

Supplementary Materials for
Enantioselective Intermolecular Radical C–H Amination

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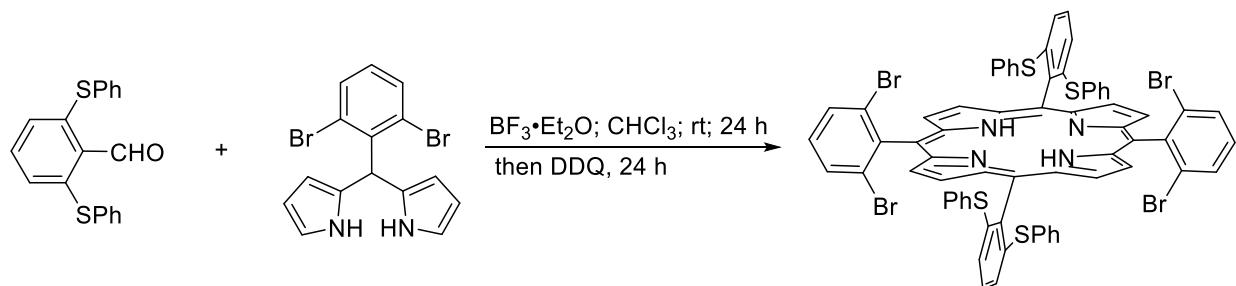
1. General Considerations

Proton nuclear magnetic resonance (^1H NMR) spectra and carbon nuclear magnetic resonance (^{13}C NMR) spectra were recorded on a Varian 600-MHz or Bruker 500-MHz or Bruker 400-MHz instrument. Chemical shifts for protons are reported in parts per million downfield from tetramethylsilane and are referenced to residual protium in the NMR solvent ($\text{CHCl}_3 = 7.26$ ppm, $(\text{CH}_3)_2\text{CO} = 2.05$ ppm, $(\text{CH}_3)_2\text{SO} = 2.5$ ppm). Chemical shifts for carbon are reported in parts per million downfield from tetramethylsilane and are referenced to the carbon resonances of the solvent residual peak ($\text{CDCl}_3 = 77.00$ ppm). High-resolution mass spectrometry was performed on a Micromass LCT ESI-MS and JEOL Accu TOF Dart at the Mass Spectrometry Facility, Boston College. Infrared spectra were measured with a Nicolet Avatar 320 spectrometer with a Smart Miracle accessory. Optical rotations were measured on a Rudolph Research Analytical AUTOPOL® IV digital polarimeter. HPLC measurements were carried out on a Shimadzu HPLC system with Chiralcel OD-H, OJ-H, AD-H, IA, IB, IC, ID, IE, IF and Whelk columns. The UV-Vis absorption spectra in the range 200–700 nm were measured with an Evolution 300 UV-VIS spectrophotometer using quartz cuvettes with 1.0 dm optical path length.

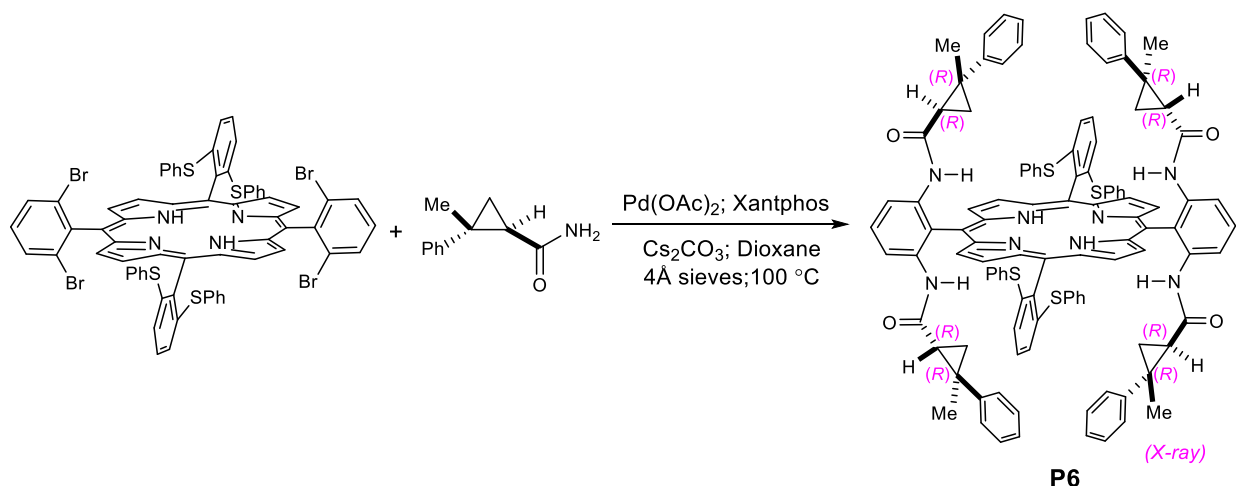
Unless otherwise noted, all catalytic reactions were performed under an atmosphere of N_2 , in oven-dried glassware following standard vacuum line techniques. Gas tight syringes were used to transfer liquid reagents and solvents in catalytic reactions. Anhydrous solvents as well as other commercial reagents were purchased from Sigma-Aldrich, Acros, Alfa Aesar, Strem, Oakwood Products Inc., TCI, or Matrix Scientific and used as received unless otherwise stated. All azides were synthesized according to the reported procedure (1, 2). Thin layer chromatography was performed on Merck TLC plates (silica gel 60 F254). Flash column chromatography was performed with ICN silica gel (60 Å, 230-400 mesh, 32-63 μm).

2. Synthesis of Catalysts and Substrates

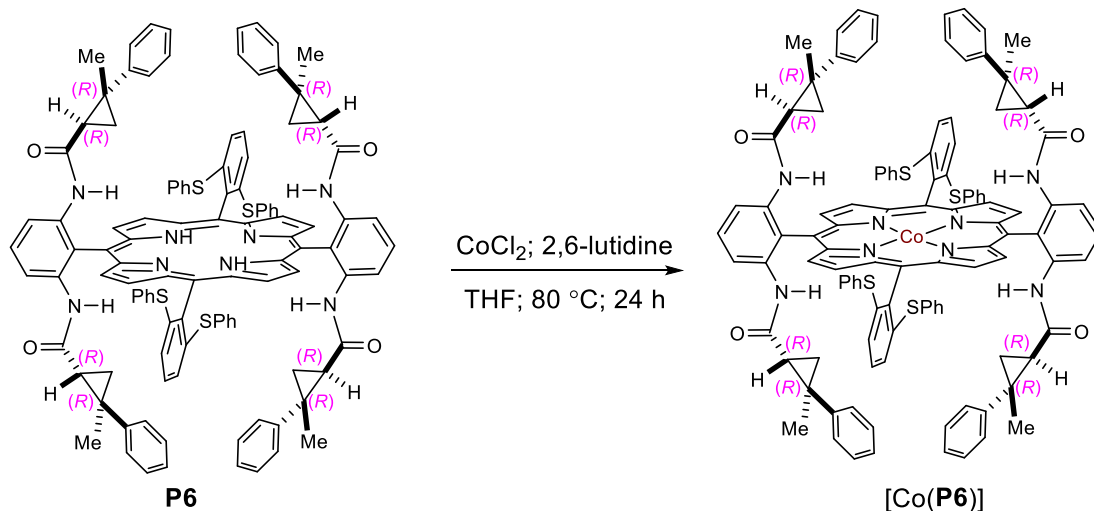
2.1. Synthesis of [Co(P6)] (P6 = 2,6-DiPhS-QingPhyrin)



5,15-bis(2,6-bis(phenylthio)phenyl)-10,20-bis(2,6-dibromophenyl)porphyrin was synthesized through modified procedure of our reported method (3) : A mixture of 2,6-bis(phenylthio)benzaldehyde (4) (828 mg, 2.6 mmol), *meso*-(2,6-dibromophenyl)dipyrromethane (1 g, 2.6 mmol) in chloroform (300 mL) was purged with nitrogen for 5 min. Boron trifluoride diethyl etherate (0.26 mL) was added dropwise via a syringe and the flask was wrapped with aluminum foil to shield it from light. The solution was stirred under a nitrogen atmosphere at room temperature for 24 h, and then 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) (700 mg, 3 mmol) was added as powder at one time. After 24 h, 1 mL of triethylamine was added. The reaction solution was then allowed to sit in refrigerator for overnight to precipitate the product. The precipitate was then filtrated out through Büchner funnel. The residue was washed several times with methanol and then hexanes to afford the pure compound **5,15-bis(2,6-bis(phenylthio)phenyl)-10,20-bis(2,6-dibromophenyl)porphyrin** (510 mg, 29% yield). ¹H NMR (600 MHz, DMSO-*d*₆): δ 8.77 (brs, 4H), 8.62 (m, 4H), 8.21 (d, *J* = 8.3 Hz, 4H), 7.76 (t, *J* = 8.4 Hz, 2H), 7.67 (t, *J* = 8.3 Hz, 2H), 7.30 (d, *J* = 8.3 Hz, 4H), 7.27 – 7.19 (m, 20H), -2.48 (s, 2H). ¹³C NMR spectrum could not be obtained due to poor solubility of the compound. UV–vis (CHCl₃), λ_{max} nm (log ε): 432 (5.42), 524 (4.37), 560 (3.72), 598 (3.95), 654 (3.36). HRMS (ESI) ([M+H]⁺) Calcd. for C₆₈H₄₃Br₄N₄S₄: 1358.9104; Found: 1358.9092.

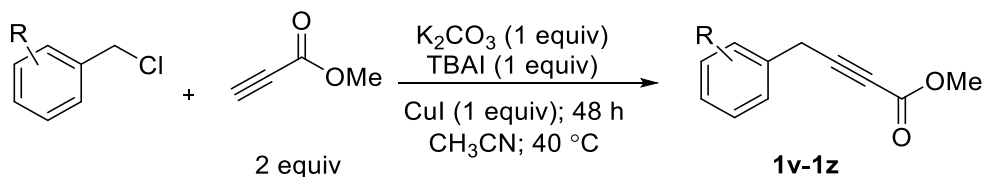


P6 was synthesized according to our previously reported procedure (3). The 5,15-bis(2,6-bis(phenylthio)phenyl)-10,20-bis(2,6-dibromophenyl)porphyrin (360 mg, 0.26 mmol), (1*R*,2*R*)-2-methyl-2-phenylcyclopropane-1-carboxamide (5) (528 mg, 3 mmol), Pd(OAc)₂ (27 mg, 0.12 mmol), Xantphos (138 mg, 0.24 mmol), 4 Å molecular sieves and Cs₂CO₃ (1.95 g, 6 mmol) were placed in an oven-dried, re-sealable Schlenk tube. The tube was capped with a Teflon screw cap, evacuated, and backfilled with nitrogen for three times. The screw cap was replaced with a rubber septum, and dioxane (24 mL) was added via syringe. The tube was purged with nitrogen for 1 min, and then the septum was replaced with the Teflon screw cap. The tube was sealed, and its contents were heated at 100 °C for 72 h with stirring. The resulting mixture was cooled to room temperature, diluted with ethyl acetate, filtered through a short pad of silica gel and concentrated in vacuo. The crude product was then purified by flash chromatography (silica gel, EtOAc/hexanes = 1/3, v/v) to give the desired product **P6** (300 mg, 66% yield). ¹H NMR (600 MHz, CDCl₃): δ 8.96 – 8.91 (m, 8H), 8.51 (brs, 4H), 7.91 (brs, *J* = 8.0 Hz, 2H), 7.50 (t, *J* = 7.8 Hz, 2H), 7.21 – 7.15 (m, 24H), 6.83 (s, 4H), 6.33 (brs, 4H), 6.14 (brs, 8H), 6.01 (brs, 8H), 1.12 (brs, 12H), 0.71 (brs, 4H), 0.54 (brs, 4H), 0.36 (brs, 4H), -2.06 (s, 2H). ¹³C NMR (150 MHz, CDCl₃) δ 168.74, 144.36, 143.09, 138.88, 137.20, 133.54, 133.47, 130.21, 129.88, 129.28, 128.31, 127.31, 125.67, 125.14, 121.72, 118.26, 117.06, 108.19, 30.65, 28.83, 19.30, 18.96. UV–vis (CHCl₃), λ_{max} nm (log ε): 436 (5.28), 524 (4.29), 564 (3.70), 596 (3.9), 650 (3.47). HRMS (ESI) ([M+H]⁺) Calcd. for C₁₁₂H₉₁N₈O₄S₄: 1739.6046; Found: 1739.6077.

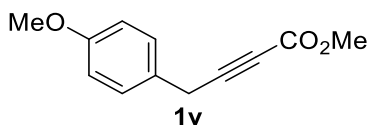


[Co(**P6**)] was synthesized according to our previously reported procedure (3). Free base porphyrin **P6** (230 mg, 0.13 mmol) and anhydrous CoCl_2 (170 mg, 1.3 mmol) were placed in an oven-dried, re-sealable Schlenk tube. The tube was capped with a Teflon screw cap, evacuated, and backfilled with nitrogen. The screw cap was replaced with a rubber septum, 2,6-lutidine (139 mg, 1.3 mmol) and dry THF (5 mL) were added via syringe. The tube was purged with nitrogen for 1 min, and then the septum was replaced with the Teflon screw cap. The tube was sealed, and its contents were heated at 80 °C for 24 h with stirring. The resulting mixture was cooled to room temperature, diluted with ethyl acetate, and transferred to a separatory funnel. The mixture was washed with water 3 times and concentrated in vacuo. The pure compound [Co(**P6**)] was obtained after flash column chromatography (200 mg, 86% yield). UV–vis (CHCl_3), λ_{max} nm (log ϵ): 426 (5.18), 532 (4.2), 570 (3.94). HRMS (ESI) ($[\text{M}]^+$) Calcd. for $\text{C}_{112}\text{H}_{89}\text{CoN}_8\text{O}_4\text{S}_4$: 1796.5222, Found: 1796.5177.

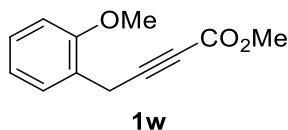
2.2. Synthesis of Substrates



1v-1z were synthesized according to reported procedure (6). An oven-dried round-bottom flask with a stir bar was charged with CuI (5 mmol), K_2CO_3 (5 mmol) and tetrabutylammonium iodide (5 mmol). The flask was placed under a nitrogen atmosphere, and then anhydrous acetonitrile (25 mL) was added via syringe. Next, the benzyl chloride (5 mmol) was added via syringe (if the benzyl chloride was a solid, it was added before the flask was placed under nitrogen), followed by methyl propiolate (10 mmol). The reaction mixture was stirred at $40\text{ }^\circ\text{C}$ for 48 h, and then the reaction was quenched with saturated aqueous NH_4Cl . The reaction mixture was extracted twice with Et_2O , and the combined organic extracts were dried over anhydrous Na_2SO_4 , filtered, and concentrated. The resulting residue was purified by column chromatography (hexanes/ethyl acetate, 40:1 to 20:1 gradient) to afford the products **1v-1z**.

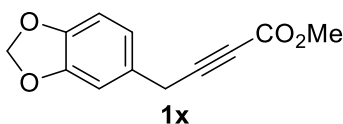


Methyl 4-(4-methoxyphenyl)but-2-ynoate 1v. Yellow oil. Yield: 81%. 1H NMR (500 MHz, $CDCl_3$) δ 7.22 (d, $J = 8.3$ Hz, 2H), 6.86 (d, $J = 8.1$ Hz, 2H), 3.79 (s, 3H), 3.77 (s, 3H), 3.68 (s, 2H). ^{13}C NMR (125 MHz, $CDCl_3$) δ 158.76, 154.12, 129.01, 125.95, 114.18, 87.19, 74.27, 55.29, 52.60, 24.14. This compound is known.

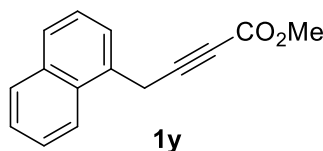


Methyl 4-(2-methoxyphenyl)but-2-ynoate 1w. Yellow oil. Yield: 73%. 1H NMR (600 MHz, $CDCl_3$) δ 7.40 (d, $J = 7.3$ Hz, 1H), 7.25 (t, $J = 7.1$ Hz, 1H), 6.94 (t, $J = 7.5$ Hz, 1H), 6.85 (d, $J = 8.2$ Hz, 1H), 3.83 (s, 3H), 3.76 (s, 3H), 3.70 (s, 2H). ^{13}C NMR (150 MHz, $CDCl_3$) δ 156.64, 154.18,

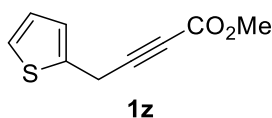
128.91, 128.47, 122.44, 120.56, 110.11, 87.19, 74.21, 55.28, 52.59, 19.45. This compound is known.



Methyl 4-(benzo[d][1,3]dioxol-5-yl)but-2-ynoate 1x. Yellow oil. Yield: 80%. ^1H NMR (600 MHz, CDCl_3) δ 6.80 (s, 1H), 6.80 – 6.75 (m, 2H), 5.94 (s, 2H), 3.77 (s, 3H), 3.64 (s, 2H). ^{13}C NMR (150 MHz, CDCl_3) δ 154.03, 147.94, 146.74, 127.55, 121.02, 108.52, 108.36, 101.24, 86.75, 74.41, 52.62, 24.65. IR (neat, cm^{-1}): 2898, 2237, 1708, 1487, 1241, 1034, 750. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{12}\text{H}_{11}\text{O}_4$: 219.06519, Found: 219.06505.



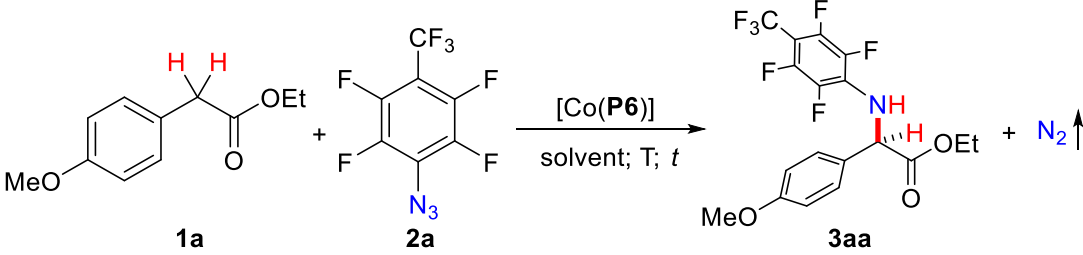
Methyl 4-(naphthalen-1-yl)but-2-ynoate 1y. Yellow oil. Yield: 73%. ^1H NMR (500 MHz, CDCl_3) δ 7.95 (d, $J = 8.4$ Hz, 1H), 7.89 (d, $J = 8.4$ Hz, 1H), 7.81 (d, $J = 8.2$ Hz, 1H), 7.61 – 7.56 (m, 2H), 7.53 (m, 1H), 7.45 (dd, $J = 8.3, 7.0$ Hz, 1H), 4.13 (s, 2H), 3.77 (s, 3H). ^{13}C NMR (150 MHz, CDCl_3) δ 154.08, 133.72, 131.17, 129.81, 128.83, 128.20, 126.53, 126.10, 125.97, 125.53, 122.99, 86.35, 75.27, 52.64, 22.88. This compound is known.



Methyl 4-(thiophen-2-yl)but-2-ynoate 1z. Yellow oil. Yield: 49%. ^1H NMR (600 MHz, CDCl_3) δ 7.18 (d, $J = 5.0$ Hz, 1H), 6.96 (m, 1H), 6.93 (m, 1H), 3.89 (s, 2H), 3.76 (s, 3H). ^{13}C NMR (150 MHz, CDCl_3) δ 153.79, 135.77, 126.93, 125.86, 124.72, 85.34, 74.04, 52.60, 19.73. IR (neat, cm^{-1}): 2953, 2238, 1707, 1433, 1249, 1078, 699. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_9\text{H}_9\text{O}_2\text{S}$: 181.03178, Found: 181.03123.

3. Optimization of Reaction Conditions

Table S1. Optimization of Reaction Conditions using [Co(P6)]^a



entry	temp (°C)	solvent	yield (%) ^b	ee (%) ^c
1	40	benzene	43	97
2	60	benzene	80	88
3	40	fluobenzene	12	92
4	40	hexane	26	95
5	40	α,α,α -trifluorotoluene	25	96
6 ^d	40	α,α,α -trifluorotoluene	68	97
7 ^e	40	α,α,α -trifluorotoluene	95	97
8 ^e	20	α,α,α -trifluorotoluene	38	97

^a Carried out with **1a** (0.10 mmol) and **2a** (0.15 mmol) in the presence of 4Å MS by [Co(P6)] (2 mol %) in solvent (0.5 mL) for 24 h. ^b Isolated yields. ^c Determined by chiral HPLC. ^d [Co(P6)] (4 mol %) and 24 h. ^e [Co(P6)] (4 mol %) and 48 h.

4. DFT-Generated Stereochemical Models for [Co(P6)]-Catalyzed C–H Amination

4.1. Computational Details and Simplifications

The calculations were performed using the Gaussian 09 D.01 program. Considering the time cost and computing resource for the large system with [Co(P6)], all the structures were optimized at the BP86 (7, 8) level of density functional theory with the basis set LANL2DZ (9, 10) in the gas phase at room temperature. Frequency calculations were performed to verify the transition states. Intrinsic reaction coordinate (IRC) (11) calculations were carried out to make sure that every transition state links relevant intermediates. Due to the large size and conformational complexity of the system under investigation, no efforts have been made to improve the accuracy of single point energies at the B3LYP6 (10)/def2-tzvp (9, 10) level of theory along with Grimme's dispersion correction (D3BJ) (12) and SMD (13) model (in trifluorotoluene).

4.2. Transition States of Radical Substitution Step

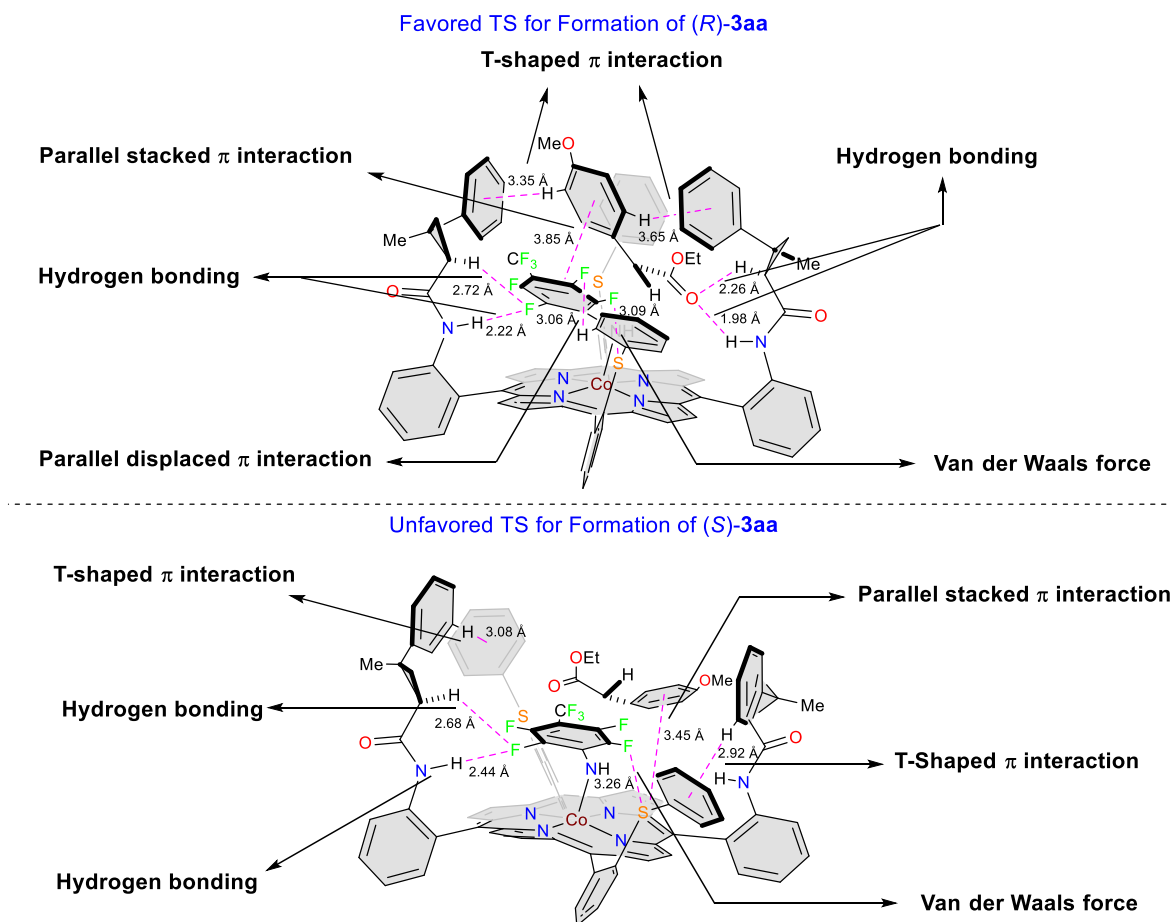


Figure S1. DFT-generated stereochemical models for [Co(P6)]-catalyzed C–H amination. Substituents at Bottom Side Omitted for Clarity

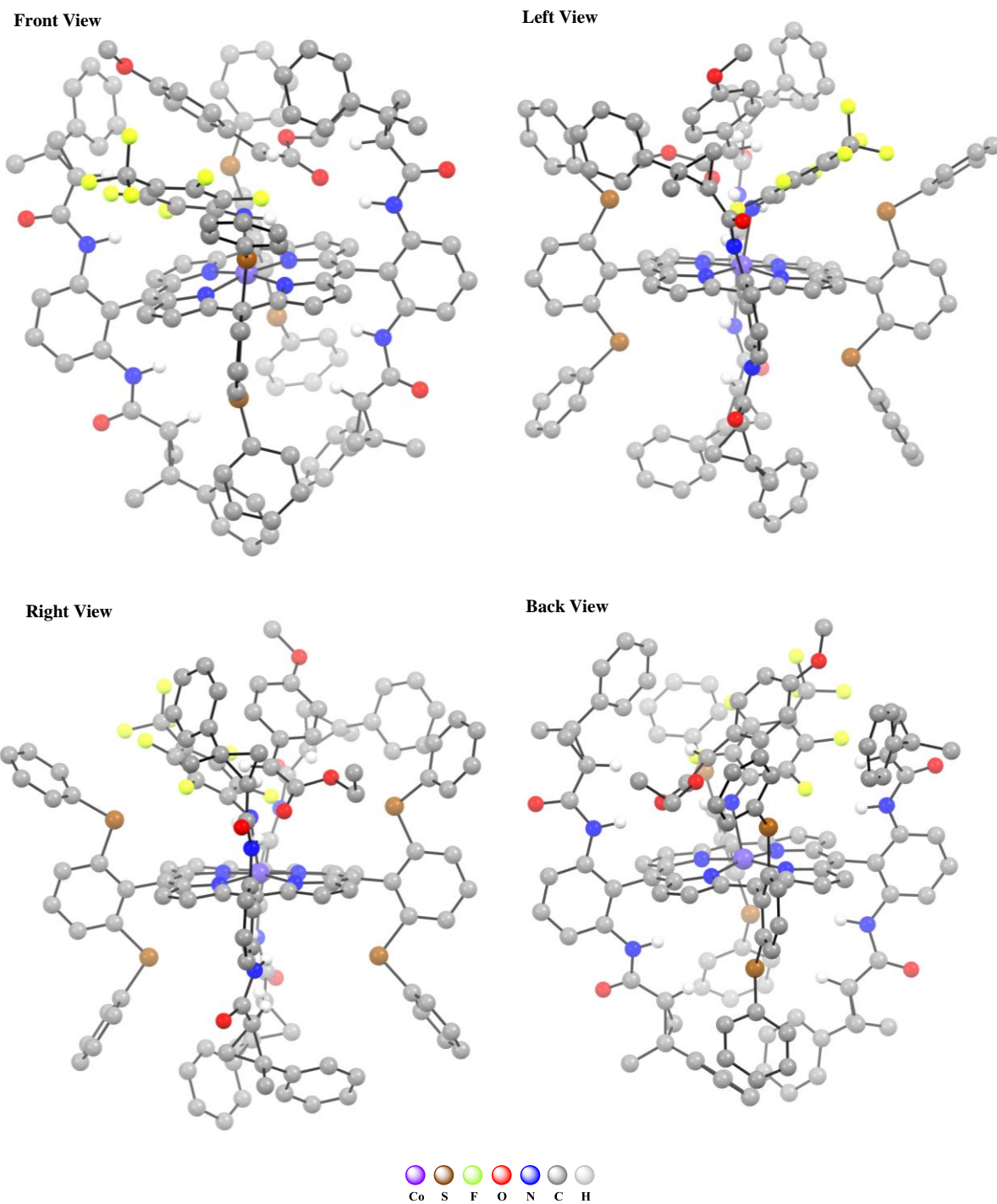
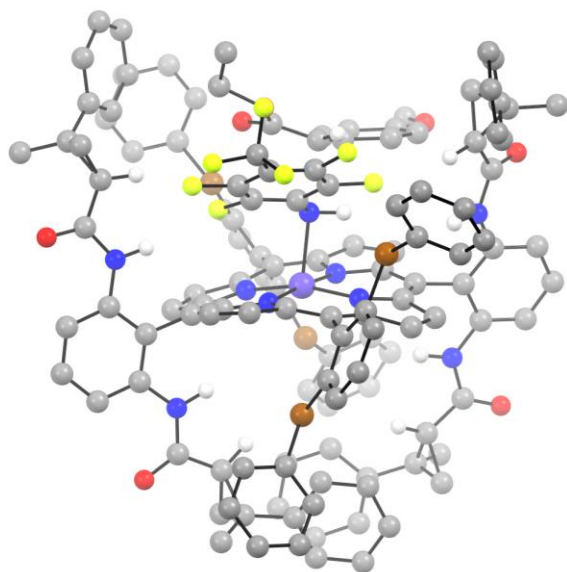
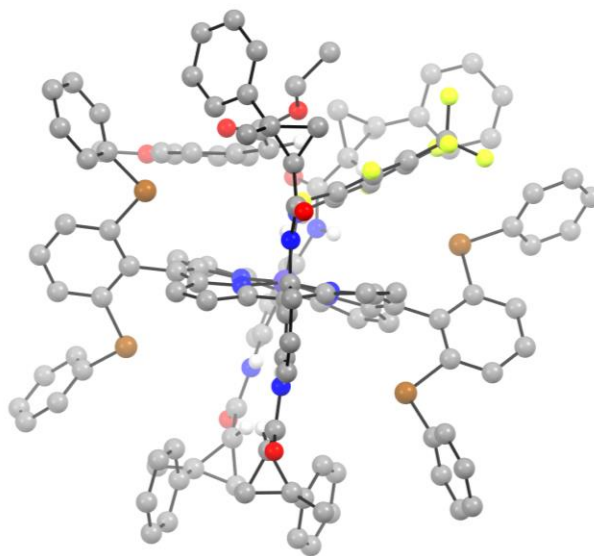


Figure S2. Favored transition state structure for formation of (*R*)-**3aa**; ball and stick model; most hydrogen atoms have been omitted for clarity.

