Supplementary Information

Stem Cell Characteristics Promote Aggressiveness of Diffuse Large B-Cell Lymphoma

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Supplementary Figure S1. The purified tumor cells are well recognized and account for more than 90% of all cells.





Supplementary Figure S3. The sequence of Vector Myc-DDK



Cloning sites used for ORF Shuttling:

* The last codon before the Stop codon of the ORF

Supplementary Figure S4. Vector pLKO.1-TRC1

(http://rnai.genmed.sinica.edu.tw/searchDatabase)



Description:

pLKO.1-Puro is a cloning vector of shRNA, originally derived from HIV-1. It contains all the necessary *cis*-elements for packaging, reverse transcription, and integration for subsequent production of the lentiviral particles.

Cloning site:

Agel and EcoRI (only these two sites available)

Supplementary Figure S5. Vector pLKO.1-TRC2 (http://rnai.genmed.sinica.edu.tw/searchDatabase)



Description:

pLKO_TRC005 (TRC2 vector) was derived from pLKO_TRC001 with the following modifications:

- (i) A *Kpn*I restriction enzyme site was created next to *Age*I, thereby two combination sites for shRNA cloning can be used, *Kpn*I-*Eco*RI or *Age*I-*Eco*RI.
- (ii) A post-transcription regulation element from woodchuck hepatitis virus (WPRE) was inserted behind PAC gene (puromycin acetyltransferase gene).

Location of Features (for other features, please refer to pLKO.1-puro):

- Human U6 promoter: nt7004-7255
- Stuffer : nt7256-9124
- Human PGK promoter: nt4-508
- PAC (Puro; Puromycin acetyltransferase): nt529-1128
- WPRE (woodchuck post-transcription regulation element): nt1144-1732

Note:

DNA sequences within lentiviral genome including both 5'LTR and 3'LTR (not included the whole stuffer sequence) had been verified by TRC.



Supplementary Figure S6. IPA analysis of HOXA9 and NANOG HOXA9 & NANOG-2

The relationship between NANOG and HOXA9 is linked to the several properties of stem cells such as pluripotency, proliferation, self-renewal, and differentiation.

Supplementary Figure S7. IPA analysis reveals the cross-link pathways between



the 14 genes listed in Table 1.

The top 5 pathways are cell-to-cell signaling and interaction, cellular movement (migration), cellular function and maintenance, cellular development, and cell growth and proliferation.

Supplementary Figure S8. RNA *in situ* hybridization (upper panel) and immunofluorescence (other panels) confirm the mRNA expression of NANOG and HOXA9 in clinical cases (upper panel) and protein expression in variable cell lines (other panels) of DLBCL.



Upper panel (left, CD20 stain [inset, HE]; middle, NANOG-RNA; right, HOXA9-RNA) shows RNAscope detection of NANOG (middle) and HOXA9 (right) mRNA transcripts in ~20% of tumor cells in BLS-type DLBCL. Immunofluorescence demonstrates the cytoplasmic localization of NANOG in DLBCL cell lines (HT and U2932), and nuclear localization of HOXA9 in DLBCL cell lines (SU-DHL-5 and HT).



Supplementary Figure S9. Survival curves of patients with DLBCL.

The poorer prognostic factors for overall survival are as follows. (A) presence of B symptoms; (B) high IPI score (3-5); (C) NANOG expression; (D) HOXA9 expression; (E) CCR6 expression; and (F) BLS type DLBCL. The survival time was measured in months.

Supplementary Figure S10. Transfection of HOXA9 decreases cell apoptosis and decreases G2/M phase cell cycle arrest in SU-DHL-5 cells.



(A) SU-DHL-5 cells were transfected with HOXA9 and cultured for 48 hrs, which was evaluated by Western blotting with significantly higher expression. (B) SU cells were stained with annexin V (Q2+Q4) and analyzed by flow cytometry. Representative histograms depict decrease of cell apoptosis with HOXA9 transfection (MYC, 56.2% vs HOXA9, 44.8%). (C) Representative flow histograms depict decreased G2/M cell cycle arrest after HOXA9 transfection (MYC, 10.2% vs HOXA9, 7.1%). Quantitation of G1/S and G2/M fractions in SU cells after 48 hrs culture post HOXA9 transfection. The experiment was repeated in triplicate and merged data from all the experiments are shown. * p<0.05, ** p<0.01, *** p<0.001, Student paired t-test.

