Supplementary Figures and Tables for Mukherjee et al., *Molecular estimation of neurodegeneration pseudotime in older brains.*

Supplemental Figures

Supplementary Figure 1 - Comparison between trajectories inferred using different gene sub-set selection methods: i) Differential Expression with an FDR cut-off of 0.1, ii) High variance gene selection.





Tissue.Diagnosis • TCX.AD • TCX.CONTROL • TCX.OTHER



Tissue.Diagnosis • TCX.AD • TCX.CONTROL • TCX.OTHER



Supplementary Figure 2 – Trajectories with DE genes at FDR p-value <= 0.01 in TCX and DLFPC brain regions.



Supplementary Figure 3 – Patient trajectory maps for TCX data by adjustment A) Adjusted for PMI, B) Adjusted for first 10 PCs, C) Adjusted for RIN, D) Adjusted for RIN, PMI, and first 10 PCs.



Supplementary Figure 4 – Patient trajectory maps for DLPFC data by adjustment A) Adjusted for PMI, B) Adjusted for first 10 PCs, C) Adjusted for RIN, D) Adjusted for RIN, PMI, and first 10 PCs.



Supplementary Figure 5 – Absolute value of Pearson correlations between pseudotime estimated with all samples in both ROSMAP (DLPFC) and Mayo RNAseq (TCX), and pseudotime estimated with leave one out data-sets.



Supplementary Figure 6 – Pseudotime by AD case-control status for 218 independent samples from two independent studies. A) Adjusted for PMI, B) Adjusted for first 10 PCs, C) Adjusted for RIN, D) Adjusted for PMI, RIN, and first 10 PCs. Box plots have lower and upper hinges at the 25th and 75th percentiles and whiskers extending to at most 1.5xIQR (interquartile range).



Supplementary Figure 7 – Lineage inference in the Mayo eGWAS expression array data-set for 186 independent samples from one independent study. A) Monocle2 inferred manifold, B) disease state as a function of disease pseudotime, C) APOE e4 dosage as a function of disease pseudotime, D) resistant individuals on the disease pseudotime manifold. Box plots have lower and upper hinges at the 25th and 75th percentiles and whiskers extending to at most 1.5xIQR (interquartile range).



Supplementary Figure 8 - Manifold learning infers disease states from RNA-seq samples, samples from males only for 143 independent samples from two independent studies. A) Estimated cell trajectory from A) TCX and B) DLPFC. C) Distribution of pseudotime for AD cases and controls for DLPFC and TCX. D) Distribution of expression correlation with pseudotime for both LOAD GWAS genes and non-LOAD GWAS genes. Box plots have lower and upper hinges at the 25th and 75th percentiles and whiskers extending to at most 1.5xIQR (interquartile range).



Supplementary Figure 9 - Manifold learning and measures of staging in LOAD in DLPFC samples, males only for 143 independent samples from two independent studies. A-C) Samples colored by three external measures of LOAD staging: Braak score, CERAD score, and cognitive diagnosis. D-F) Distribution of samples by inferred stage for different distinct stages in each of the three methods of measuring LOAD severity. Box plots have lower and upper hinges at the 25th and 75th percentiles and whiskers extending to at most 1.5xIQR (interquartile range).



Supplementary Figure 10 – Manifold learning infers disease states from RNA-seq samples, samples from males and females combined for 361 independent samples from two independent studies. A) Estimated cell trajectory from A) TCX and B) DLPFC. C) Distribution of pseudotime for AD cases and controls for DLPFC and TCX. D) Distribution of expression correlation with pseudotime for both LOAD GWAS genes and non-LOAD GWAS genes. Box plots have lower and upper hinges at the 25th and 75th percentiles and whiskers extending to at most 1.5xIQR (interquartile range).



Supplementary Figure 11 – Manifold learning and measures of staging in LOAD in DLPFC samples, male and females combined for 537 independent samples from one study. A-C) Samples colored by three external measures of LOAD staging: Braak score, CERAD score, and cognitive diagnosis. D-F) Distribution of samples by inferred stage for different distinct stages in each of the three methods of measuring LOAD severity. Box plots have lower and upper hinges at the 25th and 75th percentiles and whiskers extending to at most 1.5xIQR (interquartile range).



