Suppl. Material and Method

EDTA-whole blood samples were collected at Cycle 1, Cycle 5, Cycle 9 and Cycle 13 (in Part B only) prior to treatment (therefore two weeks after the previous dosing) and EOT. Direct blood staining with BD Multitest™ 6-Color TBNK (FITC-labeled CD3, clone SK7; PE-labeled CD16, clone B73.1, and PE-labeled CD56, clone NCAM16.2; PerCP-Cy5.5–labeled CD45, clone 2D1; PE-Cy7–labeled CD4, clone SK3; APC-labeled CD19, clone SJ25C1 and APC-Cy7–labeled CD8, clone SK1) and with BD Multitest™ CD8-FITC (clone SK1)/CD38-PE (clone HB7)/CD3-PerCP (clone SK7)/HLA-DR-APC (clone L343) was performed to enumerate ex vivo the absolute counts of T and NK cells populations and activated T $CD3^+CD8^+$ or $CD3^+CD8^-$ cells co-expressing HLA-DR and CD38 in BD Trucount™ tubes with a Lyse/No Wash procedure, avoiding any bias induced by centrifugation steps. Cells were acquired on BD FACSCanto™ II flow cytometer. The paired non-parametric Wilcoxon signed rank test was used to compare the immuno-monitoring results obtained at the different time points to the pre-treatment baseline using JMP® version 12 software.

Suppl. Figure 1 legend

Suppl. Figure 1: Combined treatment effect of T/NK cells compartment.

Absolute counts of circulating $CD45^+$ $CD3^+$ (T cell), $CD45^+$ $CD3^+$ CD4+ (CD4 T cell), $CD45^+$ $CD3^+$ $CD8^+$ (CD8 T cell), $CD3^+$ $CD8^+$ $HLA\text{-}DR^+$ $CD38^+$ (Activated CD8 T cells), $CD3^+$ $CD8^-$ $HLA\text{-}DR^+$ $CD38^+$ (Activated CD4 T cells) in blood samples collected pre-dosing at the start of combined therapy (Cycle 1), at Cycle 5, Cycle 9 and Cycle 13 (for part B only) are shown for each dosing group in Part A and in Part B (top panels). Relative fold changes from the baseline calculated for all cell population are shown (bottom panels) Significant changes assessed by paired non-parametric Wilcoxon signed rank test are displayed with a horizontal bar ($p<0.05$).