

Beyond the Nanomaterials Approach: Influence of culture conditions on the stability and antimicrobial activity of silver nanoparticles

Supplementary material

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Figure S1. Culture media alter the UV-Vis spectra of the AgNPs.

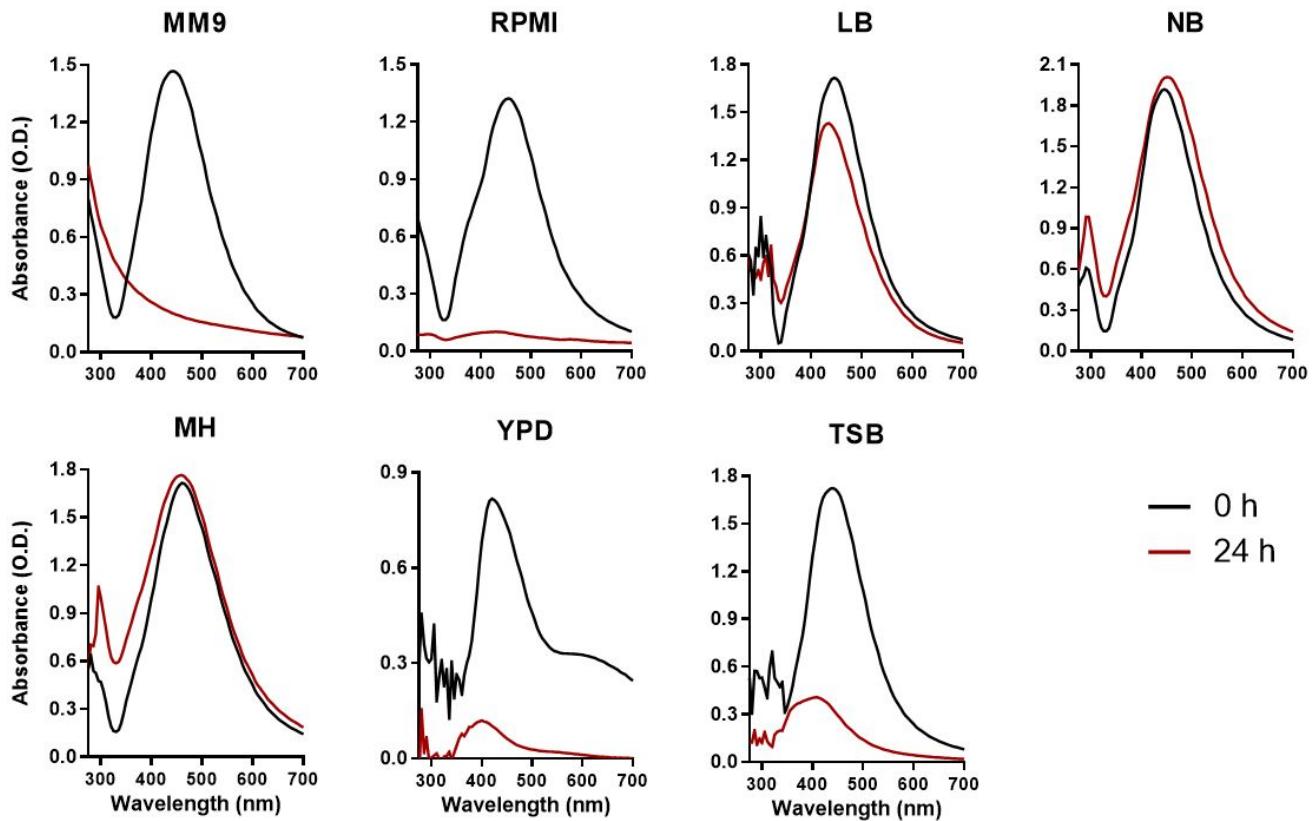


Figure S1. Culture media alter the UV-Vis spectra of the AgNPs. UV-Vis spectra profile of AgNPs when exposed to different culture media, at $t = 0$ h and $t = 24$ h.

