- 174 ZMZ conceptualised the paper, cleaned and analysed the data and drafted the manuscript, JT and TC
- reviewed the manuscript for intellectual content, EM guided the whole paper structure development. Allauthors reviewed the final manuscript for submission.

177 **Conflict of Interest**

178 The authors declare no competing interests for this manuscript.

179 Appendix 1: Statistical software and challenges

180 In addition to the use of msm package in R which can handle maximum likelihood estimation (MLE) multistate models, the other library within R software is the *tdc.msm* library developed by Meira-Machado 181 et al. (2009) which can fit five different multistate models including time-homogeneous and non-182 homogeneous Markov multistate models; and Cox Markov and Cox semi-Markov multistate models (1). 183 The tdc.msm library is a comprehensive package for modelling multistate longitudinal data since different 184 models can be fitted within one library, and model comparison can be done easily (1). For nonparametric 185 estimation, the msSurv library within R can be used to estimate state occupation probabilities, initial and 186 exiting time in a state, and the marginal integrated transition rate for the non-Markov multistate process 187 188 (49). The other R package is the *mstate* developed by Wreede *et al.* (2011) for both the competing risk models and multistate models (50). Additional R libraries are the *etm* library Allignol *et al.* (2011) for 189 190 empirical transition probabilities and the *changeLOS* (change length of hospital stay) library introduced by 191 Wangler et al. 2006) for the Aalen-Johansen Estimator is implemented within R software. The limitation 192 of the *changeLOS* library is that it does not support the inclusion of covariates in the multistate model and 193 left truncated data (51). However, the *mvna* library can handle both left truncated and right censored 194 multistate data (51).

- 195 The STATA software (licensed for use) (52) can fit the MLE multistate models using the multistate model 196 ado files developed by Cowther and Lambert (2016) which restructures and declares the multistate data as survival and any survival model within STATA can be used (53). This package can estimate each transition 197 198 rate by its unique model structure, assuming either a Markov or semi-Markov process (53). Uniquely to the 199 STATA models is the ability to estimate each transition rate assuming different hazard functions which best 200 fit the transition as compared to the R msm models which assumes the same hazard function on all the 201 model transition processes. Another option in STATA is using *illdprep* and *stpm2illd* commands which can 202 perform a similar analysis as described elsewhere (54).
- The Bayesian estimation (BE) multistate models can be implemented in BayesX software using the *bayesreg* object. This package can handle several data features which the MLE cannot handle like nonlinear effects of continuous covariates, time-varying effects, adjust for individual and spatial random effects accounting for unobserved heterogeneity. The WINBUGS software which estimates using Gibbs Sampling also can handle BE multistate model through the use of Kolmogorov-Chapman forward equations. This modelling approach promotes the use of partially observed aggregated data which the MLE approach cannot do as illustrated in the main text.

210 Most of these statistical packages which can handle multistate models, both from the MLE and BE are free while few may require licensing. Here we highlight a couple of strengths and limitations associated with 211 each software type which handles multistate models, some of which might not have been discussed in detail 212 in this manuscript. Firstly, most of the existing software assumes the Markov property and time-213 homogeneous by default which makes it difficult if these assumptions are violated. Secondly, the Markov 214 215 assumption can be difficult to test, and in most cases, studies are silent on pre-model assumption testing of 216 the Markov property; however, markovchain library implemented in R is one of the packages which can test for Markov property (26). Thirdly, not all software types are freely available for use like BayesX, 217

218 WinBUGS and R, some of the software types require a license to be granted access, for instance, STATA 219 and SAS. Fourthly, data argumentation (structure) is different in each of the software, which may be a major 220 drawback if one wishes to make a comparison across the software. In addition to this, at times convergence 221 is an issue in multistate models depending on the size of the data, the model complexity of the proposed model (number of states, reversible transition and number of covariates included) and the preferred method 222 223 of estimation (BE or MLE) as the models may take much longer (hours or days) to converge or may fail to converge at all. Lastly and most importantly, the result outputs vary moving from one software package to 224 another; hence, methodological background theory for each package is crucial to enable results comparison. 225 226 For instance, in the R *msm* library, one gets the exact transition rate values which can be interpreted directly while in BayesX outputs, the estimates are based on the flexible predictor on a log scale (39). 227

