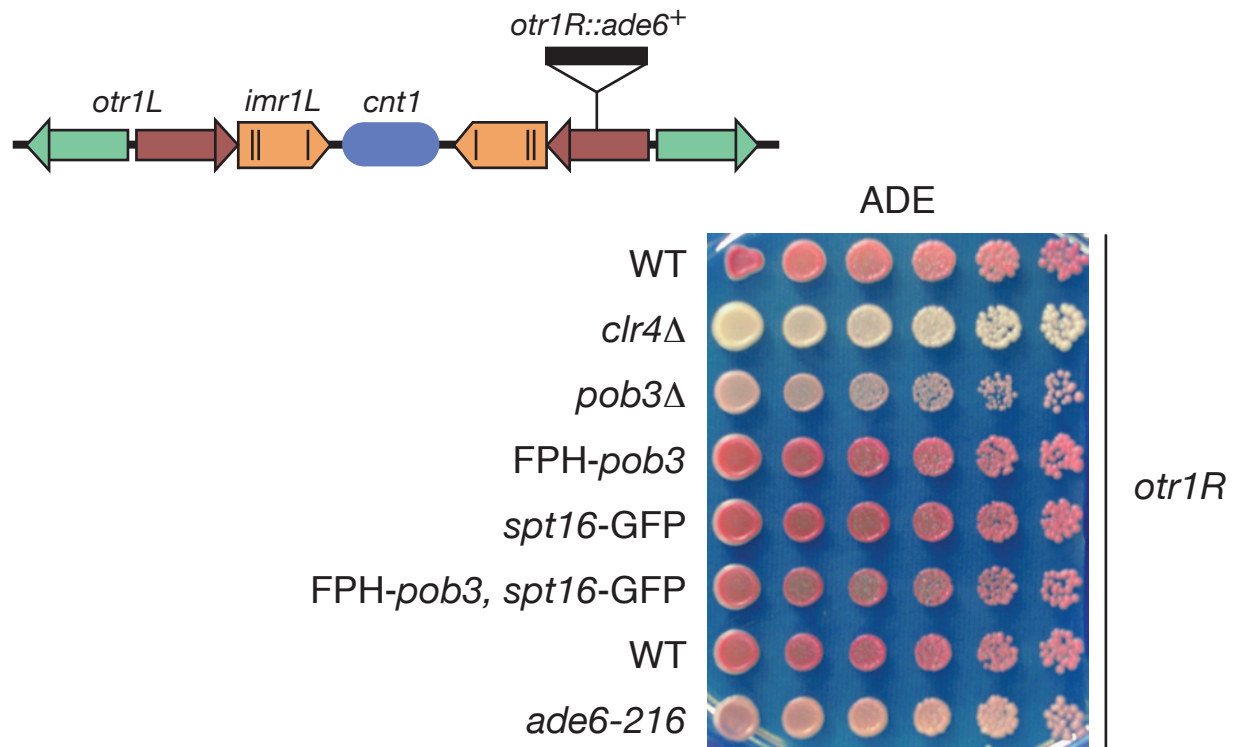


Figure S1. (B) Primary sequence alignments between *S. pombe* Pob3 (SSRP1) and select eukaryotic orthologues

