

GPI-anchor synthesis in plants – a glycobiology perspective

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Supplementary Material

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Figure S1. LC-ESI-MS/MS analysis of the C-terminal peptide from *N. benthamiana* RFP-COB1.

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Supplementary Methods



Figure S1. LC-ESI-MS/MS analysis of the C-terminal peptide from *N. benthamiana* RFP-COB1. RFP-COB1 was transiently expressed by agroinfiltration of leaves, captured by binding to RFP-Trap resin and subjected to PI-PLC and trypsin digestion. A representative analysis from two biological replicates is shown.



Figure S2. LC-ESI-MS analysis of α -mannosidase or β -galactosidase digested GPI-anchor derived from *N. benthamiana* expressed RFP-COB1. The β -galactosidase digestion was carried out two times with identical results. A representative analysis is shown.



Figure S3. LC-ESI-MS/MS analysis of β -galactosidase digested and mock-incubated GPI-anchor derived from RFP-COB1 expressed in *A. thaliana* seedlings. A representative analysis from two technical replicates is shown.

Supplementary Methods

The C-terminal peptide of COBRA_ARATH was amplified from genomic DNA of A. thaliana using cob1 5F (5'-TATAGGATCCCCTTTTCTTCCCAACGGTGGTTCC-3')/cob1 6R primers (5'-TATACTCGAGTTAGGCAGAGAAGAAGAAGAAGAAC-3') and cloned into a modified version of the pPT2-vector (Strasser et al. 2005) containing the Arabidopsis ubiquitin10-promotor and N-terminal mRFP. Reporter proteins were either purified from leaves of N. benthamiana following agroinfiltration or from 14-day old seedings of stably transformed A. thaliana. Plant samples were snap frozen, grinded and the membrane fraction collected by centrifugation in 100 mM Tris, 25 %(w/v) sucrose and 5 % (v/v) glycerol buffer (pH 7.4) at 65000 g for 1 h at 4 °C. After washing in 100 mM Tris, 150 mM NaCl and 0.5 mM EDTA buffer (pH 7.4), the membrane pellet was resuspended in 20 mM Tris, 10 mM NaCl, 0.1 mM CaCl₂, 1% (v/v) Phosphoinositide Phospholipase C (PIPLC, P6466, Thermo Fisher Scientific) (pH7.4) and incubated for 15 min. After cleavage of lipids from RFP-GPI, the protein was purified using RFP trap (Chromotek), separated by SDS-PAGE, excised from the gel, reduced, Scarbamidomethylated and subjected to tryptic digest as described previously (Kolarich and Altmann 2000). The resulting GPI-bearing peptide fragment was analyzed via LC-ESI-MS/MS using a Bruker Maxis 4G Q-TOF instrument. For enzymatic deglycosylation of the GPI anchor, either β-galactosidase from Aspergillus oryzae (Zeleny et al. 1997) or α-mannosidase from Canavalia ensiformis (M7257, Sigma-Aldrich) was used. RFP-GPI was purified once from A. thaliana and two times from N. benthamiana.

References

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