



Figure S1

## Supplementary Information

### A functional microRNA screen uncovers O-linked N-acetylglucosamine transferase as a host factor modulating hepatitis C virus morphogenesis

Katharina Herzog<sup>1,2\*</sup>, Simonetta Bandiera<sup>1,2\*</sup>, Sophie Pernot<sup>1,2</sup>, Catherine Fauvelle<sup>1,2</sup>, Frank Jühling<sup>1,2</sup>, Amélie Weiss<sup>2,3,4,5</sup>, Anne Bull<sup>6</sup>, Sarah C. Durand<sup>1,2</sup>, Béatrice Chane-Woon-Ming<sup>2,7</sup>, Sébastien Pfeffer<sup>2,7</sup>, Marion Mercey<sup>8</sup>, Hervé Lerat<sup>8</sup>, Jean-Christophe Meunier<sup>6</sup>, Wolfgang Raffelsberger<sup>2,3,4,5</sup>, Laurent Brino<sup>2,3,4,5</sup>, Thomas F. Baumert<sup>1,2,9,#</sup>, Mirjam B. Zeisel<sup>1,2,10,#</sup>

<sup>1</sup>Inserm, U1110, Institut de Recherche sur les Maladies Virales et Hépatiques, Strasbourg, France; <sup>2</sup>Université de Strasbourg, Strasbourg, France; <sup>3</sup>Institut de Génétique et de Biologie Moléculaire et Cellulaire, Illkirch, France; <sup>4</sup>CNRS, UMR7104, Illkirch, France; <sup>5</sup>Inserm, U1258, Illkirch, France; <sup>6</sup>Inserm U1259, Faculté de Médecine, Université François Rabelais and CHRU de Tours, Tours, France; <sup>7</sup>Architecture et Réactivité de l'ARN – UPR 9002, Institut de Biologie Moléculaire et Cellulaire du CNRS, Strasbourg, France; <sup>8</sup>Institute for Advanced Biosciences, Centre de Recherche UGA - Inserm U1209 - CNRS UMR 5309, Grenoble, France; <sup>9</sup>Institut Hospitalo-Universitaire, Pôle Hépato-digestif, Hôpitaux Universitaires de Strasbourg, Strasbourg, France; <sup>10</sup>Inserm, U1052, CNRS UMR 5286, Centre Léon Bérard (CLB), Cancer Research Center of Lyon (CRCL), Université de Lyon (UCBL), Lyon, France

\*Authors contributed equally to this work

#### Supplementary Material and methods

**Cells and cell culture conditions.** The source and culture conditions of Huh7.5.1 cells have been described[1]. HeLa cells were purchased from ATCC and cultured in Dulbecco's modified Eagle medium (Gibco® DMEM GlutaMAX™, ThermoFisher Scientific) containing 1% sodium pyruvate as described for Huh7.5.1 cells[1].

1  
2  
3 1 **Viruses and infectivity assays.** Cell culture-derived recombinant cell culture-derived  
4 hepatitis C virus (HCVcc) Jc1 (genotype 2a/2a chimera), H77R2a (genotype 1a/2a chimera  
5 engineered for *Renilla* luciferase expression), Con1R2a (genotype 1b/2b chimera engineered  
6 for *Renilla* luciferase expression), and JcR2a (genotype 2a/2a chimera engineered for *Renilla*  
7 luciferase expression) were generated in Huh7.5.1 cells as described[1, 2, 3, 4]. HCVcc  
8 infectivity was determined by calculating the 50% tissue culture infectious dose (TCID<sub>50</sub>)  
9 using anti-NS5A antibody as described[5, 6] or by assessing luciferase activity. HCVcc were  
10 used at 10<sup>5</sup>-10<sup>6</sup> TCID<sub>50</sub>/mL throughout the study. HCV RNA was purified using a QIAmp viral  
11 RNA minikit (Qiagen) and analyzed by one-step RT-qPCR using a Sensi Fast NO ROX kit  
12 (Bioline) according to the manufacturer's instructions. Standard curves were performed using  
13 10-fold dilution series of HCV RNA.

14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28 13 **Purification of HCVcc particles using sucrose cushion or iodixanol density gradient.**  
29 HCVcc (JcR2a) were concentrated 10-fold using a Vivaspin column (GE Healthcare). For  
30 sucrose cushion purification, HCVcc were purified by overlaying 3.5 mL of culture media on  
31 1.5 mL of 20% sucrose, and by ultracentrifuging samples for 4h at 40,000 rpm on a SW-55  
32 rotor (Beckman Coulter). Purified HCVcc were resuspended in 30 µL of PBS for analysis via  
33 immunocapture and electron microscopy. Density distributions of infectious HCVcc were  
34 determined by overlaying 0.5 mL culture media on a 5 mL, 4%-40% iodixanol step gradient,  
35 and ultracentrifuging samples for 16h at 40,000 rpm on a SW-55 rotor (Beckman Coulter): 625  
36 µL fractions were carefully harvested from the top of each tube, and density was determined  
37 by weighing. Infectivity of each fraction was quantified by TCID<sub>50</sub> using anti-NS5A antibody  
38 as described[5, 6], while HCV RNA of fractions was purified and analyzed as described above.  
39 ApoB and ApoE concentrations of fractions were determined by enzyme-linked  
40 immunosorbent assay (Human Apolipoprotein B or E ELISA<sup>PRO</sup> kit, Mabtech) undiluted or in a  
41 1:50 dilution, respectively, according to the manufacturer's instructions (Mabtech).

52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 1 **miRNA mimics and siRNAs.** Non-targeting control miRNA, miR-501-3p mimic, miR-619-3p  
4 mimic, antagomiR-122, antagomiR-501-3p, non-targeting control antagomiR, non-targeting  
5 control siRNA, siRNAs targeting *OGT*, *CD81* or *apolipoprotein E (ApoE)* and a library of 28  
6 custom ON-TARGETplus smart pool siRNAs were purchased from Dharmacon (GE  
7 Healthcare).

8  
9  
10 7 **miRNA expression analysis.** Total RNA (100 ng) was purified from control or HCV-infected  
11 Huh7.5.1 cells using Tri reagent® (Thermo Scientific) and Direct-zol™ RNA purification kit  
12 (Zymo Research). Total RNA was first polyadenylated and reverse transcribed using a  
13 miScript II RT system (Qiagen) according to the manufacturer's instructions. The obtained  
14 cDNA was subjected to RT-qPCR using miScript SYBR Green kit (Qiagen). Primers were the  
15 mature miRNA sequence for the forward primer (Thermo Scientific) and the universal miScript  
16 primer (Qiagen) for the reverse primer. Data were analyzed by the  $\Delta\Delta C_t$  method using small  
17 nucleolar RNA, C/D box 61 (SNORD61) as an endogenous reference and the non-infected  
18 samples as a calibrator[7].

19  
20  
21 17 **Antibodies.** Rabbit anti-OGT antibodies DM-17 and AL24 were purchased from Sigma or  
22 kindly provided by Dr. G. W. Hart and Dr. S. Hardivillé (Johns Hopkins University School of  
23 Medicine, Baltimore, MD)[8], respectively. Mouse anti- $\beta$ -actin antibody was purchased from  
24 Abcam and mouse, rabbit or sheep HRP-conjugated secondary antibodies (A9044, A0545  
25 and A3415, respectively) were purchased from Sigma. Sheep anti-NS5A serum for  
26 determination of TCID50 was a kind gift from M. Harris[9]. Human anti-E2 (AR3A) antibody[10]  
27 for electron microscopy analysis was kindly provided by Mansun Law (SCRIPPS, California,  
28 USA).

29  
30  
31 26 **Western blotting.** OGT and actin protein expression in human cells was assessed by  
32 Western blot as described[8] with some modifications. Briefly, cells were lysed in lysis buffer  
33 no. 6 (R&D Systems) according to the manufacturer's instructions. Equal amounts of protein

1 (40 µg) were size-separated through a Mini PROTEAN® TGX Stain-Free™ gel electrophoresis  
2 (Bio-Rad) and transferred to PVDF membranes (Bio-Rad). Immunoblots were performed  
3 using rabbit anti-OGT (1:2000) and mouse anti-β-actin (1:1000) antibodies[8, 11]. Antigen-  
4 antibody complexes were detected by incubating the membrane with the appropriate HRP-  
5 conjugated secondary antibodies (1:5000; 1:10,000) and imaged by enhanced  
6 chemiluminescence with a ChemiDoc MP imager (Bio-Rad). Quantification of protein  
7 expression was performed using ImageLab™ 5.2.1 software (BioRad). For analysis of OGT  
8 and GAPDH expression in liver tissue from HCV transgenic (FL-N/35) or wild-type mice[12],  
9 crude protein extracts were prepared by homogenization of frozen mouse livers (50–100 µg)  
10 in tissue lysis buffer from the Ambion PARIS RNA (Thermo Scientific) and protein isolation kit,  
11 supplemented with protease inhibitors (cOmplete™ EDTA-free protease inhibitor mixture,  
12 Sigma-Aldrich) and phosphatase inhibitors (PhosSTOP™, Sigma-Aldrich), using a tissue  
13 homogenizer (MP Fast Prep24, MP Biomedicals, Santa Ana, CA) and MP Lysing Matrix A  
14 tubes. Proteins were quantified using the BCA assay (Thermo Fisher Scientific). Western  
15 blotting was performed as described above.

16  
17 **Immunocapture and electron microscopy analysis of viral particles.** Sucrose-cushion  
18 purified or iodixanol gradient fractionated HCVcc (JcR2a) produced in cells transfected with a  
19 non-targeting siRNA control or a pool of siRNA against OGT were transferred via anti-E2  
20 antibody AR3A on electron microscopy (EM) grids through immunocapture (IC) as  
21 described[13]. Particles were stained with uranyl acetate dihydrate and observed in a JEOL  
22 1230 electron microscope. Series of electron micrographs were acquired at random from IC  
23 EM grids. The images were then analyzed with Image-J software, to determine the particle  
24 size distribution.

25  
26 **Gene expression analysis in patient-derived liver tissue.** For OGT expression analysis in  
27 patient's samples, raw data were retrieved from the Gene Expression Omnibus (GSE84346)  
28 and re-analyzed by quality-trimming (cutadapt) and mapping (HISAT2) to human genome

1 assembly hg19. Reads mapping to Gencode v.19 genes were counted using htseq-count and  
2 normalized applying DESeq2. Activity and fibrosis scores as well as viral load were taken from  
3 the supplemental data published in[14]. To analyze OGT expression in liver tissue of chronic  
4 hepatitis B or C patients, FPKM values and clinical data were retrieved from The Cancer  
5 Genome Atlas (TCGA, [https://www.cancer.gov/about-](https://www.cancer.gov/about-nci/organization/ccg/research/structural-genomics/tcga)  
6 [nci/organization/ccg/research/structural-genomics/tcga](https://www.cancer.gov/about-nci/organization/ccg/research/structural-genomics/tcga)). This data set includes samples from  
7 39 HCV-infected patients, 83 hepatitis B virus (HBV)-infected, 80 patients with alcoholic liver  
8 disease (ALD) and 13 patients with non-alcoholic fatty liver disease (NAFLD).

### 10 **Supplementary References**

- 11 1 Zhong J, Gastaminza P, Cheng G, *et al.* Robust hepatitis C virus infection in vitro. Proc  
12 Natl Acad Sci U S A 2005;**102**:9294-9.
- 13 2 Reiss S, Rebhan I, Backes P, *et al.* Recruitment and activation of a lipid kinase by  
14 hepatitis C virus NS5A is essential for integrity of the membranous replication compartment.  
15 Cell Host Microbe 2011;**9**:32-45.
- 16 3 Da Costa D, Turek M, Felmlee DJ, *et al.* Reconstitution of the entire hepatitis C virus  
17 life cycle in non-hepatic cells. J Virol 2012;**86**:11919-25.
- 18 4 Fauvelle C, Felmlee DJ, Crouch E, *et al.* Apolipoprotein E Mediates Evasion From  
19 Hepatitis C Virus Neutralizing Antibodies. Gastroenterology 2016;**150**:206-17 e4.
- 20 5 Lindenbach BD, Evans MJ, Syder AJ, *et al.* Complete replication of hepatitis C virus in  
21 cell culture. Science 2005;**309**:623-6.
- 22 6 Bandiera S, Pernet S, El Saghire H, *et al.* Hepatitis C Virus-Induced Upregulation of  
23 MicroRNA miR-146a-5p in Hepatocytes Promotes Viral Infection and Deregulates Metabolic  
24 Pathways Associated with Liver Disease Pathogenesis. J Virol 2016;**90**:6387-400.
- 25 7 Schmittgen TD, Livak KJ. Analyzing real-time PCR data by the comparative C(T)  
26 method. Nat Protoc 2008;**3**:1101-8.
- 27 8 Iyer SP, Akimoto Y, Hart GW. Identification and cloning of a novel family of coiled-coil  
28 domain proteins that interact with O-GlcNAc transferase. J Biol Chem 2003;**278**:5399-409.

