

## Supplementary Materials for

### **YAP-mediated mechanotransduction tunes the macrophage inflammatory response**

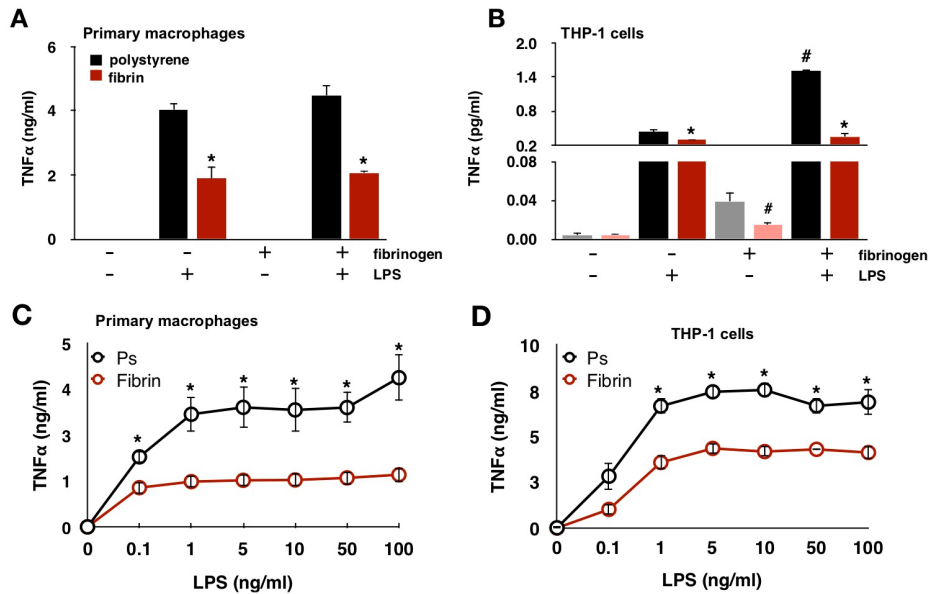
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