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### **Supplemental Information**

### Depression of Accumbal to Lateral

#### Hypothalamic Synapses Gates Overeating

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**Supplementary Figure 1** 

## Figure S1. Related to Figure 1.FSK induced i-LTP is absent 7 days after an acute food restriction episode.

(a) Schematic representing the maximum and minimum infection with the AAV-DIO-ChR2 from animal that were kept in the study.

(**b**) Body weight from mice tested 7 days after a single 24h food restriction episode. Mice returned to their baseline weight.

(c) IPSC expressed as a percent of baseline (BL). Bath application of forskolin (FSK) was unable to potentiate the inhibitory transmission.

(d) PPR ratio was not changed by FSK.

(e) The coefficient of variation was not affected by FSK



## Figure S2. Related to Figure 2. Quantification of NeuN expression in mice injected with CTB in the LH.

(a) To control for CTB-induced lesion in the LH as a limitation of our strategy, we quantified the expression of NeuN in control or CTB-injected mice.

(b) NeuN expression density was similar in the LH of control and CTB-injected mice: t(19) = 0.8260, p=0.4190.

(c) 82.8% of CTB positive colocalized with NeuN, further demonstrating that CTB did induce a lesion in 10 days after the injection.

Plots show mean ± SEM.



# Figure S3. Related to Figure 2. Dual retrograde tracing from anterior and posterior lateral hypothalamus in Drd1a-tdTomato mice

(a) Schematic of experiment (left), with example images of Cholera toxin subunit-B (CTB) injection sites in anterior and posterior lateral hypothalamus (aLH/pLH) and CTB-labeled cell bodies visible in the nucleus accumbens. NAcC: accumbens core, NAcSh: accumbens shell, LS: lateral septum. All images are from the same animal. Scale bars: left & center: 500µm; Right, 200µm.

(**b**) From the animal in (A), confocal images from anterior (left) and posterior (right) NAcSh showing colocalization of CTB with tdTomato (i.e. D1R-MSNs). Arrowheads indicate non-colocalizing neurons located in posterior NAcSh. Scale bars: 50µm.

(c) Schematic indicating imaging sites across anterior/posterior NAcSh (left), with proportion of all CTB cells identified in NAcSh (n=3219) as aLH (green), pLH (light blue) or dual projecting (dark blue; center) and density distribution of each CTB type across NAcSh (right; grey points show group mean total CTB densities  $\pm$  SEM).

(d) Proportion of all aLH or pLH projecting MSNs that colocalize with tdTomato (i.e. D1R-MSNs; left). Significantly fewer MSNs projecting to aLH vs. pLH were D1R+ ( $80.9 \pm 3.51\%$  vs.  $89.5 \pm 0.7\%$ ; paired t-test, \*p < 0.05). Distribution of CTB/tdTomato overlap across anterior/posterior NAcSh (right). Significantly fewer MSNs located in the most posterior quarter of NAcSh and projecting to either aLH or pLH were D1R+ (area 1 vs. 4, Bonferroni corrected t-test, \*p < 0.016).

Plots shows means ± SEM.



## Figure S4. Related to Figure 6. Ex vivo 100Hz high frequency optogenetic stimulation at D1-MSN to LH synapse.

(a) IPSC example recorded from a cell in the LH in food restricted mice. Stimulation occurred at 10 min.

(b) IPSC example recorded from a cell in the LH in food restricted mice. In this preparation, the D1 agonist SKF38393 was present in the bath during the entire recording.

(c) IPSC expressed as a percentage of baseline for all recorded cell from control, AFR and AFR + SKF38393 groups.

(d) Grouped IPSC. The IPSC was significantly increased in AFR mice when SKF38393 was bath applied. ANOVA: F(2,16) = 4.073, p<0.05. Post-hoc Bonferonni t-test: \* p<0.05.