- 1  
2  
3 1 9 Macdonald A, Crowder K, Street A, *et al.* The hepatitis C virus non-structural NS5A  
4 protein inhibits activating protein-1 function by perturbing ras-ERK pathway signaling. *J Biol*  
5 *Chem* 2003;**278**:17775-84.  
6  
7  
8  
9 4 10 Giang E, Dorner M, Prentoe JC, *et al.* Human broadly neutralizing antibodies to the  
10 envelope glycoprotein complex of hepatitis C virus. *Proc Natl Acad Sci U S A* 2012;**109**:6205-  
11 10.  
12  
13  
14  
15 7 11 Verrier ER, Colpitts CC, Bach C, *et al.* A targeted functional RNA interference screen  
16 uncovers glypican 5 as an entry factor for hepatitis B and D viruses. *Hepatology* 2016;**63**:35-  
17 48.  
18  
19  
20  
21  
22 10 12 Lerat H, Honda M, Beard MR, *et al.* Steatosis and liver cancer in transgenic mice  
23 expressing the structural and nonstructural proteins of hepatitis C virus. *Gastroenterology*  
24 2002;**122**:352-65.  
25  
26  
27  
28 13 13 Piver E, Boyer A, Gaillard J, *et al.* Ultrastructural organisation of HCV from the  
29 bloodstream of infected patients revealed by electron microscopy after specific  
30 immunocapture. *Gut* 2017;**66**:1487-95.  
31  
32  
33  
34 16 14 Boldanova T, Suslov A, Heim MH, *et al.* Transcriptional response to hepatitis C virus  
35 infection and interferon-alpha treatment in the human liver. *EMBO Mol Med* 2017;**9**:816-34.  
36  
37  
38  
39  
40

#### 19 **Supplementary figure legends**

20 **Figure S1.** (A) Effect of miR-501-3p inhibition on HCV infectivity. Huh7.5.1 cells were  
21 transfected with control antagomiR (Ctrl), antagomiR-122 as loss-of-function control to perturb  
22 HCV replication and antagomiR-501-3p, prior to infection with HCVcc (JcR2a) according to  
23 the two-step protocol depicted in Fig. 1A. After 48h, supernatants were transferred onto naive  
24 Huh7.5.1 cells. After 72h, Renilla Luciferase activity of infected Huh7.5.1 cells was  
25 determined. Data are expressed as mean percentage as compared to Ctrl  $\pm$  s.d. Results are  
26 from four independent experiments in quadruplicate. The dashed line indicates values from  
27 vehicle-treated cells set at 100%. Statistics: \*,  $p$ -value < 0.05, Mann-Whitney test. (B) miR-  
28 501-3p expression upon HCV infection. Huh7.5.1 cells were infected with HCVcc (JcR2a).

1  
2  
3 1 After 72h, RNA was purified and miR-501-3p expression analyzed by RT-qPCR. Percentage  
4  
5 2 of miR-501-3p expression relative to uninfected Huh7.5.1 cells (Ctrl). Results are presented  
6  
7 3 as mean  $\pm$  s.d. from three independent experiments in duplicate. The dashed line indicates  
8  
9 4 values from uninfected Huh7.5.1 cells set at 100%. Statistics: \*,  $p$ -value < 0.05, Mann-Whitney  
10  
11 5 test.  
12  
13  
14 6

15  
16 7 **Supplementary Table 1. A genome-wide miRNA mimic screen identifies cellular**  
17  
18 8 **miRNAs modulating HCV infection.** Log<sub>2</sub>(FC), lfd<sub>r</sub> and effect on HCV infection in part 1 and  
19  
20 9 part 2 of the screen are shown for the individual miRNAs of the miRNA mimic library. In red:  
21  
22 10 proviral effect, in blue: antiviral effect. FC: fold change, lfd<sub>r</sub>: local false discovery rate  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60





Table with columns: ID, Gene Name, Coordinates, RefSeq ID, Accession, Strand, Type, Log P Value, Odds Ratio, and other metrics.



1	hsa-miR-6504-3p	MIMAT0025465	C-302672-00	CAUUACAGCACAGCCAUUCU	-0.219268298279075	0.49143	FALSE	-2.3682663259571	0.08542	TRUE
2	hsa-miR-6505-3p	MIMAT0025467	C-302675-00	UGACUUCUACCCUUCUCAAAG	-1.41016990839787	0.00054279	FALSE	-2.92045878806631	0.040079	TRUE
3	hsa-miR-6507-5p	MIMAT0025470	C-302679-00	GAAGAAUAGGAGGACUUUGU	-0.637513482741369	0.03164	FALSE	-2.6469629700191	0.057617	TRUE
4	hsa-miR-6514-3p	MIMAT0025485	C-302693-00	CUGCCUGUUCUCCACUCCAG	-1.57253367763337	0.00023985	TRUE	-0.857463656453242	0.72813	FALSE
5	hsa-miR-652-3p	MIMAT0022709	C-301928-00	CAACCCUAGGAGAGGUGCCAUUCA	-0.590578760859739	0.01741	FALSE	-2.0413301671663	0.11887	TRUE
6	hsa-miR-654-5p	MIMAT0003330	C-300988-01	UGUGGGCCGGAACAUGUGC	-1.24102088307823	0.00054279	FALSE	-4.37321715649902	0.0018396	TRUE
7	hsa-miR-659	MIMAT0003337	C-300994-01	CUUGUUCAGGAGGGUCCCA	-0.865338332757254	0.0053924	FALSE	-2.2970254587848	0.12226	TRUE
8	hsa-miR-660-3p	MIMAT0022711	C-301897-00	ACCUCUGUGGCAUGGUAUA	-0.914364872105563	0.0018747	FALSE	-2.7989676696545	0.028781	TRUE
9	hsa-miR-661	MIMAT0003324	C-300981-01	UGCCUGGUCUCUGCCUGCGCGU	-0.966061272920722	0.0046172	FALSE	-2.04789510522588	0.11887	TRUE
10	hsa-miR-664b-3p	MIMAT0022772	C-302539-00	UUGAAUUGCCUCCAGCCUACA	-0.63588937093853	0.061384	FALSE	-2.38779084523873	0.057617	TRUE
11	hsa-miR-671-5p	MIMAT0003880	C-301000-03	AGGAAGCCUUGGAGGGGUGGAG	-1.01268622344544	0.00023985	TRUE	-0.995050180707598	0.5811	FALSE
12	hsa-miR-6721-5p	MIMAT0025852	C-302707-00	UGGGCAGGGGCUUUGGAGGAG	-1.0522716652067	0.0021076	FALSE	-2.86092601806193	0.028781	TRUE
13	hsa-miR-7	MIMAT0000252	C-300546-07	UGGAAGACUAGUAGUUUGUUGU	-0.833080606802588	0.0014577	FALSE	-2.34382127982937	0.11887	TRUE
14	hsa-miR-718	MIMAT0012735	C-301486-00	CUUCGCGCCGCGGCGUCCG	-0.712476270533885	0.0014577	FALSE	-2.51688917044679	0.08542	TRUE
15	hsa-miR-744	MIMAT0004945	C-301242-01	UGCGGGCUAGGCUUACAGCA	-1.31292964878657	0.0027607	FALSE	-2.17130853543699	0.057617	TRUE
16	hsa-miR-744*	MIMAT0004946	C-301241-01	CUUGUCCGCUAACCCUAAACCU	-0.309425861300587	3.9917e-05	TRUE	-2.41920375944258	0.11887	TRUE
17	hsa-miR-764	MIMAT0010367	C-301640-00	GCAGGUGUCACUUGUCCUCCU	-1.15960521227396	6.1693e-05	TRUE	-0.963991799367289	0.5811	FALSE
18	hsa-miR-767-3p	MIMAT0003883	C-301003-01	UCUGUCUACUCCUAGGUAUUCU	-0.875815222995269	0.0021076	FALSE	-2.87852305265755	0.009387	TRUE
19	hsa-miR-801	MIMAT0005202	C-301014-01	GAUUGCUGGCGGCGGAUUCGAC	-1.02838938735747	0.00023985	TRUE	0.128066636696993	1	FALSE
20	hsa-miR-873-3p	MIMAT0022717	C-301921-00	GGAGACUGAUGAUUCCCGGGA	-0.45500701580676	0.050037	FALSE	-2.17559760678154	0.08542	TRUE
21	hsa-miR-888*	MIMAT0004917	C-301227-01	GACUGACACCUUUGGUGUAA	-0.798719536763374	0.015067	FALSE	-1.83668949541677	0.11887	TRUE
22	hsa-miR-940	MIMAT0004983	C-301269-01	AAGGCAGGGCCCGCCUCCCC	-1.31504996360232	0.00054279	FALSE	-2.52010733193133	0.11887	TRUE
23	hsa-miR-96	MIMAT0000095	C-300514-07	UUUGCCAGUAGCACAUUUUGCU	-0.788957233607577	0.0074787	FALSE	-2.19234338350236	0.11527	TRUE
24	hsa-miR-99a	MIMAT0000097	C-300516-03	AACCGUAGAUCCGAAUUCUGU	-0.986802247020601	0.0030028	FALSE	-2.42723567464877	0.11887	TRUE

**Legend**

Antiviral effect

Proviral effect

Note: Hits were selected using the following Local False Discovery Rate (fdr) thresholds: 0.00027 in screen part 1 and 0.1226 in screen part 2. Positive log<sub>2</sub> fold change values indicate a proviral effect of the miRNA on the HCV life cycle, while negative log<sub>2</sub> fold change values indicate an antiviral effect.

1  
2  
3 1 **A functional microRNA screen uncovers O-linked N-acetylglucosamine transferase as**  
4  
5 2 **a host factor modulating hepatitis C virus morphogenesis and infectivity**  
6  
7 3

9 4 Katharina Herzog<sup>1,2\*</sup>, Simonetta Bandiera<sup>1,2\*</sup>, Sophie Pernot<sup>1,2</sup>, Catherine Fauvelle<sup>1,2</sup>, Frank  
11 5 Jühling<sup>1,2</sup>, Amélie Weiss<sup>2,3,4,5</sup>, Anne Bull<sup>6</sup>, Sarah C. Durand<sup>1,2</sup>, Béatrice Chane-Woon-Ming<sup>2,7</sup>,  
13 6 Sébastien Pfeffer<sup>2,7</sup>, Marion Mercey<sup>8</sup>, Hervé Lerat<sup>8</sup>, Jean-Christophe Meunier<sup>6</sup>, Wolfgang  
15 7 Raffelsberger<sup>2,3,4,5</sup>, Laurent Brino<sup>2,3,4,5</sup>, Thomas F. Baumert<sup>1,2,9,#</sup>, Mirjam B. Zeisel<sup>1,2,10,#</sup>  
17 8

19 9 <sup>1</sup>Inserm, U1110, Institut de Recherche sur les Maladies Virales et Hépatiques, Strasbourg,  
21 10 France; <sup>2</sup>Université de Strasbourg, Strasbourg, France; <sup>3</sup>Institut de Génétique et de Biologie  
23 11 Moléculaire et Cellulaire, Illkirch, France ; <sup>4</sup>CNRS, UMR7104, Illkirch, France ; <sup>5</sup>Inserm,  
25 12 U1258, Illkirch, France ; <sup>6</sup>Inserm U1259, Faculté de Médecine, Université François Rabelais  
27 13 and CHRU de Tours, Tours, France ; <sup>7</sup>Architecture et Réactivité de l'ARN – UPR 9002,  
29 14 Institut de Biologie Moléculaire et Cellulaire du CNRS, Strasbourg, France; <sup>8</sup>Institute for  
31 15 Applied Biosciences, Centre de Recherche UGA - Inserm U1209 - CNRS 5309, Grenoble,  
33 16 France <sup>9</sup>Institut Hospitalo-Universitaire, Pôle Hépatodigestif, Hôpitaux Universitaires de  
35 17 Strasbourg, Strasbourg, France; <sup>10</sup>Inserm, U1052, CNRS UMR 5286, Centre Léon Bérard  
37 18 (CLB), Cancer Research Center of Lyon (CRCL), Université de Lyon (UCBL), Lyon, France

39 19 \*Authors contributed equally to this work  
41 20

43 21 **Word count.** Abstract: 196 words; main manuscript: 3991 words; 55 references; 7 figures; 1  
45 22 table; Supplementary information (including 1 Supplementary Table and 1 Supplementary  
47 23 Figure)  
49 24

51 25 **#Corresponding authors.** Dr. Mirjam B. Zeisel, Inserm U1052 – CRCL, 151 cours Albert  
53 26 Thomas, 69424 Lyon Cedex 03, France, Phone: +33472681970, Fax: +33472681971, E-  
55 27 mail: mirjam.zeisel@inserm.fr and Prof. Thomas F. Baumert, Inserm U1110, Institut de  
59 60

1  
2  
3 1 Recherche sur les Maladies Virales et Hépatiques, 3 rue Koeberlé, 67000 Strasbourg,  
4  
5 2 France, Phone: +33368853703, Fax: +33368853724, Email: thomas.baumert@unistra.fr  
6  
7 3

8  
9 4 **Financial support.** This work was supported by the European Union (INTERREG-IV-Rhin  
10  
11 5 Supérieur-FEDER-Hepato-Regio-Net 2012 to T.F.B. and M.B.Z., ERC-AdG-2014-671231-  
12  
13 6 HEPCIR, EU H2020-667273-HEPCAR to T.F.B.), ANRS (2012/239 to T.F.B., M.B.Z. and  
14  
15 7 L.B.), ARC, Paris and Institut Hospitalo-Universitaire, Strasbourg (TheraHCC IHUARC  
16  
17 8 IHU201301187 to T.F.B.), the Impulsion Program of the IDEXLYON (to M.B.Z.), Ligue contre  
18  
19 9 le cancer (to M.B.Z.), Inserm, and University of Strasbourg. This work has been published  
20  
21 10 under the framework of the LABEX ANR-10-LABX-0028\_HepSYS, ANR-10-LABX-36  
22  
23 11 NetRNA and Inserm Plan Cancer 2019-2023 and benefits from funding from the state  
24  
25 12 managed by the French National Research Agency as part of the Investments for the future  
26  
27 13 program. S.P. and K.H. were supported by PhD fellowships from the French Ministry of  
28  
29 14 Research and the IdEx program of the University of Strasbourg, respectively.  
30  
31 15

32  
33 16 **Author contribution.** M.B.Z. coordinated and supervised research. K.H., S.B., S.P., C.F.,  
34  
35 17 A.W., L.B., J-C.M. and M.B.Z. designed experiments. K.H., S.B., S.P, C. F., A.W., A.B.,  
36  
37 18 S.C.D., M.M., H.L. and J-C.M. performed experiments. K.H., S.B., S.P., C. F., F.J., A.W.,  
38  
39 19 A.B., B.C.W.M., S.P., J-C.M., W.R., L.B., T.F.B. and M.B.Z. analyzed data. K.H., S.B., and  
40  
41 20 M.B.Z. wrote the paper.  
42  
43 21

44  
45 22 **Competing interests:** The authors do not have competing interest.  
46  
47 23  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 **Abstract**  
4  
5  
6

7 **Objective:** Infection of human hepatocytes by the hepatitis C virus (HCV) is a multistep  
8 process involving both viral and host factors. microRNAs (miRNAs) are small non-coding  
9 RNAs that post-transcriptionally regulate gene expression. Given that miRNAs were  
10 indicated to regulate between 30% and 75% of all human genes, we aimed to investigate the  
11 functional and regulatory role of miRNAs for the HCV life cycle.  
12  
13  
14  
15  
16

17 **Design:** To systematically reveal human miRNAs affecting the HCV life cycle, we performed  
18 a two-step functional high-throughput miRNA mimic screen in Huh7.5.1 cells infected with  
19 recombinant cell culture-derived HCV. miRNA targeting was then assessed using a  
20 combination of computational and functional approaches.  
21  
22  
23  
24  
25

26 **Results:** We uncovered miR-501-3p and miR-619-3p as novel modulators of HCV  
27 assembly/release. We discovered that these miRNAs regulate O-linked N-acetylglucosamine  
28 (O-GlcNAc) transferase (OGT) protein expression and identified OGT and O-GlcNAcylation  
29 as regulators of HCV morphogenesis and infectivity. Furthermore, increased OGT  
30 expression in patient-derived liver tissue was associated with HCV-induced liver disease and  
31 cancer.  
32  
33  
34  
35  
36  
37  
38

39 **Conclusion:** miR-501-3p and miR-619-3p and their target OGT are previously undiscovered  
40 regulatory host factors for HCV assembly and infectivity. In addition to its effect on HCV  
41 morphogenesis, OGT may play a role in HCV-induced liver disease and  
42 hepatocarcinogenesis.  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

## Significance of this study

1

### What is already known about this subject?

- ◆ To establish chronic infection, the hepatitis C virus (HCV) hijacks cellular factors including microRNAs (miRNAs), known to post-transcriptionally regulate gene expression.
- ◆ miRNAs may positively or negatively modulate HCV infection either by directly targeting the viral genome or indirectly by regulating virus-associated cellular pathways[1, 2].

