

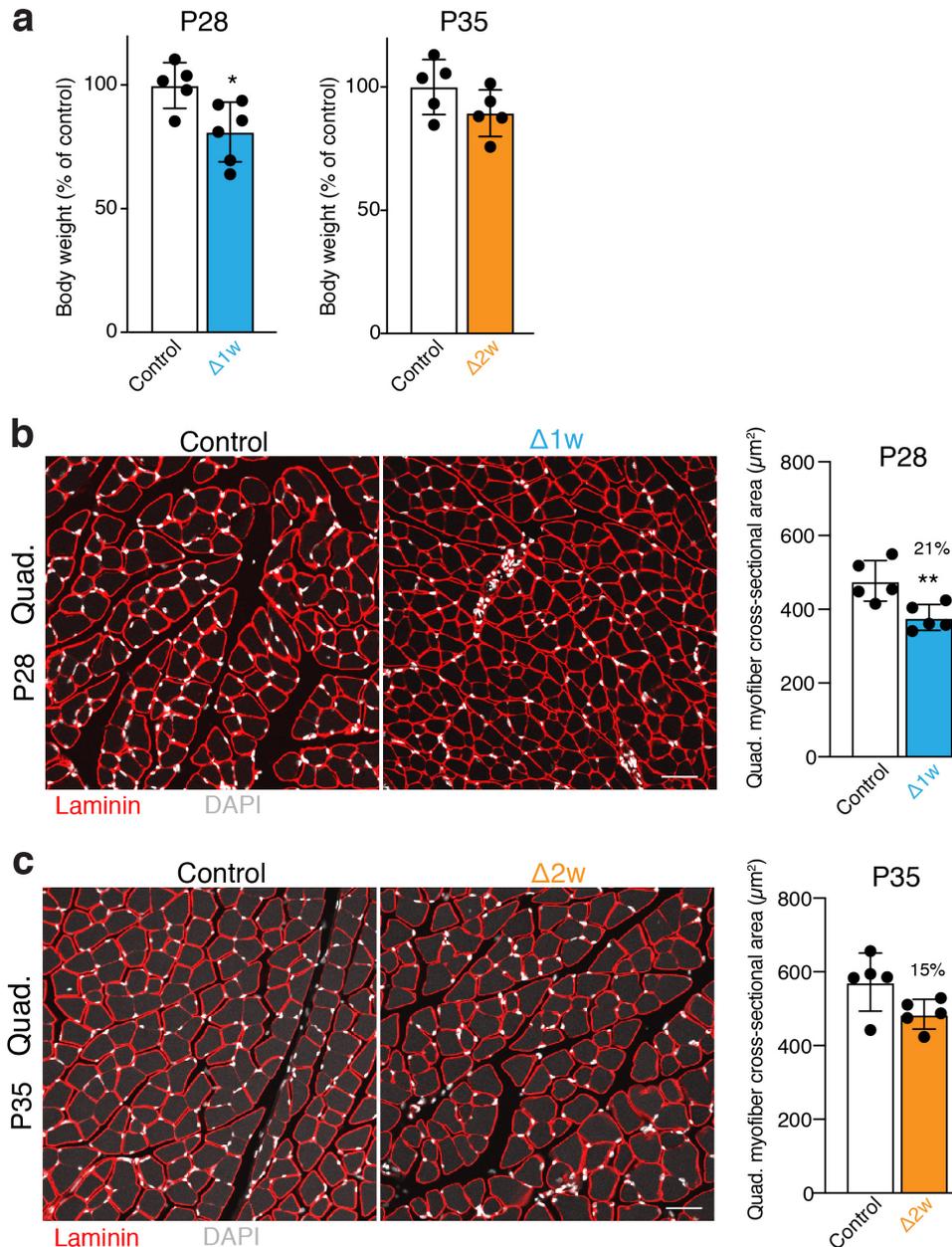
Supplementary Information

Nuclear numbers in syncytial muscle fibers promote size but limit flexibility

Cramer AW. et al.

