

**Supplementary material for “The Phylogenomics of CRISPR-Cas System and
revelation of its features in *Salmonella*”**

Simran Krishnakant Kushwaha^{1*}, Narra Lakshmi Sai Bhavesh^{1#}, Bahaa Abdella^{2,3#}, Chandrajit
Lahiri² and Sandhya Amol Marathe^{1*}

Supplementary materials include:

Table S1-S8

Supplementary figure legends

Figures S1-S13

Supplementary Methodology

Supplementary Table S1. Name, accession number for whole genome sequence and spacer details for the strains analysed in the study.

STRAIN NAME	ACCESSION NUMBER	No. OF SPACERS- CRISPR1	No. OF SPACERS- CRISPR2	STRAIN NAME	ACCESSION NUMBER	No. OF SPACERS- CRISPR1	No. OF SPACERS- CRISPR2
<i>Salmonella enterica</i> subsp. <i>enterica</i>							
Serovar Anatum	Broad-host range			Serovar Paratyphi A	Host-restricted-Primates		
USDA-ARS-USMARC-1175	CP007483.2	2	26	AKU 12601	FM200053.1	7	3
USDA-ARS-USMARC-1676	CP014620.1	2	19	ATCC 11511	CP019185.1	7	3
USDA-ARS-USMARC-1677	CP014663.2	2	21	ATCC 9150	CP000026.1	5	3
USDA-ARS-USMARC-1727	CP014621.2	2	12	Serovar Gallinarum	Host-restricted- Poultry and other avians		
USDA-ARS-USMARC-1728	CP014664.1	2	25	28791	AN931173.1	2	10
USDA-ARS-USMARC-1735	CP007584.2	8	8	9184	CP019035.1	2	10
USDA-ARS-USMARC-1736	CP014657.1	8	26	Serovar Pullorum	Host-restricted- Poultry		
USDA-ARS-USMARC-1765	CP014659.2	2	21	ATCC 9120	CP012347.1	2	6
USDA-ARS-USMARC-1766	CP014663.1	8	24	S06084	CP006576.1	2	2
USDA-ARS-USMARC-1781	CP014666.2	8	20	Serovar Gallinarum/Pullorum	Host-restricted- Poultry		
USDA-ARS-USMARC-1783	CP014661.1	2	25	CDC 1983-67	CP003786.1	2	6
ATCC BAA-1592	CP007531.1	8	20	RKSS078	CP003047.1	2	6
CDC 06-0532	CP007211.2	8	20	Serovar Typhimurium	Broad-host range		
Serovar Enteritidis	Broad-host range			USDA-ARS-USMARC-1808	CP014869.1	10	26
77-1427	CP007598.1	9	8	USDA-ARS-USMARC-1810	CP014882.2	21	31
CDC 2010K_0968	CP007528.1	9	11	USDA-ARS-USMARC-1880	CP014981.1	11	35
EC20090641	CP007249.2	9	11	USDA-ARS-USMARC-1896	CP014977.1	28	24
EC20090699	CP007248.1	9	11	USDA-ARS-USMARC-1898	CP014971.2	22	32
EC20110221	CP007247.1	9	11	USDA-ARS-USMARC-1899	CP007235.2	8	19
EC20110354	CP007175.1	9	12	str. 14028S	CP001363.1	22	26
EC20110355	CP007250.1	9	12	str. 798	CP003386.1	22	22
EC20110356	CP007262.1	9	12	CDC 2009K-1640	CP014975.1	10	26
EC20110357	CP007261.1	9	11	CDC 2009K-2020	CP014983.1	28	24
EC20110358	CP007260.1	9	11	CDC 2010K-1587	CP014985.1	27	24
EC20110360	CP007258.1	9	12	CDC 2011K-0870	CP007523.1	11	34
EC20110361	CP007263.1	9	12	CDC 2011K-1702	CP014967.1	10	26
EC20111095	CP007254.1	9	12	D23580	LS997973.1	21	18
EC20111174	CP007253.1	9	12	DT104	HG37208.1	10	26
EC20111175	CP007252.1	9	12	DT2	HG326213.1	20	26
EC20120002	CP007329.2	9	12	L-3553	AP014565.1	22	15
EC 20120005	CP007267.2	9	12	SARA13	CP017228.1	17	25
EC20120008	CP007243.1	9	11	SL1244	FG031003.1	14	23
EC20121175	CP007269.2	9	12	SI474	CP002487.1	14	23
P125109	AN931172.1	8	10	T000240	AP011957.1	20	32
Serovar Saintpaul	Broad-host range			UK-1	CP002614.1	14	23
CFSAN004173	CP019204.1	15	20	CFSAN001921	CP006048.1	27	24
CFSAN004174	CP019206.1	15	20	Serovar Senftenberg	Host-restricted-Primates		
CFSAN004175	CP019172.1	15	20	NC15772	NZ_LGV_R01000002.1	4	1
SGB23	CP023166.1	11	22	Serovar Tennessee	Broad-host range		
SARA26	CP017277.1	41	22	ATCC 10722	CP025218.1	63	23
Serovar Heidelberg	Broad-host range			CFSAN001387	CP014994.1	47	23
CFSAN002064	CP005993.1	25	18	CFSAN070643	CP024168.1	63	23
41578	CP006086.1	25	16	CFSAN076210	CP033345.1	47	21
B182	CP003416.1	25	18	PIR00537	CP025217.1	47	23
CFSAN002069	CP006590.2	25	18	CFSAN070645	CP024164.1	49	21
SARA35	CP019176.1	25	18	TXSC - TXSC 08-19	CP007505.1	51	23
SL476	CP001128.1	25	18	Serovar Dublin	Host-adapted/cattle		
Serovar Typhi	Host-restricted- Primates			CT_02821853	CP001144.1	2	5
Pstrv-12	CP003278.1	6	1	Serovar Paratyphi C	Host-restricted- Primates (not frequent) and known to infect other animals like pigs		
Ty2	AE014613.1	6	1	RKS4594	CP000857.1	10	9
CT18	AL513382.1	6	1	Serovar Montevideo	Broad-host range		
1016889	LF040893.1	6	1	CFSAN051296	CP029336.1	24	17
1036491	LF040883.1	6	1	S31954	CP029035.1	24	17
1290235-M	LF040883.1	6	1	507440-20	CP007530.1	24	17
ERL034151	LF040867.1	7	1	CDC 07-4954	CP017974.1	11	25
Serovar Newport II	Broad-host range			CDC 08-1942	CP017975.1	6	21
CVM 21558	CP010282.1	26	19	CDC 2010K-0257	CP020912.1	24	17
CVM 21550	CP010283.1	26	19	CDC 2011K-1674	CP017976.1	11	21
CVM 22425	CP010279.1	26	19	CDC 2013K-1544	CP017977.1	24	17
CVM 22462	CP010280.1	26	19	CDC 2013K-0218	CP017978.1	6	21
CVM 22513	CP010281.1	20	19	FCC0123	CP040379.1	4	22
CVM N1543	CP010284.1	26	19	CDC 2009K-0792	CP020752.1	35	20
CVM N18486	CP009561.1	17	12	USDA-ARS-USMARC-1900	CP017970.1	36	22
SI_254	CP001113.1	26	19	USDA-ARS-USMARC-1901	CP017971.1	34	20
USMARC-S3124.1	CP006631.1	26	19	USDA-ARS-USMARC-1904	CP017972.1	33	20
WA_14882	CP010357.1	25	19	USDA-ARS-USMARC-1912	CP017973.1	35	16
Serovar Newport III	Broad-host range			USDA-ARS-USMARC-1903	CP007222.1	35	23
CFSAN001660	CP016010.1	15	20	USDA-ARS-USMARC-1921	CP007540.2	35	23
CDC 2009K-1331	CP025248.1	13	16	<i>Salmonella enterica</i> subsp. <i>arizonae</i>	Host-cold-blooded animals, mainly reptiles and sheeps		
CDC 2012K-0938	CP025246.1	4	20	NC121047	LR134156.1	23	ABSENT
Levine 1	CP015922.1	13	19	<i>Salmonella enterica</i> subsp. <i>diarizonae</i>	Host-cold-blooded animals, mainly reptiles and sheeps		
USDA-ARS-USMARC-1927	CP007216.2	18	10	SI_MZ0080	CP022142.1	29	ABSENT
Levine 15	CP015924.1	5	16	<i>Salmonella bongori</i>	Host-cold-blooded animals, have been isolated from humans but very rare		
Serovar Agona	Broad-host range			CFSAN005010	CP045766.1	20	17
SI_483	CP001138.1	18	8	NC121419	LR134137.1	20	17
Serovar Bovismorbificans	Broad-host range			66 SA19983605	CP022120.1	20	17
3114	HF969015.2	24	15				

The strains represented in bold were considered for further analysis. The strains highlighted in grey for serovars Montevideo and Saintpaul represent the ones having distinct spacer sets.

Supplementary Table S2. The statistics of the spacer index for the serovars under consideration.

			No. of strains analysed	CRISPR1*			CRISPR2*		
				Minimum	Maximum	Average	Minimum	Maximum	Average
<i>Salmonella enterica</i> subsp. <i>enterica</i>	Host-restricted	Typhi	7	6	7	6.14	1	1	1
		Paratyphi A	3	5	7	6.33	3	3	3
		Gallinarum	2	2	2	2	10	10	10
		Pullorum	2	2	2	2	2	6	4
		Gallinarum/Pullorum	2	2	2	2	6	6	6
		Sendai	1	4	4	4	1	1	1
	Host-adapted	Dublin	1	2	2	2	5	5	5
	Broad-host range	Anatum	13	2	8	4.77	8	26	20.25
		Paratyphi C	1	10	10	10	9	9	9
		Heidelberg	6	25	25	25	16	18	17.67
		Newport II	10	17	26	24.4	12	19	18.3
		Newport III	6	4	18	11.33	10	20	16.83
		Enteritidis	20	8	9	8.95	8	12	11.35
		Typhimurium	23	8	28	17.95	15	35	25.39
		Bovismorbificans	1	24	24	24	15	15	15
		Tennessee	7	41	63	52.43	21	23	22.14
		Montevideo	17	4	36	23.59	16	25	19.94
		Saintpaul	5	11	41	19.4	20	22	20.8
		Agona	1	18	18	18	8	8	8
Cold-blooded hosts	<i>Salmonella enterica</i> subsp. <i>arizonae</i>	1	23	23	23		-	-	
	<i>Salmonella enterica</i> subsp. <i>diarizonae</i>	1	29	29	29		-	-	
	<i>Salmonella bongori</i>	3	20	20	20	17	17	17	
Total					15.29			12.58	
* Each serovar has one CRISPR1 and one CRISPR2 array except <i>S. enterica</i> subsp. <i>arizonae</i> and <i>diarizonae</i>									

Supplementary Table S3. Name and accession number for whole genome sequence for the strains analysed in the study.

SPECIES	ACCESSION NUMBER	ABBREVIATION
<i>Salmonella bongori</i> str. 66 SA19983605	CP022120.1	SB
<i>Salmonella enterica</i> subsps. <i>arizonae</i> str. NCTC10047	LR134156.1	SE-A
<i>Salmonella enterica</i> subsps. <i>diarizonae</i> 50 MZ0080	CP022142.1	SE-D
<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar Typhimurium str. 14028s	CP001363.1	STM1
<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar Typhimurium str. SARA13	CP017728.1	STM2
<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar Typhi str. Ty2	AE014613.1	STY1
<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar Typhi str. CT18	AL513382.1	STY2
<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar Newport str. CVM21538	CP010282.1	SE-NP1
<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar Newport str. CVM22513	CP010281.1	SE-NP2
<i>Klebsiella pneumoniae</i> subsp. <i>pneumoniae</i> strain TGH10	CP012744.1	KP1
<i>Klebsiella pneumoniae</i> strain INF235-sc-2280127	CP031817.1	KP2
<i>Shigella boydii</i> strain ATCC 49812	CP026836.1	SB1
<i>Citrobacter freundii</i> complex sp. CFNIH3	CP026235.1	C1
<i>Citrobacter freundii</i> complex sp. CFNIH9	CP026238.1	C2
<i>Citrobacter</i> sp. 30_2	CP022311.1	C3
<i>Escherichia coli</i> strain 52148	CP050382.1	EC1
<i>Escherichia coli</i> strain SCU-118	CP051716.1	EC2

The CRISPR1 leader sequences of these strains were used for the phylogenetic analysis. The abbreviations are the key to the figure.

Supplementary Table S4. Strains of *Klebsiella*, *Escherichia* and *Shigella* matching with CRISPR1-STM leader and *cas*-STM.

	<i>Escherichia</i>	<i>Klebsiella</i>	<i>Shigella</i>
CRISPR1-STM Leader	491	105	12
Cas-STM	~1000	~100	~50

Values indicate number of strains.

