

Deletion of the nuclear receptor ROR α in macrophages does not modify the development of obesity, insulin resistance and NASH

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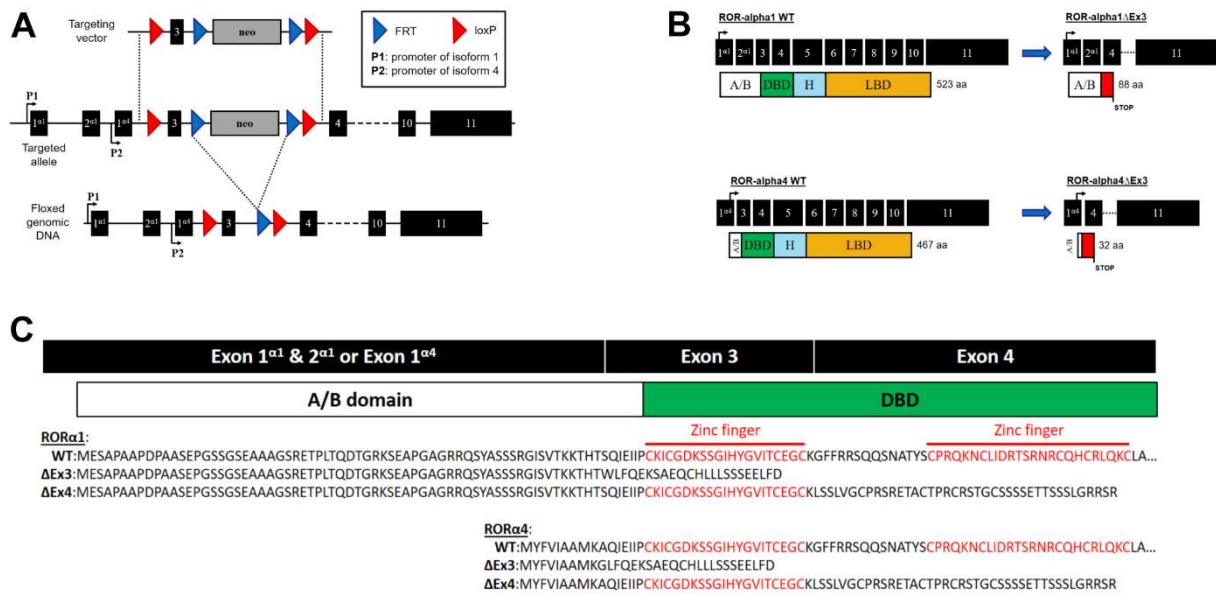


Figure S1: Characterization of ROR α MKO mice (Related to Figure 1).

(A) Schematic representation of the strategy to target exon 3 of *Rora*. **(B)** mRNA structures of ROR α 1 and ROR α 4 and the resulting proteins in both WT and MKO (Δ Ex3) mice. Exons are represented in black. Protein predictions are illustrated below mRNA and functional domains are indicated with colors. **(C)** Comparison of *Rora* exon 3 and exon 4 deletion on protein translation. Zinc finger motifs are indicated in red. Protein predictions were generated with Translate tool from ExPASy (<https://web.expasy.org/translate>) by using GenBank sequences for ROR α 1 (NM_013646.2) and ROR α 4 (NM_001289916.1).

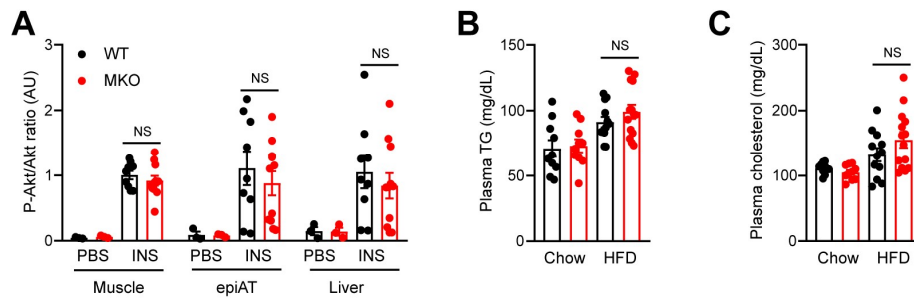


Figure S2: Effect of ROR α deletion in macrophages on HFD-induced obesity and IR (Related to Figure 2).

Ten weeks old WT and MKO mice were fed with either a chow or a HFD for 12 weeks. **(A)** Quantification of western blot for phospho-Akt in skeletal muscle, epididymal adipose tissue (epiAT) and liver. Quantification were performed with the Image studio Lite Version 4.0.21 software (LI-COR, <https://www.licor.com/bio/image-studio-lite/>) **(B)** Plasma triglyceride (TG). **(C)** Plasma cholesterol. Data are shown as mean \pm SEM. 2-way ANOVA followed by Sidak's multiple comparisons test was performed. All statistical analyses were carried out using GraphPad Prism 8 for Windows (GraphPad Software). n=10-15 mice per group. NS: Not significant; AU: Arbitrary unit.