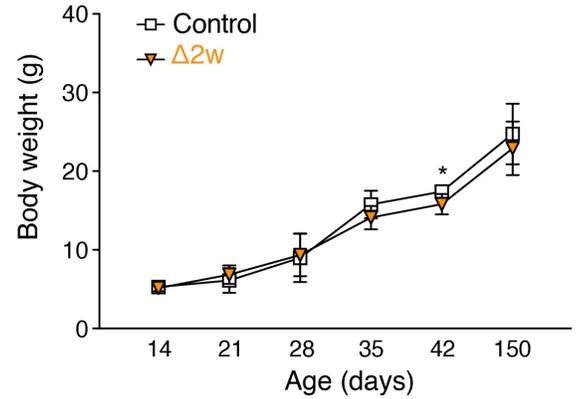
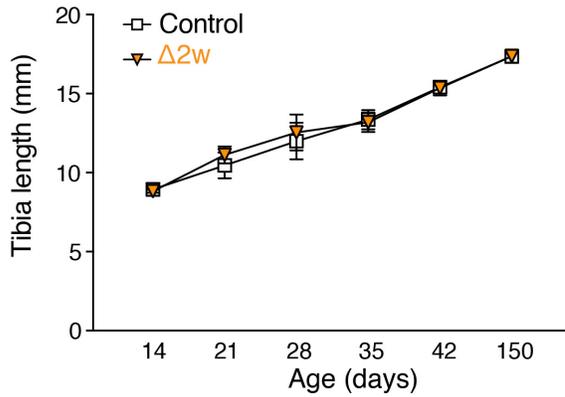
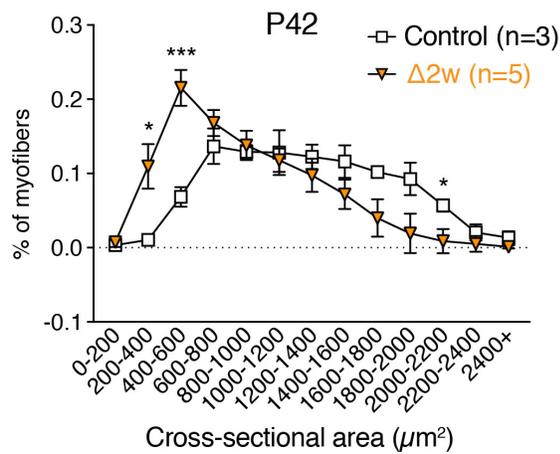
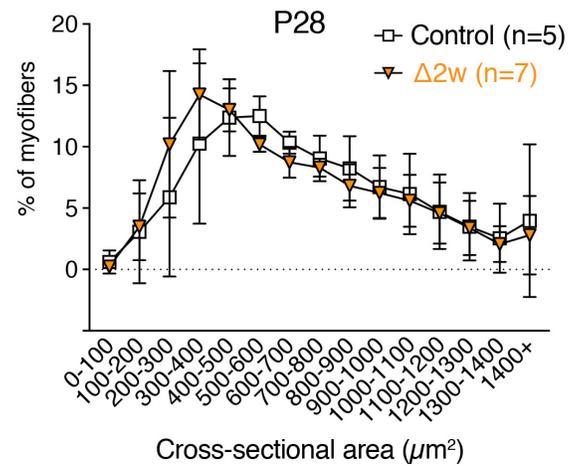
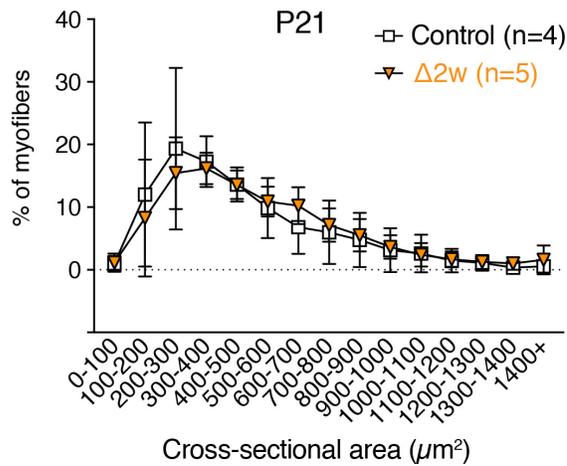
This PDF file includes:

- Supplementary Figure 1
- Supplementary Figure 2
- Supplementary Figure 3
- Supplementary Figure 4
- Supplementary Figure 5
- Supplementary Figure 6
- Supplementary Figure 7



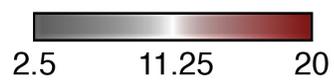
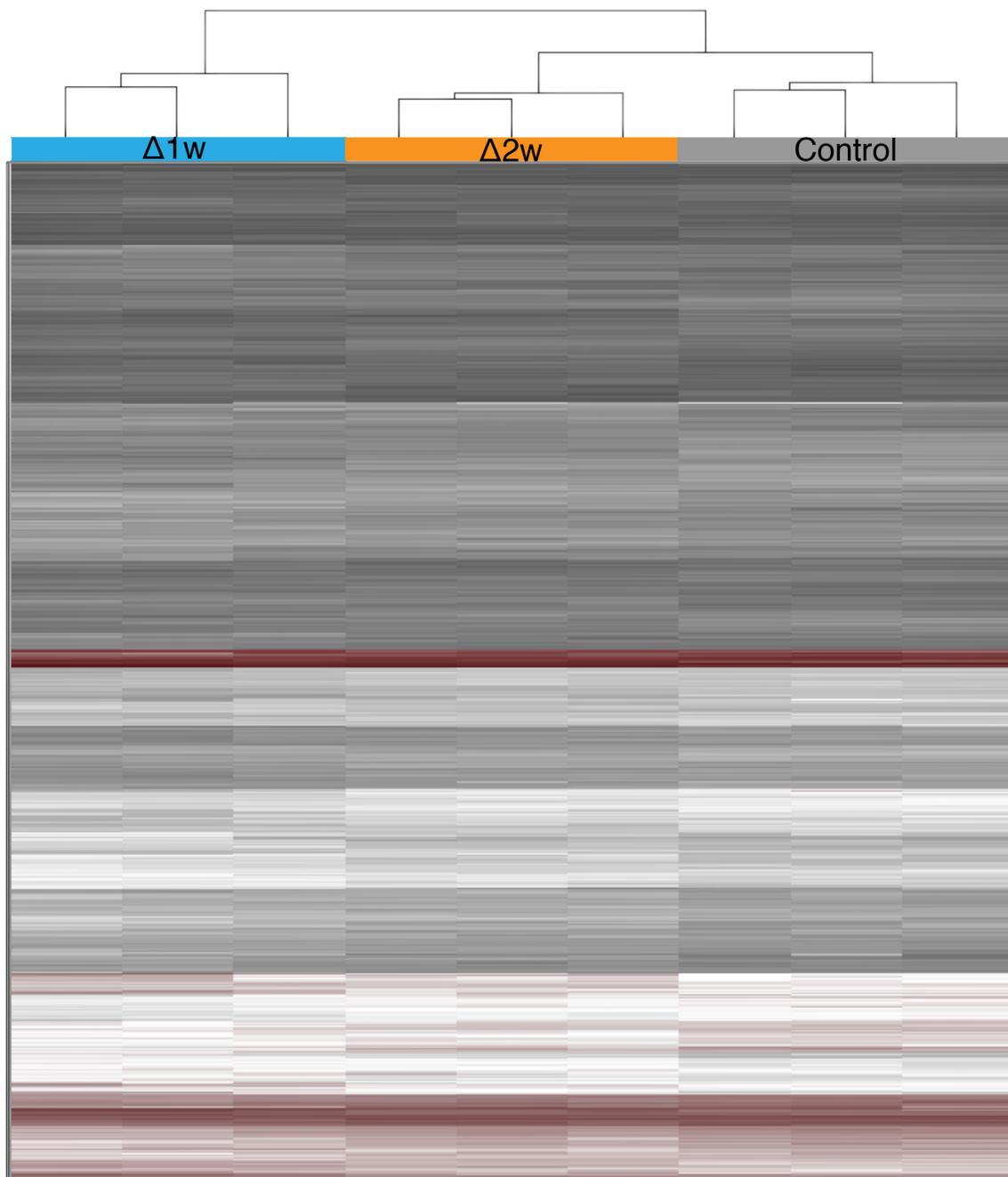
Supplementary Fig. 1 Effects of reduced myonuclear number on cross-sectional area of quadriceps. **a** Body weights of $\Delta 1w$ (left) and $\Delta 2w$ (right) mice expressed as a percent of their respective control mice 4 weeks after tamoxifen administration ($n = 5-6$ biologically independent animals). **b** Sections of quadriceps (Quad.) muscle of $\Delta 1w$ and control mice at P28 (left). Analysis of cross-sectional area (right) revealed a 21% reduction in $\Delta 1w$ myofiber cross-sectional area compared to controls. **c** Sections of quadriceps muscle of $\Delta 2w$ and control mice at P35 (left). Cross-sectional area analysis (right) showed $\Delta 2w$ myofibers are 15% smaller than controls in the quad. Sections in **b** and **c** were immunostained with laminin antibodies (red) and stained with DAPI (white). At least 3 20x images were analyzed per mouse in **b** and **c** ($n = 5$ biologically independent animals). Statistical analyses and data presentation: **a-c**, two-sided unpaired t -test; * $p < 0.05$,

