## nature research

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## **Reporting Summary**

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our Editorial Policies and the Editorial Policy Checklist.

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For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	$\mathbf{x}$ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	🕱 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
x	The statistical test(s) used AND whether they are one- or two-sided  Only common tests should be described solely by name; describe more complex techniques in the Methods section.
×	A description of all covariates tested
x	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
x	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
x	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
×	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
x	$\square$ Estimates of effect sizes (e.g. Cohen's $d$ , Pearson's $r$ ), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.
So:	ftware and code

Policy information about <u>availability of computer code</u>

Data collection not applicable

Data analysis

Interferometry data were processed with the Octet Data Analysis software v10.0 (FortéBio). The MST data were analyzed using the MO. Affinity Analysis software (NanoTemper Technologies). The neutralization data were fitted using Prism 8 (GraphPad).

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

## Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The scattering data and models of Sy23, RBD and their complex are deposited into SASBDB (www.sasbdb.org), entries SASDJF4, SASDJG4 and SASDJH4 respectively. The cryo-EM density maps of SARS-CoV-2 spike glycoprotein with Sb23 bound were deposited in the Electron Microscopy Data Bank (EMDB) with accession codes EMD-11616 (1-up) and EMD-11617 (2-up). The corresponding models were deposited in the Protein Data Bank (PDB) with accession codes 7A25 (1-up) and 7A29 (2up). The sequences of all the selected sybodies from this study are provided in Supplementary Data 1. The MST, BLI and neutralization data are available under https://github.com/tania-custodio/Sb23 and in the Source File. Any other experimental data that support the findings of this study are available from the corresponding authors upon request.

Field-spe	ecific re	porting			
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🗶 Life sciences	★ Life sciences				
For a reference copy of t	the document with	all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>			
Life scier	nces stu	udy design			
All studies must dis	All studies must disclose on these points even when the disclosure is negative.				
Sample size	Sample size is n	not relevant for this study, no calculations were made.			
Data exclusions	No data was ex	xcluded from any analysis			
Replication	Replicate exper	eplicate experiments are indicated in the respective legends			
Randomization	This study repo	study reports the biophysical and structural characterization of protein complex, and randomization is not relevant.			
Blinding	This study repo	rts the biophysical and structural characterization of protein complex, and blinding is not relevant.			
Reporting for specific materials, systems and methods  We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.  Materials & experimental systems    Methods					
Antibodies used	Mouse	e monoclonal anti myc, Sigma #M4439 ELISA. Dilution of 1:2000			
Validation The antibody was validated by ELISA		tibody was validated by ELISA			
Eukaryotic cell lines					
Policy information about <u>cell lines</u>					
Cell line source(s)		FreeStyle ™ 293-F purchase from Invitrogen. This cells are derived from HEK293-F cells.			
Authentication		No authentication of the FreeStyle ™ 293-F cell line was done.			
Mycoplasma contamination		We confirmed that all cell lines tested negative for mycoplasma contamination.			
Commonly misidentified lines (See ICLAC register)		No commonly misidentified cell lines were used.			