Long non-coding RNA levels can be modulated by 5-Azacytidine in *Schistosoma mansoni*

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Supplementary Figure S1: Clustering of RNA-Seq biological replicates

assessed by principal component analysis (PCA). RNA-Seq data from Geyer *et al.*, 2018 ¹ were re-analyzed using the *S. mansoni* genome PRJEA36577 (v.7) retrieved from WormBase and the recently published transcriptome that includes long non-coding RNAs ² as reference. PCA plot was obtained after normalization using the vst function followed by the plotPCA function from DESeq2. Both control and 5-AzaC treated *S. mansoni* female samples are represented by three biological replicates each (n = 3), which are separated by their first two principal components. The control samples are represented by blue dots and the 5-AzaC treated samples by red dots.



Supplementary Figure S2: Venn diagram representing the number of IncRNAs differentially expressed in different conditions of female pairing compared with females under 5-AzaC exposure. Re-analysis of RNA-seq public data from Lu et al., 2016³, focusing on IncRNAs mapping and quantification, followed by comparison with IncRNAs differentially expressed in bisex females after 5-AzaC exposure, as determined by re-analysis of data from Geyer et al., 2018¹. A. IncRNAs downregulated in bisex females after 5-AzaC exposure are compared with IncRNAs differentially expressed between the following conditions: IncRNAs enriched in bisex (paired) females compared with single-sex (unpaired) females (bF>sF); IncRNAs enriched in ovaries from bisex (paired) females compared with ovaries from single-sex (unpaired) females (bO>sO); IncRNAs enriched in ovaries from bisex (paired) females compared with bisex (paired) females (bO>bF). B. IncRNAs downregulated in bisex females after 5-AzaC exposure are compared with IncRNAs differentially expressed between the following conditions: IncRNAs enriched in single-sex (unpaired) females compared with bisex (paired) females (sF>bF); IncRNAs enriched in ovaries from single-sex (unpaired) females compared with ovaries from bisex (paired) females (sO>bO); IncRNAs enriched in bisex (paired) females compared with ovaries from bisex (paired) females (bF>bO). C. IncRNAs upregulated in bisex females after 5-AzaC exposure are compared with IncRNAs differentially expressed between the following conditions: IncRNAs enriched in bisex (paired) females compared with single-sex

(unpaired) females (bF>sF); IncRNAs enriched in ovaries from bisex (paired) females compared with ovaries from single-sex (unpaired) females (bO>sO); IncRNAs enriched in ovaries from bisex (paired) females compared with bisex (paired) females (bO>bF). **D.** IncRNAs upregulated in bisex females after 5-AzaC exposure are compared with IncRNAs differentially expressed between the following conditions: IncRNAs enriched in single-sex (unpaired) females compared with bisex (paired) females (sF>bF); IncRNAs enriched in ovaries from single-sex (unpaired) females (sF>bF); IncRNAs enriched in ovaries from single-sex (unpaired) females compared with ovaries from bisex (paired) females (sO>bO); IncRNAs enriched in bisex (paired) females compared with ovaries from bisex (paired) females (bF>bO). Samples are labeled as bF: bisex (paired) females; sF: single-sex (unpaired) females; bO: bisex (paired) ovaries; sO: single-sex (unpaired) ovaries, according to Lu *et al.*, 2016 ³.



Supplementary Figure S3: Evaluation of *S. mansoni* schistosomula viability after 5-AzaC treatment at different concentrations and incubation times. (A) ATP quantitation using a luminescent assay to assess schistosomula survival under 5-AzaC exposure. *S. mansoni* schistosomula (100-110/well) were incubated with the indicated concentrations of 5-AzaC or with medium only (control) for 24, 48, 72, 96 and 120 h. Viability was expressed as % luminescence values relative to the control (medium only). Mean \pm SEM from two replicate experiments. Two-way ANOVA was applied, and no statistically significant difference was found in any of the comparisons. (B) Light microscopy of schistosomula incubated with the indicated concentrations of 5-AzaC or with medium only (control) for 120 h. Scale bar: 100 µm.



Supplementary Figure S4: Effect of 5-AzaC treatment on adult *S. mansoni* viability and egg laying. A. 5-AzaC does not affect adult *S. mansoni* viability. Nine worm pairs were cultivated in the presence (n = 5) or absence (n = 5) of 491 µM 5-AzaC for 48 h. Worms were collected and ATP levels were measured in control and treated worm couples. Student's unpaired two-sided t test; ns: not significant. **B.** 5-AzaC significantly inhibits *S. mansoni* egg production. Thirty adult worm pairs were cultured either in the presence or absence of 491 µM 5-AzaC. Each culture condition was replicated (n = 5) and eggs were collected and counted after 48 hours. Mean ± SEM are shown. Student's unpaired two-sided t test was applied. *p-value = 0.02. **C.** Light microscopy of schistosome eggs laid by control worm pairs (control) or 5-AzaC treated worm pairs for 48 h (5-AzaC, 491µM). Scale bar: 100 µm.