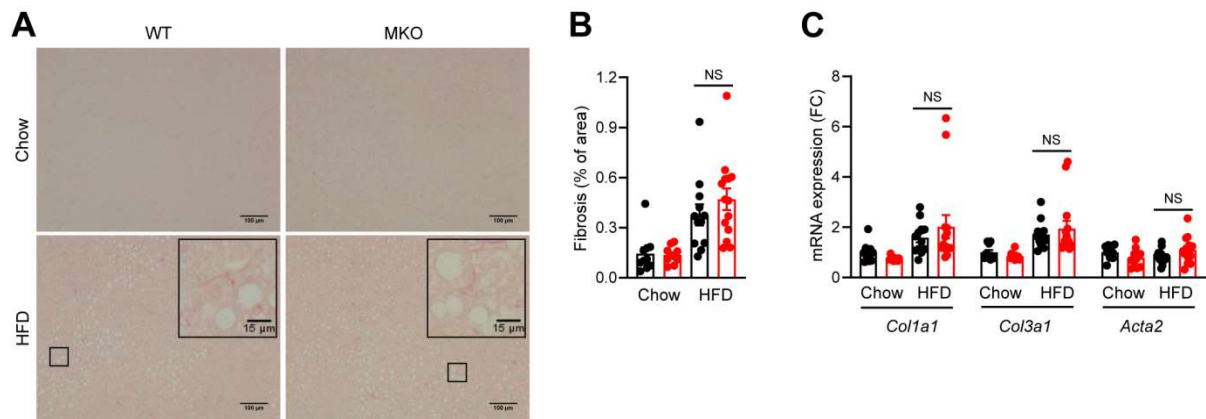


Figure S3: Effect of ROR α deletion in macrophages on HFD-induced fibrosis (Related to Figure 3).

Ten weeks old WT and MKO mice were fed with either a chow or a HFD for 12 weeks (**A-B**) Sirius red staining of liver sections (**A**) and quantification of fibrosis (**B**). Images were acquired on an Eclipse Ti-U microscope (Nikon) and quantified with Image J version 1.51j8 software (NIH, <https://imagej.nih.gov/ij/>). (**C**) mRNA expression levels measured by RT-qPCR for *Col1a1*, *Col3a1*, and *Acta2* genes in the liver. Data are shown as mean \pm SEM. 2-way ANOVA followed by Sidak's multiple comparisons test was performed. All statistical analyses were carried out using GraphPad Prism 8 for Windows (GraphPad Software). n=10-15 mice per group. NS: Not significant; FC: Fold Change

