	Number of data points	Number of outliers
FHL1 (-)	28	0
FHL1 (+)	80	1
FHL2 (-)	69	5
FHL2 (+)	142	4
FHL3 (-)	166	9
FHL3 (+)	226	2
Paxillin (-)	72	2
Paxillin (+)	134	2
HIC5 (-)	102	1
HIC5 (+)	161	1
Zyxin (-)	74	5
Zyxin (+)	140	3
FBLIM1 (-)	91	2
FBLIM1 (+)	132	1
TRIP6 (-)	50	2
TRIP6 (+)	89	4
LPP (-)	39	2
LPP (+)	101	3
LIMD1 (-)	28	1
LIMD1 (+)	37	2
Paxillin LDO (-)	198	14
Paxillin LDO (+)	244	1
HIC5 LDO (-)	111	2
HIC5 LDO (+)	170	3
Zyxin LDO (-)	71	3
Zyxin LDO (+)	115	2
FBLIM1 LDO (-)	100	2
FBLIM1 LDO (+)	146	0
TRIP6 LDO (-)	45	1
TRIP6 LDO (+)	69	1
LPP LDO (-)	129	3
LPP LDO (+)	222	2
LIMD1 LDO (-)	35	2
LIMD1 LDO (+)	93	2

Table S1. Number of data points and outliers identified by ROUT method in GraphPad Prism for SF enrichment quantification in unstretched (-) and stretched (+) MEFs. Related to Figure 2.

Table S2. Additional oligonucleotides and recombinant DNAs used in this paper. Related to the Key Resources Table.



Figure S1. Non-mechanoaccumulating LIM proteins identified in the cell-stretching screen. Related to Figure 1. (A) Epifluorescence micrographs of eGFP-labeled LIM proteins in unstretched (-) and stretched (+) MEFs stained with phalloidin to label F-actin. Double-headed arrow indicates the uniaxial stretch direction. Scale bar, 20 μ m. (B) Whole-cell actin enrichment of LIM proteins in unstretched (-) and stretched (+) MEFs ($20 \le n \le 60$). Bars represent means. Dunnett's T3 multiple comparisons test after Welch's ANOVA is performed by comparing the actin enrichments of each LIM protein in unstretched cells with those in stretched ones simultaneously for all the LIM proteins included in the screen. NS, p > 0.05; ** p < 0.01; **** p < 0.001.



Figure S2. FHL-, paxillin-, and zyxin-family proteins localize to SF strain sites with similar kinetics, supporting recognition of a common molecular mark. Related to Figure 1. (A) Left, spinning disk confocal snapshot of a U2OS cell co-expressing F-tractin-mApple (magenta) and zyxin-eGFP (green). Scale bar, 5 μ m. Box highlights an SF strain site. Right, intensity versus time plot (top), time-lapse montages (middle), and intensity line scan (bottom) of F-tractin and zyxin at an SF strain site. Scale bar, 1 μ m. (B) Intensity vs. time plots (top), time-lapse montages (middle), and intensity line scans (bottom) of representative SF strain sites marked by zyxin-FusionRed and the indicated eGFP-labeled LIM protein. Magenta, zyxin-FusionRed. Green, eGFP-labeled LIM protein. Scale bar, 1 μ m. The time of first intensity peak is indicated for each protein. (C) Timing of first intensity peak of indicated LIM proteins at $6 \le n \le 17$ SF strain sites relative to zyxin-FusionRed. Bars represent means. Dashed line indicates simultaneous recruitment of zyxin-FusionRed and the eGFP-labeled LIM protein.





Figure S3. Mechanoresponsive LIM proteins accumulate on SFs in stretched MEFs. Related to Figure 2. (A) Confocal slices of phalloidin-stained SFs and eGFP-labeled paxillin- and zyxin-family proteins in unstretched (-) and stretched (+) MEFs. Top panel, full-length (FL). Bottom panel, LIM domain only (LDO). Scale bar, 2 μ m. (B) Left panel, confocal slices of SFs stained by phalloidin (top) and endogenous FHL2 (bottom) visualized by immunofluorescence. Scale bar, 2 μ m. Right panel, SF enrichment of endogenous FHL2 in unstretched (-) and stretched (+) MEFs. Each data point is obtained from a single SF (25 \leq n \leq 38, N = 2 biological replicates). Bars represent means. Welch's unpaired t-test: ***** p < 0.0001.

