

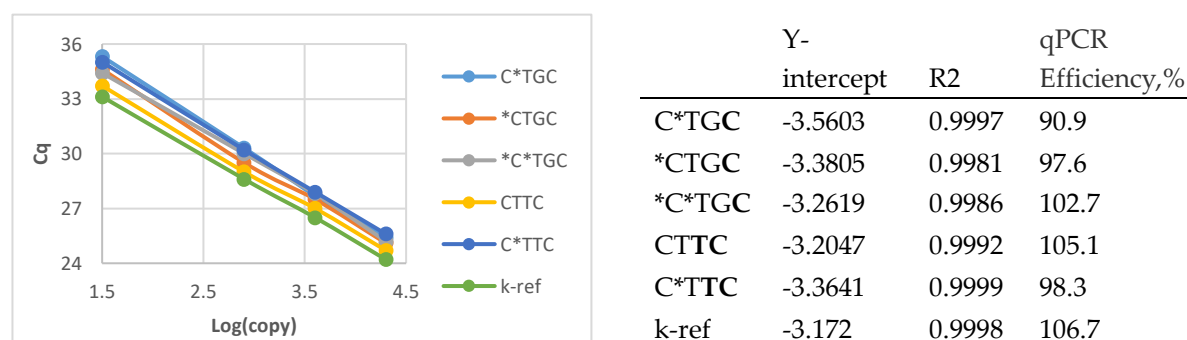
# Allele-Specific PCR for KRAS Mutation Detection Using Phosphoryl Guanidine Modified Primers

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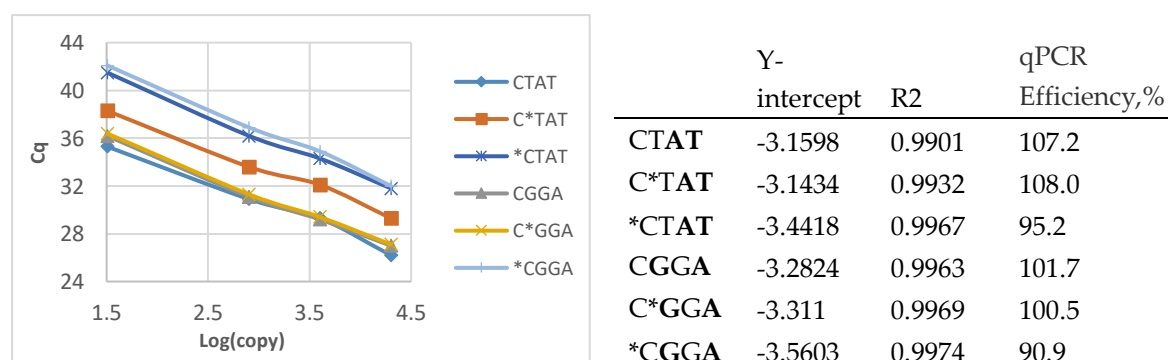
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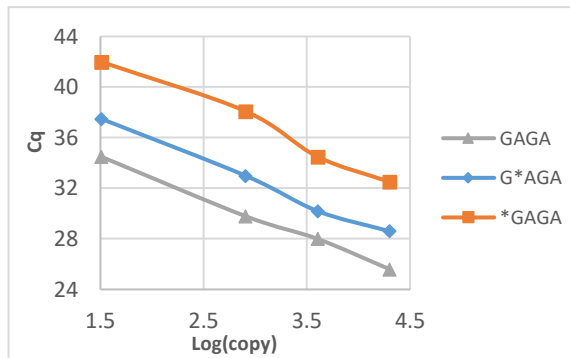
The influence of the number of PG groups and their position at 3'-terminus were investigated using AS-primers CTTC and CTGC (Table S1) and WT and KRAS G12A mutation plasmids as templates [1,2]. Bold symbols marked nucleotides means mismatched nucleotides in the relation to the wild-type DNA sequence. To determine assay reproducibility, several experiments were performed in 5 replicates with various amounts of the mutant template in the samples (Table S2). Standard curve data were generated for several native and phosphoryl guanidine (PG) modified primers for KRAS mutation detection and the PCR efficiency for each of these primers was calculated using Thermo Fisher Scientific online calculator (Figure S1-S3) [3]. All experiments had amplification efficiency more than 90% and a correlation coefficient  $R^2$  above 0.99. Allele-specific blocking PCR KRAS mutations detection assay results using blockers containing C\*T\*G\*G or G\*T\*G\*G 3'-terminal fragment are presented in Table S3.



