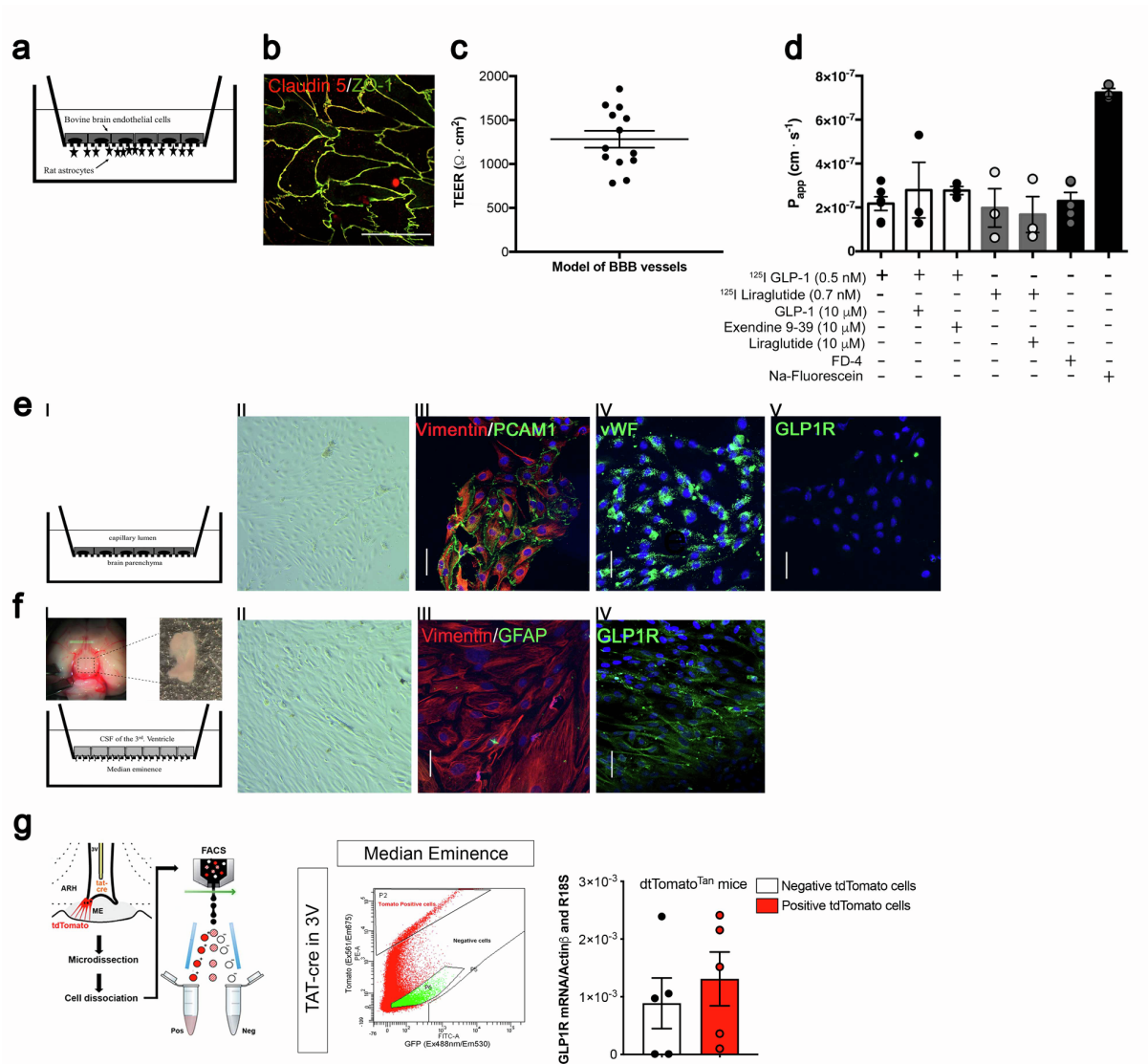


Supplemental information

**Tanycytes control hypothalamic liraglutide
uptake and its anti-obesity actions**

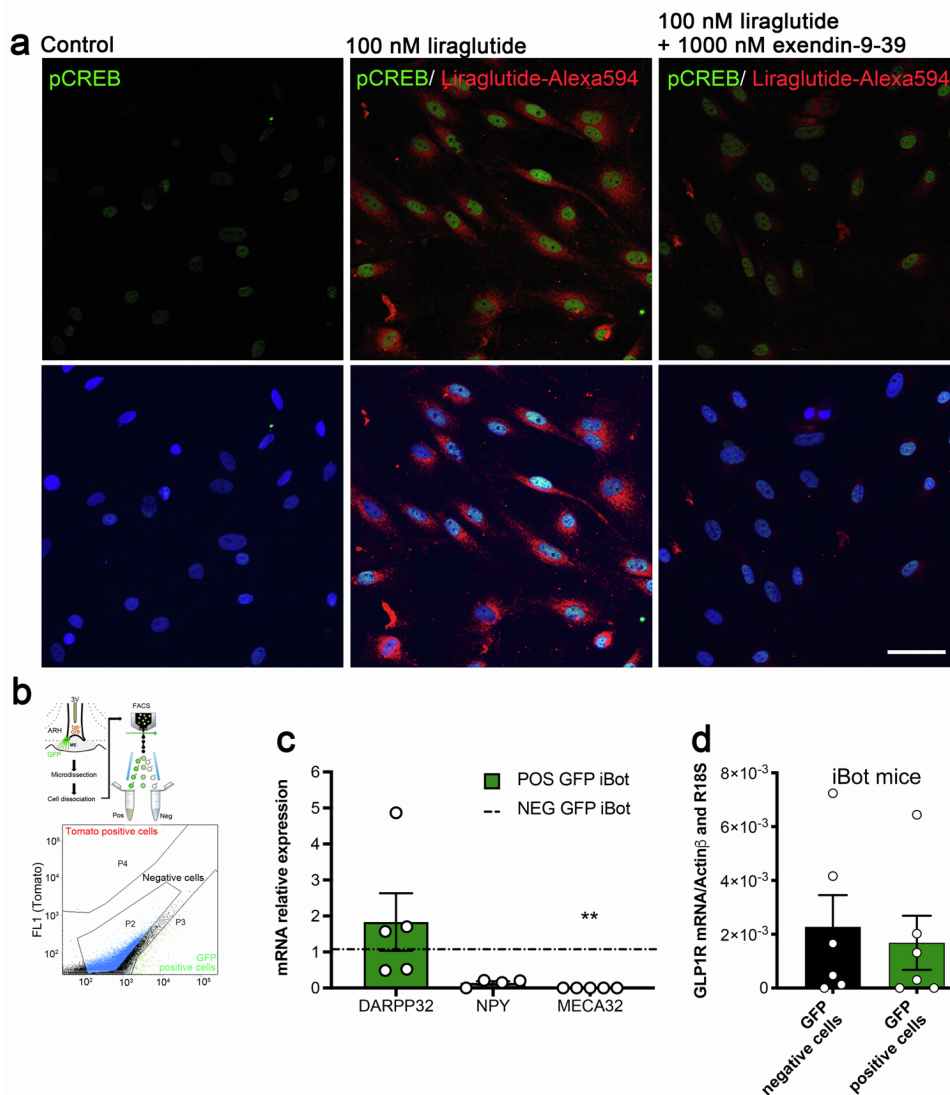
Monica Imbernon, Chiara Saponaro, Hans Christian Cederberg Helms, Manon Duquenne, Daniela Fernandois, Eleonora Deligia, Raphael G.P. Denis, Daniela Herrera Moro Chao, Sowmyalakshmi Rasika, Bart Staels, François Pattou, Frank W. Pfrieger, Birger Brodin, Serge Luquet, Caroline Bonner, and Vincent Prevot



