

## SUPPLEMENTARY DATA

### SUPPLEMENTARY MATERIALS AND METHODS

#### Yeast strains, media and growth conditions.

BY4741, MB159-4D and MJ15-9C yeast strains used in this study were described previously (1, 2). Heterozygous diploid *RPB10/rpb10Δ* (BY4743 MAT $\alpha$ /MAT $\alpha$  *ura3Δ0/ura3Δ0 leu2Δ0/leu2Δ0 his3Δ1/his3Δ1 met15Δ0/MET15 LYS2/lys2Δ0 YOR210w/YOR210w::kanMX*) and the deletion mutant *upf1Δ* (BY4741 MAT $\alpha$  *ura3Δ0 leu2Δ0 his3Δ1 met15Δ0 YMR080c::kanMX*) were obtained from Euroscarf. *RPB10/rpb10Δ* was transformed with plasmids containing wild type or mutated *RPB10* alleles and subjected to meiosis. Generated *rpb10Δ* haploids expressing *RPB10* alleles from the plasmids were subsequently crossed with the *rpc128-1007* mutant (MJ15-9C MAT $\alpha$  *SUP11 ade2-1 ura3-1 lys2-1 leu2-3,112 his3*) resulting in *rpc128-1007 rpb10Δ[RPB10]* or *rpc128-1007 rpb10Δ[RPB10 Δ3'154]*. The strain Upf1-TAP (MAT $\alpha$  *ade2-1 his3-11,15 leu2-3,112 trp1-1 ura3-52 UPF1 TAP HIS3*) kindly supported by A. Dziembowski, was crossed with *rbs1Δ* (BY4742 MAT $\alpha$  *his3Δ1 leu2Δ0 lys2Δ0 ura3Δ0 YDL189w::kanMX4*) or Rbs1-Myc (MAT $\alpha$  *SUP11 ade2-1 ura3-1 lys2-1 leu2-3,112 his3 YDL189w::13Myc::KanMX6*) (1) that resulted Upf1-TAP *rbs1Δ* or Upf1-TAP Rbs1-Myc strains.

The *rpc128-1007 upf1Δ* strain was obtained by transformation of *upf1Δ* deletion cassette, which was amplified by PCR from the genomic DNA of BY4741 *upf1Δ* by using specific Upf1For (5'-AGAAGGAAGGGCAGCAAGAC-3') and Upf1Rev (5'-GCGCTCATTTCACGGTTGAG-3') primers. Transformants were selected on YPD medium supplemented with geneticin (200 µg/ml) and the replacement of *UPF1* with the kanamycin cassette was confirmed by PCR.

The RBS1-HTP strain was obtained by transformation of the appropriate sequence encoding the HTP tag, which was amplified by PCR using the pBS1539-HIS6-TEV-ProtA plasmid as DNA template and specific primers, RBSHTP\_F (5'-CTAGGGATACTGATTGGTAGAGATGAAATTGATAAACATTAGGAGCA CCATCACCATCAC-3') and RBS1HTP\_R (5'-TCTATAAACCGTTACGTAATTTTCGCTATGTATAGTTCACTCCTCACGTACGAC TCACTATAGGG-3'). Transformants were selected on SC-ura medium. The presence of RBS1-HTP-encoding sequence was confirmed by western blot method with the PAP antibody. As required, the YPD medium was supplemented with geneticin (200 µg/ml). Sporulation medium contained 0.05% glucose, 0.1% yeast extract and 1% potassium acetate. LB medium (1% Bacto tryptone, 0.5% yeast extract, 1% NaCl, pH 7.5) was used for growing *E. coli* strains. As required, the LB medium was supplemented with ampicillin (60 µg/ml). Solid media contained 2% agar. For analysis of the phenotypes, yeast strains grown on SC-ura, SC-leu or SC-ura-leu medium were replica-plated on YPD plates and incubated for 3 days at the desired temperature. Yeast cells were transformed with plasmids by using the lithium acetate method (3).

#### Plasmids

YEpl181-*RBS1*, called here [*RBS1*], multicopy plasmid (*LEU2*, 2µ) containing the *RBS1* gene (1). YEpl181-*RBS1* R3H, called here [*RBS1* R3H], the conserved Arg57 aa and His61 aa, located within the R3H domain, were changed to alanine using primers Mut\_rev (5'-ATAGAACAGCATAATATGAATTCACTCGGTC-3') and Mut\_for (5'-CCGCCAAATAGCTGAGTACCAACAA-3'). YEpl181-*RBS1*-Myc and YEpl181-*RBS1* R3H-Myc, called here [*RBS1*-Myc] and [*RBS1* R3H-Myc] contain the wild type or mutated version

of *RBS1* tagged with Myc epitope at the 3' termini. They were obtained by transformation of BY4741 *rbs1Δ* [*RBS1*] or [*RBS1* R3H] with the His3MX6 cassette PCR amplified from pFA6a-13Myc-His3MX6 plasmid using primers RBS1\_F2 (5'-TACTGATTCGGTAGAGATGAAATTGATAAACATTACCGATCCCCGGGTTA ATTAA-3') and RBS1\_R1 (5'-ACTAGAATTCTATAAACCGTTACGTAATTTTCGCTATGTATAGTTCACTCCTC GAATTCGAGCTCGTTAAC-3') (4). Deletion of the respective restriction fragments from the YEpl181-*RBS1* resulted in derivative plasmids, called here [*RBS1* ΔC1] and [*RBS1* ΔC2], containing deletion of fragments of *RBS1* encoding 231-332 aa and 231-457 aa, respectively. Fragments containing 211 bp upstream and 154 bp, 231 bp or 253 bp downstream of the *RPB10* ORF were amplified by PCR the [*RPB10*] plasmid DNA by using RPBF (5'-GATGGCTACTACACTGGAAG-3') and RPBR (5'-CCTACAGTATGCAGAGACAC-3'), 231RpbR (5'-ACTTCCTTATCGTCTTGAAGAGT-3') or 253RpbR (5'-TCGCACGATGTAACATCTACA-3') primers, respectively. PCR products were introduced to pDrive vector (Qiagen), cut by restriction enzyme EcoRI and cloned into EcoRI site of centromeric plasmid pRS316 (*URA3 CEN6*) resulting in plasmids called here [*RPB10* Δ3'Δ3' 154], [*RPB10* Δ5'Δ3' 231] or [*RPB10* Δ5'Δ3' 253]. Both [*RPB10* Δ3' 154] and [*RPB10* Δ5'] plasmids were constructed in the same way. [*RPB10* Δ3' 154] plasmid containing 663 bp upstream and 154 bp downstream of the *RPB10* ORF was amplified by PCR from the pFL44L-*RPB10a* plasmid (5) by using MGF (5'-AGACAGCAGCATGCATCG-3') and RPBR (described above) primers. [*RPB10* Δ5'] plasmid containing 211 bp upstream and 647 bp downstream of the *RPB10* ORF was amplified by PCR from the pMJ18 plasmid (1) with RPBF (described above) and MGMR (5'-ATTGACCTCCACAGCTG-3') primers. HA-epitope sequence fused at N-terminus of *RPB10* gene with a 5' and 3' regulatory regions, 250 bp and 390 bp, respectively (synthesized by Syngen biotech company) in pEX-A128 vector was cloned to pRS316 in NotI and EcoRI sites, called here [HA-*RPB10*].