Figure S2. UV-Vis profile from the centrifuged AgNPs supernatant

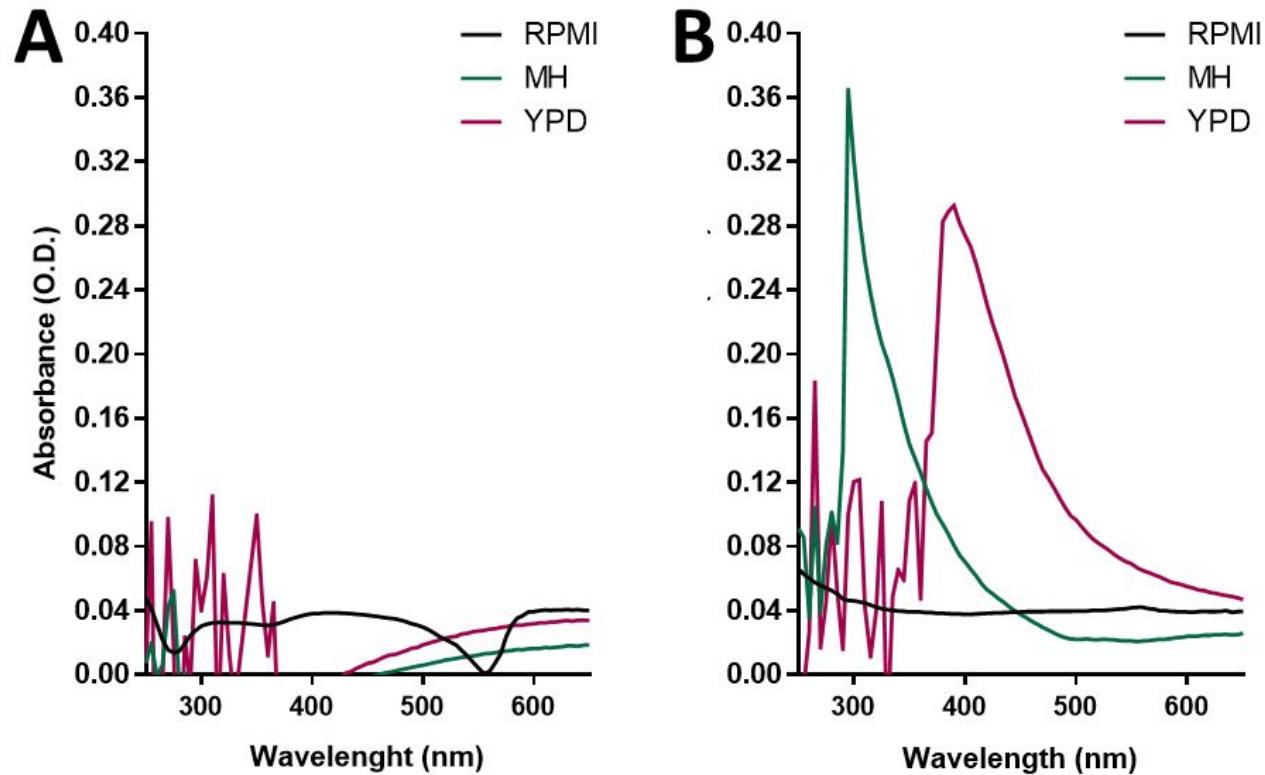


Figure S2. UV-Vis profile from the centrifuged AgNPs supernatant. The supernatant of the centrifuged AgNPs in RPMI, MH, and YPD showed different profiles at $t = 0$ h (A) and $t = 24$ h (B). In all conditions, the absorbance profile reveals the potential presence of non-AgNPs silver species $\lambda < 300$ nm. Only in YPD $t = 24$ h, a surface plasmon typical form the AgNPs was observed.

Figure S3. Transmission electron micrographs of AgNPs used in this study.