### What are the new findings?

- ◆ A functional miRNA mimic screen uncovered miR-501-3p and miR-619-3p to enhance late steps of HCV infection.
- ◆ miR-501-3p regulates the expression of O-linked N-acetylglucosamine transferase (OGT) at the protein level.
- ◆ Silencing of OGT expression or inhibition of O-linked N-acetylglucosaminylation (O-GlcNAcylation) leads to an increase in the infectivity and size of HCV particles.
- ◆ OGT expression increases in patient-derived liver tissue during liver disease progression and cancer.

### How might it impact on clinical practice in the foreseeable future?

- ◆ As upregulation of OGT and increased O-GlcNAcylation of proteins have been associated with various forms of cancer, OGT may play a dual role in HCV morphogenesis as well as pathogenesis of HCV-induced liver disease and carcinogenesis.

2

3



## 1 Introduction

2 Chronic hepatitis C is a major cause of chronic liver disease and hepatocellular carcinoma  
3 (HCC). Since the approval of pan-genotypic direct-acting antivirals (DAAs), it is considered a  
4 curable disease in more than 90% of treated patients. Nonetheless, an estimated 71 million  
5 individuals are still infected by the hepatitis C virus (HCV) and several challenges remain;  
6 viral cure reduces but does not eliminate the HCC risk in patients with advanced fibrosis[3],  
7 the majority of infected patients has limited access to therapy and DAA failure/viral  
8 resistance has been reported in a subset of patients[4, 5]. To overcome these limitations,  
9 approaches to target host factors involved in HCV infection and pathogenesis are  
10 developed[6, 7]. Interestingly, defined host factors that contribute to the establishment of  
11 chronic HCV infection and represent potential antiviral targets, e.g. epidermal growth factor  
12 receptor[8], also play a role in liver disease pathogenesis and represent candidate targets for  
13 treatment of advanced liver disease and HCC prevention[9]. Thus, uncovering host factors  
14 usurped by HCV not only contributes to a better understanding of virus-host interactions  
15 underlying the HCV life cycle but also to the identification of potential targets for treatment of  
16 liver disease and prevention of HCC.

17 The establishment of various models to study HCV infection has shed light on the  
18 molecular mechanisms that govern the HCV life cycle, which can be subdivided into early  
19 steps, including viral entry, translation and replication as well as late steps, including  
20 assembly and release of new virions. Each step of the HCV replication cycle relies on  
21 specific virus-host interactions that involve host proteins and microRNAs (miRNAs)[7], small  
22 non-coding RNAs that regulate gene expression at the post-transcriptional level. One miRNA  
23 can target numerous messenger RNAs (mRNAs) by base-pairing with a complementary site  
24 that is typically located within the 3' untranslated region (3'UTR) of the mRNA. Accumulating  
25 evidence indicates that miRNAs participate to HCV replication by exerting pro- or antiviral  
26 effects. The breakthrough discovery of the direct targeting of HCV by miR-122, the most  
27 abundant miRNA in the liver, revealed the crucial role of this miRNA for HCV  
28 translation/replication that contributes to progression to chronic HCV infection[1, 10]. miR-

1  
2  
3 1 122 antisense oligonucleotides were subsequently developed as host-targeting antivirals[11,  
4  
5 2 12]. Other miRNAs can indirectly target HCV by regulating host factors that participate in  
6  
7 3 antiviral responses and immune surveillance[2, 13, 14]. Since up to 60% of all human  
8  
9 4 protein-coding genes were reported to be under miRNA-mediated regulation and miRNAs  
10  
11 5 are involved in basically every biological process, we hypothesized that miRNAs provide a  
12  
13 6 tool for loss-of-function approaches to uncover novel HCV host factors. We performed  
14  
15 7 genome-wide high-throughput modulation of the human miRNome and analyzed their impact  
16  
17 8 on HCV infection by combining computational and functional approaches.  
18  
19 9

## 20 21 22 10 **Material and methods**

23  
24 11 **Cells, cell culture conditions, viruses, virus purification, infectivity assays, miRNAs,**  
25  
26 12 **antagomiRs, siRNAs, antibodies, immunoblot, immunocapture, electron microscopy**  
27  
28 13 **analysis of viral particles and gene expression analysis in liver tissue** are described in  
29  
30 14 the Supplementary information.  
31  
32 15

33  
34 16 **Functional miRNA/siRNA screens.** Huh7.5.1 cells were transfected with the miRIDIAN  
35  
36 17 human miRNA mimic library (miRBase 19) comprising more than 2000 mature miRNAs or 28  
37  
38 18 ON-TARGETplus smart pool siRNAs (20 nM, Dharmacon) using Interferin HTS (Polyplus) in  
39  
40 19 a 96-well format[8]. After 48h, a viability test (Presto Blue, Thermo Scientific) was performed  
41  
42 20 prior to a two-step infection assay[15, 16, 17]. During part 1 of the protocol, 50  $\mu$ L of HCV cell  
43  
44 21 culture-derived particles (HCVcc, JcR2a) were incubated with cells during 4h. The inoculum  
45  
46 22 was removed and cells were incubated with 150  $\mu$ l of medium for 48h. In part 2, supernatants  
47  
48 23 from part 1 cells were transferred onto naive Huh7.5.1 cells and part 1 cells were lysed to  
49  
50 24 determine luciferase activity[17, 18]. After 72h, part 2 cells were lysed to determine luciferase  
51  
52 25 activity[17]. siCD81 (20 nM), antagomiR-122 (100 nM) and siApoE (20 nM) were used as  
53  
54 26 positive controls[17]. A non-targeting siRNA with no sequence complementarity to any  
55  
56 27 human gene or homology to any human miRNA was used as negative control.  
57  
58  
59  
60 28

1  
2  
3 1 **Inhibitor treatment.** Four hours following HCV RNA electroporation[8], Huh7.5.1 cells were  
4 incubated with vehicle or inhibitors of OGT (peracetylated 5-thio-N-acetylglucosamine  
5 (Ac<sub>4</sub>5S-GlcNAc)[19]) or OGA (Thiamet G (Sigma))[20]. After 96h, supernatants were  
6 transferred onto naïve Huh7.5.1 cells for 72h prior to determination of luciferase activity while  
7 electroporated cells were lysed to determine luciferase activity.  
8  
9  
10  
11  
12  
13  
14  
15

16 7 **Gene expression analyses.** Total RNA was purified[17] and transcribed into cDNA using  
17 Maxima reverse transcriptase (Thermo Scientific). *GAPDH* and *OGT* mRNA was detected by  
18 real time qPCR using iTaq™ Universal Probes Supermix (Bio-Rad) and TaqMan Gene  
19 Expression Assay (Thermo Scientific). Relative *OGT/GAPDH* gene expression was  
20 calculated by the  $\Delta\Delta C_t$  method[21].  
21  
22  
23  
24  
25  
26  
27  
28

29 13 **Dual luciferase reporter gene assay.** The human *OGT* 3'UTR sequence was retrieved from  
30 NCBI (NM\_181672.2) and Ensembl genome browser (ENST00000373719.3). A fragment of  
31 the *OGT* 3'UTR (positions 3380-3837, NM\_181672.2) (Thermo Fisher Scientific GENEART)  
32 was cloned between the *NotI* and *XhoI* sites downstream of a *Renilla* luciferase cassette in a  
33 psiCHECK2 plasmid (Promega). A mutated version of this construct (9-bp substitution in the  
34 predicted miR-501-3p target site) was generated as described[22]. The functionality of the  
35 *OGT* 3'UTR was assessed as described[23]. The miRIDIAN mimic negative control 1 was  
36 used as control. *Renilla* and *firefly* luciferase activity was assessed 48h after transfection into  
37 HeLa cells using Dual-Luciferase Reporter assay (Promega).  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49

50 23 **Bioinformatic and statistical analysis.** Data analysis and statistical treatment for the  
51 miRNA mimic screen were performed in R (www.r-project.org). Cell measurement data used  
52 in further analysis were cell viability and luciferase activity. In total 26 sets of plates  
53 (performed in triplicate) were tested. The presence of multiple wells with negative and  
54 positive controls on each plate allowed stepwise normalization intra- and inter-plate. First,  
55 intra-plate zonal bias was examined and a model of median effects across the entire screen  
56  
57  
58  
59  
60

1  
2  
3 1 determined using the median-polish algorithm[24] and all plates corrected accordingly. Then  
4  
5 2 the dataset was examined for outlier plates, i.e. plates where all individual measurements  
6  
7 3 correlate very poorly with the other remaining replicates. Three and 9 plates were excluded  
8  
9 4 for part 1 and part 2 of the screen, respectively, based on poor median correlation ( $r < 0.7$ )  
10  
11 5 so that the remaining plates correlation improved substantially ( $> 40\%$ ). Next, the plates were  
12  
13 6 normalized inter replicates using the particularly robust quantile-quantile approach[25].  
14  
15 7 Finally, the data were tested using a moderated t-test (empirical Bayes shrinkage, R-  
16  
17 8 package limma[26]) for the null-hypothesis of no change of a given miRNA compared to the  
18  
19 9 negative control. The resulting  $p$ -values for independent testing of each miRNA were  
20  
21 10 corrected for the multiple testing situation and expressed as local false discovery rate (lfdr,  
22  
23 11 R-package fdrtool[27]). The testing was performed independently for part 1 and 2 of the  
24  
25 12 screen and candidate miRNAs selected for each part. For data from part 1, a lfdr threshold of  
26  
27 13 0.00027 was used. Data from part 2 were subject to increase inherent stochastic noise and  
28  
29 14 for this reason the minimum acceptable relative risk of false positives was increased to  
30  
31 15 0.1226 (i.e. maximum 15% risk for each of the retained hits).

32  
33  
34 16 Other datasets were analyzed using the two-tailed Mann-Whitney test, Wilcoxon test,  
35  
36 17 Spearman correlation or the two-tailed unpaired t-test for data with normal distribution as  
37  
38 18 assessed by D'Agostino and Pearson omnibus and Shapiro-Wilk normality tests (GraphPad  
39  
40 19 Prism v.6 package).

## 41 42 43 44 45 46 47 22 **Results**

48  
49 23 **Genome-wide identification of human miRNAs affecting the HCV life cycle.** We  
50  
51 24 performed a genome-wide screen in human hepatoma Huh7.5.1 cells using a genomic  
52  
53 25 miRNA mimics library and a two-step infection assay[17] with a luciferase reporter virus  
54  
55 26 (JcR2a), which allowed us to functionally assess the role of miRNAs during the early steps  
56  
57 27 (part 1 - viral entry/translation/replication) and the late steps (part 2 - viral  
58  
59 28 assembly/release/infectivity) of the HCV life cycle (Fig. 1A). Silencing of *CD81* and *ApoE*,

1  
2  
3 1 two essential host factors required for HCV entry or assembly, respectively, was performed  
4  
5 2 in parallel using small interfering RNA (siRNA) as controls. Silencing of *CD81* resulted in a  
6  
7 3 reduction of HCV infection in part 1 and consequently in part 2 of the screen since reduced  
8  
9 4 viral entry in the first part of the assay leads to a reduced production of viral particles (Fig.  
10  
11 5 1B)[17]. Silencing of *ApoE* resulted in a marked inhibition of HCV infection only in part 2 of  
12  
13 6 the assay, consistent with the role of ApoE in HCV assembly (Fig. 1B)[17]. The screen  
14  
15 7 identified 427 miRNAs (corresponding to about 16% of the library) that significantly  
16  
17 8 modulated HCV infection ( $\text{Ifdr} < \text{threshold}$ , Supplementary Table 1 and Fig. 1C): 186 miRNAs  
18  
19 9 affected HCV infection in part 1, 309 miRNAs affected HCV infection in part 2, including 68  
20  
21 10 hits in part 1 and part 2. The limited number of part 1 and 2 hits may be due to the fact that a  
22  
23 11 single miRNA may modulate the expression of several proteins, which may have different  
24  
25 12 roles in the viral life cycle. Most hits were observed to dampen HCV infection independently  
26  
27 13 of any significant alteration of cell viability (data not shown). The 186 miRNAs modulating the  
28  
29 14 early steps of HCV infection all decreased viral infection. Among the 309 miRNAs that had  
30  
31 15 an impact in part 2, 11 miRNAs increased HCV infection by at least 3-fold while 298 miRNAs  
32  
33 16 inhibited HCV infection by at least 2.7-fold. Hits from the screen included the let-7 family[2,  
34  
35 17 28], miR-27a[29] and miR-29 family[30] that were already shown to inhibit HCV infection, as  
36  
37 18 well as miR-21[31] and miR-146a-5p[17] that were shown to stimulate HCV infection thus  
38  
39 19 supporting the relevance of our findings. Collectively, our screen identified a set of miRNAs  
40  
41 20 whose overexpression overall impairs HCV infection by affecting viral  
42  
43 21 entry/translation/replication and/or virion assembly/egress/infectivity.