### RNA isolation and northern hybridization

Total RNA was isolated by heating and freezing the cells in the presence of SDS and phenol as described previously (6). Samples containing 20 µg of total RNA were denatured in a glyoxal reaction mixture at 55°C for 1 h (7) and were resolved by electrophoresis in 1.2 % agarose gel in 1×BPTE buffer (10 mM PIPES, 30 mM Bis-Tris, 1 mM EDTA pH 8.0). RNA samples were transferred into a Hybond-N+ membrane (Amersham) with 10×SSC using the TurboBlotter downward capillary transfer system (Schleicher & Schull) and crosslinked by UV radiation (0.14 J/cm<sup>2</sup>). The blot was prehybridized for 3 h at 65°C in buffer containing 7% SDS, 0.5 M Na<sub>2</sub>HPO<sub>4</sub> pH 7.4, 1 mM EDTA, 1% BSA, and hybridized with *RPB10*, *ACT1* and *SCR1* probes. DNA probes were amplified by PCR using oligonucleotides listed in the Supplementary Table 1 and were labeled with [ $\alpha$ -<sup>32</sup>P]-dATP by random priming using the HexaLabel DNA labeling kit (Fermentas). Hybridization was carried out overnight at 65°C in the same buffer as prehybridization. Filters were washed three times (15 min each) with 2×SSC, 0.1% SDS at 65°C. Hybridization signals were exposed to a phosphorimager screen. RNA was quantified using the PhosphorImager STORM 820 (Molecular Dynamics). Band intensities were quantified using the MultiGauge v3.0 software (Fujifilm).

**Supplementary Table 1.** Oligonucleotides for northern hybridization

Target	Forward	Reverse
RPB10N	ATCAGCACTCCGATGGCTAC	ACAAGTCTCTTATATCGCACGA
ACT1N	TTCCCATCTATCGTCGGTAG	GTGGTGGAGAAAGAGTAACC

SCR1N	TCTGGTGGGATGGGATACGT	TCAGGACACACTCCATCCCC
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### cDNA synthesis and reverse transcription-quantitative PCR (RT-qPCR)

100 nanograms of RNA was used for cDNA synthesis using a QuantiTect reverse transcriptase kit (Qiagen). cDNA for each sample was performed according to the manufacturer's instruction. RT-qPCR reactions contained 1 µL of cDNA template, 300 nM primer pairs and 5 µL of RT PCR Mix SYBR (A&A Biotechnology). Quantitative PCR was performed on a Roche LightCycler 480 System using a 5-min incubation at 95°C, followed by 40 cycles of 30 s at 95°C, 20 s at 60°C, and 20 s at 72°C (with a plate read after each cycle). A melting curve analysis was performed for each sample after PCR amplification to ensure that a single product with the expected melting curve characteristics was obtained. Each sample was loaded in triplicate. Each plate contained cDNA dilutions for the standard curve, a non-reverse transcriptase control, and a no template control. PCR efficiencies were between 90% and 100%. Data were processed in LightCycler 480 Software and then analyzed in Excel (Microsoft). Data are expressed in arbitrary units calculated from standard curve where the highest cDNA concentration was set to 1. The primer sequences are listed in the Supplementary Table 2.

**Supplementary Table 2.** Oligonucleotides for RT-qPCR

Target	Forward	Reverse
<i>RPB10</i>	TGTTGGTGACAAGTGGAAA	GATCGACGTGGTTAGAACATCA
<i>PGA1</i>	ACCACTCGGCTAGGTATCCC	CGTTCAATACGTGTGGTGCC
<i>MSH1</i>	ACTTGGTCGTGGTTCAAGCA	CGTTGCCTCCGTTGAAGTA
<i>SPO16</i>	ATTCGCAGCTAACGAGGA	GGAACCTCCAGGGCTTTCTG
<i>CNN1</i>	AAGCATTGGAATGGATCAGG	TCCTCTTATCGTCGCTTGG
<i>pre-RPL28</i>	CCATCTCACTGTTGAGACGG	CTCAGTTGCGATGGAAGAG
<i>RPL28</i>	TCACGTCTCAGCCGGTAAAG	ATGTTGACCACCGGCCATAC
<i>ACT1</i>	CATGTTCCCAGGTATTGCCGA	GTCAAAGAAGCCAAGATAGA
<i>SCR1</i>	GAGAATTCTGGCCGAGGAACAAA	TCTGCCAGGACAAATTACGA

### Protein extraction and western blot analysis

The protein extraction method was described earlier (8). Protein extracts were separated by 6% or 10% sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE). After electrophoresis proteins were transferred onto the nitrocellulose membrane (Millipore), which was then blocked in TBST (10mM Tris, 150 mM NaCl, 0.05% Tween 20) containing 5% fat-free dry milk for 30 min and subsequently incubated with the appropriate antibody: mouse monoclonal antibodies 9E10 anti-Myc (Roche) at a 1:2,000 dilution for 1 h, anti-HA (Covance) at a 1:5,000 dilution for 2h, anti-Pgk1 (Abcam) at a 1:20,000 dilution for 1 h, anti-Nab2 (gift from Torben Heick Jensen) at a 1:1,000,000 dilution for 1 h and rabbit polyclonal antibodies anti-Rbs1 (Gramsh) at a 1:1,000 dilution for 1h. Blots with TAP-tagged Upf1 protein were incubated with anti-PAP antibody at a 1:3,000 dilution. PAP is a peroxidase anti-peroxidase antibody which does not necessitate the use of a secondary antibody. Then membrane was incubated for 1 h with a secondary anti-mouse or anti-rabbit antibody coupled to horseradish

peroxidase (DAKO) at a dilution of 1:5,000. The signal on the membrane was visualized by chemiluminescence using the ECL detection kit (Bio-Rad).