Supplementary Table S5. MGE candidates flanking the CRISPR-Cas system.

	Genome location (Loci start and Loci end)			MGE Transposase/ Helicase	Percentage GC Content				
	CRISPR2	<i>cas</i>	CRISPR1		CRISPR2 loci	<i>cas</i> operon	CRISPR1 loci	CRISPR-Cas	Whole genome
Paratyphi A str. AKU_12601	2902111-2902322	2885645-2894598	2885105-2885560	2856664-2857091 and 3007579-3008037	51.3*	49.9	53.2*	49	52.18
Newport II str. SL254	3073142-3074328	3056558-3064452	3054859-3056473	3024723-3025160	54.2	50.4	55.6	51	52.22
Newport III str. USDA-ARS-USMARC-1927	975001-975639	983339-991789	991886-993012	1020178-1020414	51.3	52	57.2	51	52.18
Heidelberg str. SL476	3069137-3070263	3052976-3061435	3051217-3052879	3022734-3022874	56.3	53.1	56.2	53	52.07
Enteritidis str. EC20121175	2967508-2968269	2951453-2959906	2950779-2951356	2923727-2923816	55.8	52.8	55.7	51	52.17
Gallinarum str. 9184	1224776-1225415	1233429-1241471	1241469-1241716	1271625-1271848	56.8	53.3	47.6*	51	52.2
Pullorum str. ATCC 9120	3871330-3871722	3855356-3863734	3855036-3855283	3827981-3828070	53.6*	52.9	48.4*	51	52.19
Gallinarum/Pullorum str. CDC1983-67	2947545-2947937	2931570-2939948	2931250-2931497		53.6*	52.9	48.4*	51	52.23
Montevideo-STY str. USDA-ARS-USMARC-1900	1003049-1004420	1011949-1021097	1021182-1023406	1050505-1050672	57.7	50.4	57	52	52.35
Montevideo-STM str. CDC 2010K-0257	992948-994014	1010602-1010052	1010149-1011641	1038743-1038910	56.6	52.4	55.9	51	52.21
Bovismorbificans str. 3114	2976895-2977839	2960410-2969354	2958839-2960331		54	50	55.6	50	52.16
Anatum str. CDC 06-0532	970192-971440	979153-987612	987709-988225	1018009-1018429	56.2	52.8	55.5	51	52.18
Tennessee str. ATCC 10722	963889-965320	972914-981858	981943-985815	1015542-1015960	59.3	50.4	57.1	52	52.23
Saintpaul-STM str. CFSAN004173	946615-947863	955563-964025	964122-965066	993235-993458	57.2	52	57.4	51	52.21
Saintpaul-STY str. SARA26	944744-946114	953779-962723	962808-965337	993600-993740	58.2	50.3	56.5	51	52.05
Dublin str. CT_02021853	3137409-3137742	3121348-3129807	3121100-3121350	3090967-3091107	51.6*	53	44.6*	51	52.18
Agona str. SL483	3005517-3006033	2989328-2997778	2988105-2989231	2956649-2956789 and 2975984-2977192	54.2	52	56.3	51	52.08
Typhimurium str. CFSAN001921	473159-474652	482228-490687	490784-492564	522104-522291 and 423309-425144	55.9	50.4	56.9	51	52.17
Typhi str. CT18	2943208-2943716	2926652-2935104	2926182-2926567	2898592-2899034 and 3013615-3014073	39.8*	50.4	57.3	50	52.05
Subsp. <i>arizonae</i>		962736-971156	971253-972682			52.9	57.4	52	51.38
Subsp. <i>diarizonae</i>		1126557-1134989	1135086-1136883			52.9	56.7	53	51.54
<i>S. bongori</i>	922139-923204	930621-939089	939186-940434		53.4	52.4	54.5	50	51.33

*The lower GC content of CRISPR arrays due to the AT-rich leader sequences are represented by asterisks

Supplementary Table S6. MGE present in the whole genome of *S. enterica* subsp. *enterica* serovar Typhimurium str. LT2 and serovar Typhi str. TY2

<i>Salmonella</i> Typhimurium str. LT2			<i>Salmonella</i> Typhi str. TY2		
Gene Start	Gene End		Gene Start	Gene End	
110002	112908	Helicase	109791	112697	Helicase
218296	220783	Helicase	115562	116002	putative IS element transposase
340748	340953	Transposase	116022	117230	putative IS element transposase
341175	341460	Integrase	271067	271525	transposase for insertion sequence element IS200
369198	369998	Transposase	222741	225182	Helicase
405293	408272	Helicase	280079	280537	transposase for insertion sequence element IS200
489948	490109	Transposase	305010	306344	Helicase
537933	538874	Transposase	464716	465174	transposase for insertion sequence element IS200
613596	613797	Integrase	528278	528736	transposase for insertion sequence element IS200
613753	614167	Transposase	673526	673984	transposase for insertion sequence element IS200
781631	782197	Integrase	712019	713779	Helicase
885578	886953	Helicase	1070157	1070768	Helicase
887150	889305	Helicase	1070777	1071787	Helicase
962615	963952	Transposase Integrase	1139104	1141014	Helicase
969994	971585	Helicase	1189317	1189775	transposase for insertion sequence element IS200
1025163	1025627	Transposase	1220753	1221211	transposase for insertion sequence element IS200
1026090	1026773	Transposase Integrase	1265128	1265586	transposase for insertion sequence element IS200
1098231	1099531	Transposase Integrase	1450560	1451018	transposase for insertion sequence element IS200
1166406	1168476	Helicase	1598822	1602724	Helicase
1214284	1215158	Helicase	1647002	1647460	transposase for insertion sequence element IS200
1731328	1735230	Helicase	1745409	1745867	transposase for insertion sequence element IS200
1748282	1749655	Helicase	1802131	1802589	transposase for insertion sequence element IS200
1918512	1920433	Helicase	1895018	1895476	transposase for insertion sequence element IS200
1956851	1957174	Transposase	1898230	1898688	transposase for insertion sequence element IS200
1957832	1958200	Integrase	1903289	1905343	Helicase
1967023	1967919	Integrase	1970990	1972330	Integrase
1990051	1990071	Helicase	2473258	2473716	transposase for insertion sequence element IS200
1990070	1990693	Helicase	2534377	2534835	transposase for insertion sequence element IS200
2045732	2046196	Transposase	2735202	2736233	Integrase
2324547	2326307	Helicase	3012372	3012830	transposase for insertion sequence element IS200
2579330	2579794	Transposase	3043685	3045148	Integrase
2622291	2622652	Transposase	3052442	3053338	Integrase
2731352	2731902	Transposase	3077594	3078052	transposase for insertion sequence element IS200
2775590	2776825	Integrase	3085185	3085643	transposase for insertion sequence element IS200
2782896	2784240	Helicase	3108025	3108483	transposase for insertion sequence element IS200
2876819	2877844	Integrase	3287491	3289431	Helicase
2878160	2879237	Integrase	3440746	3442575	Helicase
2900435	2901318	Integrase	3447448	3449610	Helicase
2905473	2906075	Integrase	3450462	3451364	Integrase
2906456	2906923	Integrase	3482595	3483860	Helicase
2907070	2907343	Transposase	3485500	3487524	Helicase
2908424	2910413	Helicase	3501128	3502180	Integrase
2910673	2910984	Transposase	3894835	3896916	Helicase
2910967	2911847	Transposase	4296365	4297780	Helicase
3081922	3082490	Transposase	4395333	4396697	Helicase
3084047	3086721	Helicase	4489831	4490856	Integrase
3144282	3146128	Helicase	4511590	4512603	Integrase
3146114	3149659	Helicase	4524556	4525815	Integrase
3152718	3156100	Helicase	4666742	4668007	Integrase
3194324	3194788	Transposase	4689323	4693072	Helicase
3204431	3205339	Integrase	4693110	4695647	Helicase
3317055	3317570	Helicase			
3446192	3448132	Helicase			
3635168	3635632	Transposase			
3936873	3938974	Helicase			
4049846	4051066	Integrase			
4119776	4121800	Helicase			
4123440	4124705	Helicase			
4153785	4154702	Integrase			
4155554	4157716	Helicase			
4162571	4164418	Helicase			
4466589	4468022	Helicase			
4559426	4559890	Transposase			
4702530	4702982	Transposase			
4731324	4731619	Integrase			
4731975	4735496	Helicase			

Supplementary Table S7. Mapping protospacer sources of CRISPR spacers

		Species	CRISPR1					CRISPR2				
			No. of spacers having protospacers	No. of spacers not displaying any protospacers	Total No. of spacers	Percent spacers having protospacers	Percent spacers not displaying protospacers	No. of spacers having protospacers	No. of spacers not displaying any protospacers	Total No. of spacers	Percent spacers having protospacers	Percent spacers not displaying protospacers
<i>Salmonella enterica</i> subsp. <i>enterica</i>	Host-restricted	Typhi	5	2	7	71.43	28.57	1	0	1	100.00	0.00
		Paratyphi A	5	2	7	71.43	28.57	2	1	3	66.67	33.33
		Gallinarum	1	1	2	50.00	50.00	6	4	10	60.00	40.00
		Pullorum	2	1	3	66.67	33.33	3	3	6	50.00	50.00
		Gallinarum/Pullorum	2	0	2	100.00	0.00	3	3	6	50.00	50.00
		Sendai	4	0	4	100.00	0.00	1	0	1	100.00	0.00
	Host-adapted	Dublin	1	1	2	50.00	50.00	2	3	5	40.00	60.00
	Broad-host range	Anatum	4	4	8	50.00	50.00	18	8	26	69.23	30.77
		Paratyphi C	6	4	10	60.00	40.00	4	5	9	44.44	55.56
		Heidelberg	18	7	25	72.00	28.00	13	5	18	72.22	27.78
		Newport II	16	11	27	59.26	40.74	15	6	21	71.43	28.57
		Newport III	10	8	18	55.56	44.44	12	9	21	57.14	42.86
		Enteritidis	4	5	9	44.44	55.56	9	3	12	75.00	25.00
		Typhimurium	16	13	29	55.17	44.83	25	12	37	67.57	32.43
		Bovismorbificans	17	7	24	70.83	29.17	9	6	15	60.00	40.00
		Tennessee	37	26	63	58.73	41.27	16	7	23	69.57	30.43
		Montevideo-STM	16	9	25	64.00	36.00	12	15	27	44.44	55.56
		Montevideo-STY	26	10	36	72.22	27.78	18	5	23	78.26	21.74
		Saintpaul-STM	10	5	15	66.67	33.33	13	9	22	59.09	40.91
		Saintpaul-STY	27	14	41	65.85	34.15	13	9	22	59.09	40.91
Agona	8	10	18	44.44	55.56	5	3	8	62.50	37.50		
Cold-blooded hosts	<i>Salmonella enterica</i> subsp. <i>arizonae</i>	18	5	23	78.26	21.74	NA	NA	NA	NA	NA	
	<i>Salmonella enterica</i> subsp. <i>diarizonae</i>	18	11	29	62.07	37.93	NA	NA	NA	NA	NA	
	<i>Salmonella bongori</i>	9	11	20	45.00	55.00	9	8	17	52.94	47.06	
		AVERAGE				63.92	36.08				64.07	35.93
		STANDARD DEVIATION				14.80	14.80				15.6	15.6

Supplementary Table S8. Mapping the ChIP-seq of Cas5 on the whole genome of *S. enterica* subsp. *enterica* serovar Typhimurium str. 14028S

