



## Supporting Information

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Polysaccharide-Polyplex Nanofilm Coatings Enhance  
Nanoneedle-Based Gene Delivery and Transfection  
Efficiency

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## Supplementary Information

### **Polysaccharide-polyplex nanofilm coatings enhance nanoneedle-based gene delivery and transfection efficiency.**

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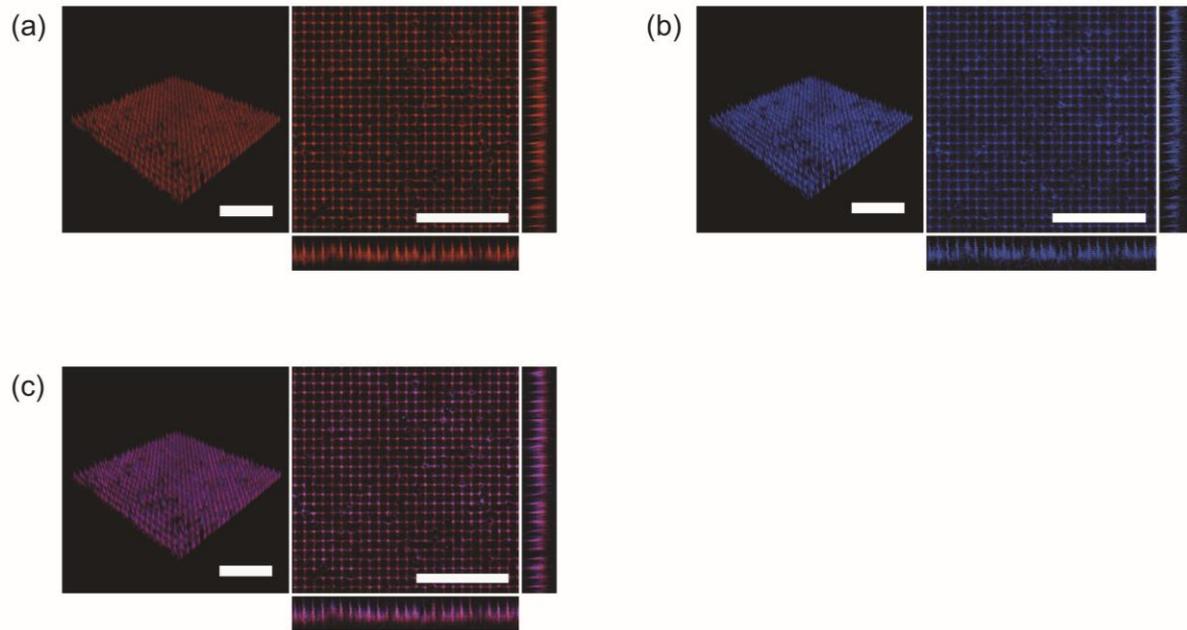
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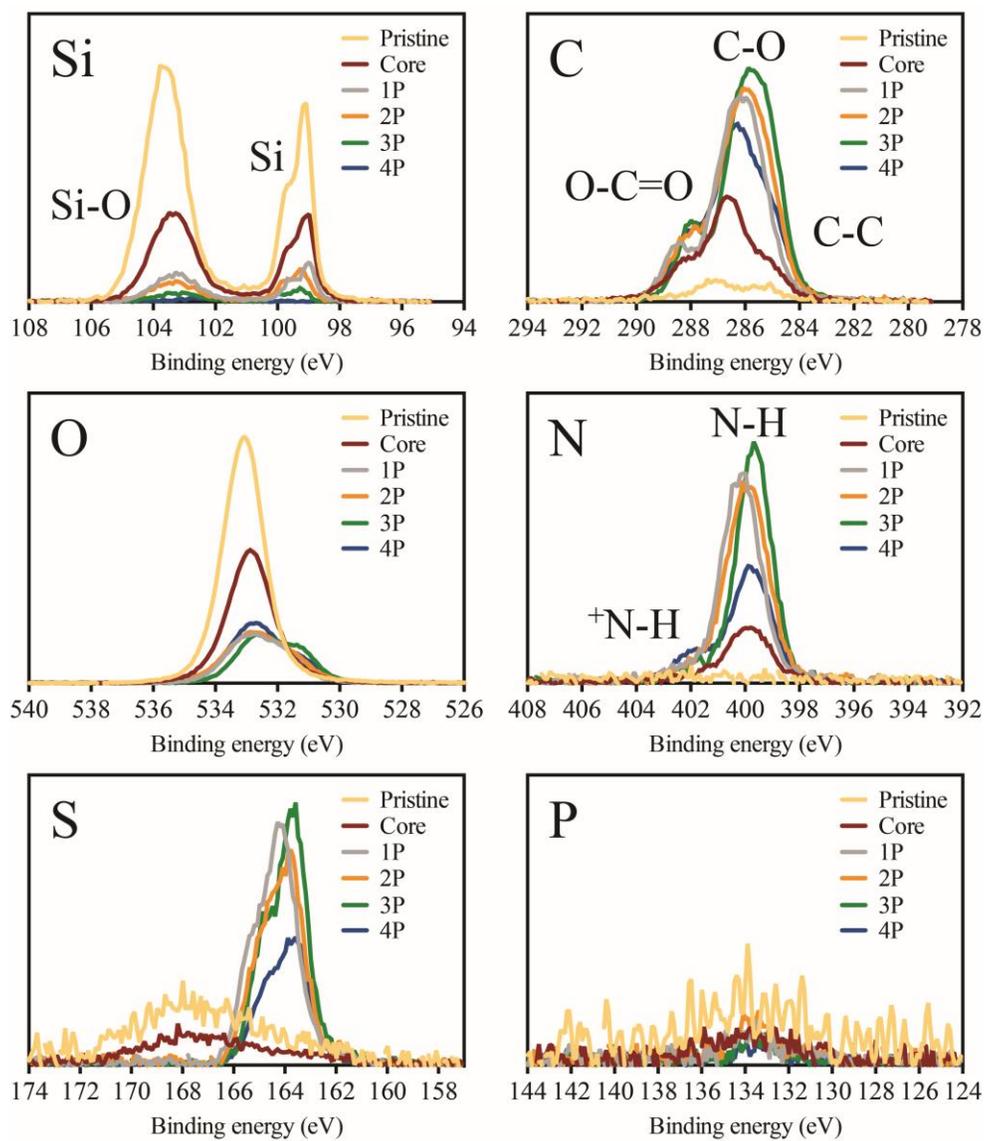
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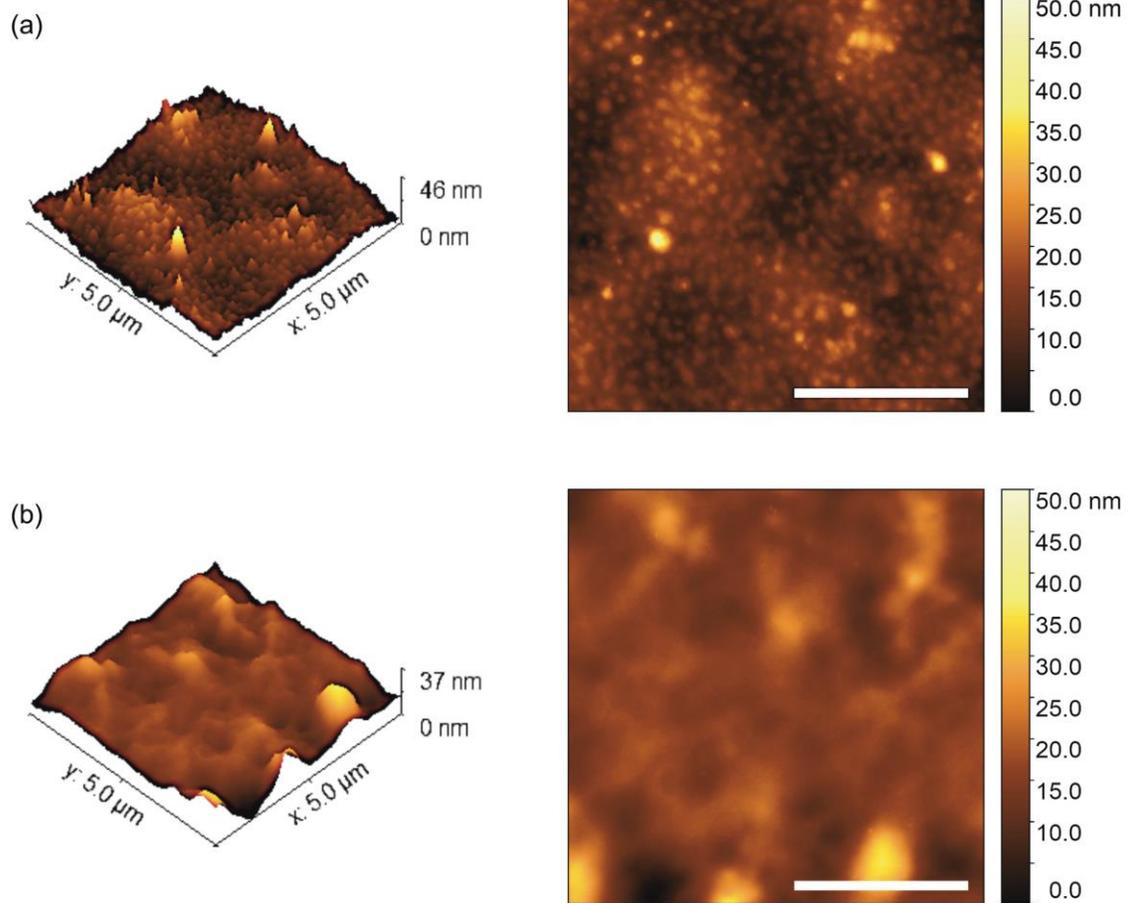
