

Supporting Online Material for

DmSAS is required for sialic acid biosynthesis in cultured *Drosophila* third instar larvae CNS neurons

Annelise E. von Bergen Granell^{*}, Karen B. Palter⁺, Ihan Akan⁺, Udayanath Aich[⊥], Kevin J. Yarema[⊥], Michael J. Betenbaugh[#], William B. Thornhill^φ and Esperanza Recio-Pinto^{*,ω}

^{*}Department of Anesthesiology, New York University Langone Medical Center, NY, NY 10016

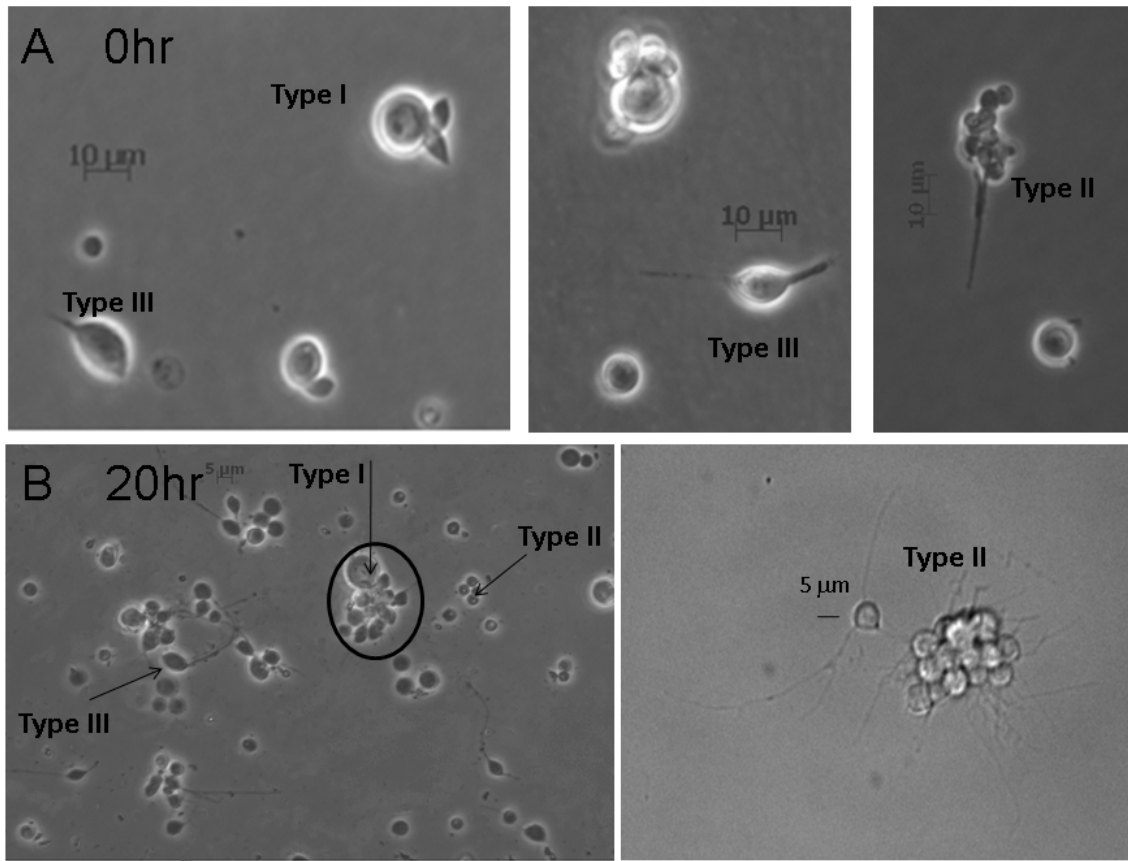
⁺Department of Biology, Temple University, Philadelphia, Pennsylvania 19122

[⊥]Department of Biomedical Engineering, The Johns Hopkins University, Baltimore, MD 21218

[#]Department of Chemical and Biomolecular Engineering, The Johns Hopkins University, Baltimore, MD 21218

^φDepartment of Biological Sciences, Fordham University, Bronx, NY 10458

^ωDepartment of Pharmacology, New York University Langone Medical Center, NY, NY 10016



Supplemental Figure S1: Neuronal cultures. Light microscopy pictures of live cultures from dissociated third instar larvae CNS (1.25CNS/coverslip) cultured in serum-containing medium (40x objective). (A) Freshly dissociated CNS cells (< 1h) in which we can identify the 3 types of cells that have been previously described(1). Type I, or Neuroblast-like, with a diameter of $\geq 8\mu\text{m}$, and we confirmed that these cells undergo asymmetrical division (data not shown). Type II, which are small cells (2-3 μm diameter) and usually display processes following dissociation. And Type III, oval or spindled-shaped cells (4-10 μm diameter), with thick and short processes. (B) After 20 h, cultures show that most of the type II and type III cells displayed neuritic outgrowth that continue to extend and branch with time.

1. Wu, C. F., Suzuki, N., and Poo, M. M. (1983) Dissociated neurons from normal and mutant *Drosophila* larval central nervous system in cell culture, *J Neurosci* 3, 1888-1899.