**p<0.01. Data are represented as mean \pm SD. Scale bar: 50 μ m. Source data are provided as a Source Data file.

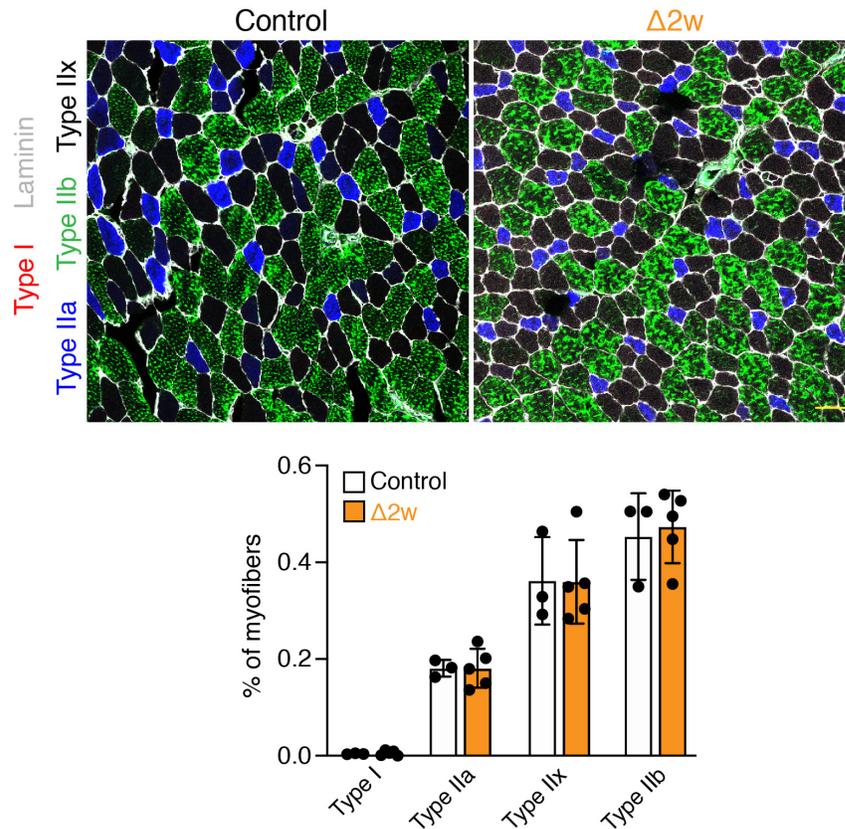
a**b**

Supplementary Fig. 2 Kinetics of muscle growth in $\Delta 2w$ mice. **a** Tibia length (left) and body weight (right) measurements at various timepoints revealed $\Delta 2w$ mice grow at the same rate as controls from P14 to P150 (P14 control n=3, $\Delta 2w$ n=4; P21 control n=6, $\Delta 2w$