ChIP-seq Peak Center	Gene name	Function
648399	<i>entF</i>	Enterobactin synthetase, component F ¹
658872	<i>entE</i>	Enterobactin synthetase, component E ¹
1482744	<i>trtS</i>	Tetrathionate sensor histidine kinase ³
1048355	<i>ompF</i>	Forms pores that allow passive diffusion of small molecules across the outer membrane ⁴
2724461	<i>seaA</i>	Type III secretion system chaperone ⁵
2747746	<i>hmpA</i>	Is involved in NO detoxification in an aerobic process, termed nitric oxide dioxygenase (NOD) reaction that utilizes O ₂ and NAD(P)H to convert NO to nitrate, which protects the bacterium from various noxious nitrogen compounds ⁶
2242087	<i>wcaI</i>	GDP-mannose mannosyl hydrolase ⁷
2937813	<i>iroC</i>	ABC transporter ATP-binding protein ⁸
3042704	<i>sptP</i>	Effector proteins function to alter host cell physiology and promote bacterial survival in host tissues ⁹
3210092	<i>sttB</i>	Fimbrial biogenesis outer membrane usher protein ¹⁰
3525028	<i>sspA</i>	Important for enteropathogenicity ¹¹
3815385	<i>bcsA</i>	Catalytic subunit of cellulose synthase. It polymerizes uridine 5'-diphosphate glucose to cellulose, which is produced as an extracellular component for mechanical and chemical protection at the onset of the stationary phase, when the cells exhibit multicellular behavior ¹²
3976737	<i>mgfB</i>	Mediates magnesium influx to the cytosol ¹³
4161855	<i>cyoA</i>	Adenylate cyclase is involved in the regulation of cyclase activity by the carbon source ¹⁴
4547246	<i>basS</i>	Member of the two-component regulatory system BasS/BasR. Autophosphorylates and activates BasR by phosphorylation ¹⁵
4861926	<i>sthD</i>	Putative fimbrial subunit ¹⁶
4862974	<i>sthB</i>	Fimbrial assembly protein ¹⁷
28623	<i>bcdF</i>	Cell adhesion
37110		putative 5'-nucleotidase
67716		putative citrate-sodium symporter
74672	<i>dispB</i>	Catalyzes the conversion of 4-hydroxy-tetrahydrodipicolinate (HTPA) to tetrahydrodipicolinate
78290	<i>carB</i>	Carbamoyl-phosphate synthase large chain
81565	<i>caif</i>	DNA binding
85304	<i>caib</i>	Catalyzes the reversible transfer of the CoA moiety from gamma-butyrobetainyl-CoA to L-carnitine to generate L-carnitinyl-CoA and gamma-butyrobetaine.
113296	<i>hepA</i>	Transcription regulator that activates transcription by stimulating RNA polymerase (RNAP) recycling in case of stress conditions such as supercoiled DNA or high salt concentrations.
114993	<i>polB</i>	DNA polymerase
134582	<i>leuL</i>	Involved in control of the biosynthesis of leucine.
136070	<i>leuO</i>	Probable HTH-type transcriptional regulator
137573	<i>ilvI</i>	Acetolactate synthase isozyme 3 large subunit
162127		
166935	<i>hpfC</i>	Protein transport protein
171781		putative cytoplasmic protein
188885	<i>acnB</i>	Acetate hydratase-Involved in the catabolism of short chain fatty acids
254924		30S ribosomal protein S2
259664	<i>dar</i>	Catalyzes the NADP-dependent rearrangement and reduction of 1-deoxy-D-xylulose-5-phosphate (DXP) to 2-C-methyl-D-erythritol 4-phosphate (MEP)
260309	<i>upsS</i>	Ditrans,poly(cis-undecaprenyl)-diphosphate synthase ⁶
270914	<i>dnaE</i>	DNA polymerase III is a complex, multichain enzyme responsible for most of the replicative synthesis in bacteria.
283323	<i>proS</i>	Proline-tRNA ligase
284920		
292686	<i>rriH</i>	-
316369		putative cytoplasmic protein
331550		putative cytoplasmic protein
354530	<i>fadE</i>	Catalyzes the dehydrogenation of acyl-coenzymes A (acyl-CoAs) to 2-enoyl-CoAs, the first step of the beta-oxidation cycle of fatty acid degradation.
360797		
385712	<i>pseudogene</i>	
391414		putative outer membrane protein
444928	<i>yafF</i>	Fructokinase
461724	<i>tgt</i>	Queuine tRNA-ribosyltransferase
503419	<i>tig</i>	Involved in protein export. Acts as a chaperone by maintaining the newly synthesized protein in an open conformation.
510542	<i>oxyD</i>	Peptidoglycan isomerase
530680	<i>acrB</i>	Multidrug efflux pump subunit drug efflux protein complex with broad substrate specificity that uses the proton motive force to export substrates.
533628	<i>acrA</i>	Multidrug efflux pump subunit drug efflux protein complex with broad substrate specificity that uses the proton motive force to export substrates.
547471		hypothetical protein
565916	<i>yhbO</i>	NAD(P)-dependent oxidoreductase
606034	<i>lincD</i>	Involved in the export and assembly of fimbrial subunits across the outer membrane.
704128	<i>rvxD</i>	Catalyzes cross-linking of the peptidoglycan cell wall
781326	<i>ybgL</i>	Catalyzes the cleavage of 5-oxoprolinone to form L-glutamate coupled to the hydrolysis of ATP to ADP and inorganic phosphate.
799842	<i>sdhA</i>	Succinate dehydrogenase flavoprotein
805852	<i>sueB</i>	E2 component of the 2-oxoglutarate dehydrogenase (OGDH) complex which catalyzes the second step in the conversion of 2-oxoglutarate to succinyl-CoA and CO ₂ .
862272	<i>bioF</i>	Biotin-7-oxononanoate synthase
911440		hypothetical protein
995967	<i>flxK</i>	Essential cell division protein that coordinates cell division and chromosome segregation.
1027515	<i>ycaI</i>	Lactamase_B domain-containing protein
1071391		
1086398		minor tail protein
1088277		attachment/invasion protein
1126386	<i>helD</i>	DNA Helicase
1146356		hypothetical protein
1173716		putative transcriptional regulator
1175296		putative sodium/glucose cotransporter
1197202	<i>mdoH</i>	Involved in the biosynthesis of osmoregulated periplasmic glucans
1230875	<i>rluC</i>	Ribosomal large subunit pseudouridine synthase C
1237741	<i>yagG</i>	Catalyzes the NADPH-dependent reduction of beta-ketoacyl-ACP substrates to beta-hydroxyacyl-ACP products, the first reductive step in the elongation cycle of fatty acid biosynthesis
1250794	<i>nagZ</i>	Plays a role in peptidoglycan recycling by cleaving the terminal beta-1,4-linked N-acetylglucosamine (GlcNAc)
1319463		phage tail component H-like protein
1339141		
1362889		
1371489	<i>yeaH</i>	-
1379980	<i>yeaA</i>	Peptide methionine sulfoxide reductase
1393323	<i>argD_1</i>	Catalyzes the transamination of N2-succinylornithine and alpha-ketoglutarate into N2-succinylglutamate semialdehyde and glutamate
1409579		hypothetical protein
1427820	<i>pheT</i>	Phenylalanine-tRNA ligase beta subunit
1435560	<i>araH</i>	Stereo-specific condensation of phosphoenolpyruvate (PEP) and D-erythrose-4-phosphate (E4P) giving rise to 3-deoxy-D-arabino-heptulosonate-7-phosphate (DAHP).
1573929	<i>ynfM</i>	Permease
1575922	<i>mic</i>	-
1599510	<i>yafG</i>	NADP-dependent dehydrogenase with broad substrate specificity acting on 3-hydroxy acids
1609053	<i>ydeA</i>	Involved in the efflux of sugars.
1613004	<i>yneH</i>	Glutaminase
1620035	<i>ylnG</i>	Formate dehydrogenase allows E. coli to use formate as major electron donor during anaerobic respiration, when nitrate is used as electron acceptor
1707334		putative benzoate membrane transport protein
1749855		
1875758	<i>narX</i>	Sensor protein
1902575		putative cytochrome oxidase subunit I
1966259		
2022871		
2152720	<i>cbpP</i>	Cobyrinic acid synthase Catalyzes amidations at positions B, D, E, and G on adenosylcobyrinic A,C-diamide. NH ₂ groups are provided by glutamine, and one molecule of ATP is hydrolyzed for each amidation.
2174789	<i>pduJ</i>	Carboxysome shell protein
2286468		
2328373		
2340192	<i>galS</i>	Repressor of the <i>galP</i> operon. Binds galactose and D-fructose as inducers. GalS binds to an operator DNA sequence within its own coding sequence
2402408	<i>cmf_1</i>	Cytochrome c type biogenesis protein F1
2454950	<i>ois</i>	Lipopolysaccharide core heptose(II)-phosphate phosphatase is involved in the pathway lipopolysaccharide metabolism, which is part of bacterial outer membrane biogenesis.
2502812	<i>pta</i>	Involved in acetate metabolism
2562245	<i>pgtC</i>	Required for PgtP expression, it may act jointly with the PgtA/PgtB signaling proteins.
2612828	<i>hemF</i>	Involved in the heme biosynthesis
2615387		putative cytoplasmic protein
2676031	<i>guoB</i>	Catalyzes the conversion of inosine 5'-phosphate (IMP) to xanthosine 5'-phosphate (XMP), the first committed and rate-limiting step in the de novo synthesis of guanine nucleotides, and therefore plays an important role in the regulation of cell growth.
2690387	<i>ratB</i>	Involved in toxin-antitoxin system
2696323	<i>ratA</i>	Ribosome association toxin
2708378	<i>ispG</i>	4-hydroxy-3-methylbut-2-en-1-yl diphosphate synthase
2723080		putative inner membrane lipoprotein
2759144	<i>yfhK</i>	Histidine Kinase
2772009		putative permease
2795645		phage tail component H-like protein
2852910	<i>rriG</i>	-
2878401	<i>recV</i>	May be involved in recombinational repair of damaged DNA
2932715		