228 Appendix 2: WinBUGs three state reversible multistate model code

```
229
        ###MODEL
230
        model {
231
        #####Multinomial likelihood for observed data
232
                for (i in 1:2)
233
                   {
234
                   r[i,1:3] ~ dmulti(P[i,1:3],n[i])
235
                   }
236
        #####Find transition probabilities (for given time) in terms of rates
237
238
        h<- sqrt(pow(lambda[1]-lambda[2], 2) + 4*G[1,2]*G[2,1])
239
        e1<-exp(-.5*(lambda[1] + lambda[2] - h)*(2*t.obs))
240
        e2 < exp(-.5*(lambda[1] + lambda[2] + h)*(2*t.obs))
241
242
        P[1,1] < -((-lambda[1]+lambda[2]+h)*e1+ (lambda[1]-lambda[2]+h)*e2)/(2*h)
243
        P[1,2]<-((-lambda[1]+lambda[2]+h)*(lambda[1]-lambda[2]+h)*(e1-e2))/(4*h*G[2,1]) P[1,3]<-1-P[1,1] - P[1,2]
244
        P[2,1]<-G[2,1]*(e1-e2)/h
245
        P[2,2] <- ((lambda[1]-lambda[2]+h)*e1+ (-lambda[1] + lambda[2] + h)*e2)/(2*h)
246
        P[2,3]<-1 - P[2,1] - P[2,2]
247
248
        #Give exponential priors for unknown transition rate parameters
249
               for (i in 1:2)
250
                  {
251
                for (j in (i+1):3)
252
253
                             G[i,j] \sim dexp(.001)
254
255
                  }
256
               for (i in 2:2)
257
                  {
258
               for (j in 1:(i-1))
259
260
                          G[i,j] ~ dexp(.001)
261
262
                  }
263
        lambda[1]<- G[1,2] + G[1,3]
264
        lambda[2]<-G[2,1]+G[2,3]
265
266
        ######Find P(t.new) for given new time of interest=1 years
267
        e1.new < exp(-.5*(lambda[1] + lambda[2] - h)*(2*t.new))
```

- 268 e2.new<-exp(-.5*(lambda[1] + lambda[2] + h)*(2*t.new))
- 269 Pt[1,1]<-((-lambda[1]+lambda[2]+h)*e1.new+ (lambda[1] -lambda[2]+h)*e2.new)/(2*h)
- 270 Pt[1,2]<-((-lambda[1]+lambda[2]+h)*(lambda[1]-lambda[2]+h)*(e1.new-e2.new))/(4*h*G[2,1])
- 271 Pt[1,3]<- 1-Pt[1,1] Pt[1,2]
- 272 Pt[2,1]<- G[2,1]*(e1.new-e2.new)/h
- 273 Pt[2,2]<- ((lambda[1]-lambda[2]+h)*e1.new+ (-lambda[1] + lambda[2] + h)*e2.new)/(2*h)
- 274 Pt[2,3]<- 1-Pt[2,1] Pt[2,2]
- 275 ##### P(t.final)=half a year (put 0.25 in the data)
- 276 e1.final<-exp(-.5*(lambda[1] + lambda[2] h)*(2*t.final))
- 277 e2.final<-exp(-.5*(lambda[1] + lambda[2] + h)*(2*t.final))
- 278 Ptn[1,1]<-((-lambda[1]+lambda[2]+h)*e1.final+ (lambda[1] -lambda[2]+h)*e2.final)/(2*h)
- 279 Ptn[1,2]<-((-lambda[1]+lambda[2]+h)*(lambda[1]-lambda[2]+h)*(e1.final-e2.final))/(4*h*G[2,1])
- 280 Ptn[1,3]<-1 Ptn[1,1] Ptn[1,2]
- 281 Ptn[2,1]<- G[2,1]*(e1.final-e2.final)/h
- 282 Ptn[2,2]<- ((lambda[1]-lambda[2]+h)*e1.final+ (-lambda[1] + lambda[2] + h)*e2.final)/(2*h)
- 283 Ptn[2,3]<-1 Ptn[2,1] Ptn[2,2]
- 284 }
- 285 ###DATA
- 286 iist(r=structure(.Data=c(2269,143,78,137,2882,87),.Dim=c(2,3)),n=c(2490,3106),t.obs=0.5,
- t.new=0.25,

- 287 t.final=0.125)
- 288
- 289 ###INITIAL VALUES
- 290 list(G=structure(.Data=c(NA,.1,.1,.1,NA,.1),.Dim=c(2,3)))
- 291 Here is an **R** code for comparable results if one has individual-level data
- 292 #StepVL1-is the name of the dataset as saving name
- 293 #ID-is the unique identifier
- 294 #States-is the variable which defines individual states
- 295 #Time1-if the cumulative time
- 296 library(haven)
- 297 StepVL1 <- read_stata("C:/Users/ WinBUGS/VL_KOL_1.dta")
- 298 statetable.msm(States, ID, data=StepVL1)
- 299 twoway3.q <- rbind(c(0.25, 0.5, 0.25), c(0.25, 0.5, 0.25), c(0, 0, 0))
- 300 crudeinits.msm(States ~ Time1, ID, data=StepVL1, qmatrix=twoway3.q)
- 301 StepVL1.msm <- msm(States ~ Time1, subject=ID, data = StepVL1,
- 302 qmatrix = twoway3.q, deathexact = 3,
- 303 control = list (fnscale = 5000, maxit= 500))
- 304 ###To get the transition rates
- 305 StepVL1.msm
- 306 ###To get the transition probabilities
- 307 P_1year<-pmatrix.msm(StepVL1.msm,t=1)
- 308 P_6months<-pmatrix.msm(StepVL1.msm,t=0.5)
- 309 P_3months<-pmatrix.msm(StepVL1.msm,t=0.25)
- 310

311 **REFERENCES**

- Meira-Machado L, Uña-álvarez J De, Cadarso-suárez C, Andersen PK. Multi-state models
 for the analysis of time-to-event data. Statistical methods in Medical Research.
 2009;18(2):195–222.
- 2. Hougaard P. Multi-state Models : A Review. Lifetime Data Analysis. 1999;264:239–64.