Supplementary Figure 12 - Correlations with pseudotime for IGAP GWAS genes for 17446 genes from two independent studies for A) PMI adjusted pseudotimes, B) top 10 PC adjusted pseudotime, C) RIN adjusted pseudotimes, and D) PMI, PC, and RIN adjusted pseudotimes. Box plots have lower and upper hinges at the 25th and 75th percentiles and whiskers extending to at most 1.5xIQR (interquartile range).



Supplementary Figure 13 – Lineages analyses adjusted for Braak score in DLPFC for 338 independent samples from one study. A) Lineage adjusted for Braak, B) Diagnosis as a function of Braak adjusted pseudotime, C) Cognitive diagnosis on Braak adjusted lineage, D) Cognitive diagnosis as a function of Braak adjusted pseudotime, E) correlation between IGAP GWAS genes and Braak adjusted pseudotime for 17446 genes from one study. Box plots have lower and upper hinges at the 25th and 75th percentiles and whiskers extending to at most 1.5xIQR (interquartile range).



Supplementary Figure 14 - Manifold learning and measures of staging in LOAD in TCX samples for 76 independent samples from one study. A) Samples colored by two external measures of LOAD staging: Braak score and Thal amyloid. B) Distribution of samples by inferred stage for different distinct stages in each of the two methods of measuring LOAD severity. Box plots have lower and upper hinges at the 25th and 75th percentiles and whiskers extending to at most 1.5xIQR (interquartile range).



Supplementary Figure 15 – Pearson correlation between Pseudtime and principal component 1 (A), 2 (B), tSNE component 1 (C), 2 (D), and UMAP component 1 (E), and 2 (F) for ROS/MAP (DLPFC).



Figure S16 - Pearson correlation between Pseuodtime and principal component 1 (A), 2 (B), tSNE component 1 (C), 2 (D), and UMAP component 1 (E), and 2 (F) for Mayo RNAseq (TCX).



Supplementary Figure 17 – Monocle 3 trajectories and associations: UMAP method (Monocle3) for 218 independent samples from two independent studies, A) Lineage learned in TCX, B) Lineage learned in DLPFC, C) Association between disease pseudotime and diagnosis, D) correlation between pseudotime and IGAP GWAS genes. Box plots have lower and upper hinges at the 25th and 75th percentiles and whiskers extending to at most 1.5xIQR (interquartile range).



Supplementary Figure 18 – Monocle 3 trajectories and neuropath associations in DLPFC: UMAP method (Monocle3) for 338 independent samples from one study. A-C) Samples colored by three external measures of LOAD staging: Braak score, CERAD score, and cognitive diagnosis. D-F) Distribution of samples by inferred stage for different distinct stages in each of the three methods of measuring LOAD severity. Box plots have lower and upper hinges at the 25th and 75th percentiles and whiskers extending to at most 1.5xIQR (interquartile range).



Supplementary Figure 19 - APOE e4 status of samples overlaid on inferred manifolds for both TCX and

DLPFC brain regions.



Supplementary Figure 20 - DLPFC manifolds with samples colored by inferred disease state.



Supplementary Figure 21 - Quantile-quantile plot for the association with pseudotime in 305 female patients in the ROS/MAP cohort. The graph shows the Q-Q plot for GWAs of pseudotime in the ROS/MAP cohort with a genomic Inflation factor (lambda) of 0.981.



Supplementary Figure 22 - Quantile-quantile plot for the association with pseudotime in 131 female patients in the Mayo cohort.