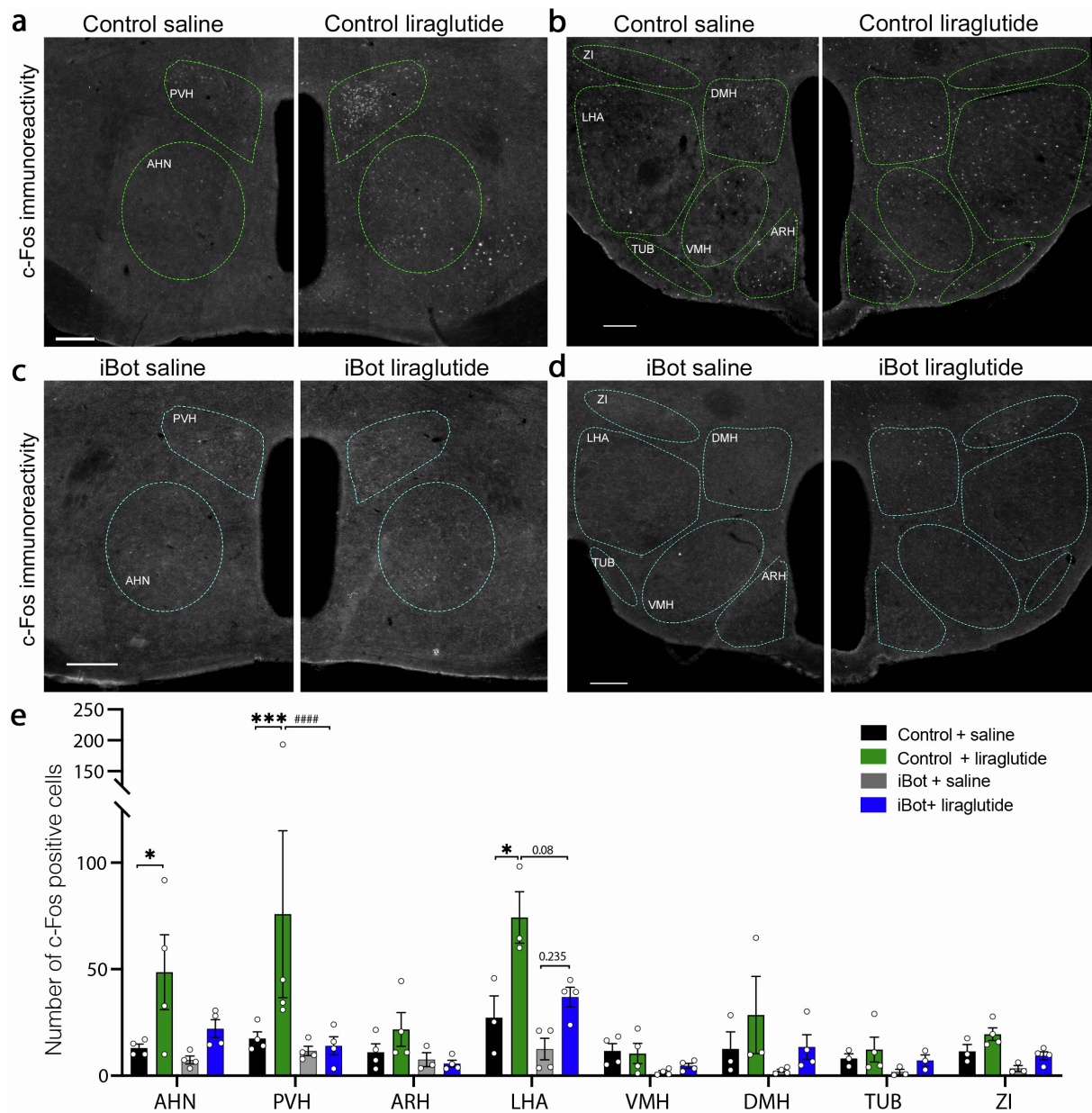
Supplementary Figure 1. Characterization and validation of *in vitro* blood-brain barrier models

