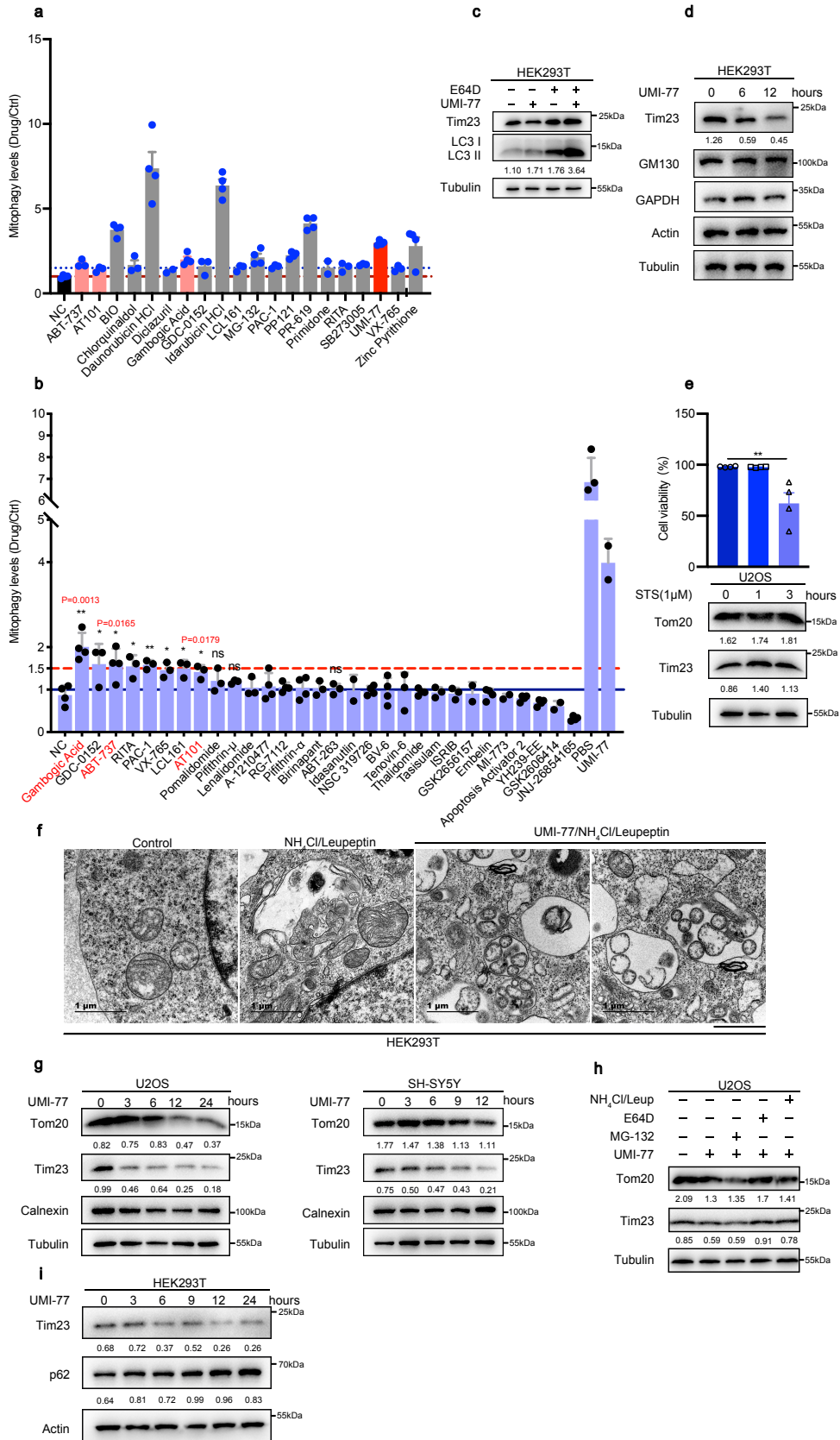
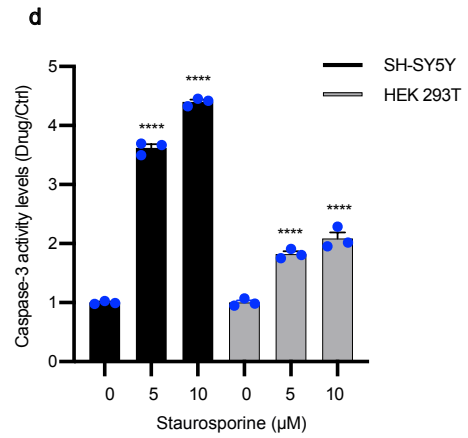
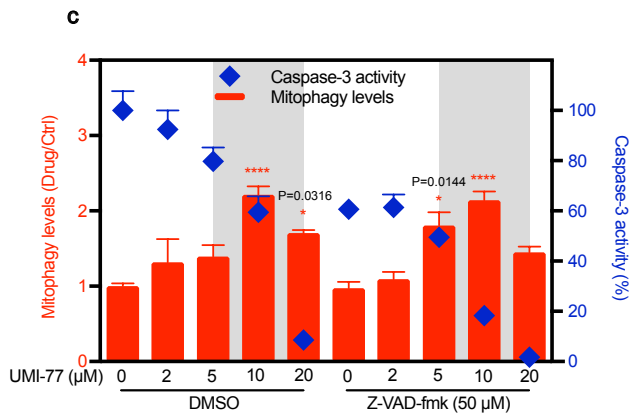
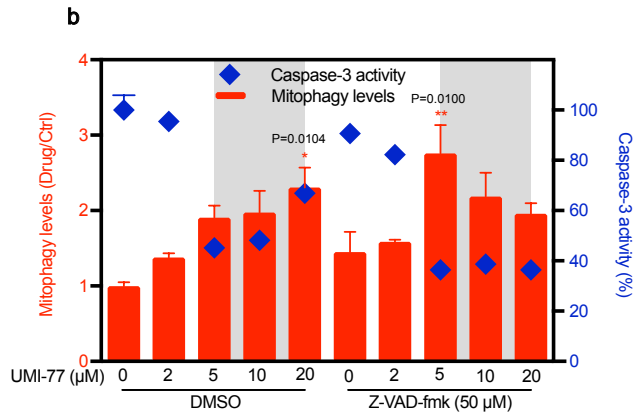
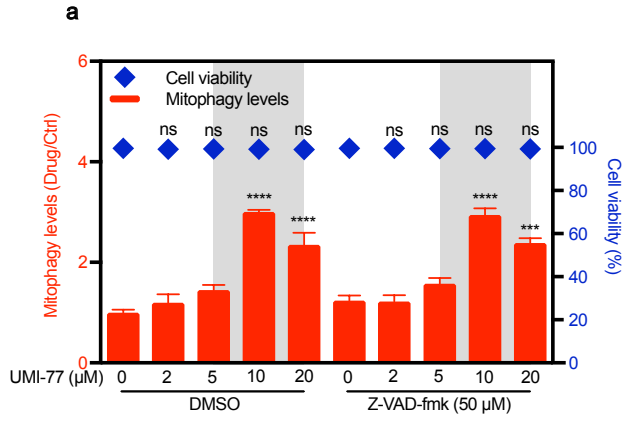