44  
45  
46  
47  
48  
49 23 **miR-619-3p, miR-501-3p and OGT play a role in late steps of the HCV life cycle.** We  
50  
51 24 focused our analysis on miRNAs that modulate late steps of the HCV life cycle, as the  
52  
53 25 molecular mechanisms of HCV assembly/release remain only partially understood. Our  
54  
55 26 screen identified 241 miRNAs that modulated late steps without affecting early steps of  
56  
57 27 infection: 11 miRNAs increased HCV infection while 230 miRNAs decreased HCV infection.  
58  
59 28 Among the miRNAs that increased HCV infection, miR-140-3p, miR-501-3p, miR-619-3p and

1  
2  
3 1 miR-4778-5p have not yet been associated with HCV. Since they enhanced HCV infection in  
4  
5 2 part 2 without affecting part 1, these miRNAs may target host genes that control virus  
6  
7 3 assembly/egress/infectivity. We first confirmed the effect of these miRNAs in independent  
8  
9 4 experiments using the same protocol as for the screen. Overexpression of miR-619-3p or  
10  
11 5 miR-501-3p consistently led to an increase in the infection of progeny virions (Fig. 1D) while  
12  
13 6 infection was decreased with progeny virions from antagomiR-transfected cells  
14  
15 7 (Supplementary Figure S1A). miR-619-3p or miR-501-3p were thus selected for further  
16  
17 8 investigation. To study the molecular mechanisms by which these miRNAs affect HCV  
18  
19 9 infection, we generated a list of predicted miRNA targets using DIANA, TargetScan Human  
20  
21 10 v6.2 and miRDB databases, and selected candidate targets based on their expression in our  
22  
23 11 Huh7.5.1 cells as assessed by microarray (data not shown). Ingenuity Pathway Analysis  
24  
25 12 enabled us to refine the gene list by selecting 28 genes involved in the following functional  
26  
27 13 networks or pathways that contribute to the HCV life cycle[32, 33, 34]: lipid metabolism and  
28  
29 14 cholesterol biosynthesis, protein maturation and processing at the endoplasmic reticulum  
30  
31 15 (ER), components of the endosomal sorting complex, adipocyte biogenesis, cellular  
32  
33 16 morphology and cell inflammation (Table 1).

34  
35  
36  
37 17 To assess whether knock-down of these 28 candidate targets affects virus  
38  
39 18 production, we performed a siRNA-based screen using siRNA pools exhibiting strong  
40  
41 19 silencing without cytotoxicity (Fig. 2). Silencing of *CD81* and antagomiR-122 served as  
42  
43 20 controls for part 1; knock-down of *ApoE* served as control for part 2 (Fig. 2). Hits were  
44  
45 21 defined as genes whose knock-down modulated HCV infection in at least one part of the  
46  
47 22 screen with high significance (Fig. 2,  $p$ -value  $\leq 0.0001$ , Mann-Whitney U-test). HCV  
48  
49 23 entry/translation/replication was significantly modulated by silencing of *PPP3CA*, *CEBPA*,  
50  
51 24 *MID1*, *WDFY3*, *DCX* and *SLC35D1*. HCV assembly/egress/infectivity was significantly  
52  
53 25 modulated by knock-down of *PPP3CA*, *CSDE1*, *GAN*, *USP37*, *CEBPA*, *MID1*, *WDFY3*, *DCX*,  
54  
55 26 *MAPK9*, *SLC35D1*, *DCC*, *RNF144A*, *PPP2R2C* and *OGT*. Strikingly, only the silencing of  
56  
57 27 *OGT* was associated with an enhancement of HCV assembly/release/infectivity ( $p$ -value =  
58  
59 28 0.0002), while that of the other hits was associated with reduced HCV infection (Fig. 2).

1  
2  
3 1 These results indicate that the down-regulation of *OGT* phenocopies the effect of miR-501-  
4 3p and miR-619-3p on HCV infection (Fig. 2) and suggest *OGT* as a novel player in the HCV  
5 2  
6 3 life cycle.  
7  
8  
9 4

10  
11 5 **miR-501-3p post-transcriptionally regulates *OGT* expression.** To study whether miR-501-  
12 6 3p and miR-619-3p target *OGT*, we analyzed *OGT* RNA and protein levels in Huh7.5.1 cells  
13 7 following overexpression of miR-501-3p or miR-619-3p. While neither miRNA had an impact  
14 8 on *OGT* RNA levels (Fig. 3A), up-regulation of miR-501-3p significantly decreased *OGT*  
15 9 protein expression by ~65% (Fig. 3B,  $p$ -value  $\leq 0.05$ , t-test). miR-619-3p also decreased  
16 10 *OGT* expression but less robustly than miR-501-3p (Fig. 3B), prompting us to focus our  
17 11 investigation on miR-501-3p. To assess whether *OGT* is a functional target of miR-501-3p,  
18 12 we subcloned a fragment of the *OGT* mRNA 3'UTR that harbors the predicted miR-501-3p  
19 13 target site in the *Renilla* luciferase expression cassette (RLuc) of a dual luciferase reporter  
20 14 construct. Co-transfection of miR-501-3p mimic with the wild-type 3'UTR reporter (RLuc wt  
21 15 *OGT* 3'UTR) significantly decreased luciferase activity as compared to the empty vector (Fig.  
22 16 3C,  $p$ -value  $< 0.05$ , t-test). In contrast, the repression of luciferase expression was lost when  
23 17 the reporter with mutated miR-501-3p binding site (RLuc mt *OGT* 3'UTR) was used (Fig. 3C).  
24 18 These data are consistent in indicating that miR-501-3p mediates post-transcriptional  
25 19 regulation of *OGT*.  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44

45 21 **O-GlcNAcylation modulates HCVcc infectivity.** To investigate whether *OGT* modulates  
46 22 HCV assembly and/or infectivity, we determined infectious virus titer (TCID<sub>50</sub>) and HCV RNA  
47 23 levels to calculate the specific infectivity of HCVcc particles generated in *OGT*-silenced  
48 24 Huh7.5.1 cells. Interestingly, *OGT*-silencing led to a significant increase in the TCID<sub>50</sub> and  
49 25 the specific infectivity of HCVcc (Fig. 4A,  $p$ -value  $\leq 0.05$ , Mann-Whitney test). Noteworthy,  
50 26 the effect of *OGT* on HCVcc infectivity was genotype-independent as demonstrated by  
51 27 increased infectivity of HCVcc bearing the envelope glycoproteins of genotypes 1a, 1b and  
52 28 2a upon *OGT*-silencing (Fig. 4B). We next sought to investigate how *OGT* could modulate

1 HCVcc infectivity. OGT is the only enzyme that catalyzes the addition of N-  
2 acetylglucosamine (O-GlcNAc) to serine and threonine residues of proteins. Moreover, OGT  
3 has a scaffold function and promotes binding of proteins in multiprotein complexes[35]. To  
4 assess whether the enzymatic activity of OGT modulates HCVcc infectivity, we used  
5 pharmacological inhibitors of OGT (Ac<sub>4</sub>5S-GlcNAc) or O-GlcNAcase (OGA) (Thiamet G), the  
6 OGT counterpart that removes O-GlcNAc (Fig. 4C). Ac<sub>4</sub>5S-GlcNAc led to a significant  
7 enhancement of HCVcc infectivity in a dose-dependent manner, while the opposite effect  
8 was observed with Thiamet G (Fig. 4D,  $p$ -value  $\leq$  0.05, Mann-Whitney test). Collectively,  
9 these results demonstrate that O-GlcNAcylation modulates HCVcc infectivity.

11 **OGT-silencing affects HCVcc biophysical properties and size distribution.** To further  
12 assess how OGT may impact HCVcc morphogenesis, we analyzed the structural and  
13 biophysical properties of HCVcc produced in siCtrl- and siOGT-transfected Huh7.5.1 cells  
14 following iodixanol gradient ultracentrifugation. Silencing of OGT led to the production of  
15 more infectious HCVcc with higher density (Fig. 5A-B) as well as higher ApoE concentrations  
16 (Fig. 5C) suggesting that OGT/O-GlcNAcylation affects the biophysical properties of HCVcc.  
17 No change in apoB concentrations were observed between HCVcc produced from siCtrl- or  
18 siOGT-transfected cells (Fig. 5D), in line with the model that HCV lipovirions contain  
19 several exchangeable ApoE molecules and one non-exchangeable apoB[36]. We also  
20 visualized HCVcc by electron microscopy (EM) following anti-E2 antibody  
21 immunocapture[36] to assess whether OGT-silencing had an impact on HCVcc size. Particle  
22 size distribution was assessed from a series of randomly acquired electron micrographs. A  
23 shift towards bigger sizes was observed for sucrose-cushion purified HCVcc generated in  
24 OGT-silenced Huh7.5.1 cells as compared to control HCVcc (Fig. 6A-B). This shift was also  
25 observed in different fractions of iodixanol gradient-separated HCVcc (Fig. 6C-F) in line with  
26 the higher infectivity and ApoE concentrations of HCVcc generated in OGT-silenced  
27 Huh7.5.1 cells (Fig. 5A-C). These data suggest that OGT-silencing affects the lipidation of  
28 HCVcc.



1 **OGT expression increases in liver disease.** Since silencing of OGT promotes HCV  
2 infectivity, we assessed whether HCV infection in turn had an effect on miR-501-3p and OGT  
3 expression. In Huh7.5.1 cells, HCV infection lead to small but significant increase of miR-  
4 501-3p and decrease of OGT levels (Fig. 7A-B and Supplementary Fig. 1B;  $p$ -value < 0.05,  
5 Mann-Whitney test), which may promote viral infection given the pro- and antiviral roles of  
6 miR-501-3p and O-GlcNAcylation, respectively (Fig. 1C-D and 4D). In contrast, no significant  
7 difference of OGT expression was observed between the livers of HCV transgenic and wild-  
8 type mice[37] (data not shown) suggesting that HCV proteins do not directly modulate OGT  
9 expression. In liver tissue from HCV-infected patients, HCV RNA levels were not correlated  
10 with OGT expression (Fig. 7C, Spearman correlation: 0.06004019,  $p$ -value = 0.7661)  
11 suggesting that in patients there is likely no direct effect of HCV on OGT expression.

12 O-GlcNAcylation has been associated with a variety of cancers, including HCC  
13 recurrence linked to increased O-GlcNAcylation after liver transplantation[38]. We therefore  
14 investigated OGT expression in chronic liver disease and HCC. While there was a trend for  
15 increased OGT expression in liver tissue from HCV-infected patients with fibrosis and  
16 inflammation (Fig. 7D-E), OGT levels were markedly and significantly elevated in the tumor  
17 liver tissue of patients chronically infected with HCV or hepatitis B virus and patients with  
18 alcoholic liver disease or non-alcoholic fatty liver disease as compared to non-tumor tissue  
19 (Fig. 7F,  $p$ -value < 0.05, Wilcoxon test). These data suggest that OGT expression increases  
20 in HCC in an etiology-independent manner. Collectively, these results suggest that OGT  
21 expression is likely increased in HCV-induced liver disease and cancer through inflammation  
22 and fibrosis rather than by HCV itself.

## 25 Discussion

26 By focusing on miRNAs affecting late steps of the viral life cycle, we uncovered that i) miR-  
27 501-3p regulates the expression of OGT; ii) silencing of OGT expression or inhibition of its  
28 enzymatic activity increases the infectivity of HCV particles; and iii) OGT knock-down leads

1  
2  
3 1 to the release of bigger HCV particles. Our data suggest that O-GlcNAcylation affects HCV  
4 morphogenesis and infectivity.

5  
6  
7 3 While we were characterizing the role of OGT/O-GlcNAcylation for HCV  
8 morphogenesis, Li and colleagues published their functional genomics study of HCV-miRNA  
9 interactions[2]. By conducting genome wide miRNA mimic and hairpin inhibitor screens, they  
10 identified a set of miRNAs exhibiting a pro- or antiviral effect on HCV. Characterization of the  
11 underlying molecular processes showed that miR-25, let-7 and miR-130 families restrict viral  
12 infection by decreasing the expression of cellular HCV co-factors[2]. Despite similarities in  
13 the cell type and HCV infection models used here and by Li and colleagues, our screen only  
14 displays a small overlap with their study (9% common miRNA hits). This is not surprising  
15 given the small overlap between previous siRNA screens to uncover HCV host factors[8, 15]  
16 and is likely due i) to the different sizes of miRNA mimic libraries as the library used here was  
17 more than 2-times larger than the one used by Li and co-workers, and ii) to the markedly  
18 distinct pipelines for hit selection that were used in the two studies. Nonetheless, both  
19 screens were consistent in confirming the proviral role of miR-146a-5p in promoting HCV  
20 assembly/egress that we previously reported[17] and the global multistep inhibitory effects of  
21 the let-7 family on HCV infection[28], further corroborating the involvement of these miRNAs  
22 in fine-tuning the HCV life cycle. Both studies also consistently indicated that miR-518a-5p,  
23 miR-517-3p, miR-185 and members of the miR-302 family inhibit early steps of HCV  
24 infection, while miR-586, miR-620 and members of the miR-200 family inhibit late steps of  
25 viral infection. Since none of these miRNAs except miR-185 has been previously associated  
26 with HCV infection[39], it might be interesting to further characterize the involvement of these  
27 miRNAs in HCV-host interactions. Interestingly, an overall proviral effect of miR-501-3p was  
28 also observed by Li and colleagues[2], however the mechanism of action was not studied. By  
29 characterizing the role of miR-501-3p in the HCV life cycle, we uncovered OGT as a miR-  
30 501-3p target in liver-derived cells and showed for the first time a link between O-  
31 GlcNAcylation and HCV infection. These results indicate that genome-wide miRNA functional

1  
2  
3 1 screens represent a powerful strategy to dissect the role of miRNAs in pathogen-host  
4  
5 2 interactions.