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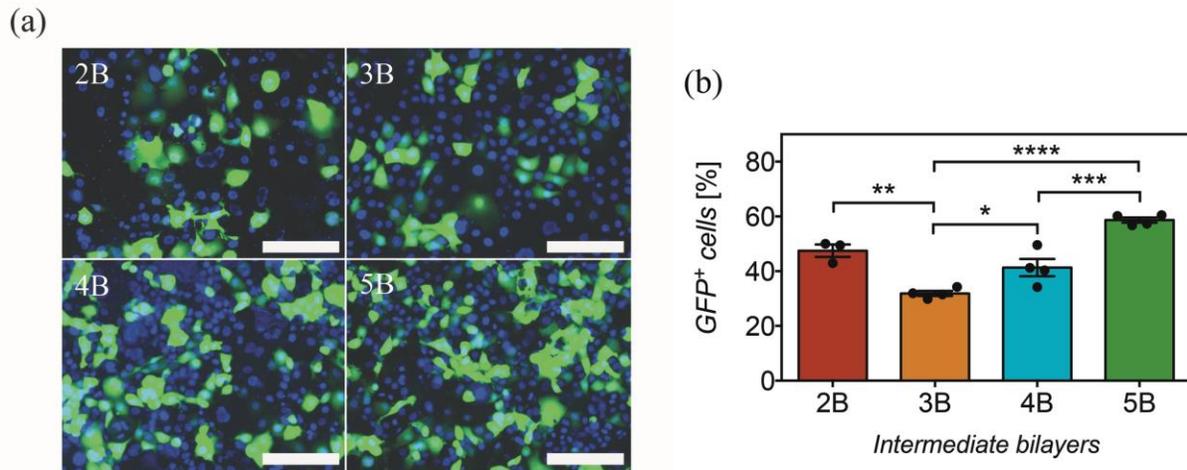
**Supplementary figure 1.** Confocal microscopy representative image of silicon nanoneedles coated with hyaluronate-chitosan-pDNA polyplex nanofilms. The nanofilm was stained with TAMRA in red **(a)** and pDNA-polyplexes were stained with DAPI **(b)**. Merged image shown in **(c)**. Scale bars represent 20 μm.



