

Cell Reports Medicine, Volume 1

Supplemental Information

**CD103⁺CD8⁺ T_{RM} Cells Accumulate in Tumors of
Anti-PD-1-Responder Lung Cancer Patients and
Are Tumor-Reactive Lymphocytes Enriched with Tc17**

Stéphanie Corgnac, Ines Malenica, Laura Mezquita, Edouard Auclin, Elodie Voilin, Jamila Kacher, Heloise Halse, Laetitia Grynszpan, Nicolas Signolle, Thibault Dayris, Marine Leclerc, Nathalie Droin, Vincent de Montpréville, Olaf Mercier, Pierre Validire, Jean-Yves Scoazec, Christophe Massard, Salem Chouaib, David Planchard, Julien Adam, Benjamin Besse, and Fathia Mami-Chouaib

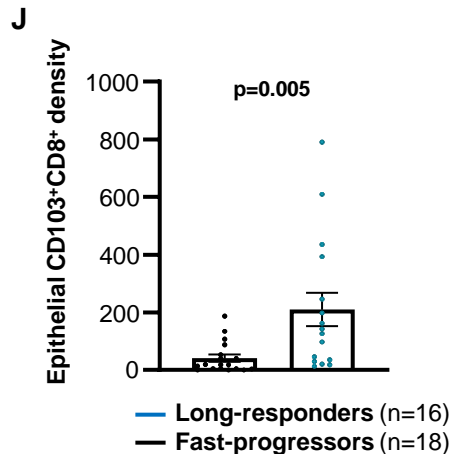
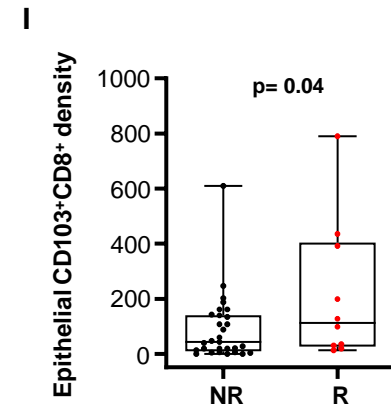
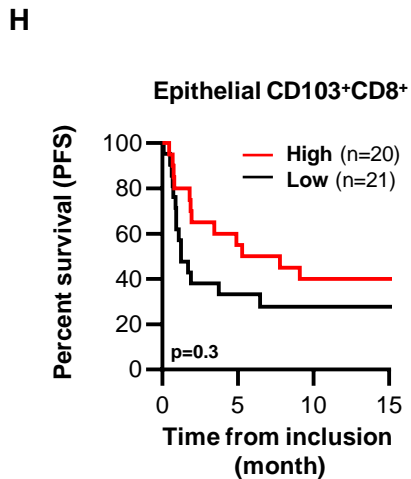
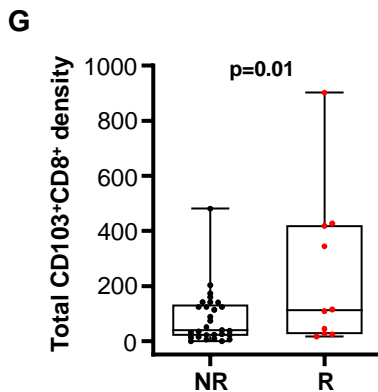
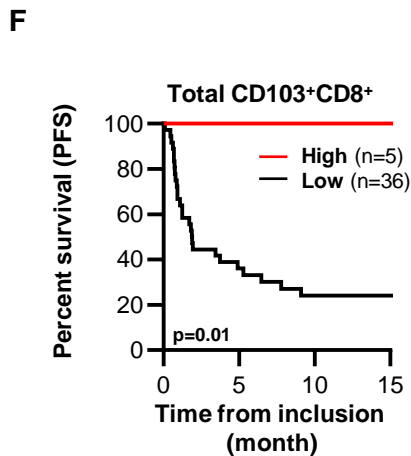
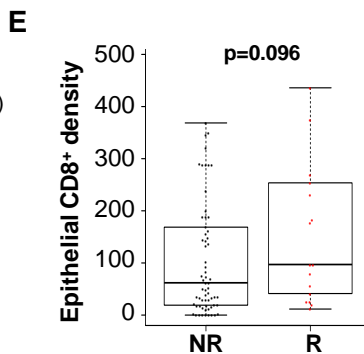
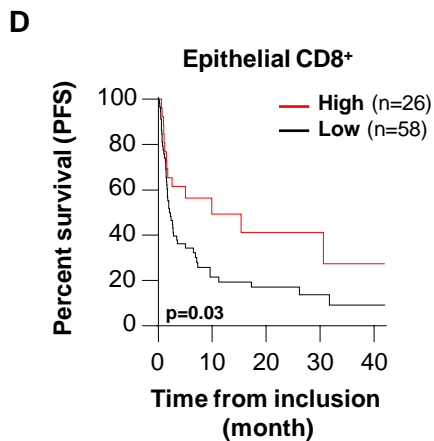
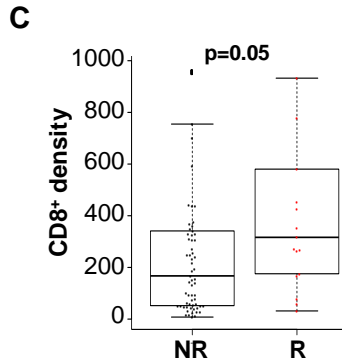
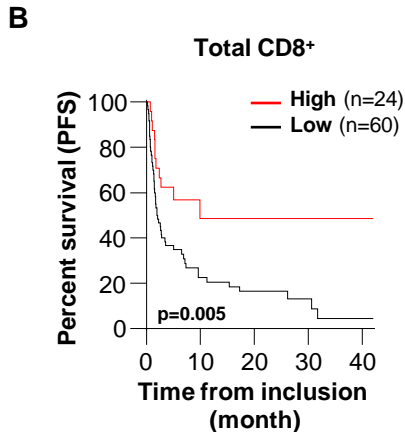
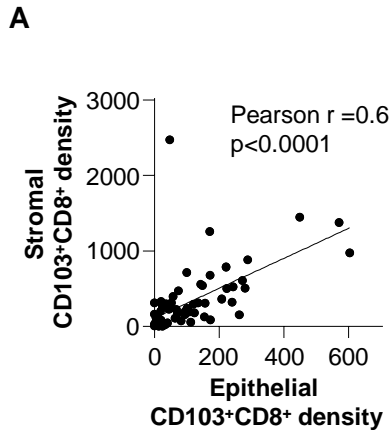


Figure S1. Density of CD8⁺ T cells in anti-PD-(L)1-treated patients. (Related to Figure 1)

(A) Correlation between the density of CD103⁺CD8⁺ cells in the stroma and in epithelial tumor regions. r-value indicates Pearson correlation coefficient. (B) Kaplan-Meier curve shows iPFS of PD-1 blockade-treated patients with tumors harboring a high density (CD8^{high}>350/mm²) or a low density (CD8^{low}<350/mm²) of CD8⁺ cells (n=86). (C) Density of CD8⁺ cells in tumors depending on iORR of non-responder (NR; n=65) and responder (R; n=17) patients to PD-1 blockade. (D) Kaplan-Meier curve shows iPFS of anti-PD-1-treated patients with epithelial tumor regions harboring a high (>169/mm²) or low (<169/mm²) density of CD8⁺ cells (n=86). (E) Density of CD8⁺ cells in epithelial tumor regions depending on iORR of NR (n=65) and R (n=17) patients to PD-1 blockade. (A-E) Discovery cohort. (F) Kaplan-Meier curve shows iPFS of PD-1 blockade-treated patients from a validation cohort with tumors harboring high (>252/mm²) or low (<252/mm²) densities of CD103⁺CD8⁺ cells (n=41). (G) Density of total CD103⁺CD8⁺ cells in tumors depending on iORR of NR (n=29) and R (n=10) patients to PD-1 blockade. (H) Kaplan-Meier curve shows iPFS of anti-PD-1-treated patients with tumor epithelial regions harboring a high (>48/mm²) or low (<48/mm²) density of CD103⁺CD8⁺ cells. (I) Density of CD103⁺CD8⁺ cells in epithelial tumor regions depending on iORR of NR (n=29) and R (n=10) patients to anti-PD-1. (J) Density of CD103⁺CD8⁺ cells in epithelial tumor regions of anti-PD-1-treated patients undergoing a long-response (PFS>6 months and OS>12 months; n=16) or a fast-progression (defined by an “early death” occurring during the first 12 weeks since the beginning of ICB; n=18). (F-J) validation cohort. p-value was determined by log-rank (B, D, F and H) or Chi2 test (C and E) or unpaired t-test (G, I and J). PFS: progression-free survival.