Supplementary Figure S11. Expression of HOXA9 shows a trend for more sphere formation.

	Мус		HO	XA9#10		HOXA9#11					
	SUST (um)	<50	~>100	<100~>	200	>200					
	Мус	2	20	31		2					
	HOXA9#10	1	19	43		5					
	HOXA9#11	4	46	50		0					
SU	HOXA9	Mean/HOXA9	Actin	Mean/Actin	∆Cp	1/(2^∆Cp)	fold change				
Мус	30	29.575	14.84	14.84	14.735	3.6671E-05	1				
	29.15		15.33								
HOXA9#10	23.38	23.26	16.19	16.19	7.07	0.007442484	202.952739				
	23.14		16.49								
HOXA9#11	16.13	15.68	15.36	15.36	0.32	0.801069878	21844.76553				
	15.23		15.64	Charles Contractor							

SU cells - Sphere formation (7ds)

The tumor sphere formation assay for the stemness function shows that in comparison with Myc control, SU lymphoma cells with stable expression of HOXA9 have a trend for more sphere formation (20 [Myc] vs. 19 [HOXA9 #10] and 46 [HOXA9 #11], and 31 [Myc] vs. 43 [HOXA9 #10] and 50 [HOXA9 #11], p=0.098, *t*-test).

Supplementary Figure S12. A recent cohort also shows that DLBCL cases with HOXA9 expression carry a poorer prognosis.



(A) This BLS type DLBCL shows nuclear expression of HOXA9 (original magnifications X400).(B) The new cohort of DLBCL cases further validate that overexpression of HOXA9 is a poorer prognostic factor (HOXA9-, n=72; HOXA9+, n=7, p=0.028).

Cell line	Source	Subtype or immunophenotyping
HT	ATCC CRL-2260	Germinal center B cell (GCB)
SU-DHL-5	ATCC CRL-2958	Germinal center B cell (GCB)
HBL2	Gift from Dr. Chen YP	Germinal center B cell (GCB)
U2932	DSMZ ACC 633	Activated B cell (ABC)
U2940	DSMZ ACC 634	Primary mediastinal large B-cell lymphoma

Supplementary Table S1. Characterization of DLBCL cell lines

All are negative for Epstein-Barr virus (EBV). The authentication of cell lines was performed by short-tandem repeat profiling, and *Mycoplasma* testing was done by conventional PCR methods, in Jan. 2019. Eight to ten passages of cell lines between collections were used in the experiments.

Gene Name	Systematic Name	Description	Fold Change	p value
S100A8	NM_002964	S100 calcium binding protein A8 (S8), mRNA (alias MRP8)	6.367	5.2*10 ⁻⁸
S100A12	NM_005621	S100 calcium binding protein A12 (S12), mRNA	6.169	6.4*10 ⁻⁸
CD86	NM_006889	CD86 molecule (CD86), transcript variant 2, mRNA	5.640	1.2*10 ⁻⁷
KRBA2	NM_213597	KRAB-A domain containing 2 (KRBA2), mRNA	5.623	1.2*10 ⁻⁷
BMP8B	NM_001720	bone morphogenetic protein 8b (BMP8B), mRNA	5.613	1.2*10 ⁻⁷
VHL	NM_000551	von Hippel-Lindau tumor suppressor (VHL), transcript variant 1, mRNA	5.393	1.7*10 ⁻⁷
HOXA9	NM_152739	homeobox A9 (HOXA9), mRNA	5.380	2.3*10 ⁻⁷
MMP9	NM_004994	matrix metallopeptidase 9 (92kDa type IV collagenase) (MMP9), mRNA	5.241	2.3*10 ⁻⁷
CCR6	NM_031409	chemokine (C-C motif) receptor 6 (CCR6), transcript variant 2, mRNA	4.917	4.3*10 ⁻⁷
FOXI2	NM_207426	forkhead (FOXI2), mRNA	4.546	1.1*10 ⁻⁶
S100A9	NM_002965	S100 calcium binding protein A9 (S9), mRNA	4.116	6.1*10 ⁻⁶
NANOG	NM_024865	Nanog homeobox (NANOG), mRNA	4.101	7.9*10 ⁻⁶
CLDN10	NM_182848	claudin 10 (CLDN10), transcript variant a, mRNA	4.081	8.4*10 ⁻⁶
NFE2	NM_006163	nuclear factor (erythroid-derived 2), 45kDa (NFE2), transcript variant 1, mRNA	4.004	2.2*10 ⁻⁵
MLL3	NM_170606	myeloid/lymphoid or mixed-lineage leukemia 3 (MLL3), mRNA	3.943	1.2*10 ⁻⁵
MMP1	NM_002421	matrix metallopeptidase 1 (MMP1), transcript variant 1, mRNA	3.932	3.3*10 ⁻⁵
INHBB	NM_002193	inhibin, beta B (INHBB), mRNA	3.658	7.9*10 ⁻⁵
ITGBL1	NM_004791	integrin, beta-like 1 (with EGF-like repeat domains) (ITGBL1), mRNA	3.299	5.9*10 ⁻⁴