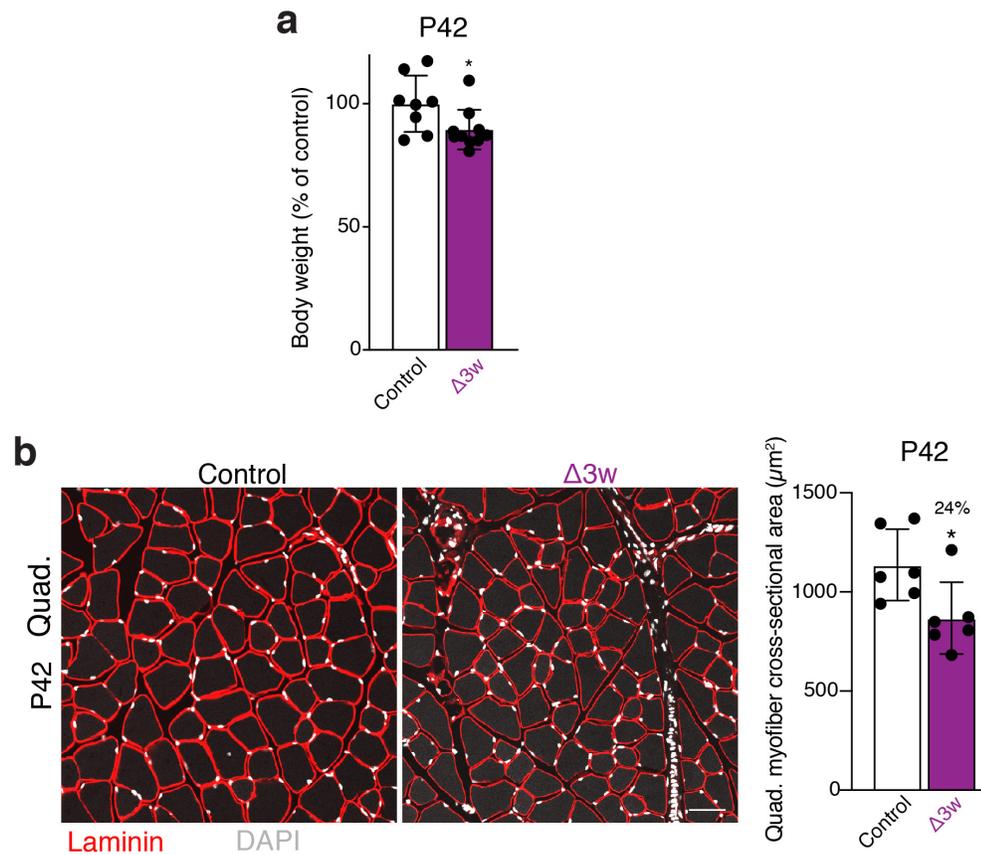
n=10; P28 control n=6, Δ 2w n=8; P35 control n=5, Δ 2w n=5; P42 control n=7, Δ 2w n=9; P150 control n=5, Δ 2w n=15). **b** Tibialis anterior myofiber cross-sectional area size distribution at P21, P28, and P42. At least 3 20x images were analyzed per mouse. Statistical analyses and data presentation: **a** and **b** multiple two-sided unpaired *t*-tests (**a**: one per timepoint, **b**: one per CSA bin); * $p < 0.05$, *** $p < 0.001$. Data are represented as mean \pm SD. Source data are provided as a Source Data file.