6  
7 3 While N-glycosylation of HCV envelope glycoproteins plays an important role for  
8  
9 4 escape from virus-neutralizing antibodies[40], so far no functional association between HCV  
10  
11 5 and O-glycosylation has been reported. In contrast to N-linked glycosylation that consists of  
12  
13 6 the attachment of a glycan to a nitrogen of an asparagine residue of proteins in the ER/Golgi  
14  
15 7 prior to their trafficking to the plasma membrane and/or their secretion, the glycosylation of  
16  
17 8 serine and threonine residues with O-GlcNAc is a post-translational modification (PTM) of  
18  
19 9 intracellular proteins that are localized in the nucleus, cytoplasm or mitochondria. The O-  
20  
21 10 glycosylation/deglycosylation of proteins is catalyzed by a single pair of nucleo-cytoplasmic  
22  
23 11 enzymes, OGT/OGA. O-GlcNAcylation is complementary to protein  
24  
25 12 phosphorylation/dephosphorylation, another more broadly known abundant protein PTM that  
26  
27 13 involves numerous kinases/phosphatases. OGT/OGA are often found in protein complexes  
28  
29 14 that also include kinases/phosphatases and a protein can be either O-GlcNAcylated or  
30  
31 15 phosphorylated on a same residue to fine-tune cellular signaling[41]. O-GlcNAcylation and  
32  
33 16 phosphorylation on the same or neighboring serine or threonine residue is known as yin yang  
34  
35 17 site[42].

36  
37  
38 18 O-GlcNAcylation plays a major role in the regulation of metabolic pathways in the  
39  
40 19 liver, including insulin signaling, bile acid metabolism and lipogenesis[35]. The large number  
41  
42 20 of OGT/OGA substrates and cellular pathways regulated by O-GlcNAcylation hampers a  
43  
44 21 detailed characterization of the role of these proteins in HCV infection. Since i) HCV  
45  
46 22 assembly takes place at ER-derived membranes, ii) OGT/OGA are not known to localize in  
47  
48 23 the ER lumen, and iii) O-GlcNAcylation of extracellular proteins containing EGF-like domains  
49  
50 24 is catalyzed by EGF domain-specific OGT (EOGT) in the ER lumen in an OGT-independent  
51  
52 25 manner[43]), OGT/OGA most likely modulate HCV infection by post-translationally modifying  
53  
54 26 one or several cellular factors required for HCV morphogenesis rather than by affecting viral  
55  
56 27 proteins, although HCV glycoproteins contain putative O-GlcNAcylation sites as determined  
57  
58 28 using OGlcNAcScan, OGTsite and YingOYang1.2 bioinformatics tools (data not shown).

1  
2  
3 1 Regarding HCV host factors that may be regulated by OGT/OGA, O-GlcNAcylation  
4  
5 2 sites have been predicted in human CLDN1[44] and OCLN at serine sites that can also be  
6  
7 3 phosphorylated and this has been suggested to potentially play a role for HCV entry[45].  
8  
9 4 However, in our experimental setting we did not observe a significant effect of OGT-silencing  
10  
11 5 on the early steps of HCV infection, suggesting that O-GlcNAcylation of CLDN1 and/or  
12  
13 6 OCLN likely does not play a major role in HCV infection. Other host factors important for the  
14  
15 7 HCV life cycle are well-known O-GlcNAcylated proteins, as for example various nuclear pore  
16  
17 8 complex proteins (Nups) including Nup98, Nup153 and Nup155 that are involved in HCV  
18  
19 9 replication and assembly and/or may be associated with viral particles[46, 47, 48]. However,  
20  
21 10 since depletion of Nups was reported to alter HCV replication and/or assembly but to have  
22  
23 11 no impact on the specific infectivity of HCV particles[46] in contrast to the depletion of OGT  
24  
25 12 as shown here, it is unlikely that a modulation of Nup O-GlcNAcylation accounts for the  
26  
27 13 effects of OGT-silencing and/or OGT/OGA inhibitors on HCVcc infectivity observed in our  
28  
29 14 study. This is in line with our observation that OGT knock-down had no effect on Dengue  
30  
31 15 virus (DENV) replication and infectivity (unpublished observations KH, MZ and Evelyne  
32  
33 16 Schaffer, IBMC, Strasbourg), although Nup98 had been suggested to potentially play a role  
34  
35 17 for DENV infection[46]. These data suggest that OGT does not broadly modulate the  
36  
37 18 infectivity of viruses of the *Flaviviridae* family.

38  
39  
40  
41 19 However, OGT and/or O-GlcNAcylation have been reported to play a role in the  
42  
43 20 infection with other viruses[49, 50, 51]. Interestingly, while OGT expression modulates the  
44  
45 21 levels of human papillomavirus 16 (HPV16) oncoproteins E6 and E7[52], E6 in turn can up-  
46  
47 22 regulate OGT to increase O-GlcNAcylation and the oncogene activities of HPV[53],  
48  
49 23 suggesting that OGT/O-GlcNAcylation could play a role in virus-induced cancer. In cell  
50  
51 24 culture, HCV infection appeared to be associated with a minor decrease in OGT expression  
52  
53 25 in line with an antiviral role of O-GlcNAcylation. In contrast, an increased OGT expression  
54  
55 26 was observed in HCC tissues of HCV-infected patients. Since OGT has been suggested to  
56  
57 27 activate oncogenic signaling pathways in non-alcoholic steatohepatitis-related HCC[54] and  
58  
59 28 O-GlcNAcylation has been associated with HCC recurrence linked to increased O-

1 GlcNAcylation after liver transplantation[38], these data suggest that in addition to their effect  
2 on the HCV life cycle, OGT/O-GlcNAcylation may also play a role in HCV-induced  
3 hepatocarcinogenesis.  
4  
5  
6  
7  
8  
9  
10  
11  
12

## 13 **Acknowledgments**

14 We wish to thank Gerald W. Hart and Stéphan Hardivillé, the CardioPEG CoreC4 (NHLBI  
15 P01 HL107153) for providing AL24 and for useful technical discussions. We are grateful to  
16 David Vocadlo (Simon Fraser University, Burnaby, Canada) for the gift of Ac<sub>4</sub>5S-GlcNAc. We  
17 also thank Ralf Bartenschlager (University of Heidelberg, Germany) for providing the  
18 plasmids for production of HCVcc and Frank Chisari (The Scripps Research Institute, La  
19 Jolla, CA) for the gift of Huh7.5.1 cells. We acknowledge Evelyne Schaeffer (CNRS  
20 UPR3572, IBMC, Strasbourg) for the DENV experiment, Charlotte Bach and Christine  
21 Thumann (Inserm, U1110, Strasbourg) for excellent technical work during the functional  
22 miRNA mimic screen, as well as Armando A. Roca-Suarez (Inserm, U1110, Strasbourg),  
23 Hussein El Saghire (Inserm, U1110, Strasbourg), Arnaud Kopp (IGBMC, Department of  
24 Functional Genomics and Cancer) and Erika Girardi (UPR 9002, IBMC, Strasbourg) for  
25 helpful discussions. We thank the INGESTEM infrastructure for access to the IGBMC high-  
26 throughput screening workstation.  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48

## 49 **References**

- 50 1 Jopling CL, Yi M, Lancaster AM, *et al.* Modulation of hepatitis C virus RNA  
51 abundance by a liver-specific MicroRNA. *Science* 2005;**309**:1577-81.
- 52 2 Li Q, Lowey B, Sodroski C, *et al.* Cellular microRNA networks regulate host  
53 dependency of hepatitis C virus infection. *Nat Commun* 2017;**8**:1789.
- 54 3 Baumert TF, Juhling F, Ono A, *et al.* Hepatitis C-related hepatocellular carcinoma in  
55 the era of new generation antivirals. *BMC Med* 2017;**15**:52.

- 1  
2  
3 1 4 Pawlotsky JM. Hepatitis C Virus Resistance to Direct-Acting Antiviral Drugs in  
4 2 Interferon-Free Regimens. *Gastroenterology* 2016;**151**:70-86.  
5  
6  
7 3 5 Dietz J, Susser S, Vermehren J, *et al.* Patterns of Resistance-Associated  
8 4 Substitutions in Patients With Chronic HCV Infection Following Treatment With Direct-Acting  
9 5 Antivirals. *Gastroenterology* 2018;**154**:976-88 e4.  
10  
11  
12 6 6 Zeisel MB, Baumert TF. Clinical development of hepatitis C virus host-targeting  
13 7 agents. *Lancet* 2017;**389**:674-5.  
14  
15  
16 8 7 Zeisel MB, Crouchet E, Baumert TF, *et al.* Host-Targeting Agents to Prevent and  
17 9 Cure Hepatitis C Virus Infection. *Viruses* 2015;**7**:5659-85.  
18  
19  
20 10 8 Lupberger J, Zeisel MB, Xiao F, *et al.* EGFR and EphA2 are host factors for hepatitis  
21 11 C virus entry and possible targets for antiviral therapy. *Nature Medicine* 2011;**17**:589-95.  
22  
23  
24 12 9 Fuchs BC, Hoshida Y, Fujii T, *et al.* Epidermal growth factor receptor inhibition  
25 13 attenuates liver fibrosis and development of hepatocellular carcinoma. *Hepatology*  
26 14 2014;**59**:1577-90.  
27  
28  
29 15 10 Masaki T, Arend KC, Li Y, *et al.* miR-122 stimulates hepatitis C virus RNA synthesis  
30 16 by altering the balance of viral RNAs engaged in replication versus translation. *Cell Host*  
31 17 *Microbe* 2015;**17**:217-28.  
32  
33  
34 18 11 Janssen HL, Reesink HW, Lawitz EJ, *et al.* Treatment of HCV infection by targeting  
35 19 microRNA. *N Engl J Med* 2013;**368**:1685-94.  
36  
37  
38 20 12 van der Ree MH, de Vree JM, Stelma F, *et al.* Safety, tolerability, and antiviral effect  
39 21 of RG-101 in patients with chronic hepatitis C: a phase 1B, double-blind, randomised  
40 22 controlled trial. *Lancet* 2017;**389**:709-17.  
41  
42  
43 23 13 Bandiera S, Pfeffer S, Baumert TF, *et al.* miR-122--a key factor and therapeutic target  
44 24 in liver disease. *J Hepatol* 2015;**62**:448-57.  
45  
46  
47 25 14 Li H, Jiang JD, Peng ZG. MicroRNA-mediated interactions between host and hepatitis  
48 26 C virus. *World J Gastroenterol* 2016;**22**:1487-96.  
49  
50  
51 27 15 Li Q, Brass AL, Ng A, *et al.* A genome-wide genetic screen for host factors required  
52 28 for hepatitis C virus propagation. *Proc Natl Acad Sci U S A* 2009;**106**:16410-5.

- 1  
2  
3 1 16 Poenisch M, Metz P, Blankenburg H, *et al.* Identification of HNRNPK as regulator of  
4 hepatitis C virus particle production. *PLoS Pathog* 2015;**11**:e1004573.  
5  
6 2  
7 3 17 Bandiera S, Pernot S, El Saghire H, *et al.* Hepatitis C Virus-Induced Upregulation of  
8 MicroRNA miR-146a-5p in Hepatocytes Promotes Viral Infection and Deregulates Metabolic  
9 Pathways Associated with Liver Disease Pathogenesis. *J Virol* 2016;**90**:6387-400.  
10  
11 4  
12 5  
13 6 18 Da Costa D, Turek M, Felmlee DJ, *et al.* Reconstitution of the entire hepatitis C virus  
14 life cycle in non-hepatic cells. *J Virol* 2012;**86**:11919-25.  
15  
16 7  
17 8 19 Gloster TM, Zandberg WF, Heinonen JE, *et al.* Hijacking a biosynthetic pathway  
18 yields a glycosyltransferase inhibitor within cells. *Nat Chem Biol* 2011;**7**:174-81.  
19  
20 9  
21 20 Yuzwa SA, Macauley MS, Heinonen JE, *et al.* A potent mechanism-inspired O-  
22 GlcNAcase inhibitor that blocks phosphorylation of tau in vivo. *Nat Chem Biol* 2008;**4**:483-90.  
23  
24 11  
25 12 21 Schmittgen TD, Livak KJ. Analyzing real-time PCR data by the comparative C(T)  
26 method. *Nat Protoc* 2008;**3**:1101-8.  
27  
28 13  
29 14 22 Jin Y, Chen Z, Liu X, *et al.* Evaluating the microRNA targeting sites by luciferase  
30 reporter gene assay. *Methods Mol Biol* 2013;**936**:117-27.  
31  
32 15  
33 16 23 Van Renne N, Roca Suarez AA, Duong FHT, *et al.* miR-135a-5p-mediated  
34 downregulation of protein tyrosine phosphatase receptor delta is a candidate driver of HCV-  
35 associated hepatocarcinogenesis. *Gut* 2018;**67**:953-62.  
36  
37 17  
38 18 24 Mosteller F, Tukey J. *Data Analysis and Regression*. Reading, MA: Addison-Wesley,  
39 1977.  
40  
41 19  
42 20 25 Amaratunga D, Cabrera J. Analysis of Data from Viral DNA Microchips. *Journal of the*  
43 *American Statistical Association* 2001;**96**:1161.  
44  
45 21  
46 22 26 Smyth GK. Linear models and empirical Bayes methods for assessing differential  
47 expression in microarray experiments. *Statistical Applications in Genetics and Molecular*  
48 *Biology* 2004;**3**:Article 3.  
49  
50 23  
51 24 27 Strimmer K. fdrtool: a versatile R package for estimating local and tail area-based  
52 false discovery rates. *Bioinformatics* 2008;**24**:1461-2.  
53  
54 25  
55 26  
56 27  
57  
58  
59  
60