(Related to Fig. 1). (a) Illustration of the bovine endothelial cell/rat astrocyte BBB contact co-culture model with endothelial cells cultured on transwell permeable supports and astrocytes covering the lower surface of the support. (b) Photomicrograph showing bovine endothelial cells in co-culture with rat astrocytes after immunocytochemical labeling with antibodies against the gap junction markers claudin-5 (green) and ZO-1 (red). Yellow color indicates co-localization of the markers. Scale bar 30 μm . (c) Trans-endothelial electrical resistance (TEER) was measured to validate the junctional integrity of the *in vitro* BBB model. Each data point represents the mean of several permeable supports (technical replicates) from an individual preparation. Line and whiskers indicate mean and SEM across biological replicates ($n = 13$). (d) Transcellular permeation of ^{125}I -labelled GLP-1 (0.5 nM) and liraglutide (0.7 nM) measured in the BBB model and compared to the permeation of 4 kDa FITC-dextran (FD-4) and sodium

fluorescein. Bars show means \pm SEM ($n = 6, 3, 3, 3, 3, 5, 3$ wells from 2 independent cultures). **(e)** Rat brain cortical endothelial cells on a transwell insert (I), bright-field photomicrograph of endothelial cell cultures from rat cortical capillaries (II), immunocytochemical labeling for the endothelial marker PECAM-1 (green), vimentin (red), a marker for tanycytes in the hypothalamus, von Willebrand's factor (green), which also labels endothelial cells, and GLP1R (green) (III-V); scale bar: 30 μ m. **(f)** Tanycytes cultured on transwell permeable supports (I), bright-field image of tanycytic cultures from median eminence explants (II), immunofluorescence labeling for vimentin (red), GFAP (green) and GLP1R (green) (III-IV) scale bar: 30 μ m. **(g)** Diagram and gating strategy for sorting Tomato-positive putative tanycytes following TAT-Cre infusion into the third ventricle (3V) of *tdTomato*^{loxP-STOP-loxP} mice and GLP1R mRNA expression in tanycytes isolated by FACS from *tdTomato* mice ($n = 5$).

