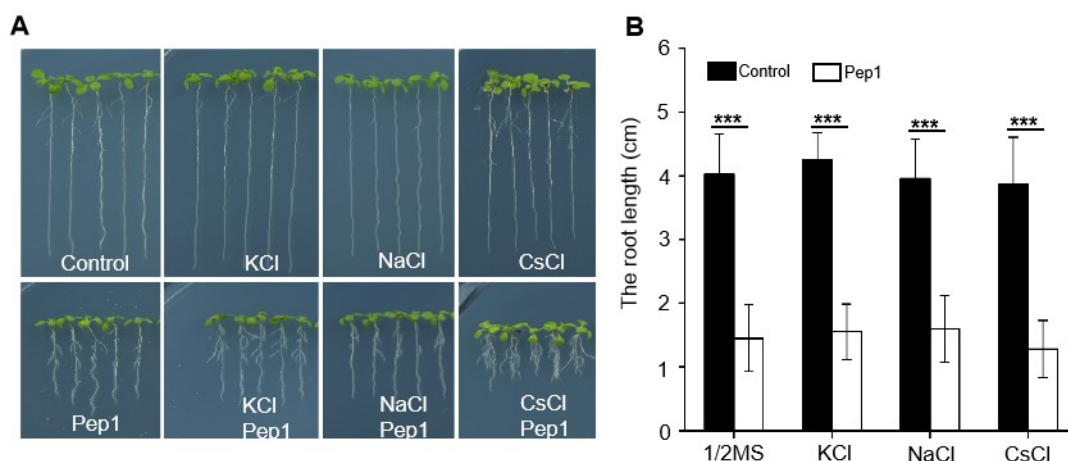
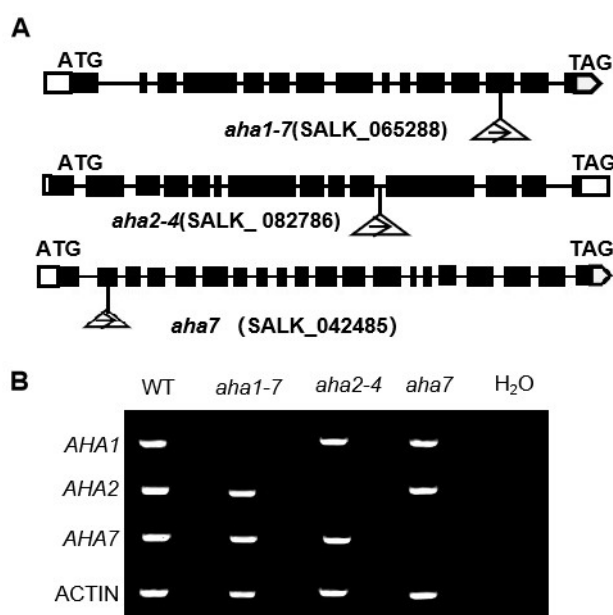


Supplementary data



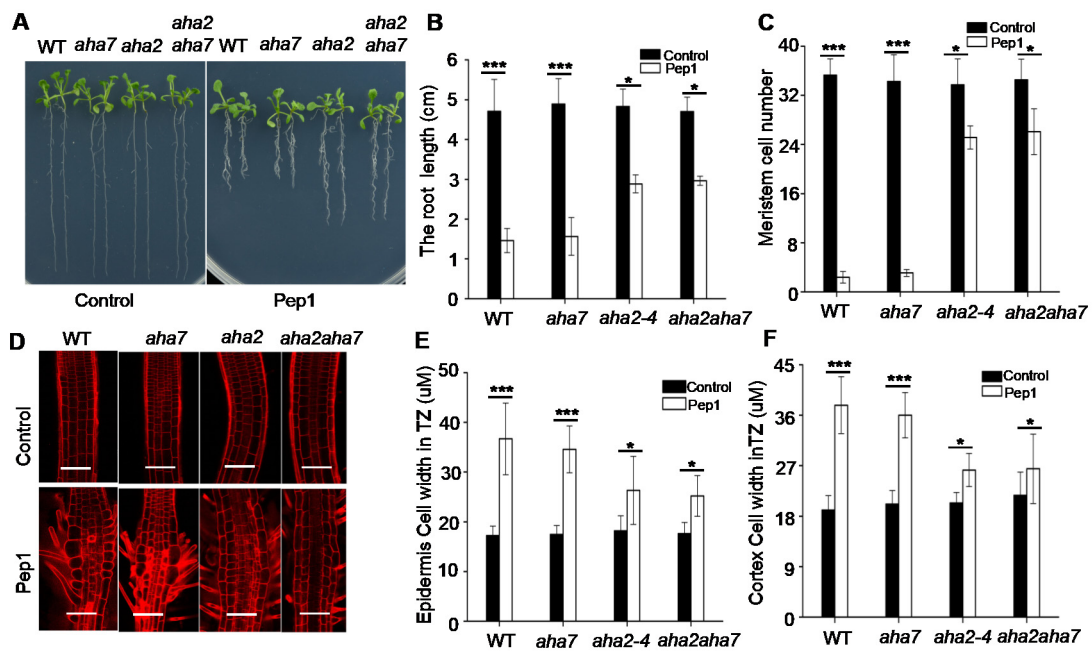
Supplementary figure 1. Pep1 inhibits the root growth independent on K^+ , Na^+ and Cs^+ cations.

