Hepatocyte nuclear factor 1 alpha influences pancreatic cancer growth and metastasis

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Supplementary materials and methods:

Antibodies and Reagents

The primary antibodies obtained and used for the experiments in this study were: $HNF1\alpha$ (12425), Notch 2 (4530), Snail (3879), Zeb (3396), Vimentin (5741), Slug (9585), N-Cadherin (4016), E-Cadherin (3195), pPI3K (4228), PI3K (4292), pmTOR (2974), mTOR (4517), pp70S6 kinase (9206), p70S6 kinase (9202), and Bim (2933), Bak (3814), Bax (2772), Cleaved Caspase 3 (9661), P53, pP53 (Cell Signaling Technology, Boston, MA, USA); Cleaved PARP (sc-8007), pAKT (sc-101629), AKT (sc-5298), pERK (sc-101760), and ERK (sc-94) (Santa Cruz, CA, USA); Caspase 8 (ab61755) and Bcl-2 (ab73985) (Abcam, Cambridge, MA, USA); β-actin (Sigma Aldrich, St. Louis, MO, USA). Respective anti-mouse and anti-rabbit secondary antibodies were acquired from Santa Cruz Biotechnology (Santa Cruz, CA, USA). The Transwell (6.5 mm) with 8.0 µm pore polycarbonate membrane inserts were purchased from Corning Incorporated (Corning, NY, USA). Both BD Matrigel (Bedford, MA, USA) and BD Pharmingen Annexin V-FITC Apoptosis Detection Kit I were obtained from BD Biosciences (San Diego, CA, USA). MTS reagent [3-(4, 5-dimethylthiazol-2-yl)-5-(3carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium] was purchased from Promega (Madison, WI, USA). Mammalian protein extraction reagent (mPER) was acquired from Thermo Scientific (Rockford, IL, USA).

Sequences for *HNF1α* siRNA subtypes "A", "B", and "C" SR304741A-rGrGrArGrUrGrCrArArUrArGrGrGrGrGrGrGrArArUrGrCrATC SR304741B-rGrGrArCrArGrGrArCrUrArArCrArCrUrCrArGrArArGrCCT SR304741C-rCrGrGrUrGrUrGrUrGrCrGrCrUrArUrGrGrArCrArGrCrCrUrGCG

Supplementary Figure S1







Supplementary Figure S3



3





Supplementary Figure S5



С





Supplementary Figure S6



Zeb

Notch-2

Vimentin

Snail

β-actin

1 Supplementary Figure Legends:

2 Supplementary Fig. S1

A. Immunofluorescence of HNF1α expression in AsPC-1 and hTERT HPNE cells (20X and 100X
magnification). B. AsPC1 cells were transfected with three different predesigned HNF1α siRNAS
at different concentrations.

6 Supplementary Fig. S2

A. Western blot analysis of HNF1α protein expression in overexpressed HNF1α HPAC pancreatic
cancer cells. B. Relative mRNA expression level of HNF1α in overexpressing HNF1α HPAC cells.
Data shown as mean +/- SEM. Experiments (n=3) were repeated three times in triplicates.
*p<0.05.

11 Supplementary Fig. S3

A. MTS assay was used to measure cell viability in response to HNF1α overexpression in HPAC
cells. D. Expression levels of pAKT, pmTOR, pERK, mTOR, pP70S6K, and β-actin were
determined by Western Blot in overexpressed HNF1α HPAC cells. Data shown as mean +/- SEM.
Experiments (n=3) were repeated three times in triplicates.

16 Supplementary Fig. S4

17 A & B. Apoptosis analysis of overexpressed HNF1α HPAC cells measured by flow cytometry.

- 18 C. Western blot analysis of apoptotic markers in siHNF1α AsPC-1 cells and overexpressed HPAC
- 19 cells. Data shown as mean +/- SEM. Experiments (n=3) were repeated three times in triplicates.

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22 Supplementary Fig. S5

A & B. Wound-healing assay was performed in HNF1α overexpressed HPAC cells; migration was
analyzed using Nikon Biostation Ct at 2h intervals for up to 96h at 4X magnification. C & D.
Invasiveness of OVHNF1α HPAC cells were observed using a Matrigel invasion assay, captured
using Nikon Eclipse TS 100 microscope. Data shown as mean +/- SEM. Experiments (n=3) were
repeated three times in triplicates. *p<0.05.

28 Supplementary Fig. S6

A & B. Colony formation assay was performed in HNF1α overexpressed HPAC cells. **C.** EMT

- 30 markers analyzed by Western blot in overexpression HNF1 α in HPAC cells. Data shown as mean
- +/- SEM. Experiments (n=3) were repeated three times in triplicates. *p<0.05

Fig. 2C



Fig. 3A





Fig. 3E



Fig. 4G

Scr siHNF-1 α

Fig. 5G

N-Cadherin

Scr si-HNF1 α

E-Cadherin

Vimentin

Scr si-HNF1α

Scr si-HNF1α

Supplementary Figure S2A

Supplementary Figure S3B

Supplementary Figure S6C