2935299	<i>hin</i>	DNA-invertase hin
2946034		
2959109	<i>ygaf</i>	Multi functional fusion protein
2968949		putative inner membrane protein
2993084	<i>alaS</i>	Catalyzes the attachment of alanine to tRNA (Ala)
3007285	<i>ygbd</i>	One of at least two accessory proteins for anaerobic nitric oxide (NO) reductase
3050276	<i>truD</i>	tRNA pseudouridine synthase D
3092114		hypothetical protein
3093634	<i>cysN</i>	Sulfate adenylyltransferase subunit 1-May be the GTPase, regulating ATP sulfurylase activity.
3110917	<i>cysI</i>	Component of the sulfite reductase complex that catalyzes the 6-electron reduction of sulfite to sulfide.
3125034	<i>rumA</i>	23S rRNA [uracil(2939)-C5]-methyltransferase
3132946	<i>ygqT</i>	Glucuronate dehydratase
3151410	<i>glcU</i>	Involved in the anomeric conversion of L-fucose.
3153348	<i>ygqE</i>	Catalyzes the 2'-O-methylation at nucleotide C2498 in 23S rRNA.
3226541	<i>ygqZ</i>	tRNA modifying protein
3227626	<i>ygfA</i>	Cytolysis
3305087		putative amino acid transporter
3335029		
3408554	<i>ygiQ</i>	DUF218 domain-containing protein
3422879	<i>yhoO</i>	amino acid transmembrane transport
3435898	<i>garL</i>	Catalyzes the reversible retro-aldol cleavage of both 5-keto-4-deoxy-D-glucarate and 2-keto-3-deoxy-D-glucarate to pyruvate and tartronic semialdehyde.
3468956	<i>hfb</i>	Translation initiation factor IF-2
3471285	<i>yhcC</i>	Ribosome maturation factor
3475670	<i>glmM</i>	Catalyzes the conversion of glucosamine-6-phosphate to glucosamine-1-phosphate
3477559	<i>fbp</i>	Catalyzes the condensation of para-aminobenzoate (pABA) with 6-hydroxymethyl-7,8-dihydropterin diphosphate (DHPI-PP) to form 7,8-dihydropterate (H2Pte)
3479495	<i>hfbB</i>	Acts as a processive, ATP-dependent zinc metalloproteinase for both cytoplasmic and membrane proteins.
3487508	<i>ispB</i>	Octaprenyl diphosphate synthase
3513963	<i>gltD</i>	Glutamate synthase
3523435	<i>yhcK</i>	Hfh-type transcriptional repressor
3558707		hypothetical protein
3560950	<i>yhdH</i>	Putative oxidoreductase
3583298	<i>rrlD</i>	"
3589371	<i>smg</i>	"
3602356		
3618205	<i>yheO</i>	Putative reulator
3619144	<i>βpA</i>	Peptidyl-prolyl cis-trans isomerase
3635929		Hypothetical protein
3658225	<i>hgfQ</i>	DNA transporter
3685793	<i>malQ</i>	Transfers a segment of a [1->4]-alpha-D-glucan to a new position in an acceptor, which may be glucose or a [1->4]-alpha-D-glucan.
3690088	<i>malP</i>	Alpha-1,4 glucan phosphorylase
3729165		
3773648	<i>pitA</i>	Phosphate transporter
3783533	<i>gor</i>	Glutathione reductase
3795727	<i>yhjD</i>	Inner membrane protein
3820332	<i>yhjU</i>	Cellulose biosynthesis protein
3850991		hypothetical protein
3851150		
3856794		putative outer membrane lipoprotein
3862386	<i>ylr</i>	Catalyzes the phosphorylation of N-acetyl-D-glucosamine (GlcNAc) derived from cell-wall degradation, yielding GlcNAc-6-P
3863537	<i>box</i>	"
3865552	<i>malS</i>	Alpha-amylase
3878721		putative regulatory protein
3904893		Hypothetical protein
3905779		putative mandelate racemase
3934110	<i>kdhA</i>	Involved in lipopolysaccharide (LPS) biosynthesis.
3940960	<i>rph</i>	Phosphorylytic exoribonuclease that removes nucleotide residues following the -CCA terminus of rRNA and adds nucleotides to the ends of RNA molecules by using nucleoside diphosphates as substrates.
3949816	<i>spoT</i>	βppGpp synthetase II
3961073	<i>yicI</i>	MFS transporter
3963291		Hypothetical protein
3990958		putative glycosyl hydrolase
4029473	<i>ccmF_2</i>	Cytochrome c type biogenesis protein F2
4043153	<i>dgoT</i>	D-galactonate transport protein
4058622	<i>dnaA</i>	Plays an important role in the initiation and regulation of chromosomal replication.
4094038	<i>atpI</i>	ATP PFO1 synthase subunit I
4094227	<i>atpI</i>	ATP PFO1 synthase subunit I
4095913	<i>glfA</i>	NAD-binding protein involved in the addition of a carboxymethylaminomethyl (cmnm) group at the wobble position (U34) of certain tRNAs, forming tRNA-cmnmS2U34.
4116354	<i>rrlC</i>	"
4134314	<i>rep</i>	ATP-dependent DNA helicase
4151518	<i>wecF</i>	Catalyzes the synthesis of Und-PP-GlcNAc-ManNAcA-Fuc4NAc (lipid III), the third lipid-linked intermediate involved in ECA synthesis.
4162972		putative inner membrane protein
4186636		putative arylsulfatase regulator
4212456	<i>rrlA</i>	"
4230132	<i>glfA</i>	Catalyzes the ATP-dependent biosynthesis of glutamine from glutamate and ammonia.
4242194	<i>yihQ</i>	Alpha-glucosidase
4248738	<i>yihV</i>	Sulfofructose kinase
4285755		
4312781	<i>hslU</i>	ATPase subunit of a proteasome-like degradation complex; this subunit has chaperone activity.
4315832	<i>cytR</i>	DNA-binding transcriptional regulator
4341382	<i>frwC</i>	PTS system fructose-like IIC component
4345376	<i>pfk</i>	Pyruvate formate lyase II activase
4353316	<i>argC</i>	Catalyzes the NADPH-dependent reduction of N-acetyl-5-glutamyl phosphate to yield N-acetyl-L-glutamate 5-semialdehyde.
4367423	<i>rrlB</i>	"
4383934	<i>rpoC</i>	DNA-dependent RNA polymerase catalyzes the transcription of DNA into RNA using the four ribonucleoside triphosphates as substrates.
4410803	<i>rrlE</i>	Probably a phage tail protein
4422051	<i>meth</i>	Methionine synthase
4422597	<i>meth</i>	Methionine synthase
4425744		
4428326		putative cytoplasmic protein
4453813	<i>pgi</i>	Glucose-6-phosphate isomerase
4498488		putative inner membrane protein
4510031		putative inner membrane protein
4512158		putative inner membrane protein
4544554		
4563616	<i>dauS</i>	Histidine kinase
4599778	<i>fadC</i>	Two distinct, membrane-bound, FAD-containing enzymes are responsible for the catalysis of fumarate and succinate interconversion; fumarate reductase is used in anaerobic growth, and succinate dehydrogenase is used in aerobic growth.
4601159	<i>yjeM</i>	Inner membrane transporter
4603171	<i>yjeP</i>	Mechanosensitive channel protein
4606105	<i>psd</i>	Catalyzes the formation of phosphatidylethanolamine (PtdEtn) from phosphatidylserine (PtdSer)
4608075	<i>orn</i>	3'-to-5' exoribonuclease specific for small oligoribonucleotides
4619855	<i>hfk</i>	HRK could encode or regulate a protease
4630083	<i>yjK</i>	Putative cytoplasmic protein
4651224	<i>yjG</i>	Nrma domain-containing protein
4659723	<i>yjM</i>	Autotransporter assembly factor
4672673		sugar transporter
4687149		putative carbohydrate kinase
4695198	<i>pmbA</i>	Metalloprotease PmbA
4696115	<i>cybC</i>	Soluble cytochrome b562-Electron-transport protein of unknown function.
4725096	<i>argI</i>	Reversibly catalyzes the transfer of the carbamoyl group from carbamoyl phosphate (CP) to the N(epsilon) atom of ornithine (ORN) to produce L-ornithine.
4743769		putative DNA helicase
4745250		putative DNA helicase
4747780		putative DNA helicase
4748007		putative DNA helicase
4778370	<i>yjE</i>	Cell density-dependent motility repressor
4798141	<i>mrr</i>	Mrr_cat domain-containing protein
4804820		putative transcriptional regulator
4821700	<i>fluF</i>	Ligase
4836394	<i>deoA</i>	The enzymes which catalyze the reversible phosphorolysis of pyrimidine nucleosides
4847924	<i>radA</i>	DNA-dependent ATPase involved in processing of recombination intermediates, plays a role in repairing DNA breaks.
4857384	<i>creB</i>	Response regulator in two-component regulatory system with CreC

Gene information was obtained from NCBI. The italicized sites don't match with any gene regions. The genes in bold correspond to virulence genes (suggested by literature).

SUPPLEMENTARY FIGURE LEGENDS

Fig. S1 Spacer conservation of various serovars of *Salmonella*. Spacer conservation in **a.** CRISPR1 array and **b.** CRISPR2 array across *Salmonella* serovars. **c & d.** Inter-serovar spacer conservation in both, **c.** the CRISPR1 and **d.** CRISPR2 arrays. The colour code for a particular column represents spacer sequences with greater than 90% nucleotide similarity. The number denotes the position of the spacer from the leader sequence. The direct repeats have been eliminated for simplicity. The asterisk (*) represents the host-specific serovars. The duplication and triplication is depicted as a pattern in the coloured box. The images were generated using Microsoft Excel 365.

Fig. S2. The sequence logo of the consensus DR sequence and last direct repeat. The DR sequence of all the strains belonging to the CRISPR1-STM, CRISPR1-STY and CRISPR2 were considered and the web-logo of these were generated. The last direct repeat (distal to the leader sequence) was observed to contain SNPs. **a.** CRISPR1-STM consensus DR, **b.** CRISPR1-STM last DR, **c.** CRISPR1-STY consensus DR, **d.** CRISPR1-STY last DR, **e.** CRISPR2 consensus DR, and **f.** CRISPR2 last DR. The sequence logos were generated using WEBLOGO ver 2.8.2 available at <https://weblogo.berkeley.edu/>

Fig. S3. The phylogenetic trees of a. CRISPR1 leader and b. CRISPR2 leader. The phylogenetic tree were constructed using PHYML version 3.1 with statistical tests for branch support. The values indicate the aLRT scores.

Fig. S4. The sequence logos of the leader sequence for each CRISPR array. The leader sequence of all the strains were aligned and a Web-logo was generated. The CRISPR1 leader sequence of *Salmonella enterica* subsp. *enterica* is of two types CRISPR1-STM and CRISPR1-STY. The CRISPR2 leader sequence is present and highly conserved among all the serovars except *Salmonella enterica* subsp. *arizonae* and subsp. *diarizonae*. **a.** CRISPR1-STM *S. enterica* subsp. *enterica*, **b.** CRISPR1 *S. enterica* subsp. *arizonae* and subsp. *diarizonae enterica* **c.** CRISPR1 *Salmonella bongori* **d.** CRISPR1-STY *S. enterica* **e.** CRISPR2 *S. enterica* subsp. *enterica*. **f.** CRISPR2 *Salmonella bongori*. The sequence logos were generated using WEBLOGO ver 2.8.2 available at <https://weblogo.berkeley.edu/>

Fig. S5. Heat map representing the spacer conservation for a. CRISPR1 array b. CRISPR2 array c. both the CRISPR arrays combined. The unique set of CRISPR spacers of each serovar was compared with the remaining serovars for their presence. The spacers showing 90% sequence similarity were considered to be same. Based on this the proportion of spacers shared across serovars was analysed and heat map was made. The heat maps were created using the tool Seaborn version 0.11 available at <https://seaborn.pydata.org/>¹.

Fig. S6. Orientation of the CRISPR array and the *cas* operon in *Salmonella*. Five types of arrangements were evident in *Salmonella*. The *cas*-STY arrangement present in strains belonging to the CRISPR1-STY clade. The *cas*-STM type operon were subdivided into four types - *S. enterica* subsp. *enterica* (*cas*-STM, belong to CRISPR1-STM clade), *S. bongori* (*cas*-STM.B), *S. enterica* subsp. *enterica*, subsp. *arizonae* (*cas*-STM.A) and subsp. *diarizonae* (*cas*-STM.D). All the strains of serovar Montevideo-STM and *S. enterica* subsp. *diarizonae* str. MZ0080 (used in our study) contain a non-sense mutation in *cas3* and is represented by an asterisk (*). *Salmonella bongori* str. SA19983605 (used in our study) does not contain the *cas7* gene and is depicted by a hash (#) and all strains of *S. enterica* subsp. *arizonae* contain a stop codon in the *cas3* operon represented by a thunderbolt.

Fig. S7. The percentage conservation of the entire *cas* operon. The values in the matrix represent the percentage nucleotide match between the categories. The reference strains used were *S. enterica* serovar Typhimurium str.14028S, Typhi str. CT18, *S. enterica* subsp. *arizonae* str. NCTC10047, *S. enterica* subsp. *diarizonae* str. MZ0080 and *S. bongori* str. SA19983605.

Fig. S8. The phylogenetic tree of *cas3* gene. The phylogenetic tree were constructed using PHYML version 3.1 with statistical tests for branch support. The values indicate the aLRT scores.

Fig. S9. The multiple sequence alignment of Cas3 and Cse1 protein by Clustal Omega. a. Cas3 The domains were predicted using InterPro (<https://www.ebi.ac.uk/interpro/>). The alignment was done using NCBI BLAST. The similarity between the three domains- HD domain, DEAD box and Helicase C-terminal are indicated by yellow, green and blue boxes respectively. **b. Cse1** The bold red color represents fully conserved residues (*) and strongly similar groups with a score >0.5 in Gonnet PAM 250 matrix (:). The red color represent groups of weekly similar properties with a score <0.5 (.) for the residues essential for PAM recognition. The residues essential for Cas3 protein recruitment is indicated by the red box

Fig. S10. The phylogeny of CRISPR1 leader. The CRISPR1 leader sequences of 128 strains were aligned using MUSCLE and the phylogenetic trees was constructed by Maximum Likelihood.

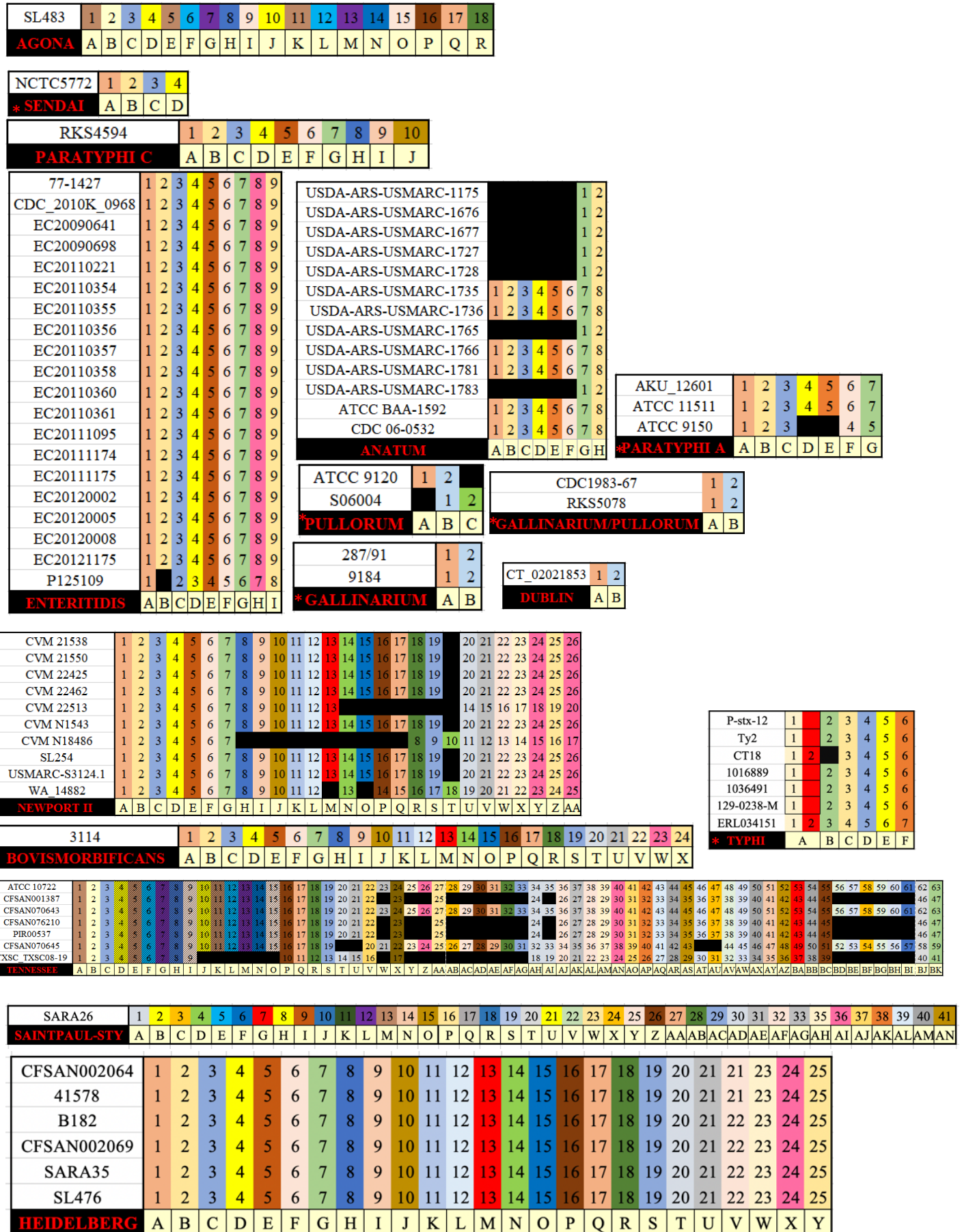
Fig. S11. Heat map for sharing of protospacer source by pairs of serovars for spacers belonging to a. CRISPR1 array and b. CRISPR2 array. The protospacer sources - phages, plasmids and DNA viruses were extracted using the tool CRISPR target. The protospacers were matched among all serovars and a map of same targets were created. The heat maps were created using the tool Seaborn version 0.11 available at <https://seaborn.pydata.org/>¹.