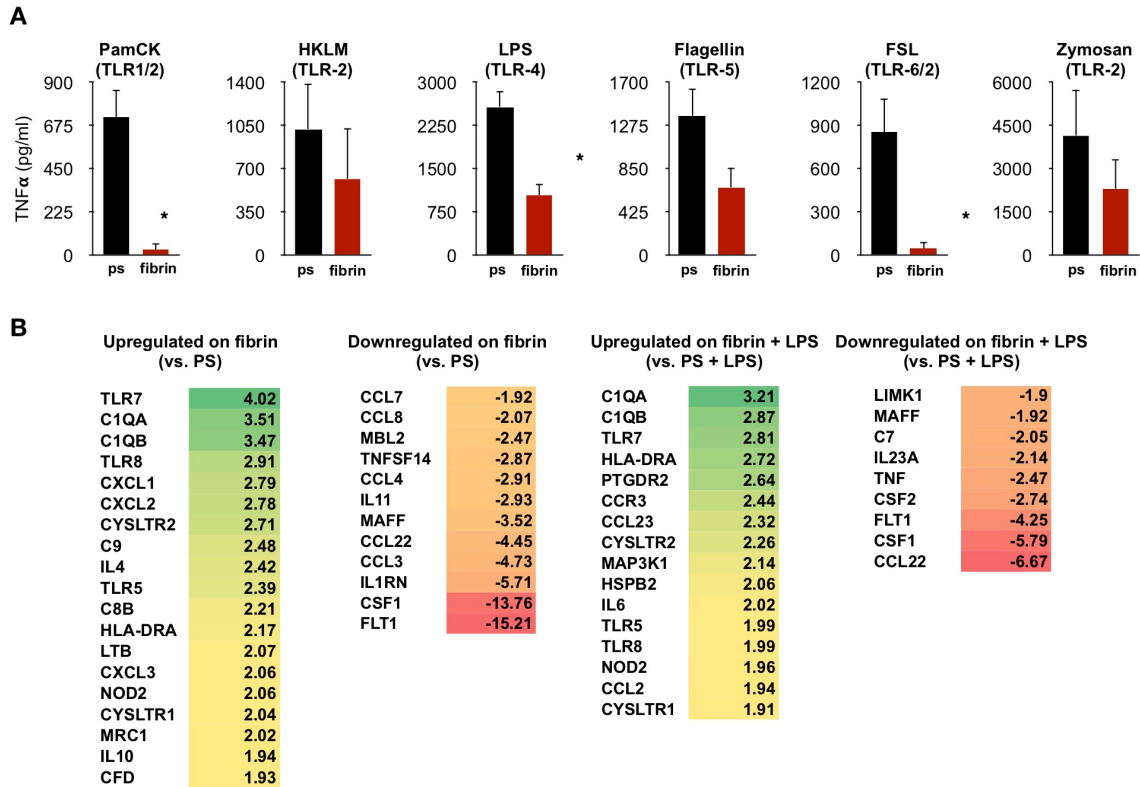
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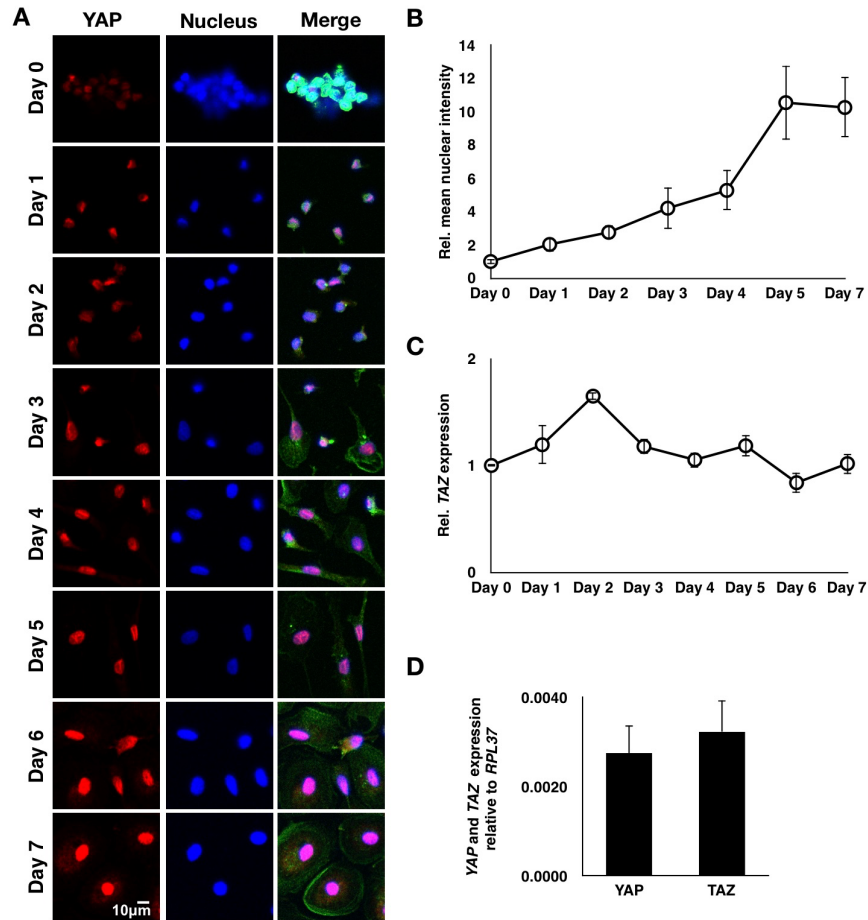
Figs. S1 to S10  
Tables S1 and S2