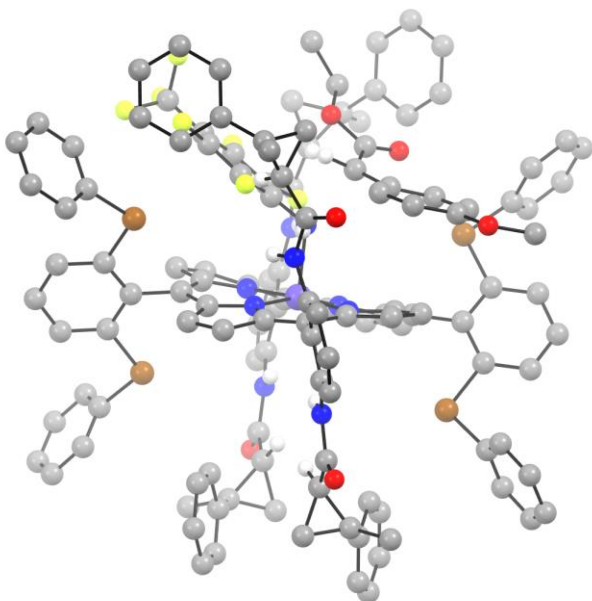
Front View



Left View



Right View



Back View

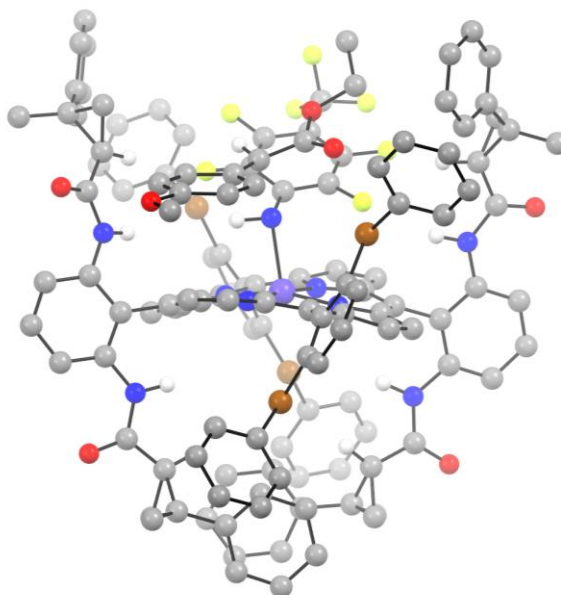


Figure S3. Unfavored transition state structure for formation of (*S*)-**3aa**; ball and stick model; most hydrogen atoms have been omitted for clarity.

XYZ Coordinate

TS for (R)-3aa

G = -6916.656735

	XYZ Coordinate		
	X	Y	Z
H	3.976490	10.686526	-1.150360
C	3.452595	9.728580	-1.051731
H	4.120114	8.975292	-2.990130
H	2.643873	10.186610	0.924963
C	3.532687	8.767128	-2.088676
H	-2.462585	8.381398	-2.408236
C	2.702126	9.448721	0.116639
H	-7.920704	5.468117	-3.940280
C	-2.146599	7.387165	-2.071996
H	-0.113768	8.016713	-1.619362
C	2.861821	7.525884	-1.963311
H	-4.094531	6.484705	-2.416280
C	-7.393807	5.356411	-2.986075
C	2.029393	8.209409	0.248964
H	-8.920198	6.398950	-1.827709
C	-0.823055	7.184263	-1.631024
C	-3.066722	6.318785	-2.082186
C	-7.956677	5.877269	-1.795890
C	2.107861	7.256476	-0.795863
C	-6.147167	4.683749	-2.948805
H	2.929211	6.772489	-2.754345
H	-5.702723	4.281310	-3.865401
H	1.453989	7.979934	1.152289
F	4.377872	4.693775	-3.973472
F	4.805698	2.748731	-5.039284
C	-7.276203	5.718735	-0.562638
C	-0.415801	5.897862	-1.205639
C	-2.653956	5.035231	-1.653464
C	-5.470589	4.531870	-1.714244
H	-0.737749	3.756086	-3.623267
C	4.811375	3.355291	-3.775792
H	-7.709877	6.116041	0.362171
C	-6.033224	5.041725	-0.517753
S	1.305305	5.592469	-0.582441
C	-1.313972	4.786838	-1.219645
F	3.777990	0.576868	-4.039908

S	-3.856971	3.607513	-1.650560
C	-0.655394	2.747471	-3.220966
H	-0.514916	1.391603	-4.983270
F	6.191309	3.455366	-3.390243
H	-1.126431	-1.832494	-8.568992
C	3.994315	2.635634	-2.742022
C	-0.544911	1.556316	-3.907124
O	3.145484	-1.919783	-6.820483
H	-7.208174	0.127533	-5.689756
C	-0.996501	-1.693265	-7.489602
H	-5.503138	4.909207	0.431451
H	1.177504	-1.916178	-7.563351
C	3.509965	1.311131	-2.886388
C	0.298806	-1.742079	-6.941698
C	-2.130908	-1.493043	-6.679813
H	-3.135793	-1.518781	-7.099678
F	4.292937	4.507153	-1.216800
C	-0.909172	3.389129	-0.797918
C	3.738711	3.254104	-1.490602
H	-7.633353	2.718209	-1.203523
C	-0.668078	2.426430	-1.800650
C	-7.350563	-0.924674	-5.378611
C	2.966286	-2.029173	-5.563944
C	-8.026706	1.776304	-1.600795
C	0.472354	-1.555400	-5.541928
H	5.663737	-1.947225	-6.071621
H	-6.188692	1.288074	-2.666764
C	-7.209709	0.965236	-2.426410
H	7.697061	4.360349	1.490944
H	-8.427476	-1.160464	-5.473482
H	-1.446208	5.060827	1.451477
C	2.784339	0.668792	-1.868642
C	-9.352216	1.382835	-1.301211
C	-1.962976	-1.283690	-5.284714
H	-6.776702	-1.575428	-6.057323
H	-9.986178	2.017755	-0.670813
N	1.747856	-1.733937	-4.930750
C	-0.482298	0.494300	-2.911693
C	-0.652210	-1.254053	-4.703189
F	2.431793	-0.669778	-2.057509
C	-7.700811	-0.255899	-2.952462
C	4.040670	-2.536647	-4.645815
C	2.987182	2.619664	-0.500129

C	5.516128	-2.401453	-5.085910
C	-6.907218	-1.117994	-3.919214
N	-3.105451	-1.113106	-4.437270
C	-9.854260	0.164756	-1.823409
H	4.329334	-3.811389	-7.169811
C	7.062568	4.098447	2.346431
C	-1.242315	4.005026	1.622203
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H	7.263668	2.800319	5.533218
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H	7.427883	-4.499566	-4.273411
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O	2.014718	2.561872	7.533978
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H	3.982687	-1.033995	5.558246
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S	2.609544	-4.706670	1.111845
C	-1.436040	-5.477979	1.610171
C	0.997340	-5.570341	1.434706
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C	-6.014185	-7.340625	1.375732
C	-4.498939	-6.028630	3.379800
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C	-1.469258	-6.870743	1.857967
C	3.827723	-5.716697	2.090533
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C	0.967477	-6.968908	1.655599
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H	2.570671	-6.021955	3.853141
C	5.104442	-5.920577	1.511858
H	-2.423835	-7.378646	2.019432
C	3.558142	-6.177751	3.404851
C	-0.267866	-7.610906	1.875221
H	-7.012060	-8.362485	3.025620

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C	4.565016	-6.858317	4.128550
H	-0.298544	-8.693334	2.044469
C	5.847547	-7.057222	3.559311
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TS for (S)-**3aa**

G = -6916.653518

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H	-9.346176	-2.086041	5.069431
C	-7.986309	-3.696712	4.504261
C	-8.305867	-2.433019	5.057083
C	-5.664486	-7.458279	2.412106
H	-5.464275	-8.259170	3.133206
C	-6.172742	-5.381230	0.546098
F	-6.478084	-2.866330	-1.982084
C	-6.639925	-4.133647	4.486475
H	-6.362846	-4.576396	-0.169538
H	-6.386412	-5.104951	4.047120
F	-7.516535	-0.870842	-1.673557
C	-7.282893	-1.615418	5.590459
C	-6.324229	-1.465610	-2.192859
F	-6.364314	-1.275924	-3.582387
C	-4.627326	-6.564643	2.050764
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F	-5.767816	-1.952505	0.559673
C	-5.620026	-3.315013	5.025729
C	-5.929405	-2.046468	5.580130
H	-3.821108	-7.058287	-0.527779
H	-3.628170	-6.665284	2.488628

H	-5.629276	-1.342029	8.271755
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C	-4.838912	-1.207982	-0.165901
H	-4.577352	-3.656932	5.006482
C	-2.891256	-6.623193	-0.903312
H	-7.075243	7.118790	-3.409169
S	-3.541023	-4.325131	0.672843
C	-4.675587	-1.507218	7.735632
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H	-4.393398	-2.568021	7.873398
C	-4.867883	-1.188164	6.245627
H	-1.542687	2.323790	6.131610
H	-6.425386	5.459736	-5.180072
C	-2.073995	-7.363413	-1.782247
F	-4.332953	0.341601	-3.495036
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C	-6.048198	6.735415	-3.449854
C	-1.495352	2.826097	5.159783
C	-2.506663	-5.317490	-0.515928
C	-5.682864	5.801029	-4.447341
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H	-4.872127	2.724355	-6.884809
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H	-3.275503	2.787145	-4.190984
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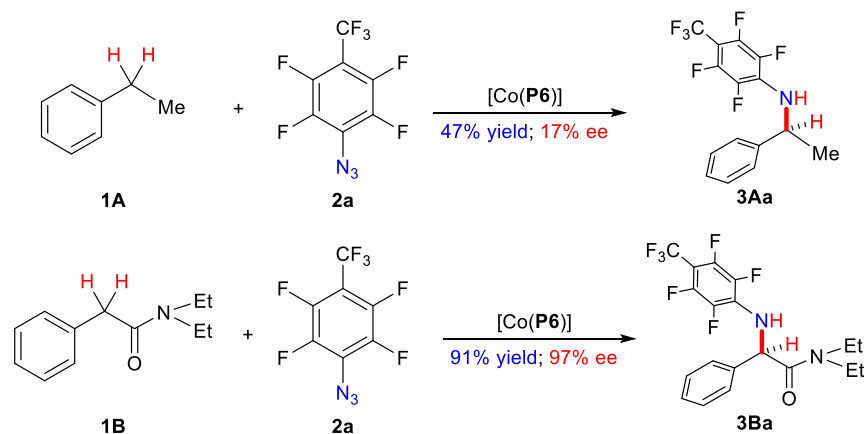
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H	-2.377560	5.358324	-3.606198
O	-4.452792	2.615370	-0.004251
H	-2.557241	5.751519	-6.620029
N	0.092760	-1.723904	0.989190
H	-1.107717	-1.483407	-4.970409
H	-7.063031	3.348557	-0.075096
S	1.135833	-4.785270	-2.429105
C	1.384414	-6.003724	-4.981163
H	0.686762	-5.228193	-5.314683
C	0.476325	-1.915899	2.335408
C	1.766287	-6.066627	-3.618588
C	-0.212441	-0.683335	-3.058388
H	-0.362835	-1.473753	7.740798
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C	0.446138	-1.492133	7.009347
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C	1.822550	2.590322	2.885555
H	2.009321	-1.857323	8.465411
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C	2.007855	-0.159498	-5.577639
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H	3.628135	-3.547021	-1.768091
N	2.910357	-0.770998	-4.655830
C	3.894900	6.832317	1.128778
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C	4.570418	-4.026565	-1.479323
C	3.973917	4.391552	1.128791
H	4.371298	7.798185	1.333667
H	3.861391	-1.932848	6.761829
H	3.181018	-0.619472	-7.345149
H	4.659639	-4.755699	-4.779442
C	4.594192	5.636137	1.390911
C	4.171795	-1.328471	-4.911185
C	5.834185	-5.420097	0.096737
C	4.907584	-1.946963	4.252376
H	5.879193	-6.011038	1.019665
H	4.508318	-1.447944	-2.722158
O	4.673629	-1.419452	-6.079119
C	5.708661	-3.896447	-2.313817
S	4.915770	2.799323	1.342761
C	5.562528	-4.121165	-4.852404
C	4.910112	-1.790364	-3.684203
C	5.656910	-3.171892	-3.648229
H	5.099922	-2.065989	2.045079
O	5.430258	-2.181487	5.391063
C	5.685193	-2.074562	2.972809
H	5.524982	-3.556827	-5.799049
H	5.615025	5.668918	1.782525
H	5.785874	0.082978	1.032909
C	6.980540	-5.292145	-0.727391
H	6.444340	-4.789755	-4.869450
C	6.915766	-4.536478	-1.920174
H	6.632220	0.993968	-1.133254
C	6.793421	-0.181941	0.691030
C	6.454602	-1.875397	-3.783866
C	6.354793	3.339156	2.389356
C	7.272910	0.333251	-0.537415
H	7.718841	3.484760	0.698017
C	7.157845	-1.539173	2.848384
C	6.901894	-3.037936	2.994427
H	7.920617	-5.778598	-0.439537
H	5.232574	3.214418	4.252354
C	7.611610	3.577946	1.783557
H	7.028881	-3.682411	2.119031
H	6.863241	-1.644016	-4.773583
C	6.204765	3.427743	3.795224
C	7.600356	-1.033682	1.485667
H	7.026112	-1.514315	-2.923313

H	7.114328	-3.488331	3.970017
H	7.807631	-4.431518	-2.551069
H	7.538537	0.306917	3.955673
C	7.791011	-0.768143	4.016501
H	7.443090	-1.161942	4.985762
C	8.573441	0.003507	-0.988215
C	8.723820	3.919496	2.591916
C	8.903103	-1.363658	1.020835
C	7.318657	3.775529	4.596799
H	8.947422	0.404969	-1.937951
C	9.387610	-0.851383	-0.203823
C	8.578026	4.022833	3.996322
H	8.892879	-0.858777	3.968209
H	9.698619	4.097158	2.123743
H	7.204991	3.839186	5.685092
H	9.533792	-2.030347	1.623099
H	10.393930	-1.118523	-0.548805
H	9.440960	4.283844	4.620000

4.3. Comparative Study on Intermolecular C–H Amination Reactions of Ethyl Benzene and Arylacetamide



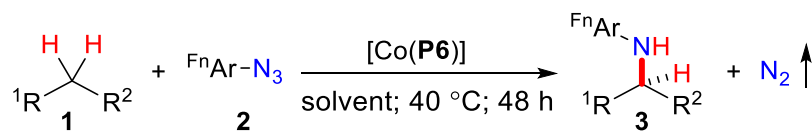
An oven dried Schlenk tube was charged with catalyst ([Co(P6)], 4 mol %) and 4 Å molecular sieves. The Schlenk tube was then evacuated and back filled with nitrogen. The Teflon screw cap was replaced with a rubber septum and **1A** or **1B** (0.1 mmol), azide **2a** (1.5 equiv) and α,α,α -trifluorotoluene (0.5 mL) were added. The Schlenk tube was then purged with nitrogen for 30 s and the rubber septum was replaced with a Teflon screw cap. The mixture was then stirred at 40 °C for 48 h. Following completion of the reaction, the reaction mixture was concentrated and then directly purified by flash chromatography give the product **3Aa** or **3Ba**.

3Aa: Yield: 47%. $[\alpha]_D^{20} = -8^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3): δ 7.34 (t, $J = 7.4$ Hz, 2H), 7.30 – 7.22 (m, 3H), 5.01 (m, 1H), 4.43 (d, $J = 8.6$ Hz, 1H), 1.58 (d, $J = 6.7$ Hz, 1H). $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 144.78 (dm, $J = 258.9$ Hz), 143.60, 136.66 (dm, $J = 240.3$ Hz), 130.29 (m), 128.88, 127.70, 125.43, 121.51 (q, $J = 272.8$ Hz), 96.52 (m), 54.54 (t, $J = 4.3$ Hz), 24.58. $^{19}\text{F NMR}$ (564 MHz, CDCl_3) δ -55.01 (t, $J = 20.8$ Hz, 3F), -143.31 (m, 2F), -158.97 (m, 2F). IR (neat, cm^{-1}): 3340, 2930, 1656, 1512, 1334, 1237, 1133, 984, 872. HRMS (DART) ($[\text{M}]^+$) Calcd. for: $\text{C}_{15}\text{H}_{10}\text{F}_7\text{N}$: 337.06960, Found: 337.06953. HPLC analysis: ee = 17%. IA (98% hexane: 2% isopropanol, 0.8 ml/min): $t_{\text{major}} = 5.57$ min, $t_{\text{minor}} = 5.38$ min.

3Ba: Yield: 91%. $[\alpha]_D^{20} = -135^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 7.39 (m, 2H), 7.35 (t, $J = 7.4$ Hz, 2H), 7.31 (m, 1H), 6.07 (br, 1H), 5.61 (d, $J = 8.6$ Hz, 1H), 3.47 (dq, $J = 13.9$, 7.1 Hz, 1H), 3.34 (m, 2H), 3.19 (dq, $J = 14.4$, 7.0 Hz, 1H), 1.10 (t, $J = 7.0$ Hz, 3H), 0.94 (t, $J = 7.0$ Hz, 3H). $^{13}\text{C NMR}$ (150 MHz, CDCl_3) δ 168.35, 144.78 (dm, $J = 256.0$ Hz), 137.34, 137.20 (dm,

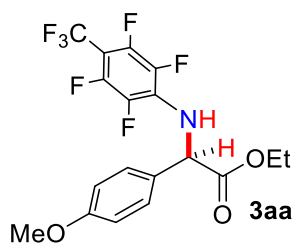
$J = 239.8$ Hz), 129.70 (m), 129.26, 128.81, 127.48, 121.50 (q, $J = 272.7, 272.3$ Hz), 97.04 (m), 58.24 (t, $J = 3.9$ Hz), 41.28, 40.91, 13.47, 12.47. ^{19}F NMR (564 MHz, CDCl_3) δ -55.07 (t, $J = 20.9$ Hz, 3F), -143.50 (m, 2F), -157.60 (m, 2F). IR (neat, cm^{-1}): 3340, 2979, 1652, 1506, 1334, 1133, 979, 883. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{19}\text{H}_{18}\text{F}_7\text{N}_2\text{O}$: 423.13033, Found: 423.13019. HPLC analysis: ee = 97%. IA (80% hexane: 20% isopropanol, 0.5 ml/min): $t_{\text{major}} = 7.60$ min, $t_{\text{minor}} = 8.19$ min.

5. General Procedure for [Co(P6)] Catalyzed C–H Amination with Fluoroaryl Azides

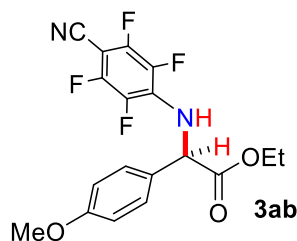


An oven dried Schlenk tube was charged with catalyst ([Co(**P6**)], 4 mol %) and 4 Å molecular sieves. The Schlenk tube was then evacuated and back filled with nitrogen. The Teflon screw cap was replaced with a rubber septum and C–H substrate (0.1 mmol), azide (1.2 or 1.5 equiv) and solvent (α,α,α -trifluorotoluene or benzene) (0.5 mL) were added. The Schlenk tube was then purged with nitrogen for 30 s and the rubber septum was replaced with a Teflon screw cap. The mixture was then stirred at 40 °C for 48 h. Following completion of the reaction, the reaction mixture was concentrated and then directly purified by flash chromatography.

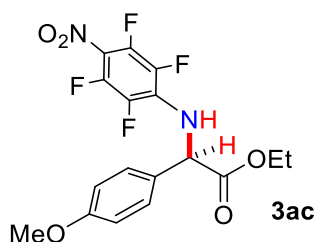
5.1. Enantioselective C–H Amination of 1a with Various Fluoroaryl Azides



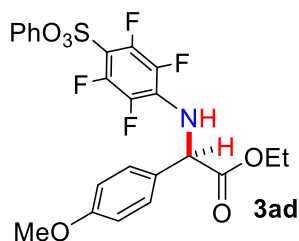
Ethyl (R)-2-(4-methoxyphenyl)-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)acetate **3aa** was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 95%. $[\alpha]_D^{20} = -90^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3): δ 7.29 (d, $J = 8.4$ Hz, 2H), 6.88 (d, $J = 7.8$ Hz, 2H), 5.42 (m, 2H), 4.25 (m, 2H), 3.80 (s, 3H), 1.22 (t, $J = 7.1$ Hz, 3H). $^{13}\text{C NMR}$ (150 MHz, CDCl_3) δ 171.03, 160.00, 144.77 (dm, $J = 249.9$ Hz), 136.93 (dm, $J = 241.6$ Hz), 129.25 (t, $J = 11.1$ Hz), 128.65, 128.15, 121.44 (q, $J = 272.8$ Hz), 114.50, 97.39 (m), 62.30, 60.06, 55.25. $^{19}\text{F NMR}$ (564 MHz, CDCl_3) δ -55.11 (t, $J = 20.9$ Hz, 3F), -143.03 (m, 2F), -158.56 (m, 2F). IR (neat, cm^{-1}): 3387, 2915, 2848, 1737, 1657, 1509, 1332, 1235, 1176, 1131, 713. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{18}\text{H}_{13}\text{F}_7\text{NO}_3$: 424.0784, Found: 424.0793. HPLC analysis: ee = 97%. Whelk (99.5% hexane: 0.5% isopropanol, 1 ml/min): $t_{\text{major}} = 7.48$ min, $t_{\text{minor}} = 9.68$ min.



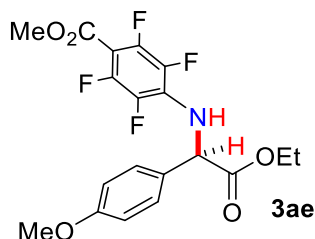
Ethyl (R)-2-((4-cyano-2,3,5,6-tetrafluorophenyl)amino)-2-(4-methoxyphenyl)acetate 3ab was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 53%. $[\alpha]_D^{20} = -200^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 7.26 (d, $J = 8.6$ Hz, 2H), 6.87 (d, $J = 8.5$ Hz, 2H), 5.66 (br, 1H), 5.42 (d, $J = 7.5$ Hz, 1H), 4.20 (m, 2H), 3.79 (s, 3H), 1.20 (t, $J = 7.1$ Hz, 3H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 170.71, 160.08, 147.58 (dm, $J = 254.7$ Hz), 135.99 (dm, $J = 245.8$ Hz), 131.30 (m), 128.30, 128.11, 114.55, 108.53 (t, $J = 3.6$ Hz), 80.82 (t, $J = 17.7$ Hz), 62.49, 59.95, 55.27, 13.93. $^{19}\text{F NMR}$ (470 MHz, CDCl_3) δ -134.81 (m, 2F), -157.67 (m, 2F). IR (neat, cm^{-1}): 3383, 2983, 2235, 1737, 1656, 1510, 1178, 837. HRMS (DART) ($[\text{M}+\text{NH}_4]^+$) Calcd. for: $\text{C}_{18}\text{H}_{18}\text{F}_4\text{N}_3\text{O}_3$: 400.12788, Found: 400.12793. HPLC analysis: ee = 90%. IC (90% hexane: 10% isopropanol, 0.8 ml/min): $t_{\text{major}} = 22.20$ min, $t_{\text{minor}} = 23.52$ min.



Ethyl (R)-2-(4-methoxyphenyl)-2-((2,3,5,6-tetrafluoro-4-nitrophenyl)amino)acetate 3ac was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 60%. $[\alpha]_D^{20} = -245^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 7.26 (d, $J = 8.2$ Hz, 2H), 6.88 (d, $J = 9.0$ Hz, 2H), 5.65 (d, $J = 8.4$ Hz, 1H), 5.41 (d, $J = 8.4$ Hz, 1H), 4.22 (m, 2H), 3.79 (s, 3H), 1.21 (t, $J = 6.9$ Hz, 3H). $^{13}\text{C NMR}$ (150 MHz, CDCl_3) δ 173.34, 162.77, 144.86 (dm, $J = 261.3$ Hz), 138.83 (dm, $J = 243.6$ Hz), 132.97 (t, $J = 10.7$ Hz), 130.86, 130.74, 122.72, 117.22, 65.19, 62.63, 57.90, 16.61. $^{19}\text{F NMR}$ (564 MHz, CDCl_3) δ -146.96 (m, 2F), -158.46 (m, 2F). IR (neat, cm^{-1}): 3379, 2983, 1737, 1640, 1549, 1334, 1177, 1027, 769. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{17}\text{H}_{15}\text{F}_4\text{N}_2\text{O}_5$: 403.09116, Found: 403.09007. HPLC analysis: ee = 96%. IC (90% hexane: 10% isopropanol, 0.8 ml/min): $t_{\text{major}} = 18.12$ min, $t_{\text{minor}} = 18.83$ min.

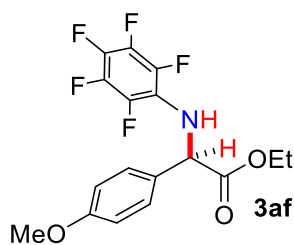


Ethyl (R)-2-(4-methoxyphenyl)-2-((2,3,5,6-tetrafluoro-4-(phenoxysulfonyl)phenyl amino) acetate 3ad was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 60%. $[\alpha]_D^{20} = -67^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.33 (t, $J = 7.5$ Hz, 2H), 7.27 – 7.32 (m, 3H), 7.13 (d, $J = 8.0$ Hz, 2H), 6.89 (d, $J = 8.2$ Hz, 2H), 5.67 (d, $J = 7.3$ Hz, 1H), 5.42 (d, $J = 7.3$ Hz, 1H), 4.21 (m, 2H), 3.81 (s, 3H), 1.21 (t, $J = 7.1$ Hz, 3H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 170.70, 160.08, 149.10, 145.45 (dm, $J = 260.8$ Hz), 136.93 (dm, $J = 246.1$ Hz), 131.71 (m), 129.92, 128.28, 128.13, 127.59, 121.70, 114.55, 102.01 (m), 62.50, 59.93, 55.29, 13.93. $^{19}\text{F NMR}$ (470 MHz, CDCl_3) δ -136.72 (m, 2F), -157.89 (m, 2F). IR (neat, cm^{-1}): 3383, 2936, 1737, 1636, 1501, 1200, 1144, 777. HRMS (DART) ($[\text{M}+\text{NH}_4]^+$) Calcd. for: $\text{C}_{23}\text{H}_{23}\text{F}_4\text{N}_2\text{O}_6\text{S}$: 531.12075, Found: 531.12103. HPLC analysis: ee = 87%. ODH (95% hexane: 5% isopropanol, 0.8 ml/min): $t_{\text{major}} = 25.48$ min, $t_{\text{minor}} = 27.93$ min.

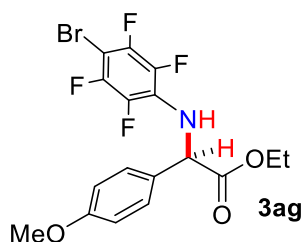


Methyl (R)-4-((2-ethoxy-1-(4-methoxyphenyl)-2-oxoethyl)amino)-2,3,5,6-tetrafluorobenzoate 3ae was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 87%. $[\alpha]_D^{20} = -212^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 7.27 (d, $J = 8.6$ Hz, 2H), 6.86 (d, $J = 8.6$ Hz, 2H), 5.40 (m, 2H), 4.19 (m, 2H), 3.87 (s, 3H), 3.78 (s, 3H), 1.20 (t, $J = 7.1$ Hz, 3H). $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 171.07, 160.90 (t, $J = 3.0$ Hz), 159.92, 146.17 (dm, $J = 253.4$ Hz), 136.84 (dm, $J = 238.9$ Hz), 129.07 (m), 128.75, 128.14, 114.43, 99.86 (t, $J = 14.3$ Hz), 62.22, 60.13, 55.24, 52.52, 13.94. $^{19}\text{F NMR}$ (564 MHz, CDCl_3) δ -140.24 (m, 2F), -159.03 (m, 2F). IR (neat, cm^{-1}): 3385, 2957, 2840, 1731, 1652, 1512, 1232, 1180, 1026. HRMS (DART) ($[\text{M}+\text{NH}_4]^+$) Calcd. for: $\text{C}_{19}\text{H}_{21}\text{F}_4\text{N}_2\text{O}_5$: 433.13811, Found: 433.13813. HPLC

analysis: ee = 96%. IC (90% hexane: 10% isopropanol, 0.8 ml/min): t_{major} = 18.16 min, t_{minor} = 19.37 min.

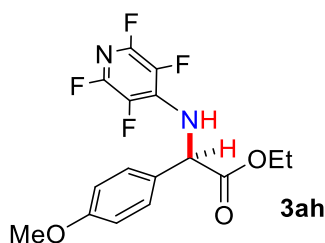


Ethyl (*R*)-2-(4-methoxyphenyl)-2-((perfluorophenyl)amino)acetate 3af was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 72%. $[\alpha]_D^{20}$ = -60° (c = 0.5, CHCl_3). ^1H NMR (500 MHz, CDCl_3): δ 7.26 (d, J = 8.5 Hz, 2H), 6.85 (d, J = 8.5 Hz, 2H), 5.26 (d, J = 9.0 Hz, 1H), 4.88 (br, 1H), 4.19 (m, 2H), 3.78 (s, 3H), 1.21 (t, J = 7.0 Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3): δ 171.50, 159.81, 138.32 (dm, J = 238.8 Hz), 137.95 (dm, J = 246.0 Hz), 134.22 (dm, J = 244.1 Hz), 128.86, 128.19, 121.56 (m), 114.35, 62.01, 60.88 (t, J = 3.6 Hz), 55.22, 13.96. ^{19}F NMR (376 MHz, CFCl_3 , CDCl_3): δ . -158.03 (m, 2F), -164.50 (m, 2F), -170.12 (m, 1F). IR (neat, cm^{-1}): 3379, 2938, 1736, 1611, 1511, 1247, 1176, 1022, 967, 836. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{17}\text{H}_{13}\text{F}_5\text{NO}_3$: 374.0816, Found: 374.0826. HPLC analysis: ee = 95%. Whelk (99% hexane: 1% isopropanol, 1 ml/min): t_{major} = 6.59 min, t_{minor} = 7.90 min.

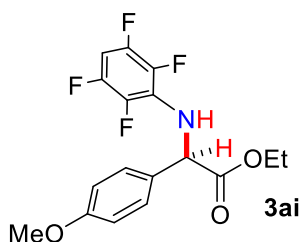


Ethyl (*R*)-2-((4-bromo-2,3,5,6-tetrafluorophenyl)amino)-2-(4-methoxyphenyl)acetate 3ag was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 64%. $[\alpha]_D^{20}$ = -54° (c = 1.0, CHCl_3). ^1H NMR (500 MHz, CDCl_3): δ 7.28 (d, J = 8.5 Hz, 2H), 6.86 (d, J = 8.0 Hz, 2H), 5.34 (d, J = 8.5 Hz, 1H), 5.13 (d, J = 8.5 Hz, 1H), 4.24 (m, 2H), 3.78 (s, 3H), 1.21 (t, J = 7.0 Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3): δ 171.35, 159.84, 145.08 (dm, J = 243.0 Hz), 138.01 (dm, J = 241.7 Hz), 131.11, 128.91, 128.18, 125.34 (t, J = 12.3 Hz), 114.37,

86.91 (t, $J = 23.0$ Hz), 62.07, 60.50 (t, $J = 3.9$ Hz), 55.22, 13.96. ^{19}F NMR (376 MHz, CFCl_3 , CDCl_3): δ -136.17 (m, 2F), -156.76 (m, 2F). IR (neat, cm^{-1}): 3382, 2936, 1735, 1641, 1494, 1248, 1177, 1032, 947, 833, 796, 622, 529. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{17}\text{H}_{13}\text{BrF}_4\text{NO}_3$: 434.0015, Found: 434.0026. HPLC analysis: ee = 95%. ADH (99% hexane: 1% isopropanol, 1 ml/min): $t_{\text{major}} = 7.79$ min, $t_{\text{minor}} = 8.46$ min.