#### SUPPLEMENTARY TABLES

Experiment & Figure	Mouse line	Virus / Tracer	Injection site	n
CTB in LH and VP Fig. 2	Drd1a-tdTomato	CTB-Alexa488, CTB-Alexa567	LH, VP	5
CTB in LH and VTA Fig. 2	Drd1a-tdTomato	CTB-Alexa488, CTB-Alexa567	LH, VTA	6
Rabies tracing Fig.3	Drd1a-tdTomato X VGluT2Cre	AAV5-Flex-TVA-mCherry AAV8-Flex-RG SAD∆G-EFG(EnvA)	LH	2
aLH and pLH Fig. S3	Drd1a-tdTomato	CTB-Alexa488, CTB-Alexa567	aLH, pLH	10

### Table S1. Related to Figure 2 and Figure 3. Details of mice used in tracing experiments.

Experiment & Figure	Mouse line	Virus / Tracer	Injection site	n
HFS, FSK and Control cells Fig. 1	D1Cre	AAV5-EF1A-DIO-ChR2(H134R)- eYFP	NAcSh	22
FSK in the VP and Control cells Fig. 1	D1Cre	AAV5-EF1A-DIO-ChR2(H134R)- eYFP	NAcSh	5
FSK and Control cells Fig. 3	VGATCre	AAV5-hsyn-ChR2(H134R)-eYFP AAV5-DIO-EF1A-mCherry	NAc (Chr2) LH (mCherry)	6
LH VGlut2 connectivity Fig. 3	VGluT2Cre	AAV5-hsyn-ChR2(H134R)-eYFP AAV5-DIO-EF1A-tdTomato	NAcSh (ChR2) LH (TdTomato)	5
FSK and Control cells Fig. 3	VGluT2Cre	AAV5-hsyn-ChR2(H134R)-eYFP AAV5-DIO-EF1A-mCherry	NAc (Chr2) LH (mCherry)	6
Acute food restriction and FSK Fig. 1	D1Cre	AAV5-EF1A-DIO-ChR2(H134R)- eYFP	NAcSh	12
7 days after acute food restriction Fig. S1	D1Cre	AAV5-EF1A-DIO-ChR2(H134R)- eYFP	NAcSh	14
Acute food restriction and FSK in LH GABA cells Fig. 3	VGATCre	AAV5-hsyn-ChR2(H134R)-eYFP AAV5-DIO-EF1A-mCherry	NAc (Chr2) LH (mCherry)	12
Acute food restriction and FSK in LH glutamate cells Fig. 3	VGluT2Cre	AAV5-hsyn-ChR2(H134R)-eYFP AAV5-DIO-EF1A-tdTomato	NAcSh (ChR2) LH (TdTomato)	10
WIN55212-2 and Control cells in ad libitum fed mice Fig. 4	D1Cre	AAV5-EF1A-DIO-ChR2(H134R)- eYFP	NAcSh	16
Acute food restriction and CB1R antagonist on slice Fig. 4	D1cre	AAV5-EF1A-DIO-ChR2(H134R)- eYFP	NAcSh	15

Acute food restriction and CB1R antagonist Fig. 5	D1Cre	AAV5-EF1A-DIO-ChR2(H134R)- eYFP	NAcSh	32
High fat exposure Fig. 1 and 5	D1Cre	AAV5-EF1A-DIO-ChR2(H134R)- eYFP	NAcSh	16

Table S2. Related to Figure 1, Figure 3, Figure 4 and Figure S1. Details of mice used for whole-cell patch clamp recordings.

Experiment & Figure	Mouse line	n
Acute food restriction and systemic CB1R antagonist Fig. 5	C57BL/6J	14
High fat exposure and CB1R antagonist Fig. 5	C57BL/6J	15
Acute food restriction and HFS in vivo Fig. 6	D1cre	7
Local infusion of CB1R agonists and antagonist Fig. 5	C57BL/6J	12

Table S3. Related to Figure 5 and Figure 6. Details for the mice used for behavioural experiments.