**Fig. S1. Adhesion to fibrin hydrogels inhibits on inflammatory activation of macrophages.** (A) TNF $\alpha$  secretion by human peripheral blood monocyte-derived macrophages and (B) PMA-treated THP-1 cells (right) cultured on 2 mg/ml fibrin gels or polystyrene, stimulated with 10 ng/ml ultrapure lipopolysaccharide (LPS) and/or 2 mg/ml soluble fibrinogen, or left unstimulated as a control, as measured by ELISA. (C) Secretion of TNF $\alpha$  by monocyte-derived macrophages cultured on 2 mg/ml fibrin gels or polystyrene, stimulated with the indicated concentrations of LPS and analyzed by ELISA (D) Secretion of TNF $\alpha$  by PMA-treated THP-1 cells cultured on 2 mg/ml fibrin gels or polystyrene, stimulated with the indicated concentrations of LPS and analyzed by ELISA. Data presented as mean  $\pm$  SEM of 3 biological replicates and \*  $p < 0.05$  when comparing fibrin vs. polystyrene with the same soluble condition and #  $p < 0.05$ , when comparing fibrinogen vs. no fibrinogen, assessed by two-tailed Student's *t*-test.

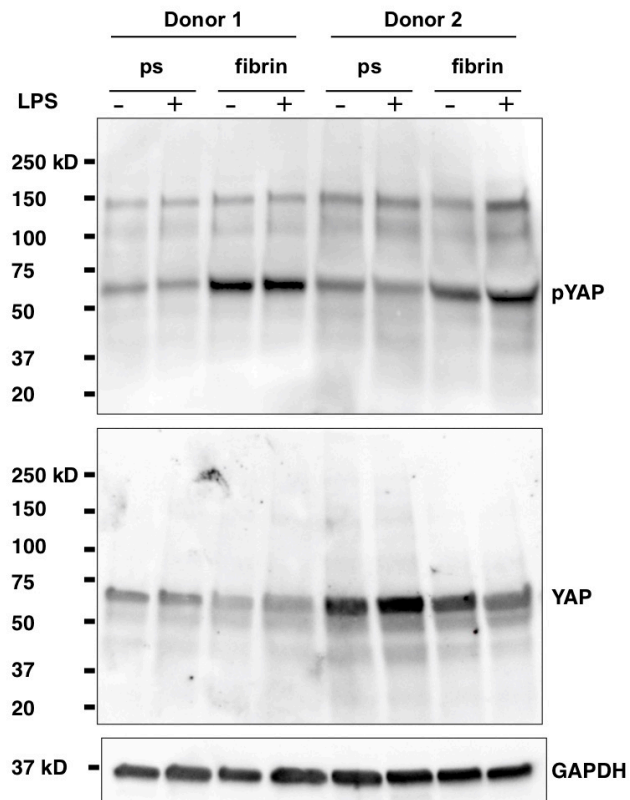


**Fig. S2. Adhesion to fibrin broadly suppresses inflammatory activation of macrophages in response to Toll-like receptor (TLR) ligands.** (A) Secretion of TNF $\alpha$  by monocyte-derived macrophages cultured on 2 mg/ml fibrin gels or polystyrene, stimulated with indicated TLR agonists, analyzed by ELISA. Values are the mean  $\pm$  SEM of 3 biological replicates and \*  $p < 0.05$  comparing fibrin to polystyrene, assessed by two-tailed student's *t*-test. (B) Gene expression of monocyte-derived macrophages cultured on 2 mg/ml fibrin gels or polystyrene, stimulated with LPS for 4h, analyzed using Nanostring inflammation gene expression panel. Values shown are the mean fold change across 2 biological replicates. Transcripts that were greater than 1.9 fold up or downregulated are shown.