### CRAC analysis

Samples were processed as previously described (PMID: 30718516, PMID: 32585128) with modifications. Cells were lysed in TNMC100 (50 mM Tris-HCl pH 7.5, 150 mM NaCl, 0.1% NP-40, 5 mM MgCl<sub>2</sub>, 10 mM CaCl<sub>2</sub>, 20U of DNase RQ1 and a protease-inhibitor cocktail (1 tablet / 50 mL) with zirconia beads in a 50 mL conical. The cells were lysed with five one-minute pulses, with cooling on ice in between. The supernatant was spun for 20 minutes at 21,000g. The cleared lysate was incubated with the IgG Sepharose for two hours at 4°C, with nutating. Subsequently, the beads were washed two times with TMN600 (50 mM Tris-HCl pH 7.5, 600 mM NaCl, 0.1% NP-40, 1.5 mM MgCl<sub>2</sub>) and two times TMN100 (50 mM Tris-HCl pH 7.5, 100 mM NaCl, 0.1% NP-40, 5 mM MgCl<sub>2</sub>). For RNA digestion beads were resuspended in 600 µL TMN100 containing 0.04U of RNase-IT and samples were incubated for 10 minutes at 23 °C 1000 rpm to fragment protein-bound RNA. RNase digestion was slowed down by incubation on ice for 3 min.

The beads were washed three times TMN100 and resuspended in 600 µL TMN100 containing 5 µL HaloTEV (Promega). Protein: RNA complexes were eluted by incubation with HaloTEV for 2h at 18°C with shaking. The supernatant was separated and adjusted for nickel affinity purification with the addition of 400 mg guanidine hydrochloride, 45 µL NaCl (3M) and 7 µL imidazole (1 M) and added to 50 µL of washed nickel beads (Qiagen).

Following 2h incubation, the nickel beads were washed three times with WBI (6.0 M guanidine hydrochloride, 50 mM Tris-HCl pH 7.5, 300 mM NaCl, 0.1% NP-40, 10 mM imidazole, 1.5 mM MgCl<sub>2</sub>), three times with C buffer (50mM Tris-HCl pH 7.5, 50 mM NaCl, 1.5 mM MgCl<sub>2</sub>, 0.1% IGEPAL CA-630) and twice with 50 mM Bis-Tris pH 6.5 and transferred to a spin column. Subsequent reactions (80 µL total volume for each) were performed in the columns, and afterward washed once with WBI and three times with PNK buffer:

1. Dephosphorylation (70 mM Bis-Tris pH 6.5, 10 MgCl<sub>2</sub>, 40 U T4 PNK (NEB), 2 U TSAP (Promega); 37°C 30 min)
2. 3' linker ligation (70 mM Tris-HCl pH 7.5, MgCl<sub>2</sub>, 12.5% PEG8000, 40U T4 RNA Ligase II truncated K227Q, 80U RNasin, 80 pmol preadenylated 3' miRCat-33 linker (IDT); 16°C overnight).
3. 5' end phosphorylation and radiolabeling (70 mM Tris-HCl pH 7.5, MgCl<sub>2</sub>, 40 U T4 PNK (NEB), 80U RNasin, 40 µCi <sup>32</sup>P-γATP; 37°C for 45 min, with addition of 100 nmol of ATP after 30 min).
4. 5' linker ligation (70 mM Tris-HCl pH 7.5, MgCl<sub>2</sub>, 12.5% PEG8000, 40 U T4 RNA ligase I (NEB), 80 U RNasin, linker, 200 pmol 5' linker, 1 mM ATP; 16°C for 3h and 25°C for 2h).

The beads were washed twice with WBI and three times with C buffer. Protein: RNA complexes were eluted 100 µL of elution buffer (50 mM Tris-HCl pH 7.5, 500 mM NaCl, 0.1% Triton X-100, 6M guanidine hydrochloride and 300 mM imidazole), supplemented 3 µL GlycoBlue and ethanol precipitated overnight (9 volumes of ethanol). RNPs were pelleted at 21000g for 20 minutes, washed twice 80% acetone, rehydrated in 12 µL water. 4 µL 4X NuPAGE sample loading buffer supplemented with 8% β-mercaptoethanol. The sample was denatured by incubation at 65°C for 10 minutes, and run on a 4%–12% Bis-tris NuPAGE gel at 130 V. The protein: RNA complexes were transferred to Hybond-C nitrocellulose membranes with NuPAGE MOPS transfer buffer for 2 h at 100V.

Labelled RNA was detected by autoradiography. The appropriate region was excised from the membrane and treated with 0.2 µg/µL Proteinase K (50 mM Tris-HCl pH 7.5, 50 mM NaCl,

0.1% IGEPAL CA-630, 1% SDS, 5 mM EDTA; 2 hr 55°C with shaking) in a 500 µL reaction. The RNA component was isolated with a standard phenol:chloroform extraction followed by ethanol precipitation with 2 µL of GlycoBlue. The RNA was reverse transcribed using Superscript IV and the miRCat-33 RT oligo (IDT) for 15 min at 50°C in a 20µL reaction. Subsequently 2 µL ExoI (NEB) were added, incubated 30 min at 37°C and heat inactivated 20 min at 80°C. The resulting cDNA was amplified by PCR in 50 µL reactions using Phusion (Thermo) (2 µL template, 19 cycles) PCR reactions were combined, purified and concentrated using AmpureXP beads, and resolved on a 3% Metaphore agarose gel. A region corresponding to 150 to 250 bp was excised from the gel and extracted using the Zumoclean Gel DNA Recovery Kit (Zymo Research). Libraries were measured with Qbit and sequenced using Illumina NextSeq 550 with 150bp single-end reads.

## Quantification and statistical analysis of CRAC data

### *Pre-processing and data alignment*

Illumina sequencing data were demultiplexed using in-line barcodes and in this form were submitted to GEO. First quality control step was performed using FastQC software (<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>) considering specificity of CRAC data. Raw reads were trimmed with flexbar v3.4.0 (Dodd et al., 2012) with parameters –q TAIL –qf i1.8 –qt 20 to remove bases with QC<20. Subsequently reads were collapsed to remove PCR duplicates using FASTX-collapser v0.0.14 ([http://hannonlab.cshl.edu/fastx\\_toolkit/](http://hannonlab.cshl.edu/fastx_toolkit/)) then inline barcodes were removed using pyBarcodeFilter.py script from pyCRAC package v3.0 (9). The 3' adapter were removed using flexbar v3.4.0 (10) with parameters –as TGGAATTCTCGGGTGCCAAGGC -u 3 -m 17 -n 16 -bt RIGHT.