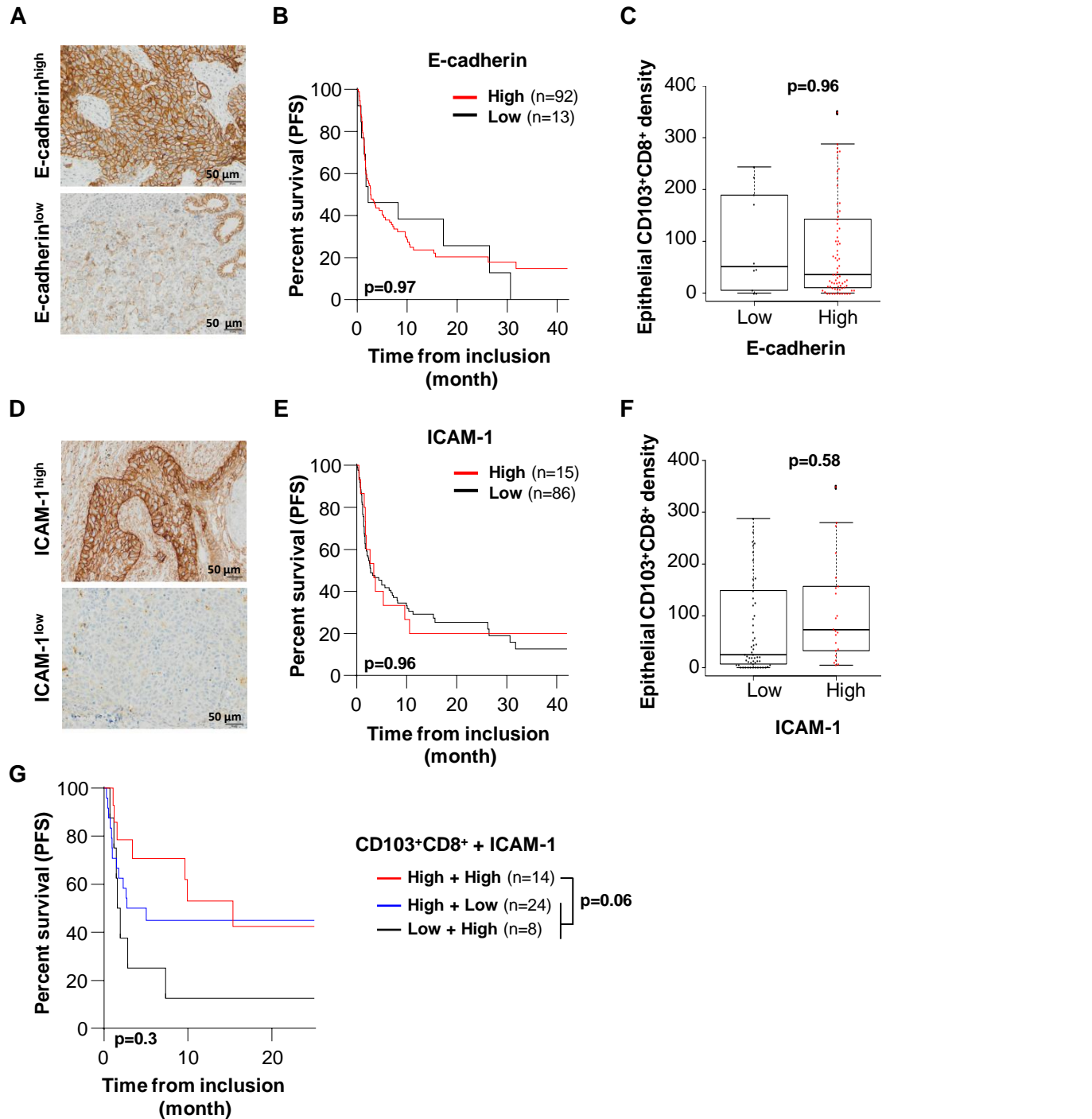


Figure S2. E-cadherin and ICAM-1 expression on tumor cells and patient survival. (Related to Figure 1)

(A) Representative images of positive and negative expression of E-cadherin by tumor cells. Scale bar 50 μ m (B) Kaplan-Meier curve shows iPFS of PD-1 blockade-treated patients with tumors harboring high and low expression of E-cadherin (n=105). (C) Density of CD103⁺CD8⁺ cells in epithelial tumor regions depending on E-cadherin expression on tumor cells (low, n=13; high, n=92). (D) Representative images of high and low expression of ICAM-1 on tumor cells. Scale bar 50 μ m (E) Kaplan-Meier curve shows iPFS of anti-PD-1-treated patients with tumors harboring high or low expression of ICAM-1 (n=101). (F) Density of CD103⁺CD8⁺ cells in epithelial tumor regions depending on ICAM-1 expression on tumor cells (low, n=86; high, n=15). (G) Kaplan-Meier curve shows iPFS of anti-PD-1-treated patients with epithelial tumor regions harboring high CD103⁺CD8⁺ infiltration and high ICAM-1 expression (red) or high CD103⁺CD8⁺ infiltration and weak expression of ICAM-1 (blue) or low CD103⁺CD8⁺ infiltration and high ICAM-1 expression (n=84). p-value was determined by log-rank test (B, E and G) or Chi2 test (C and F). The number of analyzed samples for each patient group is shown in parentheses.

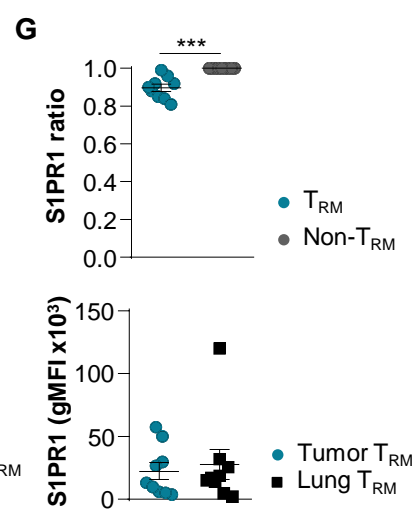
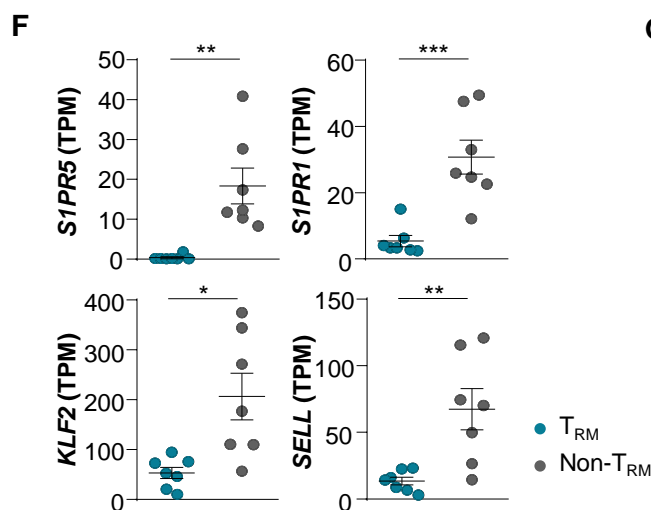
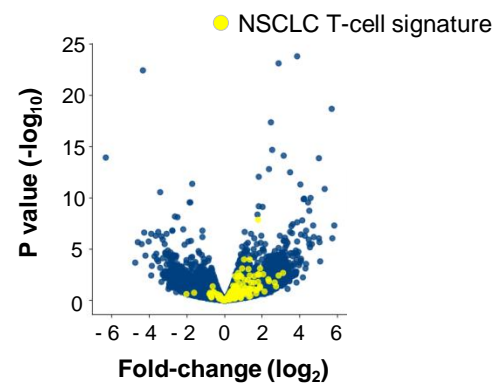
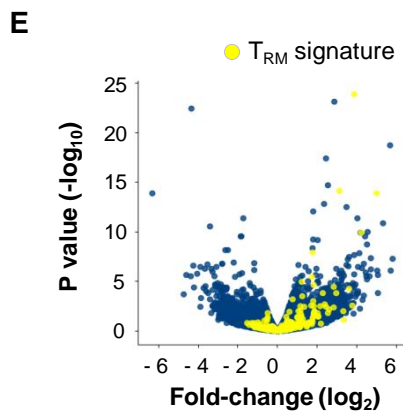
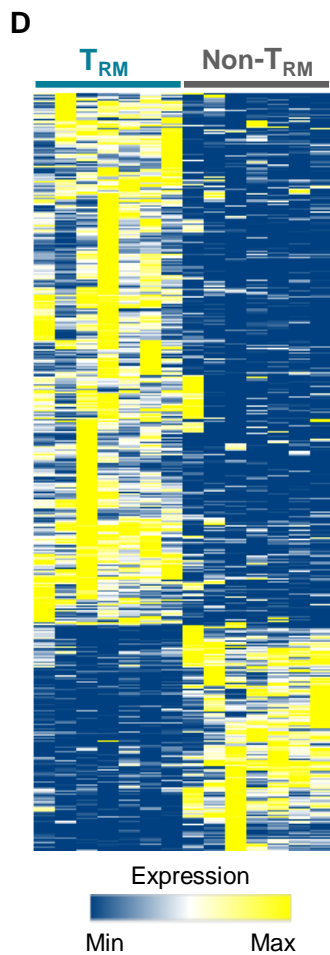
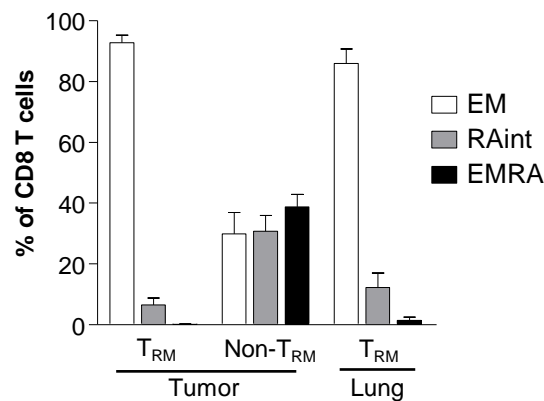
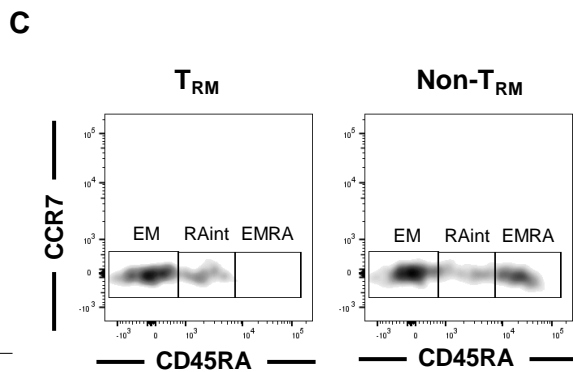
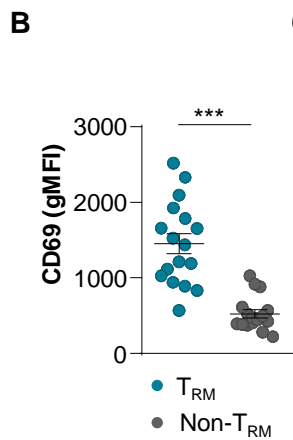
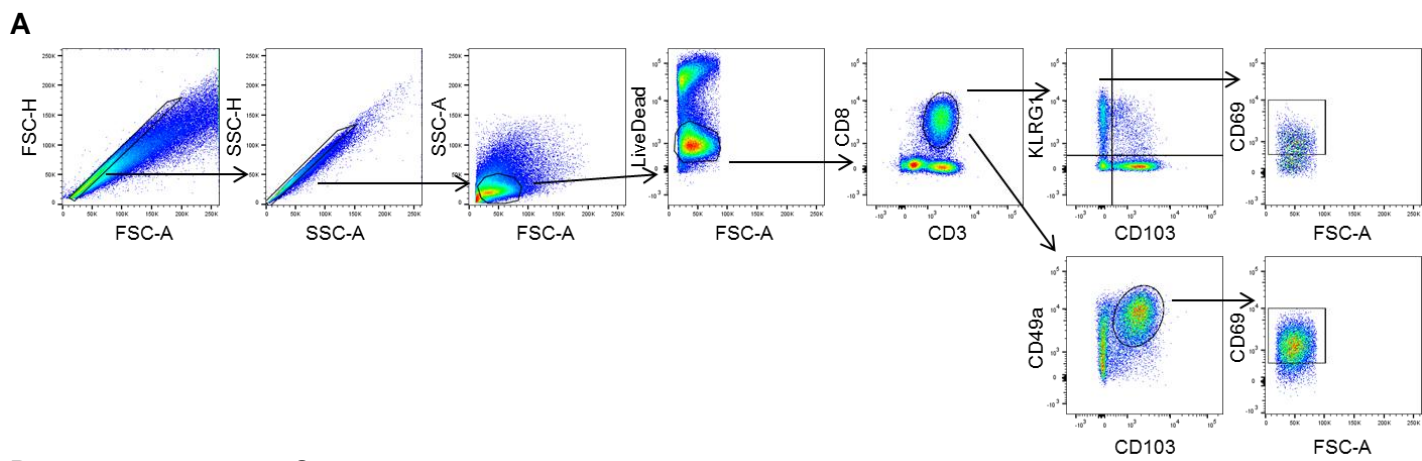