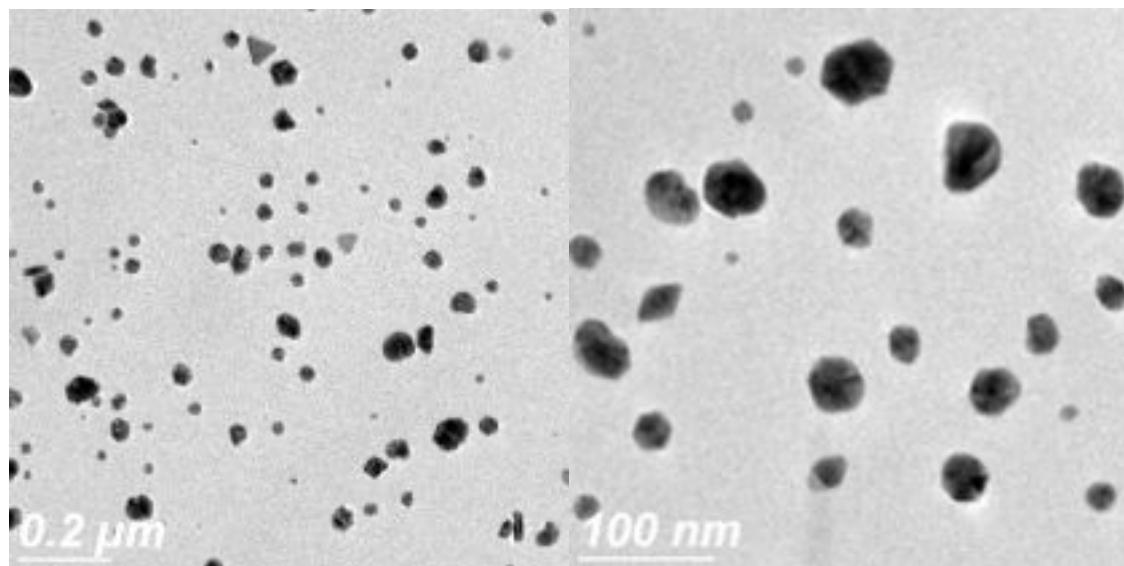


Figure S3. TEM micrographs of silver nanoparticles showed an aspect ratio close to 1, with a metallic core diameter of 35 ± 15 nm.

Figure S4. Size distribution of AgNPs in different culture media.

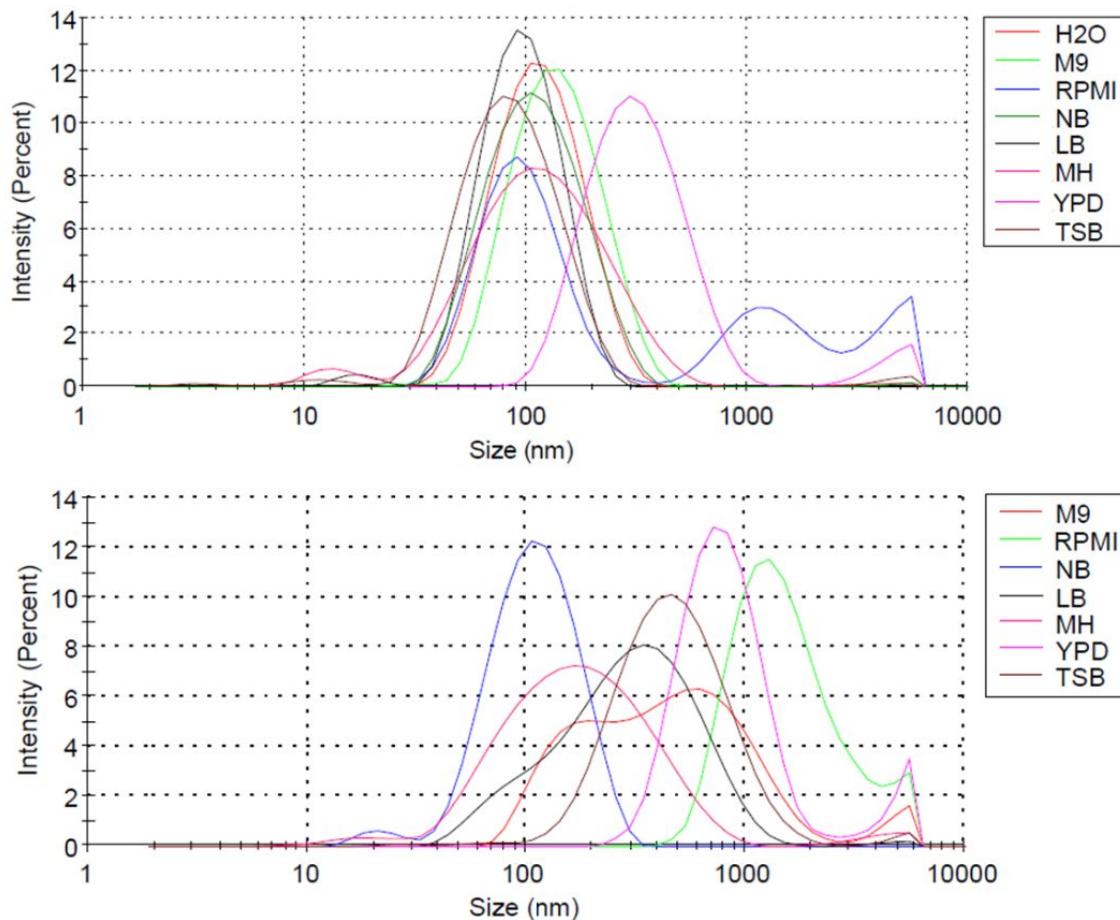


Figure S4. Hydrodynamic Size distribution graphs of AgNPs suspended in different culture media, at t = 0 h (top) and t = 24 h (bottom). The graphs reveal that AgNPs hydrodynamic size changes over time in different culture media.