(A) The growth phenotype of wild type (WT) root. Three-day-old seedlings were transferred onto half-strength MS medium supplemented with or without (Control) 50nM Pep1 in the presence or in the absence of 30mM KCl, 50mM NaCl or 2mM CsCl for 6 days. (B) The statistical analysis of the primary root length as indicated in (A). Data are means \pm SD from three replicate experiments (n=24 plants per treatment).



Supplementary figure 2. The identification of *aha1-7*, *aha2-4* and *aha7* mutants.

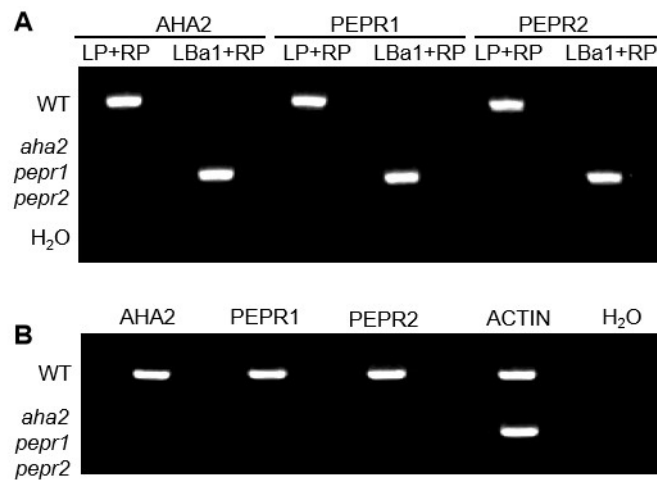
(A) Schematic map of T-DNA insertion locations of *aha1-7*, *aha2-4*, *aha7* mutants. Black boxes, lines and triangles represented exons, introns, and the position of the T-DNA insertion, respectively. The 5'- and 3'-UTRs are indicated by open boxes. ATG, translation start codon; TGA, stop codon. (B) RT-PCR detection of the transcriptional level of *AHA1*, *AHA2* and *AHA7* expression in wild type, *aha1-7*, *aha2-4*, and *aha7* mutants. Water was used as a negative control.



Supplementary figure 3. AHA2 does not function redundant with AHA7 to regulate the Pep1 signaling in root.

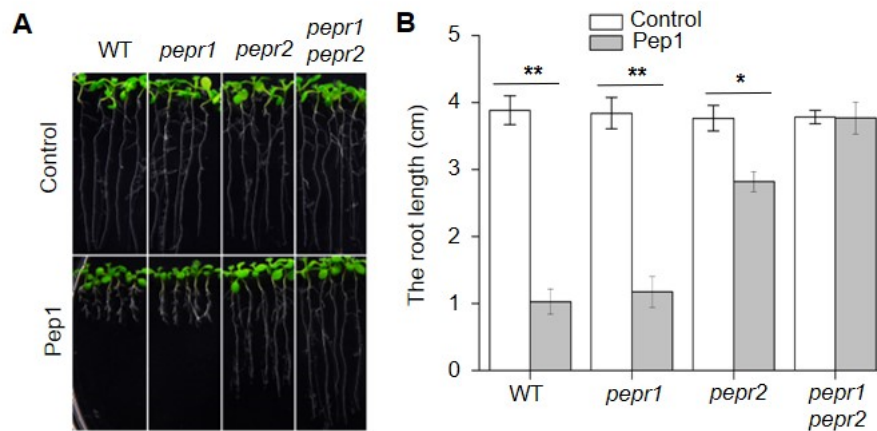
(A) The growth phenotype of wild type (WT), *aha7*, *aha2* and *aha2 aha7* roots. Three-day-old seedlings were transferred on half-strength MS agar medium supplemented with or without (Control) 50 nM Pep1 for 6 days. (B) The statistical analysis of the primary root length in WT, *aha7*, *aha2* and *aha2 aha7* plants as indicated (A). Data are means \pm SD from three independent experiments, (n=15). (C) Quantitative analysis of numbers of meristematic cortex cells as indicated in (A). Data are means \pm SD from three replicate experiments (n=24). (D) Longitudinal structures of roots in WT, *aha7*, *aha2* and *aha2 aha7* plants. 5-day-old seedlings were transferred on half-strength MS agar medium supplemented with or without (Control) 100nM Pep1 for 24h, the roots were stained with 5 μ M PI for 15s and photographed