All datasets were aligned to the yeast genome using Novoalign v2.07.00 (<http://www.novocraft.com>) with parameter –r random and saved in novo format to calculate classes of bound RNAs and BigWig files for visual inspection in IGV genome browser. Second quality control step was performed using pyReadCounters script (pyCRAC package) which calculates overlaps between aligned cDNAs and yeast genomic features. The 1 nt resolution BigWig files were generated using bamCoverage v3.1.3 script from deepTools package (11). Sam file operations were performed using SAMtools v1.9 (12).

Additionally all datasets were aligned to yeast transcriptome with parameter using STAR aligner v.2.7.3a (13) and only unambiguously mapped reads were used to generate binding profiles.

Downstream analyses were performed using python 3.6.10 Jupiter notebooks, python libraries (pandas v0.19.2, numpy v1.16.0, scipy v1.2.0, matplotlib v2.2.3) and in-house scripts submitted available s an update of gwide toolkit v0.5.27 (<https://github.com/tturowski/gwide>) (14).

### *Identifying Rbs1 enriched genes*

To compare CRAC data and RNA-seq all reads mapping to mRNAs were normalized to reads per million (RPM) and converted to log2 RPM. For each transcript p-value was calculated using a two-sided T-test. High confidence Rbs1 targets were selected using following criteria: p-value <0.01, ratio between Rbs1 binding and RNA-seq >1.5 and >128 uniquely mapped RPM.

### *Rbs1 metagene representation*

For metagene analysis data were processed independently for each replicate. Reads mapped to each transcript were summed up to 1 and fraction of reads was used further. This excluded risk that the obtained profile is biased by the most abundant transcripts, as each transcript has a value of 1. To combine transcripts of different length, for metagene representation fraction of

reads was binned as follows: 5'UTR 10 bins, CDS 100 bins and 3'UTR 10 bins. Average fraction of reads was plotted for each bin.

### ***Binding to mature mRNA***

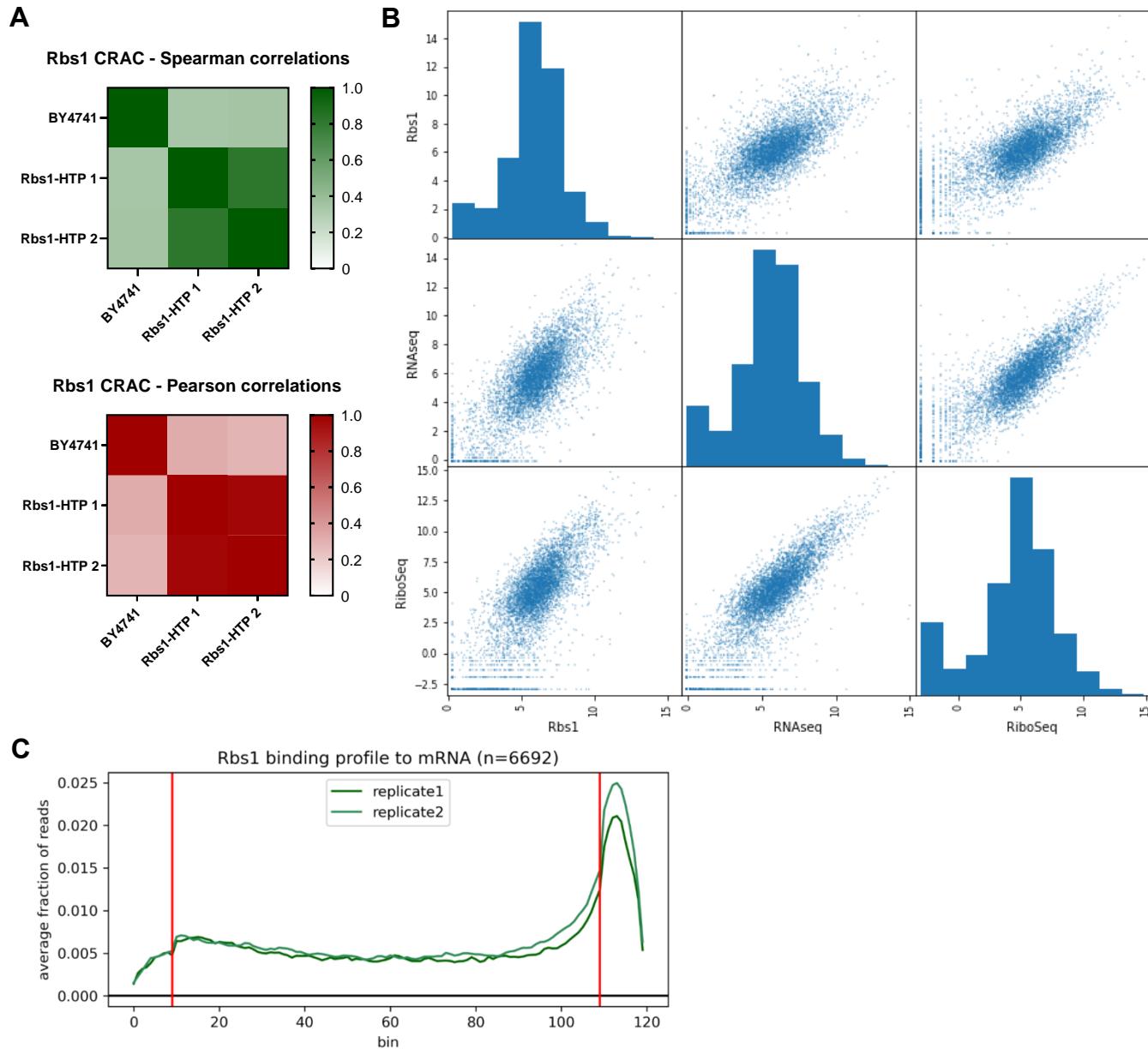
Reads mapped to each transcript were summed up to 1 and fraction of reads was used further. Fraction of reads mapping to each feature was counted. Boxplots present 2<sup>nd</sup> and 3<sup>rd</sup> quartile, line marks median and whiskers range between 5<sup>th</sup> and 95<sup>th</sup> percentile.

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## SUPPLEMENTARY FIGURES



**Supplementary Figure S1.** Statistical analysis of CRAC data for Rbs1 over all mature mRNAs. (A) Spearman and Pearson correlations between datasets. (B) Scatter plots comparing the Rbs1 RNA binding to RNA-seq and Ribo-seq data. Reads uniquely mapping to mRNAs are presented (log2 RPM). (C) Metagene representation of read density over all mature mRNAs (n=6692). Data were separated into 120 bins: 10 for the 5' UTR, 100 for the CDS and 10 for the 3' UTR. Horizontal lines indicate where CDS starts and stops. Metagene analysis performed for mRNA containing at least 100 reads in Rbs1 CRAC data (n=6692).