Figure S3. Expression of T_{RM} markers on CD8⁺ TIL and transcriptional profiles of CD103⁺ (T_{RM}) and KLRG1⁺ (non-T_{RM}) CD8⁺ T cells. (Related to Figures 2 and 3)

(A) Gating strategy for analysis of KLRG1, CD103, CD49a and CD69 by CD3⁺CD8⁺ TIL. (B) Expression of CD69 on tumor T_{RM} and non-T_{RM} (gMFI). (C) Dot plot showing expression of CCR7 and CD45RA on T_{RM} and non-T_{RM} from one representative NSCLC patient. Right, percentages of effector memory (EM), effector memory CD45RA intermediate (RAint) and effector memory CD45RA⁺ (EMRA) cells among T_{RM} and non-T_{RM} from tumor and paired T_{RM} from healthy lung (n=6). (D) Heat map of transcripts differentially expressed in paired CD103⁺ and KLRG1⁺ CD8⁺ TIL (n=7). Genes of interest (n=500) were selected using fold-change (FC) ≥2 and p-value ≤0.05. Data were bi-clustered by transcript using Pearson's coefficient as the distance metric. The magnitude of relative expression of a particular transcript is shown in color. Each column represents an individual patient. (E) Differential gene expression profiles in CD103⁺ and paired KLRG1⁺ CD8 T cells. T_{RM} up-gene signature: all transcripts (blue), upregulated gene sets related to T_{RM} (yellow)¹³. Right, differential expression of NSCLC T-cell signature genes in tumor T_{RM} and paired non-T_{RM} (n=7). All transcripts (blue) and upregulated genes related to NSCLC T-cell signature¹⁸ (yellow) are included. (F) Expression of *SIPR5*, *SIPR1*, *KLF2* and *SELL* genes in CD103⁺ and paired KLRG1⁺ CD8 T cells (n=7). Transcripts per million (TPM) are shown. (G) Ratio/non-T_{RM}) of geometric mean fluorescence intensity (gMFI) of S1pr1 in paired samples of tumor T_{RM} and non-T_{RM} (n=9). Down, expression of S1pr1 (gMFI) in T_{RM} cells from tumors or paired healthy lung tissues. Histograms represent mean ± SEM. * P<0.05, ** P<0.01, *** P<0.001 (paired t-test).

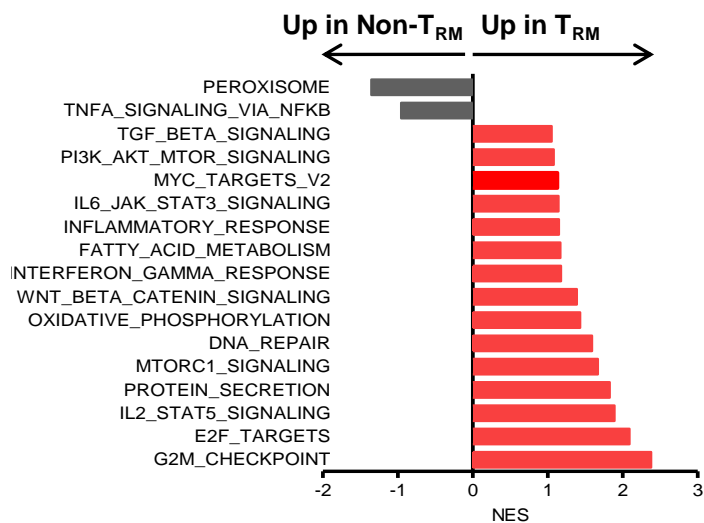
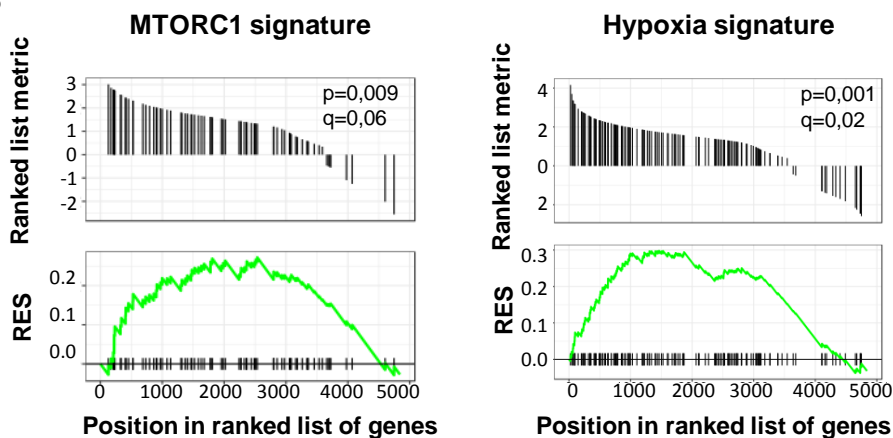
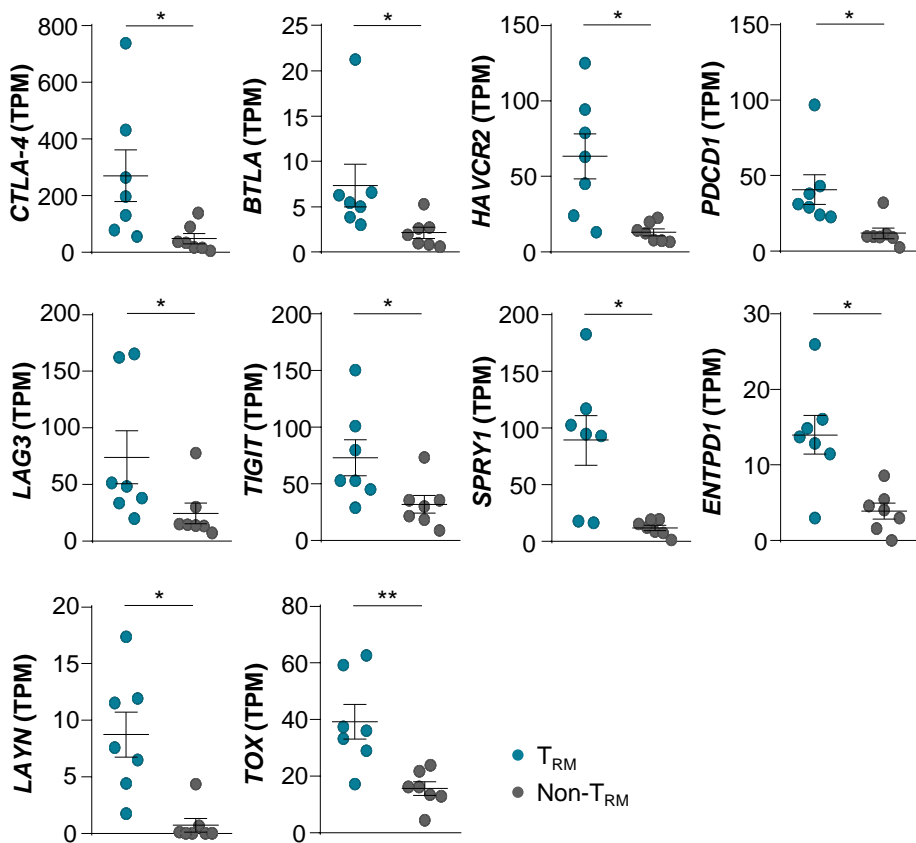
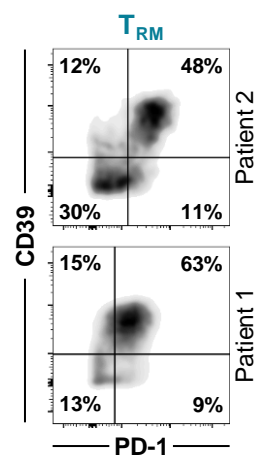
A**B****C****D**

Figure S4. Transcriptional and phenotypic profiles of CD8⁺ T_{RM} and non-T_{RM}. (Related to Figure 3)

(A) Gene set enrichment analysis (GSEA) of paired tumor T_{RM} and non-T_{RM} cells showing hallmark gene sets enriched in T_{RM} subset. Normalized enrichment scores (*NES*) are shown. (B) GSEA of mTORC1 (left) signature genes (Gene set M5924) and GSEA of hypoxia (right) signature genes (Gene set M48562) upregulated in the transcriptome of tumor T_{RM} relative to non-T_{RM}. Running enrichment score (RES) for gene set, as analysis ‘walks down’ ranked list of genes; position of the gene set members (black vertical lines) in ranked list of genes and value of ranking metric are shown. (C) Expression of *CTLA4*, *BTLA*, *HAVCR2*, *PDCD1*, *LAG3*, *TIGIT*, *SPRY1*, *ENTPD1*, *LAYN* and *TOX* genes in CD103⁺ and KLRG1⁺ CD8⁺ TIL. Values are TPM. (D) Representative dot plots of co-expression of CD39 and PD-1 on T_{RM} from two NSCLC patients (patients 1 and 2). Horizontal lines are mean ± SEM. * P<0.05, ** P<0.01 (paired t-test).