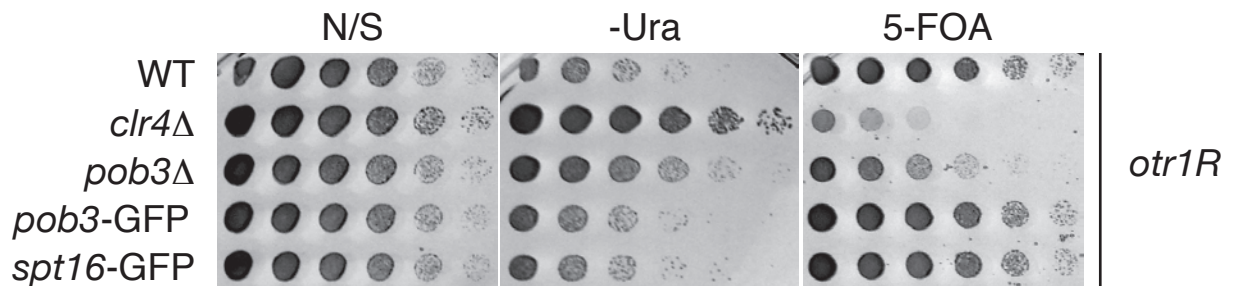


## Figure S2

**A**



**B**

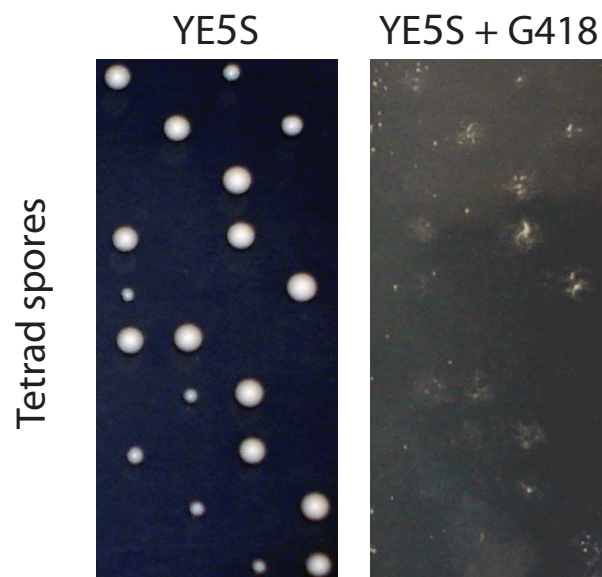


**Figure S2. Tagged isoforms of the *S. pombe* Pob3 and Spt16 proteins are functional.**

(A) Tagged FPH-Pob3, Spt16-GFP and double tagged strains are functional in the ADE-colorimetric centromere silencing assay.

(B) Pob3-GFP and Spt16-GFP fusion proteins do not affect centromeric silencing. Serial dilutions of wild-type (WT), *clr4Δ*, *pob3Δ*, *pob3*-GFP and *spt16*-GFP strains.

## Figure S3

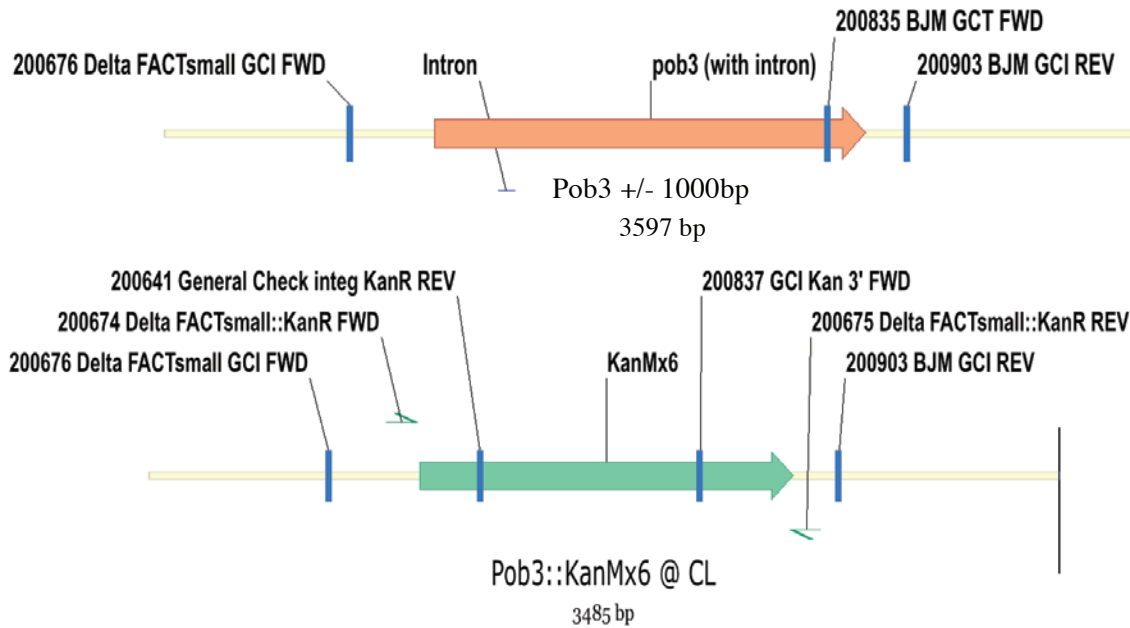


### Figure S3. *spt16+* is an essential gene.

Tetrads dissected from the diploid *spt16::kanMX6/spt16+* were grown on YE5S media at 32 degrees Celsius for 2 days. All growing spore clones were G418 sensitive.