**A**

XP\_010757305.1 hypothetical protein PADG\_02291 [Paracoccidioides brasiliensis Pb18]  
RMJ22032.1 R3H domain protein [Aspergillus sp. HF37]  
QGA13340.1 hypothetical protein EYB26\_000988 [Talaromyces marneffei]  
OQE15951.1 hypothetical protein PENSTE\_c026G04122 [Penicillium steckii]  
XP\_002585083.1 predicted protein [Uncinocarpus reesii 1704]  
PGH19146.1 hypothetical protein A780\_04226 [Polytolypa hystricis UAMH7299]  
XP\_033404196.1 uncharacterized protein G1Q15\_05635 [Arthroderma uncinatum]  
KAF192399143.1 hypothetical protein EJ06DRAFT\_72354 [Trichodelitschia bisporula]  
KAF1939774.1 hypothetical protein EJ02DRAFT\_407933 [Clathrospora elynae]  
ELQ33482.1 hypothetical protein OOU\_Y34sccaffold00936g9 [Pyricularia oryzae Y34]  
TQS37745.1 hypothetical protein Golomagni\_01771 [Golovinomyces magnicellulatus]  
SZF03220.1 unnamed protein product [Blumeria graminis f. sp. hordei]  
KFZ15736.1 hypothetical protein V502\_05427 [Pseudogymnoascus sp. VKM F-4520 (FW-2644)]  
XP\_016645040.1 R3H domain-containing protein [Scedosporium apiospermum]  
KHF40507.1 R3H domain-containing protein-like protein [Acromonium chrysogenum ATCC 11550]  
CCE34041.1 uncharacterized protein CPUR\_07972 [Claviceps purpurea 20.1]  
CEJ80227.1 hypothetical protein VHEMI00424 [Torrubiella hemipterigena]  
KAB5577609.1 hypothetical protein GE09DRAFT\_1213834 [Coniochaeta sp. 2T2.1]  
XP\_014168719.1 r3h domain protein [Grosmannia clavigera kw1407]  
PSR74187.1 hypothetical protein BD289DRAFT\_226619 [Coniella lustricola]  
VBB75180.1 Putative protein of unknown function [Podospora comata]  
XP\_009848197.1 hypothetical protein NEUTE1DRAFT\_144342 [Neurospora tetrasperma FGSC 2508]  
KIH88101.1 r3h domain protein [Sporothrix brasiliensis 5110]  
XP\_030994999.1 uncharacterized protein EOL32\_006261 [Phialemoniopsis curvata]  
KXJ91167.1 hypothetical protein MicholqCDRAFT\_119375 [Microdochium bolleyi]  
KKA30932.1 hypothetical protein TD95\_004645 [Thielaviopsis punctulata]  
AJV06700.1 Rbs1p [Saccharomyces cerevisiae YJM1418]  
GCF00085.1 hypothetical protein ZYGM\_002314 [Zygosaccharomyces mellis]  
OVA05989.1 Single-stranded nucleic acid binding R3H [Macleaya cordata]  
GEU59184.1 R3H domain-containing protein 1-like [Tanacetum cinerariifolium]  
OWM87646.1 hypothetical protein CDL15\_Pgr022759 [Punica granatum]  
XP\_008808576.1 uncharacterized protein LOC103720587 [Phoenix dactylifera]  
MQL81330.1 hypothetical protein Taro\_013800, partial [Colocasia esculenta]  
XP\_020699935.1 R3H domain-containing protein 1 [Dendrobium catenatum]  
MQM11394.1 hypothetical protein Taro\_044302 [Colocasia esculenta]  
KMZ62657.1 Single-stranded nucleic acid binding R3H protein [Zostera marina]  
PKA67145.1 hypothetical protein AXF42\_Ash004637 [Apostasia shenzhenica]  
XP\_009386294.1 R3H domain-containing protein 2 [Musa acuminata subsp. malaccensis]  
XP\_020254643.1 uncharacterized protein LOC109831676 isoform X1 [Asparagus officinalis]  
XP\_020254456.1 cAMP-regulated phosphoprotein 21-like isoform X2 [Asparagus officinalis]  
OAY67162.1 R3H domain-containing protein 1 [Ananas comosus]  
KAF3787699.1 R3H domain-containing protein 1 [Nymphaea thermarum]  
XP\_023741289.1 R3H domain-containing protein 2 isoform X2 [Lactuca sativa]  
KAE9465814.1 hypothetical protein C3L33\_02274, partial [Rhododendron williamsianum]  
KAE8697742.1 Single-stranded nucleic acid binding R3H protein isoform 2 [Hibiscus syriacus]  
XP\_006402961.1 R3H domain-containing protein 2 [Eutrema salugineum]  
XP\_010552904.1 uncharacterized protein LOC104823164 [Tarenaya hassleriana]  
XP\_022966122.1 uncharacterized protein LOC111465900 isoform X1 [Cucurbita maxima]  
GER51925.1 single-stranded nucleic acid binding R3H protein [Striga asiatica]  
XP\_020114255.1 uncharacterized protein LOC109728293 isoform X1 [Ananas comosus]  
ERM99095.1 hypothetical protein AMTR\_s00101p00122520 [Amborella trichopoda]  
XP\_006654401.1 uncharacterized protein LOC102704805 isoform X1 [Oryza brachyantha]  
KAF3341400.1 R3H domain-containing protein 1 [Carex littledalei]  
XP\_034191896.1 R3H domain-containing protein 1-like isoform X3 [Osmia lignaria]  
XP\_024218457.1 cAMP-regulated phosphoprotein 21 isoform X2 [Halymorpha halys]  
GFG30713.1 hypothetical protein Cfor\_07482, partial [Coptotermes formosanus]  
XP\_015115923.1 R3H domain-containing protein 1 isoform X3 [Diachasma alloeum]  
XP\_029828169.1 R3H domain-containing protein 2 [Ixodes scapularis]  
XP\_022254084.1 cAMP-regulated phosphoprotein 21-like isoform X2 [Limulus polyphemus]  
XP\_015903829.1 cAMP-regulated phosphoprotein 21-like [Parasteatoda tepidariorum]  
XP\_023222768.1 R3H domain-containing protein 2-like [Centruroides sculpturatus]  
XP\_002423078.1 R3H domain-containing protein, putative [Pediculus humanus corporis]  
XP\_019865105.1 cAMP-regulated phosphoprotein 21 [Aethina tumida]  
XP\_022901769.1 cAMP-regulated phosphoprotein 21 isoform X2 [Ontophagus taurus]  
XP\_017778862.1 cAMP-regulated phosphoprotein 21 isoform X4 [Nicrophorus vespilloides]  
VEN60483.1 unnamed protein product [Callosobruchus maculatus]  
KAF2898948.1 hypothetical protein ILUMI\_07232 [Ignelater luminosus]  
XP\_030766515.1 cAMP-regulated phosphoprotein 21 isoform X3 [Sitophilus oryzae]  
XP\_018019452.1 cAMP-regulated phosphoprotein 21-like [Hyalella azteca]  
XP\_014672054.1 cAMP-regulated phosphoprotein 21-like [Priapulus caudatus]  
XP\_014675087.1 cAMP-regulated phosphoprotein 21-like, partial [Priapulus caudatus]  
XP\_030385838.1 protein encore [Scaptodrosophila lebanonensis]  
XP\_029725418.1 protein encore isoform X2 [Aedes albopictus]  
XP\_032599088.1 protein encore isoform X4 [Drosophila grimshawi]  
KNC31935.1 Protein encore, partial [Lucilia cuprina]  
XP\_020714208.1 protein encore isoform X4 [Ceratitis capitata]  
XP\_023246188.1 R3H domain-containing protein 1 [Copidosoma floridanum]  
XP\_023329389.1 cAMP-regulated phosphoprotein 21-like [Eurytemora affinis]  
XP\_029201356.1 cAMP-regulated phosphoprotein 21-like [Acropora millepora]  
XP\_021368844.1 cAMP-regulated phosphoprotein 21-like [Mizuhoplecten yessoensis].  
XP\_034305772.1 cAMP-regulated phosphoprotein 21 isoform X6 [Crassostrea gigas]  
XP\_013815153.1 R3H domain-containing protein 1 isoform X3 [Apteryx mantelli mantelli]  
XP\_016861070 ARPP-21/cAMP-regulated phosphoprotein 21 isoform X3 [Homo sapiens]