Supplementary Figure 23 - Manifold learning identified potential genetic factors of stage progression and subtypes of LOAD. A-B) GWA analysis was performed on the Mayo (A) and ROSMAP (B) cohorts using whole genome sequenced data and LOAD pseudotime as the phenotype. Despite the small sample sizes of both analyses (N = 131 in Mayo, N = 306 in ROSMAP), several genomic loci were identified harboring SNPs with a genome wide suggestive p-value (p < $1x10^{-5}$). These include several loci that were previously associated with LOAD or LOAD related endophenotypes (red labels; see also **Table S5**)



Supplementary Figure 24 - UpSet plot of branch differentially expressed genes from a two-sided Tukey's honest significant test (FDR < 0.05) with branch one as reference branch in both studies respectively.





Supplementary Figure 25 – UpSet plot of comparison of clusters from Figure 4b from Mayo RNAseq lineage and differentially expressed genes from resistant individuals from the Mayo eGWAS study.



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Supplementary Figure 26 - Comparison of different manifold learning methods for TCX brain region.





Supplementary Figure 27 - Comparison of different manifold learning methods for DLPFC brain region.





Supplementary Figure 28 - Correlation between pseudotimes estimated by different manifold learning approaches on both TCX and DLPFC brain region.



Supplemental Tables

Supplementary Table 1

Patient characteristics table

	Мауо	(TCX)	Rosmap (DLPFC)			
	Females Males		Females	Males		
Characteristic ^a	N=134	N=128	N=338	N=199		
Combined Diagnosis, n (%)						
Control	35 (26.1)	36 (28.1)	42 (12.4)	37 (18.6)		
AD	49 (36.6)	31 (24.2)	92 (27.2)	39 (19.6)		
Other	ŇA	NA	204 (60.4)	123 (61.8)		
Path. Aging	17 (12.7)	13 (10.2)	ŇA	ŇA		
PSP	33 (24.6)	48 (37.5)	NA	NA		
Age of death, mean (sd)	ŇA	ŇA	89.6 (6.4)	86.3 (6.5)		
Braak stage, n (%)				. ,		
0	1 (0.7)	6 (4.7)	2 (0.6)	5 (2.5)		
0.5-2	29 (21.6)	22 (17.2)	46 (13.6)	49 (24.6)		
2.5-4	19 (14.2)	30 (23.4)	216 (63.9)	115 (57.8)		
4.5-6	49 (36.6)	31 (24.2)	74 (21.9)	30 (15.0)		
Missing	36 (26.9)	39 (30.5)	0	0		
CERAD score, n (%)		. ,				
1	NA	NA	101 (29.9)	47 (23.6)		
2			116 (34.3)	70 (35.2)		
3			37 (10.9)	23 (11.6)		
4			84 (24.9)	59 (29.6)		
Cognitive diagnostic category, n						
(%)						
1	NA	NA	102 (30.2)	69 (34.7)		
2			83 (24.6)	47 (23.6)		
3			3 (0.9)	6 (3.0)		
4			130 (38.5)	58 (29.1)		
5			14 (4.1)	13 (6.5)		
6			6 (1.8)	6 (3.0)		
Thal amyloid stage						
0	21 (15.7)	23 (18.0)	NA	NA		
1-2	13 (9.7)	19 (14.8)				
3	6 (4.5)	9 (7.0)				
4-5	36 (26.9)	17 (13.3)				
Missing	58 (43.3)	60 (46.9)				
APOE4 status ^b , n (%)						
0	92	98	257 (76.0)	146 (73.4)		
1	36	28	79 (23.4)	50 (25.1)		
2	6	2	2 (0.6)	3 (1.5)		
RIN ^c , median (min, max)	8.3 (5.3, 10.0)	8.3 (5.3, 10.0)	7.3 (5.0, 9.9)	7.3 (5.0, 9.2)		
DE genes ^d , n	72	34	28	20		

^aSee methods for detailed descriptions of clinical and neuropathological characteristics ^bNumber of APOE E4 alleles

CRNA integrity number

^dNumber of differentially expressed genes used in the manifold analysis (based on FDR<0.1).

Supplementary Table 2 - Results of logistic regression for the association between unadjusted and adjusted pseudotime calculations (scaled) and AD case-control status.

