Supplementary Information for Kim et al.: Improving the informativeness of Mendelian disease-derived pathogenicity scores for common disease

Supplementary Tables

Supplementary Table 1. List of 41 independent diseases and complex traits analyzed. Analogous to a previous study^{[1](#page-15-0)}, we considered 89 GWAS summary association statistics, including 34 traits from publicly available sources and 55 traits from the UK Biobank (up to $N = 459K$); summary association statistics were computed using BOLT-LMM v2.3^{[2,](#page-15-1)[3](#page-15-2)}. We obtained 41 independent traits (average $N = 320K$) with genetic correlation less than 0.9 (computed using cross-trait LDSC^{[4](#page-15-3)}). For 6 traits, we analyzed two different sources (both publicly available and UK Biobank), resulting in total 47 summary statistics analyzed. For each trait, we report a trait identifier, trait description, reference, sample size, and heritability z-score.

Supplementary Table 2. Expected heritability enrichments of binary annotations derived from boosted Mendelian disease-derived missense scores with significantly negative τ^* . We report the heritability enrichment that is expected based on the boosted annotation's overlap with the baseline-LD model and corresponding published annotations, by assuming that the τ of the annotation is zero. In general, expected enrichments were significantly less than observed enrichments.

Supplementary Figures

Assess informativeness for common disease

Supplementary Figure 1. Overview of AnnotBoost framework. We describe the AnnotBoost model training. AnnotBoost requires only one input, a pathogenicity score to boost, and generates a genome-wide (probabilistic) boosted pathogenicity score. From the input pathogenicity score (e.g. CADD as shown here), we built a classification model, each for even and odd chromosome SNPs using 75 baseline-LD annotations [18](#page-17-0)[,19](#page-17-1) as features. We assessed informativeness of annotations derived from published scores (input) and boosted scores (output) for common disease using S-LDSC^{[20](#page-17-2)}.

Supplementary Figure 2. Feature importance of boosted Mendelian disease-derived missense pathogenicity scores. We applied SHAP^{[21](#page-17-3)} to assess which features from the baseline-LD drives the prediction of 11 boosted missense scores by AnnotBoost. We report the signed impact of top 20 features for each of 11 predictive models: (A) PolyPhen-2, (B) PolyPhen-2-HVAR, (C) MetaLR, (D) MetaSVM, (E) PROVEAN, (F) SIFT 4G, (G) REVEL, (H) M-CAP, (I) Primate-AI, (J) MPC, and (K) MVP. We obtained similar results for even/odd chromosome classifiers; we report odd chromosome results here (see full results online; see URLs).

Supplementary Figure 3. Excess overlap between gene scores derived from input pathogenicity scores and 165 reference gene sets of biological importance. We report the excess overlap of genes linked to published and boosted scores in existing gene sets of biological importance (summarized in Supplementary Data 8): (A) PolyPhen- $2^{22,23}$ $2^{22,23}$ $2^{22,23}$ $2^{22,23}$ gene quintiles from published and boosted scores, (B) $\text{CADD}^{24,25}$ $\text{CADD}^{24,25}$ $\text{CADD}^{24,25}$ $\text{CADD}^{24,25}$ gene quintiles from published and boosted scores, and (C) CCR^{26} CCR^{26} CCR^{26} gene quintiles from published and boosted scores. Error bars represent 95% confidence intervals. Numeric results for excess overlap and correlaton among gene scores are shown in Supplementary Data 9. Numeric results for odds ratios and p-values from Fisher's exact test (two-sided) between published gene quintiles and boosted gene quintiles are reported in Supplementary Data 10.

Supplementary Figure 4. Feature importance of boosted genome-wide Mendelian diseasederived pathogenicity scores. We applied $S HAP²¹$ $S HAP²¹$ $S HAP²¹$ to assess which features from the baseline-LD drives the prediction of 11 boosted missense scores by AnnotBoost. We report the signed impact of top 20 features for each of 6 genome-wide Mendelian disease-derived pathogenicity scores: (A) CADD, (B) Eigen, (C) Eigen-PC, (D) ReMM, (E) NCBoost, and (F) ncER. We obtained similar results for even/odd chromosome classifiers; we report odd chromosome results here (see full results online; see URLs).

Supplementary Figure 5. Feature importance of boosted scores derived from 18 additional genome-wide scores and 47 baseline-LD model annotations. We applied $SHAP^{21}$ $SHAP^{21}$ $SHAP^{21}$ to assess which features from the baseline-LD drives the prediction of 18 boosted additional scores by AnnotBoost. We report the signed impact of top 20 features for each of 18 additional scores: (A) CDTS, (B) CCR, (C-I) DeepSEA-CTCF, -DNase, -H3K27ac, -H3K4me1, -H3K4me2, -H3K4me3, -H3K9ac, (J-K) DIS-DNA, -RNA, (L) pLI, (M) LIMBR, (N-Q) Gene network connectivity-Saha, Greene, InWeb, Sonawane, (R) EDS. We obtained similar results for even/odd chromosome classifiers; we report odd chromosome results here (see full results online; see SHAP results of 47 boosted baseline-LD scores online; see URLs).