**Pharmacological targeting of MCL-1 promotes mitophagy and improves disease pathologies in an Alzheimer's disease mouse model**



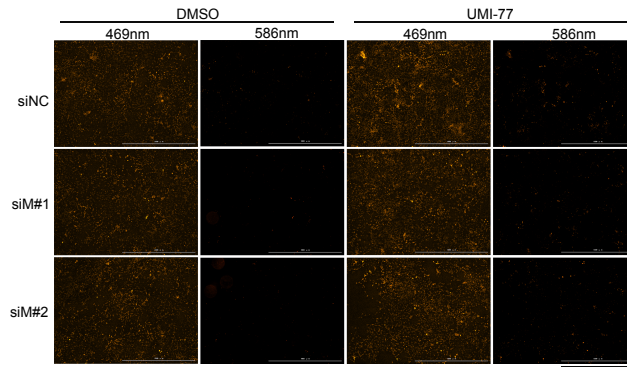
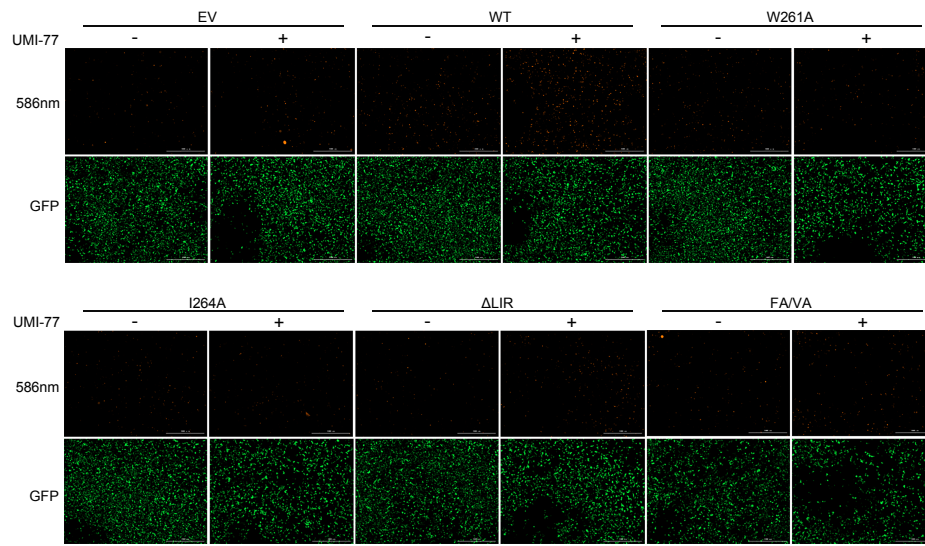
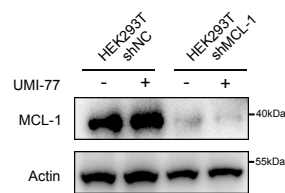
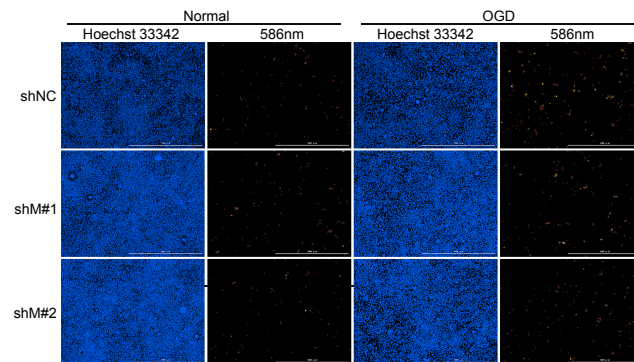
**Supplementary Fig. 1. UMI-77 induces mitochondrial degradation through the lysosomal pathway.**