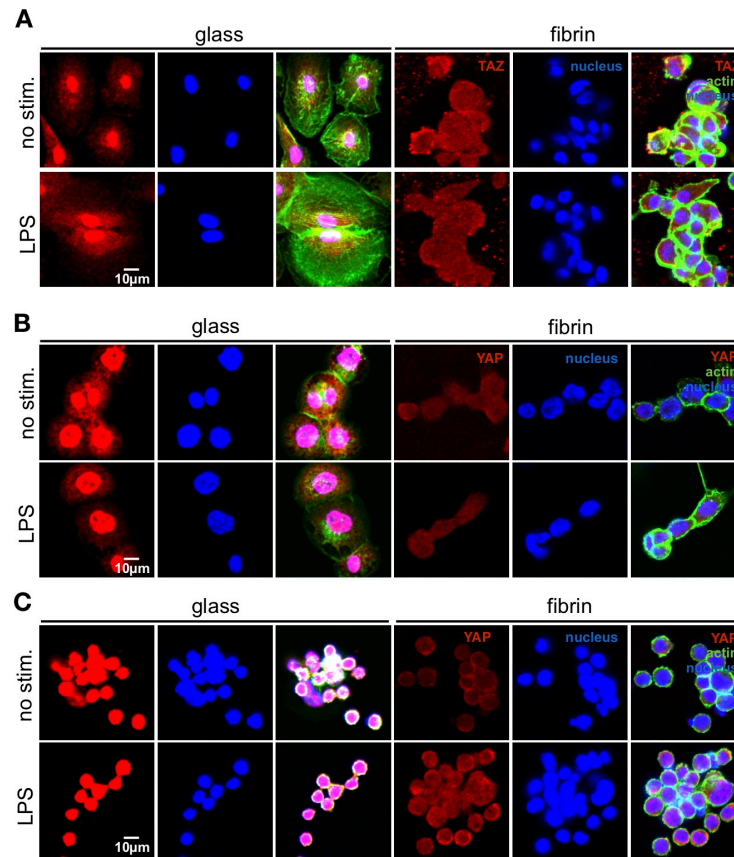


**Fig. S3. YAP and TAZ expression with macrophage differentiation.** (A) Confocal immunofluorescence images of YAP in monocytes cultured on glass with macrophage colony stimulating factor (M-CSF) to induce differentiation and stained for YAP during differentiation for seven days. (B) Graph shows average mean nuclear YAP intensity for two biological replicates normalized to Day 0. (C) Relative gene expression of *TAZ* in monocytes undergoing differentiation analyzed by qPCR. Graph shows mean expression for two biological replicates normalized to Day 0. (D) Gene expression of *YAP* and *TAZ* analyzed by qPCR in monocyte-derived macrophages differentiated on polystyrene for 7 days from three donors. Expression was normalized to *RPL37A* housekeeping gene. Values are the mean  $\pm$  SEM of 3 biological replicates.