Table S1. DLS analysis.**Table S1.** DLS characterization of AgNPs in different culture media

Culture media	Hydrodynamic size (nm)		Polydispersity Index		ζ -potential (mV)	
	t= 0 h	t= 24 h	t= 0 h	t= 24 h	t= 0 h	t= 24 h
Milli-Q H ₂ O	95.3	92.2	0.247	0.296	-14.1	-20.5
M9	124.5	332.4	0.196	0.443	-16.5	-17.9
RPMI	136.2	784.2	0.621	0.327	-16.9	-16
NB	87.8	215.1	0.266	0.222	-14.3	-10.8
LB	77.7	90.1	0.229	0.404	-10	-16.1
MH	89.8	113.6	0.369	0.417	-8.16	-24.4
YPD	299.6	314	0.293	0.384	-16.4	-18.1
TSB	67.4	373.5	0.291	0.315	-14.8	-15.6

Table S2. Culture media broth major components.**Table S2. Culture media broth major components**

Culture media	Grams per liter of solution (g/L)				pH
	Carbon source	Amino acids/ proteins	Inorganics Salts	Others	
M9	4	0	11.67	Biotin .001 Trace elements Thiamine .001	7.5
RPMI *	2	0.99	9.35	Vitamins 0.04 Glutathione 0.001 Phenol Red 0.0053	7.2
NB	0 #	8	0 #	0 #	6.8
LB	0 #	15	5 #	0 #	7.5
MH	1.5 #	19.5	0 #	0 #	7.4
YPD	20	30	0 #	0 #	6.5
TSB	2.5 #	20	7.5 #	0 #	7.3

#= It can be inherently added by the other components, due to the non-defined composition nature of the culture media's formulations.

* RPMI supplemented with L-Glu+, and NaCOOH-

** Thiamin

Table S3. Silver content estimated by ICP-OES

Table S3. The silver content varies over time in the different culture media

Culture media	Silver content (ppm)	
	t = 0 h	t = 24 h
RPMI	0.34	0.17
MH	1.16	0.89
YPD	1.41	1.70

Table S4. AgNPs MIC on *E. coli*, on different culture media**Table S4. AgNPs MIC on *E. coli*, on different culture media, as determined by our study and the meta-analysis of the literature.**

Culture media	Our Data	MIC ranges ($\mu\text{g ml}^{-1}$) [#]		References
		All culture conditions**	Similar conditions***	
M9	N/A*	0.11 – 0.21	0.11 – 0.21	1,2
RPMI 1640	0.5	1.25	1.25	3
LB	1	1 – 40	1 – 40	2,3,13–23
NB	2.5	0.25 - 180	1 – 8	4–12
MH	12.5	0.78 – 31.25	1.6 – 31.25	1,22,24–34
TSB	15	27 – 100	-	22,35
BHI	N/A*	0.5 - 2	1.7 – 2	36–38

We provide the MIC values published, still, most reports do not specify if those values are based on the total concentration of AgNPs or only to the silver content.

* The AgNPs MIC values for M9 and BHI were not determined.

** For “All culture conditions” we mean all the MIC values, regardless of initial inoculum size, exposition time, and temperature.

*** For “similar conditions”, we considered those similar to the CLSI A09 protocol with a wider range in the inoculum size: only MIC values, an initial inoculum of 10^5 to 10^6 cells ml^{-1} , 24 h cultures, at 37 °C.

List S1. Culture media broth formulations.

The followed standards recipes for the different culture media are described next. These were used for the preparation of culture media. The quantities of components are for a 1-liter 1X solution and were diluted in 1 distilled water; pH was adjusted using NaOH or HCL, as needed. Culture media was sterilized by autoclaving or filtration, following the seller recommendations.

<u>CULTURE MEDIA REAGENTS</u>	<u>QUANTITY</u>
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M9 media

M9 salt solution	11.52 g
• Na ₂ HPO ₄ 3.37 mM (7.52 g/L)	
• KH ₂ PO ₄ 2.20 mM (3 g/L)	
• NaCl 0.855 mM (0.5 g/L)	
• NH ₄ Cl 0.935 mM (0.5 g/L)	
Glucose	4 g
1 mM MgSO ₄	120 mg
0.3 mM CaCl ₂	33 mg/L
Biotin	1 µg
Thiamin	1 µg
Trace elements	0.1%
pH 7.5	

RPMI 1640 (R0883)

Inorganic Salts	9.75 g
• Calcium Nitrate • 4H ₂ O (0.1 g)	
• Magnesium Sulfate (anhydrous) (0.05 g)	
• Potassium Chloride (0.4 g)	
• Sodium Bicarbonate (2 g)	
• Sodium Chloride (6 g)	
• Sodium Phosphate Dibasic (anhydrous) (0.8 g)	
Amino Acids	0.69403 g
• L-Alanyl-L-Glutamine (0)	
• L-Arginine (0.2)	
• L-Asparagine (anhydrous) (0.05)	
• L-Aspartic Acid (0.02)	
• L-Cystine •2HCl (0.0652)	
• L-Glutamic Acid (0.02)	
• L-Glutamine (—)	
• Glycine (0.01)	
• L-Histidine (0.015)	
• Hydroxy-L-Proline (0.02)	
• L-Isoleucine (0.05)	
• L-Leucine (0.05)	