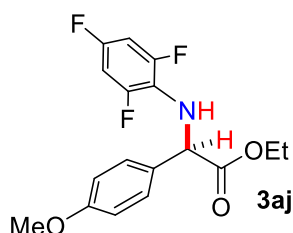


Ethyl (R)-2-(4-methoxyphenyl)-2-((perfluoropyridin-4-yl)amino)acetate 3ah was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 87%. $[\alpha]_D^{20} = -97^\circ$ ($c = 1.0$, CHCl_3). ^1H NMR (500 MHz, CDCl_3): δ 7.29 (d, $J = 9.0$ Hz, 2H), 6.89 (d, $J = 8.5$ Hz, 2H), 5.74 (m, 1H), 5.47 (d, $J = 7.5$ Hz, 1H), 4.27 (m, 2H), 3.80 (s, 3H), 1.22 (t, $J = 7.5$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3): δ 170.69, 160.07, 144.05 (td, $J = 15.5$, 235.4 Hz), 135.76 (m), 131.47 (dm, $J = 246.4$ Hz), 128.38, 128.13, 114.54, 62.45, 59.61 (t, $J = 3.8$ Hz), 55.28, 13.93. ^{19}F NMR (376 MHz, CFCl_3 , CDCl_3): δ -93.85 (m, 2F), -162.63 (m, 2F). IR (neat, cm^{-1}): 3386, 2939, 1736, 1646, 1479, 1248, 1151, 1030, 940, 836, 727, 624. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{16}\text{H}_{13}\text{F}_4\text{N}_2\text{O}_3$: 357.0862, Found: 357.0873. HPLC analysis: ee = 93%. Whelk (97% hexane: 3% isopropanol, 1 ml/min): $t_{\text{major}} = 8.47$ min, $t_{\text{minor}} = 9.62$ min.



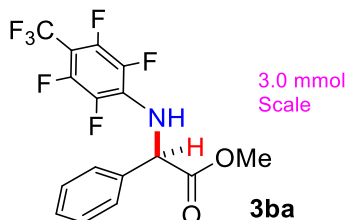
Ethyl (R)-2-(4-methoxyphenyl)-2-((2,3,5,6-tetrafluorophenyl)amino)acetate 3ai was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 34%. $[\alpha]_D^{20} = -110^\circ$ ($c = 0.5$, CHCl_3). ^1H NMR (500 MHz, CDCl_3): δ 7.29 (d, $J = 8.5$ Hz, 2H), 6.86 (d, $J = 8.0$ Hz, 2H), 6.39 (m, 1H), 5.36 (d, $J = 9.0$ Hz, 1H), 5.08 (d, $J = 8.8$ Hz, 1H), 4.25 (dq, $J =$

10.8, 7.1 Hz, 1H), 4.14 (dq, $J = 10.8, 7.1$ Hz, 1H), 3.78 (s, 3H), 1.21 (t, $J = 7.0$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3): δ 171.54, 159.76, 146.32 (dm, $J = 243.0$ Hz), 137.70 (dm, $J = 238.6$ Hz), 129.19, 128.21, 126.28 (m), 114.32, 94.68 (t, $J = 23.1$ Hz), 61.97, 60.59 (t, $J = 4.1$ Hz), 55.24, 13.98. ^{19}F NMR (376 MHz, CFCl_3 , CDCl_3): δ -141.45 (m, 2F), -158.70 (m, 2F). IR (neat, cm^{-1}): 3384, 2937, 1735, 1651, 1610, 1510, 1459, 1248, 1176, 1031, 920, 797, 711. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{17}\text{H}_{14}\text{F}_4\text{NO}_3$: 356.0910, Found: 356.0918. HPLC analysis: ee = 91%. Whelk (99% hexane: 1% isopropanol, 1 ml/min): $t_{\text{major}} = 7.23$ min, $t_{\text{minor}} = 8.94$ min.

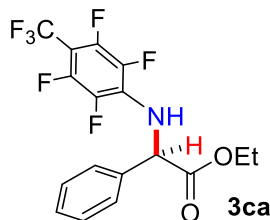


Ethyl (R)-2-(4-methoxyphenyl)-2-((2,4,6-trifluorophenyl)amino)acetate 3aj was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 30%. $[\alpha]_D^{20} = -76^\circ$ ($c = 0.5$, CHCl_3). ^1H NMR (500 MHz, CDCl_3): δ 7.28 (d, $J = 7.8$ Hz, 2H), 6.84 (d, $J = 7.5$ Hz, 2H), 6.55 (t, $J = 7.8$ Hz, 2H), 5.23 (d, $J = 8.8$ Hz, 1H), 4.64 (d, $J = 9.4$ Hz, 1H), 4.17 (m, 2H), 3.77 (s, 3H), 1.20 (t, $J = 7.0$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3): δ 172.01, 159.58, 156.15 – 154.11 (m, 2C), 129.53, 128.24, 120.23 (m), 114.14, 100.18 (m), 61.66, 61.44 (t, $J = 3.8$ Hz), 55.20, 14.00. ^{19}F NMR (376 MHz, CFCl_3 , CDCl_3): δ -120.48 (t, $J = 8.6$ Hz, 1F), -124.21 (m, 2F). IR (neat, cm^{-1}): 3457, 2962, 1735, 1609, 1508, 1442, 1247, 1175, 1114, 1032, 995, 833, 571. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{17}\text{H}_{15}\text{F}_3\text{NO}_3$: 338.1004, Found: 338.1000. HPLC analysis: ee = 49%. OJH (99% hexane: 1% isopropanol, 1 ml/min): $t_{\text{major}} = 12.51$ min, $t_{\text{minor}} = 18.37$ min.

5.2. Enantioselective C–H Amination of Various Arylacetate Esters with 2a

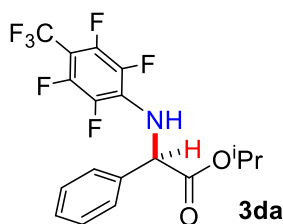


Methyl (*R*)-2-phenyl-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)acetate 3ba was obtained through the general procedure on 3 mmol scale using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 75%. $[\alpha]_D^{20} = -97^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (500 MHz, CDCl_3): δ 7.36 (m, 5H), 5.49 (m, 2 H), 3.76 (s, 3H). $^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 171.29, 144.86 (dm, $J = 254.75$ Hz), 137.03 (dm, $J = 239.38$ Hz), 136.50, 129.20, 129.07, 126.93, 121.41 (q, $J = 271.1$ Hz), 97.60 (m), 60.54 (t, $J = 4.1$ Hz), 53.21. $^{19}\text{F NMR}$ (376 MHz, CFCl_3 , CDCl_3): δ -55.60 (t, $J = 20.7$ Hz, 3F), -143.31 (m, 2F), -158.98 (m, 2F). IR (neat, cm^{-1}): 3390, 2959, 1742, 1656, 1508, 1331, 1293, 1236, 1177, 1129, 1069, 967, 875, 698. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{16}\text{H}_9\text{F}_7\text{NO}_2$: 380.0522, Found: 380.0527. HPLC analysis: ee = 96%. ODH (99% hexane: 1% isopropanol, 1 ml/min): $t_{\text{major}} = 5.99$ min, $t_{\text{minor}} = 7.16$ min.

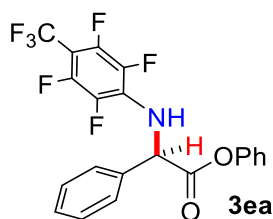


Ethyl (*R*)-2-phenyl-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)acetate 3ca was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 82%. $[\alpha]_D^{20} = -84^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (500 MHz, CDCl_3): δ 7.38 – 7.34 (m, 5H), 5.48 (m, 2H), 4.27 (m, 2H), 1.22 (t, $J = 7.0$ Hz, 3H). $^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 170.77, 144.84 (dm, $J = 257.5$ Hz), 136.96 (dm, $J = 241.3$ Hz), 136.65, 129.12, 128.96, 126.88, 121.42 (q, $J = 269.9$ Hz), 62.43, 60.61 (t, $J = 4$ Hz), 13.92. $^{19}\text{F NMR}$ (376 MHz, CFCl_3 , CDCl_3): δ -55.58 (t, $J = 20.7$ Hz, 3F), -143.38 (m, 2F), -159.05 (m, 2F). IR (neat, cm^{-1}): 3396, 2973, 1736, 1505, 1291, 1134, 954, 696. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{17}\text{H}_{11}\text{F}_7\text{NO}_2$: 394.0678, Found:

394.0687. HPLC analysis: ee = 95%. ODH (97% hexane: 3% isopropanol, 1 ml/min): t_{major} = 8.38 min, t_{minor} = 9.16 min.

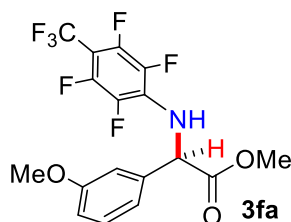


Isopropyl (*R*)-2-phenyl-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)acetate 3da was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 61%. $[\alpha]_D^{20} = -199^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (500 MHz, CDCl_3): δ 7.40 – 7.31 (m, 5H), 5.52 – 5.40 (m, 2H), 5.06 (p, $J = 6.2$ Hz, 1H), 1.29 (d, $J = 6.3$ Hz, 3H), 1.06 (d, $J = 6.2$ Hz, 3H). $^{13}\text{C NMR}$ (62.5 MHz, CDCl_3): δ 170.26, 144.86 (dm, $J = 253.9$ Hz), 136.95 (dm, $J = 240.7$ Hz), 136.76, 129.27 (m), 129.04, 128.85, 126.80, 121.44 (q, $J = 271.3$ Hz), 97.30 (m), 70.34, 60.72, 21.60, 21.15. $^{19}\text{F NMR}$ (376 MHz, CFCl_3 , CDCl_3): -55.56 (t, $J = 21.1$ Hz, 3F), -143.46 (m, 2F), -159.13 (m, 2F). IR (neat, cm^{-1}): 3391, 2990, 1733, 1652, 1506, 1330, 1291, 1139, 1099, 957, 878, 710. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{18}\text{H}_{13}\text{F}_7\text{NO}_2$: 408.0835, Found: 408.0842. HPLC analysis: ee = 97%. Whelk (100% hexane, 1 ml/min): t_{major} = 8.14 min, t_{minor} = 8.74 min.

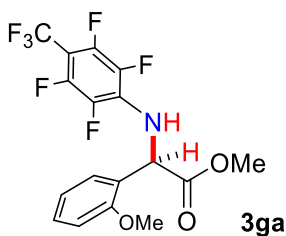


Phenyl (*R*)-2-phenyl-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)acetate 3ea was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 50%. $[\alpha]_D^{20} = -92^\circ$ ($c = 0.5$, CHCl_3). $^1\text{H NMR}$ (500 MHz, CDCl_3): δ 7.52 – 7.40 (m, 5H), 7.37 – 7.33 (m, 2H), 7.24 (t, $J = 7.5$ Hz, 1H), 6.99 – 6.93 (m, 2H), 5.74 (d, $J = 8.0$ Hz, 1H), 5.39 (d, $J = 8.0$ Hz, 1H). $^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 169.63, 150.20, 144.83 (dm, $J = 240.9$ Hz), 137.03 (dm, $J = 240.5$ Hz), 135.99, 129.55, 129.44, 129.41, 129.01 (m), 127.11, 126.46, 121.37 (q, $J = 272.9$ Hz), 120.90, 97.89 (m), 60.88. $^{19}\text{F NMR}$ (376 MHz, CDCl_3): δ -55.12 (t, $J =$

20.9 Hz, 3F), -142.60 (m, 2F), -158.48 (m, 2F). IR (neat, cm^{-1}): 3394, 2924, 1759, 1656, 1509, 1332, 1236, 1190, 1130, 979, 958, 880, 713, 687. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{21}\text{H}_{11}\text{F}_7\text{NO}_2$: 442.0678, Found: 442.0683. HPLC analysis: ee = 94%. ADH (99% hexane: 1% isopropanol, 0.7 ml/min): $t_{\text{major}} = 17.11$ min, $t_{\text{minor}} = 10.23$ min.

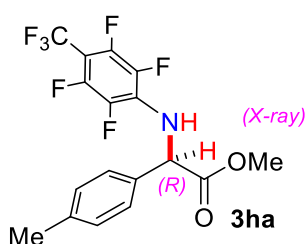


Methyl (R)-2-(3-methoxyphenyl)-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)acetate 3fa was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 61%. $[\alpha]_D^{20} = -93^\circ$ ($c = 1.0$, CHCl_3). ^1H NMR (500 MHz, CDCl_3): δ 7.29 (t, $J = 7.5$ Hz, 1H), 6.95 (d, $J = 7.5$ Hz, 1H), 6.89 (m, 2H), 5.45 (m, 2H), 3.81 (s, 3H), 3.77 (s, 3H). ^{13}C NMR (125 MHz, CDCl_3): δ 171.18, 160.09, 144.91 (dm, $J = 250.8$ Hz), 137.93, 136.94 (dm, $J = 223.4$ Hz), 130.26, 129.04 (m), 121.39 (q, $J = 272.6$ Hz), 119.09, 114.28, 112.79, 97.62 (m), 60.47 (t, $J = 4.1$ Hz), 55.30, 53.24. ^{19}F NMR (376 MHz, CFCl_3 , CDCl_3): δ -55.60 (t, $J = 20.7$ Hz, 3F), -143.29 (m, 2F), -158.94 (m, 2F). IR (neat, cm^{-1}): 3390, 2959, 1743, 1657, 1508, 1330, 1235, 1129, 970, 714. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{17}\text{H}_{11}\text{F}_7\text{NO}_3$: 410.0627, Found: 410.0634. HPLC analysis: ee = 93%. ADH (99% hexane: 1% isopropanol, 1 ml/min): $t_{\text{major}} = 6.40$ min, $t_{\text{minor}} = 7.14$ min.

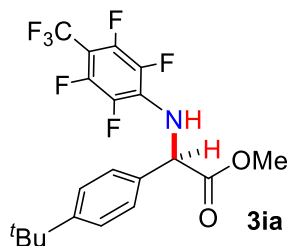


Methyl (R)-2-(2-methoxyphenyl)-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)acetate 3ga was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 52%. $[\alpha]_D^{20} = -152^\circ$ ($c = 0.6$, CHCl_3). ^1H NMR (500 MHz, CDCl_3): δ 7.34 (dt, $J = 2.0, 7.8$ Hz, 1H), 7.28 (d, $J = 7.5$ Hz, 1H), 6.97 (dt, $J = 0.5, 7.5$ Hz, 1H),

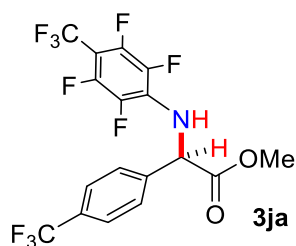
6.93 (d, $J = 8.0$ Hz, 1H), 5.69 (d, $J = 9.5$ Hz, 1H), 5.46 (d, $J = 9.0$ Hz, 1H), 3.87 (s, 3H), 3.73 (s, 3H). ^{13}C NMR (125 MHz, CDCl_3): δ 171.45, 156.99, 144.80 (dm, $J = 255.4$ Hz), 137.18 (dm, $J = 239.1$ Hz), 130.37, 130.02 (m), 129.57, 124.92, 121.48 (q, $J = 271.4$ Hz), 121.12, 111.38, 97.42 (m), 57.73 (t, $J = 4.5$ Hz), 55.62, 52.93. ^{19}F NMR (376 MHz, CFCl_3 , CDCl_3): δ -55.55 (t, $J = 20.9$ Hz, 3F), -143.77 (m, 2F), -159.09 (m, 2F). IR (neat, cm^{-1}): 3389, 2956, 2844, 1746, 1655, 1509, 1331, 1236, 1128, 967, 874, 754, 713. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{17}\text{H}_{11}\text{F}_7\text{NO}_3$: 410.0627, Found: 410.0622. HPLC analysis: ee = 95%. Whelk (99.5% hexane: 0.5% isopropanol, 1 ml/min): $t_{\text{major}} = 8.27$ min, $t_{\text{minor}} = 11.14$ min.



Methyl (R)-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)-2-(p-tolyl)acetate 3ha was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 70%. $[\alpha]_D^{20} = -88^\circ$ ($c = 0.5$, CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ 7.25 (d, $J = 8.4$ Hz, 2H), 7.17 (d, $J = 8.0$ Hz, 2H), 5.45 (m, 2H), 3.75 (s, 3H), 2.34 (s, 3H). ^{13}C NMR (125 MHz, CDCl_3): δ 171.46, 144.75 (dm, $J = 258.8$ Hz), 139.06, 136.89 (dm, $J = 237.8$ Hz), 133.51, 129.88, 129.27 (m), 126.82, 121.42 (q, $J = 269.8$ Hz), 97.35 (m), 60.29 (t, $J = 4.0$ Hz), 53.15, 21.13. ^{19}F NMR (376 MHz, CFCl_3 , CDCl_3): δ -55.59 (t, $J = 20.7$ Hz, 3F), -143.39 (m, 2F), -158.97 (m, 2F). IR (neat, cm^{-1}): 3374, 2957, 2924, 1738, 1656, 1505, 1456, 1291, 1176, 1139, 1010, 968, 874, 711, 619. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{17}\text{H}_{11}\text{F}_7\text{NO}_2$: 394.0678, Found: 394.0692. HPLC analysis: ee = 94%. OJH (99% hexane: 1% isopropanol, 1 ml/min): $t_{\text{major}} = 8.52$ min, $t_{\text{minor}} = 5.25$ min.

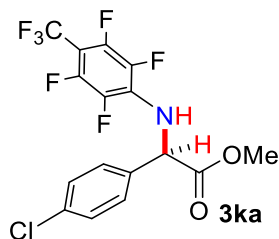


Methyl (*R*)-2-(4-(*tert*-butyl)phenyl)-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)acetate **3ia** was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 57%. $[\alpha]_D^{20} = -76^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (500 MHz, CDCl_3): δ 7.39 (d, $J = 8.5$ Hz, 2H), 7.29 (d, $J = 8.5$ Hz, 2H), 5.49 (d, $J = 8.0$ Hz, 1H), 5.37 (d, $J = 8.0$ Hz, 1H), 3.76 (s, 3H), 1.32 (s, 9H). $^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 171.46, 152.16, 144.88 (dm, $J = 256.2$ Hz), 136.99 (dm, $J = 239.3$ Hz), 133.36, 129.26 (m), 126.58, 126.19, 121.46 (q, $J = 271.0$ Hz), 97.22 (m), 60.27 (t, $J = 4.1$ Hz), 53.09, 34.64, 31.19. $^{19}\text{F NMR}$ (376 MHz, CFCl_3 , CDCl_3): δ -55.55 (t, $J = 21.1$ Hz, 3F), -143.42 (m, 2F), -159.16 (m, 2F). IR (neat, cm^{-1}): 3408, 2974, 1738, 1656, 1546, 1505, 1462, 1328, 1133, 1082, 1008, 962, 875, 712, 667. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{20}\text{H}_{17}\text{F}_7\text{NO}_2$: 436.1147, Found: 436.1165. HPLC analysis: ee = 88%. Whelk (100% hexane, 0.8 ml/min): $t_{\text{major}} = 10.65$ min, $t_{\text{minor}} = 11.36$ min.

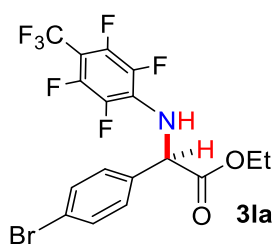


Methyl (*R*)-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)-2-(4-(trifluoromethyl)phenyl)acetate **3ja** was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 41%. $[\alpha]_D^{20} = -65^\circ$ ($c = 0.7$, CHCl_3). $^1\text{H NMR}$ (500 MHz, CDCl_3): δ 7.64 (d, $J = 8.0$ Hz, 2H), 7.51 (d, $J = 8.0$ Hz, 2H), 5.67 – 5.49 (m, 2H), 3.79 (s, 3H). $^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 170.55, 144.79 (dm, $J = 255.0$ Hz), 140.50, 136.85 (tdd, $J = 4.9, 15.2, 239.4$ Hz), 131.28 (q, $J = 32.6$ Hz), 128.57 (m), 127.34, 126.19 (q, $J = 3.8$ Hz), 123.73 (q, $J = 270.6$ Hz), 121.31 (q, $J = 273.5$ Hz), 98.05 (m), 60.05, 53.56. $^{19}\text{F NMR}$ (376 MHz, CFCl_3 , CDCl_3): δ -55.66 (t, $J = 20.9$ Hz, 3F), -63.32 (s, 3F), -142.80 (m, 2F), -159.00 (m, 2F). IR (neat, cm^{-1}): 3392, 2963,

1745, 1658, 1509, 1322, 1124, 1067, 968, 876, 713. HRMS (ESI-negative) ($[M-H]^-$) Calcd. for: $C_{17}H_8F_{10}NO_2$: 448.0395, Found: 448.0407. HPLC analysis: ee = 92%. ADH (99% hexane: 1% isopropanol, 1 ml/min): $t_{major} = 5.94$ min, $t_{minor} = 5.67$ min.

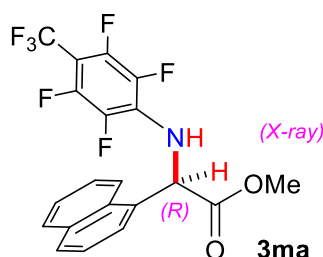


Methyl (R)-2-(4-chlorophenyl)-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl) amino) acetate 3ka was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 60%. $[\alpha]_D^{20} = -125^\circ$ ($c = 0.8$, $CHCl_3$). 1H NMR (500 MHz, $CDCl_3$): δ 7.36–7.30 (m, 4H), 5.53 (br, 1H), 5.47 (d, $J = 8.0$ Hz, 1H), 3.77 (s, 3H). ^{13}C NMR (125 MHz, $CDCl_3$): δ 170.89, 144.83 (dm, $J = 254.6$ Hz), 136.90 (dm, $J = 240.6$ Hz), 135.04, 129.41, 128.68 (m), 128.25, 121.32 (q, $J = 272.7$ Hz), 97.96 (m), 59.79 (t, $J = 4.1$ Hz), 53.41. ^{19}F NMR (376 MHz, $CFCl_3$, $CDCl_3$): δ -55.64 (t, $J = 20.7$ Hz, 3F), -143.00 (m, 2F), -158.92 (m, 2F). IR (neat, cm^{-1}): 3390, 2959, 1743, 1656, 1509, 1332, 1131, 968, 876, 770, 712. HRMS (ESI-negative) ($[M-H]^-$) Calcd. for: $C_{16}H_8ClF_7NO_2$: 414.0132, Found: 414.0145. HPLC analysis: ee = 93%. OJH (99% hexane: 1% isopropanol, 1 ml/min): $t_{major} = 12.07$ min, $t_{minor} = 7.60$ min.

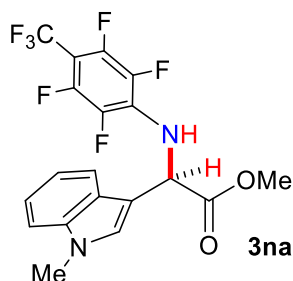


Ethyl (R)-2-(4-bromophenyl)-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)acetate 3la was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 67%. $[\alpha]_D^{20} = -178^\circ$ ($c = 0.5$, $CHCl_3$). 1H NMR (600 MHz, $CDCl_3$): δ 7.50 (d, $J = 8.0$ Hz, 2H), 7.25 (d, $J = 8.0$ Hz, 2H), 5.53 (br, 1H), 5.42 (d, $J = 8.0$ Hz, 1H), 4.22 (m, 2H), 1.22 (t, $J = 7.0$ Hz, 3H). ^{13}C NMR (125 MHz, $CDCl_3$): δ 170.30, 144.80 (dm, $J = 243.9$ Hz), 136.77 (dm, $J = 248.0$ Hz), 132.31, 128.80 (m), 128.51, 123.11, 121.33 (q, $J =$

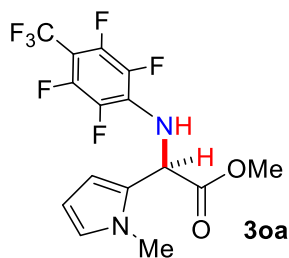
265.1 Hz), 97.82 (m), 62.72, 59.98 (t, $J = 4.0$ Hz), 13.92. ^{19}F NMR (376 MHz, CFCl_3 , CDCl_3): δ -55.62 (t, $J = 21.1$ Hz, 3F), -143.05 (m, 2F), -158.99 (m, 2F). IR (neat, cm^{-1}): 3391, 2990, 1733, 1652, 1506, 1456, 1330, 1291, 1139, 1099, 957, 878, 710, 630. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{17}\text{H}_{10}\text{BrF}_7\text{NO}_2$: 471.9783, Found: 471.9796. HPLC analysis: ee = 92%. Whelk (99.5% hexane: 0.5% isopropanol, 1 ml/min): $t_{\text{major}} = 6.48$ min, $t_{\text{minor}} = 6.99$ min.



Methyl (R)-2-(naphthalen-1-yl)-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)acetate 3ma was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 68%. $[\alpha]_D^{20} = -68^\circ$ ($c = 1.0$, CHCl_3). ^1H NMR (500 MHz, CDCl_3): δ 8.20 (d, $J = 8.5$ Hz, 1H), 7.92 (d, $J = 8.0$ Hz, 1H), 7.90 – 7.88 (m, 1H), 7.63 – 7.56 (m, 2H), 7.48 (m, 2H), 6.31 (d, $J = 8.0$ Hz, 1H), 5.36 (br, 1H), 3.75 (s, 3H). ^{13}C NMR (125 MHz, CDCl_3): δ 171.75, 144.82 (dm, $J = 255.1$ Hz), 136.94 (dm, $J = 241.0$ Hz), 134.19, 132.20, 130.84, 129.98, 129.28 (m), 129.12, 127.20, 126.30, 125.33, 125.15, 122.82, 121.43 (q, $J = 272.5$ Hz), 97.46 (m), 57.56, 53.19. ^{19}F NMR (376 MHz, CFCl_3 , CDCl_3): δ -55.61 (t, $J = 20.7$ Hz, 3F), -143.22 (m, 2F), -159.29 (m, 2F). IR (neat, cm^{-1}): 3396, 2967, 1748, 1660, 1510, 1331, 1285, 1121, 998, 961, 792, 775, 714. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{20}\text{H}_{11}\text{F}_7\text{NO}_2$: 430.0678, Found: 430.0687. HPLC analysis: ee = 88%. Whelk (99.5% hexane: 0.5% isopropanol, 1 ml/min): $t_{\text{major}} = 8.91$ min, $t_{\text{minor}} = 10.80$ min.

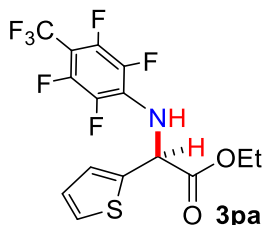


Methyl (R)-2-(1-methyl-1H-indol-2-yl)-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)acetate 3na was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 90%. $[\alpha]_D^{20} = -53^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (500 MHz, CDCl_3): δ 7.70 (d, $J = 8.0$ Hz, 1H), 7.34 (d, $J = 8.0$ Hz, 1H), 7.29 (t, $J = 7.5$ Hz, 1H), 7.19 (t, $J = 7.5$ Hz, 1H), 7.15 (s, 1H), 5.80 (d, $J = 8.0$ Hz, 1H), 5.35 (br, 1H), 3.79 (s, 3H), 3.76 (s, 3H). $^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 171.74, 144.76 (dm, $J = 244.3$ Hz), 137.02 (dm, $J = 240.6$ Hz), 137.19, 129.60 (m), 128.28, 125.36, 122.48, 121.45 (q, $J = 271.0$ Hz), 120.25, 118.87, 109.78, 109.43, 97.01 (m), 54.25 (t, $J = 4.5$ Hz), 53.00, 32.95. $^{19}\text{F NMR}$ (376 MHz, CFCl_3 , CDCl_3): δ -55.06 (t, $J = 21.1$ Hz, 3F), -143.12 (m, 2F), -158.69 (m, 2F). IR (neat, cm^{-1}): 3367, 2954, 1732, 1655, 1508, 1330, 1237, 1125, 943, 876, 742, 713. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{19}\text{H}_{12}\text{F}_7\text{N}_2\text{O}_2$: 433.0787, Found: 433.0792. HPLC analysis: ee = 93%. ADH (99% hexane: 1% isopropanol, 1 ml/min): $t_{\text{major}} = 9.05$ min, $t_{\text{minor}} = 15.23$ min.



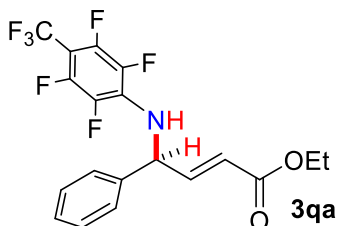
Methyl (R)-2-(1-methyl-1H-pyrrol-2-yl)-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)acetate 3oa was obtained through the general procedure using 1.5 equiv of azide with α,α,α -trifluorotoluene as solvent. Yield: 75%. $[\alpha]_D^{20} = -106^\circ$ ($c = 0.5$, CHCl_3). $^1\text{H NMR}$ (500 MHz, CDCl_3): δ 6.65 (t, $J = 2.0$ Hz, 1H), 6.08 (m, 2H), 5.59 (d, $J = 9.0$ Hz, 1H), 4.97 (m, 1H), 3.79 (s, 3H), 3.71 (s, 3H). $^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 170.55, 144.87 (dm, $J = 254.3$ Hz), 137.19 (dm, $J = 240.5$ Hz), 129.30 (m), 126.63, 124.30, 121.38 (q, $J = 271.9$ Hz), 108.14, 107.53, 98.04 (m),

53.72 (t, $J = 4.8$ Hz), 53.13, 33.92. ^{19}F NMR (376 MHz, CFCl_3 , CDCl_3): δ -55.63 (t, $J = 20.7$ Hz, 3F), -143.12 (m, 2F), -158.84 (m, 2F). IR (neat, cm^{-1}): 3389, 2958, 1746, 1655, 1509, 1332, 1232, 1129, 1080, 965, 876, 713. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{15}\text{H}_{10}\text{F}_7\text{N}_2\text{O}_2$: 383.0630, Found: 383.0637. HPLC analysis: ee = 98%. ADH (99% hexanes: 1% isopropanol, 1 ml/min): $t_{\text{major}} = 5.91$ min, $t_{\text{minor}} = 6.54$ min.