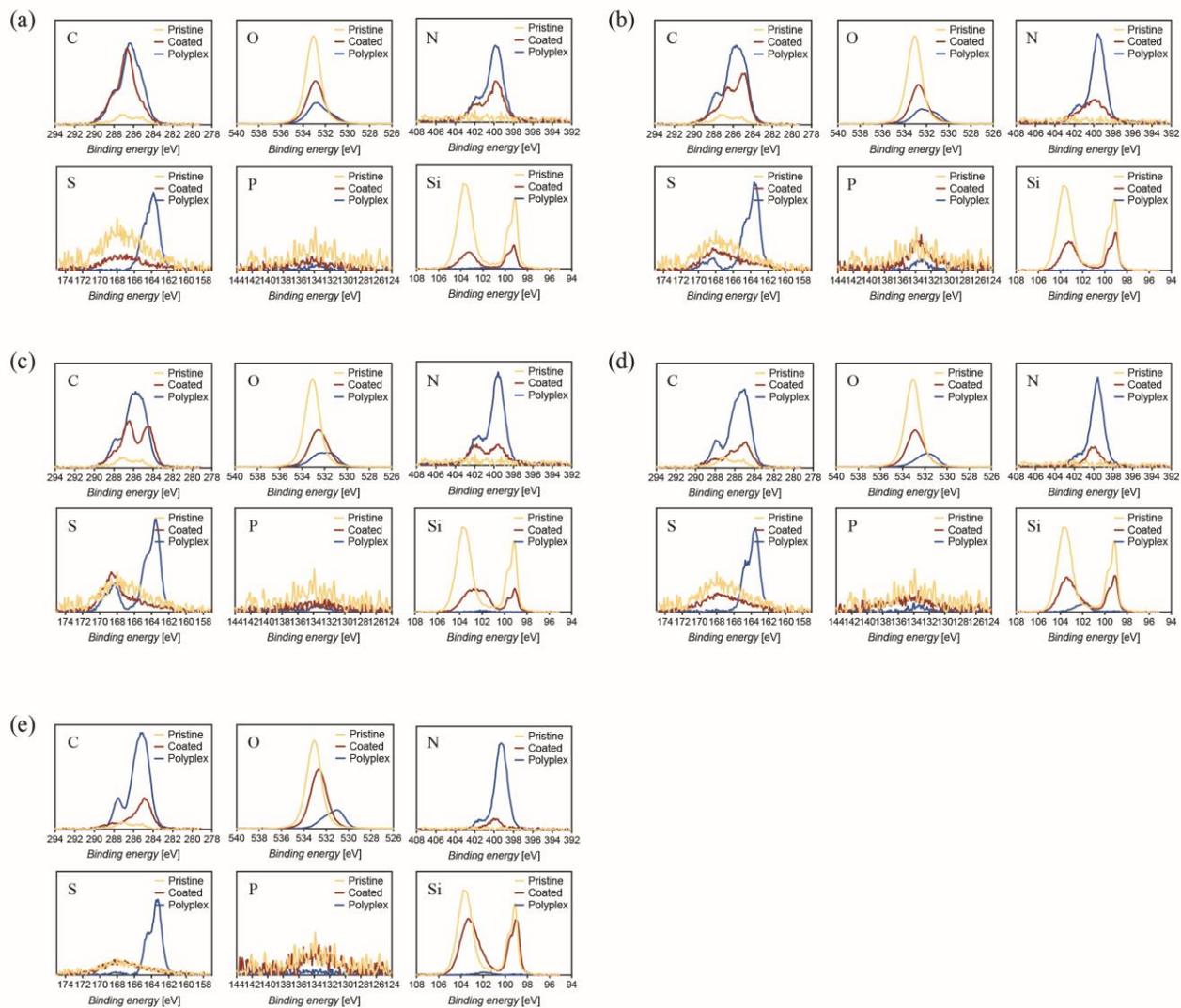
**Supplementary figure 2.** Carbon, oxygen, nitrogen, sulfur, phosphorus and silicon spectra obtained via XPS from pristine, coated (no polyplex) and nanoneedles coated with 1 to 4 polyplex layers (1P to 4P).