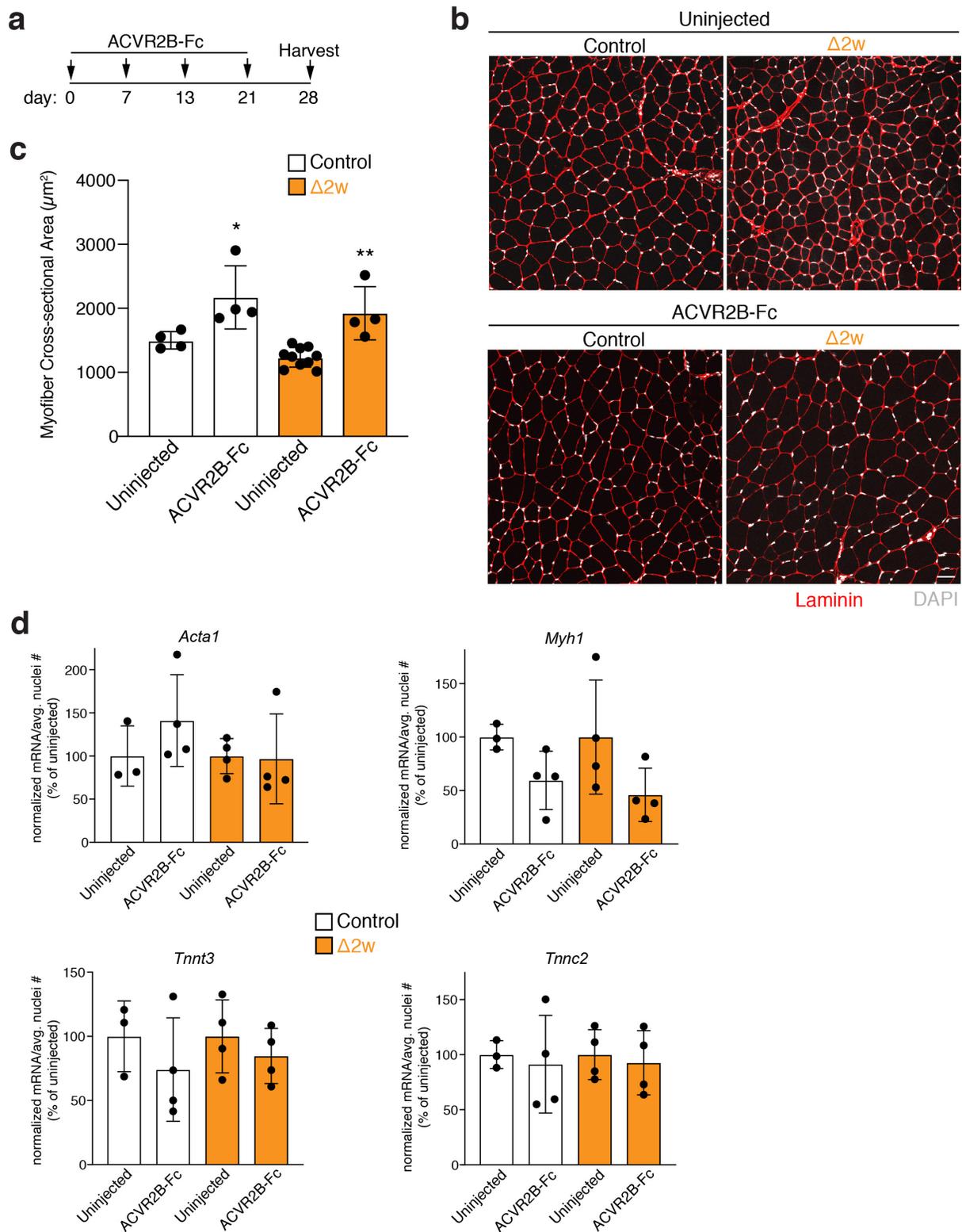
Supplementary Fig. 3 Hierarchical cluster analysis of control, $\Delta 1w$, and $\Delta 2w$ muscle at P28.



Supplementary Fig. 4 Tibialis anterior fiber type distribution in $\Delta 2w$ muscle. Sections of tibialis anterior (TA) muscle in control and $\Delta 2w$ mice at 5-7 months of age immunostained with antibodies for MYH7 (Type I; red), MYH2 (Type IIa; blue), MYH4 (Type IIb; green), and laminin (white) antibodies. Unstained fibers were counted as Type Ix myofibers. 3 20x images were analyzed per mouse ($n = 3-5$ biologically independent animals). Data presentation: Data are represented as mean \pm SD. Scale bar: 50 μ m. Source data are provided as a Source Data file.