Α Mechanoresponder FBLIM1_LIM3 LIMD1_LIM3 Zyxin_LIM3 TRIP6_LIM3 LPP_LIM3 FBLIM1_LIM2 LIMD1_LIM2 Zyxin_LIM2 TRIP6_LIM2 TRIP6_LIM2 FHL1_LIM2 FHL1_LIM2 FHL3_LIM2 DE I I FAED DFKI DAGD EFPI EAGD DFPVEAGD LIMCD1 LIM2 RCSG KCHG ICHG CGRU CGRU CGRU KGAAPPDSE CK IVYPTE CK TVYPME CQK TVYPAE RMGEEGSELPV FHL3_LIM2 FHL1_LIM1

FHL2 LIM1 FHL3 LIM1

LIMD1_LIM1 FBLIM1_LIM1 FBLIM1_LIM1 TRIP6_LIM1 FHL1_LIM4 FHL2_LIM4 FHL2_LIM3 FHL2_LIM3 FHL2_LIM3 FHL2_LIM3 FHL2_LIM3 Paxilin_LIM1 HIC5_LIM1 Paxilin_LIM1 HIC5_LIM2 HIC5_LIM2

D

Phalloidin eGFP

(F-actin) LIM protein paxillin

В

Phalloidin

Zyxin LDO-eGFP

Anti-

Paxillin

WT

Non-mechanoresponder



Figure S4. Role of the divergent LIM3 of zyxin in SF mechanoaccumulation and role of actin mechanoaccumulation in LIM protein recruitment to FAs. Related to Figure 4. (A) Multiple sequence alignments of mechanoresponder (left) and non-mechanoresponder (right) LIM domains, colored by conservation (shades of purple). Green boxes highlight the conserved phenylalanine in mechanoresponder LIM domains (left) and residues at the same position in non-mechanoresponder LIM domains (right). Red box highlights the phenylalanine next to the conserved position in zyxin LIM3. Orange boxes highlight extended insertions in LIM3 of zyxin-family members. (B) Epifluorescence micrographs of MEFs expressing eGFP-labeled WT zyxin LDO (left), zyxin LDO F(1,2)A (middle), and zyxin LDO F(1-3)A (right), stained with phalloidin to label F-actin. Scale bar, 20 μ m. (C) Whole-cell actin enrichments of wild-type (n = 62), F(1,2)A (n = 102), and F(1-3)A (n = 56) zyxin LDO. N = 2 biological replicates. Games-Howell's multiple comparison test after Welch's ANOVA: *** p < 0.001; **** p < 0.0001. (D) Epifluorescence micrographs of FAs in MEFs expressing eGFP-labeled LIM proteins, stained with phalloidin and anti-paxillin antibody. Scale bar, 2 μ m. (E) Whole-cell focal adhesion and actin enrichments of wild-type and F(1-4)/(1-3)A full-length and LDO constructs. n = 81-115. N = 2 biological replicates. Welch's unpaired t-test: **** p < 0.0001.



Figure S5. Purified LIM proteins featuring phenylalanine mutations preserve their folded

structures. Related to Figures 5, 6. (A) Size-exclusion chromatograms of Halo-tagged wild-type and mutant FHL3 (left), HIC5 (middle), and zyxin (right). (B) Coomassie-stained SDS-PAGE (3 µg) and (C) circular dichroism spectra of the indicated purified proteins.



Figure S6. Additional analyses of LIM protein patches. Related to Figures 5, 6. (A) Two additional examples of the cumulative projections of wild-type FHL3 (top), HIC5 (middle), and zyxin (bottom) in the absence and presence of ATP. (B) Montages showing the rebinding of FHL3 (left) and zyxin (right) after filament breakage. Scale bars, 2 μ m. (C) Scatter plots of maximum area (top) and maximum normalized average fluorescence intensity (bottom) of patches versus their lifetime for the indicated proteins.



Figure S7. Strained actin binding is the dominant upstream process that mediates FHL2 mechanosensation in MEFs, which does not require the phosphorylation of FHL2 by FAK. Related to Figure 7. (A) Top panel, maximum intensity projections (MIPs) of expressed FHL2-eGFP localization in a MEF before (left) and after (middle) Y-27632 treatment, and after washout (right). MIPs were constructed from three confocal z-slices. Bottom panel, zoomed-in views of the boxed regions. Scale bars, 10 μ m. (B) Average nuclear (top) and actin (bottom) enrichments obtained from n = 16 cells from N = 3 biological replicates as a function of time, aligned by the timepoint of Y-27632 addition. Error bar represents SEM. Dash lines indicate the time points of Y-27632 addition (10 minutes) and washout (100 minutes). (C) Replotting of average nuclear enrichment versus average actin enrichment, displayed as a scatterplot where points are color-coded by time. (D) Nuclear (left) and actin (right) enrichment of eGFPtagged wild-type (n = 106), F(1-4)A (n = 110), Y93F (n = 41), and F(1-4)A-Y93F FHL2 (n = 50) expressed in MEFs plated on glass. N = 2 biological replicates. Games-Howell's multiple comparison test after Welch's ANOVA: NS, p > 0.05; * p < 0.05; **** p < 0.001. (E) Nuclear (left) and actin (right) enrichment of eGFP-tagged FHL2 F(1-4)A expressed in MEFs plated on glass with (n = 34) and without (n = 36) FAK inhibition. N = 2 biological replicates. Welch's unpaired t-test: NS, p > 0.05. (F) Nuclear (left) and actin (right) enrichment of eGFP-tagged wild-type FHL2 expressed in MEFs plated on 12-kPa hydrogels with (n = 31) and without (n = 32) FAK inhibition. N = 2 biological replicates. Welch's unpaired t-test: NS, p > 0.05.