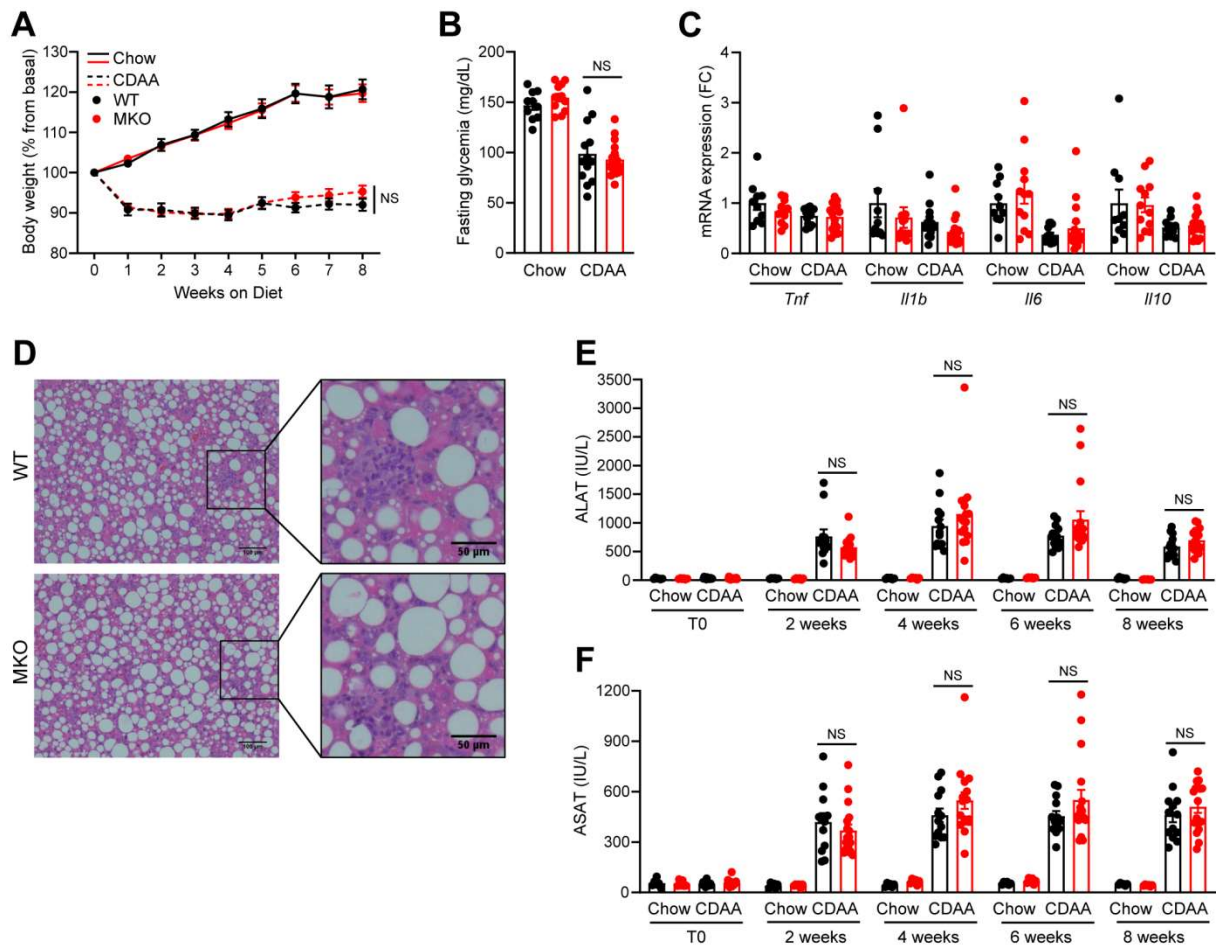


Figure S4: Effect of ROR α deletion in macrophages upon CDAA diet feeding (Related to Figure 4).

Ten weeks old WT and MKO mice were fed with either a chow or a CDAA diet for 8 weeks. **(A)** Body weight gain. **(B)** Fasting glycemia. **(C)** mRNA expression levels measured by RT-qPCR for *Tnf*, *Il1b*, *Il6*, and *Il10* genes in epiAT. **(D)** Hematoxylin & Eosin staining of liver sections. Images were acquired on an Eclipse Ti-U microscope (Nikon) and prepared with Image J version 1.51j8 software (NIH, <https://imagej.nih.gov/ij/>). **(E-F)** Kinetics of plasma transaminases activity. Data are shown as mean \pm SEM. 2-way ANOVA followed by Sidak's multiple comparisons test was performed. All statistical analyses were carried out using GraphPad Prism 8 for Windows (GraphPad Software). n=10-17 mice per group. NS: Not significant; FC: Fold Change.