**a** Twenty positive compounds were re-validated in HEK293T-mt-Keima cells. Bcl-2 family protein inhibitors were indicated in red. **b** Pro-apoptotic compounds were re-validated in HEK293T mt-Keima cells, with three replicates per drug. The mitophagy levels were analyzed using two-tailed t-test (data represent mean  $\pm$  S.E.M.; n=3. \*\*\* p<0.001, \*\* p<0.01, \* p<0.05). PBS was used as a positive control to induce autophagy. **c** HEK293T cells were treated with UMI-77 and E64D, cell lysates were immunoblotted with indicated antibodies. The numbers under the blots represent the gray scale quantification (LC3II/Tubulin). **d** HEK293T cells were treated with 5  $\mu$ M UMI-77 for the indicated times and cell lysates were immunoblotted with indicated antibodies. The numbers under the blots represent the gray scale quantification (Tim23/Tubulin). **e** U2OS cells were treated with 1  $\mu$ M staurosporine (STS) for the indicated times, cell lysates were immunoblotted with indicated antibodies and cell viability were estimated by using LIVE/DEAD™ cell imaging kit. Data were analyzed by one-way ANOVA (data represent mean  $\pm$  S.E.M.; n=4. \*\* p<0.01 (P=0.0062)). The numbers under the blots represent the gray scale quantification (Tom20/Tubulin, Tim23/Tubulin). **f** HEK293T cells were treated with UMI-77 (5  $\mu$ M) for 24 h, treated with NH<sub>4</sub>Cl (20 mM) and Leupeptin (100nM) for 12 h, and the analysis by electron microscopy was performed. Scale bars, 1  $\mu$ m. **g** U2OS and SH-SY5Y cells were treated with 5  $\mu$ M UMI-77 for the indicated times and cell lysates were immunoblotted with indicated antibodies. The numbers under the blots represent the gray scale quantification (Tom20/Tubulin, Tim23/Tubulin). **h** U2OS cells were treated with 5  $\mu$ M UMI-77 in the presence or absence of MG-132, E64D and NH<sub>4</sub>Cl/Leupeptin (Leup) for 12 h, and cell lysates were immunoblotted with indicated antibodies. The numbers under the blots represent the gray scale quantification (Tom20/Tubulin, Tim23/Tubulin). **i** HEK293T cells were treated with 5  $\mu$ M UMI-77 for the indicated times, and cell lysates were immunoblotted with indicated antibodies. The numbers under the blots represent the gray scale quantification (Tim23/Tubulin, p62/Tubulin). Source data are provided as a Source Data file.



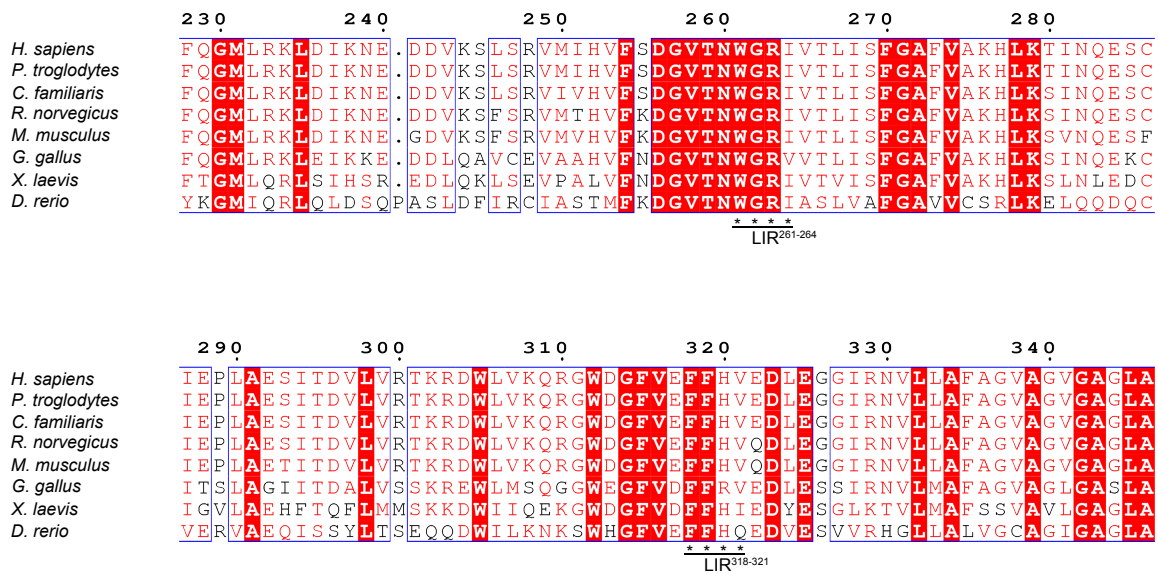
**Supplementary Fig. 2. UMI-77 induces mitophagy independent of apoptosis.**