**Figure S1.** Cq values of KRAS G12A mutation detection using C\*TGC, \*CTGC, \*C\*TGC, CTTC, C\*TTC, k-ref primers and various amount of mutant DNA per reaction. Standard curves showing the PCR efficiency values for the several primers generated from a dilution series of mutant DNA.



**Figure S2.** Cq values of KRAS G12V, G12D mutations detection using CTAT, C\*TAT, \*CTAT, CGGA, C\*GGA, \*CGGA primers and various amount of mutant DNA per reaction. Standard curves showing the PCR efficiency values for the several primers generated from a dilution series of mutant DNA.



	Y-intercept	R2	qPCR Efficiency, %
<b>GAGA</b>	-3.1516	0.9979	107.6
<b>G*AGA</b>	-3.2579	0.9944	102.7
<b>*GAGA</b>	-3.4786	0.9852	93.9

**Figure S3.** Cq values of *KRAS* G13D mutations detection using **GAGA**, **G\*AGA**, **\*GAGA** primers and various amount of mutant DNA per reaction. Standard curves showing the PCR efficiency values for the several primers generated from a dilution series of mutant DNA.

**Table S1.** AS-PCR *KRAS* G12A mutation detection using CTGC and CTTC primers with PG-modification in the different phosphates from the 3'-end and mutant DNA mixed with wild-type DNA (total  $2 \times 10^5$  copies per reaction) at various ratios (% of mutant DNA)

Primers	Cq		
	WT	1%	100%
CTGC	28.1±0.1	27.2±0.1	24.0±0.1
C*TGC	33.8±0.2	28.3±0.1	21.9±0.1
*CTGC	37.4±0.2	28.1±0.05	22.2±0.2
*C*TGC	N/A	29.8±0.2	23.4±0.1
CTG*C	36.9±0.2	30.3±0.2	24.1±0.1
CT*GC	36.8±0.4	31.4±0.1	25.3±0.1
*CTG*C	N/A	37.2±0.2	27.7±0.2
C*TG*C	N/A	34.9±0.5	29.5±0.2
CTTC	34.25±0.05	28.7±0.1	21.72±0.02
C*TTC	N/A	29.65±0.07	23.8±0.2
CT*TC	N/A	33.24±0.01	26.8±0.2
CTT*C	N/A	33.5±0.1	26.67±0.01
C*TT*C	N/A	36.4±0.2	29.2±0.1
*C*TTC	N/A	37.9±0.1	32.0±0.1
C*T*TC	N/A	40.9±0.2	34.8±0.1
*CTT*C	N/A	N/A	40.1±0.3
CT*T*C	N/A	N/A	N/A

NTC (No Template Control) was undetermined in all the reactions, N/A indicates that no Cq was obtained for a typical 45-cycle reaction. Symbol "\*" means PG modification location. Boldly marked nucleotides mean mismatched nucleotides in relation to the wild-type DNA sequence

**Table S2.** AS-PCR reproducibility G12A *KRAS* mutation detection using wild-type DNA (total  $2 \times 10^4$  copies per reaction) and 1% mutant DNA in the background of wild-type DNA

Primers	Cq		
	WT	1%	
k-ref	1	25.73	25.21
	2	25.53	25.23
	3	25.72	25.17
	4	25.66	24.86
	5	25.20	24.76
	aver	25.57	25.05
	SD	0.22	0.22
CTTC	1	36.55	31.43
	2	35.79	31.64
	3	36.17	31.66
	4	35.42	31.22
	5	36.26	31.29
	aver	36.04	31.45
	SD	0.44	0.20
C*TTC	1	N/A	33.13
	2	N/A	33.11
	3	N/A	32.70
	4	N/A	32.67
	5	N/A	33.00
	aver	-	32.92
	SD	-	0.22

**Table S3.** ASB-PCR KRAS mutations detection of various mutant/wild-type DNA ratio (total 2×10<sup>4</sup> or 2×10<sup>5</sup> copies per reaction)