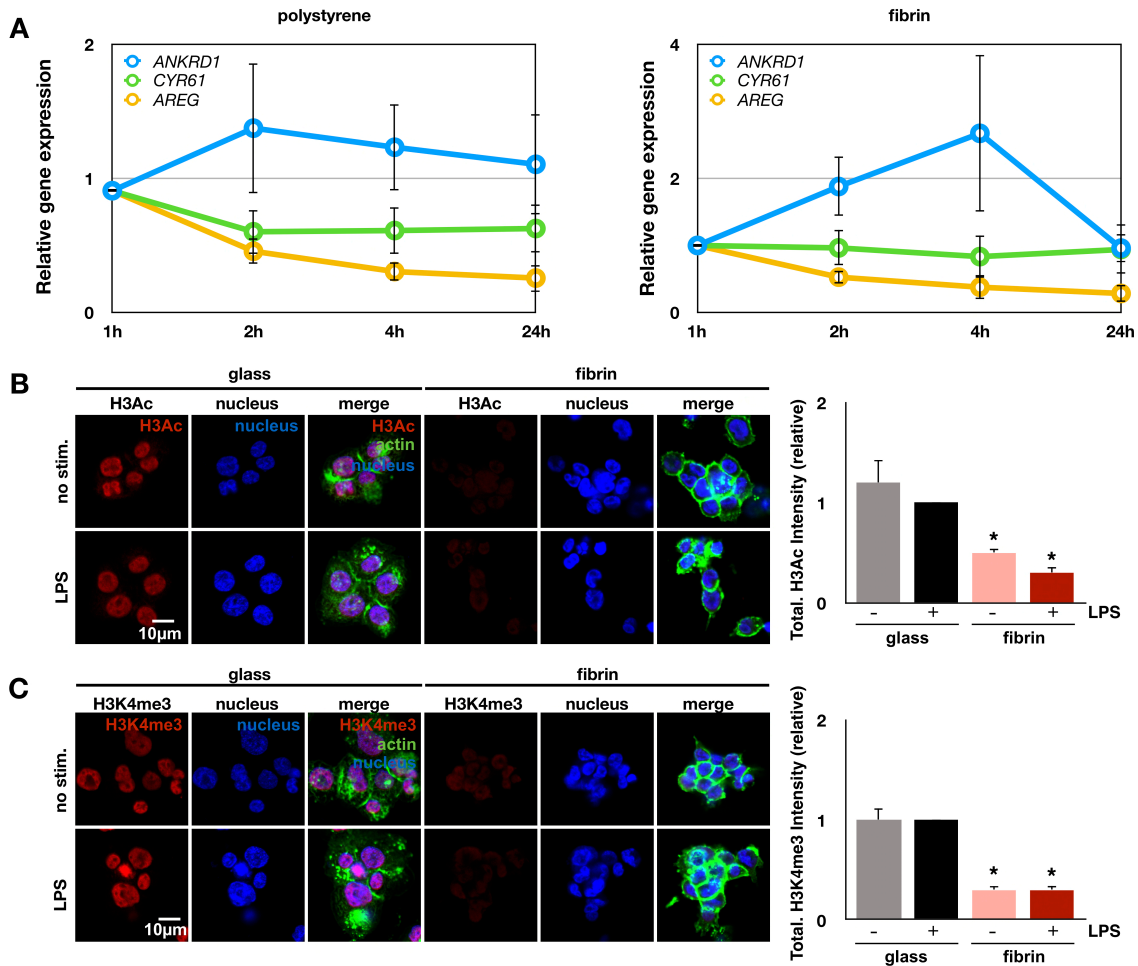




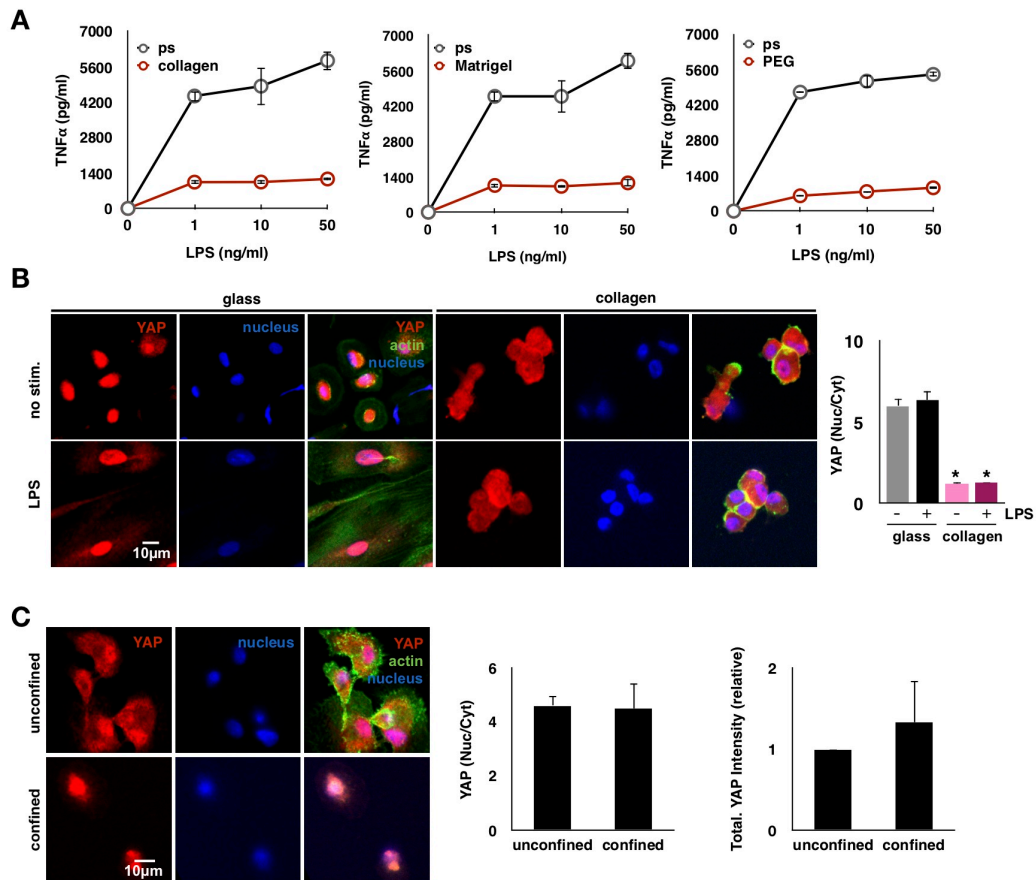
**Fig. S4. Total and phosphorylated YAP is regulated by substrate.** Representative complete Western blots of phosphorylated YAP (at serine 127, pYAP), total YAP, and GAPDH in monocyte-derived macrophages cultured on polystyrene or 2 mg/ml fibrin hydrogels for 24 h, and then stimulated with 10 ng/ml of LPS for 1 h. Two blood donors are shown.