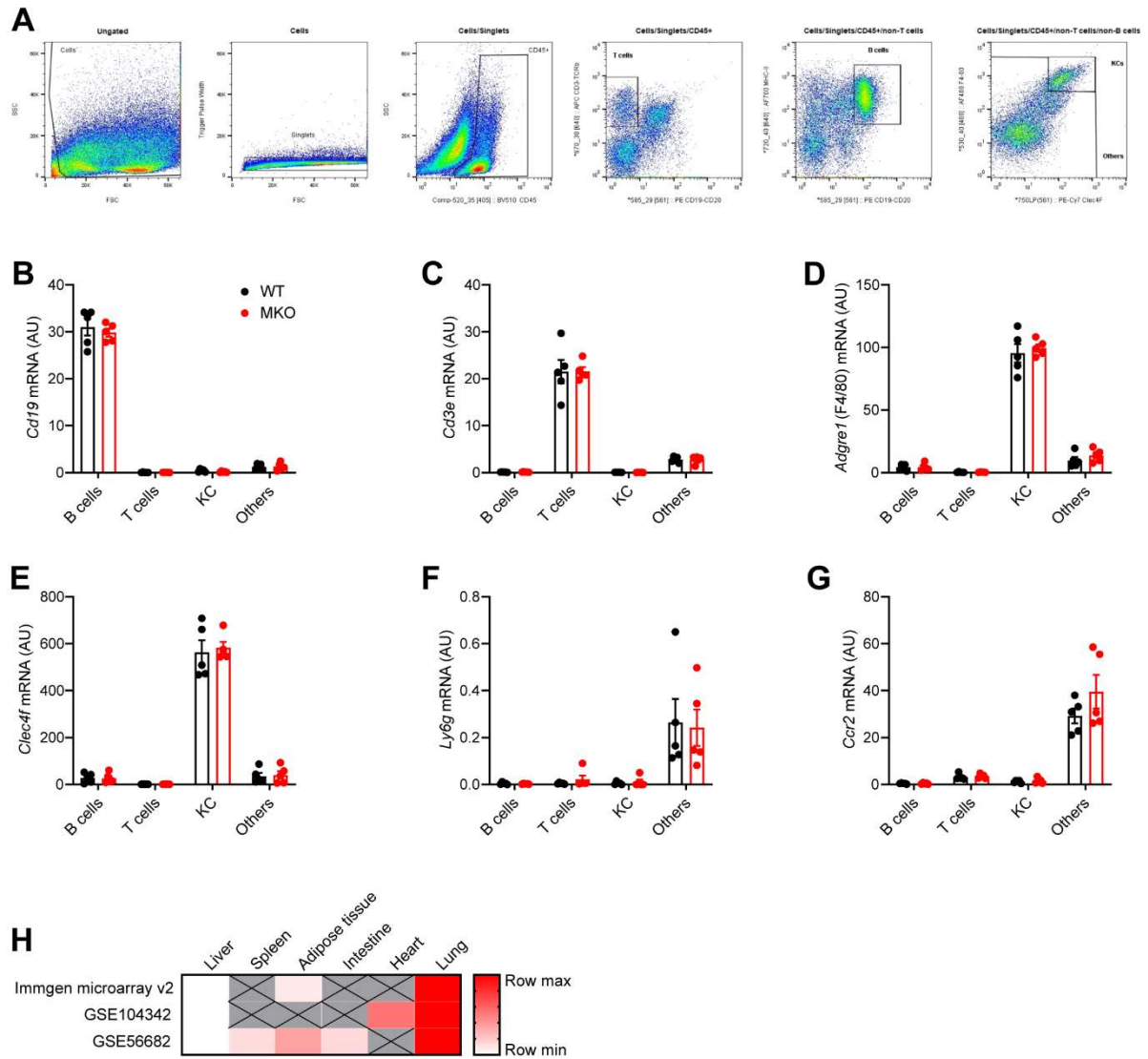


Figure S5: Quality control of Kupffer Cells (KCs) sorting (Related to Figure 5).

(A) Representative gating strategy for cell sorting of T cells, B cells, KCs and other cells (CD45⁺ non-T cells, -B cells and -KCs) from liver. (B-G) mRNA expression levels measured by RT-qPCR for the *Cd19* (B), *Cd3e* (C), *Adgre1* (D), *Clec4f* (E), *Ly6g* (F) and *Ccr2* (G) genes in B cells, T cells, KCs and Others sorted from the liver of WT and MKO mice. (H) Heatmap of *Rora* expression in different macrophage populations from public databases. Data are shown as mean \pm SEM. n=5 mice per genotype. AU: Arbitrary unit.

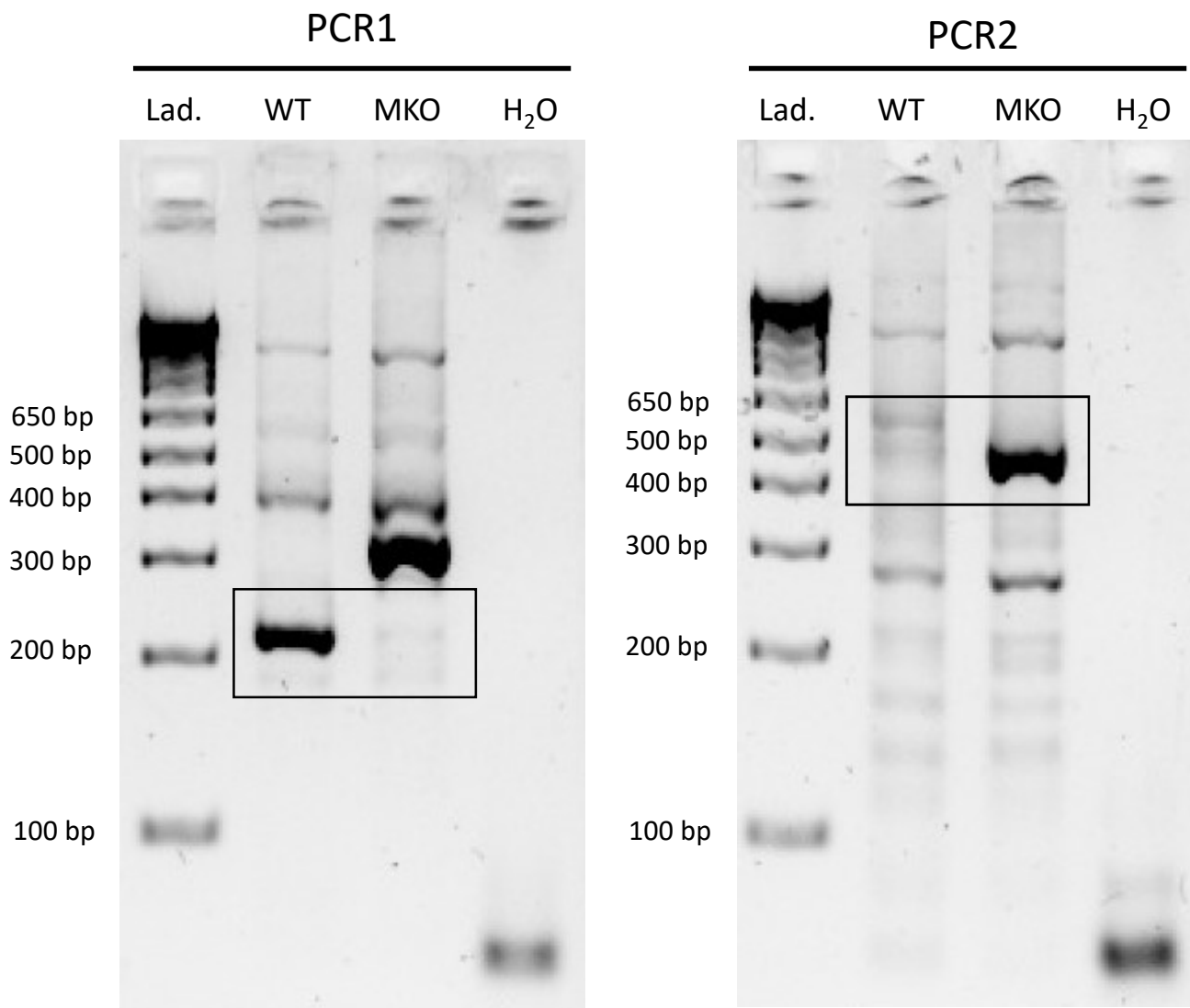


Figure S6: Uncropped agarose gel (related to Fig. 1B)

Lad., DNA Ladder.