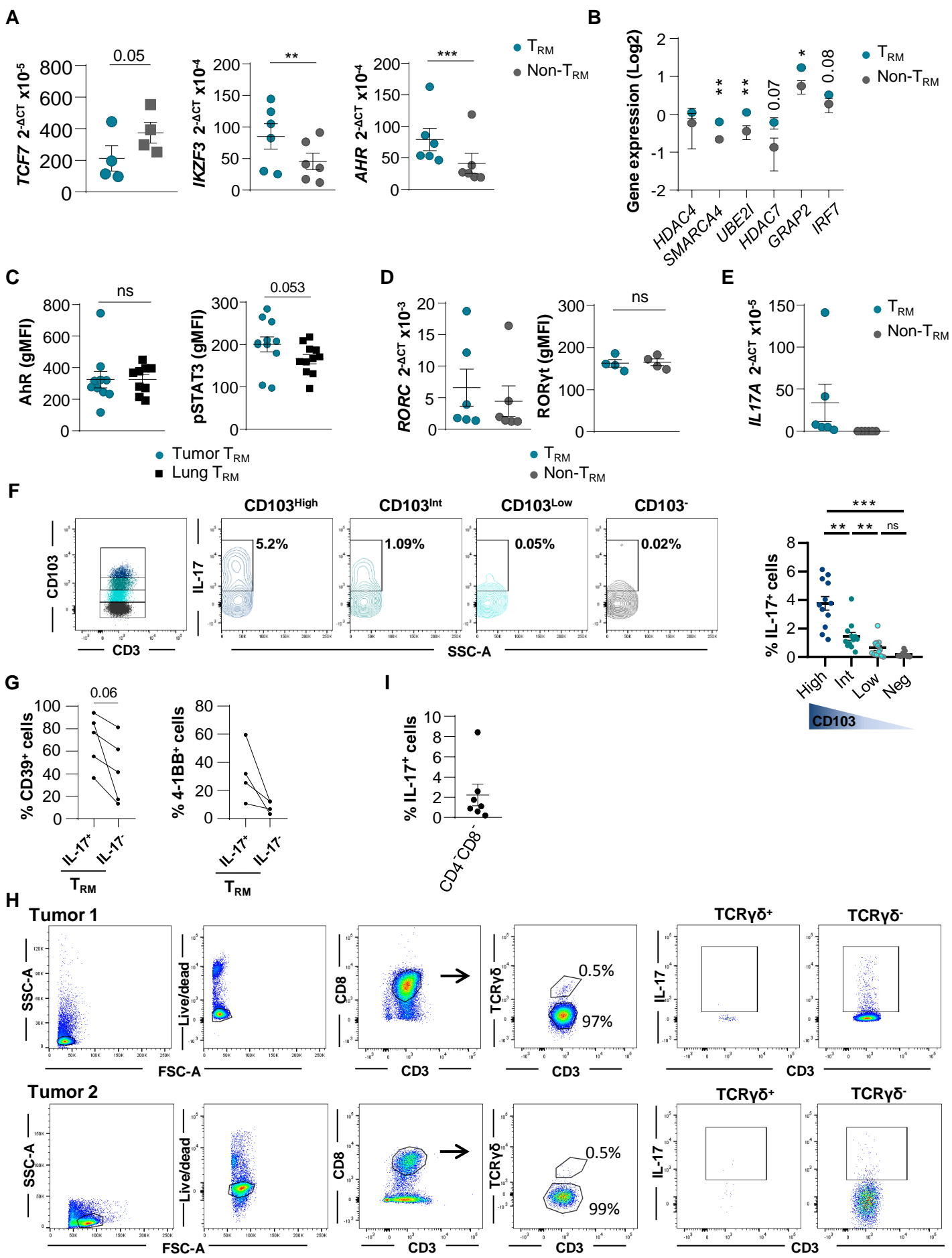


Figure S5. Lung tumor CD103⁺CD8⁺ T_{RM} display a Tc17-polarized pattern. (Related to Figures 4 and 5)

(A) *TCF7* (left), *IKZF3* (middle) and *AHR* (right) gene expression in T_{RM} and paired non-T_{RM} cells determined by RT-qPCR (n=4-6). Delta-CT values are shown. (B) Expression of genes involved in the Aiolos pathway in tumor T_{RM} and paired non-T_{RM} (n=7). Mean ± SEM of gene expression (Log2) are shown. (C) Expression (gMFI) of AhR (left) or pSTAT3 (right) in T_{RM} cells from tumor and paired healthy lung tissues (n=10). (D) Left, *RORC* gene expression (Delta-CT values) determined by RT-qPCR (n=6) and right, expression (gMFI) of RORγt in tumor T_{RM} and paired non-T_{RM} (n=4). (E) *IL17A* gene expression in tumor T_{RM} and paired non-T_{RM} determined by RT-qPCR (n=6). Delta-CT values are shown. (F) Dot plots of IL-17 expression in CD103⁺CD8⁺ TIL displaying CD103^{high}, CD103^{int} and CD103^{low} phenotype, and CD103⁻CD8⁺ TIL from one representative tumor. Right, percentages of IL-17-producing cells among CD103^{high}, CD103^{int}, CD103^{low} and CD103⁻ TIL (n=12). (G) Percentages of CD39⁺ (n=5) and 4-1BB⁺ (n=4) cells among IL17-producing (IL-17⁺) and IL17-non-producing (IL-17⁻) CD103⁺CD8⁺ T cells from NSCLC tumors. (H) Dot plots of TCRγδ expression on CD3⁺CD8⁺ TIL from two representative tumors. Right, IL-17 production by TCRγδ⁺ and TCRγδ⁻ CD8⁺ T cells stimulated 3h with PMA plus ionomycin. Two representative tumor samples are shown. (I). Percentages of IL-17⁺ cells among CD8⁻CD4⁻ T cells from NSCLC tumors after 4h stimulation with PMA/ionomycin (n=7). Each symbol represents one individual patient, horizontal lines are the mean ± SEM. * P<0.05 ** P<0.01, *** P<0.001 (paired t-test, A, B, C, and G or ANOVA with Bonferroni post-hoc test, F).

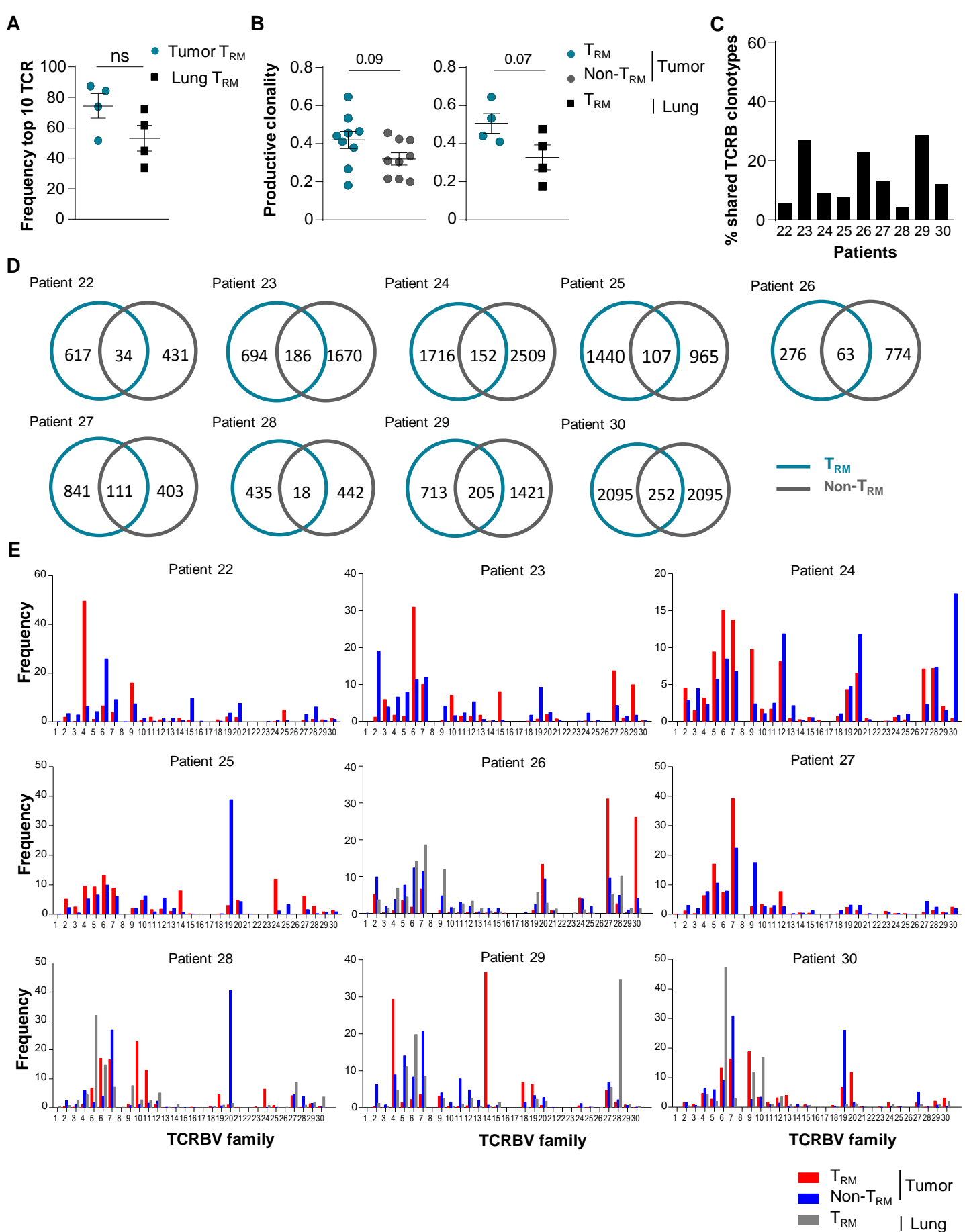


Figure S6. Tumor CD103⁺CD8⁺ T_{RM} displays a more oligoclonal TCR repertoire than non-T_{RM} counterparts. (Related to Figure 7)