Supplementary Table S2. Genes expressed more highly in BLS-type DLBCL compared with conventional DLBCL by cDNA microarray

		Liver	Spleen		Tumor	Tumor N	о.	Tumor s	ize					
Cell line		tumor	tumor		nodules			mm						
LCL	fO	Death*												
	f1	+	+	100%	+	7		5.5	6.5	3.5	6	11	9.1	5
	f3	+	+	100%	+	3		9	10	4.5				
	f5	+	+	100%	+	2		4	4					
Tumor in liver/spleen		6/6	100%			mean	4	mean	6.5					
HBL2	fO	+	-	50%	+	1		2.5						
	f1	-	-	0%	+	1		6.1						
	f3	+	-	50%	+	1		9						
	f5	+	-	50%	+	1		6						
Tumor in liver/spleen		3/8	38%			mean	1	mean	5.9					
HT	f0	+	-	50%	+	3		3.5	3.5	7				
	f1	-	-	0%	-	0								
	f3	-	-	0%	-	0								
	f5	+	-	50%	+	1		6.5						
Tumor in liver/spleen		2/8	25%			mean	1	mean	5.1					
SU	fO	Death*												
	f1	-	-	0%	+	1		3						
	f3	+	-	50%	+	1		4						

Supplementary Table S3. The result of xenograft mice injected with LCL and DLBCL cell lines

	f5	+	+	100%	+	3		2	4	5				
Tumor in liver/spleen		3/6	50%			mean	1.7	mean	3.6					
U2932	fO	+	-	50%	+	1		5						
	f1	+	-	50%	+	2		8	3					
	f3	+	+	100%	+	2		11.5	2					
	f5	+	-	50%	+	2		5	3					
Tumor in liver/spleen		5/8	63%			mean	1.8	mean	5.4					
U2940	fO	+	+	100%	+	2		9	7					
	f1	+	+	100%	+	3		6	5	3				
	f3	+	-	50%	+	1		5.5						
	f5	+	+	100%	+	7		11	10	5	4	3	2	2
Tumor in liver/spleen		7/8	88%			mean	3.3	mean	5.6					

*The two early dead mice showed no tumor formation and were excluded for calculation.

Original blotting figures of Figure 2B



Original blotting figures of Fig. 3A, 3D, 3G, and 3J Original Fig. 3A Original Fig. 3D 13 13 HT 10/24 4 + 10/31 YA HT C3 C9 NEN9 03 C9 CION 649 NIH2 HIHZ ZA ent ent ent m C <HOXA9 10/31 HT . C3 C9 NENA C3 C9 CIONENA %13 %A 14T 18T 141 182 141 182 141 182 141 182 3A M2 41 M3 M9 41 4 日本常語 \bigcirc SAMON?

Original Fig. 3G



Original Fig. 3J

< Nanog

- KGAPPH



Original blotting figures of Figure 5



Original blotting figures of Figure 6





Original blotting figures of supplementary Fig. S10