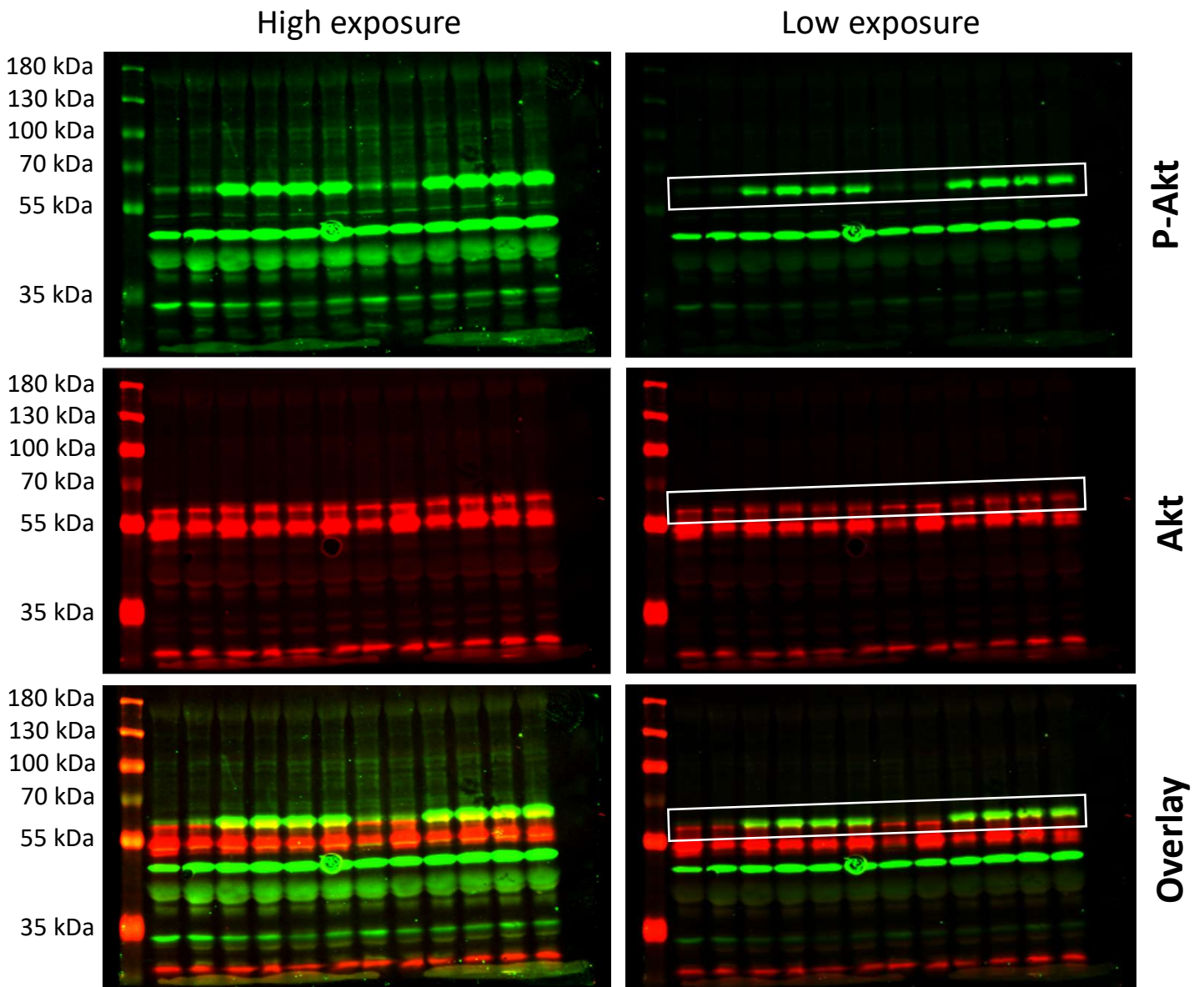


Figure S7: Uncropped WB images for muscle (related to Fig. 2H)

Akt molecular weight: 60kDa

Rabbit monoclonal anti-phospho-Akt (Ser473) (Clone 193H12) (Cell Signaling, #4058)

Mouse monoclonal anti-Akt (Pan) (Clone 40D4) (Cell Signaling, #2920)