(A) Frequency of the top ten TCR sequences in T_{RM} from tumors and healthy lung counterparts (n=4). (B) Productive clonality of tumor T_{RM} and paired non-T_{RM} (n=9). Right, productive clonality of T_{RM} from tumors and healthy lung tissues (n=4). Each symbol represents one individual patient; horizontal lines are the mean ± SEM. p-values are calculated with paired t-test. (C) Percentages of shared TCRβ clonotypes between tumor T_{RM} and non-T_{RM} in each patient (n=9). (D) Number of individual and overlap sequence diversities of TCRβ CDR3 repertoires from tumor T_{RM} and paired non-T_{RM} (n=9). (E) Histograms represent productive frequencies of the 30 TCRBV gene segments for tumor T_{RM} and paired non-T_{RM} (n=9), and paired healthy lung T_{RM} (n=4).

Table S1: Population description and IHC data of the discovery cohort (Related to Figure 1)

Population description		Overall population (n=86)	Density CD103 ⁺ CD8 ⁺ low (n=67)	Density CD103 ⁺ CD8 ⁺ high (n=19)	p-value
Age	Median, range	63 (30-84)	63 (30-84)	63 (43-73)	0.98
Gender	Female	27 (32%)	20 (31%)	7 (37%)	0.62
	Male	57 (68%)	45 (69%)	12 (63%)	
Smoking status	Current	28 (33%)	22 (34%)	6 (32%)	0.75
	Former	47 (56%)	35 (54%)	12 (63%)	
	Non-smoker	9 (11%)	8 (12%)	1 (5%)	
Histology	Adenocarcinoma	51 (61%)	41 (63%)	10 (53%)	0.65
	NSCLC, other	6 (7%)	5(8%)	1 (5%)	
	Squamous	27 (32%)	19 (29%)	8 (42%)	
Stage at diagnosis	I-IIIa	29 (35%)	21 (32%)	8 (44%)	0.34
	IIIB	14 (17%)	11 (17%)	3 (17%)	
	IVA- IVB	40 (48%)	33 (51%)	7 (39%)	
	NA	0	0	0	
Molecular status	<i>EGFR</i> mutation	5 (6%)	4 (6%)	1 (5%)	0.81
	<i>ALK</i> fusion	1 (1%)	1 (2%)	0	0.42
	<i>KRAS</i> mutation	19 (23%)	14 (22%)	5 (26%)	0.58
	<i>BRAF</i> mutation	2 (2%)	2 (3%)	0	0.87
Previous therapies	Platinum-based chemo	33 (41%)	25 (40%)	8 (44%)	0.037
	Second-line chemo	14 (17%)	14 (11%)	4 (22%)	
	Other therapies	17 (21%)	17 (27%)	4 (22%)	
	No prior therapy	6 (7%)	5 (8%)	1 (6%)	
	Chemo-radiation	11 (14%)	6 (10%)	5 (28%)	
Line of immunotherapy	≤ 2	45 (54%)	32 (49%)	13 (68%)	0.14
	> 2	39 (46%)	33 (51%)	6 (32%)	
Number of metastatic sites	≤ 2	46 (55%)	35 (54%)	11 (58%)	0.75
	> 2	38 (45%)	30 (46%)	8 (42%)	
	NA	2	2	0	
Type of immunotherapy	PD-1 inhibitor	69 (82%)	54 (83%)	15 (79%)	0.74
	PD-L1 inhibitor	15 (18%)	11 (17%)	4 (21%)	
Performance status	0-1	68 (82%)	50 (78%)	18 (95%)	0.17
	≥ 2	15 (18%)	14 (22%)	1 (5%)	
	NA	3	3	0	
Response rate	Complete response	2 (3%)	1 (2%)	1 (6%)	0.024
	Partial response	15 (18%)	8 (13%)	7 (39%)	
	Stable disease	28 (34%)	22 (34%)	6 (33%)	
	Progressive disease	37 (45%)	33 (51%)	4 (22%)	
	NA	4	3	1	
Objective response rate	CR + PR	17 (21%)	9 (14%)	8 (44%)	0.005
NA	4	3	1		
ICAM-1 expression	Median, range	80 (0-300)	70 (0-300)	140 (0-300)	0.23
	NA	3	3	0	
ICAM-1 expression by category	Low	61 (73%)	49 (77%)	12 (63%)	0.25
	High	22 (27%)	15 (23%)	7 (37%)	
	NA	0	0	0	
E-Cadherin expression by category	0-1	10 (12%)	7 (11%)	3 (16%)	0.73
	2-3	74 (88%)	58 (89%)	16 (84%)	
	NA	2	2	0	
PD-L1 expression	Median, range	28 (0-100)	10 (0-95)	80 (10-100)	0.023
	NA	46	36	10	
Epithelial CD8 ⁺ density	Low	59 (69%)	57 (85%)	2 (11%)	<0.001
	High	27 (31%)	10 (15%)	17 (89%)	
Density stromal CD8 ⁺	Low	60 (71%)	60 (90%)	0	<0.001
	High	24 (29%)	7 (10%)	17 (100%)	
	NA	2	0	2	
Total CD8 ⁺ density	Low	62 (72%)	61 (91%)	1 (5%)	<0.001
	High	24 (28%)	6 (9%)	18 (95%)	
Epithelial CD103 ⁺ CD8 ⁺ density	Low	45 (52%)	44 (66%)	1 (5%)	<0.001
	High	41 (48%)	23 (34%)	18 (95%)	
Stromal CD103 ⁺ CD8 ⁺ density	Low	61 (71%)	60 (90%)	1 (5%)	<0.001
	High	25 (29%)	7 (10%)	18 (95%)	
Total CD103 ⁺ CD8 ⁺ density	Low	67 (78%)	67 (100%)	0	<0.001
	High	19 (22%)	0	19 (100%)	
Epithelial CD103 ⁺ CD8 ⁺ density	Low	41 (48%)	36 (54%)	5 (26%)	0.01
	High	45 (52%)	31 (46%)	14 (74%)	
Stromal CD103 ⁺ CD8 ⁺ density	Low	50 (58%)	48 (72%)	2 (11%)	0.001
	High	36 (42%)	19 (28%)	17 (89%)	
Total CD103 ⁺ CD8 ⁺ density	Low	70 (81%)	62 (93%)	8 (42%)	0.001
	High	16 (19%)	5 (7%)	11 (58%)	
Total CD103 ⁺ CD8 ⁺ density (number)		102.59 (0;1320.75)	65.96 (0;246.8)	466.21 (252.42;1320.75)	<0.001
Epithelial CD103 ⁺ CD8 ⁺ density (number)		44.56 (0;603.82)	21.56 (0;262.13)	240.23 (48.14;603.82)	<0.001
Stromal CD103 ⁺ CD8 ⁺ density (number)		152.75 (0;2477.96)	93.81 (0;478.15)	678.6 (283.59;2477.96)	<0.001
Percentage of CD103 ⁺ among CD8 ⁺ in epithelial regions		76.19 (0;100)	67.48 (0;100)	83.33 (29.87;100)	0.001
Percentage of CD103 ⁺ among CD8 ⁺ in stromal regions		47.62 (0;94.44)	42.6 (0;94.44)	58.61 (26.47;87.69)	0.046

NA: not available; CR: complete response; PR: partial response.