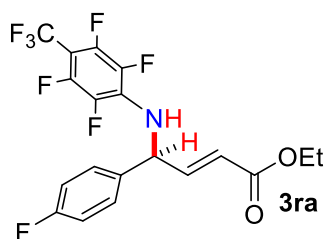


Methyl (S)-2-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)-2-(thiophen-2-yl)acetate 3pa was obtained through the general procedure using 1.5 equiv of azide with PhCF_3 as solvent. Yield: 88%. $[\alpha]_D^{20} = -120^\circ$ ($c = 0.2$, CHCl_3). ^1H NMR (500 MHz, CDCl_3): δ 7.30 (dd, $J = 1.0, 5.0$ Hz, 1H), 7.10 (d, $J = 3.5$ Hz, 1H), 6.98 (dd, $J = 3.5, 5.0$ Hz, 1H), 5.74 (d, $J = 8.5$ Hz, 1H), 5.38 (d, $J = 8.0$ Hz, 1H), 4.28 (m, 2H), 1.28 (t, $J = 7.5$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3): δ 169.85, 144.86 (dm, $J = 254.1$ Hz), 139.18, 137.18 (dm, $J = 240.9$ Hz), 128.91 (m), 127.24, 126.46, 126.38, 121.38 (q, $J = 272.1$ Hz), 97.98 (m), 62.78, 56.44 (t, $J = 4.6$ Hz), 13.97. ^{19}F NMR (376 MHz, CFCl_3 , CDCl_3): δ -55.16 (t, $J = 21.1$ Hz, 3F), -142.63 (m, 2F), -157.89 (m, 2F). IR (neat, cm^{-1}): 3384, 2984, 1740, 1655, 1508, 1334, 1231, 1132, 978, 713. HRMS (ESI-negative) ($[\text{M}-\text{H}]^-$) Calcd. for: $\text{C}_{15}\text{H}_9\text{F}_7\text{NO}_2\text{S}$: 400.0242, Found: 400.0252. HPLC analysis: ee = 91%. ODH (99% hexane: 1% isopropanol, 0.7 ml/min): $t_{\text{major}} = 8.04$ min, $t_{\text{minor}} = 11.78$ min.

5.3. Enantioselective C–H Amination of Various Arylcrotonate Esters with 2a

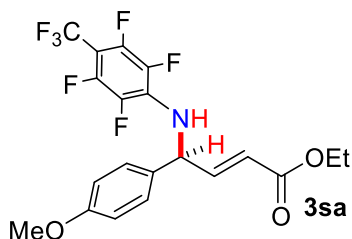


Ethyl (*S,E*)-4-phenyl-4-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)but-2-enoate 3qa was obtained through the general procedure using 1.5 equiv of azide with benzene as solvent. Yield: 64%. $[\alpha]_D^{20} = -55^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 7.41 – 7.39 (m, 2H), 7.36 (m, 1H), 7.31–7.29 (m, 2H), 7.10 (dd, $J = 15.6, 5.6$ Hz, 1H), 5.99 (dd, $J = 15.7, 1.5$ Hz, 1H), 5.60 (dd, $J = 8.4, 5.9$ Hz, 1H), 4.49 (d, $J = 8.6$ Hz, 1H), 4.20 (qd, $J = 7.2, 1.3$ Hz, 2H), 1.29 (t, $J = 7.1$ Hz, 3H). $^{13}\text{C NMR}$ (150 MHz, CDCl_3) δ 165.64, 146.03, 144.66 (dm, $J = 264.9$ Hz), 138.64, 136.48 (dm, $J = 241.3$ Hz), 129.38 (m), 129.29, 128.78, 126.89, 122.60, 121.30 (q, $J = 272.9$ Hz), 97.65 (m), 60.77, 59.42 (t, $J = 4.6$ Hz), 14.08. $^{19}\text{F NMR}$ (470 MHz, CDCl_3) δ -55.10 (t, $J = 20.8$ Hz, 3F), -142.46 (m, 2F), -158.06 (m, 2F). IR (neat, cm^{-1}): 3339, 2986, 1715, 1657, 1512, 1334, 1234, 1134, 974, 713. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{19}\text{H}_{15}\text{F}_7\text{NO}_2$: 422.09855, Found: 422.09960. HPLC analysis: ee = 94%. ODH (95% hexane: 5% isopropanol, 0.8 ml/min): $t_{\text{major}} = 13.13$ min, $t_{\text{minor}} = 17.29$ min.

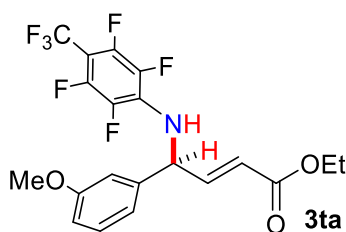


Ethyl (*S,E*)-4-(4-fluorophenyl)-4-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)but-2-enoate 3ra was obtained through the general procedure using 1.5 equiv of azide with benzene as solvent. Yield: 53%. $[\alpha]_D^{20} = -69^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 7.28 – 7.30 (m, 2H), 7.12 – 7.05 (m, 3H), 5.98 (dd, $J = 15.8, 1.4$ Hz, 1H), 5.60 (m, 1H), 4.45 (d, $J = 8.7$ Hz, 1H), 4.21 (qd, $J = 7.3, 1.2$ Hz, 2H), 1.30 (t, $J = 7.1$ Hz, 3H). $^{13}\text{C NMR}$ (150 MHz, CDCl_3) δ 165.53, 162.69 (d, $J = 248.5$ Hz), 145.65, 144.81 (dm, $J = 255.2$ Hz), 136.79 (dm, $J = 241.4$ Hz), 134.49

(d, $J = 3.4$ Hz), 129.20 (m), 128.75, 128.69, 122.90, 121.25 (q, $J = 272.6$ Hz), 116.35, 116.20, 97.90 (m), 60.85, 58.72 (t, $J = 4.6$ Hz), 14.07. ^{19}F NMR (564 MHz, CDCl_3) δ -55.12 (t, $J = 20.9$ Hz, 3F), -112.42 (m, 1F), -142.28 (m, 2F), -157.92 (m, 2F). IR (neat, cm^{-1}): 3339, 2987, 1715, 1657, 1510, 1335, 1232, 1135, 975, 714. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{19}\text{H}_{14}\text{F}_8\text{NO}_2$: 440.08913, Found: 440.08805. HPLC analysis: ee = 97%. ODH (95% hexane: 5% isopropanol, 0.8 ml/min): $t_{\text{major}} = 12.78$ min, $t_{\text{minor}} = 16.42$ min.

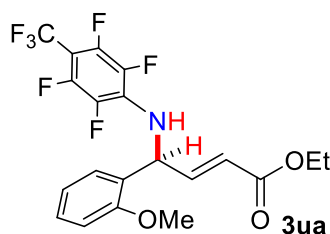


Ethyl (S,E)-4-(4-methoxyphenyl)-4-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)but-2-enoate 3sa was obtained through the general procedure using 1.5 equiv of azide with benzene as solvent. Yield: 70%. $[\alpha]_D^{20} = -75^\circ$ ($c = 1.0$, CHCl_3). ^1H NMR (500 MHz, CDCl_3) δ 7.22 (d, $J = 8.4$ Hz, 2H), 7.10 (dd, $J = 15.6, 5.5$ Hz, 1H), 6.92 (d, $J = 8.4$ Hz, 2H), 5.98 (dd, $J = 15.6, 1$ Hz), 5.56 (m, 1H), 4.43 (br, 1H), 4.21 (q, $J = 7.1$ Hz, 2H), 3.81 (s, 3H), 1.29 (t, $J = 7.0$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 165.82, 159.06, 146.40, 145.10 (dm, $J = 252.1$ Hz), 136.59 (dm, $J = 245.95$ Hz), 135.62, 129.59 (m), 126.74, 126.32, 121.40 (q, $J = 272.6$ Hz), 114.68, 97.90, 60.80, 58.95 (q, $J = 4.7$ Hz), 55.33, 14.16. ^{19}F NMR (470 MHz, CDCl_3) δ -55.08 (t, $J = 20.8$ Hz, 3F), -142.58 (m, 2F), -158.12 (m, 2F). IR (neat, cm^{-1}): 3341, 1716, 1656, 1511, 1334, 1234, 1177, 1134, 974, 713. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{20}\text{H}_{17}\text{F}_7\text{NO}_3$: 452.10912, Found: 452.11023. HPLC analysis: ee = 95%. ODH (95% hexane: 5% isopropanol, 0.8 ml/min): $t_{\text{major}} = 16.13$ min, $t_{\text{minor}} = 18.83$ min.



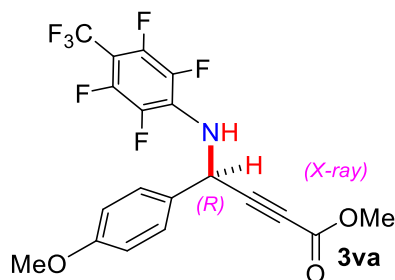
Ethyl (S,E)-4-(3-methoxyphenyl)-4-((2,3,5,6-tetrafluoro-4-trifluoromethyl)phenyl) amino)but-2-enoate 3ta was obtained through the general procedure using 1.5 equiv of azide with

benzene as solvent. Yield: 63%. $[\alpha]_D^{20} = -83^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 7.32 (t, $J = 8.0$ Hz, 1H), 7.09 (dd, $J = 15.6, 5.6$ Hz, 1H), 6.89 (d, $J = 8.1$ Hz, 2H), 6.83 (s, 1H), 6.00 (d, $J = 15.8$ Hz, 1H), 5.57 (dd, $J = 5.4, 9.0$ Hz, 1H), 4.50 (d, $J = 9.1$ Hz, 1H), 4.21 (q, $J = 7.0$ Hz, 2H), 3.82 (s, 3H), 1.29 (t, $J = 7.1$ Hz, 3H). $^{13}\text{C NMR}$ (150 MHz, CDCl_3) δ 165.72, 160.25, 146.03, 144.92 (dm, $J = 257.9$ Hz), 140.27, 136.84 (dm, $J = 241.5$), 130.51, 122.64, 121.39 (q, $J = 272.5$ Hz), 119.01, 113.82, 112.99, 97.67 (m), 60.85, 59.47, 55.24, 14.14. $^{19}\text{F NMR}$ (564 MHz, CDCl_3) δ -55.12 (t, $J = 20.8$ Hz, 3F), -142.51 (m, 2F), -158.06 (m, 2F). IR (neat, cm^{-1}): 3342, 2940, 1715, 1657, 1512, 1334, 1234, 1134, 713. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{20}\text{H}_{17}\text{F}_7\text{NO}_3$: 452.10912, Found: 452.10866. HPLC analysis: ee = 95%. ODH (95% hexane: 5% isopropanol, 0.8 ml/min): $t_{\text{major}} = 16.19$ min, $t_{\text{minor}} = 20.91$ min.

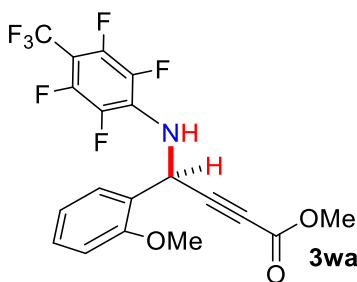


Ethyl (S,E)-4-(2-methoxyphenyl)-4-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)but-2-enoate 3ua was obtained through the general procedure using 1.5 equiv of azide with benzene as solvent. Yield: 70%. $[\alpha]_D^{20} = -97^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 7.33 (m, 1H), 7.19 (d, $J = 7.5$ Hz, 1H), 7.15 (dd, $J = 15.7, 5.5$ Hz, 1H), 7.01 – 6.90 (m, 2H), 5.97 (d, $J = 16.1$ Hz, 1H), 5.71 (dd, $J = 9.5, 5.4$ Hz, 1H), 5.29 (d, $J = 9.1$ Hz, 1H), 4.19 (q, $J = 7.1$ Hz, 2H), 3.87 (s, 3H), 1.28 (t, $J = 7.2$ Hz, 3H). $^{13}\text{C NMR}$ (150 MHz, CDCl_3) δ 166.14, 156.96, 146.38, 144.59 (dm, $J = 247.0$ Hz), 137.08 (dm, $J = 241.1$ Hz), 130.18 (m), 130.08, 128.80, 126.54, 121.85, 121.51 (q, $J = 272.4$ Hz), 121.25, 111.48, 97.10 (m), 60.70, 57.68 (t, $J = 5.1$ Hz), 55.50, 14.21. $^{19}\text{F NMR}$ (564 MHz, CDCl_3) δ -55.06 (t, $J = 20.8$ Hz, 3F), -143.07 (m, 2F), -158.58 (m, 2F). IR (neat, cm^{-1}): 3369, 2982, 1718, 1656, 1512, 1332, 1234, 1132, 876, 713. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{20}\text{H}_{17}\text{F}_7\text{NO}_3$: 452.10912, Found: 452.10854. HPLC analysis: ee = 95%. ODH (95% hexane: 5% isopropanol, 0.8 ml/min): $t_{\text{major}} = 11.60$ min, $t_{\text{minor}} = 12.89$ min.

5.4. Enantioselective C–H Amination of Various Aryltetrolate Esters with 2a

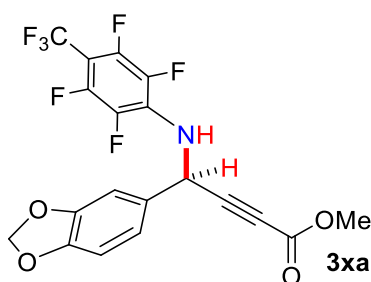


Methyl (R)-4-(4-methoxyphenyl)-4-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)but-2-ynoate 3va was obtained through the general procedure using 1.2 equiv of azide with benzene as solvent. Yield: 83%. $[\alpha]_D^{20} = 35^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 7.46 (d, $J = 8.8$ Hz, 2H), 6.95 (d, $J = 8.7$ Hz, 2H), 5.79 (d, $J = 9.2$ Hz, 1H), 4.43 (d, $J = 9.2$ Hz, 1H), 3.83 (s, 3H), 3.79 (s, 3H). $^{13}\text{C NMR}$ (150 MHz, CDCl_3) δ 163.02, 156.01, 147.47 (dm, $J = 241.2$ Hz), 140.04 (dm, $J = 239.0$ Hz), 131.42 (m), 131.16, 130.84, 123.95 (q, $J = 271.4$ Hz), 117.26, 101.53 (m), 87.02, 80.20, 58.08, 55.51, 53.03, 32.34. $^{19}\text{F NMR}$ (564 MHz, CDCl_3) δ -55.23 (t, $J = 21.3$ Hz, 3F), -142.12 (m, 2F), -156.66 (m, 2F). IR (neat, cm^{-1}): 3344, 2957, 2845, 2240, 1721, 1595, 1512, 1336, 1229, 1033, 990, 715. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{19}\text{H}_{13}\text{F}_7\text{NO}_3$: 436.07782, Found: 436.07846. HPLC analysis: ee = 93%. ASH (90% hexanes: 10% isopropanol, 0.8 ml/min): $t_{\text{major}} = 11.99$ min, $t_{\text{minor}} = 14.42$ min.

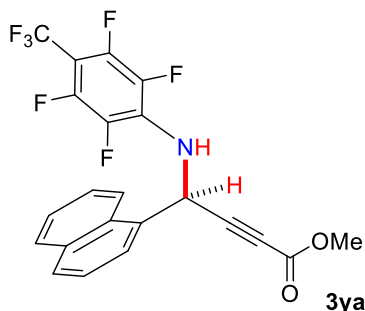


Methyl (S)-4-(2-methoxyphenyl)-4-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)but-2-ynoate 3wa was obtained through the general procedure using 1.2 equiv of azide with benzene as solvent. Yield: 89%. $[\alpha]_D^{20} = 9^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 7.41

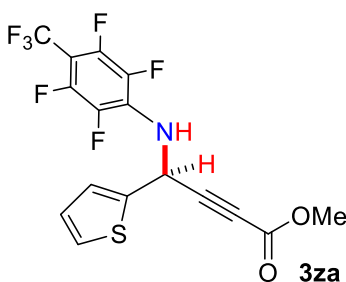
(d, $J = 7.5$ Hz, 1H), 7.37 (t, $J = 7.8$ Hz, 1H), 6.99 (t, $J = 7.5$ Hz, 1H), 6.95 (d, $J = 8.2$ Hz, 2H), 5.98 (d, $J = 9.6$ Hz, 1H), 5.03 (d, $J = 10.2$ Hz, 1H), 3.90 (s, 3H), 3.76 (s, 3H). ^{13}C NMR (150 MHz, CDCl_3) δ 156.92, 153.58, 144.78 (dm, $J = 256.5$ Hz), 137.42 (dm, $J = 242.0$ Hz), 130.81, 129.16 (m), 128.70, 124.48, 121.37 (q, $J = 273.4$ Hz), 121.08, 111.52, 98.45 (m), 84.89, 76.08, 55.67, 52.78, 47.47. ^{19}F NMR (564 MHz, CDCl_3) δ -55.18 (t, $J = 21.0$ Hz, 3F), -142.60 (m, 2F), -157.45 (m, 2F). IR (neat, cm^{-1}): 2935, 2237, 1729, 1599, 1495, 1337, 1142, 992, 876, 716. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{19}\text{H}_{13}\text{F}_7\text{NO}_3$: 436.07782, Found: 436.07780. HPLC analysis: ee = 90%. OJH (90% hexane: 10% isopropanol, 0.8 ml/min): $t_{\text{major}} = 18.90$ min, $t_{\text{minor}} = 16.36$ min.



Methyl (R)-4-(benzo[d][1,3]dioxol-5-yl)-4-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino) but-2-ynoate 3xa was obtained through the general procedure using 1.2 equiv of azide with benzene as solvent. Yield: 81%. $[\alpha]_{\text{D}}^{20} = 17^\circ$ ($c = 1.0$, CHCl_3). ^1H NMR (600 MHz, CDCl_3) δ 7.02 (d, $J = 8.1$ Hz, 1H), 7.00 (s, 1H), 6.83 (d, $J = 7.9$ Hz, 1H), 6.01 (s, 2H), 5.74 (d, $J = 9.3$ Hz, 1H), 4.45 (d, $J = 9.3$ Hz, 1H), 3.79 (s, 3H). ^{13}C NMR (150 MHz, CDCl_3) δ 153.29, 148.56, 148.48, 144.80 (dm, $J = 261.4$ Hz), 137.51 (dm, $J = 242.3$ Hz), 129.90, 128.63 (m), 121.28 (q, $J = 273.3$ Hz), 121.01, 108.71, 107.50, 101.64, 99.08 (m), 84.05, 77.63, 52.99, 50.64. ^{19}F NMR (564 MHz, CDCl_3) δ -55.25 (t, $J = 20.9$ Hz, 3F), -142.02 (m, 2F), -156.60 (m, 2F). IR (neat, cm^{-1}): 2917, 2240, 1724, 1596, 1495, 1335, 1221, 1140, 991, 715. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{19}\text{H}_{11}\text{F}_7\text{NO}_4$: 450.05708, Found: 450.05914. HPLC analysis: ee = 94%. ASH (90% hexane: 10% isopropanol, 0.8 ml/min): $t_{\text{major}} = 16.53$ min, $t_{\text{minor}} = 13.61$ min.



Methyl (*S*)-4-(naphthalen-1-yl)-4-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)but-2-ynoate **3ya** was obtained through the general procedure using 1.2 equiv of azide with benzene as solvent. Yield: 90%. $[\alpha]_D^{20} = 10^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 8.10 (d, $J = 8.4$ Hz, 1H), 8.01 – 7.86 (m, 3H), 7.67 – 7.50 (m, 3H), 6.58 (d, $J = 9.4$ Hz, 1H), 4.50 (d, $J = 8.4$ Hz, 1H), 3.80 (s, 3H). $^{13}\text{C NMR}$ (150 MHz, CDCl_3) δ 155.98, 147.53 (dm, $J = 262.3$ Hz), 140.26 (dm, $J = 258.0$ Hz), 136.79, 133.68, 133.32, 132.89, 131.84, 131.47 (m), 130.04, 129.14, 128.65, 127.95, 125.31, 123.96 (q, $J = 273.3$ Hz), 101.82 (m), 86.65, 81.01, 55.65, 51.21. $^{19}\text{F NMR}$ (564 MHz, CDCl_3) δ -55.22 (t, $J = 20.9$ Hz, 3F), -141.89 (m, 2F), -156.81 (m, 2F). IR (neat, cm^{-1}): 3344, 2957, 2240, 1719, 1659, 1511, 1335, 1230, 778, 714. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{16}\text{H}_9\text{F}_7\text{SNO}_2$: 456.08290, Found: 456.08188. HPLC analysis: ee = 59%. ASH (95% hexane: 5% isopropanol, 0.8 ml/min): $t_{\text{major}} = 9.96$ min, $t_{\text{minor}} = 7.30$ min.



Methyl (*S*)-4-((2,3,5,6-tetrafluoro-4-(trifluoromethyl)phenyl)amino)-4-(thiophen-2-yl)but-2-ynoate **3za** was obtained through the general procedure using 1.2 equiv of azide with benzene as solvent. Yield: 80%. $[\alpha]_D^{20} = 57^\circ$ ($c = 1.0$, CHCl_3). $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.38 (dd, $J = 5.1$, 1.2 Hz, 1H), 7.28 (dd, $J = 3.6$, 1.2 Hz, 1H), 7.03 (dd, $J = 5.1$, 3.6 Hz, 1H), 6.06 (d, $J = 9.7$ Hz, 1H), 4.58 (d, $J = 9.7$ Hz, 2H), 3.80 (s, 2H). $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 153.15, 144.85 (dm, $J = 257.8$ Hz), 138.97, 137.60 (dm, $J = 242.3$ Hz), 128.22 (m), 127.23, 127.18, 127.14, 121.24 (q, $J =$

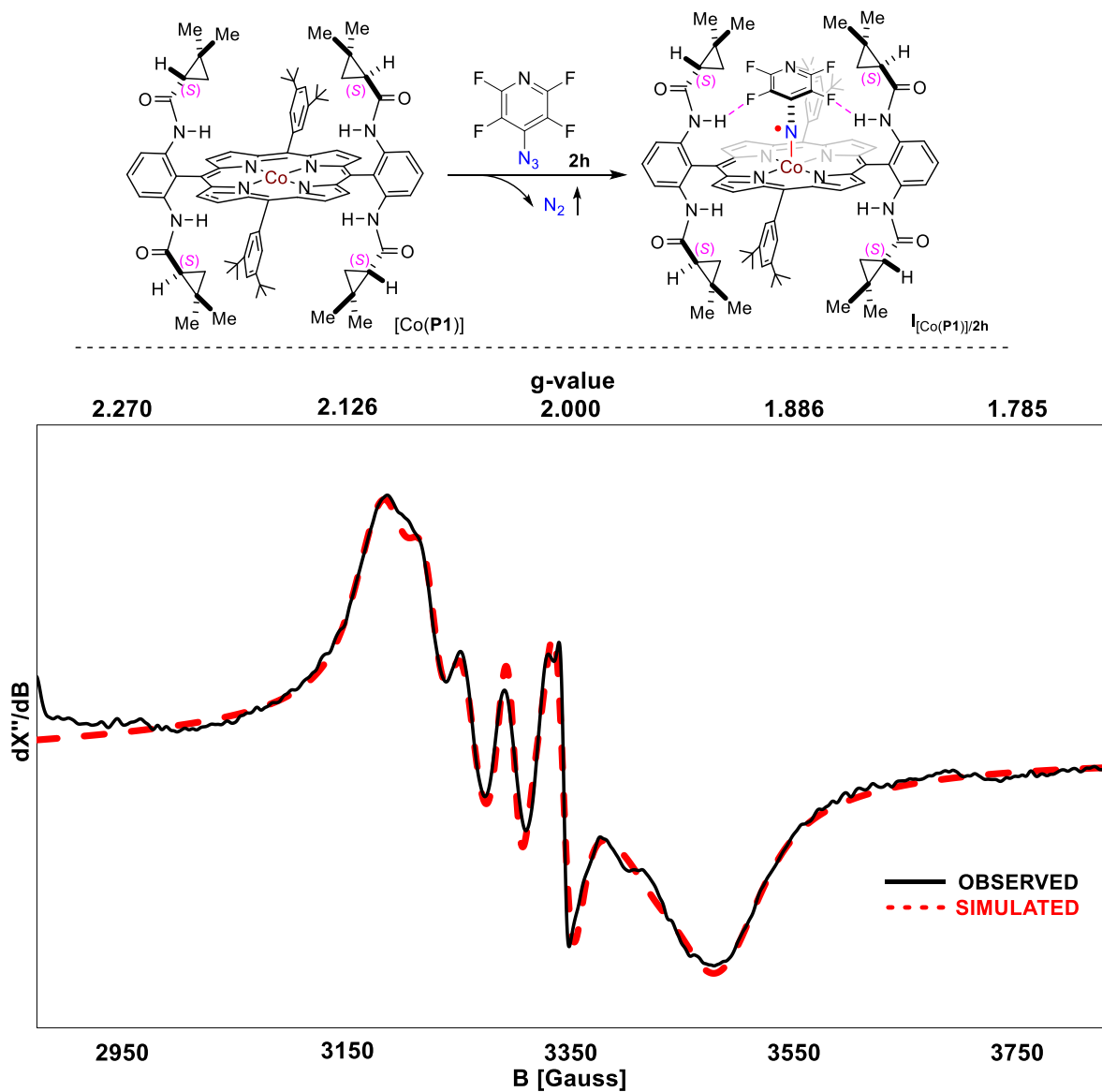
273.3 Hz), 99.66 (m), 83.17, 76.92, 53.05, 46.66. ^{19}F NMR (470 MHz, CDCl_3) δ -55.37 (t, J = 21.0 Hz, 3F), -141.82 (m, 2F), -156.20 (m, 2F). IR (neat, cm^{-1}): 3337, 2958, 2246, 1720, 1511, 1336, 1231, 971, 714. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{16}\text{H}_9\text{F}_7\text{SNO}_2$: 412.02367, Found: 412.02432. HPLC analysis: ee = 99%. ODH (95% hexane: 5% isopropanol, 0.8 ml/min): t_{major} = 15.90 min, t_{minor} = 18.87 min.

6. Characterization of α -Co(III)-Aminyl Radicals by EPR and HRMS

6.1. Procedure for EPR Experiment

Catalyst [Co(Por)] (0.001 mmol) was placed into an oven-dried EPR tube. This EPR tube was then capped with a rubber septum and was fasten with Parafilm. The tube was evacuated and backfilled with nitrogen for three times. Then azide **2** (0.05 mmol) in 0.5 mL benzene was added into this tube through gas-tight syringe. The cap of EPR tube was further sealed with vacuum grease. The reaction mixture was shaken well at r.t. for 10 min. Then the sample was ready for EPR experiment.

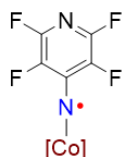
X-band EPR spectra were recorded with a Bruker EMX-Plus spectrometer (Bruker BioSpin). Simulations of the EPR spectra were performed by iteration of the isotropic g-values and line widths using the EPR simulation software SpinFit in Xenon. Experimental X-band EPR isotropic spectra of α -Co(III)-Aminyl Radical **I** in benzene was recorded at r.t. (Freq = 9.42 GHz; mod. amp. = 4 G; microwave power = 20 mW).



Experimental and simulated X-Band EPR spectra for α -Co(III)-aminyl radical $I_{[\text{Co}(\text{P1})]/2\text{h}}$ in benzene at r.t.

EPR simulation details:

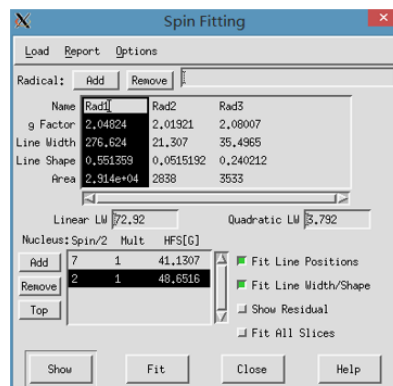
$$N\alpha I_{[Co(P1)]/2h}: 29140/(29140 + 2838 + 3533) = 82\%$$



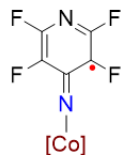
$$g = 2.04824$$

$$A_{(Co)} = 41.1307 \times 2.04824 \times 1.399611451 = 117.9 \text{ MHz}$$

$$A_{(N)} = 48.6516 \times 2.04824 \times 1.399611451 = 139.5 \text{ MHz}$$



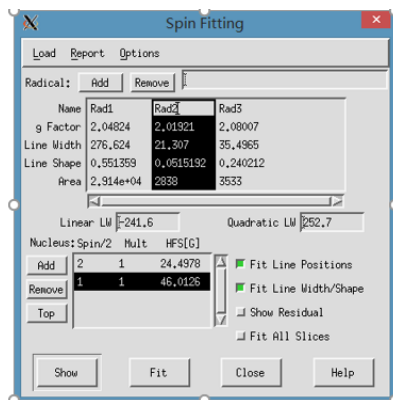
$$C\gamma I_{[Co(P1)]/2h}: 2838/(29140 + 2838 + 3533) = 8\%$$



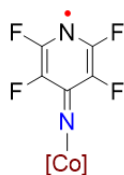
$$g = 2.01921$$

$$A_{(N)} = 24.4978 \times 2.01921 \times 1.399611451 = 69.2 \text{ MHz}$$

$$A_{(F)} = 46.0126 \times 2.01921 \times 1.399611451 = 130.0 \text{ MHz}$$



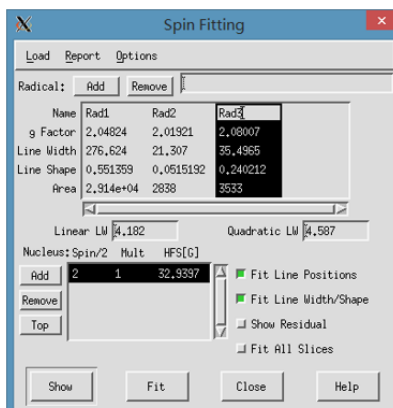
$$N\epsilon I_{[Co(P1)]/2h}: 3533/(29140 + 2838 + 3533) = 10\%$$

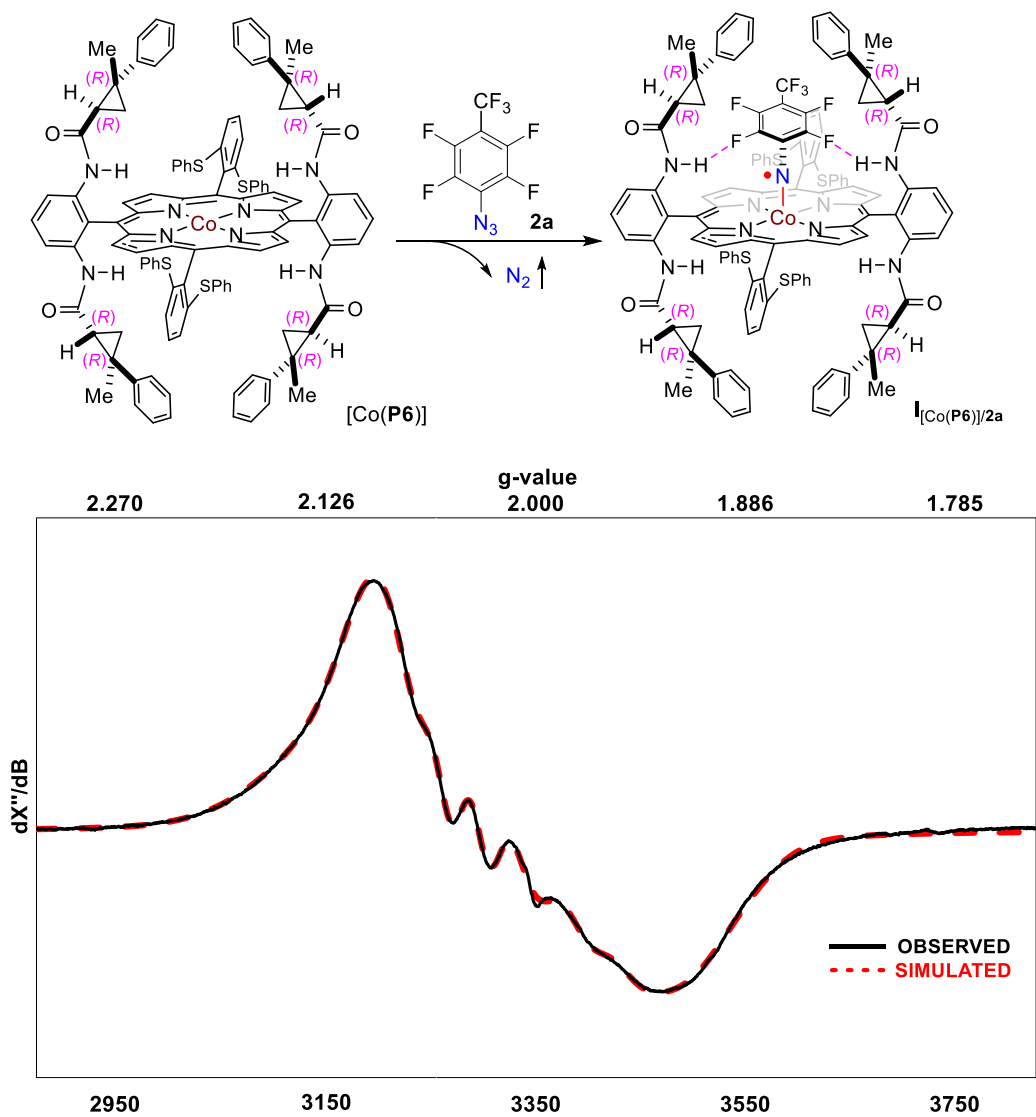


$$g = 2.08007$$

$$A_{(N)} = 32.9397 \times 2.08007 \times 1.399611451 = 95.9 \text{ MHz}$$

$$A_{(F)} = 0 \text{ MHz}$$

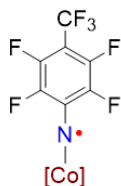




Experimental and simulated X-Band EPR spectra for α -Co(III)-aminyl radical $\mathbf{I}_{[\text{Co}(\text{P6})]/2\text{a}}$ in benzene at RT.