Primers	Cq				$\Delta Cq$ Cq(WT) - Cq(1%)		
	2×10 <sup>4</sup> copies				2×10 <sup>4</sup>	2×10 <sup>5</sup>	
	1%	0.1%	0.01%	WT	copies	copies	
G12A	No blocker CTGG	24.6±0.2	24.8±0.2	25.0±0.3	25.0±0.2	0.4	-
	Blocker C*T*G*G	N/A	N/A	N/A	N/A	-	-
	No blocker CTTC	31.5±0.1	33.9±0.1	34.2±0.2	36.7±0.2	5.2	5.7
	CTTC/C*T*G*G = 1/5, 1/2	N/A	N/A	N/A	N/A	-	-
	CTTC/C*T*G*G = 1/1	N/A	N/A	N/A	N/A	-	-
	CTTC/C*T*G*G = 1/0.5	41.6±0.3	N/A	N/A	N/A	-	-
	CTTC/C*T*G*G = 1/0.25	37.0±0.6	40.1±1.1	N/A	N/A	-	-
	CTTC/C*T*G*G = 1/0.1	34.5±0.1	38.0±0.2	N/A	N/A	10.5 <sup>a</sup>	10.7
	C*TTC/C*T*G*G = 1/5, 1/2	N/A	N/A	N/A	N/A	-	-
	C*TTC/C*T*G*G = 1/1	41.7±1.5	N/A	N/A	N/A	-	-
	C*TTC/C*T*G*G = 1/0.5	33.0±0.2	37.3±0.5	41.8±1.0	N/A	12.0 <sup>a</sup>	10.1
	C*TTC/C*T*G*G = 1/0.25	32.5±0.1	36.3±0.3	38.2±0.4	N/A	12.5 <sup>a</sup>	14.2
	C*TTC/C*T*G*G = 1/0.1	31.7±0.1	34.9±0.1	38.0±0.3	39.8±0.9	8.1	-
	C*TGC/C*T*G*G = 1/1	37.4±0.5	N/A	N/A	N/A	-	-
	C*TGC/C*T*G*G = 1/0.5	34.8±0.1	39.3±0.4	43.0±1.8	N/A	10.2 <sup>a</sup>	9.7
C*TGC/C*T*G*G = 1/0.25	33.1±0.1	37.5±0.2	40.1±0.5	42.4±1.5	9.3	9.2	
C*TGC/C*T*G*G = 1/0.1	32.3±0.1	36.4±0.3	38.7±0.7	39.0±1.0	6.7	-	
G12V	No blocker CTAT	30.6±0.1	30.8±0.2	31.4±0.2	31.6±0.2	1.0	-
	No blocker C*TAT	31.5±0.1	35.3±0.2	36.7±0.4	37.3±0.5	5.8	-
	C*TAT /C*T*G*G = 1/0.5	41.3±1.0	N/A	N/A	N/A	-	-
	C*TAT /C*T*G*G = 1/0.25	39.3±0.8	N/A	N/A	N/A	-	8.8
	C*TAT /C*T*G*G = 1/0.1	36.0±0.2	37.5±0.4	40.3±1.0	N/A	9.0 <sup>a</sup>	8.8
G12D	No blocker CGGA	33.2±0.1	36.0±0.3	37.0±0.5	37.1	3.8	-
	C*GGA/C*T*G*G = 1/0.5	35.5±0.2	39.8±1.0	N/A	N/A	10.5 <sup>a</sup>	-
	C*GGA/C*T*G*G = 1/0.25	33.7±0.1	37.3±0.5	N/A	N/A	11.3 <sup>a</sup>	9.8
	C*GGA/C*T*G*G = 1/0.1	33.6±0.1	37.1±0.6	39.2±0.9	N/A	11.4 <sup>a</sup>	9.8
G13D	No blocker GTGG	23.5±0.1	24.0±0.2	24.1±0.1	24.2±0.3	0.7	-
	Blocker G*T*G*G	N/A	N/A	N/A	N/A	-	-
	No blocker GAGA	31.1±0.1	33.1±0.2	33.6±0.3	35.2±0.2	4.1	-
	GAGA/G*T*G*G = 1/0.1	31.7±0.1	33.8±0.2	N/A	N/A	13.3 <sup>a</sup>	8.8
	G*AGA/ G*T*G*G = 1/0.5	38.8±0.7	N/A	N/A	N/A	-	9.3
	G*AGA/ G*T*G*G = 1/0.25	36.2±0.2	43.0±1.5	N/A	N/A	8.8 <sup>a</sup>	9.5
	G*AGA/G*T*G*G = 1/0.1	35.3±0.4	39.2±0.8	41.0±1.3	N/A	9.7 <sup>a</sup>	10.9

NTC (No Template Control) was undetermined in all the reactions, N/A indicates that no Cq was obtained for a typical 45-cycle reaction. Symbol “\*” means PG modification location. Boldly marked nucleotides mean mismatched nucleotides in relation to the wild-type DNA sequence. Real-time PCR assay was done using a constant 450 nM AS-primers concentration and several blocker primers excess. <sup>a</sup>If Cq(WT) = N/A to calculate  $\Delta Cq = Cq(WT) - Cq(1\%)$  value Cq(WT) = 45 was used.

## References

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