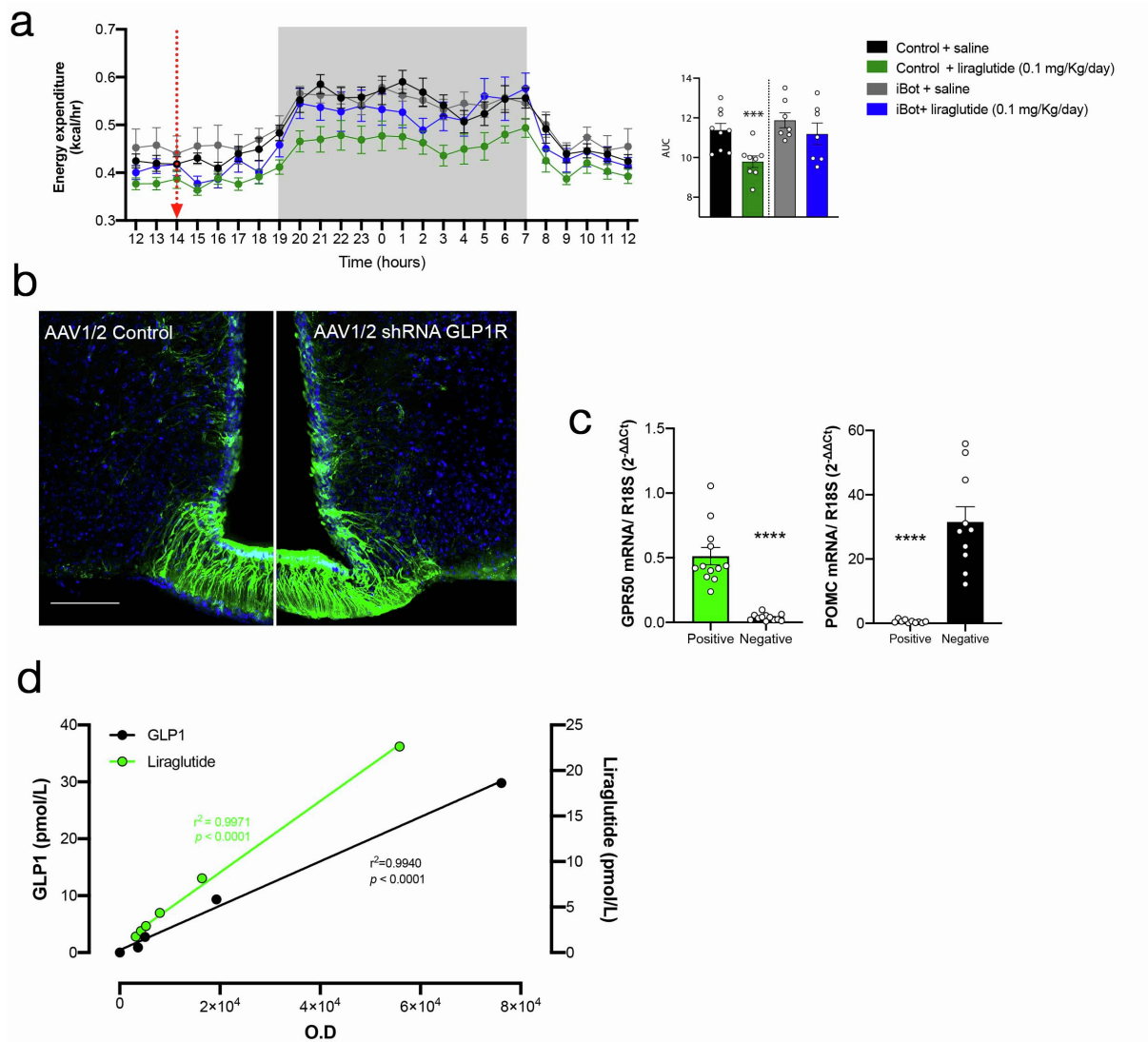


Supplementary Figure 2. Liraglutide⁵⁶⁴ is taken up by tanycytes, which express GLP1R *in vivo*, in a GLP1R-dependent manner and promotes CREB phosphorylation *in vitro* (Related to Fig. 1 and Fig. 2). (a) Photomicrographs of cultured tanycytes following immunocytochemical staining for pCREB (red) after treatment with control uptake buffer (left panel), 100 nM liraglutide⁵⁶⁴ (middle panel) and 1000 nM exendin 9-39 and subsequently with 100 nM liraglutide⁵⁶⁴; Scale bar: 30 μ m. (b) Diagram and gating strategy for sorting EGFP-positive putative tanycytes following TAT-Cre infusion into the third ventricle (3V) of BoNTB-EGFP^{loxP-STOP-loxP} mice (iBot mice). (c) EGFP-positive cells (green) express DARPP-32 mRNA (tanycytic marker), but not NPY mRNA (neuronal marker) or MECA32 mRNA (fenestrated endothelial cell marker). Expression in EGFP-negative cells has been normalized to 1 (dotted line). Mann-Whitney U test, ** $p=0.0079$, $n = 5$ mice. (d) GLP1R mRNA expression in EGFP-negative (black bar) and EGFP-positive (green bar) cells isolated from iBot mice; data are expressed as means \pm SEM. ** Mann-Whitney test, $p=0.0079$, $n = 6$ mice.



