# The temperature-regulation of *Pseudomonas aeruginosa cmaX-cfrX-cmpX* operon reveals an intriguing molecular network involving the sigma factors AlgU and SigX

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## SUPPLEMENTARY DATA

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			Genetic background		
Target	Treatmont	<b>U103</b>		AgiaV	
genes	Treatment	11105	ΔaigU	ΔsigA	
cmaX	HS	5.9 ± 1	$0.9 \pm 0.2$		
	CS	1 ± 0.4			
cfrX	HS	$1.7 \pm 0.3$	$1.3 \pm 0.4$		
•	CS	$3.6 \pm 0.08$		1.3 ± 0.1	
стрХ	HS	2.1 ± 0.4	$1.3 \pm 0.4$		
- 1	CS	3.1 ± 0.3		1. ± 0.3	
alaU	HS	2.9 ± 1.7			
- 3 -	CS	$1.4 \pm 0.7$			
alqD	HS	$19.4 \pm 6.3$	6 ± 0.5		
<u>9</u> _	CS	$1.5 \pm 0.3$			
rроН	HS	4 ± 1.2	$1.4 \pm 0.3$		
	CS	$1.3 \pm 0.4$			
oprF	HS	1.6 ± 0.1	$0.8 \pm 0.2$		
•	CS	1 ± 0.1			
amrZ	HS	0.4 ± 0.1	$0.3 \pm 0.03$		
	CS	1± 0.3			
sigX	HS	0.8 ± 0.1	0.6 ± 0.1		
C C	CS	2 ± 0.6		$0.9 \pm 0.2$	
accA	HS	1 ± 0.1			
	CS	1.8 ± 0.2		1.4 ± 0.1	
accB	HS				
	CS	$2.7 \pm 0.3$		1.9 ± 0.8	
PA5174	HS				
	CS	$2.5 \pm 0.8$		$1.2 \pm 0.5$	
psrA	HS				
	CS	$0.5 \pm 0.1$		$0.2 \pm 0.02$	

Table S1: Normalized expression of the studied genes under HS or CS in the indicated genetic backgrounds relative to *P. aeruginosa* H103 at 37°C.

Table S2: Characteristics of proteins identified by Maldi-TOF analysis.

Spot number	RefSeq	PA number	Protein name	Theorical MW(kDa)/pl	method	Protein Score	Number of Matched peptides	Sequence coverage (%)
1	NP_251641.1	PA2951	EtfA	31.4/4.75	MS/MS	54	1	5
2	NP_259243.1	PA0552	Pgk	40.4/5.06	MS/MS	123	2	5
3	NP_251641.1	PA2953	EtfD	59.9/5.6	MS/MS	42	1	1
4	NP_253858.1	PA5171	ArcA	46.4/5.5	MS	167	12	32
5	NP_249556.1	PA0865	Hpd	39.9/4.89	MS	133	12	29
6	NP_252329.1	PA3639	AccA	34.9/5.15	MS	84	7	26
7	NP_252345	PA3655	Tsf	30.7/4.95	MS	135	9	25
8	NP_252955	PA4265	TufA	43.3/5.03	MS	133	9	29
9	NP_253428.1	PA4740	Pnp	75.4/4.81	MS	112	9	17

### Table S3: Bacterial strains and plasmids used in this study.

Strains/plasmids	Relevant characteristic(s)	Sources
Strains		
E. coli		
JM109	Cloning host	Promega
S17.1	Conjugation strain	Simon et al., 1983
Pseudomonas aeruginosa		
H103	PAO1 derivative	Hancock, R. E. & Carey, 1979
mPAO1	PAO1 derivative	Jacobs, M. A. et al., 2003
<i>∆sigX</i> (PAOSX)	PAO1 H103 <i>∆sigX</i>	Bouffartigues et al., 2012
∆rpoH	Transposon mutant <i>rpoH</i> ::IS; <i>phoA</i> /hah-Tc <sup>r</sup>	Jacobs, M. A. et al., 2003
∆algU	PAO1 H103 ⊿algU	This study
Plasmids		
pAB133	Vector for transcriptional fusion analyzes. Gmr	Bazire et al., 2005
pAB <i>cmaX</i> L	pAB-P <i>cmaX-luxCDABE</i> . Gm <sup>r</sup>	This study
pAB <i>cfrX</i> L	pAB-P <i>cfrX-luxCDABE</i> . Gm <sup>r</sup>	This study
pAB <i>cmpX</i> L	pAB-P <i>cmpX-luxCDABE</i> . Gm <sup>r</sup>	This study
pEXUGL	pEXalgU with <i>lox-aacC1-lox</i> cassette. Apr, Gmr	Bazire et al., 2010
pCM157	<i>cre</i> expression vector, Tc <sup>r</sup>	Quenée et al., 2005

Cb<sup>r</sup>, carbenicillin resistance ; Gm<sup>r</sup>, gentamycin resistance ; Tc<sup>r</sup>, tetracycline resistance.

#### **References for Table S3**

Bouffartigues, E. *et al.* Transcription of the oprF gene of Pseudomonas aeruginosa is dependent mainly on the SigX sigma factor and is sucrose induced. *J. Bacteriol.* **194**, 4301–4311 (2012).

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Bazire, A. *et al.* The sigma factor AlgU plays a key role in formation of robust biofilms by nonmucoid Pseudomonas aeruginosa. *J. Bacteriol.* **192**, 3001–3010 (2010).