DLPFC

PS adjustment	Coefficient	Std. Error	z value	P value	
None	1.7398	0.4075	4.27	1.96E-05	
RIN number	1.3975	0.5732	2.438	0.0148	
PMI	1.4923	0.3721	4.01	6.06E-05	
1st 10 PCs	1.1616	0.4485	2.59	0.00961	
ALL	1.5486	0.5223	2.965	0.00303	

TCX

PS Adjustment	Coefficient	Std. Error	z value	P value	
None	1.0118	0.435	2.326	0.02	
RIN number	1.6223	0.5035	3.222	0.00127	
PMI	1.6097	0.4865	3.309	0.000938	
1st 10 PCs	1.545	0.498	3.103	0.00192	
ALL	2.0967	0.5278	3.973	7.10E-05	

Supplementary Table 4 – Associations between Braak, CERAD, and cogdx with features from alternative dimensionality reduction approaches.

Feature	Braak	CERAD	cogdx
PCA1	1.91x10 ⁻⁴	4.51x10 ⁻⁴	3.97x10 ⁻⁵
PCA2	0.800	0.201	1.07x10 ⁻²
tSNE1	0.857	0.979	0.461
tSNE2	1.02x10 ⁻³	1.11x10 ⁻⁴	1.23x10 ⁻⁵
UMAP1	0.0219	9.63x10 ⁻⁴	1.34x10 ⁻⁴
UMAP2	1.77x10 ⁻⁶	2.98x10 ⁻⁶	3.27x10 ⁻⁷
Pseudotime (DDRTree)	1.01x10 ⁻⁵	1.77x10 ⁻⁵	3.48x10 ⁻⁷
Pseudotime (UMAP)	1.34x10 ⁻⁴	6.64x10 ⁻⁴	2.18x10 ⁻⁵

DLPFC (P-value from logistic ordinal regression)

Supplementary Table 6 - Association between mean expression of cell specific signatures and inferred disease severity (pseudotime).

Study (Brain Region)	Cell Signature	P-value	R ²
Mayo RNAseq (TCX)	Neuronal	3.6x10 ⁻⁴²	0.76
	Microglial	9.1x10 ⁻²⁹	0.61
	Oligodendroglial	6.7x10 ⁻¹¹	0.28
	Astrocytic	6.7x10 ⁻²²	0.51
ROS/MAP (DLPFC)	Neuronal	1.6x10 ⁻⁷⁸	0.65
	Microglial	1.5×10^{-31}	0.33
	Oligodendroglial	1.4x10 ⁻⁴⁴	0.44
	Astrocytic	1.0×10^{-50}	0.48

Supplementary Table 7 - Overview of suggestive ($p < 10^{-5}$) results from single variant association with pseudotime. Unadjusted p-values for a two sided likelihood ratio test in a linear regression model are shown.

SNP				A1		Allele					
(dbSNP	Location	Nearest		(Effect		Freq.	Beta	SE			Previous
150)	(hg19)	Gene(s)	region	Ållele)	A2	(A1)	(Pseudotime)	(beta)	Р	Cohort	Association
ma4421010			intergeni							ROS/MA	
184421019	4:40309851	CHRNA9	С	Т	Α	0.35	-6.18	1.31	3.44E-06	Р	LOAD
			intergeni							ROS/MA	
rs12216400	6:96292130	intergenic	С	А	G	0.24	6.86	1.46	4.17E-06	Р	/
										ROS/MA	
rs1573618	7:142244415	TCRBV	intronic	Т	С	0.44	-6.22	1.29	2.43E-06	Р	/
										ROS/MA	Tangle
rs7870388	9:8660693	PTPRD	intronic	G	С	0.21	-6.40	1.42	1.32E-06	Р	burden
										ROS/MA	
rs4746059	10:72465488	ADAMTS14	intronic	G	Α	0.42	5.85	1.21	2.20E-06	Р	/
		ZNF490;	intergeni			.				ROS/MA	
rs55786848	19:12669655	ZNF564	С	C	Т	0.15	8.01	1.71	4.16E-06	Р	/
			intergeni								Plaque
rs12136200	1:240138130	CHRM3	С	C	Т	0.39	-16.61	3.36	2.42E-06	Mayo	burden
rs73818121	4:57397157	THEGL	exonic	G	С	0.07	33.19	6.63	1.81E-06	Mayo	/
			intergeni								
rs7809318	7:136419969	CHRM2	С	С	Т	0.07	-34.03	7.37	9.41E-06	Mayo	/
			intergeni								
rs3808616	8:79868493	IL7	С	G	А	0.35	-17.70	3.59	2.51E-06	Mayo	/
		ACCS;	intergeni								
rs11037791	11:44022056	ACCSL	С	A	G	0.49	-16.41	3.38	3.39E-06	Mayo	/
											LOAD,
											Tangle
		PVRL2;									burden,
60 57	10 45202254	TOMM40;		G	-	0.15	10.00	2.05	0.105.07		Plaque
rs6857	19:45392254	APOE	intronic	C	$ \mathbf{T} $	0.17	-18.23	3.95	9.18E-06	Mayo	burden