EPR Simulation Details:

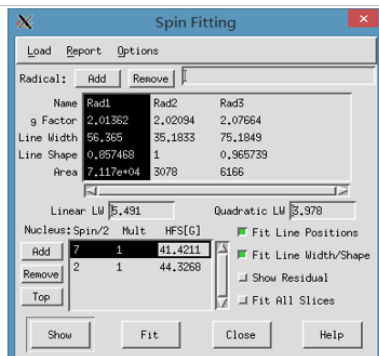
$$N\alpha I_{[Co(P6)]2a}: 71170/(71170 + 3078 + 6166) = 88\%$$



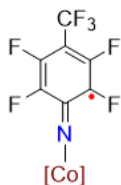
$$g = 2.01362$$

$$A(\text{Co}) = 41.4211 \times 2.01362 \times 1.399611451 = 116.7 \text{ MHz}$$

$$A(\text{N}) = 44.3268 \times 2.01362 \times 1.399611451 = 124.9 \text{ MHz}$$



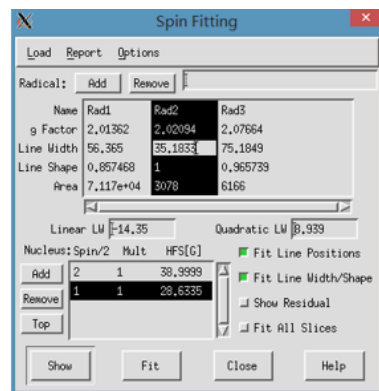
$$C\gamma I_{[Co(P6)]2a}: 3078/(71170 + 3078 + 6166) = 4\%$$



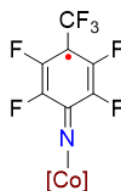
$$g = 2.02094$$

$$A(\text{N}) = 38.9999 \times 2.02094 \times 1.399611451 = 110.3 \text{ MHz}$$

$$A(\text{F}) = 28.6335 \times 2.02094 \times 1.399611451 = 81.0 \text{ MHz}$$



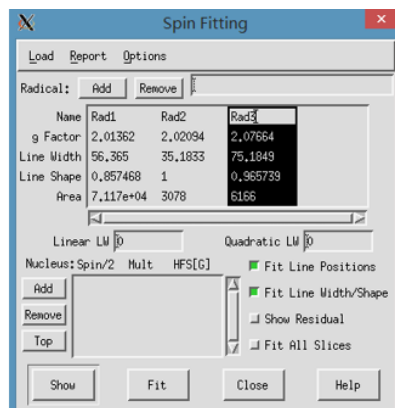
$$C\delta I_{[Co(P6)]}: 6166/(71170 + 3078 + 6166) = 8\%$$



$$g = 2.07664$$

$$A(\text{N}) = 0 \text{ MHz}$$

$$A(\text{F}) = 0 \text{ MHz}$$



6.2. Procedure for HRMS Experiment

$\mathbf{I}_{[\text{Co}(\text{P1})]/2\text{h}}$ was further detected by high resolution mass spectrometry (HRMS) with ESI ionization. Through gas-tight syringe, the same EPR solution was transferred to a sealed HRMS sample vial, which was pre-evacuated and backfilled with nitrogen. The high-resolution mass spectra (LC-HRMS) (ESI) in the absence of any additives that commonly act as electron carriers for ionization allowed for the detection of the molecular ion signals corresponding to the α -Co(III)-aminyl radical $\mathbf{I}_{[\text{Co}(\text{P1})]/2\text{h}}$ ($[\text{M}]^+$ $m/z = 1503.6881$ (observed)), by the loss of one electron (Figure S4).

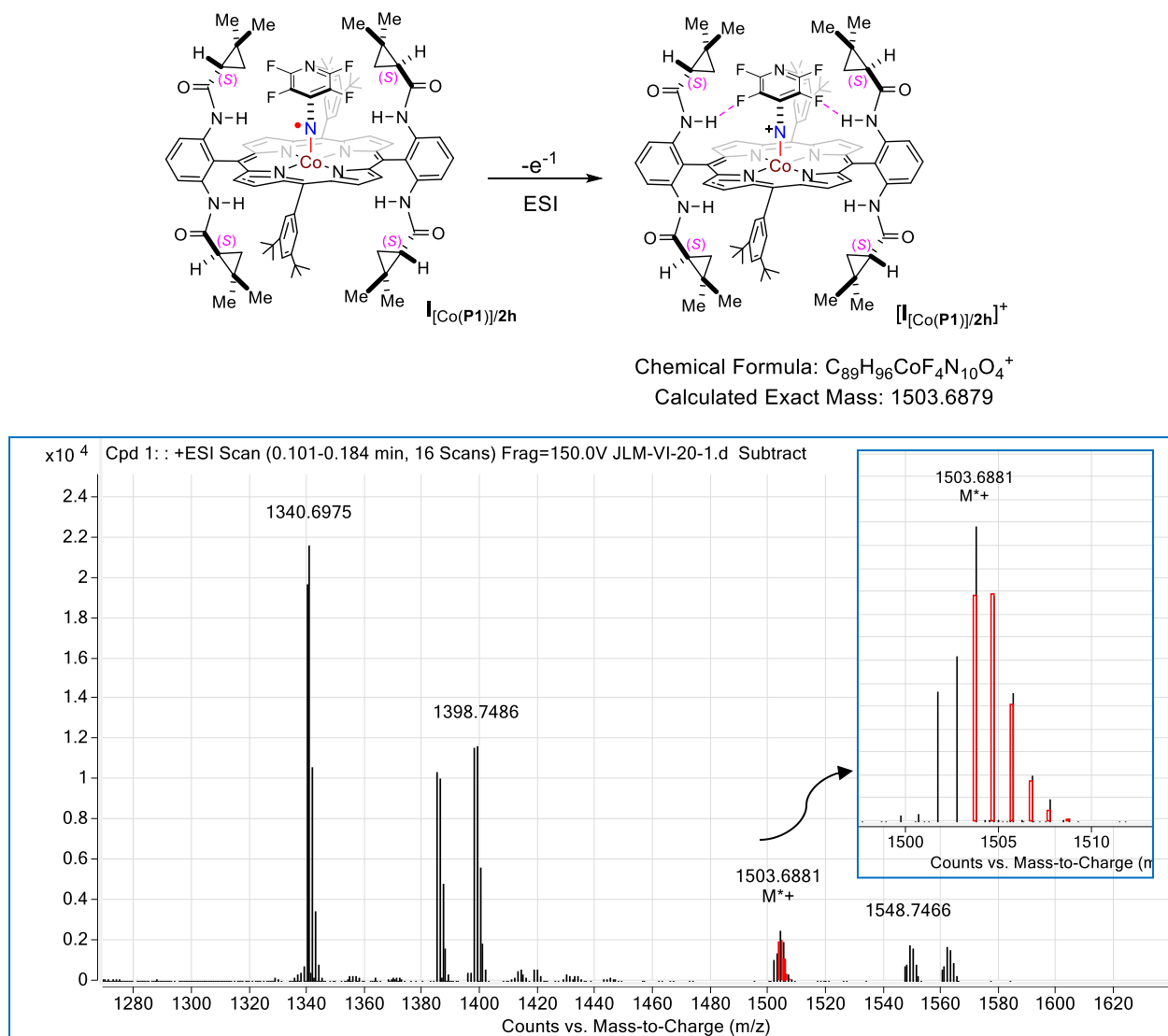
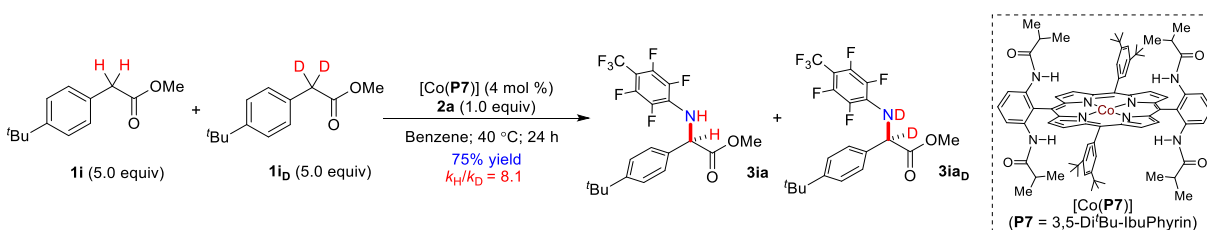


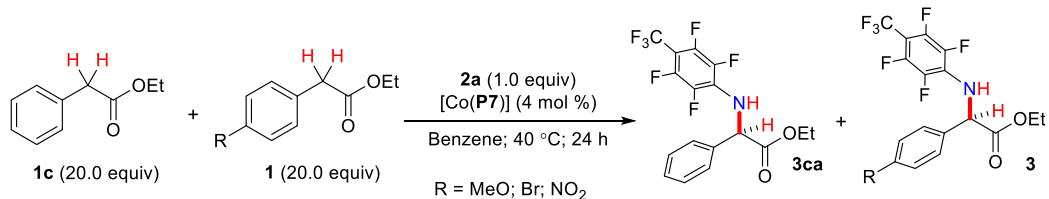
Figure S4. HR-ESI-MS spectrum of reaction mixture of azide **2h** and $[\text{Co}(\text{P1})]$ (the inset red one is the simulated mass spectrum of $\mathbf{I}_{[\text{Co}(\text{P1})]/2\text{h}}$).

7. Kinetic Isotope Effect (KIE) Experiment

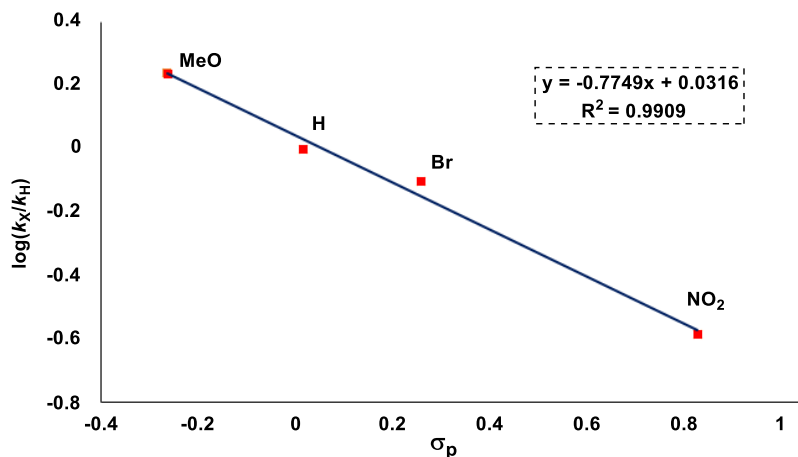


An oven dried Schlenk tube was charged with **[Co(P7)]** (4 mol %) and 4 Å molecular sieves. The Schlenk tube was then evacuated and back filled with nitrogen for three times. The Teflon screw cap was replaced with a rubber septum and methyl 2-(4-(*tert*-butyl)phenyl)acetate (**1i**, 1 mmol, 5.0 equiv), deuterated methyl 2-(4-(*tert*-butyl)phenyl)acetate (**1i_D**, 1 mmol, 5.0 equiv), 4-trifluoromethyl-2,3,5,6-tetrafluorophenyl azide (**2a**, 0.2 mmol, 1 equiv) and benzene (0.5 mL) were added. The Schlenk tube was then purged with nitrogen for 30 s and the rubber septum was replaced with a Teflon screw cap. The mixture was then stirred at 40 °C for 24 h. After completion of the reaction, the reaction mixture was purified by flash column chromatography on silica gel to afford **3ia** together with **3ia_D** in 75% overall yield. KIE (k_H/k_D) was calculated as 8.1 based on ¹H-NMR analysis.

8. Evaluation of Electronic Effect of C–H Substrates via Hammett Study

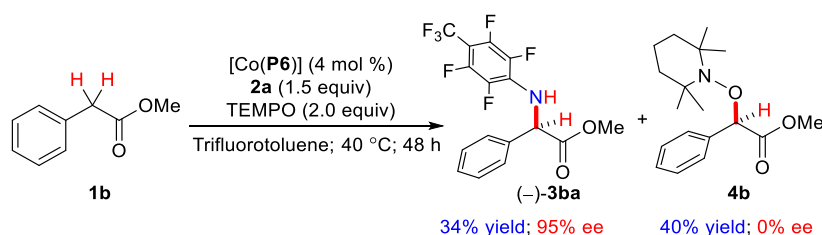


An oven dried Schlenk tube was charged with $[\text{Co}(\text{P7})]$ (4 mol %) and 4 Å molecular sieves. The Schlenk tube was then evacuated for 30 s and back filled with nitrogen. The Teflon screw cap was replaced with a rubber septum and ethyl phenylacetate **1c** (1 mmol, 20 equiv), *para*-substituted methyl phenylacetate **1** (1 mmol, 20 equiv), 4-trifluoromethyl-2,3,5,6-tetrafluorophenyl azide (**2a**, 0.05 mmol, 1 equiv) and benzene (0.5 mL) were added. The Schlenk tube was then purged with nitrogen for 30 s and the rubber septum was replaced with a Teflon screw cap. The mixture was then stirred at 40 °C for 24 h. Following completion of the reaction, the distribution of the aminated products was determined by ¹⁹F NMR: (1) $k_{\text{H}}/k_{\text{Br}} = 100/79$; (2) $k_{\text{MeO}}/k_{\text{NO}_2} = 100/15$; (3) $k_{\text{H}}/k_{\text{NO}_2} = 100/26$. Based on the eqs (1) – (3), the plot of linear free-energy correlation of $\log(k_{\text{X}}/k_{\text{H}})$ versus σ_{p} was obtained as shown below.



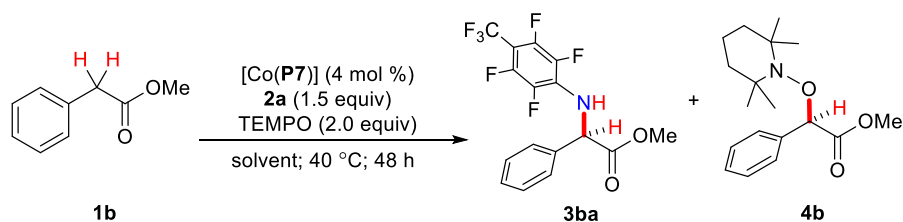
Correlation of $\log(k_{\text{X}}/k_{\text{H}})$ versus σ_{p} plot for amination of *para*-substituted phenylacetates with azide **2a**.

9. Trapping of Alkyl Radical Intermediate



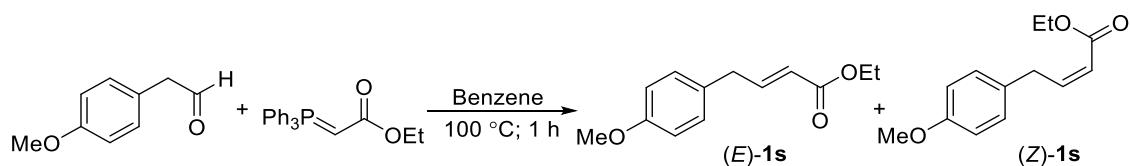
An oven-dried Schlenk tube was charged with [Co(P6)] (4 mol %) and 4 Å molecular sieves. The Schlenk tube was then evacuated and back filled with nitrogen. The Teflon screw cap was replaced with a rubber septum and methyl phenylacetate (**1b**, 0.1 mmol, 1 equiv), 4-trifluoromethyl-2,3,5,6-tetrafluorophenyl azide (**2a**, 0.15 mmol, 1.5 equiv), TEMPO (2 equiv) and α,α,α -trifluorotoluene (0.5 mL) were added. The Schlenk tube was then purged with nitrogen for 30 s and the rubber septum was replaced with a Teflon screw cap. The mixture was then stirred at 40 °C for 48 h. Following completion of the reaction, the mixture was purified by column chromatography to give amide **3ba** (34% yield, 95% ee) and TEMPO trapped product **4b** (40% yield, 0% ee). **4b**: $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 7.44 (m, 2H), 7.33 (m, 2H), 7.29 (m, 1H), 5.21 (s, 1H), 3.65 (s, 3H), 1.51 – 1.25 (m, 6H), 1.23 (s, 3H), 1.13 (s, 3H), 1.07 (s, 3H), 0.71 (s, 3H). $^{13}\text{C NMR}$ (150 MHz, CDCl_3) δ 172.45, 138.14, 128.31, 127.93, 126.87, 88.57, 59.89, 51.82, 40.21, 40.08, 33.53, 32.84, 20.13, 17.07. IR (neat, cm^{-1}): 2930, 1752, 1738, 1160, 724, 696. HRMS (ESI) ($[\text{M}+\text{H}]^+$) Calcd. for $\text{C}_{18}\text{H}_{28}\text{NO}_3$: 306.2064 found: 306.2067.

Table S2. Investigation of Solvent Viscosity Effect using [Co(P7)]

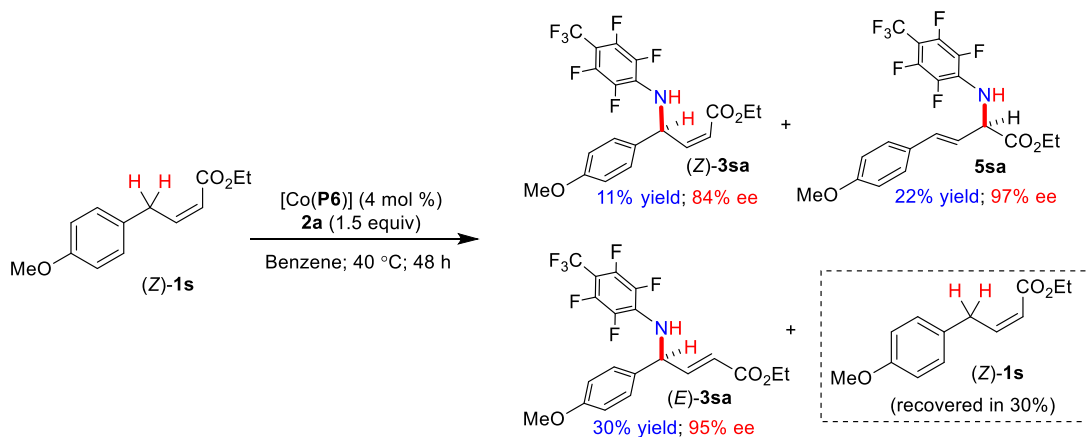


Solvent	Viscosity	Yield of 3ba	Yield of 4b
Hexane	0.31 (20 °C)	19%	48%
Benzene	0.65 (20 °C)	20%	60%
Trifluorotoluene	--	19%	42%
Chlorobenzene	0.80 (20 °C)	15%	42%
1,2-Dichlorobenzene	1.32 (25 °C)	21%	46%
1,4-Dioxane	1.44 (15 °C)	0%	7%

10. Probing Allylic Radical via Olefin Isomerization



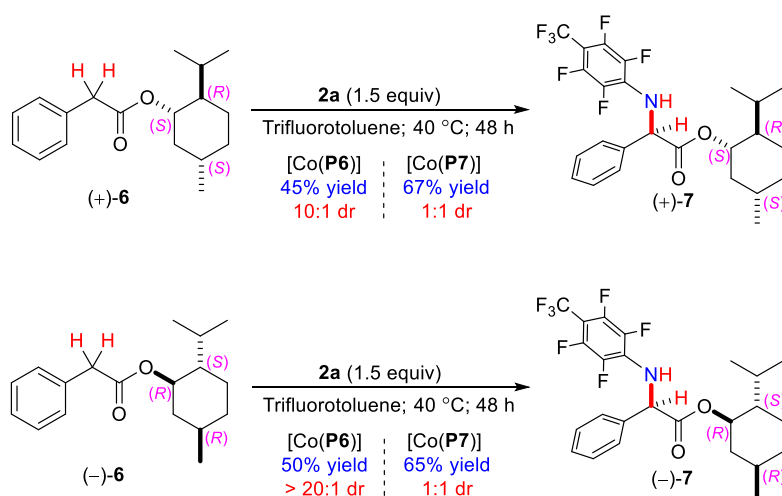
(Z)-1s was synthesized through Wittig reaction. To a solution of the 2-(4-methoxyphenyl)acetaldehyde (1.2 g, 8.0 mmol) in dry benzene (5 mL) was added $\text{Ph}_3\text{P}=\text{CHCO}_2\text{Et}$ (2.79 g, 8.0 mmol) and stirred at reflux for 1 h. After completion of the reaction, the solvent was removed under reduced pressure and the residue purified by column chromatography eluted with hexane/EtOAc (40/1 v/v) to afford **1s** as a mixture of geometrical isomers. The mixture was partially separated by silica gel column chromatography to give **(E)-1s** (1.50 g, 85%) and pure **(Z)-1s** (150 mg, 9%). **(Z)-1s**: ^1H NMR (600 MHz, CDCl_3) δ 7.15 (d, $J = 8.6$ Hz, 2H), 6.85 (d, $J = 8.6$ Hz, 2H), 6.32 (dt, $J = 11.3, 7.6$ Hz, 1H), 5.83 (dt, $J = 11.4, 1.8$ Hz, 1H), 4.22 (q, $J = 7.2$ Hz, 2H), 3.96 (dd, $J = 7.7, 1.8$ Hz, 2H), 3.79 (s, 3H), 1.32 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (150 MHz, CDCl_3) δ 166.34, 158.11, 148.31, 131.44, 129.49, 119.52, 113.97, 59.90, 55.19, 34.18, 14.23. IR (neat, cm^{-1}): 2932, 1714, 1510, 1245, 1160, 823, 756. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for $\text{C}_{13}\text{H}_{17}\text{O}_3$: 221.11722 found: 221.11652.



An oven dried Schlenk tube was charged with $[\text{Co}(\text{P6})]$ (4 mol %) and 4 Å molecular sieves. The Schlenk tube was then evacuated and back filled with nitrogen for three times. The Teflon screw cap was replaced with a rubber septum and **(Z)-1s** (0.10 mmol, 1 equiv), 4-trifluoromethyl-2,3,5,6-tetrafluorophenyl azide (**2a**, 0.15 mmol, 1.5 equiv) and benzene (0.5 mL) were added. The Schlenk tube was then purged with nitrogen for 30 s and the rubber septum was replaced with a Teflon

screw cap. The mixture was then stirred at 40 °C. After 48 h, the reaction mixture was concentrated and the yields of all products are calculated based on crude ¹H NMR with 1,1,2,2-tetrachloroethane as internal standard ((*Z*)-**3sa**: 11% yield; **5sa**: 22% yield; (*E*)-**3sa**: 30% yield; (*Z*)-**1s**: 30% recovered). Crude mixture was purified by PTLC. For product (*Z*)-**3sa**: $[\alpha]_D^{20} = -25^\circ$ ($c = 0.3$, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 7.36 (d, $J = 8.7$ Hz, 2H), 6.92 (d, $J = 8.8$ Hz, 2H), 6.65 (m, 1H), 6.31 (dd, $J = 11.5, 9.0$ Hz, 1H), 5.92 (dd, $J = 11.4, 1.1$ Hz, 1H), 4.64 (d, $J = 8.4$ Hz, 1H), 4.22 (m, 2H), 3.81 (s, 3H), 1.31 (t, $J = 7.1$ Hz, 3H). ¹⁹F NMR (470 MHz, CDCl₃) δ -55.04 (t, $J = 20.8$ Hz, 3F), -142.91 (m, 2F), -158.65 (m, 2F). ¹³C NMR (101 MHz, CDCl₃) δ 165.44, 159.54, 147.04, 144.86 (dm, $J = 245.3$ Hz), 136.85 (dm, $J = 243.4$ Hz), 132.27, 129.95, 127.81, 121.49 (q, $J = 270.0$ Hz), 120.97, 114.50, 60.64, 55.29 (2C), 14.12. IR (neat, cm⁻¹): 3352, 2917, 1716, 1655, 1510, 1333, 1234, 1176, 972, 714. HRMS (DART) ([M+H]⁺) Calcd. for: C₂₀H₁₇F₇NO₃: 452.10912, Found: 452.10941. HPLC analysis: ee = 84%. IA (95% hexane: 5% isopropanol, 0.8 ml/min): $t_{major} = 8.24$ min, $t_{minor} = 7.81$ min. For product **5sa**: $[\alpha]_D^{20} = -50^\circ$ ($c = 0.5$, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 7.32 (d, $J = 7.8$ Hz, 2H), 6.87 (d, $J = 7.7$ Hz, 2H), 6.69 (d, $J = 15.8$ Hz, 1H), 6.04 (dd, $J = 15.8, 6.6$ Hz, 1H), 5.12 (m, 2H), 4.30 (m, 2H), 3.82 (s, 3H), 1.32 (t, $J = 7.1$ Hz, 3H). ¹⁹F NMR (470 MHz, CDCl₃) δ -55.04 (t, $J = 20.8$ Hz, 3F), -142.91 (m, 2F), -158.65 (m, 2F). ¹³C NMR (100 MHz, CDCl₃) δ 170.76, 159.95, 144.81 (dm, $J = 266.2$ Hz), 136.77 (dm, $J = 243.8$ Hz), 134.00, 128.04, 121.43 (q, $J = 272.6$ Hz), 121.37, 114.09, 94.04 (m), 62.37, 58.82, 55.32, 14.09. IR (neat, cm⁻¹): 3391, 2938, 1737, 1511, 1334, 1133, 993, 713. HRMS (ESI) ([M-H]⁻) Calcd. for: C₂₀H₁₅F₇NO₃: 450.0946, Found: 450.0951. HPLC analysis: ee = 97%. IA (95% hexane: 5% isopropanol, 0.8 ml/min): $t_{major} = 7.46$ min, $t_{minor} = 8.22$ min.

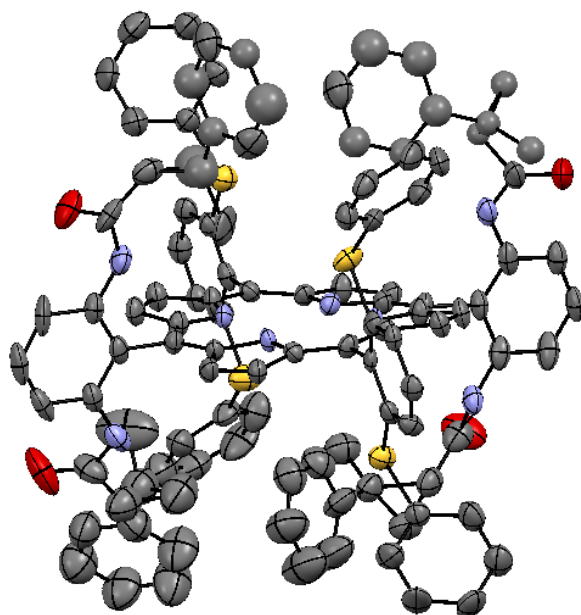
11. Evaluation of Diastereoselectivity with Chiral C–H Substrates



First, the reactions were performed with (+)-**6** (**14**) using [Co(**P6**)] or [Co(**P7**)] as catalyst respectively following the general procedure using 1.5 equiv of azide **2a** with α,α,α -trifluorotoluene as solvent. After 48 hours, the dr of products were determined by crude ^{19}F NMR. The results indicated that when [Co(**P6**)] is used as catalyst, (+)-**7** could be obtained in 45% yield with the dr of 10/1. When [Co(**P7**)] was used as catalyst, (+)-**7** could be obtained in 67% yield with the dr of 1/1. Similar results were achieved when using (-)-**6** as starting material using [Co(**P6**)] or [Co(**P7**)] as catalyst respectively. When [Co(**P6**)] was used as catalyst, (-)-**7** could be obtained in 50% yield with the dr of >20/1. When [Co(**P7**)] was used as catalyst, (-)-**7** could be obtained in 65% yield with the dr of 1/1. (-)-**7**: ^1H NMR (500 MHz, CDCl_3) δ 7.39 – 7.31 (m, 5H), 5.58 (d, $J = 7.5$ Hz, 1H), 5.42 (d, $J = 7.8$ Hz, 1H), 4.65 (td, $J = 11.0, 4.4$ Hz, 1H), 2.03 (d, $J = 11.0$ Hz, 1H), 1.71 – 1.62 (m, 1H), 1.53 – 1.41 (m, 1H), 1.29 – 1.19 (m, 2H), 1.05 (q, $J = 11.9$ Hz, 1H), 0.97 – 0.76 (m, 6H), 0.55 (d, $J = 7.0$ Hz, 3H), 0.33 (d, $J = 6.9$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 170.43, 144.73 (dm, $J = 257.6$ Hz), 136.95, 136.91 (dm, $J = 242.0$ Hz), 129.21 (m), 128.91, 128.88, 126.96, 121.42 (q, $J = 272.8$ Hz), 97.29 (m), 60.76, 47.09, 40.67, 34.03, 31.40, 25.29, 22.88, 21.92, 20.41, 15.45. ^{19}F NMR (376 MHz, CDCl_3): δ -55.08 (t, $J = 20.9$ Hz), -143.01 (m, 3F), -158.67 (m, 2F). IR (neat, cm^{-1}): 2954, 2927, 2869, 1497, 1333, 1256, 1137, 986, 695. HRMS (DART) ($[\text{M}+\text{H}]^+$) Calcd. for: $\text{C}_{25}\text{H}_{27}\text{F}_7\text{NO}_2$: 506.19245, Found: 506.18992.