Supplementary Figure 6. Classification of fine-mapped disease SNPs using aggregated scores. We report the true positive rate, false positive rate, precision, and recall along with the classification accuracy (AUPRCs and AUROCs) of four aggregated scores on classifying 5 different independent SNP sets: (A, F) 7,333 fine-mapped for 21 autoimmune diseases from Farh et al. [27](#page-18-0), (B, G) 3,768 fine-mapped SNPs for inflammatory bowel disease from Huang et al.^{[28](#page-18-1)}, (C, H) 1,851 fine-mapped SNPs for 49 traits from UK Biobank [29](#page-18-2), (D, I) 1,379 fine-mapped SNPs without functional data for 49 traits from UK Biobank [29](#page-18-2), and (E, J) 14,807 GWAS significant SNPs [30,](#page-18-3)[31](#page-18-4), from 10 LD-, MAF-, and genomic element-matched control SNPs. We report the average AUPRCs and AUROCs of even/odd-chromosome classifiers. Differences for AUROCs and AUPRCs attained between (1) baseline-LD and baseline-LD+joint model, (2) baseline-LD and baseline-LD+marginal model, and (3) baseline-LD+joint and baseline-LD+marginal were largely significant (p-val < 0.008). Numerical results, including results using the most matched control SNPs (instead of 10), are reported in Supplementary Data 21.

Supplementary Figure 7. Feature importance of baseline-LD+joint model in predicting fine**mapped or GWAS significant SNPs.** We applied $SHAP²¹$ $SHAP²¹$ $SHAP²¹$ to assess which features from the baseline-LD+joint drive the prediction of fine-mapped or GWAS significant SNPs from 10 matched control SNPs for each positive SNP. We report the signed impact of top 20 feature for each of 4 fine-mapped SNPs and GWAS significant SNPs: (A) Farh et al., (B) Huang et al., (C) Weissbrod et al., (D) Weissbrod et al. (fine-mapped without functional data), (E) GWAS significant SNPs. We obtained similar results for even/odd chromosome classifiers; we report odd chromosome results here (see full results online; see URLs).

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Supplementary Figure 8. Feature importance of baseline-LD+marginal model in predicting **fine-mapped or GWAS significant SNPs.** We applied $SHAP²¹$ $SHAP²¹$ $SHAP²¹$ to assess which features from the baseline-LD+marginal drive the prediction of fine-mapped or GWAS significant SNPs from 10 matched control SNPs for each positive SNP. We report the signed impact of top 20 feature for each of 4 fine-mapped SNPs and GWAS significant SNPs: (A) Farh et al., (B) Huang et al., (C) Weissbrod et al., (D) Weissbrod et al. (fine-mapped without functional data), (E) GWAS significant SNPs. We obtained similar results for even/odd chromosome classifiers; we report odd chromosome results here (see full results online; see URLs).

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Supplementary Figure 9. Classification of fine-mapped disease SNPs in single-score analysis using published and boosted scores. We report the classification accuracy (AUROC and AUPRC) of each of the 82 boosted scores compared to the corresponding published score. We report AUROC (resp. AUPRC) on (A, F) 7,333 fine-mapped for 21 autoimmune diseases from Farh et al.^{[27](#page-18-0)}, (B, G) 3,768 finemapped SNPs for inflammatory bowel disease from Huang et al. [28](#page-18-1), (C, H) 1,851 fine-mapped SNPs for 49 traits from UK Biobank [29](#page-18-2), (D, I) 1,379 fine-mapped SNPs without functional data for 49 traits from UK Biobank^{[29](#page-18-2)}, and (E, J) 14,807 GWAS significant SNPs^{[30,](#page-18-3)[31](#page-18-4)} from 10 LD-, MAF-, and genomic elementmatched control SNPs. Numerical results, including results using the most matched control SNPs (instead of 10), are reported in Supplementary Data 23.

Supplementary Figure 10. Informativeness of the baseline-LD model before and after adding 11 jointly significant binary annotations. We report meta-analyzed τ^* of the baseline-LD model annotations, across 41 independent traits, from two different S-LDSC analyses: (1) the baseline-LD model + 8 Roadmap annotations and (2) the baseline-LD model + 8 Roadmap annotations + 11 jointly significant annotations. Error bars represent 95% confidence intervals. Numerical results are reported in Supplementary Data 25.

Supplementary Figure 11. Informativeness for common disease of binary annotations derived from boosted CDTS scores with or without MAF features. We applied AnnotBoost to CDTS annotation using all baseline-Ld features and all features excluding MAF bins. Then, we applied S-LDSC, conditioning on published binary CDTS annotations (five thresholds from 90th percentile to 99.9th percentile) and baseline-LD model annotations; and meta-analyzed results across 41 independent traits. We report meta-analyzed enrichments and τ^* . Error bars represent 95% confidence intervals.

Supplementary Figure 12. Informativeness for common disease of binary annotations derived from boosted CDTS scores using imbalanced data. We applied AnnotBoost to CDTS annotation of varying training data. Then, we applied S-LDSC, conditioning on published binary CDTS annotations (five thresholds from 90th percentile to 99.9th percentile) and baseline-LD model annotations; and meta-analyzed results across 41 independent traits. We report meta-analyzed enrichments and τ^* . Error bars represent 95% confidence intervals.

Supplementary Figure 13. Informativeness for common disease of binary and probabilistic annotations derived from published CDTS scores. We constructed binary and probabilistic annotations for published CDTS [32](#page-18-5). We applied S-LDSC, conditional on baseline-LD model annotations and meta-analyzed results across 41 independent traits. We report meta-analyzed enrichments and τ^* . To construct probabilistic annotations of varying proportion of SNPs, we performed the following transformation to upweight the upper percentile and downweight the lower percentile SNPs: $e^{\alpha * annot}$ with α varied from 3 to 2000. Error bars represent 95% confidence intervals.

Supplementary References

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