Table S2: Immunotherapy progression-free survival (iPFS) median (Discovery cohort) (Related to Figure 1)

Variables			Total patient number	Percentage (%)	Events number	Median (95%CI) (month)	Log-rank p-value
			84	100	62	2.69 (1.74-7.13)	
CD8 ⁺ cells	Total	Low	60	71.4	51	2.05 (1.51-5.03)	0.005
		High	24	28.6	11	9.92 (2.73-NR)	
	Stromal	Low	58	69	50	2.05 (1.51-5.03)	0.001
		High	24	28.6	11	30.62 (3.42-NR)	
	Epithelial	Low	58	69	48	2.22 (1.64-4.99)	0.03
		High	26	31	14	9.92 (1.74-NR)	
CD103 ⁺ CD8 ⁺ cells	Total	Low	65	77.4	54	2.3 (1.68-5.03)	0.008
		High	19	22.6	8	30.6 (2.73-NR)	
	Stromal	Low	59	70.2	50	2.14 (1.51 -5.03)	0.006
		High	25	29.8	12	30.62 (2.30-NR)	
	Epithelial	Low	44	52.4	37	1.76 (1.48-4.99)	0.01
		High	40	47.6	25	9.63 (2.56-NR)	
CD103 ⁻ CD8 ⁺ cells	Total	Low	68	81	55	2.5 (1.68-6.54)	0.02
		High	16	19	7	NR (1.58-NR)	
	Stromal	Low	49	58.3	43	2.14 (1.64-5.03)	0.001
		High	35	41.7	19	9.92 (1.81-NR)	
	Epithelial	Low	41	48.8	27	6.90 (2.76-26.22)	0.03
		High	43	51.2	35	1.68 (1.31-3.42)	
CD103/CD8 score	Good		12	14.3	5	26.22 (15.34-NR)	<0.0001
	Intermediate		57	67.9	42	2.76 (1.70-7.33)	
	Poor		15	17.9	15	1.38 (0.72-1.97)	
ICAM-1	Low		86	85.1	66	2.79 (1.97-7.33)	0.96
	High		15	14.9	12	3.42 (1.81-NR)	
E-cadherin	Low		13	12.4	11	2.23 (1.41-NR)	0.97
	High		92	87.6	73	2.74 (1.94-5.68)	
Epithelial CD103 ⁺ CD8 ⁺ cells and ICAM-1	High/High		14	16.6	8	15.34	0.3
	High/Low		24	28.5	15	3.87	
	Low/High		8	9.5	7	1.75	

NR: not reached, CI: confidence interval

Table S3: Multivariate analysis for progression-free survival (Discovery cohort) (Related to Figure 1)

Variables	HR (95% CI)	p-value
Age \geq 65	1.79 (0.73-4.37)	0.20
Histology squamous	1.5 (0.83-2.73)	0.18
Smoker	0.35 (0.14-0.84)	0.02
Performance status \geq 2	1.12 (0.57-2.21)	0.73
Number of metastatic sites	1.83 (1.02-3.29)	0.04
High CD103 ⁺ CD8 ⁺ density	0.39 (0.18-0.85)	0.02

HR: hazard ratio; CI: confidence interval.

Table S4: Population description and immunohistochemistry data of the validation cohort (Related to Figure 1)

Population description		Overall population (n=41)
Age	Median, range	64 (42-85)
Gender	Female	20 (49%)
	Male	21 (51%)
Smoking status	Current	18 (44%)
	Former	19 (46%)
	Non-smoker	4 (10%)
Histology	Adenocarcinoma	34 (83%)
	NSCLC, others	1 (2.5%)
	Squamous	6 (14.5%)
Stage at diagnosis	IIA	2 (5%)
	IIB	1 (2.5%)
	IIIA	3 (7%)
	IIIB	2 (5%)
	IVA	7 (17%)
	IVB	26 (63.5%)
Molecular status	<i>EGFR</i> mutation	2 (5%)
	<i>ALK</i> fusion	2 (5%)
	<i>KRAS</i> mutation	16 (42%)
	<i>BRAF</i> mutation	2 (5%)
Line of immunotherapy	≤ 2	33 (80%)
	> 2	8 (20%)
Number of metastatic sites	≤ 2	12 (29%)
	> 2	29 (71%)
Type of immunotherapy	PD-1 inhibitor	41 (100%)
Performance status	0-1	26 (63%)
	≥ 2	15 (37%)
Response rate	Complete response	2 (5%)
	Partial response	8 (20%)
	Stable disease	10 (26%)
	Progressive disease	19 (49%)
	NA	2
Objective response rate	CR + PR	10 (26%)
	NA	2
PD-L1 expression	Median, range	10 (0-100)
Epithelial CD8 ⁺ density	Low	21 (51%)
	High	20 (49%)
Stromal CD8 ⁺ density	Low	26 (63%)
	High	15 (37%)
Total CD8 ⁺ density	Low	24 (59%)
	High	17 (41%)
Epithelial CD103 ⁺ CD8 ⁺ density	Low	21 (51%)
	High	20 (49%)
Stromal CD103 ⁺ CD8 ⁺ density	Low	34 (83%)
	High	7 (17%)
Total CD103 ⁺ CD8 ⁺ density	Low	36 (88%)
	High	5 (12%)
Epithelial CD103 ⁻ CD8 ⁺ density	Low	7 (17%)
	High	34 (83%)
Stromal CD103 ⁻ CD8 ⁺ density	Low	20 (49%)
	High	21 (51%)
Total CD103 ⁻ CD8 ⁺ density	Low	23 (56%)
	High	18 (44%)
Total CD103 ⁺ CD8 ⁺ density (number)	Median (range)	120.4 (0;902.4)
Epithelial CD103 ⁺ CD8 ⁺ density (number)	Median (range)	118 (0;789.85)
Stromal CD103 ⁺ CD8 ⁺ density (number)	Median (range)	150.95 (0;1147.28)

CR: complete response; PR: partial response; NA: not available

Table S5: Patient characteristics and density of cells pre- and post-treatment (Related to Figure 1)

Group of patients	Patient N°	Number of ICB cycles	RECIST/Radiologic response (tumor volume reduction)	Responder/Non-responder	Stage diagnosis	OS (months)	Site of pre-biopsy	Site of post-biopsy	Days from last ICB to post-biopsy	Density of intra-epithelial CD103 ⁺ CD8 ⁺ cells		Density of total CD103 ⁺ CD8 ⁺ cells		Density of total CD8 ⁺ cells	
										Pre-ICB	Post-ICB	Pre-ICB	Post-ICB	Pre-ICB	Post-ICB
1	1	7	SD	R	IIIA	11.5	Lung	Lung	24	161	682	204	931	417	1779
	2	NA	SD	R	IVB	13.9	Lung	Lung	135	358	511	129	518	493	1081
	3	NA	SD	R	IVB	14.3	Kidney meta	Kidney meta	174	162	134	173	159	729	666
	4	1	PD	NR	IVA	2	Lung	Lung	14	19	0	25	0	58	30
	5	NA	PD	NR	IIIA	18.4	Lung	Lung	34	18	6	44	21	221	600
2	6	1	-17%	R	NA	25.3	Lung	Lung	28	172	555	159	534	322	895
	7	1	-17%	R	IIIA	31.5	Lung	Lung	27	25	155	33	155	135	342
	8	1	-5%	NR	IB	37.1	Lung	Lung	21	230	133	258	322	1691	1775
	9	1	0%	NR	IIA	31.2	Lung	Lung	25	267	95	208	225	781	538

ICB: immune checkpoint blockade, OS : overall survival ; SD: stable disease; PD: progressive disease; R: responder; NR: non-responder; Meta: metastasis; NA: not available

Table S6: Population description of ICAM and E-cadherin cohort (Discovery cohort) (Related to Figure 1)

Population description		Overall population (N=111)
Age	Median, range	63 (30-92)
Gender	Female	35 (32%)
	Male	76 (68%)
Smoking status	Current	36 (32%)
	Former	64 (58%)
	Non-smoker	11 (10%)
Histology	Adenocarcinoma	66 (60%)
	NSCLC, other	16 (14%)
	Squamous	29 (26%)
Stage at diagnosis	I-III A	4 (4%)
	III B	11 (10%)
	IV A- IV B	95 (86%)
	Missing	1
Molecular status	<i>EGFR</i> mutation	5 (6%)
	<i>ALK</i> fusion	2 (2%)
	<i>KRAS</i> mutation	24 (30%)
	<i>BRAF</i> mutation	3 (4%)
Previous therapies	Platinum-based chemo	55 (50%)
	Second-line chemo	14 (13%)
	Other therapies	22 (20%)
	No prior therapy	6 (5%)
	Chemo-radiation	13 (12%)
Line of immunotherapy	≤ 2	64 (58%)
	> 2	47 (42%)
Number of metastatic sites	≤ 2	54 (49%)
	> 2	57 (51%)
	Missing	0
Type of immunotherapy	PD-1 inhibitor	96 (86%)
	PD-L1 inhibitor	15 (14%)
Performance status	0-1	90 (82%)
	≥2	20 (18%)
	Missing	1
Response rate	Complete response	2 (2%)
	Partial response	23 (22%)
	Stable disease	38 (36%)
	Progressive disease	44 (40%)
	Missing	4
Objective response rate	CR + PR	63 (58%)
	Missing	3
ICAM-1 expression	Median range	80 (0-300)
	Missing	10
ICAM-1 expression by category	Low	86 (85%)
	High	15 (15%)
	Missing	10
E-Cadherin expression by category	0-1	13 (12%)
	2-3	92 (88%)
	Missing	6

CR: complete response; PR: partial response.

Table S7: Frequencies of T_{RM} and non-T_{RM} cells in lung tumors and autologous adjacent normal lung tissues (Related to Figure 2)

Patient	Lung tumor				Adjacent normal lung			
	CD103 ⁺ CD49a ⁺ CD49a ⁺		KLRG1 ⁺		CD103 ⁺ CD49a ⁺ CD49a ⁺		KLRG1 ⁺	
	%	Number/mg	%	Number/mg	%	Number/mg	%	Number/mg
1	55	320	28	164	56	44	28	22
2	52	2340	31	1271	31	44	38	47
3	44	614	37	515	8	76	85	845
4	27	354	53	706	25	369	44	656
5	36	57	41	67	7	48	57	394
6	37	374	40	401	28	12	46	20
7	62	781	20	254	13	315	77	1881
8	30	3013	37	3767	27	64	61	144
9	25	143	50	287	5	6	79	111
10	81	540	10	70	41	49	30	36
11	35	314	32	310	43	126	31	90
12	87	3823	5	204	52	44	24	22
13	51	547	23	249	2	17	82	623
14	80	1727	4	93	59	640	6	60
15	48	368	31	238	16	28	51	89
16	59	797	15	204	28	85	21	63
17	40	133	38	126	2	1	91	37
18	67	499	18	135	49	81	13	22

The total number of T_{RM} per milligram of tissue was included

Table S8: Differential gene expression profiles of CD103⁺CD8⁺ TIL and autologous KLRG1⁺CD8⁺ TIL (Related to Figures 3, 4 and 5)