**a** SH-SY5Y cells were transfected with pcDNA3.1-mt-Keima plasmid for 24 h and treated with UMI-77 (0  $\mu$ M, 2  $\mu$ M, 5  $\mu$ M, 10  $\mu$ M, 20  $\mu$ M) with or without Z-VAD-fmk (50  $\mu$ M) for 12 h. Cell viability was estimated by using LIVE/DEAD™ cell imaging kit. One-way ANOVA (mean  $\pm$  S.E.M.; n=4. \*\*\*\* p<0.0001, \*\*\* p<0.001. ns, not significant). **b** HEK293T cells were transfected with pcDNA3.1-mt-Keima plasmid for 24 h and treated with UMI-77 (0  $\mu$ M, 2  $\mu$ M, 5  $\mu$ M, 10  $\mu$ M, 20  $\mu$ M) with or without Z-VAD-fmk (50  $\mu$ M) for 24 h. Apoptosis levels was estimated using Caspase-Glo® 3/7 Assay. One-way ANOVA (data represent mean  $\pm$  S.E.M.; n=4. \*\* p<0.01, \* p<0.05). **c** As in (**b**), except SH-SY5Y cells were used. Apoptosis levels was estimated using Caspase-Glo® 3/7 Assay. One-way ANOVA (data represent mean  $\pm$  S.E.M.; n=4. \*\*\*\* p<0.0001, \* p<0.05). **d** Cells were treated with staurosporine (0  $\mu$ M, 5  $\mu$ M, 10  $\mu$ M) for 3 h. Apoptosis levels were estimated as in (**b**). One-way ANOVA (data represent mean  $\pm$  S.E.M.; n=4. \*\*\*\* p<0.0001). Source data are provided as a Source Data file.

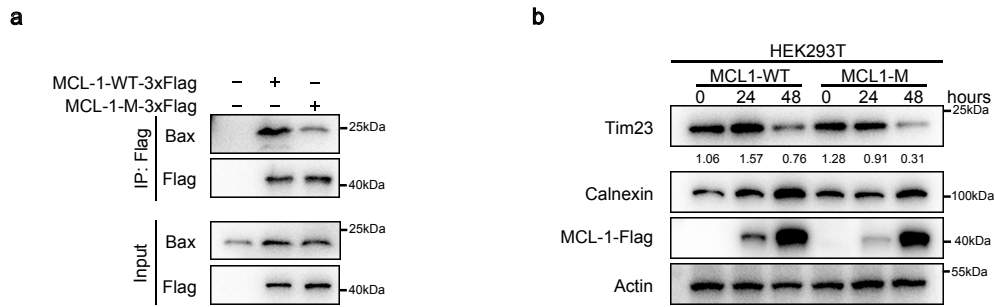
**a****b****c****d**

**Supplementary Fig. 3. Representative images of Figure 2b, 3g, 5a.**

**a** Representative image of Figure 2b. 469nm shows mitochondria and 586nm shows mitochondria in lysosome. **b** Representative image of Figure 3g. GFP shows MCL-1 knockdown cells and 586nm shows mitochondria in lysosome. **c** HEK293T-MCL-1-knockdown (HEK293T-shMCL-1) and HEK293T-control-knockdown cells (HEK293T-shNC) were treated with UMI-77 as Figure 3g shown and cell lysates were immunoblotted with indicated antibodies. Source data are provided as a Source Data file. **d** Representative image of Figure 5a. Hoechst 33342 shows nucleus and 586nm shows mitochondria in lysosome. OGD: oxygen-glucose deprivation.

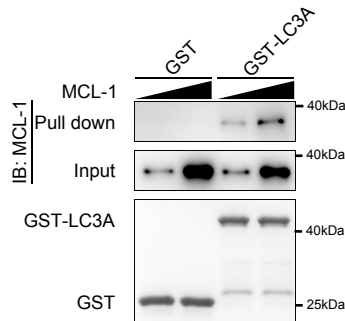


**Supplementary Fig. 4. The LIR<sup>261-264</sup> motif of MCL-1 is strongly conserved.** MCL-1 amino acid sequences were obtained from NCBI and aligned using clustalW software (<http://www.genome.jp/tools-bin/clustalw>). The image was generated using ESPrir 3.0 (<http://esprir.ibcp.fr/ESPrir/cgi-bin/ESPrir.cgi>).