• L-Lysine •HCl (0.04)	
• L-Methionine (0.015)	
• L-Phenylalanine (0.015)	
• L-Proline (0.02)	
• L-Serine (0.03)	
• L-Threonine (0.02)	
• L-Tryptophan (0.005)	
• L-Tyrosine •2Na •2H ₂ O (0.02883)	
• L-Valine (0.02)	
Vitamins	0.042655
• D-Biotin 0.0002	
• Choline Chloride 0.003	
• Folic Acid 0.001	
• myo-Inositol 0.035	
• Niacinamide 0.001	
• p-Aminobenzoic Acid 0.001	
• D-Pantothenic Acid (hemicalcium) 0.00025	
• Pyridoxine • HCl 0.001	
• Riboflavin 0.0002	
• Thiamine • HCl 0.001	
• Vitamin B12 0.000005	
Other	
• D-Glucose 2	
• Glutathione (reduced) 0.001	
• Phenol Red •Na 0.0053	
• L-Glutamine 0.3	
• Sodium Bicarbonate 0	
pH 7.2 ±0.2	

Nutrient Broth (NB)

Beef Extract	3 g
Peptone	5 g
pH 6.8 ±0.2	

Luria Bertani broth (LB)

Peptone	10 g
Yeast Extract	5 g
Sodium Chloride	5 g
pH 7.5 ±0.2	

MH

Acid Digest of Casein	17.5 g
Soluble Starch	1.5 g
Beef Extract	2 g
pH 7.4 ±0.2	

YPD

Yeast extract	10 g
Peptone	20 g
Dextrose (glucose)	20 g
pH 6.5 ±0.2	

TSB

Pancreatic Digest of Casein	17 g
Sodium Chloride	5 g
Papaic Digest of Soybean Meal	3 g
Dextrose	2.5 g
Dipotassium Phosphate	2.5 g
pH 7.3 ±0.2	