12. X-ray Crystallographic Information

12.1. Single-crystal X-ray Structure of P6



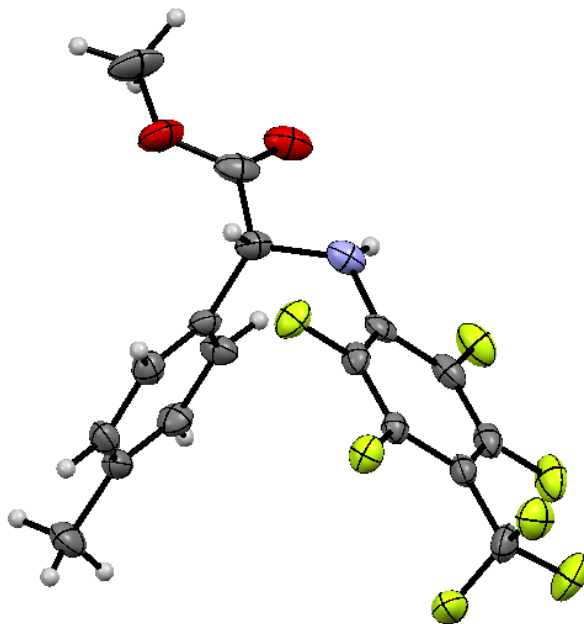
Single-crystal X-ray structure of P6 with thermal ellipsoids drawn at 50% probability

P6: The majority of non-hydrogen atoms were refined anisotropically. Several side arms of porphyrin molecules are disordered over two positions with 1:1 ratio of disordered parts (fixed). Some of disordered atoms have been refined isotropically. Disordered parts have been refined using distance and angular restraints (DFIX, DANG) and constraints (AFIX 66) to keep the geometry chemically feasible. All disordered solvent molecules of toluene, and 1,4-dioxane have been refined using geometry restraints and constraints. There are two symmetrically independent porphyrin molecules in the crystal structure.

Crystal Data and Structure Refinement for P6.

Identification code	P6
Empirical formula	C ₂₅₆ H ₂₂₂ N ₁₆ O ₁₁ S ₈
Formula weight	3954.98
Temperature/K	100(2)
Crystal system	triclinic
Space group	P1
a/Å	13.1087(5)
b/Å	15.9083(5)
c/Å	26.9265(10)
α/°	90.115(2)
β/°	101.243(2)
γ/°	102.434(2)
Volume/Å ³	5372.6(3)
Z	1
ρ _{calc} /mm ³	1.222
m/mm ⁻¹	1.285
F(000)	2086.0
Crystal size/mm ³	0.11 × 0.1 × 0.08
Radiation	CuKα (λ = 1.54178)
2θ range for data collection	6.7 to 131.96°
Index ranges	-15 ≤ h ≤ 15, -18 ≤ k ≤ 18, -31 ≤ l ≤ 31
Reflections collected	66612
Independent reflections	32728 [R _{int} = 0.0795, R _{sigma} = 0.1105]
Data/restraints/parameters	32728/285/2534
Goodness-of-fit on F ²	1.020
Final R indexes [I ≥ 2σ (I)]	R ₁ = 0.0958, wR ₂ = 0.2392
Final R indexes [all data]	R ₁ = 0.1442, wR ₂ = 0.2766
Largest diff. peak/hole / e Å ⁻³	0.62/-0.46
Flack parameter	0.09(2)

12.2. Single-crystal X-ray Structure of 3ha



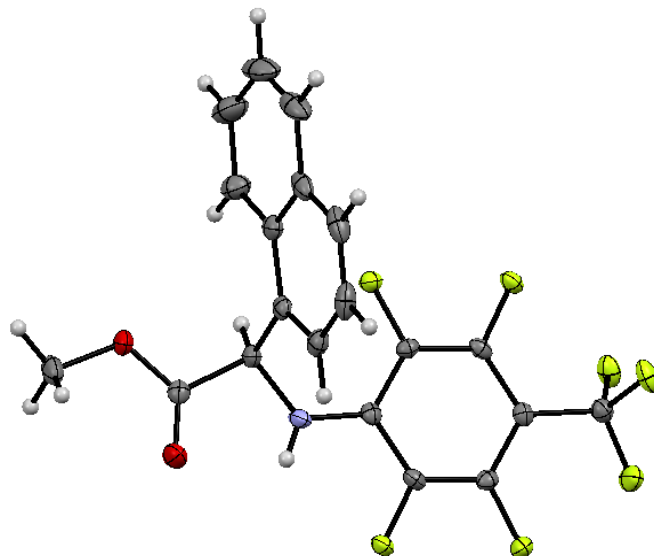
Single-crystal X-ray structure of **3ha** with thermal ellipsoids drawn at 50% probability

3ha: All non-hydrogen atoms were refined anisotropically. Hydrogen atom of –NH groups has been found from Fourier difference map and were freely refined. Remaining hydrogen atoms were placed in geometrically calculated positions and included in the refinement process using riding model with isotropic thermal parameters: $U_{iso}(H) = 1.2[1.5]U_{eq}(-CH[-CH_3])$. Disorderd -CF₃ group has been refined with SADI restraints.

Crystal Data and Structure Refinement for 3ha

Identification code	3ha
Empirical formula	C17 H12 F7 N O2
Formula weight	395.28
Temperature	100(2) K
Wavelength	1.54178 Å
Crystal system, space group	Triclinic, P1
Unit cell dimensions	a = 9.1989(3) Å alpha = 92.451(2) deg. b = 9.2630(3) Å beta = 103.071(2) deg. c = 9.9868(3) Å gamma = 94.018(2) deg.
Volume	825.38(5) Å ³
Z, Calculated density	2, 1.590 Mg/m ³
Absorption coefficient	1.406 mm ⁻¹
F(000)	400
Crystal size	0.12 x 0.06 x 0.03 mm
Theta range for data collection	4.55 to 68.70 deg.
Limiting indices	-11<=h<=11, -11<=k<=11, -12<=l<=11
Reflections collected / unique	10750 / 5264 [R(int) = 0.0374]
Completeness to theta = 66.60	97.7 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.9590 and 0.8494
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	5264 / 31 / 536
Goodness-of-fit on F ²	1.057
Final R indices [I>2sigma(I)]	R1 = 0.0403, wR2 = 0.0837
R indices (all data)	R1 = 0.0506, wR2 = 0.0888
Absolute structure parameter	0.08(11)
Largest diff. peak and hole	0.345 and -0.203 e.Å ⁻³

12.3. Single-crystal X-ray Structure of **3ma**



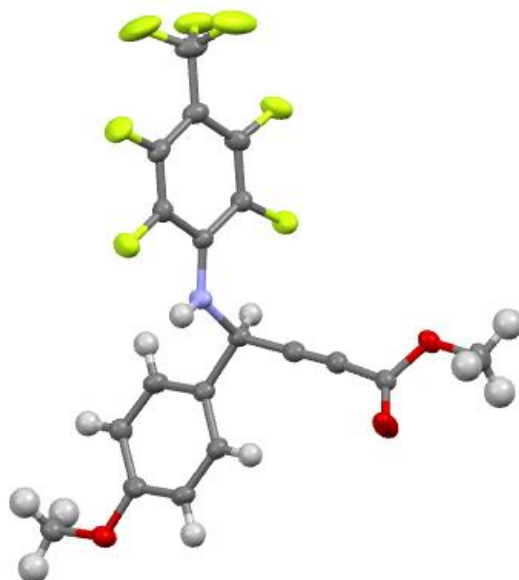
Single-crystal X-ray structure of **3ma** with thermal ellipsoids drawn at 50% probability.

3ma: All non-hydrogen atoms were refined anisotropically. The hydrogen atom of –NH group has been found from difference Fourier map and was freely refined. Remaining hydrogen atoms were placed in geometrically calculated positions and included in the refinement process using riding model with isotropic thermal parameters: $U_{iso}(H) = 1.2U_{eq}(-CH)$ and $U_{iso}(H) = 1.5U_{eq}(-CH_3)$.

Crystal Data and Structure Refinement for 3ma

Identification code	3ma
Empirical formula	C ₂₀ H ₁₂ F ₇ NO ₂
Formula weight	431.31
Temperature/K	100(2)
Crystal system	orthorhombic
Space group	P212121
a/Å	7.9791(2)
b/Å	8.3129(2)
c/Å	26.4307(6)
α/°	90
β/°	90
γ/°	90
Volume/Å ³	1753.13(7)
Z	4
ρ _{calc} /mm ³	1.634
m/mm ⁻¹	1.386
F(000)	872.0
Crystal size/mm ³	0.12 × 0.03 × 0.02
Radiation	CuKα (λ = 1.54178)
2θ range for data collection	6.688 to 137.756°
Index ranges	-9 ≤ h ≤ 9, -9 ≤ k ≤ 10, -29 ≤ l ≤ 31
Reflections collected	23084
Independent reflections	3224 [R _{int} = 0.0406, R _{sigma} = 0.0218]
Data/restraints/parameters	3224/0/276
Goodness-of-fit on F ²	1.064
Final R indexes [I ≥ 2σ (I)]	R ₁ = 0.0256, wR ₂ = 0.0596
Final R indexes [all data]	R ₁ = 0.0294, wR ₂ = 0.0613
Largest diff. peak/hole / e Å ⁻³	0.20/-0.20

12.4. Single-crystal X-ray Structure of 3va



Single-crystal X-ray structure of 3va with thermal ellipsoids drawn at 50% probability

Crystal Data and Structure Refinement for 3va

Identification code	3va
Empirical formula	C ₁₉ H ₁₂ F ₇ N O ₃
Formula weight	435.30
Temperature	173(2) K
Wavelength	1.54178 Å
Crystal system	Monoclinic
Space group	P2 ₁
Unit cell dimensions	a = 5.5696(4) Å b = 9.1428(7) Å c = 17.2564(13) Å
Volume	878.62(11) Å ³
Z	2
Density (calculated)	1.645 Mg/m ³
Absorption coefficient	1.431 mm ⁻¹
F(000)	440
Crystal size	0.580 x 0.140 x 0.100 mm ³
Theta range for data collection	5.127 to 66.601°
Index ranges	-6<=h<=6, -10<=k<=10, -20<=l<=20
Reflections collected	16507
Independent reflections	3075 [R(int) = 0.0309]
Completeness to theta = 66.601°	99.3 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.7528 and 0.5842
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	3075 / 2 / 276
Goodness-of-fit on F ²	1.053
Final R indices [I>2sigma(I)]	R1 = 0.0289, wR2 = 0.0765
R indices (all data)	R1 = 0.0291, wR2 = 0.0769
Absolute structure parameter	-0.02(4)
Extinction coefficient	n/a
Largest diff. peak and hole	0.226 and -0.231 e.Å ⁻³

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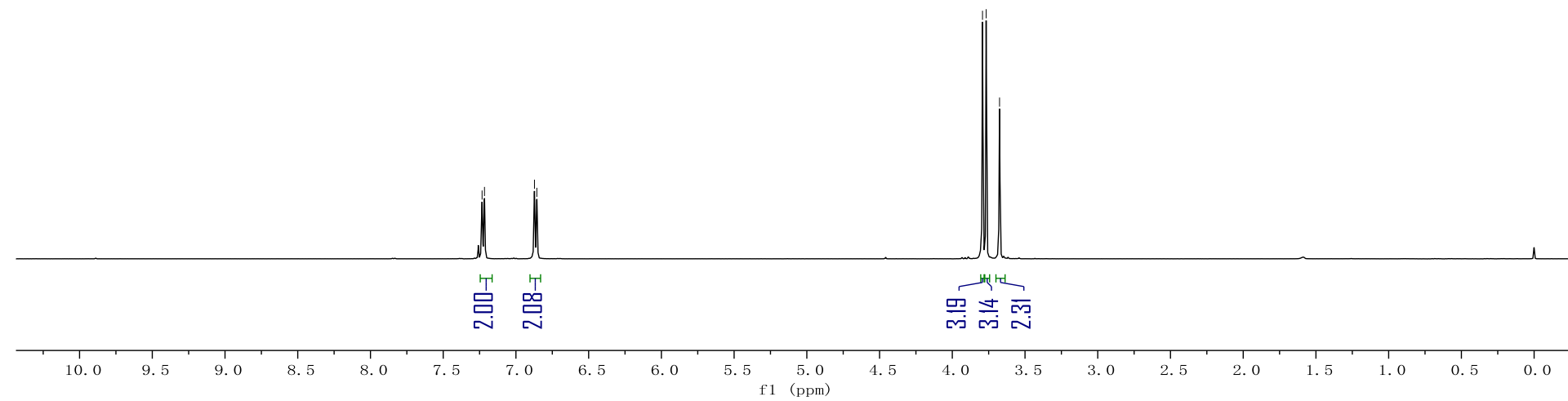
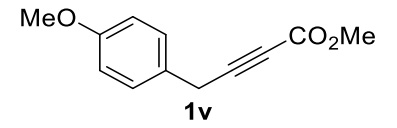
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Copies of NMR/HPLC Spectra

¹H NMR

7.233
7.216
6.872
6.856

3.792
3.767
3.675



^{13}C NMR

— 158.760

— 154.123

— 129.010

— 125.945

— 114.182

— 87.195

77.265

77.011

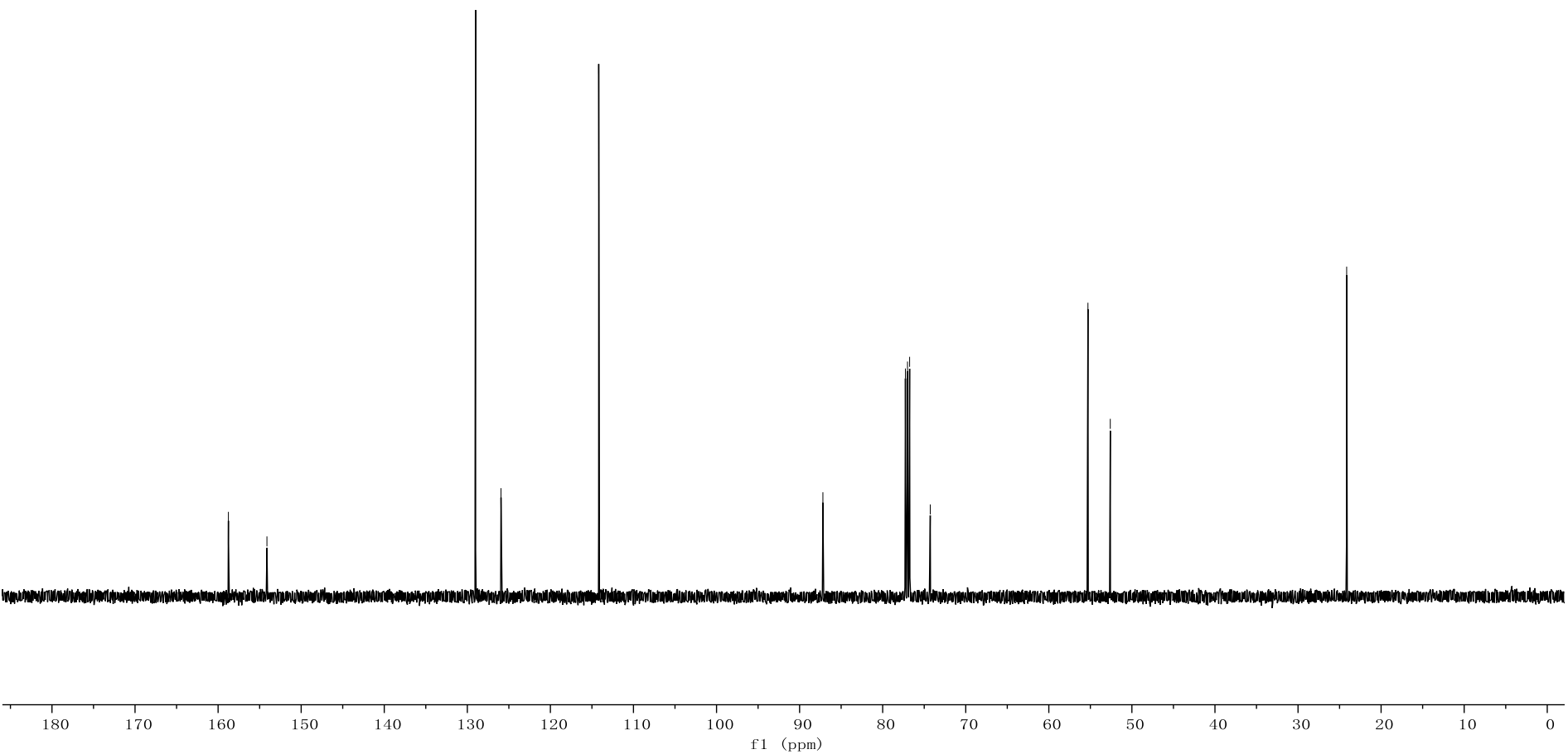
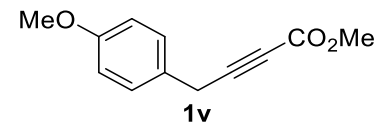
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74.272

55.291

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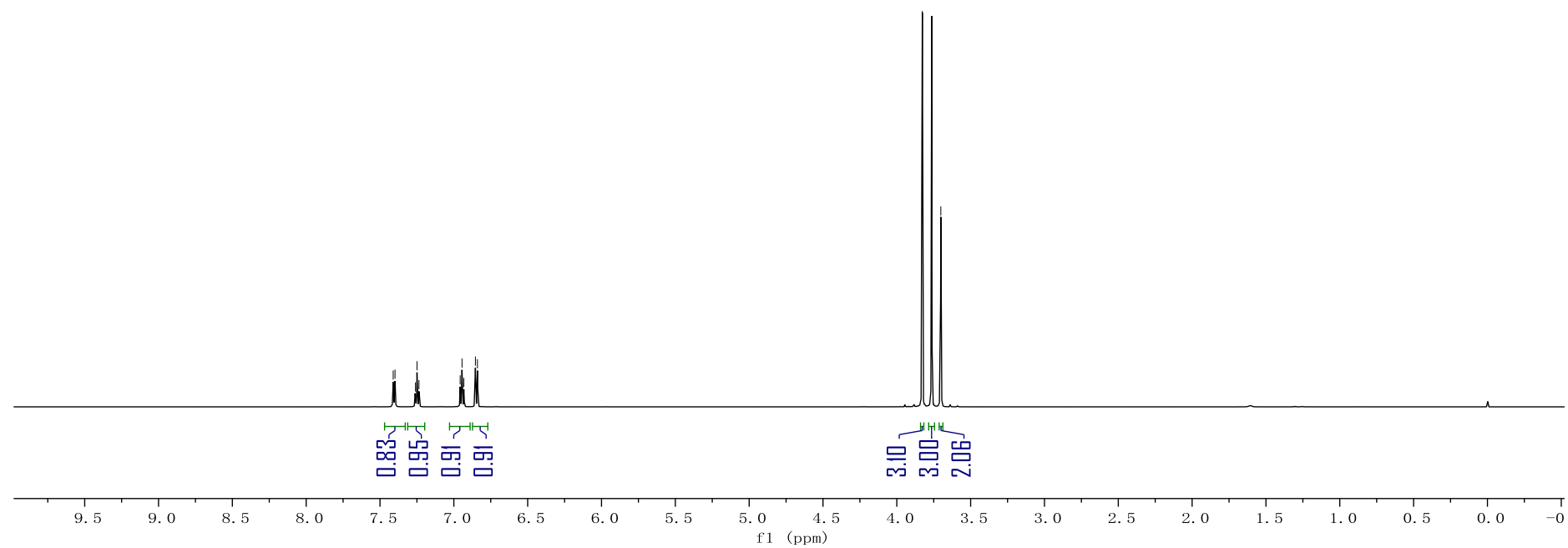
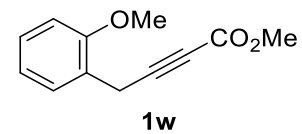
— 24.145



^1H NMR

7.411
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7.236
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6.932
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3.827
3.764
3.702



¹³C NMR

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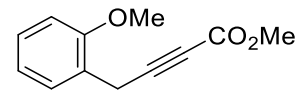
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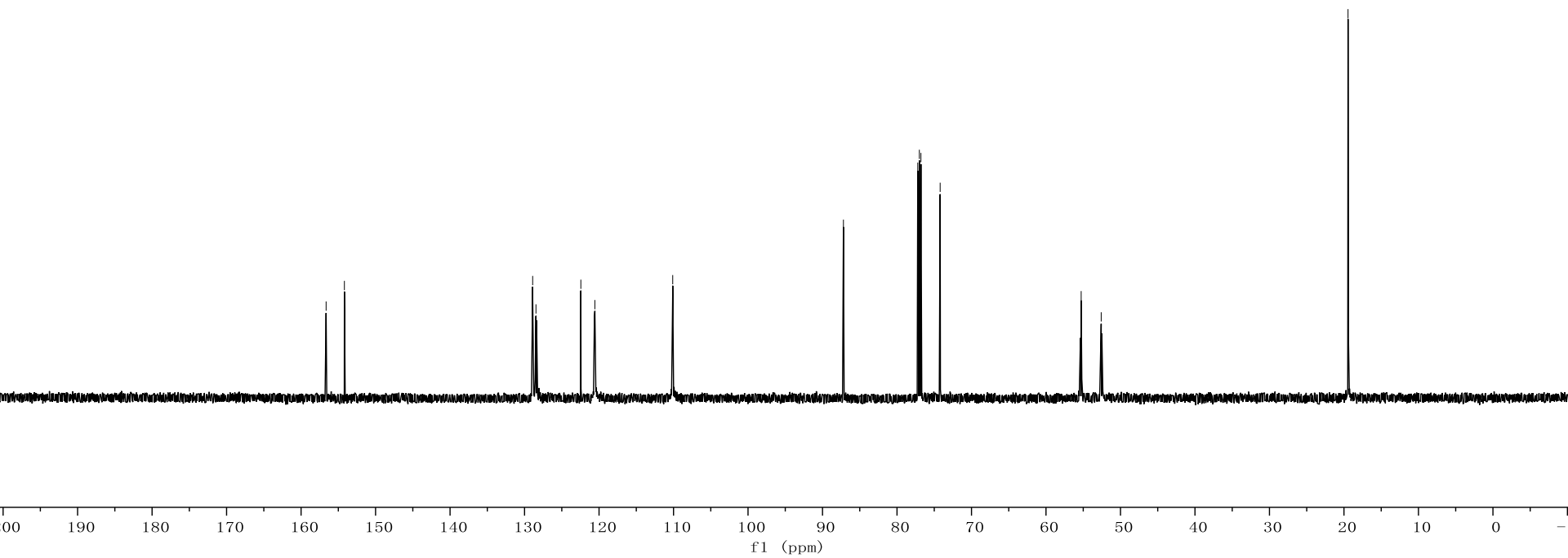
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19.454



1w

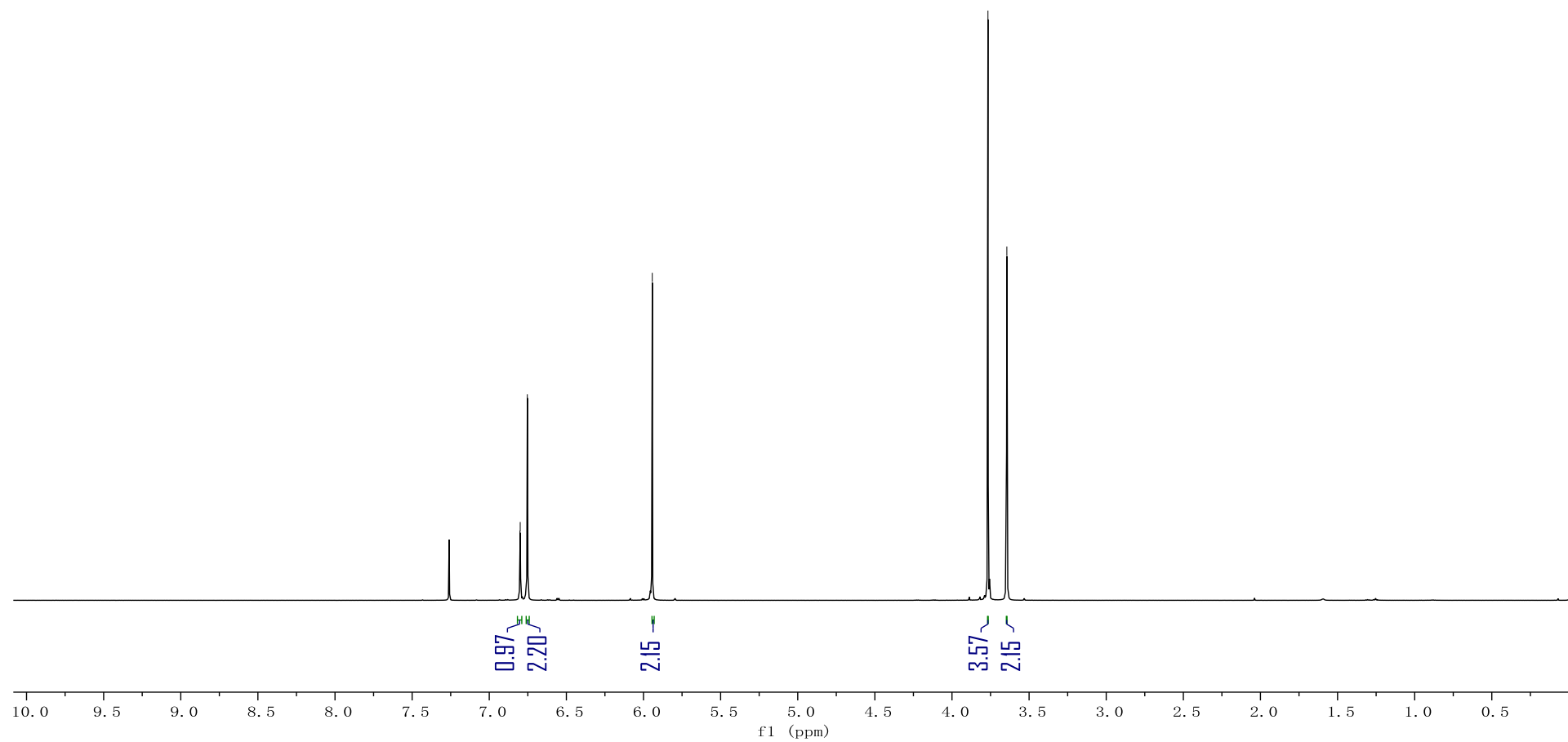
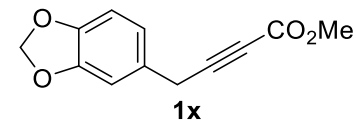


¹H NMR

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5.943

3.768
3.767
3.644



¹³C NMR

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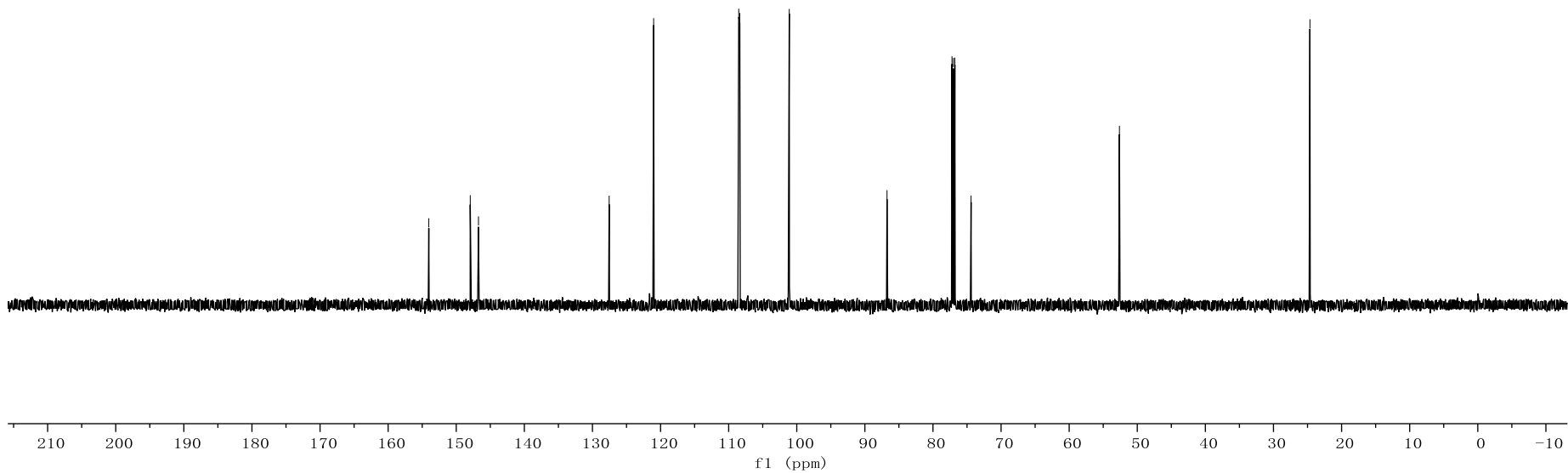
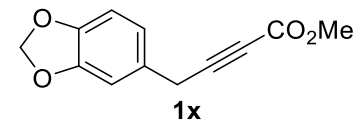
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77.001
76.788
74.408

52.619

24.647

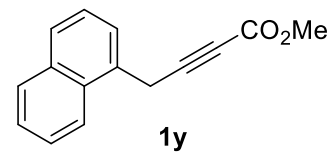


¹H NMR

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7.544
7.533
7.530
7.528
7.516
7.514
7.469
7.455
7.452
7.438

4.129

3.772



1.00
0.98
1.00
1.99
1.03
1.04

2.16

3.22

10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0

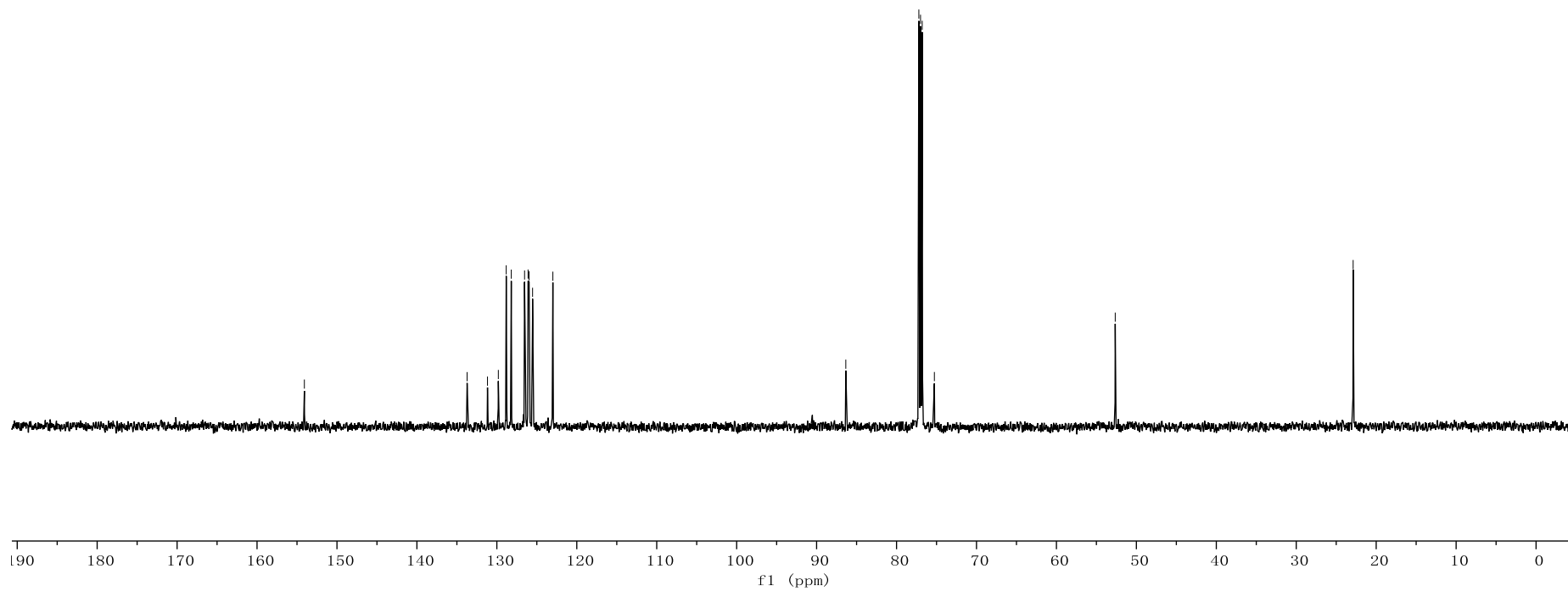
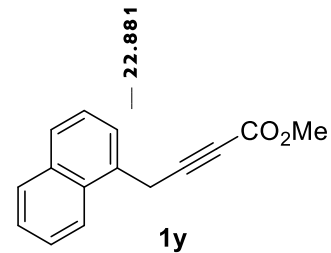
f1 (ppm)