under a confocal laser-scanning microscope. Similar results were obtained in three independent replicates. Bars=100um. **(E)** and **(F)** Quantitative analysis of epidermal and cortex cell width in TZ as indicated in **(D)**. Data are means \pm SD from three independent experiments (n = 32 cells from 8 roots per treatment). Asterisks in **(B)**, **(C)**, **(E)** and **(F)** indicate statistically significant differences (Tukey's test; *, P<0.05, **, P<0.01, ***, P<0.001).



Supplementary figure 4. The identification of *aha2 pepr1 pepr2* triple mutant.

(A) Confirmation of the T-DNA insertion in the *aha2 pepr1 pepr2* triple mutant by PCR. **(B)** RT-PCR detection of *AHA1*, *AHA2* and *AHA7* expression in wild type and *aha2 pepr1 pepr2* triple mutant. Water was used as a negative control.



Supplementary figure 5. PEPR2 primarily perceives the Pep1 signaling in roots.

(A) The growth phenotype of wild type (WT), *pepr1*, *pepr2* and *pepr1 pepr2* roots.

Three-day-old seedlings were transferred on half-strength MS agar medium supplemented with or without (Control) 50 nM Pep1 for 6 days. **(B)** The statistical analysis of the primary root length in WT, *pepr1*, *pepr2* and *pepr1 pepr2* plants as indicated **(A)**. Data are means \pm SD from three independent experiments, (n=15). Asterisks indicate statistically significant differences (Tukey's test; *, P<0.05, **, P<0.01).

Supplementary Table 1 Primers used in this study

Primer name	Primer sequences (5'-3')
(1) qRT-PCR analyze	
<i>Actin2</i> -F	CTGTTCTCTCCTTGTACGCCAGT
<i>Actin2</i> -R	CGGGTAATTCATAGTTCTTCTCGAT
<i>AHA1</i> -F	TTGAAGTTTGCCATTCGGTA
<i>AHA1</i> -R	GCCCATTGAGCTTCTCTTTC
<i>AHA2</i> -F	TTGTTGAACGTCCTGGAGCA
<i>AHA2</i> -R	AATTCCCAGTTGGCGTAAACC
<i>AHA7</i> -F	AGAAATAGCGCAACGGAACT
<i>AHA7</i> -R	TTGCAGCTGATTCAACCTTC
(2) BiFC analysis	
<i>AHA2</i> -BiFC-F	GACTAGTATGTCGAGTCTCGAAGATATCAAG
<i>AHA2</i> -BiFC-R	CGGGATCCCACAGTGTAG TGACTGGGAGT
<i>PEPR2</i> -BiFC-F	GACTAGTATGAGGAATC TTGGGTTACT CGA
<i>PEPR2</i> -BiFC-R	CCCCCGGGTGAAGTGA CCCGAAGTGC
(3) Yeast two-hybrid analysis	
<i>PEPR2</i> -F	GGAATTCCCGGTGCAAAAGAGAACCAAC
<i>PEPR2</i> -R	CGGGATCCAAATCTTTCACCACATCTCTCATC
<i>AHA2</i> -F	GGAATTCTTCAAGTTTGCCATTCGATACAT
<i>AHA2</i> -R	CGGGATCCCACATGTAGTGACTGGGAGTTTCA
(4) Identification of <i>aha1-7</i>, <i>aha2-4</i>, <i>aha7</i> mutant	
LBa1	TGGTTCACGTAGTGGGCCATCG
<i>aha1-7</i> -LP	ATGTCAGGTC TCGAAGATAT
<i>aha1-7</i> -RP	CTACACAGTG TAGTGATGTC
<i>aha2-4</i> -LP	ACCTCTGGCTCAAAATTGTCC
<i>aha2-4</i> -RP	CTCCAGGACGTTCAACAAAAG
<i>aha7</i> -LP	ATGACGGACATAGAAGCTCTC
<i>aha7</i> -RP	TTCGCTTTAG GGGCTAACTG
<i>pepr1</i> -LP	ACATCAGACGGACGTAAAACG
<i>pepr1</i> -RP	TGCAATTAGGTGATCCGAAAC
<i>pepr2</i> -LP	TCCAATGTGAGGCTCCATATC
<i>pepr2</i> -RP	TTCTCAAAACAAACTCACGGG