- 1  
2  
3 1 28 Cheng M, Si Y, Niu Y, *et al.* High-throughput profiling of alpha interferon- and  
4  
5 2 interleukin-28B-regulated microRNAs and identification of let-7s with anti-hepatitis C virus  
6  
7 3 activity by targeting IGF2BP1. *J Virol* 2013;**87**:9707-18.  
8  
9 4 29 Shirasaki T, Honda M, Shimakami T, *et al.* MicroRNA-27a regulates lipid metabolism  
10  
11 5 and inhibits hepatitis C virus replication in human hepatoma cells. *J Virol* 2013;**87**:5270-86.  
12  
13 6 30 Bandyopadhyay S, Friedman RC, Marquez RT, *et al.* Hepatitis C virus infection and  
14  
15 7 hepatic stellate cell activation downregulate miR-29: miR-29 overexpression reduces  
16  
17 8 hepatitis C viral abundance in culture. *J Infect Dis* 2011;**203**:1753-62.  
18  
19 9 31 Chen Y, Chen J, Wang H, *et al.* HCV-induced miR-21 contributes to evasion of host  
20  
21 10 immune system by targeting MyD88 and IRAK1. *PLoS Pathog* 2013;**9**:e1003248.  
22  
23 11 32 Ariumi Y, Kuroki M, Maki M, *et al.* The ESCRT system is required for hepatitis C virus  
24  
25 12 production. *PLoS One* 2011;**6**:e14517.  
26  
27 13 33 Paul D, Madan V, Bartenschlager R. Hepatitis C virus RNA replication and assembly:  
28  
29 14 living on the fat of the land. *Cell Host Microbe* 2014;**16**:569-79.  
30  
31 15 34 Meyers NL, Fontaine KA, Kumar GR, *et al.* Entangled in a membranous web: ER and  
32  
33 16 lipid droplet reorganization during hepatitis C virus infection. *Curr Opin Cell Biol* 2016;**41**:117-  
34  
35 17 24.  
36  
37 18 35 Yang X, Qian K. Protein O-GlcNAcylation: emerging mechanisms and functions. *Nat*  
38  
39 19 *Rev Mol Cell Biol* 2017;**18**:452-65.  
40  
41 20 36 Piver E, Boyer A, Gaillard J, *et al.* Ultrastructural organisation of HCV from the  
42  
43 21 bloodstream of infected patients revealed by electron microscopy after specific  
44  
45 22 immunocapture. *Gut* 2017;**66**:1487-95.  
46  
47 23 37 Lerat H, Honda M, Beard MR, *et al.* Steatosis and liver cancer in transgenic mice  
48  
49 expressing the structural and nonstructural proteins of hepatitis C virus. *Gastroenterology*  
50  
51 2002;**122**:352-65.  
52  
53 25  
54  
55 26 38 de Queiroz RM, Carvalho E, Dias WB. O-GlcNAcylation: The Sweet Side of the  
56  
57 27 Cancer. *Front Oncol* 2014;**4**:132.  
58  
59  
60



- 1  
2  
3 1 39 Singaravelu R, O'Hara S, Jones DM, *et al.* MicroRNAs regulate the immunometabolic  
4  
5 2 response to viral infection in the liver. *Nat Chem Biol* 2015;**11**:988-93.  
6  
7 3 40 Lavie M, Hanouille X, Dubuisson J. Glycan Shielding and Modulation of Hepatitis C  
8  
9 4 Virus Neutralizing Antibodies. *Front Immunol* 2018;**9**:910.  
10  
11 5 41 Hart GW, Slawson C, Ramirez-Correa G, *et al.* Cross talk between O-GlcNAcylation  
12  
13 6 and phosphorylation: roles in signaling, transcription, and chronic disease. *Annu Rev*  
14  
15 7 *Biochem* 2011;**80**:825-58.  
16  
17 8 42 Hart GW, Greis KD, Dong LY, *et al.* O-linked N-acetylglucosamine: the "yin-yang" of  
18  
19 9 Ser/Thr phosphorylation? Nuclear and cytoplasmic glycosylation. *Adv Exp Med Biol*  
20  
21 10 1995;**376**:115-23.  
22  
23 11 43 Sakaidani Y, Nomura T, Matsuura A, *et al.* O-linked-N-acetylglucosamine on  
24  
25 12 extracellular protein domains mediates epithelial cell-matrix interactions. *Nat Commun*  
26  
27 13 2011;**2**:583.  
28  
29 14 44 Butt AM, Khan IB, Hussain M, *et al.* Role of post translational modifications and novel  
30  
31 15 crosstalk between phosphorylation and O-beta-GlcNAc modifications in human claudin-1, -3  
32  
33 16 and -4. *Mol Biol Rep* 2012;**39**:1359-69.  
34  
35 17 45 Butt AM, Feng D, Nasrullah I, *et al.* Computational identification of interplay between  
36  
37 18 phosphorylation and O-beta-glycosylation of human occludin as potential mechanism to  
38  
39 19 impair hepatitis C virus entry. *Infect Genet Evol* 2012;**12**:1235-45.  
40  
41 20 46 Neufeldt CJ, Joyce MA, Levin A, *et al.* Hepatitis C virus-induced cytoplasmic  
42  
43 21 organelles use the nuclear transport machinery to establish an environment conducive to  
44  
45 22 virus replication. *PLoS Pathog* 2013;**9**:e1003744.  
46  
47 23 47 Lussignol M, Kopp M, Molloy K, *et al.* Proteomics of HCV virions reveals an essential  
48  
49 24 role for the nucleoporin Nup98 in virus morphogenesis. *Proc Natl Acad Sci U S A*  
50  
51 25 2016;**113**:2484-9.  
52  
53 26 48 Zhu Y, Liu TW, Madden Z, *et al.* Post-translational O-GlcNAcylation is essential for  
54  
55 27 nuclear pore integrity and maintenance of the pore selectivity filter. *J Mol Cell Biol* 2016;**8**:2-  
56  
57 28 16.  
58  
59  
60

- 1  
2  
3 1 49 Jochmann R, Thureau M, Jung S, *et al.* O-linked N-acetylglucosaminylation of Sp1  
4  
5 2 inhibits the human immunodeficiency virus type 1 promoter. *J Virol* 2009;**83**:3704-18.  
6  
7 3 50 Groussaud D, Khair M, Tollenaere AI, *et al.* Hijacking of the O-GlcNAcZYME complex  
8  
9 4 by the HTLV-1 Tax oncoprotein facilitates viral transcription. *PLoS Pathog*  
10  
11 5 2017;**13**:e1006518.  
12  
13 6 51 Angelova M, Ortiz-Meoz RF, Walker S, *et al.* Inhibition of O-Linked N-  
14  
15 7 Acetylglucosamine Transferase Reduces Replication of Herpes Simplex Virus and Human  
16  
17 8 Cytomegalovirus. *J Virol* 2015;**89**:8474-83.  
18  
19 9 52 Kim M, Kim YS, Kim H, *et al.* O-linked N-acetylglucosamine transferase promotes  
20  
21 10 cervical cancer tumorigenesis through human papillomaviruses E6 and E7 oncogenes.  
22  
23 11 *Oncotarget* 2016;**7**:44596-607.  
24  
25 12 53 Zeng Q, Zhao RX, Chen J, *et al.* O-linked GlcNAcylation elevated by HPV E6  
26  
27 13 mediates viral oncogenesis. *Proc Natl Acad Sci U S A* 2016;**113**:9333-8.  
28  
29 14 54 Xu W, Zhang X, Wu JL, *et al.* O-GlcNAc transferase promotes fatty liver-associated  
30  
31 15 liver cancer through inducing palmitic acid and activating endoplasmic reticulum stress. *J*  
32  
33 16 *Hepatol* 2017;**67**:310-20.  
34  
35 17 55 Boldanova T, Suslov A, Heim MH, *et al.* Transcriptional response to hepatitis C virus  
36  
37 18 infection and interferon-alpha treatment in the human liver. *EMBO Mol Med* 2017;**9**:816-34.  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

## 1 **Figure legends**

2 **Figure 1. High-throughput screen identifies human miRNAs that regulate the HCV life**  
3 **cycle.** (A) Schematic outline of the miRNA mimic screen strategy. Huh7.5.1 cells were  
4 transfected with miRNA mimics or controls prior to infection with *Renilla* luciferase HCVcc  
5 (JcR2a) two days later (part 1). Cell supernatants of part 1 were used to inoculate naïve  
6 Huh7.5.1 cells (part 2). Cells from part 1 and part 2 were lysed at the end of each infection  
7 step (2 and 3 days post infection, respectively) to determine luciferase activity. (B)  
8 Modulation of HCV entry and replication (part 1) and/or assembly and infectivity (part 2) upon  
9 transfection of control non-targeting siRNA (siCtrl, negative control), siCD81 (inhibiting viral  
10 entry) or siApoE (inhibiting viral assembly). By inhibiting HCV entry, siCD81 impacts part 1  
11 as well as part 2. In contrast, by specifically impairing late steps of HCV replication cycle,  
12 siApoE inhibits HCV infection only in part 2. The box plots show the sample lower quartile  
13 (25th percentile; bottom of the box), the median (50th percentile; horizontal line in box) and  
14 the upper quartile (75th percentile; top of the box) of relative light units (RLU) in each lysate.  
15 The whiskers indicate s.d. Data are from three independent experiments. (C) Effects of  
16 miRNA overexpression on each part of the HCV life cycle. Data were tested using a  
17 moderated t-test (empirical Bayes shrinkage, R-package limma[26]) for the null-hypothesis of  
18 no change of a given miRNA compared to the negative control. The resulting p-values for  
19 independent testing of each miRNA were corrected for the multiple testing situation and  
20 expressed as local false discovery rate (lfdr, R-package fdrtool[27]). miRNAs having a  
21 significant effect on either part 1 or 2 of the screen are below the thresholds indicated by  
22 dashed lines ( $lfdr < 0.00027$  or  $0.1226$ , respectively). miRNAs that were previously reported  
23 to impact on HCV infection as well as miR-140-3p, miR-501-3p, miR-619-3p and miR-4778-  
24 5p are highlighted in blue ( $Log_2(FC) < 0$ ) or red ( $Log_2(FC) > 0$ ). Data are from three  
25 independent experiments. (D) Effect of miR-140-3p, miR-501-3p, miR-619-3p and miR-4778-  
26 5p on the HCV life cycle. Huh7.5.1 cells were transfected with siCtrl (Ctrl), miR-140-3p, miR-  
27 501-3p, miR-619-3p or miR-4778-5p and infection experiments were carried out as described  
28 in A. HCV infection was determined as luciferase activity. Results represent mean

1  
2  
3 1 percentage  $\pm$  s.d. from three independent experiments in triplicate. The dashed line indicates  
4  
5 2 values from control-transfected cells set at 100%. Statistics: \*,  $p$ -value  $\leq$  0.05, Mann-Whitney  
6  
7 3 test.  
8  
9 4

10  
11 **Figure 2. OGT is a novel host cell factor involved in the late steps of the HCV life cycle.**

12  
13 Huh7.5.1 cells were transfected with a set of siRNAs against 28 predicted targets of miR-  
14  
15 501-3p and/or miR-619-3p, and infected with HCVcc JcR2A according to the two-step  
16  
17 protocol depicted in Fig. 1A. siCD81, antagomiR-122 and siApoE were used as loss-of-  
18  
19 function controls to perturb HCV entry, translation/replication and assembly, respectively.  
20  
21 miR-501-3p and miR-619-3p, which were ineffective in part 1 of the screen but enhanced  
22  
23 HCV infection in part 2, were transfected in parallel. HCV infection was quantified as fold  
24  
25 change of luciferase activity with respect to negative control (siCtrl). Results for different  
26  
27 replicates are shown as individual points. For each gene, median fold change of luciferase  
28  
29 activity  $\pm$  s.d. is shown as black horizontal lines. The dashed line indicates a fold change of  
30  
31 1. Data are from three independent experiments in triplicate. Results for miR-501-3p, miR-  
32  
33 619-3p and siOGT that increase HCV infection in part 2 are depicted in red. Results for  
34  
35 siRNA targeting *PPP3CA*, *CEBPA*, *MID1*, *WDFY3*, *DCX*, *SLC35D1*, *CSDE1*, *GAN*, *USP37*,  
36  
37 *MAPK9*, *DCC*, *RNF144A*, or *PPP2R2C* that significantly modulated HCV infection in part 1  
38  
39 and/or part 2 but did not phenocopy the effect of miR-501-3p and miR-619-3p are depicted in  
40  
41 blue.  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

21  
22 **Figure 3. miR-501-3p mediates post-transcriptional regulation of OGT by decreasing**

23 **its expression at the protein level.** Huh7.5.1 cells were transfected with siCtrl (Ctrl), a pool  
24  
25 of siRNA against *OGT*, miR-501-3p or miR-619-3p. After 96h, RNA and proteins were  
26  
27 purified, and *OGT* expression analyzed by RT-qPCR and Western blot. (A) Percentage of  
28  
29 *OGT* mRNA expression in miRNA-transfected cells as compared to negative control. Results  
30  
31 are presented as mean  $\pm$  s.d. and are from three independent experiments in triplicate. The  
32  
33 dashed line indicates values from control-transfected cells set at 100%. Statistics: \*,  $p$ -value

1 ≤ 0.05, t-test (B) OGT protein expression. Left: percentage of OGT protein expression in  
2 siRNA- or miRNA-transfected cells as assessed by quantification of Western blots. OGT  
3 levels were normalized to actin levels using ImageLab™ 5.2.1 software (BioRad). Results  
4 are presented as mean ± s.d. and are from three independent experiments. The dashed line  
5 indicates values from control-transfected cells set at 100%. Statistics: \*,  $p$ -value ≤ 0.05, t-  
6 test. Right: representative Western blot analysis. (C) Analysis of miRNA targeting of *OGT*  
7 expression by dual luciferase reporter assay. Left: HeLa cells were co-transfected with a  
8 miR-501-3p mimic and a dual luciferase reporter plasmid containing either wild type miR-  
9 501-3p (RLuc wt *OGT* 3'UTR) or mutated miR-501-3p binding site (RLuc mt *OGT* 3'UTR) to  
10 modulate RLuc expression. Co-transfection of the miR-501-3p mimic and empty RLuc vector  
11 was used as control. Data are expressed as mean percentage of *Renilla* luciferase activity ±  
12 s.d. normalized to *firefly* luciferase, and relative to co-transfection of the vectors with non-  
13 targeting miRNA (miR-Ctrl). Results are from three independent experiments in triplicate.  
14 The dashed line indicates values from control-transfected cells set at 100%. Statistics: \*,  $p$ -  
15 value ≤ 0.05, t-test. Right: Schematic representation of the used constructs.