¹³C NMR

— 154.079
133.721
131.172
129.814
128.829
128.197
126.533
126.097
125.966
125.532
122.992

— 86.349
77.211
77.000
76.788
75.268

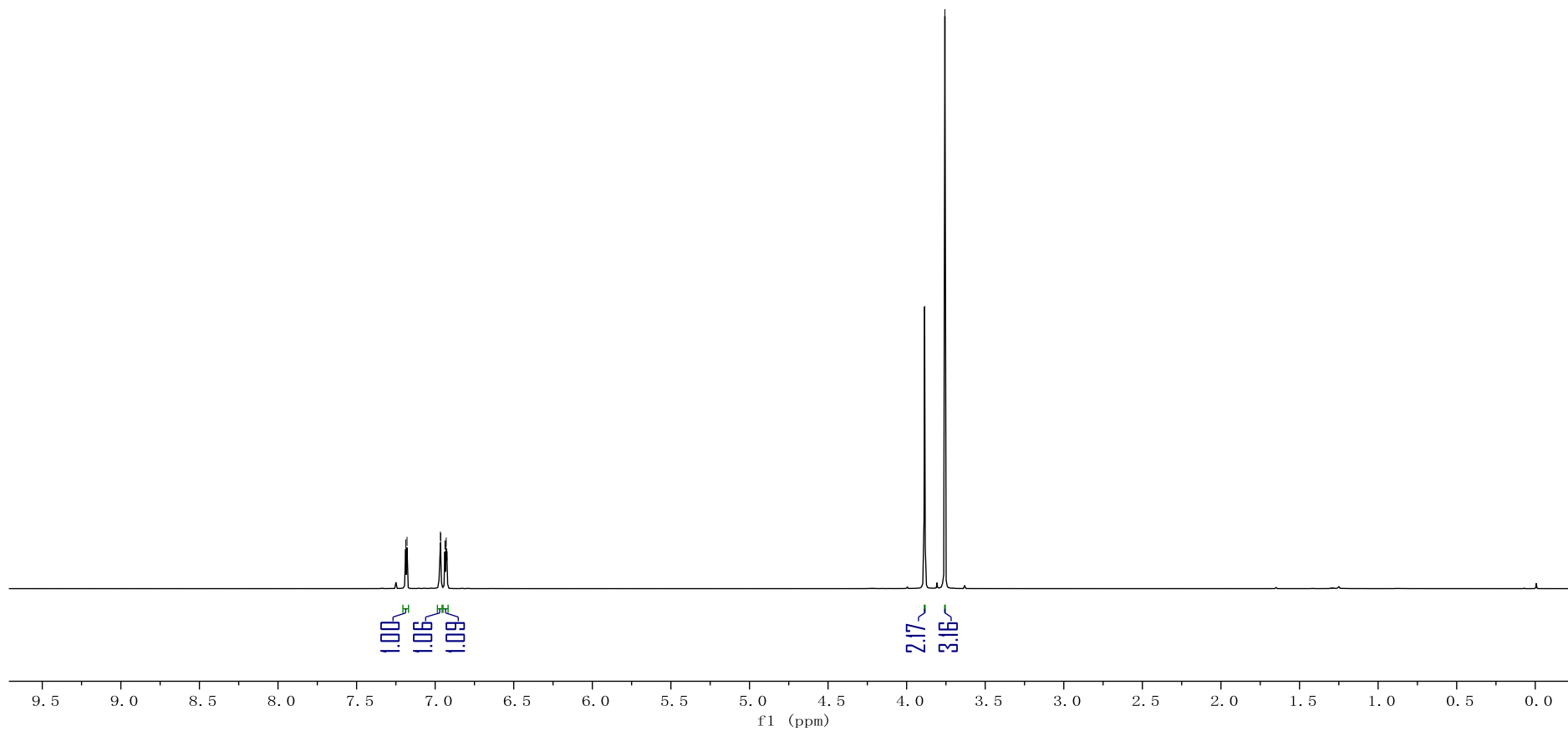
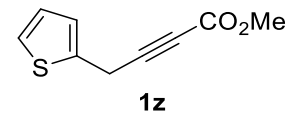
— 52.644



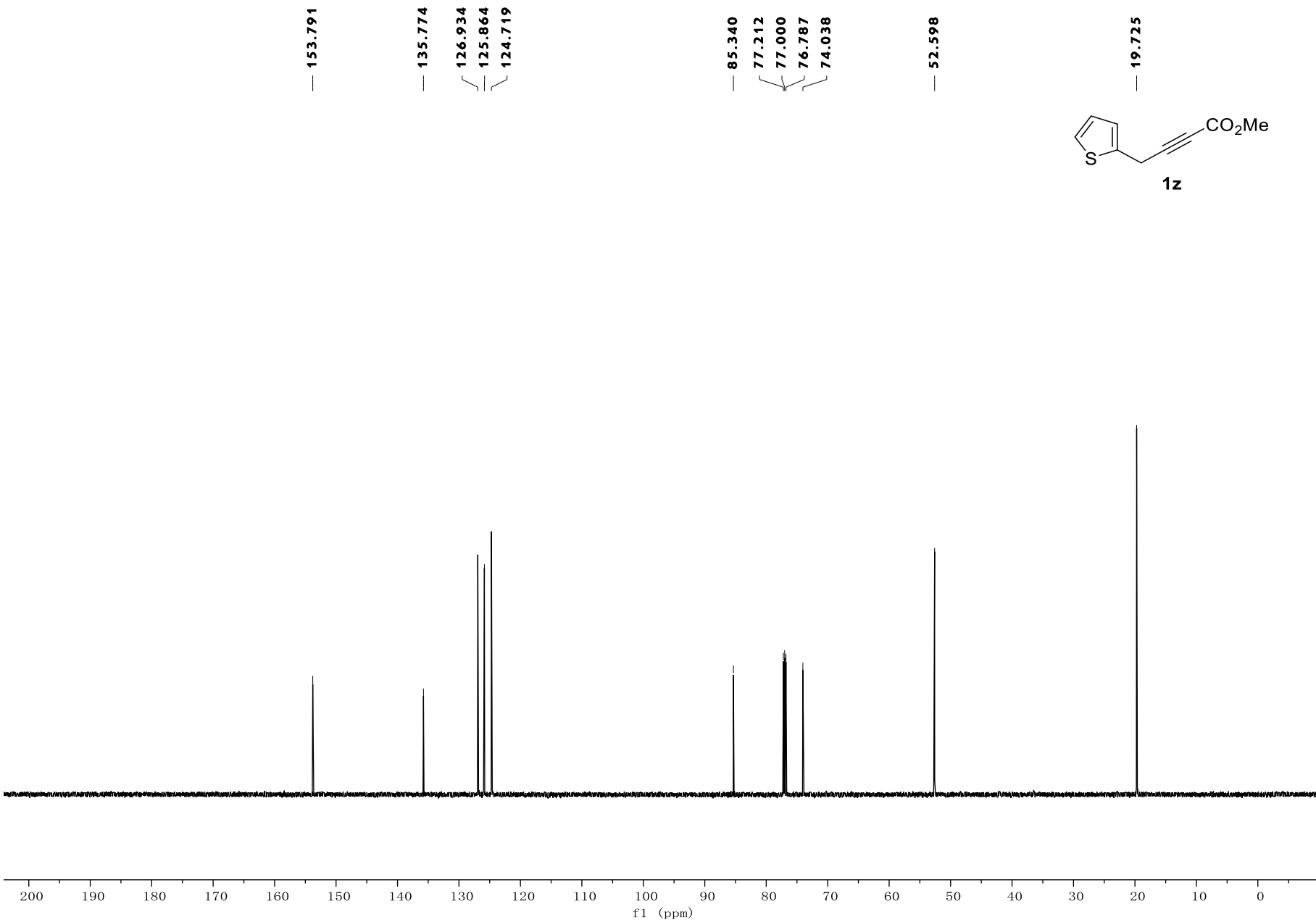
^1H NMR

7.187
7.179
6.967
6.965
6.938
6.932
6.930
6.924

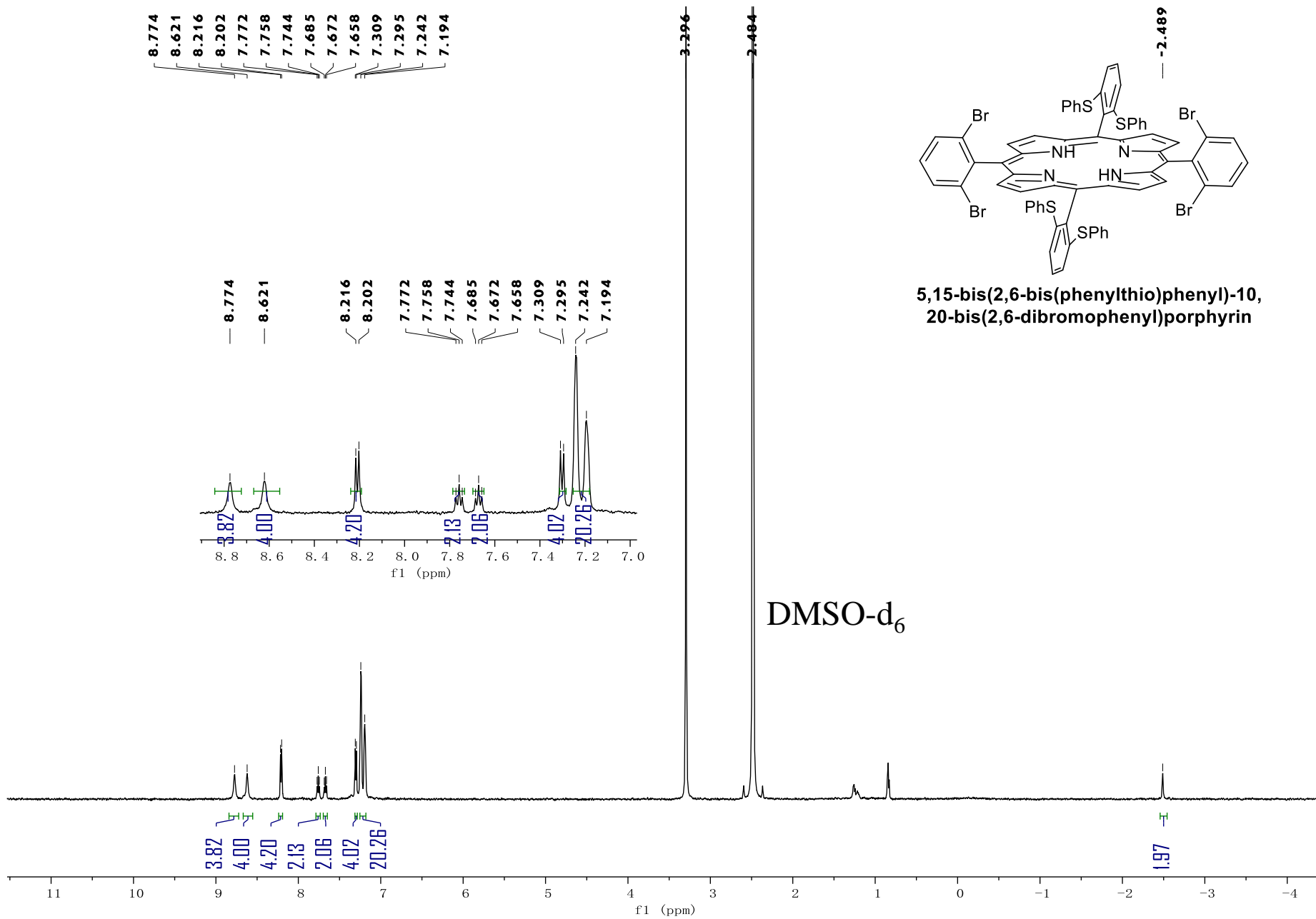
3.885
3.756



¹³C NMR



¹H NMR

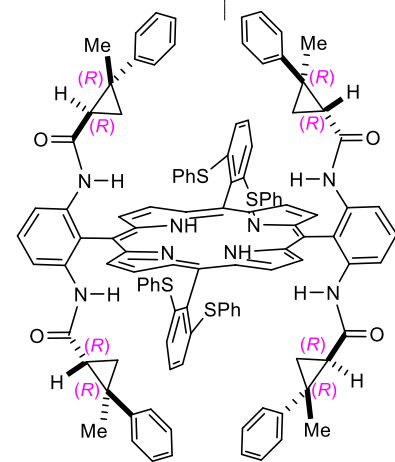


¹H NMR

8.960
8.952
8.914
8.512
7.910
7.511
7.498
7.484
7.273
7.268
7.259
7.253
7.248
7.243
7.230
7.218
7.213
7.205
7.199
7.183
6.833
6.330
6.137
6.012

1.119
0.718
0.539
0.359

-2.061



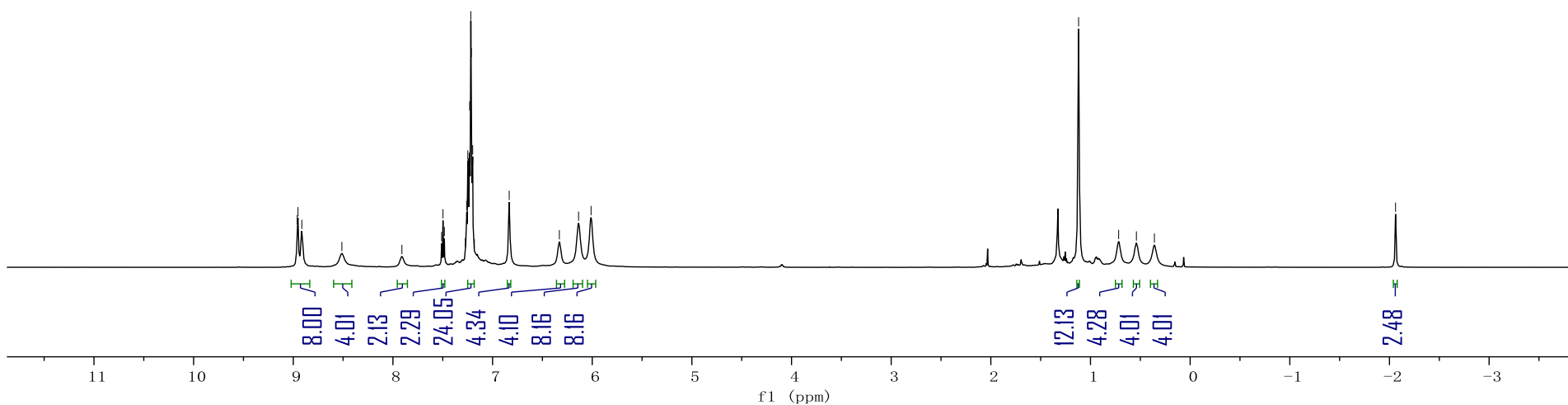
[H₂(P6)]

low solubility in CDCl₃

8.00
4.01
2.13
2.29
24.05
4.34
4.10
8.16
8.16

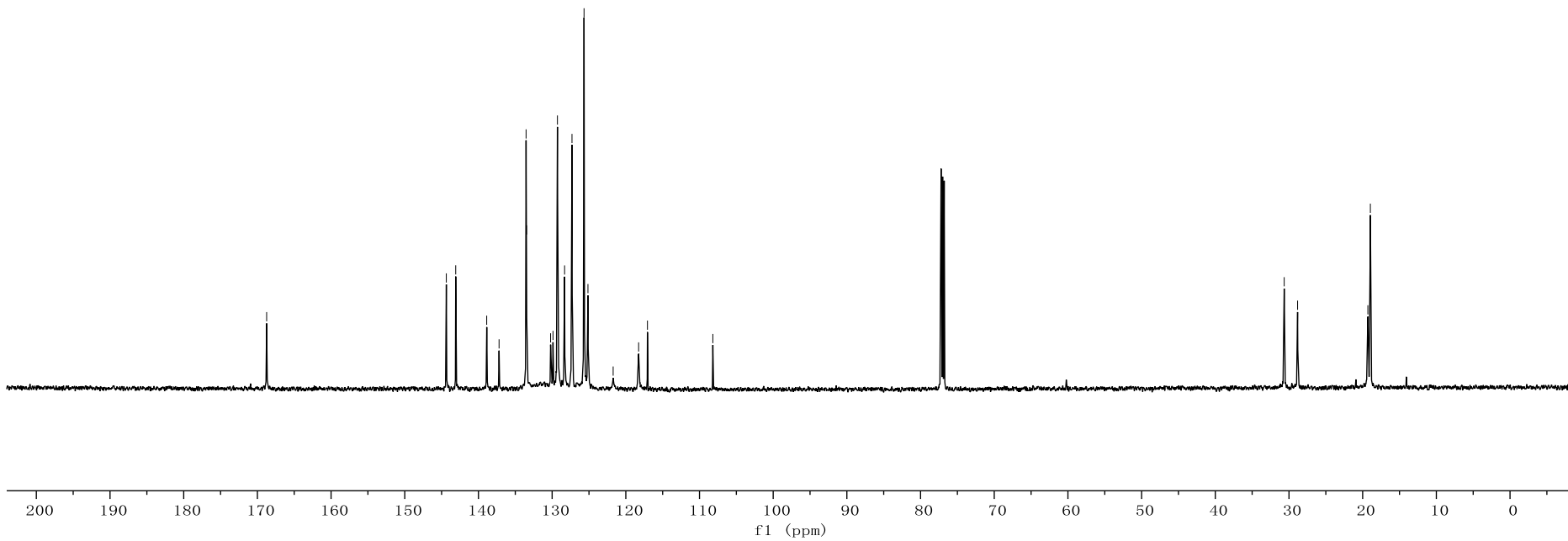
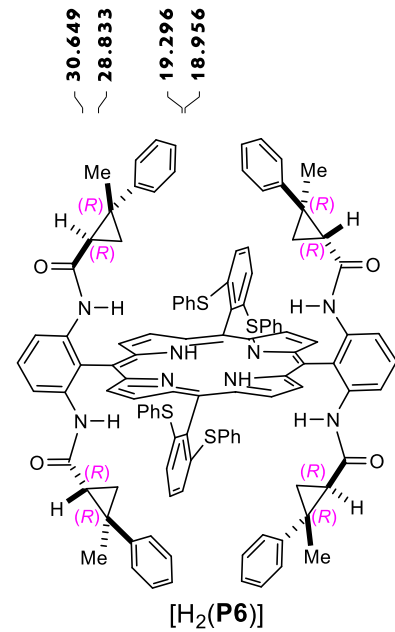
12.13
4.28
4.01
4.01

2.48



¹³C NMR

168.744
144.361
143.085
138.878
137.202
133.536
133.467
130.209
129.882
129.282
128.309
127.306
125.669
125.138
121.717
118.262
117.064
108.189

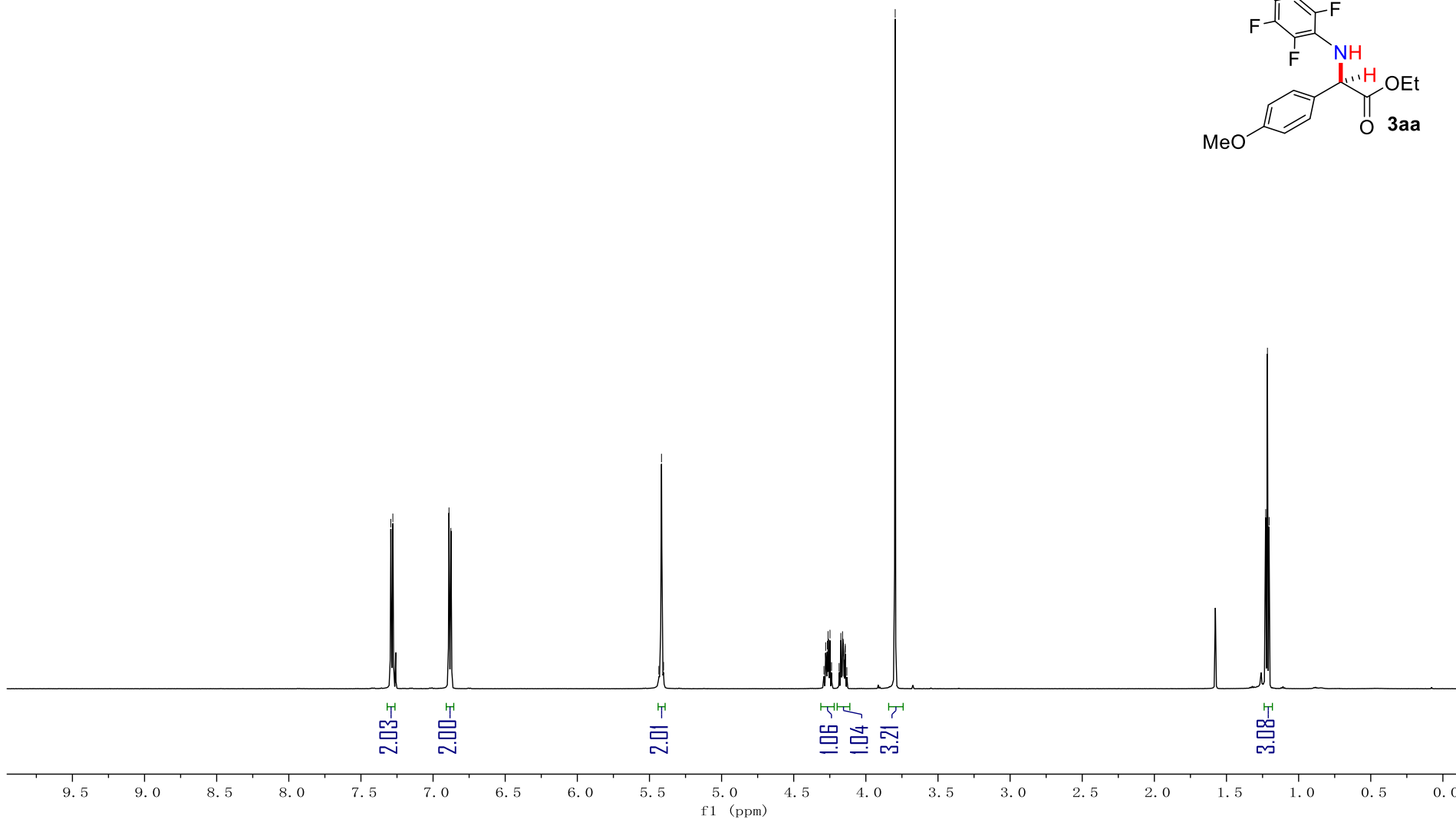
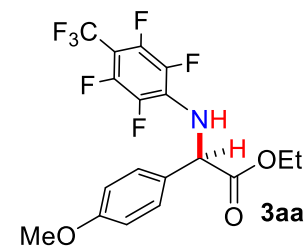


¹H NMR

7.293
7.279
6.890
6.877

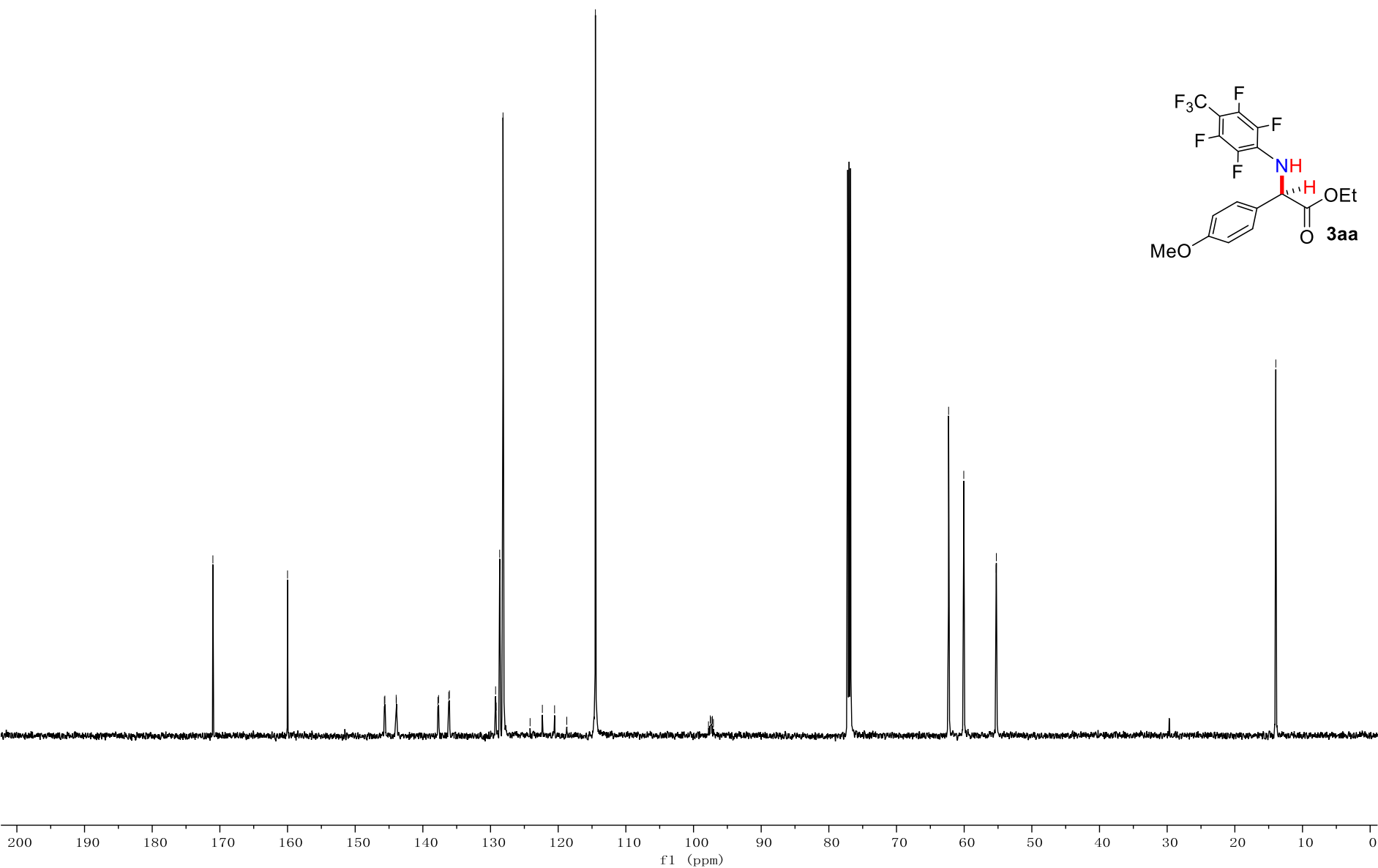
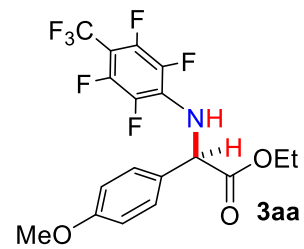
5.435
5.417
5.402
4.290
4.278
4.272
4.267
4.262
4.256
4.256
4.249
4.238
4.185
4.173
4.162
4.156
4.155
4.150
4.144
4.143
4.132
3.797

1.228
1.216
1.204

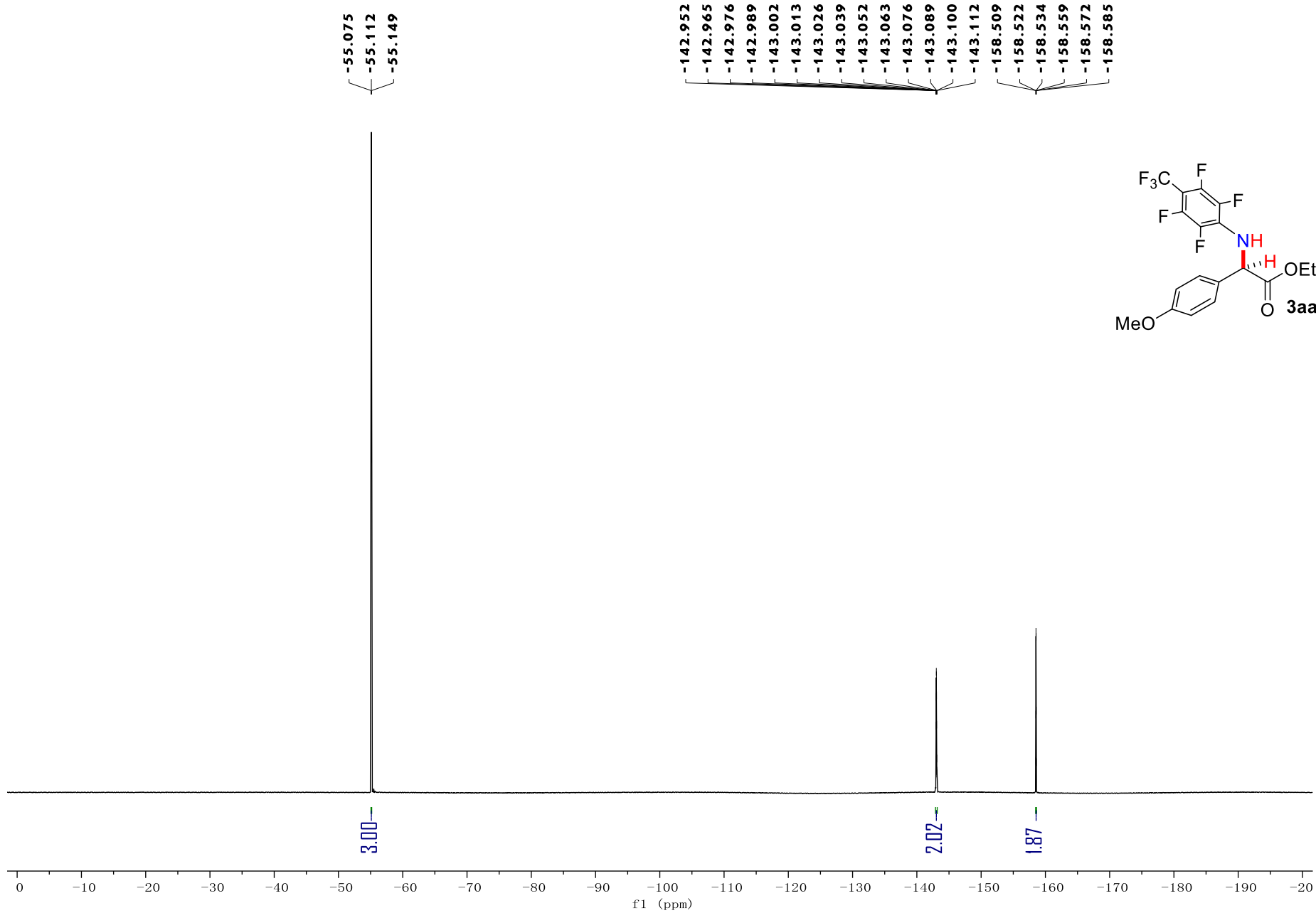


¹³C NMR

- 171.030
- 159.998
- 145.686
- 145.613
- 143.944
- 143.914
- 137.780
- 137.675
- 136.178
- 136.072
- 129.320
- 129.251
- 129.173
- 128.647
- 128.153
- 124.154
- 122.340
- 120.532
- 118.728
- 114.497
- 97.789
- 97.520
- 97.379
- 97.203
- 97.059
- 62.303
- 60.088
- 60.060
- 60.033
- 55.268
- 55.247
- 13.941