Fig. S12. Graph representing the relationship between the no. of spacers and no. of protospacer for a. CRISPR1 array and b. CRISPR2 array. The X-axis values is in the increasing number of spacers. Y-axis represent the values of obtained protospacer matches corresponding to all the spacers of the array. The graphs were created using GraphPad Prism 8 available at <https://www.graphpad.com/>

Fig. S13. Sequence alignment of the region between the transposase and CRISPR1 in *S. enterica* subsp. *enterica* serovar Typhimurium and *Citrobacter freundii* complex sp. CFNIH3. The region between the transposase and CRISPR1 was extracted from NCBI database. The sequences were then aligned using NCBI nucleotide BLAST. 81.98% identity was obtained for 51% query cover yielding nearly 40% sequence similarity.

Figure S1

a.



b.

USDA-ARS-USMARC-1175	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26
USDA-ARS-USMARC-1676	1	2								3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19
USDA-ARS-USMARC-1677				1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21		
USDA-ARS-USMARC-1727	1	2	3															4	5	6	7	8	9	10	11	12
USDA-ARS-USMARC-1728	1	2	3	4	5	6	7	8	9		10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25
USDA-ARS-USMARC-1735																		1	2		3	4	5	6	7	8
USDA-ARS-USMARC-1736	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26
USDA-ARS-USMARC-1765	1	2	3	4	5	6		7	8	9	10	11					12	13	14	15	16	17	18	19	20	21
USDA-ARS-USMARC-1766	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24		
USDA-ARS-USMARC-1781	1	2	3	4	5	6	7	8	9	10							11	12	13	14	15	16	17	18	19	20
USDA-ARS-USMARC-1783	1	2	3	4	5	6	7	8	9		10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25
ATCC BAA-1592	1	2	3	4	5	6	7	8	9	10							11	12	13	14	15	16	17	18	19	20
CDC 06-0532	1	2	3	4	5	6	7	8	9	10							11	12	13	14	15	16	17	18	19	20
ANATUM	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U	V	W	X	Y	Z

77-1427	1			2	3	4	5	6	7	8																	
CDC_2010K_0968	1	2	3	4	5	6	7	8	9	10	11																
EC20090641	1	2	3	4	5	6	7	8	9	10	11																
EC20090698	1	2	3	4	5	6	7	8	9	10	11																
EC20110221	1	2	3	4	5	6	7	8	9	10	11																
EC20110354	1	2	3	4	5	6	7	8	9	10	11	12															
EC20110355	1	2	3	4	5	6	7	8	9	10	11	12															
EC20110356	1	2	3	4	5	6	7	8	9	10	11	12															
EC20110357	1	2	3	4	5	6	7	8	9	10	11	12															
EC20110358	1	2	3	4	5	6	7	8	9	10	11	12															
EC20110360	1	2	3	4	5	6	7	8	9	10	11	12															
EC20110361	1	2	3	4	5	6	7	8	9	10	11	12															
EC20111095	1	2	3	4	5	6	7	8	9	10	11	12															
EC20111174	1	2	3	4	5	6	7	8	9	10	11	12															
EC20111175	1	2	3	4	5	6	7	8	9	10	11	12															
EC20120002	1	2	3	4	5	6	7	8	9	10	11	12															
EC20120005	1	2	3	4	5	6	7	8	9	10	11	12															
EC20120008	1	2	3	4	5	6	7	8	9	10	11	12															
EC20121175	1	2	3	4	5	6	7	8	9	10	11	12															
P125109	1	2		3	4	5	6	7	8	9	10																
ENTERITIDIS	A	B	C	D	E	F	G	H	I	J	K	L															

CFSAN002064	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18									
41578	1	2	3	4	5		6	7	8	9	10	11	12	13	14	15	16	17	18								
B182	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18									
CFSAN002069	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18									
SARA35	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18									
SL476	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18									
HEIDELBERG	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R									

CVM 21538	1	2	3	4	5	6	7	8	9	10	11	12	13	14		15	16	17	18	19							
CVM 21550	1	2	3	4	5	6	7	8	9	10	11	12	13	14		15	16	17	18	19							
CVM 22425	1	2	3	4	5	6	7	8	9	10	11	12	13	14		15	16	17	18	19							
CVM 22462	1	2	3	4	5	6	7	8	9	10	11	12	13	14		15	16	17	18	19							
CVM 22513	1	2	3	4	5	6	7	8	9	10	11	12	13	14		15	16	17	18	19							
CVM N1543	1	2	3	4	5	6	7	8	9	10	11	12	13	14		15	16	17	18	19							
CVM N18486	1	2	3	4	5	6	7	8													9	10	11	12			
SL254	1	2	3	4	5	6	7	8	9	10	11	12	13	14		15	16	17	18	19							
USMARC-S3124.1	1	2	3	4	5	6	7	8	9	10	11	12	13	14		15	16	17	18	19							
WA_14882	1	2	3	4	5	6	7	8			9	10	11	12	13	14	15	16	17	18	19						
NEWPORT II	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U						

NCTC5772	1																										
*SENDAI	A																										
287/91	1	2	3	4	5	6	7	8	9	10																	
9184	1	2	3	4	5	6	7	8	9	10																	
*GALLINARIUM	A	B	C	D	E	F	G	H	I	J																	

ATCC 9120	1	2	3	4	5	6																					
S06004	1					2																					
*PULLORUM	A	B	C	D	E	F																					

CT_02021853	1	2	3	4	5																						
DUBLIN	A	B	C	D	E																						

CDC1983-67	1	2	3	4	5	6																					
RKS5078	1	2	3	4	5	6																					
* GALLINARIUM/PULLORUM	A	B	C	D	E	F																					

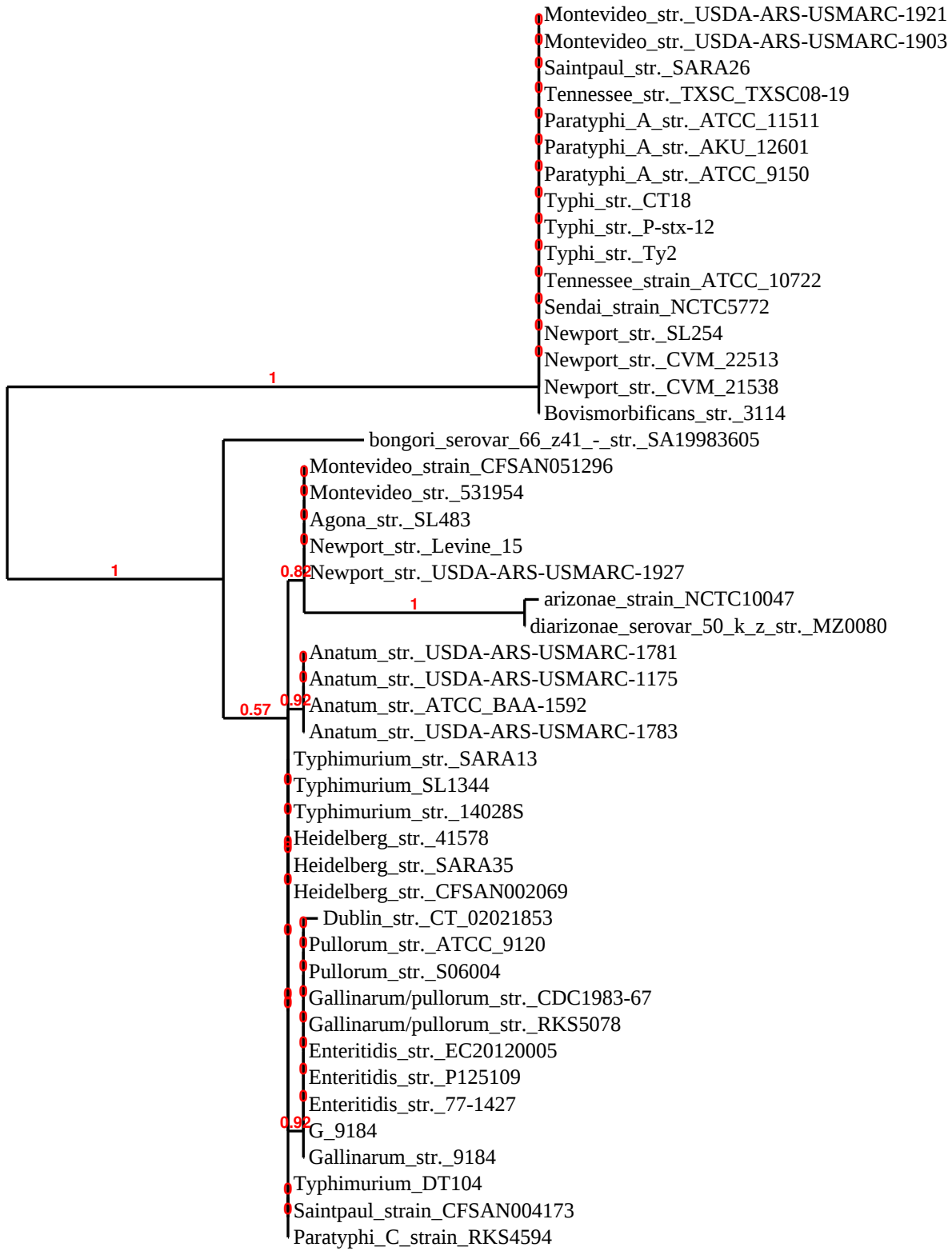
AKU_12601	1	2	3																								
ATCC 11511	1	2	3																								
ATCC 9150	1	2	3																								
* PARATYPHI A	A	B	C																								

CFSAN004173	1	2	3	4	5	6	7	8	9	10		11	12	13	14	15	16	17	18	19	20							
CFSAN004174	1	2	3	4	5	6	7	8	9	10		11	12	13	14	15	16	17	18	19	20							
CFSAN004175	1	2	3	4	5	6	7	8	9	10		11	12	13	14	15	16	17	18	19	20							
SGB23	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22						
SAINTPAUL-STM	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U	V						

SARA26	1	2	3	4	5	6	7	8	9	10	11	12	13	14
--------	---	---	---	---	---	---	---	---	---	----	----	----	----	----

Figure S3

a.



b.