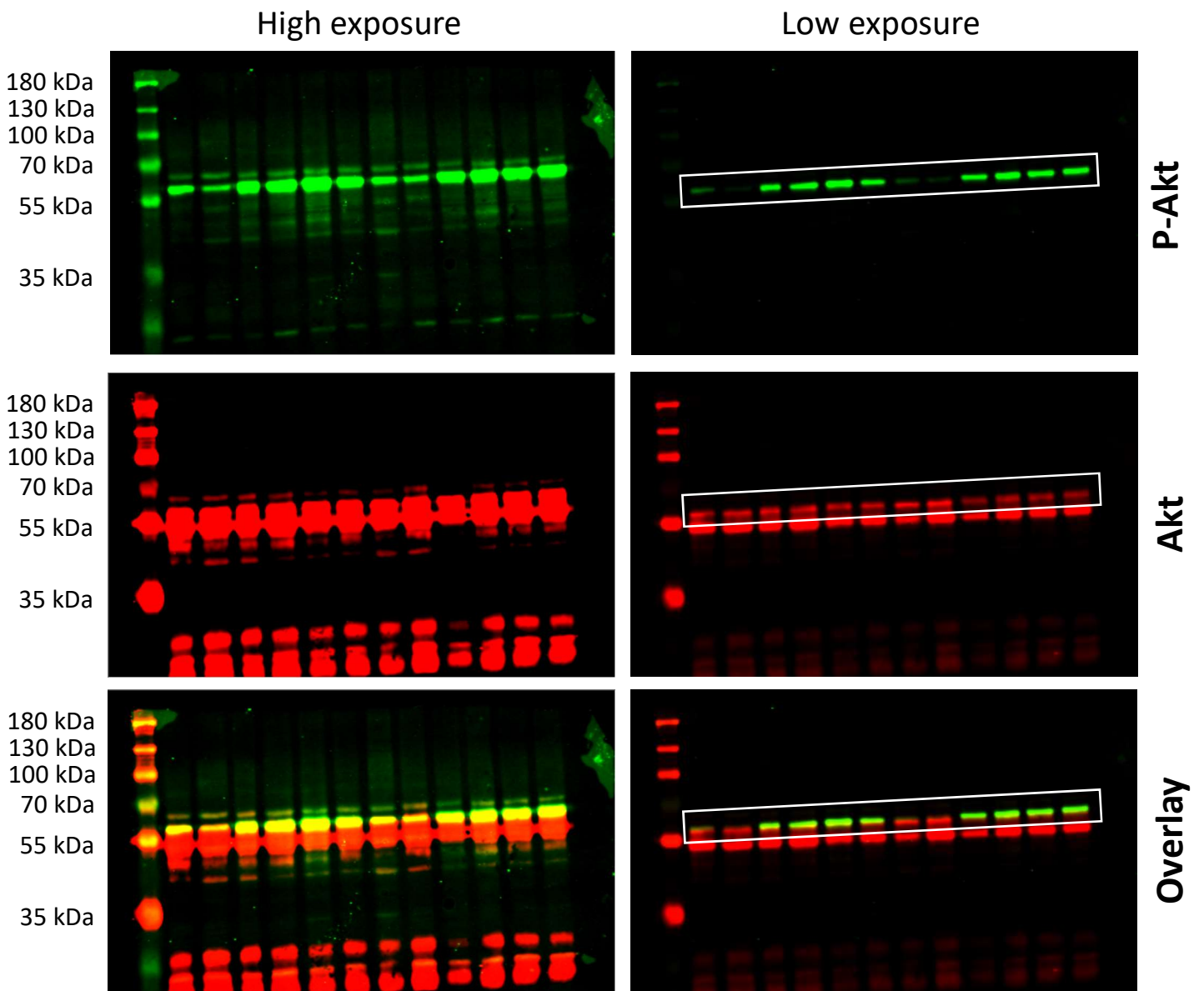


Figure S8: Uncropped WB images for epiAT (related to Fig. 2H)

Akt molecular weight: 60kDa

Rabbit monoclonal anti-phospho-Akt (Ser473) (Clone 193H12) (Cell Signaling, #4058)

Mouse monoclonal anti-Akt (Pan) (Clone 40D4) (Cell Signaling, #2920)