**Figure S4**

**Strain EL44 = *pob3+::KanMX6* construction & verification**



(a) Primers to amplify the KanR deletion cassette for *pob3+::KanMX6* strain

**200674 Delta FACTsmall::KanR FWD**

AATGTGTTAATGTGTGAATTTGAGAATAACCTGGTATAGCTGTTTTCTGACCA  
 AATACTTAATACTTCACCAGTTATAAAAAATGCGGATCCCCGGGTTAATTAA  
 (includes the *pob3*'s ATG) anneals on the Kanamycin-containing plasmid

**200675 Delta FACTsmall::KanR REV**

AATTTGTAATAGCAAAAGACAAGGCTGAAAAGTTGATCCAAATAGACTTCCA  
 ATTTAACATTGCTCAATTTTATAGATCATTAGAATTCGAGCTCGTTTAAAC  
 (includes the *pob3*'s TAA) anneals on the Kanamycin-containing plasmid

(b) Primers to check *pob3+::KanMX6* deletion by PCR on the 5' end and 3' end

**200676 Delta FACTsmall GCI FWD**

GCT GAA CCT CGT TCC TCA G

**200641 General Check integ KanR REV**

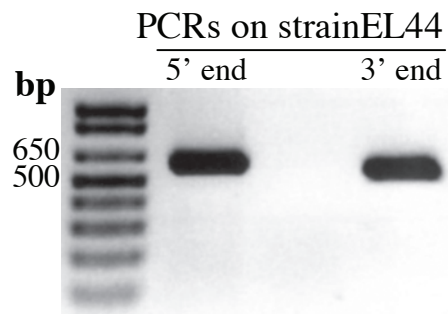
CAA TTC AAC GCG TCT GTG AGG

**200837 GCI KAN 3' FWD**

TAT GGA ACT GCC TCG GTG AG

**200903 BJM GCI REV**

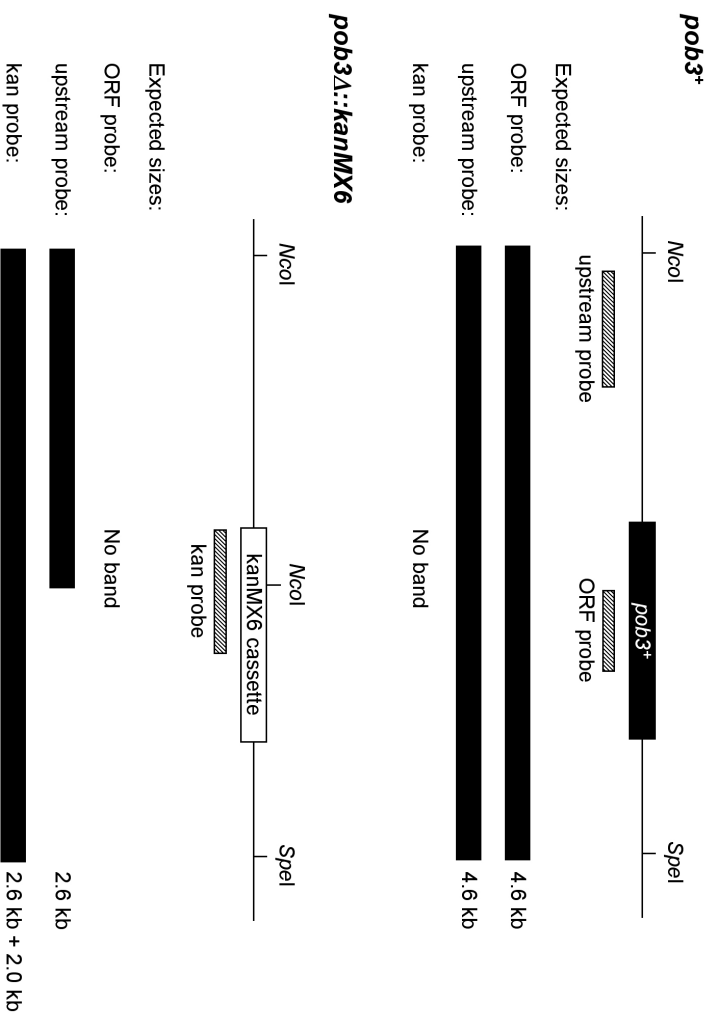
TTT ACC ACT CGG ATT CAT CG



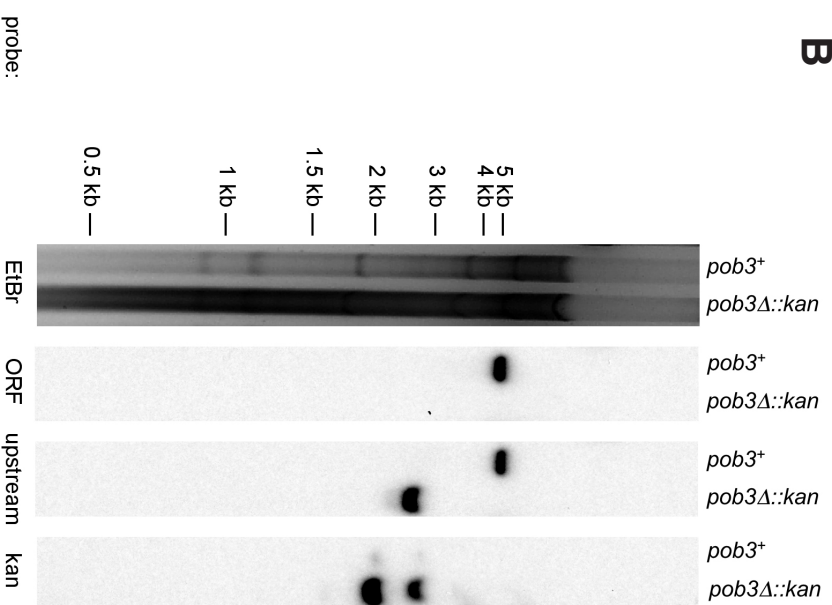
5' end (200676/200641) expected amplicon: 604bp

3' end (200903/200837) expected amplicon: 552bp

A



B

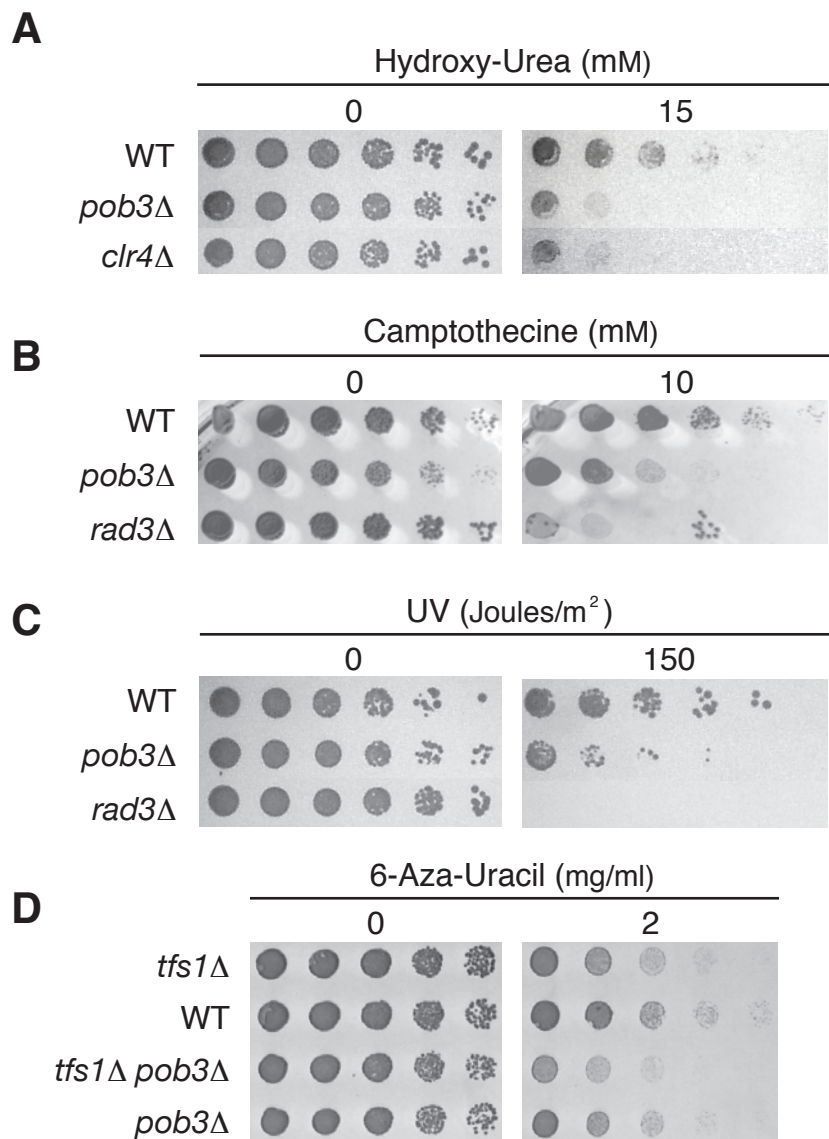


**Figure S5. Southern blot verification of *pob3+* gene deletion in *S. pombe*.**

(A) Diagram showing the wild-type *pob3+* strain and *pob3Δ::kanMX6* knockout strain, including the probes and expected sizes of DNA fragments obtained by restriction enzymes *NcoI* and *SpeI*.