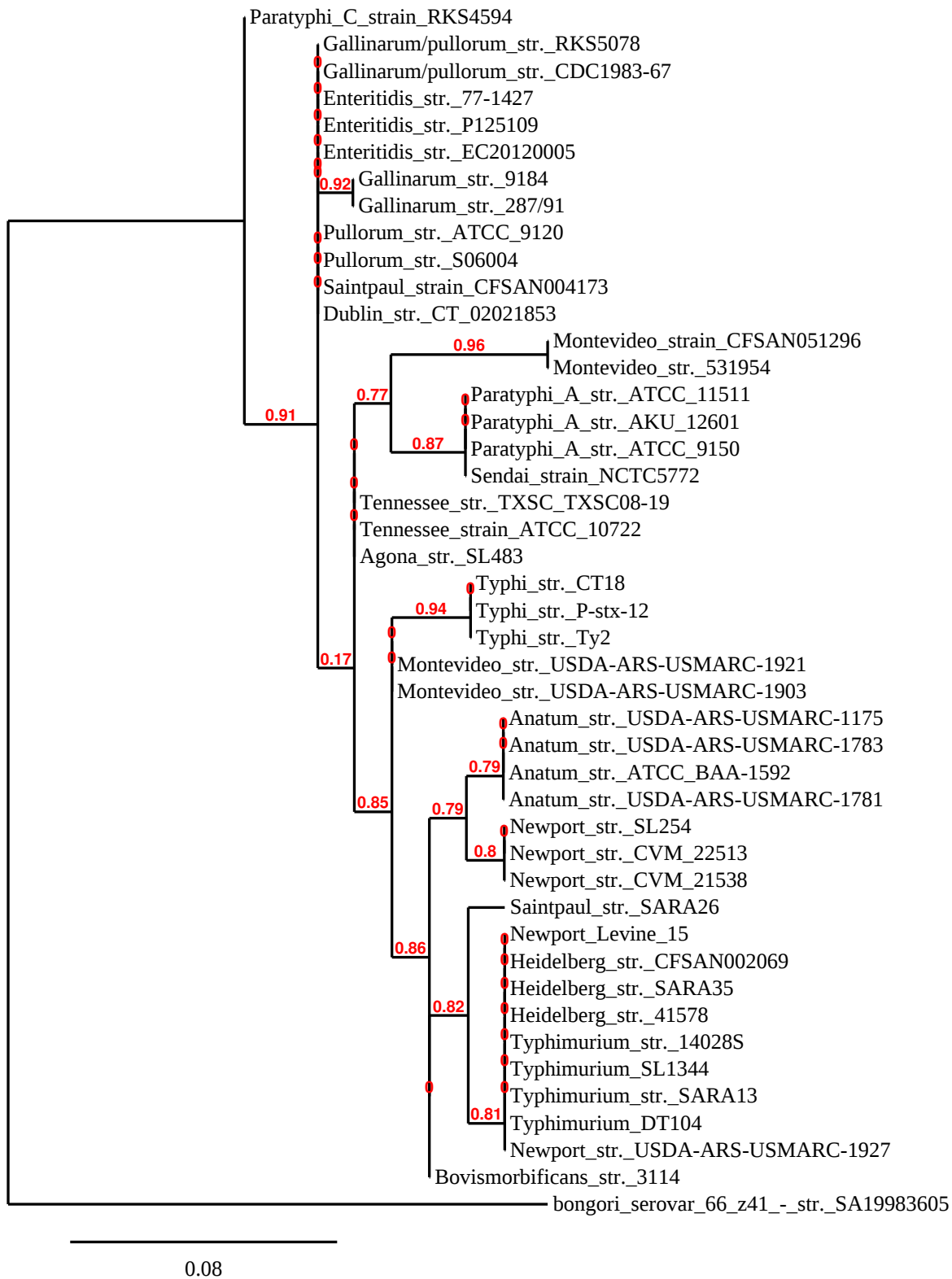
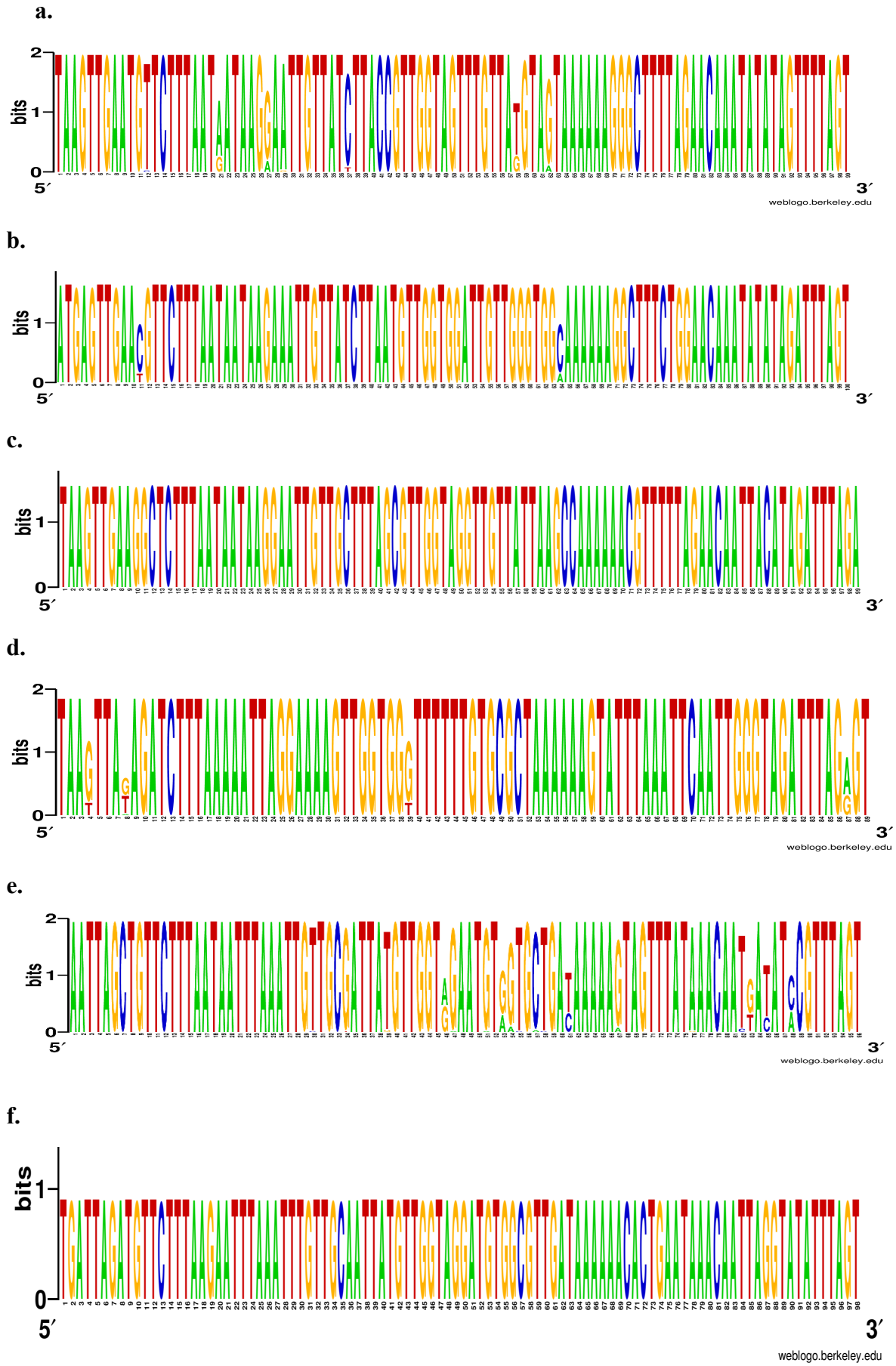


Figure S4



C.

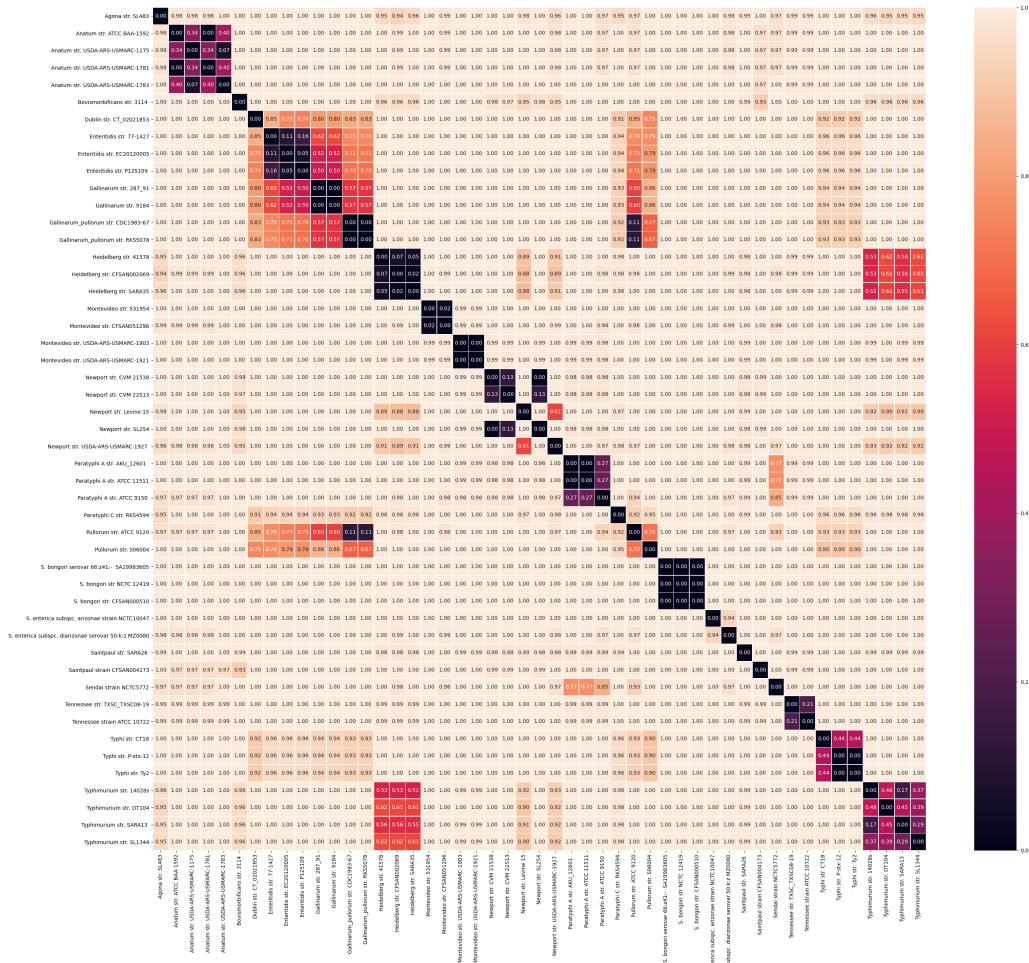


Figure S6

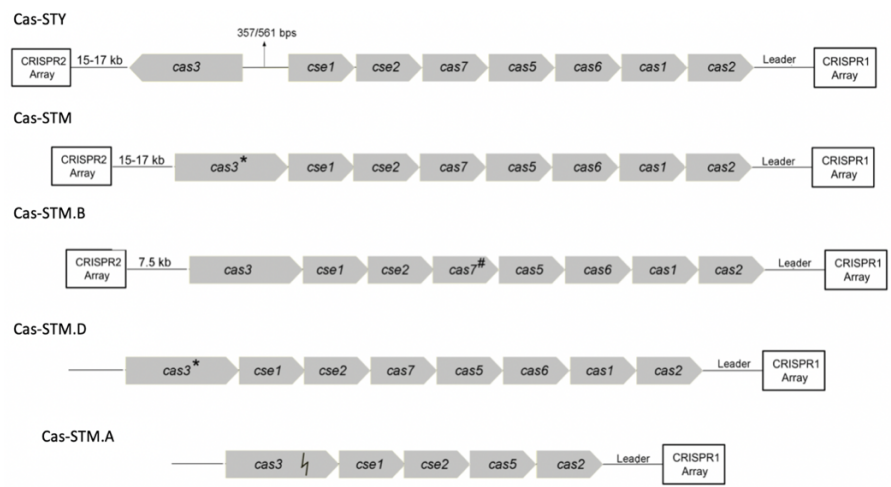
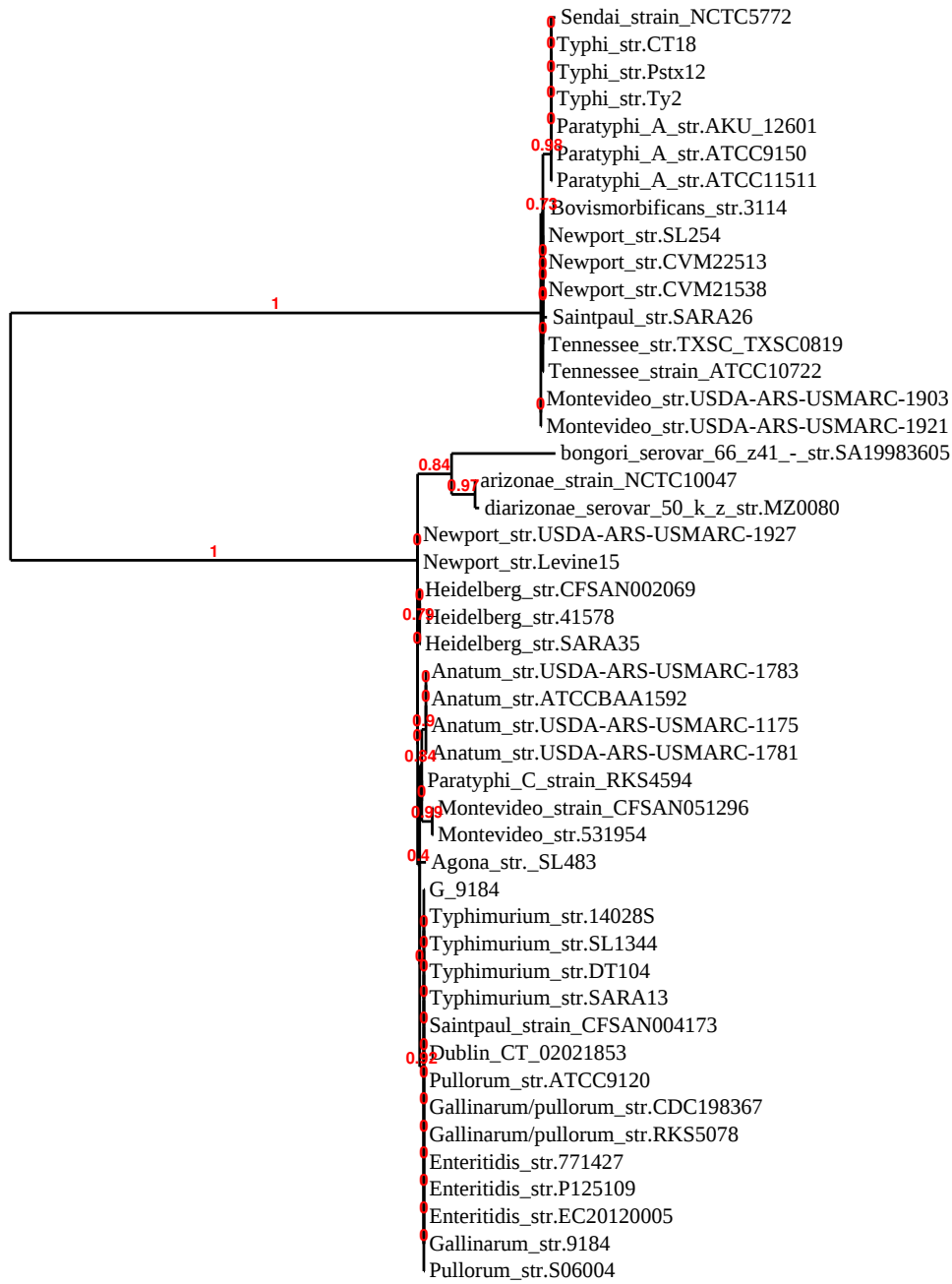