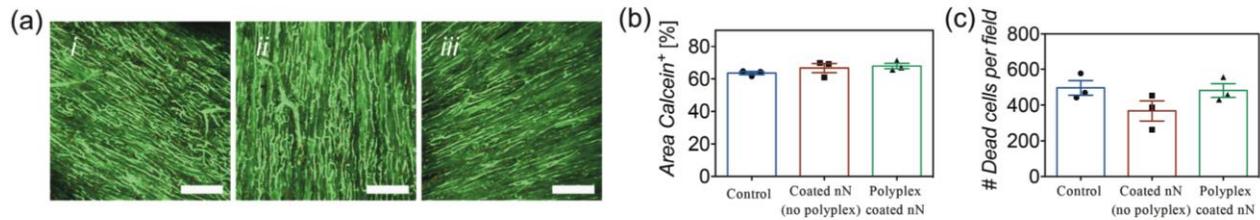
**Supplementary figure 3.** Surface topography characterization via atomic force microscopy (AFM) for flat silicon substrates coated with **(a)** 4-polyplex nanofilms and **(b)** 5-polyplex nanofilms. Scale bars represent 2  $\mu\text{m}$ .



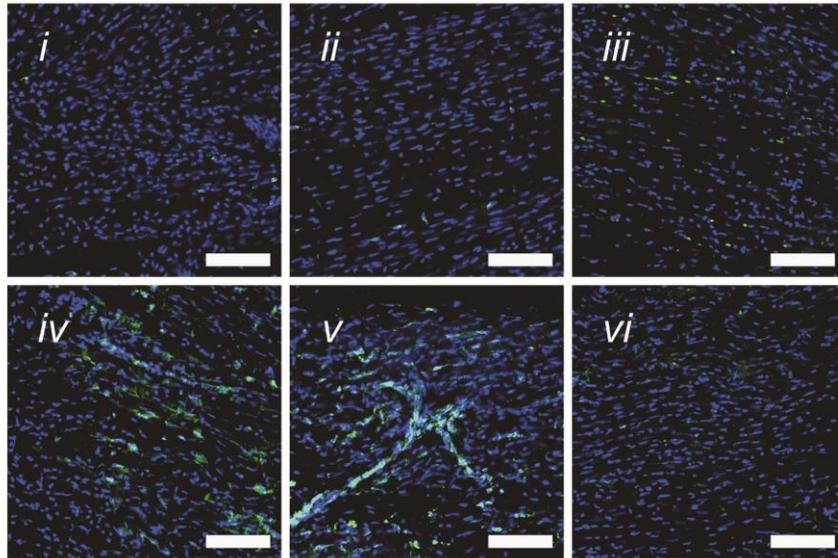
**Supplementary figure 4.** (a) Fluorescent microscopy images of COS-7 cells transfected and expressing pCAG-GFP plasmid (green) using nanofilms containing 4 polyplex layers with 2 to 5 intermediate bilayers between each polyplex layer (2B to 5B). DAPI (blue) was used as counterstaining. Scale bars represent 100  $\mu\text{m}$ . (b) Quantification of the percentage of GFP<sup>+</sup> cells and via image analysis of COS-7 cells transfected with coated nanoneedles containing 2 to 5 intermediate bilayers (2B to 5B) after 24 hours of incubation. Bars represent the mean  $\pm$  SEM (N=3). Statistical significance as (\*)  $p < 0.05$ , (\*\*)  $p < 0.01$  (\*\*\*),  $p < 0.001$  and (\*\*\*\*)  $p < 0.0001$ , using one-way ANOVA and Sidak's test.



**Supplementary figure 5.** Carbon, oxygen, nitrogen, sulfur, phosphorus and silicon spectra obtained via XPS from nanoneedles coated with 4 polyplex layers and chitosan - alginate **(a)**, chitosan - chondroitin sulfate **(b)**, chitosan - heparin **(c)**, chitosan - poly(glutamic) acid **(d)** and chitosan -  $\gamma$  poly(glutamic) acid **(e)**.



**Supplementary figure 6.** (a) Confocal microscopy images showing the viability of cardiac slices 24 hours post-transfection using calcein-AM for live cells (green) and ethidium homodimer for dead cells (red). Representative images correspond to control no chip (*i*), coated (no polyplex nanoneedles (*ii*) and polyplex-coated nanoneedles (*iii*). Polyplexes contained pDNA coding for luciferase, to eliminate green fluorescence. Bars represent 200  $\mu\text{m}$ . (b) Quantification via image analyses of calcein expression in the slice as area percentage per field and (c) number of dead cells per field for control no chip, coated (no polyplex nanoneedles and polyplex-coated nanoneedles. Bars represent the mean  $\pm$  SEM of three samples (N=3), from an average of three separate images. Differences were not statistically significant, using one-way ANOVA ( $p < 0.05$ ).



**Supplementary figure 7.** Confocal microscopy images of cardiac slices at 24 hours of transfection and immunolabelled with anti-GFP (green) and DAPI (blue). Representative images correspond to control no chip (*i*), coated (no polyplex) nanoneedles (*ii*), polyplex-coated flat substrates (*iii*) polyplex-coated nanoneedles (*iv*, *v*) and the non-interfacing side of a slice with polyplex-coated nanoneedles (*vi*). Bars represent 100  $\mu\text{m}$ .