(B) Southern blot for *pob3+* and *pob3Δ::kanMX6* strains using the indicated radiolabelled ORF, upstream and kan probes.

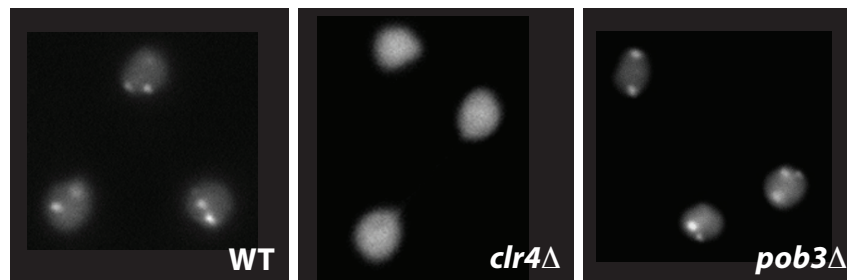
## Figure S6



**Figure S6. The small subunit of the *S. pombe* chromatin remodeling complex FACT (Pob3) has a role in genome stability.**

Sensitivity of wild-type, *pob3*Δ deletion and reference yeast strains to:  
(A) Hydroxyurea (HU), which reveals defects in DNA replication.  
(B) camptothecine (CPT) and (C) ultra-violet (UV) radiation, both of which reveal defects in controlling induced DNA damage.  
(D) 6-aza-uracil (6AU), a small molecule that affects intracellular GTP levels and is commonly used to screen for mutants affecting transcript elongation.

## Figure S7



**Figure S7.** GFP-Swi6 delocalization assay.

GFP-Swi6 is functional and a good marker for centromeres and telomeres (Pidoux *et al.*, 2000).

GFP-Swi6 fluorescence images of wild-type (WT), *clr4Δ* and *pob3Δ* cells. GFP-Swi6 is delocalized from centromeres and telomeres in *clr4Δ* mutants, as opposed to WT and *pob3Δ* cells.



**Table S1. Gene expression profiling of a *pob3Δ* strain relative to wild-type *S. pombe*.**

<b>LOWER expression in <i>pob3Δ</i> versus WT. 2-fold cut-off (3 of 4 data points):</b>	
15 genes (total)	
<b>Gene Name</b>	<b>Gene dB Product description</b>
SPCC965.14c	putative cytidine and deoxycytidylate deaminase zinc-binding protein
SPAC644.05c	putative deoxyuridine 5'-triphosphate nucleotidohydrolase (EC 3.6.1.23); dUTP pyrophosphatase
<i>rps10-2</i>	40S ribosomal protein S10
<i>soul</i>	sorbitol utilization protein soul1; short chain dehydrogenase
SPAC926.05c	dnaj protein
SPBC609.05 = <i>pob3</i>	similar to <i>S. cerevisiae</i> POB3 protein that binds to DNA polymerase I; putative structure specific recognition protein
SPAC1B3.16c	MFS transporter of unknown specificity
SPAC5D6.05	Protein that plays a role in cell separation, mating and differentiation
SPAC694.03	hypothetical protein; similar to <i>S. cerevisiae</i> YCL047C
SPAC29B12.04	putative pyridoxine (vitamin B6) biosynthetic enzyme; by similarity to yeast <i>snz1</i>
SPCC622.19	hypothetical protein; (conserved in fungi- Gaillardin et al)
SPBC146.08c	putative elongation initiation factor eIF1a
SPBP23A10.03c	hypothetical protein; similar to <i>S. cerevisiae</i> YDR511W
<i>vti1</i>	putative vesicle transport V-snare protein
SPBC1709.06	conserved hypothetical protein; UPF0034 family
<b>HIGHER expression in <i>pob3Δ</i> versus WT. 2-fold cut-off (3 of 4 data points):</b>	
44 genes (total)	
<b>Gene Name</b>	<b>Gene dB Product description</b>
<i>fiol1</i>	iron transport multi-copper oxidase precursor
<i>hsp9</i>	heat shock protein hsp9
<i>hsp16</i>	heat shock protein hsp16
SPAC29A4.06c	hypothetical protein; contains Pfam-B_33047; shares conserved region with <i>Drosophila</i> CG15747; <i>Arabidopsis</i> F12K2.14; and <i>C. elegans</i> C16C10.6; predicted coiled-coil
<i>cox1</i>	
<i>lys7</i>	putative alpha-aminoadipate reductase small chain, sixth step in lysine biosynthesis pathway- from ACPS domain
<i>fip1</i>	iron permease
<i>frp1</i>	ferric reductase transmembrane componen
SPAC212.06c	pseudogene; truncated C-terminal of DNA helicase in rearranged telomeric region
SPAC212.08c	hypothetical protein; possibly <i>S. pombe</i> specific; duplicated in telomeric region; contains N-term predicted signal sequence
SPAC212.11	DNA helicase possibly pseudo
SPAC1F8.03c	MFS efflux transporter of unknown specificity
SPAC750.07c	very hypothetical protein
SPAC1834.10c	hypothetical protein; sequence orphan

