

Supplementary Results & Discussion:

Results:

NMPP-elicited I κ B α -loss is independent of HRI eIF2 α -kinase activation: The concurrence of I κ B α -loss and NF- κ B activation and its attenuation by heme-treatment led us to consider whether NMPP-elicited heme depletion was responsible for these findings. Conceivably, heme depletion could activate the hepatic heme-sensor HRI eIF2 α -kinase resulting in the translational suppression of I κ B α and consequent NF- κ B-activation. Similar I κ B α translational suppression and consequent NF- κ B-activation has been documented upon specific activation of the other cellular eIF2 α kinases (1, 2). Indeed, in parallel with I κ B α -loss (Fig. S2A), NMPP treatment increased the relative ratio of phosphorylated eIF2 α (eIF2 α P) over basal eIF2 α levels, indicating HRI activation (3). However, NMPP-treatment of cultured hepatocytes from HRI WT (HRI^{+/+}) and knockout (KO; HRI^{-/-}) mice elicited a comparable I κ B α -loss, excluding causal HRI activation (Fig. 1D).

ZnPP-elicited I κ B α -loss is independent of autophagy, calpain-mediated proteolysis and reactive oxygen species (ROS): Given the reported I κ B α -degradation via ALD (4) and the known p62-role in this process (5), we conclusively excluded ALD in this process by documenting that knockout of the essential autophagic gene ATG5 failed to mitigate ZnPP-elicited I κ B α loss (Fig. S2B). Because of possible calpain-mediated I κ B α -degradation (6, 7), we also excluded its involvement by documenting that knockout of the calpain-degradation pathway (capn4 KO MEF cells) failed to abrogate ZnPP-elicited I κ B α -loss (Fig. S2C). These findings coupled with those with ALD and calpain inhibitors (Fig. S4B) revealed that neither ALD nor calpain degradation played any role in this I κ B α loss. Furthermore, through the use of various ROS-quenchers (Fig. S3), we excluded any possible PPIX-photoactivation and consequent ROS-mediated oxidative stress in this I κ B α -loss (8).

Discussion:

The precise mechanism of the ZnPP-elicited I κ B-protein sequestration is currently unknown, but some plausible mechanisms are presented. The fact that newly synthesized I κ B α is particularly vulnerable to such ZnPP-sequestration implicates the involvement of its intrinsic disordered domains at a stage when it has not yet reached its mature folded state.

In search of clues on potential ZnPP-mechanisms of protein sequestration, we sought the advice of various internationally recognized expert heme/porphyrin chemists and biochemists and porphyria experts on the possible mechanistic causes. Not a single investigator was aware that ZnPP caused protein aggregation, much less I κ B α -sequestration. ZnPP (along with SnPP), is actually believed to be safe and is recommended for the treatment of neonatal jaundice due to its effective inhibition of microsomal heme oxygenase (HO1), the key rate-limiting enzyme in the conversion of heme to biliverdin (9). However, *in vivo*, overproduction of protoporphyrin IX (PPIX) due to defects in ferrochelatase (congenital erythropoietic protoporphyria) or PPIX-overproduction in X-linked protoporphyria, as well as iron-deficiency or lead-poisoning induced anemias (that can exhaust iron-stores) leads to ZnPP-generation (10-12). Our findings suggest that under these conditions, these patients may easily succumb to ZnPP-elicited I κ B α -sequestration and consequently unabated hepatic NF- κ B-elicited activation of cytokines and chemokines. We believe, these findings would for the first-time alert physicians of this pathological potential of ZnPP and are clinically relevant not just in MDB-inducing diseases but also in clinical protoporphyrias and iron-deficiency/lead-induced anemias.

References:

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