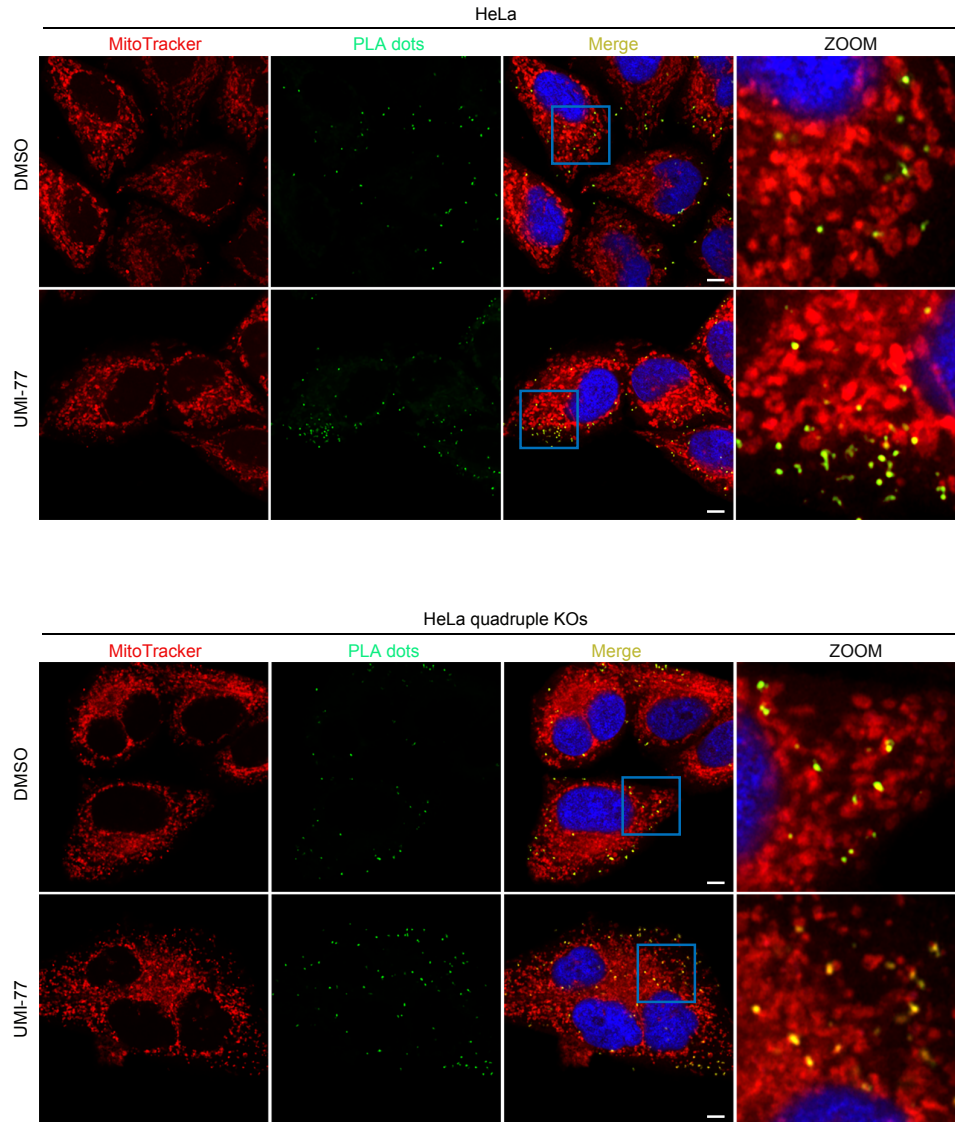
**Supplementary Fig. 5. The L213A/D218A mutant of MCL-1 promotes mitophagy.**

**a** HEK293T cells were transfected with MCL-1-WT-3xFlag or MCL-1-M-3xFlag (L213A/D218A) for 24 h, and the interaction between MCL-1 and Bax was analyzed by immunoprecipitation. **b** HEK293T cells were transfected with pcDNA3.1-MCL-1-WT (wild-type) or pcDNA3.1-MCL-1-M (L213A/D218A) plasmid for the indicated times and cell lysates were immunoblotted with indicated antibodies. The numbers under the blots represent the gray scale quantification (Tim23/Tubulin). Source data are provided as a Source Data file.