SPAC1782.01	similar to <i>S. cerevisiae</i> Ecm29 cell wall stucture/biosynthesis protein
SPAC22F8.12c	hypothetical protein; serine-rich protein; sequence orphan; predicted N-terminal signal sequence
<i>pim1</i>	putative peptide transporter
SPAC1002.19	putative GTP cyclohydrolase; riboflavin biosynthesis
SPCC338.12	putative proteinase precursor
SPAC9E9.04	conserved hypothetical protein; putative endoplasmic reticulum transmembrane protein; similar to <i>S. cerevisiae</i> YET1; contains predicted N-term signal sequence; contains 2 predicted transmembrane helices; contains predicted C-term coiled-coil
SPCC965.07c	Glutathione S-transferase
<i>srp14</i>	signal recognition particle 14 kda protein
SPAC11H11.05c	very hypothetical protein
SPAC1002.17c	probable uracil phosphoribosyltransferase
SPBPJ4664.02	hypothetical protein; sequence orphan; low similarity to glycoamylases and other cell surface proteins; contains ~250-270 copies of a 13 AA repeat, NSSTPITSSSIL; contains predicted N-terminal signal sequence
SPAC22F8.05	putative alpha,alpha-trehalose-phosphate synthase
SPBC1921.06c	hypothetical protein; sequence orphan; contains predictd N-term signal sequence
SPBC27B12.03c	putative c-5 sterol desaturase
SPCC1739.08c	short chain dehydrogenase; putative sorbitol utilization
SPAPB1A11.03	putative FMN dependent dehydrogenase; similar to lactate dehydrogenase
SPAC19G12.09	putative aldose reductase
SPBC557.06c	
<i>leu2</i>	3-isopropylmalate dehydratase
<i>rpn10</i>	19s proteasome regulatory subunit
<i>klp8</i>	kinesin-like protein
SPAC12B10.10	hypothetical protein; sequence orphan; predicted C-term coiled-coil region
<i>sec24</i>	putative component of the COPII coat; involved in endoplasmic reticulum to Golgi transport
<i>cdc16</i>	cell division control protein 16; spindle assembly checkpoint component; GTPase-activating protein (GAP) for Spg1p GTPase; essential;two-component GEF for the GTPase spg1 (with byr4)
SPBPB2B2.01	putative amino-acid permease
SPAC637.03	hypothetical protein; sequence orphan; contains 8 predicted transmembrane segments contains predicted N-term signal sequence
SPBC21D10.10	bromodomain protein
SPBC887.16	very hypothetical protein
SPAC27D7.05c	hypothetical protein; sequence orphan; has transcript from microarray
SPAC27D7.06	putative electron transfer flavoprotein alpha-subunit precursor