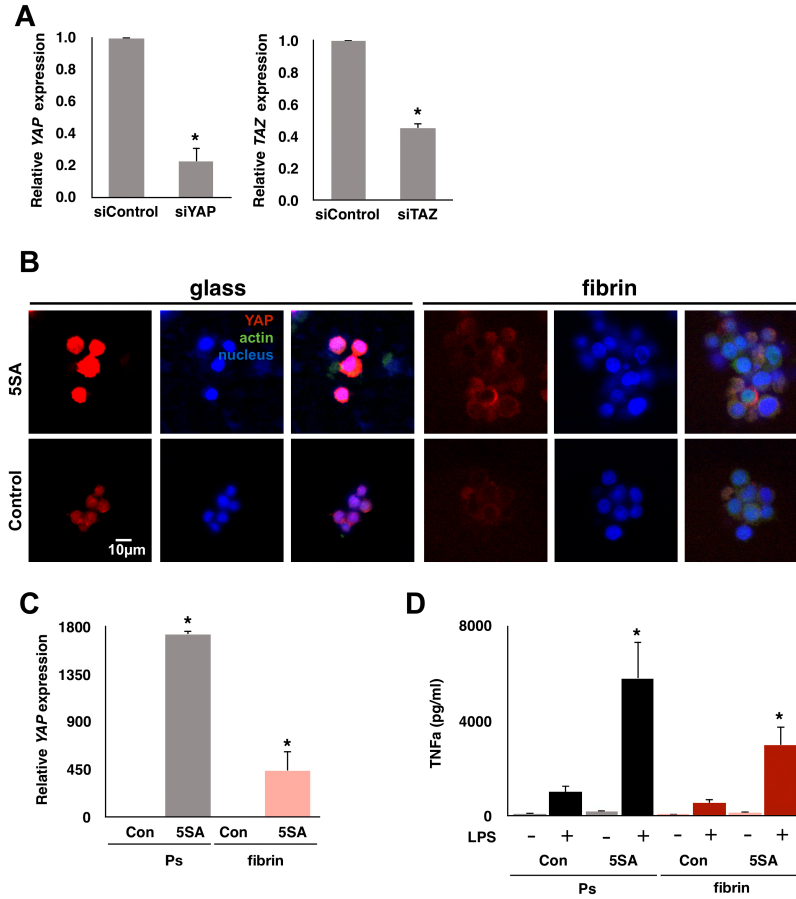
**Fig. S5. TAZ localization in PBMC-derived macrophages and YAP localization in THP-1 and U937 human monocyte/macrophage cell lines.** (A) Confocal immunofluorescence images of TAZ in monocyte-derived macrophages cultured on glass or 2 mg/ml fibrin for 24 h and stimulated with LPS for 1 h. Images are representative from 2 biological replicates. (B) Confocal immunofluorescence images of YAP in THP-1 cells cultured with phorbol myristate acetate (PMA) on 2 mg/ml fibrin gels or polystyrene for 24 h and then stimulated with LPS for 1 h (C) Confocal immunofluorescence images of YAP in U937 cells cultured with PMA on 2 mg/ml fibrin gels or polystyrene for 24 h and then stimulated with LPS for 1 h. Images are representative from 2 biological replicates.



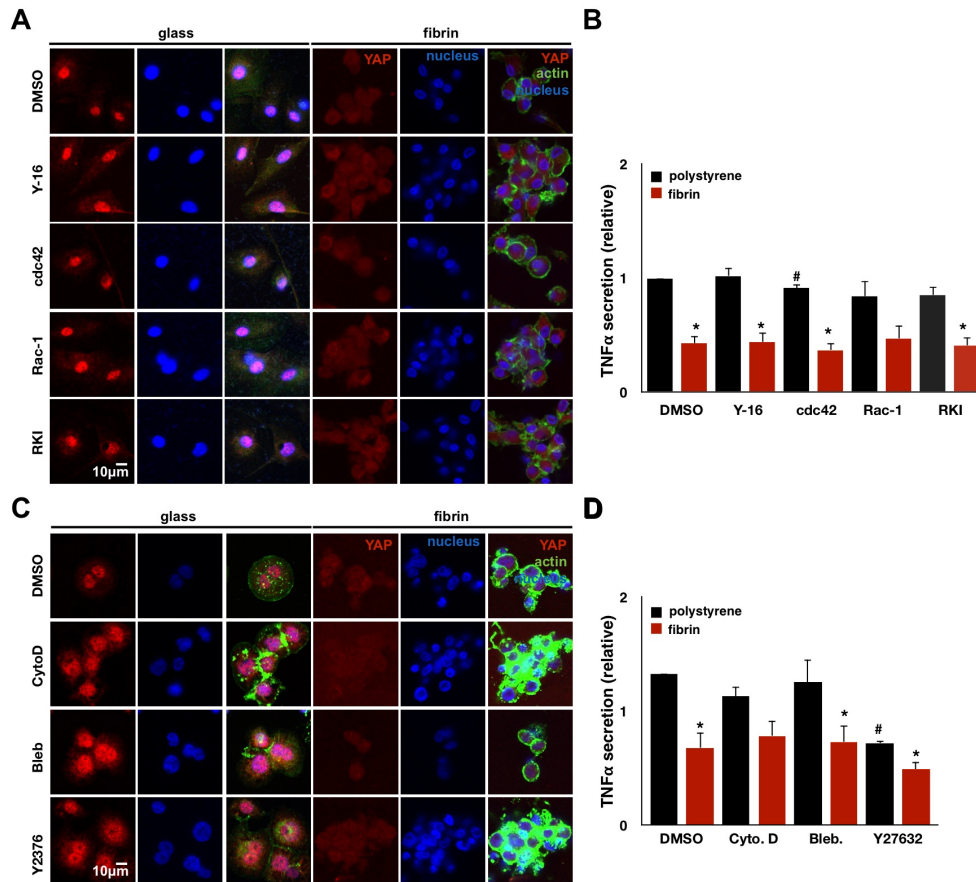
**Fig. S6. Canonical YAP target gene expression and epigenetic modifications in macrophages cultured on polystyrene or glass and fibrin.** (A) Expression of canonical YAP target genes including *ANKRD1*, *CYR61* and *AREG* from monocyte-derived macrophages cultured on polystyrene or 2 mg/ml fibrin for 1, 2, 4 and 24 h. Values shown are expression normalized to *RPL37A* and then normalized to transcript level of the respective gene at 1 h. (B) Immunofluorescence confocal images of H3Ac in THP-1 cells treated with PMA and cultured on glass or 2 mg/ml fibrin for 42 h and stimulated with 10 ng/ml LPS for 6 h, and quantification of total intensity (right). (C) Immunofluorescence confocal images of H3K4me3 in THP-1 cells treated with PMA and cultured on glass or 2 mg/ml fibrin for 42 h and stimulated with 10 ng/ml LPS for 6 h, and quantification of total intensity (right). All images are representative across at least 3 biological replicates. For intensity, analysis was performed across 3 biological replicates and on at least 150 cells (for imaging). All the values are the mean  $\pm$  SEM and \*  $p < 0.05$  when comparing fibrin to glass assessed by two-tailed Student's *t*-test in (B) and (C).