Figure S7

	Cas-STM	Cas-STY	Cas-STM.A	Cas-STM.D	Cas-STM.B
Cas-STM		28.6	87	87	85
Cas-STY			23.5	24	29
Cas-STM.A				99	88
Cas-STM.D					88.2
Cas-STM.B					

Figure S8



0.3

Figure S9

a.

HD Domain - 48%

YWGKSRGETDGGD **DYHLLCWHSLDVA**AVGYWMVINNIYFIDHYLKKLGIQDKEQAAQFF Typhimurium
YWGK+ + + +YHLL +H LDVAA G +++ NN + + LK+ I A +
YWGKTDKSTENITC **NYHLLAYHCLDVA**ACGEYIIKNNTFNSCNILKECNISGT-NAENWI Typhi

AWILCWHDIGKFAHSFQQLYRHEALNIFNEPTRHYEKIAHTTLGYMLWN----SWLSECP Typhimurium
W L HDIGKFA FQ+ + + + H +LG+ LW +W S
IWFLASHDIGKFARGFQKYAEFPDSPLVSPISGVSAFERHDSLGFYLVGKLF~~EA~~WSSGSN Typhi

ELFPSSLSVR-KSKRVMALWMPVTTGHHGRPPE **AIQELDHFRQ**QDKD--AARDFLLRIK Typhimurium
E+ R + + + WM ++TGHHG PP+ ++ D+D AA +L +
EIIAGIEPENRARFESALNSWMLISTGHHGIPPD **TMKNRSSLAFT**DEDIVAATHYLEALS Typhi

ALFPLITLPEAWDEDEGIDQFQQLSWFISA **AVVLADWTGSASRYF**PRTAEKMPVDTYWQQ Typhimurium
LFP TLP+ W G +Q SWF + V LADW GS FP + MP+ YW
ELFPF-TLPQEWKTKAGRKCLKQHSWFFAGL **VTLADW**MGSD~~ESQ~~FPLLSSAMPLKDYWPL Typhi

DEAD Box - 77%

ALAKAQTAITLFP~~SAANVSAFTGIETLFPFIQHP~~ **TPLQOKALELDINVDGAQL**FILEDVT Typhimurium
A KAQ AI P + S + LFPFIQ **TPLQO**+A ELDI+ GAQL +LEDVT
ACEKAQQAILRMPPLSQHSHYQDHRALFPFIQ **TPLQQRASELDISAPGAQL**VVLEDVT Typhi

GAGKTEAALILAHRLMAAGKAQGLYFGLPTMATANAMFERMANTWLALYQPDSRPSLILA Typhimurium
GAGKTEAALIL HRLM+A K GLY GLPTMATANAM++R+A+ + AL+ +SRPSLILA
GAGKTEAALILTHRLMSANKGHGLYVGLPTMATANAMYQRLASAYHALFTDES~~SRPSLILA~~ Typhi

HSARRLMDRFNQS IWSVTL~~SGTEE--PDEAQPYSQ~~CAAWFADSNKKALLAEVGVGTL~~DQ~~ Typhimurium
H R + D F QSIW T + E+ D+ ++ C WFADS KKALLAEVGVGTL~~DQ~~
HGGREMSDSFRQSIWQPTENIAEDYARDDGNATTE-CHTW~~FADSRK~~KALLAEVGVGTL~~DQ~~ Typhi

AMMAVMPFKHNNLRL~~LGLSNKILLADEIHACDAWMSRILEGLIERQASNGNATILLSATL~~ Typhimurium
+MAVMPF+H +LRLLG+ +KILL DE+HA D +M ++LEGL+ A+ G + I+LSATL
LLMAVMPFRHQSLRL~~LGM~~RDKILL~~LDEVHAYDGYMVKLLEGLLRFHAAQGGSAI~~ILSATL Typhi

SQQORDKLVA~~AFSRGVR~~SVQAPLLGHDD--YPWLTQVTQTELISQ~~RVDTRKEVERCVDI~~ Typhimurium
R+KL+ AFS G G D+ YPWL+ +T + L+ Q + TR EV+R V +
PAALREKLLNAFSDGAEFMSAG---GSDNAGYPWLSHLTSSGLLEQPLATRPEVQRTVAV Typhi

Helicase C-terminal - 81%

GWLHSEEACLERIGEAVEKEGNCIA **WIRNSVDDAIRIYRQLQ**LSKVVVTENLLLFHSRFAF Typhimurium
W+ + L+ I V G CI WIRN+VDDA+ Y+QL +V ++LLLFHSRFAF
NWIQQRQEALDIIYRVVATGQCIC **WIRNTVDDALD**TYQQLLHEGIVPEQDLLLFHSRFAF Typhi

YDRQRIESQTLNLF~~GKQSG-AQRAGKVI~~IATQVIEQSLDIDCEMISDLAPVDLLIQ~~RAG~~ Typhimurium
DR IE++TLN FG + ++R GKV+IATQV+EQSLD+D D MI+DLAP+DLLIQ~~RAG~~
IDRIA IENKTLNWFGNAPVSERRGKVL IATQVVEQSLDLD~~DFDWMITDLAPIDLLIQ~~RAG Typhi

RLQRH **IRDRNGLVKKSGQDERETPVL**RILAPEWDDAPR Typhimurium
RLQRH **IRD +G K + DER+ PVL I P A R**
RLQRH **IRDAHGQRKSTLPDERQPPVL**HISGPSLAGR Typhi

b.

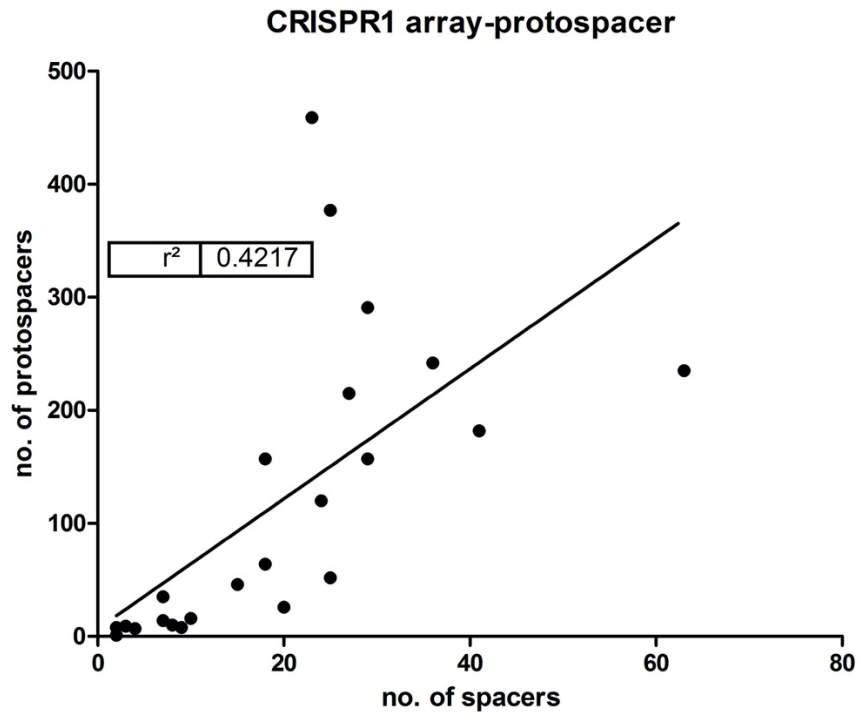
E.coli	---MNLIDNWIPVRPRNGGKQVINLQSLYCSRQWRLSLPRDDMELALALLVCIGQI	57
Typhimurium	MDNFSLLTTPWLPVRFKDGSTGKLAPVDLA--DENVVDAATRADLQGAAWQFLGLLQC	58
Typhi	---MDLTKEKWLPIVFSNGDKKKISLRDLL--DNRIQDLAYPRADFQGAAWQLIGILQC	55
	:.* ** :*. : : .. : : * **: ** :* : *	
E.coli	IAPAKDDVEFRHRIMNPLTEDEFQQLIAPWIDMFYLNHAEHPFMQTKGVKANDVTPMEKL	117
Typhimurium	SIAPKRYKNWEDIWFDGLHADVLHKALAPLEHAFQFGAETPSFMQDFEPLSSEKVSIASL	118
Typhi	TVAPEDKEEWADIWHESIIEFEQWEKALNTISLALQFGEQKPSFLQSFDPDSEYGSIAGL	115
	: :. : : : : : : : * : * : : *	
E.coli	LAGVSGATNCA---FVNQPQGEALCGGCTAIALFNQANQAPGFGGFKSGLRGGTPVT	173
Typhimurium	LPEIPGAQTTFKNDHFVKRGVTERFCPHCAALALFSLQLNAPAGGKGYRTGLRGGGPLT	178
Typhi	LVDAPGGNALKLNKDFVKRGNVEQICPHCAAIALFAIQTNSPAGGAGYRVGMRGGGPLT	175
	* * . . : * * : * * : * * : * * : * * : * *	
E.coli	TFVRGIDLRS-----TVLLNVLTPRLQKQFPNESHTENQPTWIKPIKSNEIPA--	223
Typhimurium	TLVELQEYQGERQTPWRKLNWVMPQDTADLPLPQC-DATVFPWLAATRTEQANAVT	237
Typhi	TLVVPQ---EEDKYLWKKLWLVLPQEEPP---NVTQ-HPLIFPWLAPTCTSEKAGNVV	228
	* : * : : : : : * : : : *	
E.coli	--SSIGFVRGLFWQPAHIELC-DPIGIGKSCCGQESNLRYTGFLKEKFTFTVNGLWPHP	280
Typhimurium	TPEQVNLQAYWGMRRIRLDFATLQSGCCDICGAESDELL-GFMTVKNYGVNYDGRWHP	296
Typhi	TPDNAHPLQAYWGMRRRIELDFHTVAGICDLGEGHESLL-LQMRSKNYGVQYDSWLHP	287
	. . : : : * : * . * * . * : : * : . . * * *	
E.coli	HSPCLVTVKRGEVEEKFLAFTTS--AP---SWTQISRVVVDKIIQENGNRVAAVVNQFR	335
Typhimurium	LTPYRAPVKDQN---AFFSVKPPGGLIWRDWLGLSQNNQTEAN---YESPAQVVKVFN	349
Typhi	FSPYRQALKDFSA--PWLAFKQPGGLSYKDWLGLMLNREDKFN---KMQPAKVVAAG	341
	: * : * . : : : . . . * : : : * * *	
E.coli	NIAPQSLELIMG--GY--RNQAS---ILERRHDVLMFNQGWQYGNVINEIVTVGLGY	388
Typhimurium	AR---SLTDVKAGIWFADGDFDNMKIRCWEHHPFLMTE---GLIPDLRKAVQTAARL	402
Typhi	QR-----NKMSLWCFAWDMNKAIVRCWYQHRIPLISVSHE-EQFLAALNIVLVLASES	393
	. : : : : : : : : : . : : .	
E.coli	----KTALRKALYFAEGFKNKDFKAGVSVHETAERHFYRQSELLIPDVLANVNFSDAD	444
Typhimurium	LSLLRSALKEAWFADAKGAR-GDFSFDIDFWNLTQGRFLNLIHDLE-----NGHKPD	454
Typhi	LSLLRNALKSAKFDCKPEAK-MDFSMVDIAFWQETEPAFRTLQEALAVDPLR--QDTQTR	450
	: * : * : : : * . : : : * . * : : :	
E.coli	EVIADLRDKLHQLCEMLFNQSVAPYAHHPKLI-STLALARATLYKHL-RELK-----P	495
Typhimurium	ERLNKQRELWLFTRHYFDDHVFTNPYESSDLE-RIMTARKKYFTTSAEKQSAAKAKK	513
Typhi	HAVSQWEAELAHYLFHVDRDALTNPCPDILQRLTARQDLAS-SYRKHKARKDVLAL	509
	. . . : * * : . . : ** . . .	
E.coli	QGGPSNG	502
Typhimurium	QEAAE--	518
Typhi	VE----	511