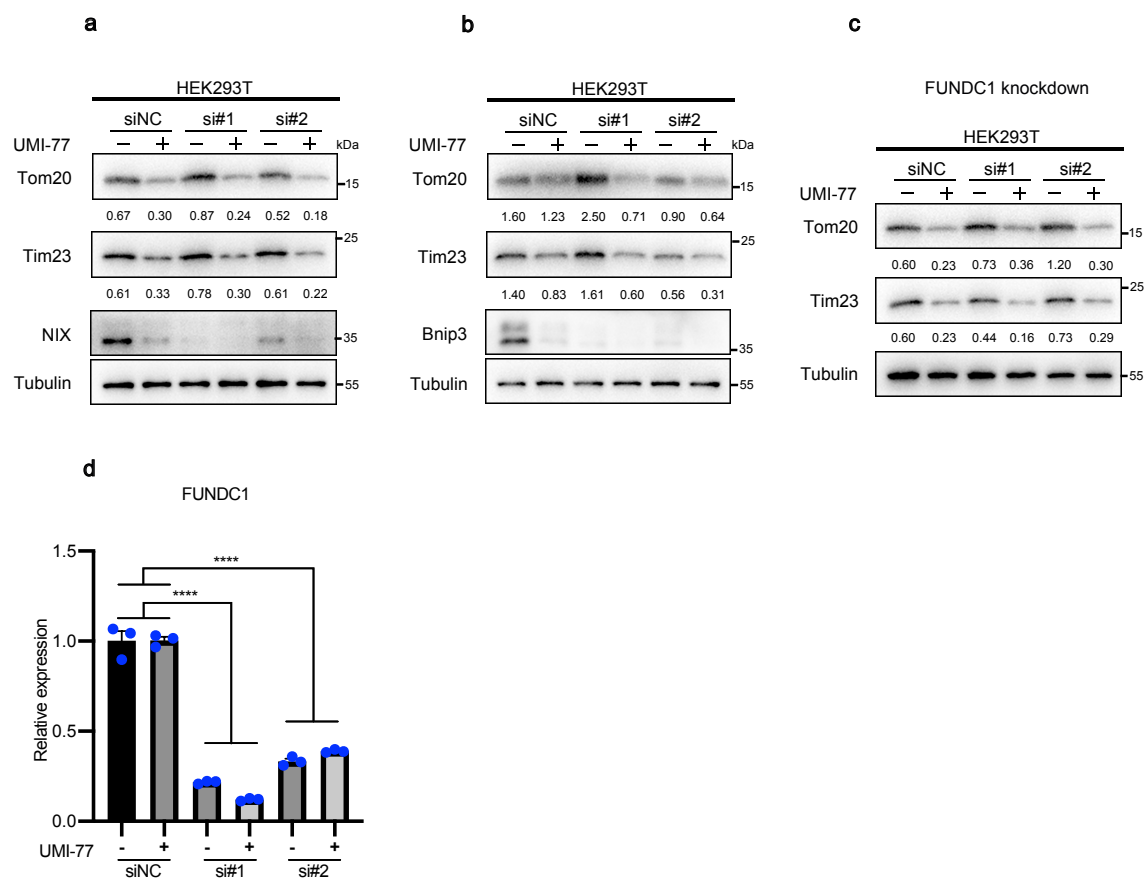


**Supplementary Fig. 6. MCL-1 directly interacts with LC3A.** Top two panels: Indicated purified proteins were incubated *in vitro* for two hours and GST pull-down was performed and analyzed by western blotting. Bottom panel: Coomassie staining was used to visualize GST-LC3A and GST proteins in the pull-down samples. Source data are provided as a Source Data file.





**Supplementary Fig. 7. MCL-1 interacts with LC3A on mitochondria.** Wild-type and quadruple KOs (NBR1, TAX1BP1, p62, and NDP52 knockout) HeLa cells were treated with UMI-77 (5  $\mu$ M) for 3h and a PLA assay for MCL-1 and LC3A was performed. Cells were counter-stained with MitoTracker™ Deep Red FM. Scale bar, 5 $\mu$ m. Source data are provided as a Source Data file.