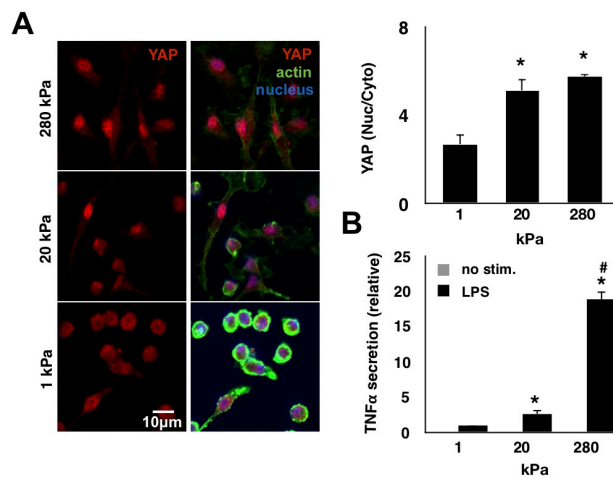
**Fig. S7. Inflammatory response and YAP nuclear localization is regulated by hydrogel environments, but not micropatterning-induced adhesive confinement.** (A) Secretion of TNF $\alpha$  by monocyte-derived macrophages cultured on 2 mg/ml of collagen, 3mg/ml Matrigel, or 5% polyethylene glycol (PEG) gels and stimulated with the indicated concentrations of LPS and analyzed by ELISA. Values are the mean  $\pm$  SEM of 3 biological replicates for collagen and Matrigel and 2 replicates for PEG gels and \*  $p < 0.05$  compared to cells seeded on polystyrene (ps) controls, assessed by two-tailed student's *t*-test. (B) Immunofluorescence confocal images of YAP (left) in monocyte-derived macrophages cultured on glass or 2 mg/ml collagen for 24 h and stimulated with 10 ng/ml LPS for 1 h, and quantification of nuclear/cytoplasmic ratio (right). Values are the mean  $\pm$  SEM of two biological replicates and \*  $p < 0.05$  compared to glass control, assessed by two-tailed Student's *t*-test. (C) Confocal images of YAP in monocyte-derived macrophages, cultured on flat (unconfined) and 10 x 10  $\mu$ m (confined) micropatterned islands for 24 h and stained for YAP (left), and quantification of nuclear to cytoplasmic ratio and total intensity of YAP (right). Values are the mean  $\pm$  SEM across 3 biological replicates.



**Fig. S8. Manipulation of YAP expression and activity.** (A) Expression of *YAP* or *TAZ* gene analyzed by qPCR in monocyte-derived macrophages treated with (siYAP) or TAZ siRNA (siTAZ), respectively, relative to siControl. (B) Confocal images of YAP in YAP-5SA-expressing U937 cells cultured with PMA on glass or fibrin for 24 h. (C) YAP gene expression in conditions shown in (B) measured by qRT-PCR. Blank lentiviral vector was used as control. (D) Secretion of TNF $\alpha$  from cells in (B) stimulated with 10 ng/ml LPS, analyzed by ELISA. Values are mean  $\pm$  SEM of 3 biological replicates and \* $p$ <0.05 when comparing YAP-5SA vs. control, assessed by two-tailed Student's *t*-test.