References

- (1) Jadallannagari, S.; Deshmukh, K.; Ramanan, S. R.; Kowshik, M. Antimicrobial Activity of Hemocompatible Silver Doped Hydroxyapatite Nanoparticles Synthesized by Modified Sol–Gel Technique. *Appl. Nanosci.* **2014**, *4* (2), 133–141. <https://doi.org/10.1007/s13204-013-0197-x>.
- (2) Lok, C.-N.; Ho, C.-M.; Chen, R.; He, Q.-Y.; Yu, W.-Y.; Sun, H.; Tam, P. K.-H.; Chiu, J.-F.; Che, C.-M. Silver Nanoparticles: Partial Oxidation and Antibacterial Activities. *JBIC J. Biol. Inorg. Chem.* **2007**, *12* (4), 527–534. <https://doi.org/10.1007/s00775-007-0208-z>.
- (3) Peetsch, A.; Greulich, C.; Braun, D.; Stroetges, C.; Rehage, H.; Siebers, B.; Köller, M.; Epple, M. Silver-Doped Calcium Phosphate Nanoparticles: Synthesis, Characterization, and Toxic Effects toward Mammalian and Prokaryotic Cells. *Colloids Surfaces B Biointerfaces* **2013**, *102*, 724–729. <https://doi.org/10.1016/J.COLSURFB.2012.09.040>.
- (4) Tahir, K.; Ahmad, A.; Li, B.; Nazir, S.; Khan, A. U.; Nasir, T.; Khan, Z. U. H.; Naz, R.; Raza, M. Visible Light Photo Catalytic Inactivation of Bacteria and Photo Degradation of Methylene Blue with Ag/TiO₂ Nanocomposite Prepared by a Novel Method. *J. Photochem. Photobiol. B Biol.* **2016**, *162*, 189–198. <https://doi.org/10.1016/j.jphotobiol.2016.06.039>.
- (5) Radzig, M. A.; Nadtochenko, V. A.; Koksharova, O. A.; Kiwi, J.; Lipasova, V. A.; Khmel, I. A. Antibacterial Effects of Silver Nanoparticles on Gram-Negative Bacteria: Influence on the Growth and Biofilms Formation, Mechanisms of Action. *Colloids Surfaces B Biointerfaces* **2013**, *102*, 300–306. <https://doi.org/10.1016/j.colsurfb.2012.07.039>.
- (6) Liu, H.-L.; Dai, S. A.; Fu, K.-Y.; Hsu, S.-H. Antibacterial Properties of Silver Nanoparticles in Three Different Sizes and Their Nanocomposites with a New Waterborne Polyurethane. *Int. J. Nanomedicine* **2010**, *5*, 1017–1028. <https://doi.org/10.2147/IJN.S14572>.
- (7) Xia, Q.; Ma, Y.; Wang, J.; Xia, Q. H.; Ma, Y. J.; Wang, J. W. Biosynthesis of Silver Nanoparticles Using Taxus Yunnanensis Callus and Their Antibacterial Activity and Cytotoxicity in Human Cancer Cells. *Nanomaterials* **2016**, *6* (9), 160. <https://doi.org/10.3390/nano6090160>.
- (8) Ninganagouda, S.; Rathod, V.; Singh, D.; Hiremath, J.; Singh, A. K.; Mathew, J.; ul-Haq, M. Growth Kinetics and Mechanistic Action of Reactive Oxygen Species Released by Silver Nanoparticles from Aspergillus Niger on Escherichia Coli. *Biomed Res. Int.* **2014**, *2014*, 753419. <https://doi.org/10.1155/2014/753419>.
- (9) Pal, S.; Tak, Y. K.; Song, J. M. Does the Antibacterial Activity of Silver Nanoparticles Depend on the Shape of the Nanoparticle? A Study of the Gram-Negative Bacterium Escherichia Coli. *Appl. Environ. Microbiol.* **2007**, *73* (6), 1712–1720. <https://doi.org/10.1128/AEM.02218-06>.
- (10) Agnihotri, S.; Mukherji, S.; Mukherji, S. Size-Controlled Silver Nanoparticles Synthesized over the Range 5–100 Nm Using the Same Protocol and Their Antibacterial Efficacy. *RSC Adv.* **2014**, *4* (8), 3974–3983. <https://doi.org/10.1039/C3RA44507K>.
- (11) Ruparelia, J. P.; Chatterjee, A. K.; Duttagupta, S. P.; Mukherji, S. Strain Specificity in Antimicrobial Activity of Silver and Copper Nanoparticles. *Acta Biomater.* **2008**, *4* (3), 707–716. <https://doi.org/10.1016/j.actbio.2007.11.006>.
- (12) Raffi, M.; Hussain, F.; Bhatti, T. M.; Akhter, J. I.; Hameed, A.; Hasan, M. M. *Antibacterial Characterization of Silver Nanoparticles against E. Coli ATCC-15224*; 2008; Vol. 24.
- (13) Shrivastava, S.; Bera, T.; Roy, A.; Singh, G.; Ramachandrarao, P.; Dash, D. Characterization of Enhanced Antibacterial Effects of Novel Silver Nanoparticles. *Nanotechnology* **2007**, *18* (22), 225103. <https://doi.org/10.1088/0957-4484/18/22/225103>.
- (14) Fayaz, A. M.; Balaji, K.; Girilal, M.; Yadav, R.; Kalaichelvan, P. T.; Venketesan, R. Biogenic Synthesis of Silver Nanoparticles and Their Synergistic Effect with Antibiotics: A Study against Gram-Positive and Gram-Negative Bacteria. *Nanomedicine Nanotechnology, Biol. Med.* **2010**, *6* (1), 103–109. <https://doi.org/10.1016/j.nano.2009.04.006>.
- (15) Li, P.; Li, J.; Wu, C.; Wu, Q.; Li, J. Synergistic Antibacterial Effects of β-Lactam Antibiotic Combined with Silver Nanoparticles. *Nanotechnology* **2005**, *16* (9), 1912–1917. <https://doi.org/10.1088/0957-4484/16/9/082>.
- (16) Zhang, Y.; Peng, H.; Huang, W.; Zhou, Y.; Yan, D. Facile Preparation and Characterization of Highly Antimicrobial Colloid Ag or Au Nanoparticles. *J. Colloid Interface Sci.* **2008**, *325*, 371–376. <https://doi.org/10.1016/j.jcis.2008.05.063>.
- (17) Ivask, A.; Elbadawy, A.; Kaweeteerawat, C.; Boren, D.; Fischer, H.; Ji, Z.; Chang, C. H.; Liu, R.; Tolaymat, T.; Telesca, D.; Zink, J. I.; Cohen, Y.; Holden, P. A.; Godwin, H. A. Toxicity Mechanisms in Escherichia Coli Vary for Silver Nanoparticles and Differ from Ionic Silver. *ACS Nano* **2014**, *8* (1), 374–386.