Gene	Fold change	p-value			
Activation/Th17					
<i>IL17A</i>	23.62	1.08 10 ⁻¹⁰⁰	<i>VCAMI</i>	5.27	1.59 10 ⁻²
<i>TNFSF4</i>	20.32	1.69 10 ⁻¹	<i>CD9</i>	5.25	1.32 10 ⁻²
<i>TMGD2</i>	9.32	8.97 10 ⁻⁴	<i>IKK</i>	4.63	5.04 10 ⁻²
<i>IL2RB2</i>	7.87	5.28 10 ⁻⁴	<i>ITGB7</i>	4.17	3.69 10 ⁻²
<i>CK1F</i>	7.74	3.74 10 ⁻³	<i>CCR6</i>	3.86	4.03 10 ⁻²
<i>CD2</i>	7.32	3.67 10 ⁻³	<i>CXCR6</i>	3.54	3.28 10 ⁻³
<i>CD82</i>	7.26	6.52 10 ⁻³	<i>ITGAD</i>	3.47	1.69 10 ⁻³
<i>CPN1</i>	7.25	5.64 10 ⁻³	<i>CD248</i>	2.64	1.61 10 ⁻²
<i>ICOS</i>	6.72	4.59 10 ⁻³	<i>KIRC2</i>	2.54	4.00 10 ⁻²
<i>CD6</i>	6.61	4.29 10 ⁻²	<i>CD44</i>	-1.61	2.73 10 ⁻²
<i>CD226</i>	6.12	1.33 10 ⁻³	<i>CXCR5</i>	-1.89	1.31 10 ⁻³
<i>IL26</i>	5.69	1.41 10 ⁻⁴	<i>CCR4</i>	-2.29	1.93 10 ⁻³
<i>CD63</i>	5.53	8.10 10 ⁻³	<i>CCR7</i>	-2.68	1.34 10 ⁻³
<i>CD109</i>	5.21	2.61 10 ⁻²	<i>ITGA5</i>	-3.62	1.36 10 ⁻²
<i>IL32</i>	5.11	2.81 10 ⁻²	<i>ITGAM</i>	-3.67	4.45 10 ⁻²
<i>CSF2</i>	4.36	3.76 10 ⁻³	<i>SELL</i>	-4.88	1.12 10 ⁻²
<i>IRF5</i>	4.31	1.02 10 ⁻²	<i>S1PR1</i>	-5.12	3.80 10 ⁻²
<i>CD101</i>	4.08	9.70 10 ⁻³	<i>KIR2F1</i>	-5.23	1.58 10 ⁻³
<i>CD70</i>	3.21	1.14 10 ⁻²	<i>CXCR2</i>	-8.84	2.02 10 ⁻³
<i>CD7</i>	2.99	3.76 10 ⁻²	<i>CXCR1</i>	-8.99	3.23 10 ⁻³
<i>RAB2A</i>	2.99	3.80 10 ⁻²	<i>CX3CR1</i>	-10.57	2.67 10 ⁻¹¹
<i>HAVCR1</i>	2.90	1.26 10 ⁻²	<i>KIR3G1</i>	-14.79	3.38 10 ⁻⁷
<i>IL17RA</i>	2.75	2.88 10 ⁻²	<i>ICAM2</i>	-19.03	2.51 10 ⁻⁷
<i>GZMB</i>	2.54	5.54 10 ⁻⁴	<i>S1PR5</i>	-20.95	3.07 10 ⁻⁹
<i>IL18RAP</i>	2.20	7.86 10 ⁻³	TRM signature		
<i>ITNG</i>	1.99	4.44 10 ⁻²	<i>GSG2</i>	8.75	4.73 10 ⁻²⁰
<i>GZMA</i>	1.68	2.89 10 ⁻²	<i>GPR34</i>	7.45	8.42 10 ⁻³
<i>CD244</i>	1.52	1.18 10 ⁻²	<i>ITGAE</i>	5.90	1.51 10 ⁻²⁸
<i>IL2RG</i>	1.45	4.89 10 ⁻²	<i>CXCR6</i>	3.29	3.89 10 ⁻⁹
<i>TNFRSF25</i>	1.39	2.34 10 ⁻²	<i>ACP5</i>	2.64	5.14 10 ⁻²
<i>TGFB3</i>	-1.64	1.81 10 ⁻²	<i>LDLRAD4</i>	2.33	1.28 10 ⁻³
<i>IL5</i>	-1.96	3.90 10 ⁻²	<i>AMICA1</i>	2.21	9.58 10 ⁻³
<i>CD28</i>	-2.13	3.97 10 ⁻³	<i>ATP10D</i>	2.15	2.28 10 ⁻³
<i>GZMK</i>	-3.44	7.07 10 ⁻³	<i>ITNG</i>	2.00	3.24 10 ⁻³
<i>IL2A</i>	-3.47	7.91 10 ⁻³	<i>ABTB2</i>	1.83	1.24 10 ⁻²
Inhibition/exhaustion			<i>QPCT</i>	1.80	6.25 10 ⁻²
<i>ENTPD1</i>	56.05	5.21 10 ⁻⁹	<i>XCL1</i>	1.67	2.53 10 ⁻²
<i>LAYN</i>	21.84	1.22 10 ⁻⁹	<i>CD244</i>	1.59	4.16 10 ⁻³
<i>SPRY1</i>	9.09	2.28 10 ⁻⁴	<i>KCNN4</i>	1.57	2.78 10 ⁻²
<i>CD200</i>	7.74	2.71 10 ⁻³	<i>RACGAP1</i>	1.53	8.43 10 ⁻²
<i>CTLA4</i>	5.14	1.48 10 ⁻²	<i>M-APKAP3</i>	1.52	6.95 10 ⁻⁴
<i>BTLA</i>	4.70	1.98 10 ⁻³	<i>P2RY10</i>	1.45	6.20 10 ⁻²
<i>HAVCR2</i>	2.78	3.66 10 ⁻³	<i>CCL5</i>	1.42	5.59 10 ⁻²
<i>PDCD1</i>	2.63	1.79 10 ⁻²	<i>PCEID1B</i>	1.41	1.04 10 ⁻²
<i>LAG3</i>	2.20	8.93 10 ⁻³	<i>VOPP1</i>	1.31	2.05 10 ⁻²
<i>TIGIT</i>	1.86	5.22 10 ⁻³	<i>ABB</i>	1.30	3.85 10 ⁻²
<i>CBLB</i>	1.43	4.85 10 ⁻²	<i>DOCK2</i>	1.27	5.22 10 ⁻²
<i>TOX</i>	1.96	3.15 10 ⁻⁴	<i>CDC25B</i>	-1.22	6.84 10 ⁻²
<i>TOX2</i>	12.96	3.71 10 ⁻⁴	<i>LITAF</i>	-1.25	3.67 10 ⁻²
<i>SPRY2</i>	-2.91	3.97 10 ⁻²	<i>NEDD4</i>	-1.36	4.25 10 ⁻²
Transcription factors			<i>BCL9L</i>	-1.38	2.85 10 ⁻²
<i>ILF3</i>	17.93	6.01 10 ⁻³	<i>STK38</i>	-1.41	1.52 10 ⁻³
<i>ATP8B4</i>	17.50	1.53 10 ⁻⁹	<i>FAM65B</i>	-1.48	6.78 10 ⁻²
<i>GF11</i>	9.24	5.20 10 ⁻⁴	<i>SAMHD1</i>	-1.50	1.78 10 ⁻³
<i>IRF3</i>	7.75	9.65 10 ⁻⁴	<i>FRMD4B</i>	-1.84	2.72 10 ⁻²
<i>ZEB1</i>	7.38	8.53 10 ⁻⁴	<i>LEF1</i>	-1.87	8.56 10 ⁻³
<i>ZNF683</i>	6.55	4.10 10 ⁻⁴	<i>PDE2A</i>	-1.90	9.30 10 ⁻²
<i>ZNF384</i>	5.28	3.91 10 ⁻²	<i>ICAM2</i>	-1.94	2.59 10 ⁻⁴
<i>FOX.B</i>	5.22	1.72 10 ⁻²	<i>RASA3</i>	-2.34	1.88 10 ⁻⁷
<i>IKZF4</i>	5.05	1.10 10 ⁻²	<i>EOMES</i>	-2.35	2.18 10 ⁻³
<i>FOX P3</i>	4.80	9.39 10 ⁻⁴	<i>KIF2</i>	-2.46	1.58 10 ⁻⁴
<i>BACH1</i>	4.43	1.62 10 ⁻²	<i>TCF7</i>	-2.47	7.30 10 ⁻⁴
<i>E2F7</i>	4.00	5.09 10 ⁻²	<i>FAM9A</i>	-2.95	9.02 10 ⁻³
<i>ATF2</i>	3.90	2.29 10 ⁻²	<i>KIF3</i>	-3.57	3.29 10 ⁻⁹
<i>ZBED2</i>	3.89	5.34 10 ⁻¹	<i>S1PR1</i>	-3.62	5.05 10 ⁻¹⁰
<i>PRDM1</i>	3.87	7.71 10 ⁻³	<i>S1PR5</i>	-19.54	3.11 10 ⁻¹⁰
<i>TSC2</i>	3.77	2.20 10 ⁻²	Lung cancer T-cell signature		
<i>PRDM5</i>	3.53	1.08 10 ⁻²	<i>CTLA4</i>	3.45	3.63 10 ⁻⁹
<i>IKKKB</i>	3.40	2.84 10 ⁻²	<i>CXCR6</i>	3.29	3.89 10 ⁻⁹
<i>BAIF3</i>	3.03	2.86 10 ⁻²	<i>PDCD1</i>	2.43	5.61 10 ⁻¹⁰
<i>PRDM2</i>	2.72	4.35 10 ⁻²	<i>SIRPG</i>	2.43	8.78 10 ⁻⁴
<i>RUNX2</i>	2.50	3.70 10 ⁻²	<i>PTPN22</i>	1.98	2.52 10 ⁻²
<i>RUNX3</i>	2.24	1.95 10 ⁻³	<i>CD2</i>	1.85	7.75 10 ⁻⁴
<i>E2F2</i>	2.13	2.45 10 ⁻²	<i>SIFI</i>	1.70	2.32 10 ⁻³
<i>IKZF3</i>	2.01	1.54 10 ⁻³	<i>TIGIT</i>	1.71	1.41 10 ⁻²
<i>BAIF</i>	1.52	4.20 10 ⁻²	<i>UBASH3A</i>	1.60	4.88 10 ⁻³
<i>E2F3</i>	-1.60	2.35 10 ⁻²	<i>GPRI74</i>	1.59	6.17 10 ⁻³
<i>IKZF2</i>	-1.63	2.06 10 ⁻²	<i>CD96</i>	1.56	1.82 10 ⁻²
<i>BCL6</i>	-1.93	2.13 10 ⁻²	<i>SLAMF1</i>	1.51	6.32 10 ⁻²
<i>EOMES</i>	-2.38	1.34 10 ⁻³	<i>SLA2</i>	1.49	8.64 10 ⁻³
<i>KIF2</i>	-2.58	8.57 10 ⁻⁴	<i>CCR5</i>	1.45	5.51 10 ⁻²
<i>KIF3</i>	-3.60	1.26 10 ⁻³	<i>P2RY10</i>	1.45	6.20 10 ⁻²
<i>TCF7</i>	-4.03	1.39 10 ⁻²	<i>CCL5</i>	1.42	5.59 10 ⁻²
Adhesion/migration			<i>LCK</i>	1.41	2.99 10 ⁻²
<i>ITGAE</i>	31.99	1.37 10 ⁻¹⁰⁴	<i>SASB</i>	1.37	7.11 10 ⁻²
<i>CXCL13</i>	18.40	1.28 10 ⁻¹⁰	<i>TBC1D10C</i>	1.37	9.11 10 ⁻²
<i>ITGA7</i>	8.16	2.30 10 ⁻³	<i>IL2RG</i>	1.33	2.67 10 ⁻²
<i>KIRC1</i>	8.10	1.81 10 ⁻⁴	<i>IL2RB1</i>	1.25	7.00 10 ⁻²
<i>CCR5</i>	6.41	1.90 10 ⁻²	<i>CST7</i>	-1.54	2.73 10 ⁻²
<i>ITGA1</i>	5.96	6.27 10 ⁻³			