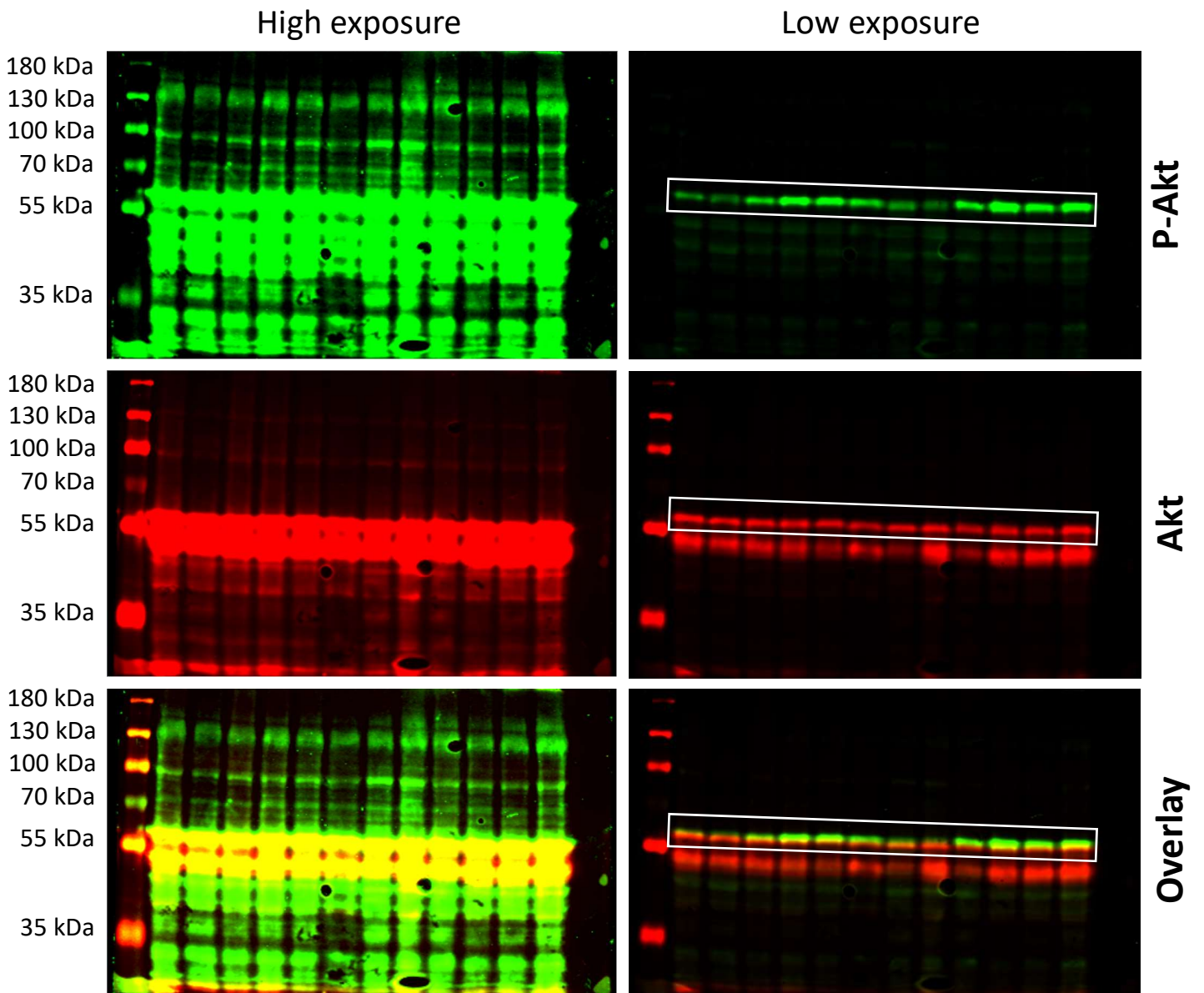


Figure S9: Uncropped WB images for liver (related to Fig. 2H)

Akt molecular weight: 60kDa

Rabbit monoclonal anti-phospho-Akt (Ser473) (Clone 193H12) (Cell Signaling, #4058)

Mouse monoclonal anti-Akt (Pan) (Clone 40D4) (Cell Signaling, #2920)

Supplementary Table 1 : Primers

	Forward	Reverse
Genotyping		
<i>Rora</i> (PCR1)	CATGCATACACCCACACACATCACAACC	CTTGATGCACTGATGTAATAACATATCC
<i>Lyz2</i>	CTTGGGCTGCCAGAATTTCTC	TTACAGTCGGCCAGGCTGAC
<i>Cre</i>	CTTGGGCTGCCAGAATTTCTC	CCCAGAAATGCCAGATTACG
Excision		
<i>Rora</i> (PCR2)	CATGCATACACCCACACACATCACAACC	CAGCCGTGTGAAACTTACGGGAAATACC
qPCR		
<i>Rplp0</i>	GGGAAGGTGTAATCAGTCTCCACAG	CATGCTGAACATCTCCCCCTTCTCC
<i>Ppia</i>	GCATACGGGTCTTGGCATCTTGTC	ATGGTGATCTTCTTGCTGGTCTTGC
<i>Rpl4</i>	CTGAACCCTTACGCCAAGAC	CCTTCTCGGATTTGGTTGCC
<i>Rps29</i>	GATCCGCAAATACGGGCTG	CCATTCAAGGTCGCTTAGTCC
<i>Rora</i> Exon 3	TTCCATGCAAGATCTGTGGAGAC	TGCAGCCTTCACACGTAAT
<i>Rorb</i>	CCCACACCTACGAGGAAATCAA	CATGAAGCCTGTTATCCGCTTG
<i>Rorc</i>	GGTACCATATGCCTCTCTGA	ATCTCCCACATTGACTTCT
<i>Lyz2</i>	CAATGTGCAAAGAGGGTGGTG	TAGAGGGGAAATCGAGGGAATG
<i>Cre</i>	CCTGGAAAATGCTTCTGTCCG	CAGGGTCTTATAAGCAATCCC
<i>Arntl</i>	GGACTTCGCCTCTACCTGTTC	ACCCGATTTCCCCGTTCC
<i>Ob</i>	GGTGTGAAAGAACCTGAGCTGAGG	CAGTGGATGCTAATGTGCCCTG
<i>Tnf</i>	AGCACAGAAAGCATGATCCG	CCCGAAGTTCAGTAGACAGAAGAG
<i>Il1b</i>	GCCACCTTTTGACAGTGATGAG	CCTGAAGCTCTTGTTGATGTGC
<i>Il6</i>	CCAGTTGCCTTCTTGGGACTG	CAGGTCTGTTGGGAGTGGTATCC
<i>Il10</i>	AGGCGCTGTGCATCGATTTCTC	TGGCCTTG TAGACACCTTGGTC
<i>Col1a1</i>	AGGCATAAAGGGTCATCGTGG	GAGACCGTTGAGTCCGTCTTT
<i>Col3a1</i>	TGGCAATGTAAAGAAGTCTC	CCCAGTGTGTTTAGTACAGC
<i>Acta2</i>	GCCGAGATCTCACCGACTAC	CAATCTCACGCTCGGCAGTA
<i>Cd19</i>	ACCTGACCATCGAGAGGCACGTG	CCTGGCGGGGTCAGTCATTTCGCTT
<i>Cd3e</i>	ACTGGAGCAAGAATAGGAAG	GGATGGGCTCATAGTCTG
<i>Adgre1</i>	CTTTGGCTATGGGCTTCCAGTC	GCAAGGAGGACAGAGTTTATCGTG
<i>Clec4f</i>	CCCAGGACCCTGCGGCACGTC	CCGAAGCAGGAGGGAGGGAGGCTC
<i>Ly6g</i>	TGATGGATTTTGC GTTGCTCTG	GTATTGTCCAGAGTAGTGGGGC
<i>Ccr2</i>	ATTCTCCACACCCTGTTTCG	CATGGCCTGGTCTAAGTGCT