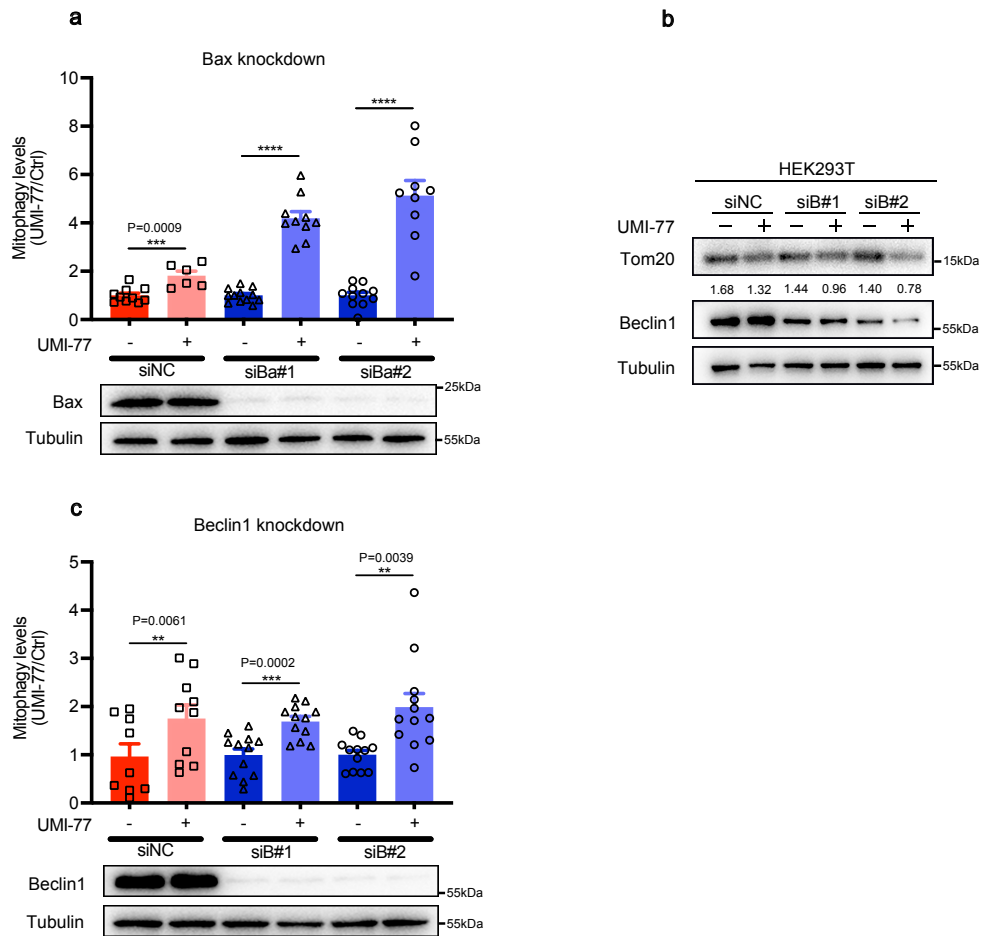
**Supplementary Fig. 8. UMI-77 induces mitophagy independent of BNIP3, NIX, FUNDC1.**

**a** HEK293T cells were transfected with NIX siRNA for 48 h and treated with 5  $\mu$ M UMI-77 for 12 h. Cell lysates were immunoblotted for mitochondrial marker proteins (Tom20, Tim23). The numbers under the blots represent the gray scale quantification (Tom20/Tubulin, Tim23/Tubulin). siNC: scrambled siRNA.

**b** As in (a), except Bnip3 siRNA was used.

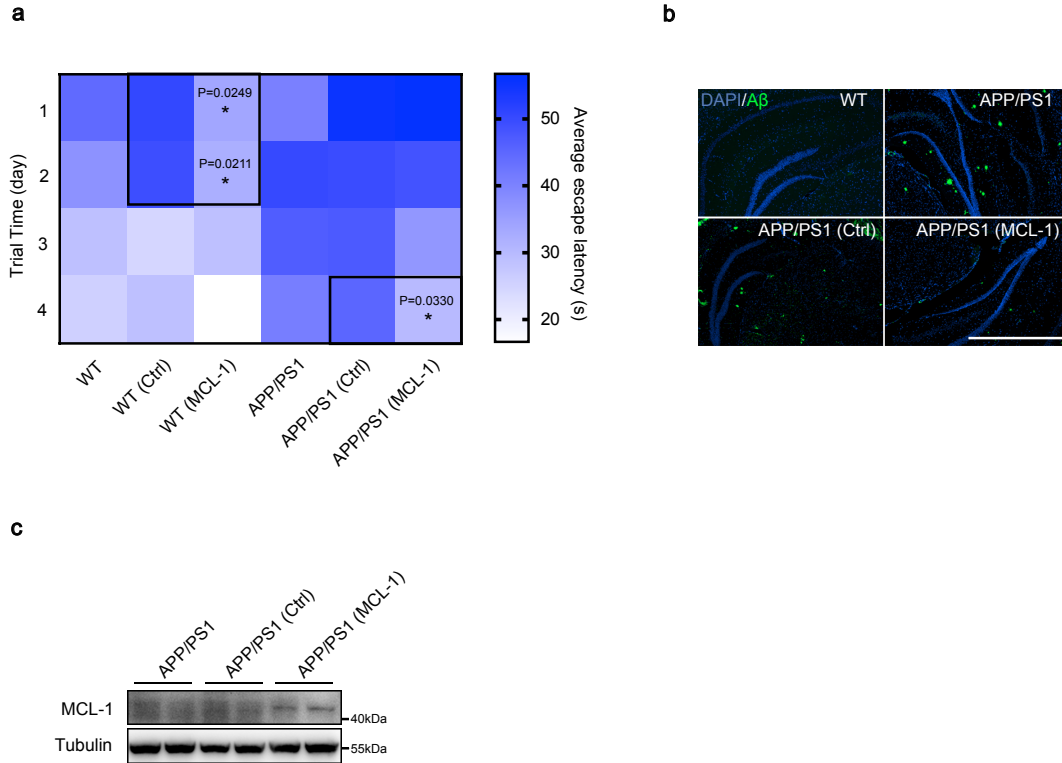
**c** As in (a), except FUNDC1 siRNA was used.