16  
17 **Figure 4. Silencing of *OGT* affects HCV morphogenesis and infectivity.** (A) Analysis of  
18 HCV infectivity. Huh7.5.1 cells were transfected with siCtrl, a pool of siRNA against *OGT* or  
19 ApoE as a loss-of-function control to perturb HCV assembly, prior to infection with HCVcc  
20 (Jc1) two days later (entry and replication). Mock-transfected cells were used as control  
21 (Ctrl). After another 48h, intra- and extracellular HCVcc particles were used to infect naïve  
22 Huh7.5.1 cells (assembly and infectivity). Virus supernatants of Huh7.5.1 cells were assayed  
23 by (left) endpoint dilution assay (TCID50). Intra- and extracellular HCV RNA was purified and  
24 analyzed by RT-qPCR to calculate (right) the specific infectivity (TCID50/RNA). Data are  
25 expressed as mean percentage as compared to control ± s.d. Results are from four  
26 independent experiments in triplicate. The dashed line indicates values from control-  
27 transfected cells set at 100%. Statistics: \*,  $p$ -value ≤ 0.05, Mann-Whitney test. (B) Genotype-  
28 independent effect of *OGT* on HCV infection. Huh7.5.1 cells were transfected with siCtrl or

1  
2  
3 1 siOGT prior to infection with HCVcc JcR2a (genotype 2a), H77R2a (genotype 1a) or  
4  
5 2 Con1R2a (genotype 1b). Experiments were carried out and analyzed as described in A. Data  
6  
7 3 are expressed as mean percentage of Renilla luciferase activity as compared to control  $\pm$  s.d.  
8  
9 4 Results are from three independent experiments in quadruplicate. The dashed line indicates  
10  
11 5 values from control-transfected cells set at 100%. Statistics: \*,  $p$ -value < 0.05, Mann-Whitney  
12  
13 6 test. (C) Activity of OGT/OGA inhibitors on O-GlcNAcylation. The activity of Ac<sub>4</sub>5S-GlcNAc  
14  
15 7 (OGT inhibitor) or Thiamet G (OGA inhibitor) on O-GlcNAcylation of proteins in Huh7.5.1  
16  
17 8 cells was demonstrated by Western blot as described in Supplementary Methods. (D) Effect  
18  
19 9 of O-GlcNAcylation on HCV infectivity. Huh7.5.1 cells were electroporated with HCVcc  
20  
21 10 (JcR2a), prior to treatment with increasing concentrations of Ac<sub>4</sub>5S-GlcNAc (OGT inhibitor,  
22  
23 11 left) or Thiamet G (OGA inhibitor, right) 4h later. After 96h, supernatants were transferred  
24  
25 12 onto naïve Huh7.5.1 cells and electroporated cells were lysed to determine luciferase  
26  
27 13 activity. Luciferase activity in infected Huh7.5.1 cells was assessed 72h later. Data are  
28  
29 14 expressed as mean percentage as compared to control  $\pm$  s.d. Results are from three  
30  
31 15 independent experiments in quadruplicate. The dashed line indicates values from vehicle-  
32  
33 16 treated cells set at 100%. Statistics: \*,  $p$ -value < 0.05, Mann-Whitney test.  
34  
35  
36  
37  
38

39 18 **Figure 5. Silencing of OGT modulates HCVcc biophysical properties.** (A) Separation of  
40  
41 19 HCVcc by iodixanol density gradient ultracentrifugation. HCVcc were produced in non-  
42  
43 20 targeting siRNA control- or siOGT-transfected Huh7.5.1 cells. After overlaying HCVcc  
44  
45 21 (JcR2A) on a 4%-40% iodixanol step gradient and ultracentrifugation for 16h, fractions of  
46  
47 22 HCV particles were used to infect naïve Huh7.5.1 cells in order to determine TCID<sub>50</sub>. HCV  
48  
49 23 RNA of each fraction was purified and analyzed by RT-qPCR. Data are expressed as mean  $\pm$   
50  
51 24 s.d. from three independent experiments. (B) Specific infectivity (TCID<sub>50</sub>/RNA) was  
52  
53 25 calculated and the density was determined by weighting each fraction. Specific infectivity of  
54  
55 26 each fraction is expressed as fold change as compared to the total infectivity of the control.  
56  
57 27 Data are expressed as mean  $\pm$  s.d. from three independent experiments. (C-D) ApoE and  
58  
59 28 ApoB concentrations in the individual fractions were determined by ELISA. The dashed lines  
60

1  
2  
3 1 indicate limits of quantification of the assays. Data are expressed as mean  $\pm$  s.d. from three  
4  
5 2 independent experiments.  
6  
7 3

9 4 **Figure 6. Silencing of OGT increases the size of HCVcc.** (A) Representative pictures of  
11 5 HCV particles generated in Huh7.5.1 cells transfected with non-targeting siRNA (siCtrl) or  
12 6 siOGT. (B-F) Comparative analysis of particle size distribution for immunocapture (IC) from  
13 7 HCV particles produced in Huh7.5.1 cells transfected with siCtrl or siOGT prior to infection  
14 8 with HCVcc (JcR2a) following sucrose-cushion purification (B) or iodixanol gradient  
15 9 fractionation (C-F) of HCVcc. HCVcc were transferred via anti-E2 antibody AR3A on electron  
16 10 microscopy (EM) grids through IC. Particle size distribution was assessed from a series of  
17 11 randomly acquired electron micrographs with Image-J software (NIH). Results from one of  
18 12 three (A-B) or two (C-F) independent experiments are shown. Black lines: size distribution of  
19 13 immunocaptured HCVcc produced in siCtrl-transfected cells. Grey lines: size distribution of  
20 14 immunocaptured HCVcc produced in siOGT-transfected cells.  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34

35 16 **Figure 7. OGT expression increases in HCC.** (A-B) Huh7.5.1 cells were infected with HCV  
36 17 (JcR2a). After 72h, RNA and proteins were purified, and OGT expression analyzed by RT-  
37 18 qPCR and Western blot. (A) Percentage of OGT mRNA expression relative to uninfected  
38 19 Huh7.5.1 cells (Ctrl). Results are presented as mean  $\pm$  s.d. from three independent  
39 20 experiments in duplicate. The dashed line indicates values from uninfected Huh7.5.1 cells  
40 21 set at 100%. Statistics: \*,  $p$ -value < 0.05, Mann-Whitney test. (B) OGT protein expression.  
41 22 Left: percentage of OGT protein expression relative to uninfected Huh7.5.1 cells (Ctrl)  
42 23 following quantification of Western blots as described in Supplementary Methods. Results  
43 24 are presented as mean  $\pm$  s.d. from three independent experiments. The dashed line  
44 25 indicates values from uninfected Huh7.5.1 cells set at 100%. Statistics: \*,  $p$ -value < 0.05,  
45 26 Mann-Whitney test. Right: representative Western blot analysis of OGT and actin. (C) OGT  
46 27 expression and viral load in liver tissue from 22 HCV-infected patients and 6 patients not  
47 28 infected with HCV described in[55]. Spearman correlation:  $\rho = 0.06004019$ ,  $p$ -value = 0.77.  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 1 (D-E) OGT expression in liver tissue from 22 HCV-infected patients and 6 patients not  
4  
5 2 infected with HCV according to fibrosis (D) or activity (E) scores described in[55]. Wilcoxon  
6  
7 3 test: F1 vs F0 p-value = 0,38; F2 vs F0 p-value = 0,18; F3 vs F0 p-value = 0,43; F4 vs F0 p-  
8  
9 4 value = 0,17; A1 vs A0 p-value = 0,28; A2 vs A0 p-value = 0,23; A3 vs A0 p-value = 0,09. (F)  
10  
11 5 OGT expression in tumor (HCC) and non-tumor (Ctrl) liver tissue from 39 HCV-infected  
12  
13 6 patients, 83 HBV-infected, 80 patients with alcoholic liver disease (ALD) and 13 patients with  
14  
15 7 non-alcoholic liver disease (NAFLD) as described in Supplementary Methods. \*, p-value <  
16  
17 8 0.05, Wilcoxon test.  
18  
19  
20 9  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60



1  
2  
3 **Table 1. Computational analysis of miR-501-3p and miR-619-3p targets and pathway**  
4  
5 **enrichment.**  
6

miRNA ID	Target gene symbol	<u>Pathway</u> or network	
miR-501-3p	<i>MEF2A; PPP3CA; PPP3CC</i>	<u>Calcium signaling</u>	
	<i>HMGCS1</i>	<u>Cholesterol biosynthesis</u>	
	<i>AFF4; CHMP1B; CUX1; DCLK1;</i> <i>LMX1A; PTBP2; RBMS1; RC3H1;</i> <i>SCN2A; SEC63; ZFHX4</i>	Inflammatory response, dermatological diseases and conditions, inflammatory disease	
	<i>CDK6; CSDE1; GLI2; HOXD10;</i> <i>LSM5; MEF2A; MYCN; OGT;</i> <i>PPP2R2C; PPP2R5E; SEMA3C;</i> <i>TFDP2</i>	Cellular development, nervous system development and function; organ morphology	
	<i>CIT; COL10A1; FNBP1L; GAN;</i> <i>HERC1; KPNA4; NONO; SHPRH;</i> <i>STRN; TARDBP; UBE2H; USP37</i>	Cell death and survival; cellular compromise; free radical scavenging	
	<i>ATXN1; CBLL1; CEBPA; DCC;</i> <i>PEX5L; RCC2; RNF144A; ZC3H12C</i>	Cell morphology, cellular assembly and organization; cellular function and maintenance	
	miR-619-3p	<i>RUNX1T1; SMAD3</i>	<u>Adipocyte biogenesis</u>
		<i>FOXG1; GPBP1; MID1; MKL2; MSI1;</i> <i>PCBP2; WDFY3</i>	Cell cycle; organismal injury and abnormalities; cancer
		<i>ACVR2B; DCX; ESRRG; MAPK9;</i> <i>OGT; PCBP1; PDE3B; SMAD3;</i> <i>SMARCC1; TGFB3; PAPOLA</i>	Carbohydrate metabolism, energy production; small molecule biochemistry
		<i>RUNX1T1; SHANK2; SLC35D1</i>	Gene expression, lipid metabolism, small molecule biochemistry

3

4

## Supplementary Information

### A functional microRNA screen uncovers O-linked N-acetylglucosamine transferase as a host factor modulating hepatitis C virus morphogenesis

Katharina Herzog<sup>1,2\*</sup>, Simonetta Bandiera<sup>1,2\*</sup>, Sophie Pernot<sup>1,2</sup>, Catherine Fauvelle<sup>1,2</sup>, Frank Jühling<sup>1,2</sup>, Amélie Weiss<sup>2,3,4,5</sup>, Anne Bull<sup>6</sup>, Sarah C. Durand<sup>1,2</sup>, Béatrice Chane-Woon-Ming<sup>2,7</sup>, Sébastien Pfeffer<sup>2,7</sup>, Marion Mercey<sup>8</sup>, Hervé Lerat<sup>8</sup>, Jean-Christophe Meunier<sup>6</sup>, Wolfgang Raffelsberger<sup>2,3,4,5</sup>, Laurent Brino<sup>2,3,4,5</sup>, Thomas F. Baumert<sup>1,2,9,#</sup>, Mirjam B. Zeisel<sup>1,2,10,#</sup>

<sup>1</sup>Inserm, U1110, Institut de Recherche sur les Maladies Virales et Hépatiques, Strasbourg, France; <sup>2</sup>Université de Strasbourg, Strasbourg, France; <sup>3</sup>Institut de Génétique et de Biologie Moléculaire et Cellulaire, Illkirch, France; <sup>4</sup>CNRS, UMR7104, Illkirch, France; <sup>5</sup>Inserm, U1258, Illkirch, France; <sup>6</sup>Inserm U1259, Faculté de Médecine, Université François Rabelais and CHRU de Tours, Tours, France; <sup>7</sup>Architecture et Réactivité de l'ARN – UPR 9002, Institut de Biologie Moléculaire et Cellulaire du CNRS, Strasbourg, France; <sup>8</sup>Institute for Advanced Biosciences, Centre de Recherche UGA - Inserm U1209 - CNRS UMR 5309, Grenoble, France, <sup>9</sup>Institut Hospitalo-Universitaire, Pôle Hépato-digestif, Hôpitaux Universitaires de Strasbourg, Strasbourg, France; <sup>10</sup>Inserm, U1052, CNRS UMR 5286, Centre Léon Bérard (CLB), Cancer Research Center of Lyon (CRCL), Université de Lyon (UCBL), Lyon, France

\*Authors contributed equally to this work

#### Supplementary Material and methods

**Cells and cell culture conditions.** The source and culture conditions of Huh7.5.1 cells have been described[1]. HeLa cells were purchased from ATCC and cultured in Dulbecco's modified Eagle medium (Gibco® DMEM GlutaMAX™, ThermoFisher Scientific) containing 1% sodium pyruvate as described for Huh7.5.1 cells[1].