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6

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Consensus ss:
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Consensus ss:

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-----CNTFYRMLAHLRLADYYLLGHVVDT-----TMTG--VKIYRTPYCRIPPPVS-
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-----CNSFCRMITHKLADYYHLTQVDA-----VAGA--VRIYRTPFCRLPSSLT-
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-----CNSFCRMITHKLADYYHMTHSYEP-----SIGA--VRIYRTPFCRVPOSIA-
-----SNSFCRMILAHLKLADYYHMTHSFEP-----NIGV--VRIYRTPFCRVPTSLA-
-----SNSFCRMITHKLADYYHMTHSYEP-----HVGS--VRIYRTPFCRVPPSLA-
-----SNSFCRMITHKLADYYHMTHSYEP-----HIGS--VRIYRTPFCRVPPSLA-
-----CNSFCRMITHKLADYYHMTHSFEA-----VAGA--VRIYRTPFCRVAPPLS-
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-----SNSFCRMITHKLADYYHMTHSFES-----QAGA--VRIYRTPFCRIPPLS-
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-----LPTSYLRAAHRVAQHYCLQSMVLDNNLPD-GSSGR--IIVRKTSCERFPVIRL-
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Consensus aa:  
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 ALHAA---NNGTPP-----QA-----MPAMKI  
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 MLHPA---NTNTTP-----PS-----APAMKI  
 GFSTP---STVANTP-----PPA-----VQARKI  
 QMVDP---AKGNTNP-----PAE-----LPARKI  
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 SIAE---STSAAAN-----TPPPV-----VIPRKI  
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 LMVPE---MSAATSS-----TPPPA-----VLPRKI  
 GTIV---NASSNS-----SPAPA-----ILPRKI  
 SLVAV---NPSSN-----TPPFT-----IQPKKI  
 SIAQ---SASTAS-----TPPPP-----VMRKI  
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 TIAP---STSNTS-----SPAPA-----VIPRKI  
 SFVPV---NTPESTN-----APPOI-----VKPRKI  
 SIAAA---NETSS-----TPPPA-----LLPRKI  
 SIMETTVPEEPEAP-----APPLA-----MLPRKI  
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 CDIVP---SKQAH-----GKNRSTTP-QSKPRQVKI  
 LQELQ---LNKKPEE-----CISSSESIEKSNNNRIFRI  
 SYGRT---DATATNT-----NS-----NKRYRI  
 ADIPI---SLPQEES-----NS-----VVKVVAI  
 ADIPV---NIPTEDI-----SSS-----VVKVVAI  
 ADIPV---NLPEEK-----SAVKFI  
 SDVPA---KQTENEK-----TE-----HFKIAI  
 SDVPA---KQQECDK-----SE-----QVKIVM  
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 SEIPV---KQLENDK-----HE-----PIKIFM  
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 SEIPT---AKQSENGK-----FE-----HMVKVS  
 SEIPA---KQPENDK-----PE-----VMKIAI  
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 SQVPA---KDSENNK-----LD-----QIKIVI  