- [https://doi.org/10.1021/nn4044047.](https://doi.org/10.1021/nn4044047)
- (18) Pallavicini, P.; Arciola, C. R.; Bertoglio, F.; Curtosi, S.; Dacarro, G.; D'Agostino, A.; Ferrari, F.; Merli, D.; Milanese, C.; Rossi, S.; Taglietti, A.; Tenci, M.; Visai, L. Silver Nanoparticles Synthesized and Coated with Pectin: An Ideal Compromise for Anti-Bacterial and Anti-Biofilm Action Combined with Wound-Healing Properties. *J. Colloid Interface Sci.* **2017**, *498*, 271–281. <https://doi.org/10.1016/j.jcis.2017.03.062>.
- (19) Goswami, S. R.; Sahareen, T.; Singh, M.; Kumar, S. Role of Biogenic Silver Nanoparticles in Disruption of Cell–Cell Adhesion in *Staphylococcus Aureus* and *Escherichia Coli* Biofilm. *J. Ind. Eng. Chem.* **2015**, *26*, 73–80. <https://doi.org/10.1016/J.JIEC.2014.11.017>.
- (20) Greulich, C.; Braun, D.; Peetsch, A.; Diendorf, J.; Siebers, B.; Epple, M.; Kölle, M. The Toxic Effect of Silver Ions and Silver Nanoparticles towards Bacteria and Human Cells Occurs in the Same Concentration Range. *RSC Adv.* **2012**, *2* (17), 6981. <https://doi.org/10.1039/c2ra20684f>.
- (21) Li, D.; Liu, Z.; Yuan, Y.; Liu, Y.; Niu, F. Green Synthesis of Gallic Acid-Coated Silver Nanoparticles with High Antimicrobial Activity and Low Cytotoxicity to Normal Cells. *Process Biochem.* **2015**, *50* (3), 357–366. <https://doi.org/10.1016/J.PROCBIO.2015.01.002>.
- (22) Zhang, S.; Liu, L.; Pareek, V.; Becker, T.; Liang, J.; Liu, S. Effects of Broth Composition and Light Condition on Antimicrobial Susceptibility Testing of Ionic Silver. *J. Microbiol. Methods* **2014**, *105*, 42–46. <https://doi.org/10.1016/J.MIMET.2014.07.009>.
- (23) Ansari, M. A.; Khan, H. M.; Khan, A. A.; Ahmad, M. K. Interaction of Silver Nanoparticles with *Escherichia Coli* and Their Cell Envelope Biomolecules. **2014**, 905–915. <https://doi.org/10.1002/jobm.201300457>.
- (24) Vazquez-Muñoz, R.; Borrego, B.; Juárez-Moreno, K.; García-García, M.; Mota Morales, J. D.; Bogdanchikova, N.; Huerta-Saquero, A. Toxicity of Silver Nanoparticles in Biological Systems: Does the Complexity of Biological Systems Matter? *Toxicol. Lett.* **2017**, *276*, 11–20. <https://doi.org/10.1016/j.toxlet.2017.05.007>.
- (25) Vazquez-Muñoz, R.; Meza-Villezcas, A.; Fournier, P. G. J.; Soria-Castro, E.; Juarez-Moreno, K.; Gallego-Hernández, A. L.; Bogdanchikova, N.; Vazquez-Duhalt, R.; Huerta-Saquero, A. Enhancement of Antibiotics Antimicrobial Activity Due to the AgNPs Impact on the Cell Membrane. *PLoS One* **2019**.
- (26) Nogueira, S. S.; de Araujo-Nobre, A. R.; Mafud, A. C.; Guimarães, M. A.; Alves, M. M. M.; Plácido, A.; Carvalho, F. A. A.; Arcanjo, D. D. R.; Mascarenhas, Y.; Costa, F. G.; Albuquerque, P.; Eaton, P.; de Souza de Almeida Leite, J. R.; da Silva, D. A.; Cardoso, V. S. Silver Nanoparticle Stabilized by Hydrolyzed Collagen and Natural Polymers: Synthesis, Characterization and Antibacterial-Antifungal Evaluation. *Int. J. Biol. Macromol.* **2019**, *135*, 808–814. <https://doi.org/10.1016/j.ijbiomac.2019.05.214>.
- (27) Orlowski, P.; Zmigrodzka, M.; Tomaszewska, E.; Ranoszek-Soliwoda, K.; Czupryn, M.; Antos-Bielska, M.; Szemraj, J.; Celichowski, G.; Grobelny, J.; Krzyzowska, M. Tannic Acid-Modified Silver Nanoparticles for Wound Healing: The Importance of Size. *Int. J. Nanomedicine* **2018**, *13*, 991–1007. <https://doi.org/10.2147/IJN.S154797>.
- (28) Annadhasan, M.; SankarBabu, V. R.; Naresh, R.; Umamaheswari, K.; Rajendiran, N. A Sunlight-Induced Rapid Synthesis of Silver Nanoparticles Using Sodium Salt of N-Cholyl Amino Acids and Its Antimicrobial Applications. *Colloids Surfaces B Biointerfaces* **2012**, *96*, 14–21. <https://doi.org/10.1016/J.COLSURFB.2012.03.009>.
- (29) Kvítek, L.; Panáček, A.; Soukupová, J.; Kolář, M.; Večeřová, R.; Prucek, R.; Holecová, M.; Zbořil, R. Effect of Surfactants and Polymers on Stability and Antibacterial Activity of Silver Nanoparticles (NPs). **2008**. <https://doi.org/10.1021/jp711616v>.
- (30) Panáček, A.; Kvítek, L.; Smékalová, M.; Večeřová, R.; Kolář, M.; Röderová, M.; Dyčka, F.; Šebela, M.; Prucek, R.; Tomanec, O.; Zbořil, R. Bacterial Resistance to Silver Nanoparticles and How to Overcome It. *Nat. Nanotechnol.* **2018**, *13* (1), 65–71. <https://doi.org/10.1038/s41565-017-0013-y>.
- (31) Suchomel, P.; Kvítek, L.; Panacek, A.; Prucek, R.; Hrbac, J.; Vecerova, R.; Zboril, R. Comparative Study of Antimicrobial Activity of AgBr and Ag Nanoparticles (NPs). *PLoS One* **2015**, *10* (3), 1–16. <https://doi.org/10.1371/journal.pone.0119202>.
- (32) Martinez-Castanon, G. A.; Niño-Martínez, N.; Martínez-Gutierrez, F.; Martínez-Mendoza, J. R.; Ruiz, F. Synthesis and Antibacterial Activity of Silver Nanoparticles with Different Sizes. *J. Nanoparticle Res.* **2008**, *10* (8), 1343–1348. <https://doi.org/10.1007/s11051-008-9428-6>.
- (33) Prucek, R.; Tuček, J.; Kilianová, M.; Panáček, A.; Kvítek, L.; Filip, J.; Kolář, M.; Tománková, K.; Zbořil, R. The Targeted Antibacterial and Antifungal Properties of Magnetic Nanocomposite of Iron Oxide and Silver Nanoparticles. *Biomaterials* **2011**, *32* (21), 4704–4713. <https://doi.org/10.1016/j.biomaterials.2011.03.039>.
- (34) Li, W. R.; Xie, X. B.; Shi, Q. S.; Zeng, H. Y.; Ou-Yang, Y. S.; Chen, Y. Ben. Antibacterial Activity and Mechanism of Silver Nanoparticles on *Escherichia Coli*. *Appl. Microbiol. Biotechnol.* **2010**, *85* (4), 1115–