Simon, R., Priefer, U. & Pühler, A. A Broad Host Range Mobilization System for In Vivo Genetic Engineering: Transposon Mutagenesis in Gram Negative Bacteria. *Nat. Biotechnol.* **1**, 784–791 (1983).

PΔ	Gene		
number	name	Primer name	Sequence (5'- 3')
	inamo		
RT-qPCR			
PA1773	cmaX	PA1773	AAAGGGCCGAAGACAGCAT
		PA1773	CGCGATCGGTCAGGTAGTG
PA1774	ofr¥	PA1774	CTGCGGGACCTCGTCAAG
	0///	PA1774	GCCGACCTGGCGATTG
PA1775	omnY	FPA1775	GGCAGATCATTGCAGGAATCTAC
	Стрх	RPA1775	TCTCTTCAATAGTGCCTTCAACGT
PA1600	fahB	F1609	CCTCGGCAATGACAAAGACA
1 A1009	TADD	R1609	GTAGGACGGATTGAAGCGAATG
DA0760	olal I	FalgU	TACCTGGCTGTATCGGATCG
FA0702	aigu	RalgU	GAAGAACTCCGCATCCTCTG
	alaD	FPA3540	GGGCTATGTCGGTGCAGTATG
PA3540	aigD	RPA3540	GCGACTTGCCCTGGTTGAT
	rooll	FrpoH	CGCAAACCTGGAAGCCTA
PA	τροπ	RrpoH	GAACAACGAAGCGCAGGT
	oprF	FPA1777	GCGTACAGCTGGACATGAAG
PAI///		RPA1777	TTCATGAAGTCAGCCAGGTTCTT
DAGGOE	amrZ	FamrZ	CGTGAGCAGATCGCAGAA
PA3385		RamrZ	GCTCGTGCAGGCTGAGTT
DA1770	sigX	FPA1776	AATTGATGCGGCGTTACCA
PA1776		RPA1776	CCAGGTAGCGGGCACAGA
<b>DA</b> 0000	accA	FaccA	TCTTCGGCAATCTGACCAGTT
PA3639		RaccA	GTAGCCGATGTAGTCGAGGGTA
DA 40.47	accB	FaccB	AAGCCATGAAGATGATGAACC
PA4847		RaccB	CGTTCTCCACCAGGATCGA
	(-1-)(	FPA5174	AGGGCGACCTGGAGATCAT
PA5174	fabY	RPA5174	GCGCGTCCTTCTTGTATACCA
PA3006		FpsrA	CGATTTCGGCGTGAACACTT
	psrA	RpsrA	CGAAGAACGGCACCATCAG
PA0668.1		F16SRNA	AACCTGGGAACTGCATCCAA
	IbSrKNA	R16SRNA	CTTCGCCACTGGTGTTCCTT
RACE-PCR			
		ASP1cmpX	CCTTCAACGTCACCCACTTT
PA1775	стрХ	ASP2cmpX	AACACCTTGGGCAGGTACAG

**Table S4:** Primer sequences of the indicated genes used for quantitative RT-qPCR reactions, transcription start sites identification and transcriptional fusions construction.

PA1774 <i>cfrX</i>	ofrV	AS1cfrX	GCAGAAGATGCGTTTTTCG	
	ASP2cfrX	GCGGAAATACGGGAAACC		
PA1773 cmaX	omaV	ASP1cmaX	CAGGTAGTGCGCGAGGTAGT	
	UndA	ASP2cmaX	CGGATTCAGGTTCACTCCAC	
Transcriptional fusions				
**PA1773	**000	FCmaX	*taataagagctcGTTCGTGTATGCCGACAACA	
	CITIAX	RCmaX	*taataactagttGCGCTTCCCTTGGTAAAAC	
**D <b>\1</b> 77 <i>1</i>	**ofrV	FCFRX	*taataagagctcGAGTTGATACGGGAGCGCAT	
FA1774	UIX	RCFRX	*taataactagttCATGTCTGTACGAGGGAGTG	
**PA1775	**cmnY	FCmpX	*taataagagctcCTGCGGGACCTGCTCAAG	
	ыпрл	RCmpX	*taataactagttCACTCAGCCTTGTCAGC	

\* Nucleotides not in the chromosomal sequence are indicated in lower case.

\*\* Gene localized downstream the PCR-amplified DNA fragment





Fig. S1: Flow cytometry analysis of *P. aeruginosa* H103 viability exposed or not to CS or HS.



Fig. S2: Effect of HS and CS on *algU* expression and activity.



Fig. S3: RpoH is not involved in the *cmaX, cfrX and cmpX* regulation in response to HS in PAO1.



Fig. S4: 2-DE total protein profiles of *P. aeruginosa* exposed to CS from 37°C to 4°C or not (control).



Fig. S5: c-di-GMP level quantification by LC-MS/MS in H103 strain exposed (square) or not to a CS (point).

#### **References for Supplementary Fig. S5**

Strehmel, J. *et al.* Sensor kinase PA4398 modulates swarming motility and biofilm formation in Pseudomonas aeruginosa PA14. *Appl Environ Microbiol.* 81(4):1274-85. (2015).

Bouffartigues, E. *et al.* The absence of the *Pseudomonas aeruginosa* OprF protein leads to increased biofilm formation through variation in c-di-GMP level. *Front Microbiol.*; 6: 630. (2015).



Fig. S6: Effect of a sublethal dose of valinomycin on membrane fluidity measured by fluorescence polarization in stationary growth phase.