Supplementary Figure S5: Expression profiles in *S. mansoni* females of selected protein-coding genes differentially expressed after 5-AzaC treatment (491 μ M). Two protein-coding genes were used as controls after reanalysis of RNA-Seq public datasets of 5-AzaC treated *S. mansoni* females from Geyer *et al.*, 2018 ¹ for validation by RT-qPCR in females. For each of the two protein-coding genes, their expression profiles in RNA-Seq are shown as TPM (transcripts per million) on the left, whereas the RT-qPCR results are shown on the right: A. Smp_151640 (*Insulin-like growth factor I*); B. Smp_121390 (*Genome polyprotein*). For the RNA-Seq data, three biological replicates were analyzed; the fold-changes and p-values represented by asterisks that are shown in the brackets were obtained using DESeq2. For the RT-qPCR data, mean ± SEM from five biological replicates are shown; *p<0.05, ****p<0.0001. Student's unpaired two-sided t test.



Supplementary Figure S6: Expression profiles in *S. mansoni* males of selected protein-coding genes differentially expressed after 5-AzaC treatment (491 μM). Two protein-coding genes were used as controls after reanalysis of RNA-Seq public datasets of 5-AzaC treated *S. mansoni* females from Geyer *et al.*, 2018 ¹ for evaluation of differential expression by RT-qPCR in males. For each of the two protein-coding genes, the expression profiles in controls and in 5-AzaC treated *S. mansoni* males by RT-qPCR are shown: **A**. Smp_151640 (*Insulin-like growth factor I*); **B**. Smp_121390 (*Genome polyprotein*). Mean ± SEM from five biological replicates are shown; Student's unpaired two-sided t test was applied. ****p<0.0001; ns: not significant.



Supplementary Figure S7: Correlation between RNA-Seq and RT-qPCR

analysis. Pearson correlation between the fold changes (FC) in expression measured by RNA-seq or RT-qPCR of six selected genes (four lincRNAs – red dots – and two protein-coding genes – black dots); fold changes were obtained by measuring the expression after treatment of females with 5-AzaC and comparing with expression in the control condition. Log2FC of the six genes obtained with the RNA-Seq assay is represented in the x-axis, and log2FC of the six genes obtained with RT-qPCR is represented in the y-axis.



Supplementary Figure S8: RNA-seq expression levels in different S. mansoni stages of protein-coding genes used as sample markers. The expression levels (shown as log2 of normalized counts) of the five proteincoding genes whose gene IDs are indicated at the top of each panel are shown. The y-axis shows the expression level for each protein-coding gene in the RNAseq assays (log2 of normalized counts) as determined at the stage indicated in the x-axis as follows: miracidia/sporocysts (M/S), cercariae (C), schistosomula (S), juveniles (J), adult males (M), adult females (F), posterior somatic tissues (P), heads (H) and tails (T). A. Smp_027920 (Tubulin alpha-1 chain, with high expression in eggs); B. Smp_044250 (STAM-binding protein, with high expression in cercariae); C. Smp 033040 (L-lactate dehydrogenase A chain, with high expression in schistosomula, juveniles and adult males); D. Smp 126730 (5-hydroxytryptamine receptor 1A, with high expression in juveniles and adult males); E. Smp_000430 (Putative eggshell protein, with high expression in adult females). Only transcripts that were upregulated in one stage/tissue when compared with all others were considered as significantly more expressed in that stage/tissue and are marked with an asterisk. *p-value < 0.05.



Supplementary Figure S9: Clustering of RNA-Seq biological replicates assessed by principal component analysis (PCA). RNA-Seq data from 26 public RNA-Seq libraries (listed in Supplementary Table S7) representing six life-cycle stages (miracidia/sporocysts, cercariae, schistosomula, juveniles, adult males, adult females) and adult worm heads, tails and posterior somatic tissues were re-analyzed using the *S. mansoni* genome PRJEA36577 (v.7) retrieved from WormBase and the recently published transcriptome that includes long non-coding RNAs ² as reference. PCA plot was obtained after normalization using the vst function followed by the plotPCA function from DESeq2.

Supplementary References

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