Supplementary Table 2 : Antibodies for cell sorting

	<u>Source</u>	<u>Identifier</u>
Spleen		
BV510Anti-CD45 (Clone 30-F11)	BioLegend	#103137
PE-CF594 Anti-CD3e (Clone 145-2C11)	BD Biosciences	#562286
PE-CF594 Anti-TCR β Chain (Clone H57-597)	BD Biosciences	#562841
PE-CF594 Anti-CD19 (Clone 1D3)	BD Biosciences	#562291
BUV395 Anti-CD11b (Clone M1/70)	BD Biosciences	#563553
BV421 Anti-Ly-6G (Clone 1A8)	BioLegend	#127627
PE/Cy7 Anti-F4/80 (Clone BM8)	BioLegend	#123113
FITC Anti-I-A/I-E (MHC class II) (Clone M5/114.15.2)	BioLegend	#107605
PE Anti-CD115 (CSF-1R) (Clone AFS98)	BioLegend	#135505
APC/Cy7 Anti-CD11c (Clone N418)	BioLegend	#117323
Liver		
BV510 Anti-CD45 (Clone 30-F11)	BioLegend	#103137
APC Anti-CD3e (Clone 145-2C11)	BD Biosciences	#553066
APC Anti-TCR β Chain (Clone H57-597)	BD Biosciences	#553174
PE Anti-CD19 (Clone 1D3)	BD Biosciences	#557399
PE Anti-CD20 (Clone AISB12)	eBioscience	#12-0201-82
AF700 Anti-I-A/I-E (MHC class II) (Clone M5/114.15.2)	BioLegend	#107622
AF488 Anti-F4/80 (Clone BM8)	BioLegend	#123120
Anti-CLEC4F/CLECSF13 (Clone 370901)*	Bio-Techne	#MAB2784
Adipose tissue		
BV510 Anti-CD45 (Clone 30-F11)	BioLegend	#103137
APC Anti-CD3e (Clone 145-2C11)	BD Biosciences	#553066
APC Anti-TCR β Chain (Clone H57-597)	BD Biosciences	#553174
PE Anti-CD19 (Clone 1D3)	BD Biosciences	#557399
PE Anti-CD20 (Clone AISB12)	eBioscience	#12-0201-82
AF700 Anti-I-A/I-E (MHC class II) (Clone M5/114.15.2)	BioLegend	#107622
AF488 Anti-F4/80 (Clone BM8)	BioLegend	#123120
BV711 Anti-CD64 (Fc γ RI) (Clone X54-5/7.1)	BioLegend	#139311
Lung		
BV510 Anti-CD45 (Clone 30-F11)	BioLegend	#103137
PE-CF594 Anti-CD3e (Clone 145-2C11)	BD Biosciences	#562286
PE-CF594 Anti-TCR β Chain (Clone H57-597)	BD Biosciences	#562841
PE-CF594 Anti-CD19 (Clone 1D3)	BD Biosciences	#562291
PE/Cy7 Anti-F4/80 (Clone BM8)	BioLegend	#123114
BV711 Anti-CD64 (Fc γ RI) (Clone X54-5/7.1)	BioLegend	#139311
BUV395 Anti-CD11b (Clone M1/70)	BD Biosciences	#563553
AF647 Anti-Siglec-F (Clone E50-2440)	BD Biosciences	#562680

* Anti-CLEC4F antibody was coupled with PE/Cy7 by using PE/Cy7 Conjugation Kit - Lightning-Link (Abcam, #ab102903).