1122. <https://doi.org/10.1007/s00253-009-2159-5>.
- (35) Kanmani, P.; Lim, S. T. Synthesis and Characterization of Pullulan-Mediated Silver Nanoparticles and Its Antimicrobial Activities. *Carbohydr. Polym.* **2013**, *97* (2), 421–428. <https://doi.org/10.1016/j.carbpol.2013.04.048>.
- (36) Paredes, D.; Ortiz, C.; Torres, R. Synthesis, Characterization, and Evaluation of Antibacterial Effect of Ag Nanoparticles against Escherichia Coli O157:H7 and Methicillin- Resistant Staphylococcus Aureus (MRSA). *Int. J. Nanomedicine* **2014**, *9* (1), 1717–1729. <https://doi.org/10.2147/IJN.S57156>.
- (37) Martinez-Gutierrez, F.; Olive, P. L.; Banuelos, A.; Orrantia, E.; Nino, N.; Sanchez, E. M.; Ruiz, F.; Bach, H.; Av-Gay, Y. Synthesis, Characterization, and Evaluation of Antimicrobial and Cytotoxic Effect of Silver and Titanium Nanoparticles. *Nanomedicine Nanotechnology, Biol. Med.* **2010**, *6* (5), 681–688. <https://doi.org/10.1016/j.nano.2010.02.001>.
- (38) Martínez-Gutierrez, F.; Thi, E. P.; Silverman, J. M.; de Oliveira, C. C.; Svensson, S. L.; Hoek, A. Vanden; Sánchez, E. M.; Reiner, N. E.; Gaynor, E. C.; Pryzdial, E. L. G.; Conway, E. M.; Orrantia, E.; Ruiz, F.; Av-Gay, Y.; Bach, H. Antibacterial Activity, Inflammatory Response, Coagulation and Cytotoxicity Effects of Silver Nanoparticles. *Nanomedicine Nanotechnology, Biol. Med.* **2012**, *8* (3), 328–336. <https://doi.org/10.1016/j.nano.2011.06.014>.