Table S9: Hallmark gene sets enriched in T_{RM} cells (Related to Figure 4)

Hallmark gene set	NES	p-value	Core enrichment
TNFA_SIGNALING_VIA_NFKB	-0,95	0,54	ZBTB10,SQSTM1,CLCF1,DENND5A,KLF2,CFLAR,IFNGR2,ICAM1,ACKR3,PLEK,NINJ1,SOD2,SLC2A6
PEROXISOME	-1,35	0,11	LONP2,SOD2,IDE,TSP0,SLC23A2
TGF_BETA_SIGNALING	1,05	0,42	NCOR2,SMURF2,CTNNB1
PI3K_AKT_MTOR_SIGNALING	1,08	0,39	CAMK4,PIK3R3,TSC2,MKNK1,RPS6KA1,CDK1,MYD88,AP2M1,MAPKAP1,CDK1,ACTR2,FASLG,ARPC3,MKNK1,RAF1,RPS6KA3,PIN1,CAB39,RPTOR
MYC_TARGETS_V2	1,13	0,32	LAS1L,UNG,PES1,MCM5,TBRG4,SLC19A1,PLK4,MCM4,MCM5,IMP4
IL6_JAK_STAT3_SIGNALING	1,14	0,29	CXCL13,CD9,IL4R,CSF2,MYD88,CSF1,IRF9,CSF1,IL17RA,ITGA4,EBI3,FAS,IL6ST,
INFLAMMATORY_RESPONSE	1,15	0,27	AB11,CD82,SRI,P2RX4,SRI,P2RX7,NLRP3,IRF7,ATP2C1,IL4R,LTA,SEMA4D
FATTY_ACID_METABOLISM	1,17	0,28	DLST,GCDH,ECI2,PPARA,ACOX1,AUH,ECI2,SDHC,CRYZ,HPGD,D2HGDH,GSTZ1,HIBCH,ECI2
INTERFERON_GAMMA_RESPONSE	1,18	0,25	P1PN6,DDX58,SRI,TRIM25,VCAM1,IRF7,IL4R,NCOA3,GPR18,RNF31,IRF5,TRAFD1,RNF31,ITGB7,MYD88,ST3GAL5,RSAD2,IFI35,IFNAR2,XAF1,IRF9
WNT_BETA_CATENIN_SIGNALING	1,39	0,11	NCOR2,RBPJ,PPARD,CTNNB1,NCSTN,JAG2,NCSTN,NUMB
OXIDATIVE_PHOSPHORYLATION	1,43	0,07	SLC25A12,ATP6V1H,DLST,COX4I1,NDUFV1,BAX,IMMT,MGST3,VDAC3,NNT,MAOB,ATP5A1,PHYH,SLC25A11,ATP6V0B,RHOT1,SDHC,ATP5G1,ATP5G2, HTRA2,MGST3,ATP5J,ATP5L
DNA_REPAIR	1,59	0,02	RNMT,NME3,LIG1,TAF6,NT5C,ERCC2,POLA2,ZWINT,TSG101,RAD51,ERCC2,ZWINT,POLD1,ITPA,PRIMI,POLD3,POLR1C,CANT1,POLD2,GTF2H3
MTORC_SIGNALING	1,67	0,01	BUB1,QDPR,AURKA,SYTL2,SYTL2,CDC25A,PIK3R3,ELOVL6,SHMT2,GSR,CD9,VLDLR,IMMT,DHFR,UNG,RRM2,SYTL2,ASNS,AURKA,UCHL5,EPRS,CCT6A,TFRC,AK4,SYTL2,PFKL,ATP5G1,PSMA4,SYTL2,TFRC,EBP,TM7SF2,NFYC,ACTR2,P4HA1,NAMPT,MTHFD2,FDXR,MCM4,G6PD,GBE1,SYTL2,MCM4,STIP1,NAMPT,HMBS,SYTL2
PROTEIN_SECRETION	1,83	0,01	STX16,ATP6V1H,SEC31A,CD63,TSG101,PAM,ARFIP1,VPS45,RAB2A,PAM,AP2M1,GOLGA4,COPB1,ICA1,SGMS1,CLTA,GOSR2,SGMS1,SNAP23,TPD52
IL2_STAT5_SIGNALING	1,89	0,001	ITGAE,CAPG,NCST1,BCL2L1,HOPX,ICOS,TNFRSF8,MYO1E,P2RX4,SNX14,CTLA4,IKZF4,RABGAP1L,IL4R,NCOA3,FLT3LG,FAH,PHTF2,MYO1E,CSF2,TNFRSF8, ST3GAL4,PLAGL1,PLEC,MYO1C
E2F_TARGETS	2,09	0,001	ILF3,BUB1B,MELK,LIG1,BIRC5,DLGAP5,AURKA,CDC25A,CIT,POLA2,ESPL1,MELK,WDR90,AURKB,MCM3,RAD51C,BRCA1,HMMR,SMC4,POLD1,UNG,POLD2,RRM2,CDK1,POLD3,STAG1,MKI67,RAD51AP1,PSMC3IP,AURKA,CDKN2A,SMC4,BRCA2,CDKN3,WEE1,CENPE,AURKB,GINS3,TFRC,EIF2S1,NUP107,MYBL2,AURKB,TMPO,RACGAP1,MCM5,TBRG4,CENPM,KIF2C,TFRC,CDC20,LIG1,ORC6,CDK1,ATAD2,CCNE1,CDCA3,EZH2,RFC2,DEPDC1,SMC4,SPAG5,CIT,CDCA3,HELLS,SMC4,STAG1,MTHFD2,RAD51AP1,POLE,TCF19,CKS1B,PRKDC,PLK4,MCM4,DNMT1,SMC6,CDKN2A,TRIP13,POLD3,DEK,EXOSC8,PLK4,KPNA2,RAD51C,BARD1
G2M_CHECKPOINT	2,38	0,001	ILF3,MTHFD2,SMC2,BUB1,PRC1,BIRC5,TPX2,AURKA,CDC25A,NUSAP1,KIF15,EXO1,TROAP,POLA2,ESPL1,NUSAP1,AURKB,MCM3,KIF23,HMMR,PRC1,SMC4,SMC2,CBX1,MLE3,STIL,TNPO2,CDK1,STAG1,CHMP1A,MKI67,KIF23,AURKA,NUSAP1,TRAIP,BRCA2,EXO1,TTK,SS18,SS18,CDKN3,ODF2,CENPE,CDK7,AURKB,DKC1,CASP8AP2,PBK,CDC27,TROAP,MYBL2,AURKB,EWSR1,TRAIP,KIF23,TMPO,RACGAP1,MCM5,SMC2,TRAIP,KIF2C,SRSF10,CDC20, ORC6,PAFAH1B1,CDK1,RBM14,EZH2

NES: Normalized enrichment score

Table S10: Primer pair sequences for RT-PCR (Related to STAR methods)

IKZF3 Forward	AGATTTGAATGTGCCCTGGT
IKZF3 Reverse	TGTCTTGCCTCCGCACTT
RORC Forward	CTTGCCGTAGGGATGTCTCG
RORC Reverse	GAAGTTCCGTCAGCCCGTT
AHR Forward	TAACCCAGACCAGATTCCTCCAGA
AHR Reverse	CCCTTGGAAATTCATTGCCAGA
IL17A Forward	AACGATGACTCCTGGGAAGA
IL17A Reverse	GGATTTTCGTGGGATTGTGAT
TCF7 Forward	CGGGACAGGACCATTACAAGTAGATCAAGGAC
TCF7 Reverse	CCACCTGCCTCGGCCTGCCAAAGT
S1PR1 Forward	CAGACAAGCAAAACAAAGTG
S1PR1 Reverse	CATCAACAAAAGTGCCAAAG
18S Forward	CGGACAGGATTGACAGATTG
18S Reverse	CAATCGCTCCACCAACTAA