 SEVSA---QQPEKCI-----AE-----KIKIAI  
 CEIPA---KMSETDK-----PE-----HVKIAI  
 SEVPS---KQTRTEH-----EAA-----EKLKFV  
 QKFLA---SSRNHD-----ETA-----EKLKFV  
 SKILP---KQSEG-----EAS-----DKKKIV  
 KEHIR---DDLILSE-----EPRRSI  
 RDFVR---DDLLPPE-----EPRRSI  
 RDHIR---EEFLFPE-----EPRRSI  
 KEHIR---DDLLTE-----EPRRSI  
 RDQIR---DDLLSE-----EPRRSI  
 NDLIH---DELVPD-----EPKKLI  
 QDFIK---DELLPE-----EPKKSI  
 REQIH---DELVPD-----EPKKSI  
 GELIR---DDFFRSE-----EPRRCI  
 KEHIK---EDIVFAE-----EPRRSI  
 ESYIR---DDIGFTE-----EPRRSI  
 KHHIK---EDIVFSE-----EPRRSI  
 KELIR---EDIVFAN-----EPRRSI  
 KTHLK---DDFFGE-----EPRRSI  
 KEHIK---EIFNDE-----PPRSI  
 REHFC---DEMPLP-----DD-----LPRRSV  
 RQLIR---DDSLGLD-----EPKRL  
 MQLIR---EDASLD-----EPKRL  
 RSLVR---VDHRDD-----SRKSI  
 KTLIN---ESFSEE-----PRKSI  
 QSLV-----RDD-----ARKSI  
 KSLVR---VDHRDD-----ARKSI  
 KSLVR---DHRDD-----SRKSI  
 KEHIR---DDLLHFV-----VDNNGKSSDEASVARQKSI  
 RDHVP---EDAVLST-----S-----EPRRLI  
 LELIP---REESEDL-----EG-----AVPRLI  
 KEHVD---ARDSQK-----KITGI  
 KEHID---NLDSAP-----KVSKI  
 CEHIK---DEKSDD-----FQKRYI  
 CEHLK---DERGEE-----SQRFI  
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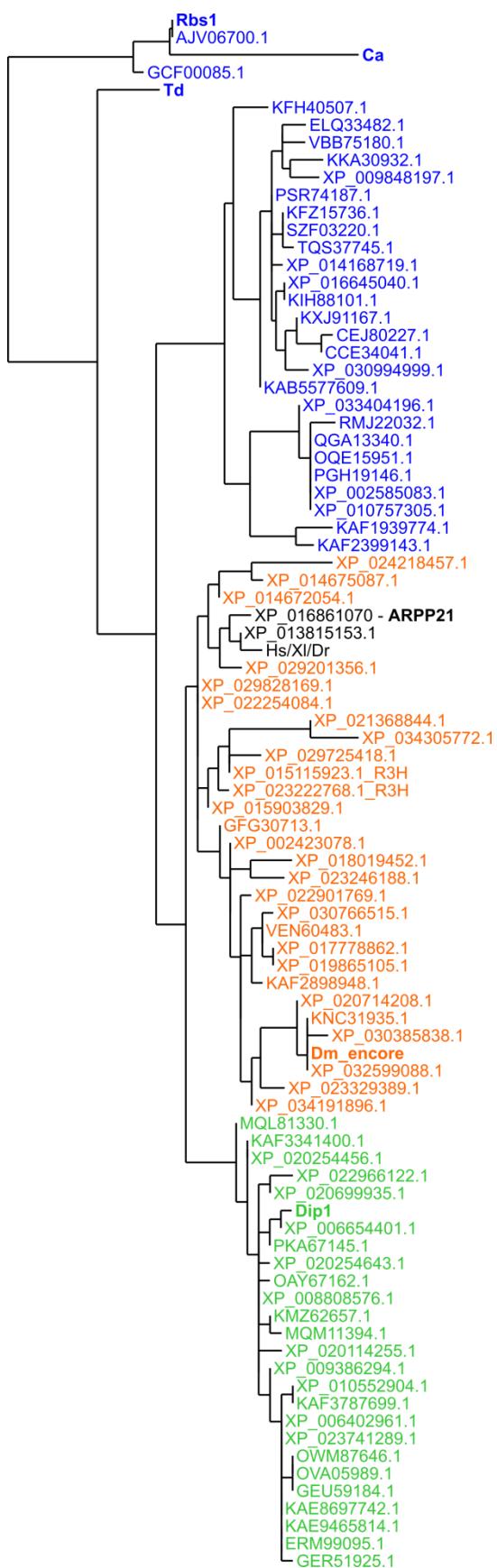
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 ELQ33482.1  
 TQS37745.1  
 SZF03220.1  
 KFZ15736.1  
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 CCE34041.1  
 CEJ80227.1  
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 XP\_014168719.1  
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 VBB75180.1  
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 Td  
 AJV06700.1  
 GCF00085.1  
 OVA05989.1  
 GEU59184.1  
 OEM87646.1  
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 XP\_022966122.1  
 GER51925.1  
 XP\_020114255.1  
 ERM99095.1  
 XP\_006654401.1  
 Dip1  
 KAF3341400.1  
 XP\_034191896.1  
 XP\_024218457.1  
 GFG30713.1  
 XP\_015115923.1  
 XP\_029828169.1  
 XP\_022254084.1  
 XP\_015903829.1  
 XP\_023222768.1  
 XP\_002423078.1  
 XP\_019865105.1  
 XP\_022901769.1  
 XP\_017778862.1  
 VEN60483.1  
 KAF2898948.1  
 XP\_030766515.1  
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 XP\_014672054.1  
 XP\_014675087.1  
 XP\_030385838.1  
 XP\_029725418.1  
 Dm\_encore  
 XP\_032599088.1  
 KNC31935.1  
 XP\_020714208.1  
 XP\_023246188.1  
 XP\_023329389.1  
 XP\_029201356.1  
 XP\_021368844.1  
 XP\_034305772.1  
 XP\_013815153.1  
 XP\_016861070\_ARPP21  
 Hs/X1/dr  
**Consensus aa:**  
**Consensus ss:**