Supplementary Figure 3. Liraglutide-induced c-Fos activation is abolished in the hypothalamus of iBot mice selectively expressing botulinum toxin in tanycytes (Related to Fig. 2). (a-d) Representative photomicrographs of c-Fos immunohistochemistry in the hypothalamus of control (a, b) and iBot (c, d) mice 10 min after intravenous injection of saline (left panels) or liraglutide (90 nmol/Kg, right panels). AHN: anterior hypothalamic nucleus; ARH: arcuate nucleus of the hypothalamus; DMH: dorsomedial hypothalamic nucleus; LHA: lateral hypothalamic nucleus; PVH: paraventricular nucleus of the hypothalamus; TUB: tuberal nucleus; ZI: zona incerta. Scale bar 200 μ m. (e) Quantification of the number of c-Fos positive cells in mouse hypothalamic sections 10 minutes after either intravenous saline (black, grey) or liraglutide (90nmol/kg) injection (green, blue) in control (black and green) and iBot mice (grey and blue); $n = 4$ animals per group; 6 to 7 sections per animal. AHN: two-way ANOVA, genotype:

$F_{(1, 12)} = 3.09, p = 0.104$; treatment: $F_{(1, 12)} = 7.55, p = 0.017$; interaction: $F_{(1, 12)} = 0.276, p = 0.276$. Fisher's LSD *post hoc* test, control saline vs. control liraglutide, $p = 0.0176$. PVH: two-way ANOVA, genotype: $F_{(1, 12)} = 2.92, p = 0.113$; treatment: $F_{(1, 12)} = 2.39, p = 0.147$; interaction: $F_{(1, 12)} = 2.03, p = 0.1794$. Fisher's LSD *post hoc* test, control saline vs. control liraglutide, $p = 0.05$; control liraglutide vs. iBot liraglutide, $p = 0.046$. LHA: two-way ANOVA, genotype: $F_{(1, 10)} = 11.3, p = 0.007$; treatment: $F_{(1, 12)} = 21.31, p = 0.001$; interaction: $F_{(1, 10)} = 2.143, p = 0.174$. Fisher's LSD *post hoc* test, control saline vs. control liraglutide, $p = 0.002$; iBot saline vs. iBot liraglutide, $p = 0.0002$; control liraglutide vs. iBot liraglutide, $p = 0.006$. Data are expressed as means \pm SEM. * $p < 0.05$; *** $p < 0.001$, control saline vs. control liraglutide; #### $p < 0.0001$, control liraglutide vs. iBot liraglutide.



Supplementary Figure 4 (Related to Fig. 3 and Fig. 4). **Effect of liraglutide on energy expenditure in iBot mice.** (a) Energy expenditure in iBot and control mice after 3 days of intraperitoneal treatment with saline (baseline, grey and black respectively) or liraglutide (blue and green respectively, 0.1 mg/Kg/day). Line plot shows mean energy expenditure at different times 3 days after liraglutide treatment compared to baseline, and AUC. AUC paired two-tailed t-test, $t_{(7)} = 5.73$, $p = 0.0007$. Dotted arrow indicates saline or liraglutide administration ($n = 9, 8, 8, 8$ mice). Two-way ANOVA with Tukey's *post hoc* test was performed on AUC (control saline vs. control liraglutide, $p < 0.0001$; control liraglutide vs. iBot liraglutide, $p < 0.0001$) ($n = 9, 8, 7, 7$ mice). Data are expressed as means \pm SEM *** $p < 0.001$. (b) Selective expression of GFP in tanycytic cell bodies and processes in animals transduced with control and shRNA viruses. (c) Expression of GPR50 mRNA (tanycytic marker) but not POMC mRNA (neuronal marker) by

GFP-positive cells (green), and vice versa in GFP-negative cells (black) FACS-isolated from mice transduced with control-GFP or shRNA-GLP1R-GFP viral vectors. Unpaired one-tailed t-test (GPR50, positive vs. negative, $t_{(22)} = 7.03$, $p < 0.0001$, $n = 12$ mice; POMC, positive vs. negative, $t_{(18)} = 6.49$, $p < 0.0001$, $n = 10$ mice). (d) Correlation between optical density (O.D.) with total GLP1 (black line) or liraglutide (green line) amounts detected with the total GLP1 ELISA kit.