Supplementary Table 8 - Associations of known AD variants associated with pseudotime in the IGAP cohort. Unadjusted p-values for a two sided likelihood ratio test in a linear regression model are shown for pseudotime.

Chr.	Position	SNP	Minor	IGAP p-	Pseudotime	Pseudotime	Gene
	(lig1))		Frequency	(Stage1+2)	Conort	p-value	
2	127887750	rs62158731	0.26	3.41E-13	Mayo	4.68E-05	BIN1
3	151018968	rs66927386	0.24	1.40E-04	ROS/MAP	0.0090	MED12L
6	32570051	rs9270823	0.25	5.77E-10	ROS/MAP	0.0068	HLA-DRB1
7	99809921	rs1727128	0.48	4.43E-06	ROS/MAP	0.0029	STAG3
9	129197516	rs887656	0.11	1.40E-04	ROS/MAP	0.0079	MVB12
10	72524413	rs2688767	0.36	1.39E-04	ROS/MAP	0.0078	ADAMTS14
11	85862728	rs72962020	0.13	8.09E-06	Mayo	0.0075	PICALM
16	11199352	rs12929596	0.13	6.43E-05	ROS/MAP	0.0067	CLEC16A
19	45392254	rs6857	0.17	1.06E-15	Mayo	9.18E-06	APOE
20	55020557	rs16979933	0.09	1.08E-07	Mayo	0.0054	CASS4

Supplementary Table 11 - Number of genes differentially expressed at an FDR of 0.05 between the control branch (Branch 1) and other branches based on a two sided Tukey's Honest significant difference test in an ANOVA model.

Study (Brain	Change in	Branch 2	Branch 3	Branch 4	Branch 5	Branch 6
Region)	expression					
ROSMAP	Increased	718	468	1121	662	1239
(DLPFC)	Decreased	781	611	1017	783	1094
MayoRNAseq	Increased	506	2067	2034	2733	1815
(TCX)	Decreased	699	1912	2441	1966	1494

Supplemental Table Legends

Supplementary Table 3: AD LOAD GWAS genes²³. Genes are from Tables 1-3 from previously published work²³.

Supplementary Table 5: Cell specific gene sets used to compute mean expression of cell signatures across the lineages, as previously described³².

Supplementary Table 9: Summary statistics from differential expression analysis in DLPFC. Tukey's honest significant difference test with branch 1 as reference is used, where unadjusted p-values from a two sided t-test for the mean difference are shown.

Supplementary Table 10: Summary statistics from differential expression analysis in TCX. Tukey's honest significant difference test with branch 1 as reference is used, where unadjusted p-values from a two sided t-test for the mean difference are shown.

Supplementary Table 12: Significant GO pathway enrichments (FDR < 0.05) for DLPFC differential expressed gene sets.

Supplementary Table 13: Significant GO pathway enrichments (FDR < 0.05) for TCX differential expressed gene sets.

Supplementary Table 14: Significant GO pathway enrichments from biclustering analysis of mean expression of six branches (states) in TCX with four clusters.