@

hh

174

Rbs1 E-----QERIEKERLYEQRKQEIFFDKLN  
 Ca E-----SQRASKEALYMKLREEIFENN  
 XP\_010757305.1 T-----MSREEREAKEYQEAKERIFRDFP  
 RMJ22032.1 I-----QTREREAKYNEVRERIFRDFP  
 QGA13340.1 T-----MTREEREAKEYQEAKERIFRDFP  
 OQE115951.1 T-----LTREEREAKEYQEAKERIFRDFP  
 XP\_002585083.1 A-----LTREEREAKEYQEAKERIFRDFP  
 PGH19146.1 A-----LTREEREAKEYQEAKERIFRDFP  
 XP\_033404196.1 N-----LTREEREAKEYQEAKERIFRDFP  
 KAF2399143.1 K-----MTREEREAKEYEARQRIFGNVD  
 KAF1939774.1 A-----LSRECREARYEARQRIFGSAE  
 ELQ33482.1 K-----LTREEREEAYNKARLIFGSSA  
 TQS37745.1 KSIPLTLSREEREEAYNKARLIFGKEE  
 SZF03220.1 K-----LSREEREAAYNKARLIFGKDE  
 KFZ15736.1 K-----LSREEREAAYNKARLIFGNT  
 XP\_016645040.1 K-----MTQEREEREEAYNRARERIFGSSE  
 KFH40507.1 K-----LTREEREEVYRAERERIFGSS--  
 CCE34041.1 K-----LSREEREEEMYKLARERIFGSS--  
 CEJ80227.1 K-----LSRECREEMYKLARERIFGNN--  
 KAB5577609.1 K-----LTREEREEKYNKARLIFGSSVE  
 XP\_014168719.1 T-----LSREEREEAYNKARLIFGSSM  
 PSR74187.1 K-----MSREEREEAYNKARLIFGTT  
 VBB75180.1 K-----MTREEREEAYNKTRQRIFGSSE  
 XP\_009848197.1 R-----MTREEREEAYNRARQRIFGNMK  
 KIH88101.1 K-----MTREEREEAYNRARERIFAM--  
 XP\_030994999.1 K-----LSREEREQKYNQARERIFGSSAA  
 KXJ91167.1 K-----LTREEREEAYQARERIFGKED  
 KKA30932.1 R-----KTREREEREEAYNEARKRIFGSSN  
 Td E-----DKLEKEERYKAARARIFQGVE  
 AJV06700.1 E-----QERIEKERLYEQRKQEIFFDKLN  
 GCF00085.1 E-----QRMEKEIQQYQRKQEIFFSAPK  
 OVA005989.1 A-----KSVEERKEEYNRARARIFSSNS  
 GEU59184.1 L-----KSVEERKEEYNRARARIFNSSS-  
 OWM87646.1 A-----KSVEERKEEYNRARARIFFNNSN  
 XP\_008808576.1 A-----RTVEERKEEYDALARIFSGSS  
 MQL81330.1 L-----RTVEEREEYDALARIFSDSS  
 XP\_020699935.1 M-----RTVEERMEEYDALARIFSCSS  
 MQM11394.1 S-----RTMEEEREEDYDALARIFSGCI  
 KMZ62657.1 V-----RSVEERKEDYDALARIFNNGS  
 PKA67145.1 L-----RTVEERKEEYDALARIFNDSF  
 XP\_009386294.1 T-----RTVEERKEEYDRARARIFSGSC  
 XP\_020254643.1 M-----RTVEERKEEYEKARARIFNS--  
 XP\_020254456.1 M-----RTVEEREEYDALARIFNGSS  
 OAY67162.1 L-----RTVEERKEEYDALARIFSGTS  
 KAF3787699.1 V-----RSVEERKEEYDALARIFSSSS  
 XP\_023741289.1 V-----RSVEERKEEYDRARARIFSTP-  
 KAE9465814.1 A-----RTVEERKEEYDRARARIFSGSS  
 KAE8697742.1 V-----RSVEERKEEYDRARARIFSSPS  
 XP\_006402961.1 L-----RSVEERKEEYDRARARIFNDLT  
 XP\_010552904.1 T-----RSVEERKEEYDALARIFNSPT  
 XP\_022966122.1 G-----RTVEERMEEYDALARIFSRHG  
 GER51925.1 V-----RSVEERREEYDALARIFNRPN  
 XP\_020114255.1 L-----RTVEERKEEYDALARIFSGSN  
 ERM99095.1 V-----RSVEERKEEYDRARARIFNGSG  
 XP\_006654401.1 A-----KTREREEREEYKARARIFNGS-  
 Dip1 P-----KTVDERIEEYNKARARIFNGSI  
 KAF3341400.1 M-----KSIEEREEEYDALARIFSGSS  
 XP\_034191896.1 N-----KSFEEREEEYERARRRIFKDS-  
 XP\_024218457.1 S-----KSFEEREEEYKIEGEIDCIDD  
 GFG30713.1 S-----KSFEEREEEYEKARKRIFNRED  
 XP\_015115923.1 S-----KSFEEREEEYEVKVRRIFKD--  
 XP\_029828169.1 S-----KSIEEREEEYEKARARIFNQD-  
 XP\_022254084.1 S-----KSIEEREEEYEKARARIFNQDS  
 XP\_015903829.1 S-----KSFEEREEEYEVKARARIFNQDE  
 XP\_023222768.1 S-----KSFEEREEEYEVKVRARIFQKDE  
 XP\_002423078.1 S-----KSFEEREEEYERARKRIFNRRDD  
 XP\_019865105.1 S-----KSFEEREEEYEVKVRRIFNRE-  
 XP\_022901769.1 S-----KSIEEREEEYEVKARRRIFNPK  
 XP\_017778862.1 S-----KSFEEREEEYEVKVRRIFNKEM  
 VEN60483.1 S-----KSFEEREEEYEVKARRRIFNREH  
 KAF2898948.1 S-----KSIEEREEEYERAKRRIFNNSRE  
 XP\_030766515.1 S-----KSFEEREEEYEVKARRRIFRQEM  
 XP\_018019452.1 S-----KSFEERQEEYHARKRMRFSQEV  
 XP\_014672054.1 N-----KSIEEREEEYQKTRARIFNQD-  
 XP\_014675087.1 T-----KSFEEREEEAYEKTAKARIYNQD  
 XP\_030385838.1 S-----KSFEEREEEDFDRSRSRIFSRST-  
 XP\_029725418.1 A-----KSFEEREEEYDKVKRRIFKNR-  
 Dm\_encore A-----KSFEEREEEDYDRARSRSRIFSRST-  
 XP\_032599088.1 A-----KSFEEREEEDYDRARSRSRIFNRTA  
 KNC31935.1 A-----KSFEEREEEDYDRARSRSRIFNRS-  
 XP\_020714208.1 A-----KSFEEREEEDYERARSRSRIFNRS-  
 XP\_023246188.1 S-----KSFEEREEAEYHMARKRIFRDEK  
 XP\_023329389.1 S-----KSIEEREEEDYERAKRRIFNDDSS  
 XP\_029201356.1 S-----KSIEEREEEYHKARARIFNQD-  
 XP\_021368844.1 A-----QSFEERQQYEVKVRQKIFSSCD  
 XP\_034305772.1 A-----RSFEERRKRYEVKRSKIFNNSCD  
 XP\_013815153.1 S-----KSIEEREEEYQRARERIFQAQDS  
 XP\_016861070\_ARPP21 S-----KSIEEREEEYQVRVRRIFAHDS  
 Hs/X1/Dr S-----KSIEEREEEYQVRERIFARES  
Consensus aa: .....o.EERC..YpcbR.RIFs...  
Consensus ss: h hhhhhhhhhhhhhhhh

**C**

**Supplementary Figure S2 (seven previous pages). Sequences of proteins comprising R3H-SUZ domain combination.** **(A)** Description of accession numbers of the retrieved sequences. The numbers are color-coded: blue for fungi, green for plants, orange for invertebrates and black for vertebrates. **(B)** Multiple sequence alignment obtained with Promals3D. Consensus secondary structure and sequence are given in the bottom of each segment. Amino acid number for Rbs1 are given on top of the alignment. For further explanation, see legend of Fig 10. **(C)** Phylogenetic tree of the analyzed sequences.