**Table S2. Hyper-geometric probability comparisons of genes either up- or down-regulated in *pob3* deletion cells.**

<b>HIGH expression in <i>pob3Δ</i> cut 2.0 (3 of 4)</b>		<b>44 genes in list</b>		
<b>P-value</b>	<b>Similar List Name</b>	<b>Genes in list</b>	<b>Genes in overlap</b>	<b>Reference for gene list</b>
1,07e-7	HIGH expression in <i>rpb7-G150D</i> cut 1.5 (6 of 8)	62	10	Djupedal <i>et al</i> (2005), <i>Genes Dev</i> , 19, 2301
2,56e-6	HIGH expression in <i>chr3D</i> cut 1.5	61	9	Hansen <i>et al</i> (2005), <i>Mol Cell Biol</i> , 25, 590
0,000136	HIGH expression in pSwi6 cut 1.5 (3 of 4)	254	13	Wiren <i>et al</i> (2005), <i>EMBO J</i> , 17, 2906
0,00162	HIGH expression in <i>chr6-1</i> cut 2.0 (6 of 10)	63	7	Wiren <i>et al</i> (2005), <i>EMBO J</i> , 17, 2906
0,00318	HIGH expression in <i>chr6-1 chr3D</i> cut 1.5	570	17	Hansen <i>et al</i> (2005), <i>Mol Cell Biol</i> , 25, 590
0,00472	HIGH expression in <i>chr1D</i> cut 1.5	47	8	Hansen <i>et al</i> (2005), <i>Mol Cell Biol</i> , 25, 590
0,00518	HIGH expression in <i>chr4-681</i> cut1.5	107	8	Hansen <i>et al</i> (2005), <i>Mol Cell Biol</i> , 25, 590
0,0109	LOW IGR WT H4K16Ac H3 Cter corr cut 1.5	1267	24	Wiren <i>et al</i> (2005), <i>EMBO J</i> , 17, 2906
0,0115	LOW IGR WT H4K5Ac H3 Cter corr cut 1.5	1110	22	Wiren <i>et al</i> (2005), <i>EMBO J</i> , 17, 2906
0,0115	HIGH expression in <i>chr6-1</i> (6 of 10) cut 1.5	253	11	Wiren <i>et al</i> (2005), <i>EMBO J</i> , 17, 2906
0,0309	LOW IGR WT H4K12Ac H3 Cter corr cut 1.5	1083	21	Wiren <i>et al</i> (2005), <i>EMBO J</i> , 17, 2906
0,0356	HIGH expression in <i>chr3-735</i> cut1.5	183	9	Hansen <i>et al</i> (2005), <i>Mol Cell Biol</i> , 25, 590

**LOW expression in *pob3Δ* cut 2.0 (3 of 4)**

**15 genes in list**

<b>P-value</b>	<b>Similar List Name</b>	<b>Genes in list</b>	<b>Genes in overlap</b>	<b>Ref. for gene list</b>
0,0229	LOW expression in <i>rpb7-G150D</i> cut 1.5 (6 of 8)	47	4	Djupedal <i>et al</i> (2005), <i>Genes Dev</i> , 19, 2301

*For methods section:* Similar gene lists were identified using the automatic hyper-geometric distribution tests in the gene list inspector function of Gene Spring v7.2 software (Silicon Genetics). The hyper-geometric distribution test calculates the probability of overlap corresponding to *k* or more genes between a list of *n* genes compared against another gene list of *m* fragments when randomly sampled from a universe of *u* genes:

$$\frac{1}{\binom{u}{m}} \sum_{i=k}^n \binom{m}{i} \binom{u-m}{n-i}$$

Table S3

## Strain list (main figures)

Strain	Genotype
EL18	h+ leu1-32,ade6-216, ura4D18, imr1R(Nco)::ura4+,
EL32	h+ clr4::kanMX6+, imr1R(Nco)::ura4+, leu1-32, ade6-216, ura4D18
EL44	h+ pob3::kanMX6+, imr1R(Nco)::ura4+, ade6-216, leu1-32, ura4D18
EL87	h- pob3::kanMX6, imr1R(Nco)::ura4+, leu1-32, ade6-210, ura4D18
EL93	h+ pob3::kanMX6, otr1R(SphI)::ura4, ade6-210, his3D1?, leu1-32, ura4DS/E
EL102	h? pob3::kanMX6, mat3M::ura4, ade6-210, arg3D4, leu1-32, ura4D18
EL105	h? pob3::kanMX6, (rDNA:ura4) Ylp2.4pUCura4.7, ade6-210, leu1-32, his3D1, ura4DS/E,
EL107	h? pob3::kanMX6, otr2:ura4+ ?, TM1:arg3+, TM3:ade6+ ?, tel:his3+, ade6-210, arg3D4, leu1-32, his3D1, ura4D18/DS/E ?
EL127	h- spt16-GFP-kanMX6, otr1R (SphI)::ura4, arg3D4, his3D1, leu1-32, ura4DS/E
EL130	h- pob3-GFP-kanMX6, otr1R (SphI)::ura4, arg3D4, his3D1, leu1-32, ura4DS/E
EL238	h- pob3::natMX6, ade6-210, arg3D4, his3D1, leu1-32, ura4DS/E
EL243	h? pob3::kanMX6, otr1R(SphI):ade6, ade6-210, leu1-32, ura4D18/DS-E?
EL306	h+ FPH-pob3, otr1R(SphI):ade6, ade6-210, leu1-32, ura4-D18
EL319	h? pob3::kanMX6, dcr1::natMX6, otr1R:ura4, ade6-210, his?, leu?, ura4D18/DS-E?
EL332	h? pob3::kanMX6, [CM3112 sup3e], ade6-704, arg3D4?, leu1-32, his3D1, ura4-D18
EL344	h? FPH-pob3, spt16-GFP-kanMX6, otr1R(SphI):ade6, ade6-210, arg3D4?, his3D1?, leu1-32, ura4D18
EL348	h? pob3::natMX6, [Ch16 ade6-216 m23::ura4], ade6-210, arg3D4?, his3D1?, leu1-32, ura4DS/E
FY511	h90 mat3-M::ura4, ade6-216, leu1-32, ura4-D18
FY521	h- [Ch16 ade6-216 m23::ura4], ade6-210, leu1-32, ura4DS/E
FY1034	h? swi6::his1, otr1R (dg-glu BamHI-Spe1 fragment) SphI::ura4, ade6-210, his1-102, leu1-32, ura4-DS/E
FY1180	h+ otr1R(dg-glu)SphI:ade6, ade6-210, leu1-32, ura4-D18
FY2116	h- [CM3112 sup3e], ade6-704, his3D1, leu1-32, ura4-D18
FY3027	h+ TM1::arg3, TM3::ade6, tel::his3, otr2::ura4, ade6-210, arg3D4, his3D1, leu1-32, ura4-D18/DS-E?
FY3606	h? rik1::leu2, TM1::arg3, TM3::ade6, otr2::ura4, his3::tel, ade6-210, arg3D4, his3D1, leu1-32, ura4D18
FY4134	h- ade6-210, arg3D4, his3D1, leu1-32, ura4DS/E
FY4841	h- otr1R(SphI)::ura4, arg3D4, his3D1, leu1-32, ura4DS/E
FY6084	h- clr4::LEU2+, otr1R(SphI):ura4 ura4D/SE, leu1-32, ade6-210/216?
FY6603	h+ Ylp2.4pUCura4.7, ura4-DS/E, leu1-32, ade6-216
FY6802	h+ tas3::kanMX6, imr1R(Nco1)::ura4+, ade6-216, leu1-32, ura4D18/DS-E?
FY7036	h- dcr1::natMX6, otr1R(dg-glu)SphI:ade6, ade6-210, his3D1?, leu1-32?, ura4D18/DS-E?

## Strain list (Supplementary figures)

Strain	Genotype
EL63	h+ ade6-210 leu1-32
EL90	h+ pob3::kanMX6, ade6-210, arg3D4, his3D1, leu1-32, ura4D18
EL98	h? pob3::kanMX6, ars1(Mlu)::pREP81X-gfp-swi6-LEU2+, ade6-210/216?, leu1-32, ura4-D18
EL111	h+ pob3::kanMX6, ade6-210, leu1-32
EL115	h? pob3::kanMX6, tfs1::LEU2+, ade6-210, leu1-32, can1-1?
EL173	h+ tfs1::natMX6, ade6-210, leu1-32
EL229	h? clr4::LEU2+ otr1R(Sph1):ade6+, ade6-210/216?, leu1-32, ura4D18/DS-E?
EL345	h? FPH-pob3, spt16-GFP-kanMX6, otr1R(dg-glu)SphI:ade6, ade6-210, arg3D4?, his3D1?, leu1-32, ura4D18
EL351	h? spt16-GFP-KanMX6, otr1R(dg-glu)Sph1:ade6, ade6-210, arg3D4?, his3D1?, leu1-32, ura4D18/DS-E?
EL355	h+/h+ spt16::KanMX6/ORF, ade6-M210/ade6-M216, ura4D18/ura4D18, leu1-32/leu1-32
FY1665	h- rad3::ura4+, ade6-704, leu1-32, ura4-D18
FY2214	h- ars1(Mlu)::pREP81Xgfpswi6 LEU2+, ade6-210, leu1-32, ura4-D18
FY3300	h? clr4 :: ura4 ars1(Mlu)::pREP81Xgfpswi6-LEU2+, ade6-210/216?, leu1-32, ura4D18/DS-E?