1  
2  
3 1 **Viruses and infectivity assays.** Cell culture-derived recombinant cell culture-derived  
4 hepatitis C virus (HCVcc) Jc1 (genotype 2a/2a chimera), H77R2a (genotype 1a/2a chimera  
5 engineered for *Renilla* luciferase expression), Con1R2a (genotype 1b/2b chimera engineered  
6 for *Renilla* luciferase expression), and JcR2a (genotype 2a/2a chimera engineered for *Renilla*  
7 luciferase expression) were generated in Huh7.5.1 cells as described[1, 2, 3, 4]. HCVcc  
8 infectivity was determined by calculating the 50% tissue culture infectious dose (TCID50)  
9 using anti-NS5A antibody as described[5, 6] or by assessing luciferase activity. HCVcc were  
10 used at 10<sup>5</sup>-10<sup>6</sup> TCID50/mL throughout the study. HCV RNA was purified using a QIAmp viral  
11 RNA minikit (Qiagen) and analyzed by one-step RT-qPCR using a Sensi Fast NO ROX kit  
12 (Bioline) according to the manufacturer's instructions. Standard curves were performed using  
13 10-fold dilution series of HCV RNA.

14  
15  
16 13 **Purification of HCVcc particles using sucrose cushion or iodixanol density gradient.**  
17 HCVcc (JcR2a) were concentrated 10-fold using a Vivaspin column (GE Healthcare). For  
18 sucrose cushion purification, HCVcc were purified by overlaying 3.5 mL of culture media on  
19 1.5 mL of 20% sucrose, and by ultracentrifuging samples for 4h at 40,000 rpm on a SW-55  
20 rotor (Beckman Coulter). Purified HCVcc were resuspended in 30 µL of PBS for analysis via  
21 immunocapture and electron microscopy. Density distributions of infectious HCVcc were  
22 determined by overlaying 0.5 mL culture media on a 5 mL, 4%-40% iodixanol step gradient,  
23 and ultracentrifuging samples for 16h at 40,000 rpm on a SW-55 rotor (Beckman Coulter): 625  
24 µl fractions were carefully harvested from the top of each tube, and density was determined  
25 by weighing. Infectivity of each fraction was quantified by TCID50 using anti-NS5A antibody  
26 as described[5, 6], while HCV RNA of fractions was purified and analyzed as described above.  
27 ApoB and ApoE concentrations of fractions were determined by enzyme-linked  
28 immunosorbent assay (Human Apolipoprotein B or E ELISA<sup>PRO</sup> kit, Mabtech) undiluted or in a  
29 1:50 dilution, respectively, according to the manufacturer's instructions (Mabtech).

1  
2  
3 1 **miRNA mimics and siRNAs.** Non-targeting control miRNA, miR-501-3p mimic, miR-619-3p  
4 mimic, antagomiR-122, antagomiR-501-3p, non-targeting control antagomiR, non-targeting  
5 control siRNA, siRNAs targeting *OGT*, *CD81* or *apolipoprotein E (ApoE)* and a library of 28  
6 custom ON-TARGETplus smart pool siRNAs were purchased from Dharmacon (GE  
7 Healthcare).

8  
9  
10 7 **miRNA expression analysis.** Total RNA (100 ng) was purified from control or HCV-infected  
11 Huh7.5.1 cells using Tri reagent® (Thermo Scientific) and Direct-zol™ RNA purification kit  
12 (Zymo Research). Total RNA was first polyadenylated and reverse transcribed using a  
13 miScript II RT system (Qiagen) according to the manufacturer's instructions. The obtained  
14 cDNA was subjected to RT-qPCR using miScript SYBR Green kit (Qiagen). Primers were the  
15 mature miRNA sequence for the forward primer (Thermo Scientific) and the universal miScript  
16 primer (Qiagen) for the reverse primer. Data were analyzed by the  $\Delta\Delta C_t$  method using small  
17 nucleolar RNA, C/D box 61 (SNORD61) as an endogenous reference and the non-infected  
18 samples as a calibrator[7].

19  
20  
21 17 **Antibodies.** Rabbit anti-OGT antibodies DM-17 and AL24 were purchased from Sigma or  
22 kindly provided by Dr. G. W. Hart and Dr. S. Hardivillé (Johns Hopkins University School of  
23 Medicine, Baltimore, MD)[8], respectively. Mouse anti- $\beta$ -actin antibody was purchased from  
24 Abcam and mouse, rabbit or sheep HRP-conjugated secondary antibodies (A9044, A0545  
25 and A3415, respectively) were purchased from Sigma. Sheep anti-NS5A serum for  
26 determination of TCID50 was a kind gift from M. Harris[9]. Human anti-E2 (AR3A) antibody[10]  
27 for electron microscopy analysis was kindly provided by Mansun Law (SCRIPPS, California,  
28 USA).

29  
30  
31 26 **Western blotting.** OGT and actin protein expression in human cells was assessed by  
32 Western blot as described[8] with some modifications. Briefly, cells were lysed in lysis buffer  
33 no. 6 (R&D Systems) according to the manufacturer's instructions. Equal amounts of protein

1 (40 µg) were size-separated through a Mini PROTEAN® TGX Stain-Free™ gel electrophoresis  
2 (Bio-Rad) and transferred to PVDF membranes (Bio-Rad). Immunoblots were performed  
3 using rabbit anti-OGT (1:2000) and mouse anti-β-actin (1:1000) antibodies[8, 11]. Antigen-  
4 antibody complexes were detected by incubating the membrane with the appropriate HRP-  
5 conjugated secondary antibodies (1:5000; 1:10,000) and imaged by enhanced  
6 chemiluminescence with a ChemiDoc MP imager (Bio-Rad). Quantification of protein  
7 expression was performed using ImageLab™ 5.2.1 software (BioRad). For analysis of OGT  
8 and GAPDH expression in liver tissue from HCV transgenic (FL-N/35) or wild-type mice[12],  
9 crude protein extracts were prepared by homogenization of frozen mouse livers (50–100 µg)  
10 in tissue lysis buffer from the Ambion PARIS RNA (Thermo Scientific) and protein isolation kit,  
11 supplemented with protease inhibitors (cOmplete™ EDTA-free protease inhibitor mixture,  
12 Sigma-Aldrich) and phosphatase inhibitors (PhosSTOP™, Sigma-Aldrich), using a tissue  
13 homogenizer (MP Fast Prep24, MP Biomedicals, Santa Ana, CA) and MP Lysing Matrix A  
14 tubes. Proteins were quantified using the BCA assay (Thermo Fisher Scientific). Western  
15 blotting was performed as described above.

16  
17 **Immunocapture and electron microscopy analysis of viral particles.** Sucrose-cushion  
18 purified or iodixanol gradient fractionated HCVcc (JcR2a) produced in cells transfected with a  
19 non-targeting siRNA control or a pool of siRNA against OGT were transferred via anti-E2  
20 antibody AR3A on electron microscopy (EM) grids through immunocapture (IC) as  
21 described[13]. Particles were stained with uranyl acetate dihydrate and observed in a JEOL  
22 1230 electron microscope. Series of electron micrographs were acquired at random from IC  
23 EM grids. The images were then analyzed with Image-J software, to determine the particle  
24 size distribution.

25  
26 **Gene expression analysis in patient-derived liver tissue.** For OGT expression analysis in  
27 patient's samples, raw data were retrieved from the Gene Expression Omnibus (GSE84346)  
28 and re-analyzed by quality-trimming (cutadapt) and mapping (HISAT2) to human genome

1 assembly hg19. Reads mapping to Gencode v.19 genes were counted using htseq-count and  
2 normalized applying DESeq2. Activity and fibrosis scores as well as viral load were taken from  
3 the supplemental data published in[14]. To analyze OGT expression in liver tissue of chronic  
4 hepatitis B or C patients, FPKM values and clinical data were retrieved from The Cancer  
5 Genome Atlas (TCGA, [https://www.cancer.gov/about-](https://www.cancer.gov/about-nci/organization/ccg/research/structural-genomics/tcga)  
6 [nci/organization/ccg/research/structural-genomics/tcga](https://www.cancer.gov/about-nci/organization/ccg/research/structural-genomics/tcga)). This data set includes samples from  
7 39 HCV-infected patients, 83 hepatitis B virus (HBV)-infected, 80 patients with alcoholic liver  
8 disease (ALD) and 13 patients with non-alcoholic fatty liver disease (NAFLD).

### 10 **Supplementary References**

- 11 1 Zhong J, Gastaminza P, Cheng G, *et al.* Robust hepatitis C virus infection in vitro. Proc  
12 Natl Acad Sci U S A 2005;**102**:9294-9.
- 13 2 Reiss S, Rebhan I, Backes P, *et al.* Recruitment and activation of a lipid kinase by  
14 hepatitis C virus NS5A is essential for integrity of the membranous replication compartment.  
15 Cell Host Microbe 2011;**9**:32-45.
- 16 3 Da Costa D, Turek M, Felmlee DJ, *et al.* Reconstitution of the entire hepatitis C virus  
17 life cycle in non-hepatic cells. J Virol 2012;**86**:11919-25.
- 18 4 Fauvelle C, Felmlee DJ, Crouchet E, *et al.* Apolipoprotein E Mediates Evasion From  
19 Hepatitis C Virus Neutralizing Antibodies. Gastroenterology 2016;**150**:206-17 e4.
- 20 5 Lindenbach BD, Evans MJ, Syder AJ, *et al.* Complete replication of hepatitis C virus in  
21 cell culture. Science 2005;**309**:623-6.
- 22 6 Bandiera S, Pernet S, El Saghire H, *et al.* Hepatitis C Virus-Induced Upregulation of  
23 MicroRNA miR-146a-5p in Hepatocytes Promotes Viral Infection and Deregulates Metabolic  
24 Pathways Associated with Liver Disease Pathogenesis. J Virol 2016;**90**:6387-400.
- 25 7 Schmittgen TD, Livak KJ. Analyzing real-time PCR data by the comparative C(T)  
26 method. Nat Protoc 2008;**3**:1101-8.
- 27 8 Iyer SP, Akimoto Y, Hart GW. Identification and cloning of a novel family of coiled-coil  
28 domain proteins that interact with O-GlcNAc transferase. J Biol Chem 2003;**278**:5399-409.

1  
2  
3 1 9 Macdonald A, Crowder K, Street A, *et al.* The hepatitis C virus non-structural NS5A  
4 protein inhibits activating protein-1 function by perturbing ras-ERK pathway signaling. *J Biol*  
5 *Chem* 2003;**278**:17775-84.

6  
7  
8  
9 4 10 Giang E, Dorner M, Prentoe JC, *et al.* Human broadly neutralizing antibodies to the  
10 envelope glycoprotein complex of hepatitis C virus. *Proc Natl Acad Sci U S A* 2012;**109**:6205-  
11 10.

12  
13  
14  
15 7 11 Verrier ER, Colpitts CC, Bach C, *et al.* A targeted functional RNA interference screen  
16 uncovers glypican 5 as an entry factor for hepatitis B and D viruses. *Hepatology* 2016;**63**:35-  
17 48.

18  
19  
20  
21 10 12 Lerat H, Honda M, Beard MR, *et al.* Steatosis and liver cancer in transgenic mice  
22 expressing the structural and nonstructural proteins of hepatitis C virus. *Gastroenterology*  
23 **2002**;**122**:352-65.

24  
25  
26  
27 13 13 Piver E, Boyer A, Gaillard J, *et al.* Ultrastructural organisation of HCV from the  
28 bloodstream of infected patients revealed by electron microscopy after specific  
29 immunocapture. *Gut* 2017;**66**:1487-95.

30  
31  
32  
33 16 14 Boldanova T, Suslov A, Heim MH, *et al.* Transcriptional response to hepatitis C virus  
34 infection and interferon-alpha treatment in the human liver. *EMBO Mol Med* 2017;**9**:816-34.

### 35 36 37 38 39 40 41 **Supplementary figure legends**

42  
43 20 **Figure S1.** (A) Effect of miR-501-3p inhibition on HCV infectivity. Huh7.5.1 cells were  
44 transfected with control antagomiR (Ctrl), antagomiR-122 as loss-of-function control to perturb  
45 HCV replication and antagomiR-501-3p, prior to infection with HCVcc (JcR2a) according to  
46 the two-step protocol depicted in Fig. 1A. After 48h, supernatants were transferred onto naive  
47 Huh7.5.1 cells. After 72h, Renilla Luciferase activity of infected Huh7.5.1 cells was  
48 determined. Data are expressed as mean percentage as compared to Ctrl  $\pm$  s.d. Results are  
49 from four independent experiments in quadruplicate. The dashed line indicates values from  
50 vehicle-treated cells set at 100%. Statistics: \*, *p*-value < 0.05, Mann-Whitney test. (B) miR-  
51 501-3p expression upon HCV infection. Huh7.5.1 cells were infected with HCVcc (JcR2a).  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 1 After 72h, RNA was purified and miR-501-3p expression analyzed by RT-qPCR. Percentage  
4  
5 2 of miR-501-3p expression relative to uninfected Huh7.5.1 cells (Ctrl). Results are presented  
6  
7 3 as mean  $\pm$  s.d. from three independent experiments in duplicate. The dashed line indicates  
8  
9 4 values from uninfected Huh7.5.1 cells set at 100%. Statistics: \*,  $p$ -value < 0.05, Mann-Whitney  
10  
11 5 test.  
12  
13  
14 6  
15

16 7 **Supplementary Table 1. A genome-wide miRNA mimic screen identifies cellular**  
17  
18 8 **miRNAs modulating HCV infection.** Log<sub>2</sub>(FC), lfd<sub>r</sub> and effect on HCV infection in part 1 and  
19  
20 9 part 2 of the screen are shown for the individual miRNAs of the miRNA mimic library. In red:  
21  
22 10 proviral effect, in blue: antiviral effect. FC: fold change, lfd<sub>r</sub>: local false discovery rate  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60