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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our Editorial Policies and the Editorial Policy Checklist.

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Fora	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.			
n/a	Confirmed			
x	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement			
x	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly			
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.			
	🗶 A description of all covariates tested			
×	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons			
x	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)			
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>			
×	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings			
x	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes			
x	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated			
,	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.			
Sof	ftware and code			
Policy information about availability of computer code				

Data collection No software was used.

Data analysis

The main results are generated by using Matlab-R2017a, Python 2.7 and R-3.6.1. The code generating the main results in this work are available at https://github.com/XuemingLiu/Robust-Multilyer-BioNetworks.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The data used in this work are collected from open-access databases.

The transcription factors and their DNA binding motifs used in the construction of the human regulatory network were downloaded from the CIS-BP database (http://cisbp.ccbr.utoronto.ca/). The three tissue-specific gene regulatory networks were downloaded from the FANTOM5 database [59], respectively "forebrain" (ID: 03), "lymphocytes" (ID: 12), and "lung" (ID: 23). The PPI network data were obtained from Supplementary Data 1 provided by Cheng et al. [60]. The human chemical-chemical (chemical.links.detailed.v4.0.tsv) and protein-biochemical (9606.protein_chemical.links.detailed.v4.0.tsv) links were downloaded from the STITCH database (version 4.0) (http://stitch.embl.de/). The human metabolites data were downloaded from the Human Metabolome Database (HMDB) (version 3.5, retrieved November 2015). The essential and essential genes were respectively collected from the Database of Essential Genes

Replication For the reasons stated above standard replication procedures were not performed. However, we validated several of our findings on alternative datasets. For example, we validated the results on the essential gene enrichment on three additional criteria for gene essentiality proposed in literature. As another example, we validated the finding on the network robustness on three additional tissue specific regulatory

networks built as described in Methods.

Randomization Since this is not a case/control study and for the reasons stated above, randomization is not relevant for this study

Blinding Since this is not a case/control study and for the reasons stated above, blinding is not relevant for this study

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems		Methods	
n/a	Involved in the study	n/a Involved in the study	
x	Antibodies	ChIP-seq	
x	Eukaryotic cell lines	Flow cytometry	
x	Palaeontology and archaeology	MRI-based neuroimaging	
x	Animals and other organisms	·	
x	Human research participants		
x	Clinical data		
×	Dual use research of concern		