Supplementary Appendix

This appendix has been provided by the authors to give readers additional information about their work.

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Table of Contents

SUPPLEMENTAL METHODS

Mouse anti–human monoclonal antibodies (MxA: Clone M143, 1:100 dilution, Millipore, Burlington, MA; MHC-1: Clone W6/32, 1:20,000 dilution, BioRad, Hercules, CA) were used for immunohistochemical studies. Cryostat muscle sections (10 μm) were fixed in cold acetone (MHC-1) or cold 1% paraformaldehyde (MxA). This was followed by soaking in cold 0.05mol/L tris(hydroxymethyl)aminomethane (Tris) buffer, pH 7.6, for 5 minutes. Sections fixed in 1% paraformaldehyde were soaked further in Tris buffer at room temperature. Slides were placed in Tris buffer supplemented with 3% porcine serum for 5 minutes before incubation with the primary antibodies. Sections then were washed with Tris saline, soaked in the same Tris buffer, and incubated with peroxidase-labeled polymer conjugated to goat anti–mouse immunoglobulin antibody (Leica Biosystems, Buffalo Grove, IL) for 30 minutes, and then washed with Tris saline. Antibody localization was affected by using a peroxidase reaction with 3,3-diaminobenzidine tetrahydrochloride (Liquid DAB+; Agilent, Santa Clara, CA) as the chromogen. The staining reaction was intensified by using 1% copper sulfate solution for 2 minutes. Slides were washed with water, counterstained with methyl green, dehydrated, and mounted with Micromount (Leica Biosystems, Buffalo Grove, IL).

SARS-CoV-2 immunohistochemistry was performed on formalin- fixed, paraffin-embedded tissue sections, following pressure cooker antigen retrieval with citrate buffer, using rabbit polyclonal antibody targeting SARS-CoV Nucleocapsid protein (NB100-56576; Novus Biologicals, Centennial, CO; 1:500 dilution) detected with Novolink Polymer DS (Leica Biosystems, Buffalo Grove, IL) creating a brown reaction product.