Durrington 38

Supplementary Fig 1. Time of HDM challenge effects AHR to methacholine,
 measured as airway resistance (area under the curve, AUC, rather than
 maximum resistance as in Fig 2) in wild type (WT) mice.

a. Airway resistance to increasing doses of methacholine was measured by area under
the curve, AUC, in mice challenged with either HDM or PBS at ZT11 or ZT23. There
was a significant time of day difference in airway resistance in mice challenged with
HDM (P= 0.005), mixed linear modelling (n=8-9, per treatment group). Mice
challenged with HDM at ZT11 showed increased airway resistance compared to those
challenged at ZT23.

b. HDM specific IgE was measured in serum from WT mice. There were significantly
 increased levels of HDM specific IgE in WT mice treated with intranasal HDM
 compared to PBS treated control mice (\* P < 0.05 at ZT23 and \*\*\* P < 0.001 at ZT11).</li>
 There were no time of challenge differences in control or HDM treated groups (Mean
 ± SEM, (n=5-8, per treatment group) Mann Whitney U).

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### 16 Supplementary Fig 2. REV-ERBα acts as a repressor of AHR

**a.** Effect of time of HDM challenge on AHR in Rev-erb $\alpha$ -/- mice. Airway resistance measured as area under the curve, AUC, rather than as maximum airway resistance as in Fig 4. There was a significant increase in airway resistance (AUC) after HDM challenge at ZT11 (\*\*\* P < 0.001) and ZT23 (\*\*\* P < 0.001), compared to control, PBS challenged mice. There was no time of challenge difference in airway resistance (AUC) after PBS challenge or after HDM challenge, mixed linear modelling, (n=7-9 per treatment group). b. Maximum airway resistance (cmH20.s/ml) was measured in PBS challenged WT
and Rev-erbα<sup>-/-</sup> mice to increasing doses of nebulised methacholine. All
measurements of maximum airway resistance were increased in the Rev-erbα<sup>-/-</sup> mice
compared to WT mice.

c. HDM specific IgE was measured in serum from Rev-erbα-/- mice. HDM specific
serum IgE was significantly increased following HDM challenge, compared to control
mice (\*\*P < 0.01 at ZT23 and \*\* P < 0.01 at ZT11). There were no time of challenge</li>
differences in control or HDM treated groups (Mean ±SEM (n=4-7 per treatment
group), Mann Whitney U).

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Supplementary Fig. 3. Genotype differences in airway and lung inflammatory
 cells.

a. BAL inflammatory cell counts measured as a percentage of the total cell count, were analysed in WT and Rev-erb $\alpha^{-/-}$  mice to determine genotype differences. There were no significant differences between the groups. Mean ± SEM (n= 8-12 per treatment group), 1 way ANOVA, followed by Tukey multiple comparison test.

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#### 18 Supplementary Fig 4. Expression of beta adrenoceptors in murine lung.

a. Quantitative PCR for myosin light chain kinase 1 (*mlck1*) in mouse lung. There were
 no time of day or genotype differences in expression between groups.

b. Quantitative PCR for smooth muscle myosin (*sm-mhc*) in mouse lung. There were
no time of day or genotype differences in expression between groups.

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- 1 c. Quantitative PCR for smooth muscle actin (*acta*) in mouse lung. There were no time
- 2 of day or genotype differences in expression between groups.
- 3 **d.** Quantitative PCR for Adrenoceptor Beta 1 (*Adrb1*) in mouse lung tissue. There were
- 4 no time of day or genotype differences in expression of Adrb1.
- 5 e. Quantitative PCR for Adrenocpetor Beta 2 (*Adrb2*) in mouse lung tissue. There was
- 6 no time of day or genotype differences in expression of Adrb2. All data presented as
- 7 mean ± SEM (n=5-9 per treatment group, in duplicate) and analysed by 1 way ANOVA,
- 8 followed by Tukey's multiple comparison test.
- 9 All QPCR data is compared to expression of the housekeeping gene *Gapdh* in WT
  10 PBS challenged mice at ZT23. Black bars indicate challenge at ZT11 and grey bars
  11 indicate challenge at ZT23.

Supplementary Fig 1.



a.

b.

## Supplementary Fig. 2







# Supplementary Fig. 3

a.





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### **Supplementary Fig. 4**