**Fig. S9. Effect of Rho GTPase inhibitors and cytoskeletal drugs on YAP localization and inflammation.** (A) Immunofluorescence confocal images of YAP in monocyte-derived macrophages, cultured on glass or 2 mg/ml fibrin hydrogel for 24 h and treated with indicated inhibitors of RhoA, Rac1, or Cdc42 and related signaling proteins for approximately 6 h. (B) Secretion of TNF $\alpha$  from cells in (A) except glass conditions were replaced with polystyrene, and stimulated with LPS for 6 h. (C) Immunofluorescence confocal images of YAP in PMA-treated THP-1 cells, cultured on glass or 2 mg/ml fibrin hydrogel for 24 h and treated with indicated cytoskeletal drugs for approximately 6 h (D) Secretion of TNF $\alpha$  secreted from cells in (C) except glass conditions were replaced with polystyrene, and stimulated with LPS for 6 h. Values are the mean  $\pm$  SEM across 3 biological replicates and \* $p$ <0.05 comparing fibrin vs. glass or polystyrene and # $p$ <0.05 comparing drug vs. DMSO, assessed by two-tailed Student's  $t$ -test.



**Fig. S10. Substrate stiffness modulates YAP nuclear localization and inflammation. (A)** Immunofluorescence confocal images of YAP in mouse bone marrow derived - macrophages cultured on polyacrylamide (PA) gels of varying stiffness for 24 h and stimulated with LPS for 6 h (left) and quantification of nuclear to cytoplasmic ratio and total intensity (right). **(B)** Secretion of TNF $\alpha$  from cells in (A) and stimulated with 10 ng/ml of LPS for 6 h, analyzed by ELISA. Cytokines are quantified across 3 biological replicates. Immunostaining analysis was performed on at least 150 cells across 3 biological replicates at each time points. Values are the mean  $\pm$  SEM and \*  $p < 0.05$  when comparing 1 to 20 or 280 kPa and #  $p < 0.05$  when comparing 1 and 20 to 280 kPa assessed by two-tailed Student's *t*-test.

**Supplementary Table 1:** List of primers used in this study for qRT-PCR analysis.

<b>Gene Name</b>	<b>Forward primer (5'-3')</b>	<b>Reverse primer (5'-3')</b>
<i>TNF</i>	AGGCGCTCCCAAGAAGACAGG	CAGCAGGCAGAAGAGCGTGGTG
<i>IL6</i>	AAGCCAGAGCTGTGCAGATGAGTA	CTTGGTCACCGACGTCTCTGT
<i>MCP1</i>	CCCAGTCCCTGCTGTTAT	TGGAATCCTGAACCCACTTC
<i>IL10</i>	TCATTCCCAACCACTTCAT	GTAGAGACGGGGTTTCACCA
<i>YAP</i>	GCAGTTGGGAGCTGTTTCTC	GCCATGTTGTTGTCTGATCG
<i>CYR61</i>	TCACCCTTCTCCACTTGACC	AGTTTTGCTGCAGTCCTCGT
<i>AREG</i>	CTGGGAAGCGTGAACCATTTT	TCTGAGTAGTCATAGTCGGCTC
<i>ANKRD1</i>	GAAGTGGTCACTGGAAAGAAGAATG	GGTGGGCTAGAAGTGTCTTCAGA
<i>RPL37A</i>	ATTGAAATCAGCCAGCACGC	AGGAACCACAGTGCCAGATCC

**Supplementary Table 2:** List of inhibitor drugs and their working concentrations used this study.

<b>Inhibitor Drug</b>	<b>Working Concentration</b>	<b>Function</b>
Cytochalasin D	10 $\mu$ M	Potent inhibitor of actin polymerization
Blebbistatin	30 $\mu$ M	Myosin inhibitor, specific for myosin II
Y-27632	30 $\mu$ M	ROCK inhibitor
Rho II Y16	15 $\mu$ M	Rho inhibitor II, targets RhoGEFs
Rac1	100 $\mu$ M	Rac1 inhibitor, inhibits Rac1 GDP/GTP exchange activity
Cdc42 III	50 $\mu$ M	Cdc42 inhibitor III, targets Cdc42 GEF