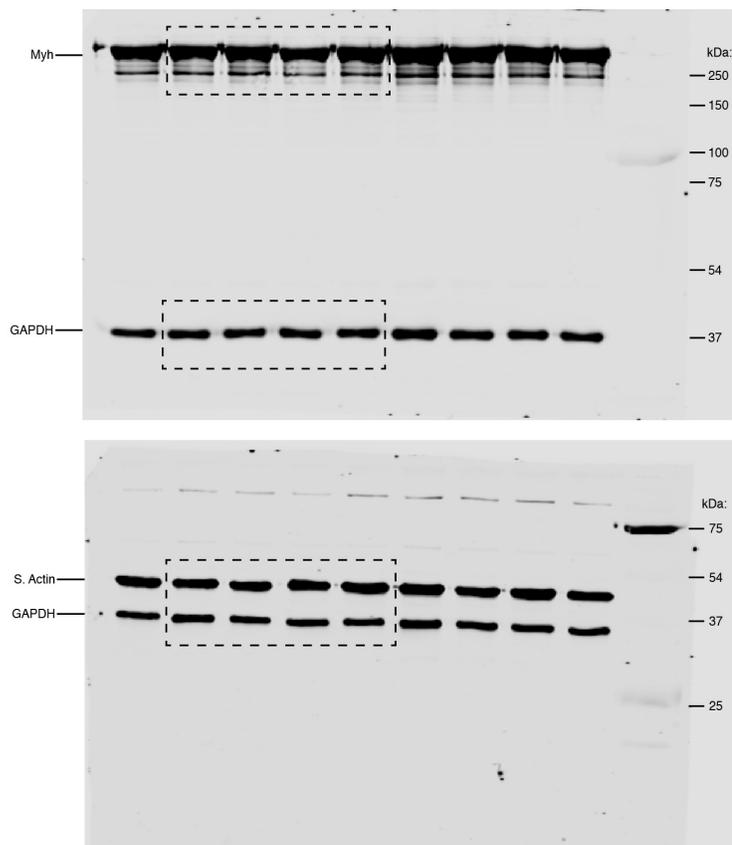


Supplementary Fig. 5 Effects of reduced myonuclear number in $\Delta 3w$ muscle. **a** Body weight of $\Delta 3w$ mice expressed as a percent of control at P42 (n = 8-9 biologically independent animals). **b** Cross-sections of quadriceps (Quad.) muscle of $\Delta 3w$ and control mice at P42 (left). Cross-sectional area analysis (right) reveals a 24% reduction in $\Delta 3w$ quadriceps compared to controls. Sections were immunostained with laminin antibodies (red) and stained with DAPI (white). At least 3 20x images were analyzed per mouse (n = 6 biologically independent animals). Statistical analyses and data presentation: **a** and **b** two-sided unpaired *t*-test; **p*<0.05. Data are represented as mean \pm SD. Scale bar: 50 μm . Source data are provided as a Source Data file.



Supplementary Fig. 6 A stimulus that increases myofiber volume does not elicit changes in mRNA concentrations. **a** Experimental design used to inhibit myostatin signaling. ACVR2B-Fc was administered to 3.5 month old control and $\Delta 2w$ mice once a week for

four weeks. **b** Representative sections of $\Delta 2w$ and control tibialis anterior muscle, uninjected (top) and ACVR2B-Fc-treated (bottom). Images show an increase in myofiber size in both groups following ACVR2B-Fc treatment for four weeks. Sections were immunostained with laminin antibodies (red) and stained with DAPI (white). **c** Analysis of cross-sectional area (CSA) using the images represented in (**b**) showed significant increases in the CSA of tibialis anterior muscle fibers in both control and $\Delta 2w$ muscle injected with ACVR2B-Fc compared to their uninjected controls. At least 3 20x images were analyzed per mouse ($n = 4-10$ biologically independent animals). **d** Concentration of transcripts coding for key skeletal muscle structural genes (*Acta1*, *Myh1*, *Tnnt3*, and *Tnnc2*) on a per nuclear basis are not significantly increased in control or $\Delta 2w$ muscle following ACVR2B-Fc treatment. To obtain these values, relative transcript levels were determined by semi-quantitative qPCR from samples, which were then normalized to total RNA and average myonuclear numbers for each genotype ($n = 3-4$ biologically independent animals). Statistical analyses and data presentation: **c** one-way ANOVA with a Tukey correction for multiple comparisons; * $p < 0.05$, ** $p < 0.01$; significance compared to uninjected groups. Data are represented as mean \pm SD. Scale bar: 50 μm . Source data are provided as a Source Data file.



Supplementary Fig. 7 Full western blots for cropped images shown in Fig. 5h.