**d** Cells were treated as in (c) and total RNA were extracted by FastPure® Cell/Tissue Total RNA Isolation Kit for FUNDC1 qPCR. Data were analyzed using one-way ANOVA (data represent mean  $\pm$  S.E.M.; n=3. \*\*\*\* p<0.0001). siNC: scrambled siRNA. Source data are provided as a Source Data file.



**Supplementary Fig. 9. UMI-77 induces mitophagy independent of Bax and Beclin1.**

**a** Bax knockdown HEK293T-mt-Keima cells were treated with 5  $\mu$ M UMI-77 for 12 h. The mitophagy levels were analyzed using two-tailed t-test (data represent mean  $\pm$  S.E.M.; The sample size was, in turn, n=10, n=6, n=12, n=12, n=10, n=11, n=9. \*\*\* p<0.001, \*\*\*\* p<0.0001.). The siRNA knockdown efficiency was shown using western blot. siNC: scrambled siRNA. **b** HEK293T cells were transfected with Beclin1 siRNA for 48 h and treated with 5  $\mu$ M UMI-77 for 12 h. Cell lysates were immunoblotted for mitochondrial marker proteins (Tom20). The numbers under the blots represent the gray scale quantification (Tom20/Tubulin). siNC: scrambled siRNA. **c** Beclin1 knockdown HEK293T-mt-Keima cells were treated with 5  $\mu$ M UMI-77 for 12 h. The mitophagy levels were analyzed using two-tailed t-test (data represent mean  $\pm$  S.E.M.; n=11. \*\* p<0.01, \*\*\* p<0.001.). The siRNA knockdown efficiency was shown using western blotting. siNC: scrambled siRNA. Source data are provided as a Source Data file.



**Supplementary Fig. 10. Overexpression of MCL-1 ameliorates cognitive decline in the APP/PS1 mouse model of Alzheimer's disease.**

**a** Six-month-old C57BL/6 and APP/PS1 mice were overexpressed MCL-1 in hippocampus for one month. Latency to escape to a hidden platform in the Morris water maze during a 4-day training period ((WT, n=6), (WT (Ctrl vector), n=5), (WT (MCL-1 overexpression), n=6), (APP/PS1, n=6), (APP/PS1 (Ctrl vector), n=6), (APP/PS1 (MCL-1 overexpression), n=5). \*  $p < 0.05$ , two-tailed t-test). **b** Mice were treated as in (a) and immunohistochemistry of hippocampus was performed to stain for amyloid plaques (6E10 antibody, green) and nuclei (DAPI, blue). Scale bar, 1000 $\mu$ m. **c** Mice were treated as in (a) and brain lysates were immunoblotted for MCL-1 and Tubulin. Source data are provided as a Source Data file.

**Supplementary Table 1.**

Primers used in this study.

Primers for pCDH-mt-Keima	
pCDH-mt-Keima-F	5'-CCGGAATTCGAAATGCTGAGCCTGCGCCAGAG-3'
pCDH-mt-Keima-R	5'-CGCGGATCCTCAACCGAGCAAAGAGTGGC-3'
Primers for MCL-1 mutant	
W261A-F	5'-TCAGCGACGGCGTAACAAACGCGGGCAGGATTGTGACTCTCAT-3'
W261A-R	5'-ATGAGAGTCACAATCCTGCCC GCGTTTGTTACGCCGTCGCTGA-3'
I264A-F	5'-GCGTAACAACTGGGGCAGGGCTGTGACTCTCATTCTTTTGG-3'
I264A-R	5'-CCAAAAGAAATGAGAGTCACAGCCCTGCCCCAGTTTGTTACGC-3'
W261A/I264A-F	5'-TCAGCGACGGCGTAACAAACGCGGGCAGGGCTGTGACTCTCATTCTTTTGG-3'
W261A/I264A-R	5'-CCAAAAGAAATGAGAGTCACAGCCCTGCCC GCGTTTGTTACGCCGTCGCTGA-3'
ΔLIR-F	5'-TCAGCGACGGCGTAACAAACGTGACTCTCATTCTTTTGG-3'
ΔLIR-R	5'-CCAAAAGAAATGAGAGTCACGTTTGTTACGCCGTCGCTGA-3'
F318A/V321A-F	5'-GCTGGGATGGGTTTGTGGAGGCCTTCCATGCAGAGGACCTAGAAGGTGGCAT-3'
F318A/V321A-R	5'-ATGCCACCTTCTAGGTCCTCTGCATGGAAGGCCTCCACAAACCCATCCCAGC-3'
Primers for GST pull down	
pET28a-mcl1-F	5'-TAGAAGCTTGCGGCCGCACT-3'
pET28a-mcl1-R	5'-TTCTAGGTCCTCTACATGGA-3'
GST-LC3A-F	5'-CGCGGATCCGATGCCCTCAGACCGGCCTTT-3'
GST-LC3A-R	5'-CCGCTCGAGTCAGAAGCCGAAGTTTCCT-3'