Figure S10

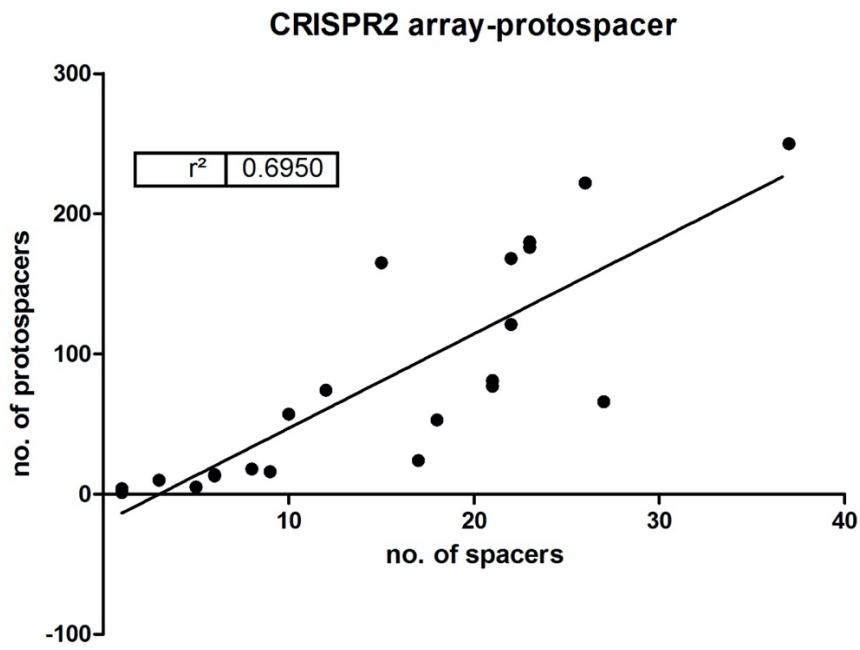


Figure S12

a.



b.



Supplementary Methodology

CRISPR loci data collection in correct orientation:

Our study comprises 133 strains belonging to two species, *S. bongori*, and *S. enterica*, including 22 serovars and three subspecies (supplementary table S1). These samples were primitively isolated from multiple sources, including primates, poultry, swine, cattle, food specimens, and natural environment (GenBank database). The complete genome sequences for all these annotated strains were obtained from the GenBank database. Only experimentally validated sequences were considered to ensure the legitimacy of the data being used. The CRISPR loci were identified in two steps - the annotation and orientation of the CRISPR array were retrieved from the online database of CRISPR-Cas⁺⁺¹⁹. The upstream and downstream regions of these arrays were aligned with the leader sequences previously reported by¹⁹ to know the correct sequence of the CRISPR array. The arrays were then classified as CRISPR1 and CRISPR2 after verifying the leader sequence and its position with respect to the *cas* operon²⁰.

Most strains of *S. enterica* subsp. *enterica* had both, the CRISPR1 and CRISPR2 arrays. However, all the analyzed strains of *S. enterica* subsp. *enterica* serovar Heidelberg, a few strains of serovar Typhimurium, and one strain of serovar Tennessee are reported to harbor more than two CRISPR arrays¹⁹. Instead, our analysis confirmed that the CRISPR1 array of serovars Typhimurium and Heidelberg were divided into two parts by a stretch of 74 nucleotides consisting of two truncated spacers and a direct repeat (DR) (supplementary fig. S1A). The two parts of the CRISPR1 array taken together in concatenation aligned well with the intact CRISPR1 array of other strains of serovar Typhimurium. Similarly, the CRISPR1 array of serovar Tennessee strain (str.) CFSAN070645 was divided into three parts (containing 19, 24, and 16 spacers) and the CRISPR2 into two parts (consisting 10 and 11 spacers) due to the presence of mutated DRs rendering a stretch of 91bp undetectable as a part of the CRISPR array. Therefore, we considered the concatenated forms of these CRISPR arrays as a single unit for further analysis. Our analysis also indicated the occurrence of CRISPR1 array with two spacers each, in the serovars Dublin, Gallinarum, Pullorum, and Gallinarum/Pullorum. However, neither of these CRISPR arrays were described as valid in the CRISPR-Cas⁺⁺ database, and the CRISPRCasFinder software allocated 27bp long DRs and 34bp long spacer sequences. Likewise, the CRISPR2 arrays of serovar Typhi and serovar Pullorum str. S06004 identified through our analysis were not detectable by this database. The CRISPR2 array of serovar Typhi possessed only one erratic spacer and that of serovar Pullorum str. S06004 had two spacers. We considered all these strains and their respective CRISPR-Cas systems in our analysis.

Protospacer analysis:

The spacer sequences were extracted from the CRISPR-Cas⁺⁺ database. The data of all the strains belonging to one particular serovar was combined. A unique spacer set was created for that serovar. The spacer sequences were then uploaded on CRISPRTarget tool²⁰. Genbank Phage, RefSeq-Plasmid and IMGVR databases were selected as the target databases. The parameters for initial BLAST screen in CRISPRTarget were kept default. Output obtained gave the accession number of the protospacer sources corresponding to these spacers. The hits obtained for Genbank Phage and RefSeq-Plasmid had accession numbers corresponding to NCBI. While, the accession number for the hits obtained from IMGVR database corresponds to IMG/VR viral resource. The accession numbers of the protospacer hits obtained were matched across serovars using a customized bash script. Based on these matches a heat map was created.

Mapping 236 spacer- and Cascade-binding sites obtained by ChIP seq of Cas5:

The ChIP analysis done by Stringer *et al.*²¹ revealed 236 binding sites. We mapped these sites on the complete genome (CP001363) available at NCBI and extracted the genes present at these sites. The functions of these gene were checked using UniProt. The genes having role in virulence (with support from literature) are marked in bold in Table S8.

References:

1. Bogomolnaya, L.M., Tilvawala, R., Elfenbein, J.R., Cirillo, J.D. & Andrews-Polymenis, H.L. Linearized Siderophore Products Secreted via MacAB Efflux Pump Protect *Salmonella enterica* Serovar Typhimurium from Oxidative Stress. *mBio*. **11**(2020).
2. Bearson, B.L. *et al.* Iron regulated genes of *Salmonella enterica* serovar Typhimurium in response to norepinephrine and the requirement of fepDGC for norepinephrine-enhanced growth. *Microbes. Infect.* **10**, 807-16 (2008).
3. Hensel, M., Hinsley, A.P., Nikolaus, T., Sawers, G. & Berks, B.C. The genetic basis of tetrathionate respiration in *Salmonella typhimurium*. *Mol. Microbiol.* **32**, 275-87 (1999).
4. Rodríguez-Morales, O. *et al.* *Salmonella enterica* serovar Typhimurium ompS1 and ompS2 mutants are attenuated for virulence in mice. *Infect. Immun.* **74**, 1398-402 (2006).
5. Coombes, B.K., Brown, N.F., Kujat-Choy, S., Vallance, B.A. & Finlay, B.B. SseA is required for translocation of *Salmonella* pathogenicity island-2 effectors into host cells. *Microbes. Infect.* **5**, 561-70 (2003).
6. Mühlig, A., Kabisch, J., Pichner, R., Scherer, S. & Müller-Herbst, S. Contribution of the NO-detoxifying enzymes HmpA, NorV and NrfA to nitrosative stress protection of *Salmonella* Typhimurium in raw sausages. *Food Microbiol.* **42**, 26-33 (2014).
7. Mariscotti, J.F. & García-del Portillo, F. Genome expression analyses revealing the modulation of the *Salmonella* Rcs regulon by the attenuator IgaA. *J. Bacteriol.* **191**, 1855-67 (2009).
8. Crouch, M.L., Castor, M., Karlinsey, J.E., Kalthorn, T. & Fang, F.C. Biosynthesis and IroC-dependent export of the siderophore salmochelin are essential for virulence of *Salmonella enterica* serovar Typhimurium. *Mol. Microbiol.* **67**, 971-83 (2008).
9. Johnson, R. *et al.* The Type III Secretion System Effector SptP of *Salmonella enterica* Serovar Typhi. *J. Bacteriol.* **199**(2017).
10. López-Garrido, J. & Casadesús, J. Crosstalk between virulence loci: regulation of *Salmonella enterica* pathogenicity island 1 (SPI-1) by products of the std fimbrial operon. *PLoS One*. **7**, e30499 (2012).
11. Tsolis, R.M. *et al.* SspA is required for lethal *Salmonella enterica* serovar Typhimurium infections in calves but is not essential for diarrhea. *Infect. Immun.* **68**, 3158-63 (2000).
12. El Hag, M. *et al.* Contribution of the csgA and bcsA genes to *Salmonella enterica* serovar Pullorum biofilm formation and virulence. *Avian. Pathol.* **46**, 541-547 (2017).
13. Choi, E. *et al.* Elongation factor P controls translation of the mgtA gene encoding a Mg. *Microbiologyopen*. **8**, e00680 (2019).
14. Velayudhan, J. *et al.* Distinct roles of the *Salmonella enterica* serovar Typhimurium CyaY and YggX proteins in the biosynthesis and repair of iron-sulfur clusters. *Infect. Immun.* **82**, 1390-401 (2014).
15. Tamayo, R., Prouty, A.M. & Gunn, J.S. Identification and functional analysis of *Salmonella enterica* serovar Typhimurium PmrA-regulated genes. *F.E.M.S. Immunol. Med. Microbiol.* **43**, 249-58 (2005).
16. Ledebor, N.A., Frye, J.G., McClelland, M. & Jones, B.D. *Salmonella enterica* serovar Typhimurium requires the Lpf, Pef, and Tafi fimbriae for biofilm formation on HEp-2 tissue culture cells and chicken intestinal epithelium. *Infect. Immun.* **74**, 3156-69 (2006).
17. Clayton, D.J. *et al.* Analysis of the role of 13 major fimbrial subunits in colonisation of the chicken intestines by *Salmonella enterica* serovar Enteritidis reveals a role for a novel locus. *B.M.C. Microbiol.* **8**, 228 (2008).
18. Waskom, M. mwaskom/seaborn. *Zendo* <https://doi.org/10.5281/zenodo.592845> (2020).
19. Couvin, D. *et al.* CRISPRCasFinder, an update of CRISRFinder, includes a portable version, enhanced performance and integrates search for Cas proteins. *Nucleic Acids Res.* **46**, W246-W251 (2018).
20. Shariat, N., Timme, R.E., Pettengill, J.B., Barrangou, R. & Dudley, E.G. Characterization and evolution of *Salmonella* CRISPR-Cas systems. *Microbiology*. **161**, 374-86 (2015).
21. Biswas, A., Gagnon, J.N., Brouns, S.J., Fineran, P.C. & Brown, C.M. CRISPRTarget: bioinformatic prediction and analysis of crRNA targets. *RNA Biol.* **10**, 817-27 (2013)
22. Stringer, A.M., Baniulyte, G., Lasek-Nesselquist, E., Seed, K.D. & Wade, J.T. Transcription termination and antitermination of bacterial CRISPR arrays. *Elife*. **9**(2020)