¹⁹F NMR

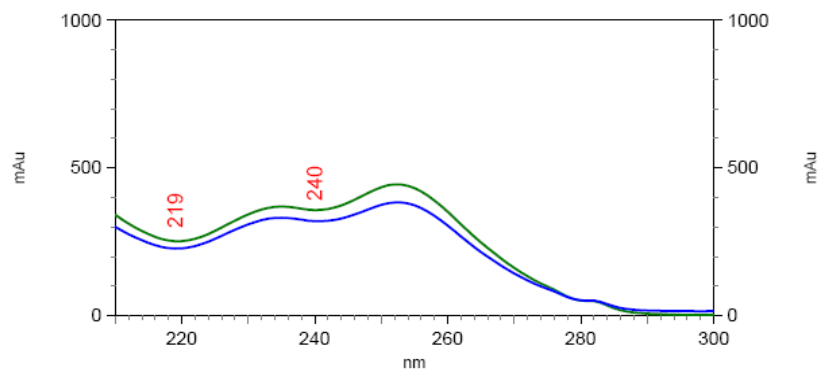
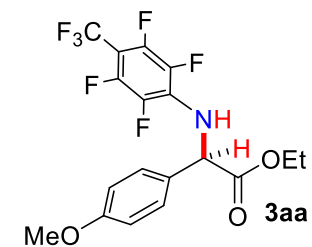
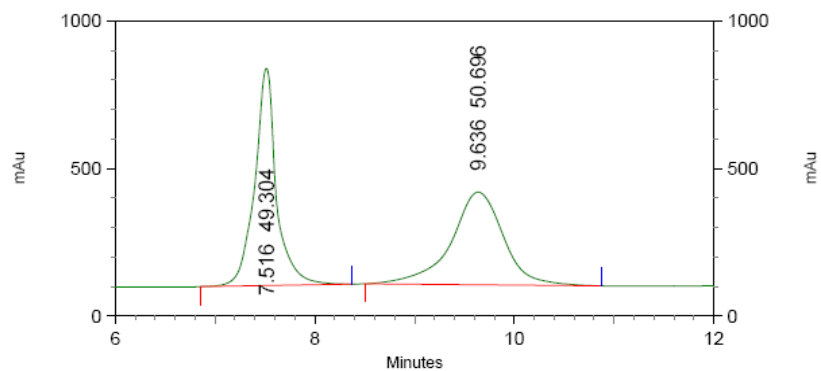


HPLC

JLM-II-198-2pure-WHELK-0.5@-1mL

C:\EZStart\Projects\Default\Method\yang-ODH 20%0.7ml premix.met

C:\EZStart\Projects\Default\Data\JLM-II-198-2pure-WHELK0.5@1ml



10: 259 nm, 4 nm

Results

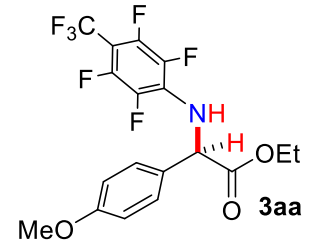
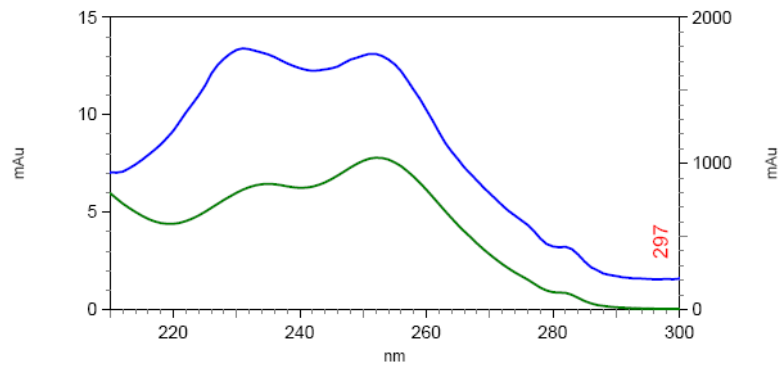
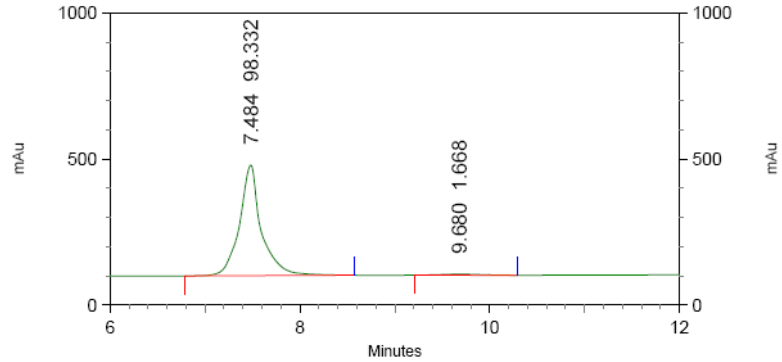
Pk #	Name	Retention Time	Area Percent
1		7.516	49.304
2		9.636	50.696
Totals			100.000

HPLC

JLM-V-122-3a1-WHELK-0.5@-1mL

C:\EZStart\Projects\Default\Method\yang-ODH 20%0.7ml premix.met

C:\EZStart\Projects\Default\Data\JLM-V-122-3a1-WHELK0.5@1ml

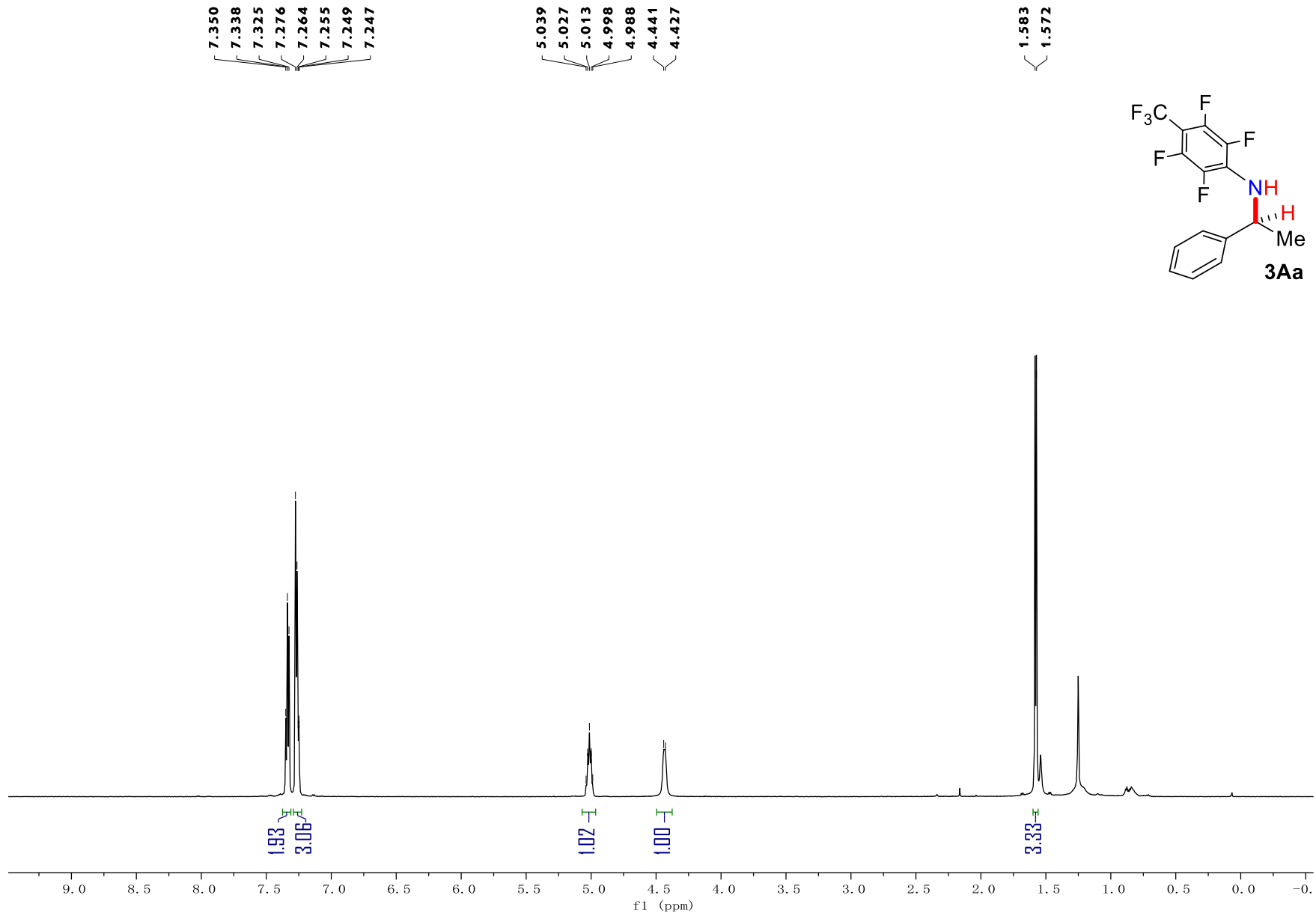


15: 270 nm, 4 nm

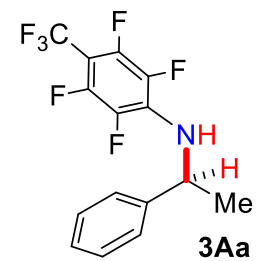
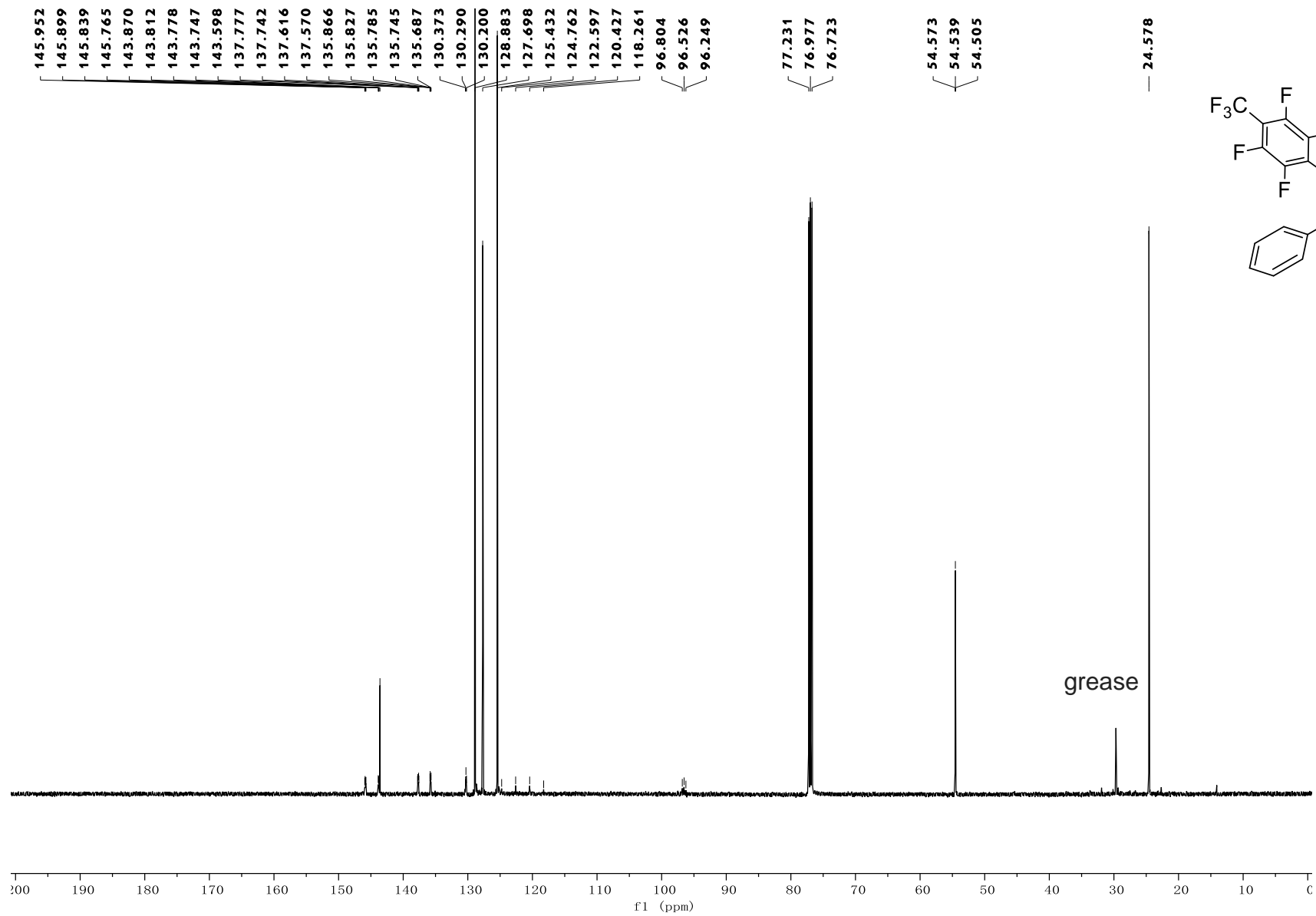
Results

Pk #	Name	Retention Time	Area Percent
1		7.484	98.332
2		9.680	1.668
Totals			100.000

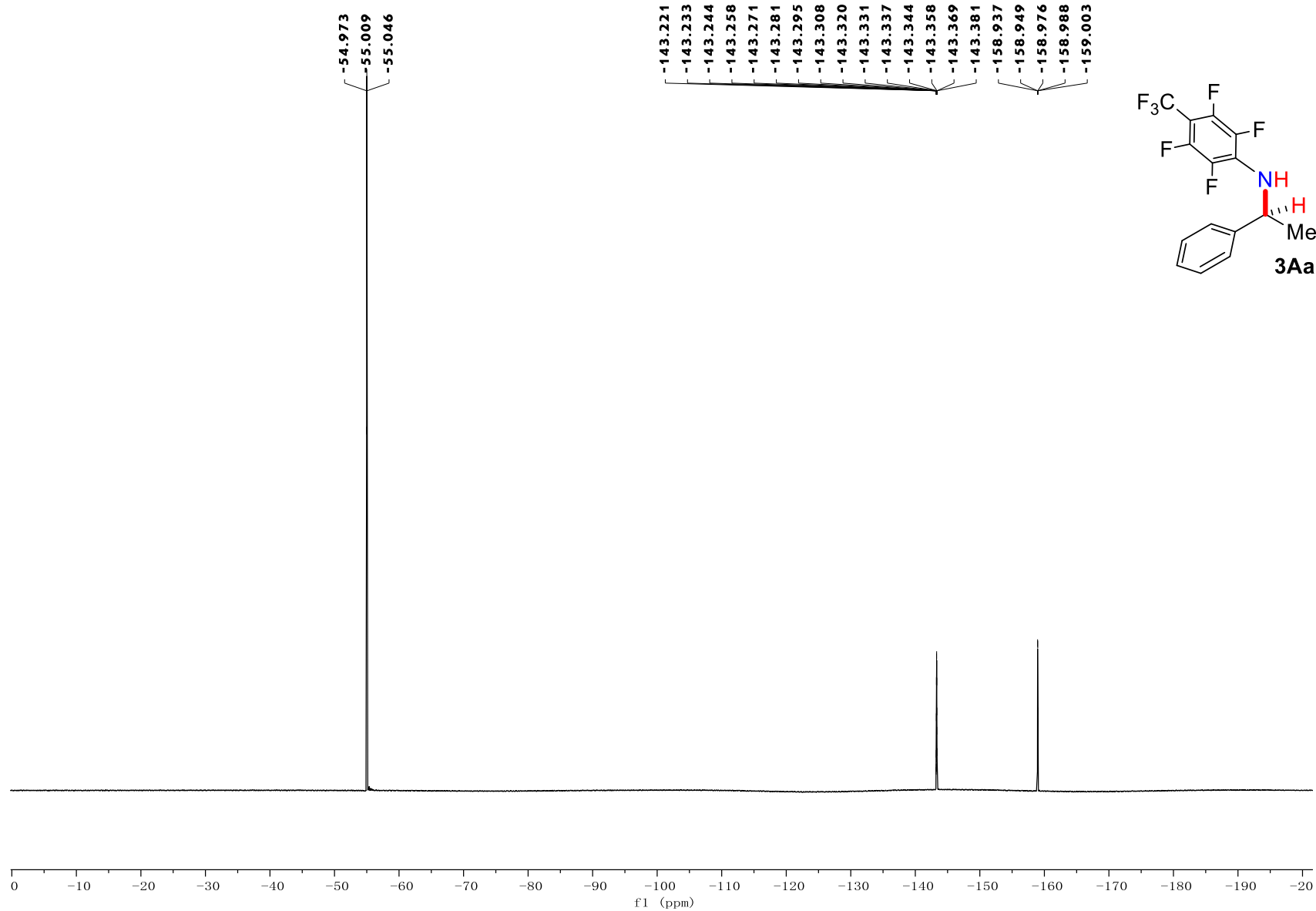
^1H NMR



¹³C NMR

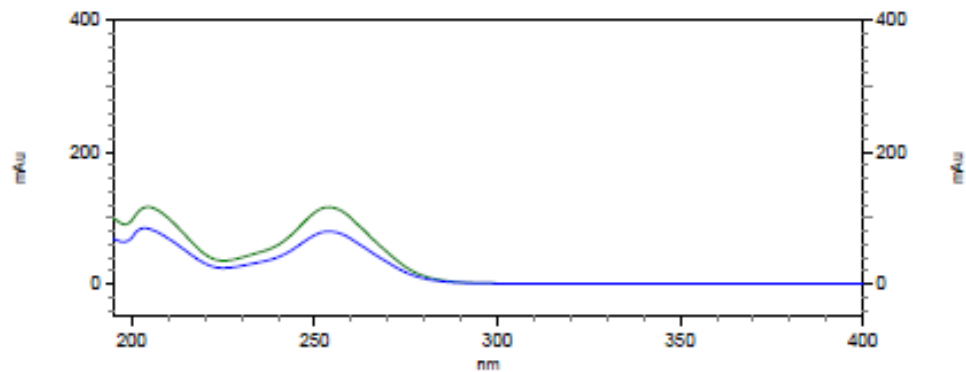
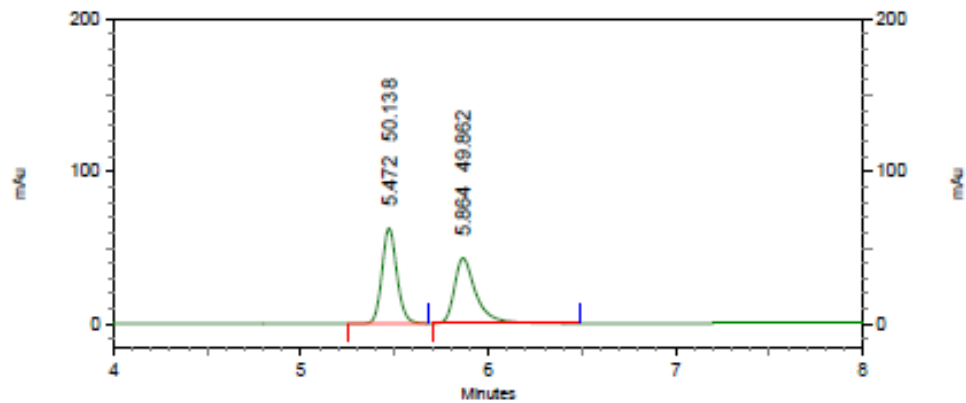


^{19}F NMR



HPLC

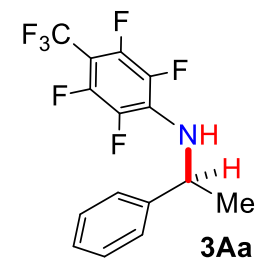
C:\EZStart\Projects\Default\Data\POX-1937-24-IA-0.8ml-PURE-1
C:\Documents and Settings\zhang\Desktop\DSW\Report-standard0311.met



4: 267 nm, 4
nm Results

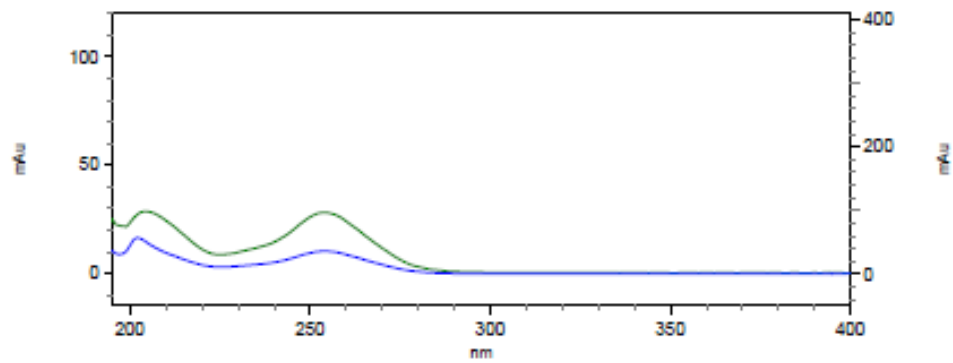
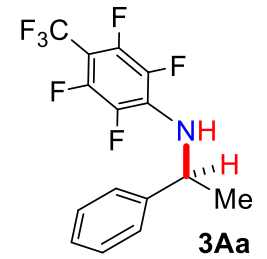
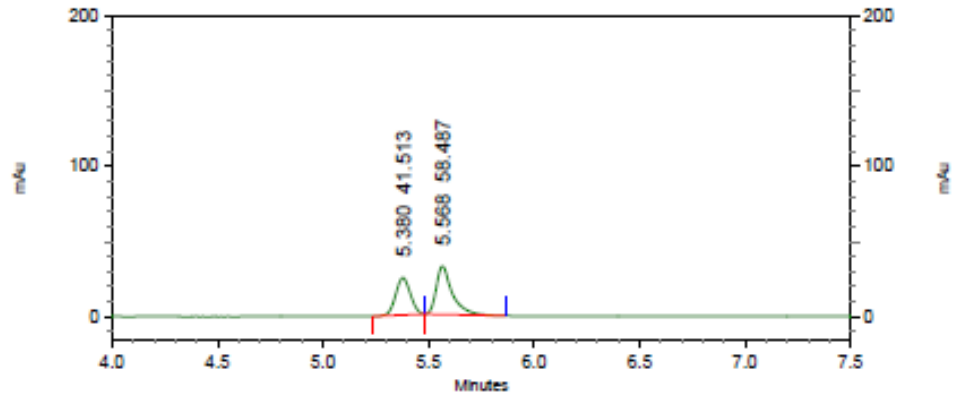
Pk #	Retention Time	Area Percent
1	5.472	50.138
2	5.864	49.862

Totals	100.000
--------	---------



HPLC

C:\EZStart\Projects\Default\Data\POX-1938-2%-IA-0.8ml
C:\Documents and Settings\zhang\Desktop\DSW\Report-standard0311.met



4: 250 nm, 4
nm Results

Pk #	Retention Time	Area Percent
1	5.380	41.513
2	5.568	58.487
Totals		100.000

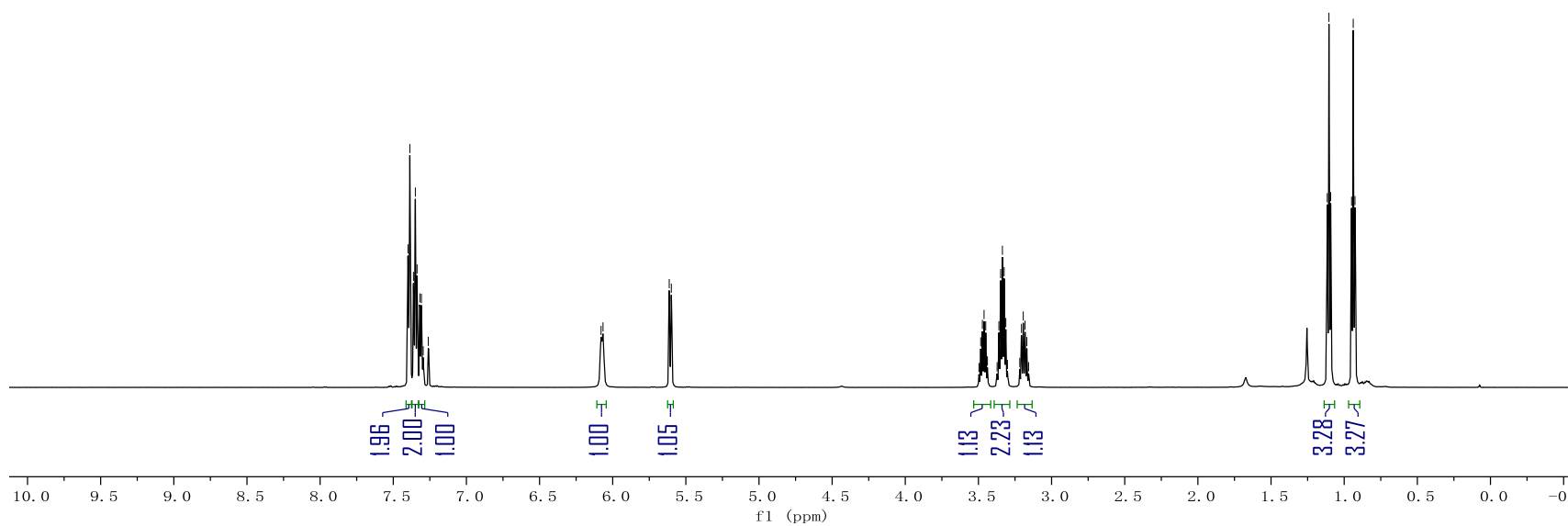
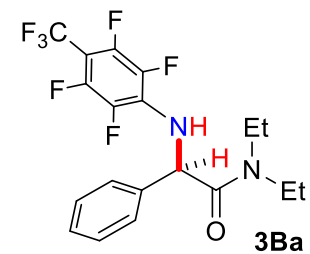
¹H NMR

7.399
7.387
7.362
7.349
7.337
7.318
7.307
7.296
7.260

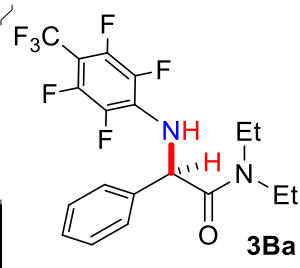
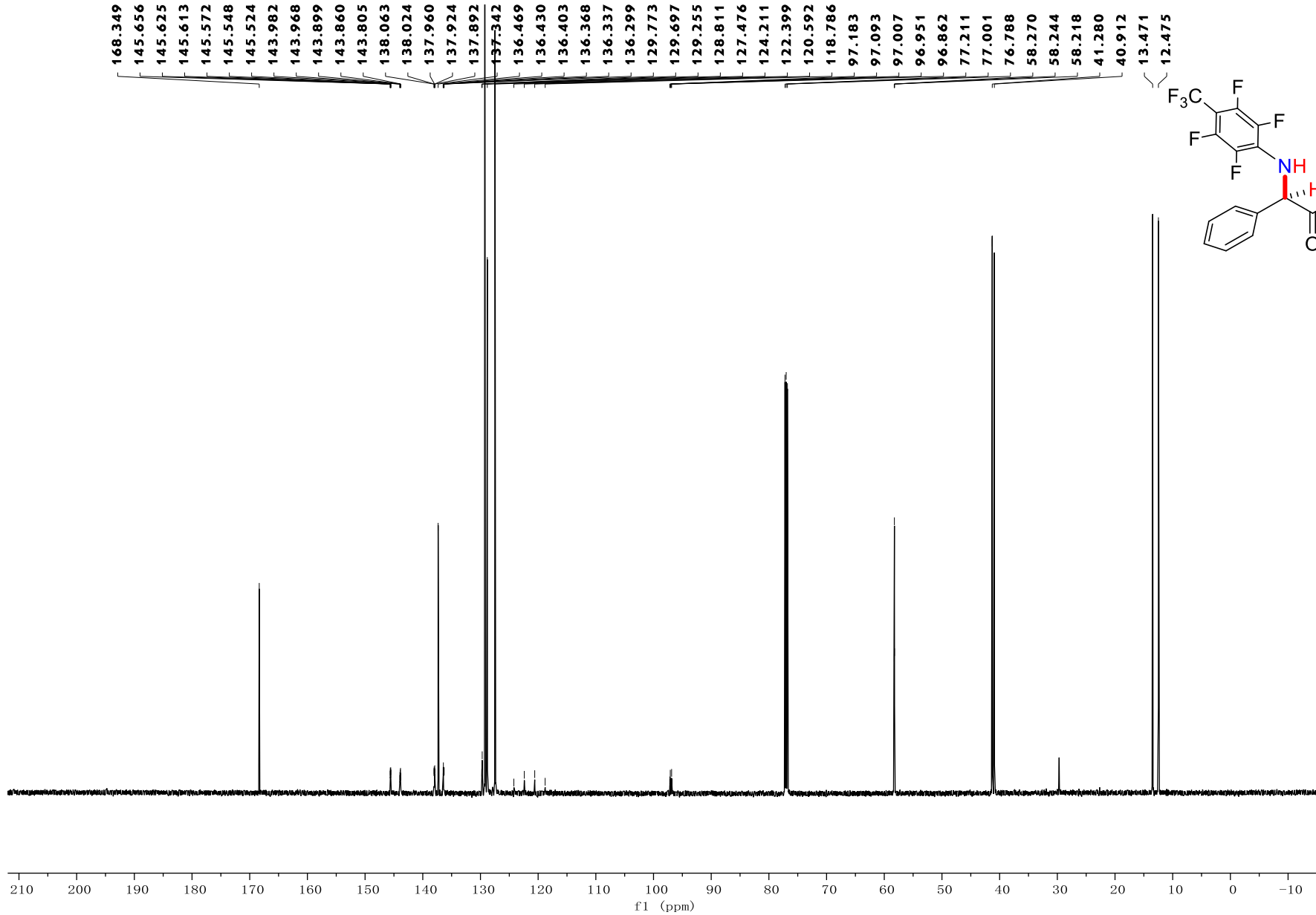
6.079
6.066
5.614
5.599

3.496
3.485
3.473
3.462
3.450
3.438
3.372
3.360
3.348
3.336
3.324
3.313
3.302
3.217
3.206
3.194
3.181
3.169
3.157

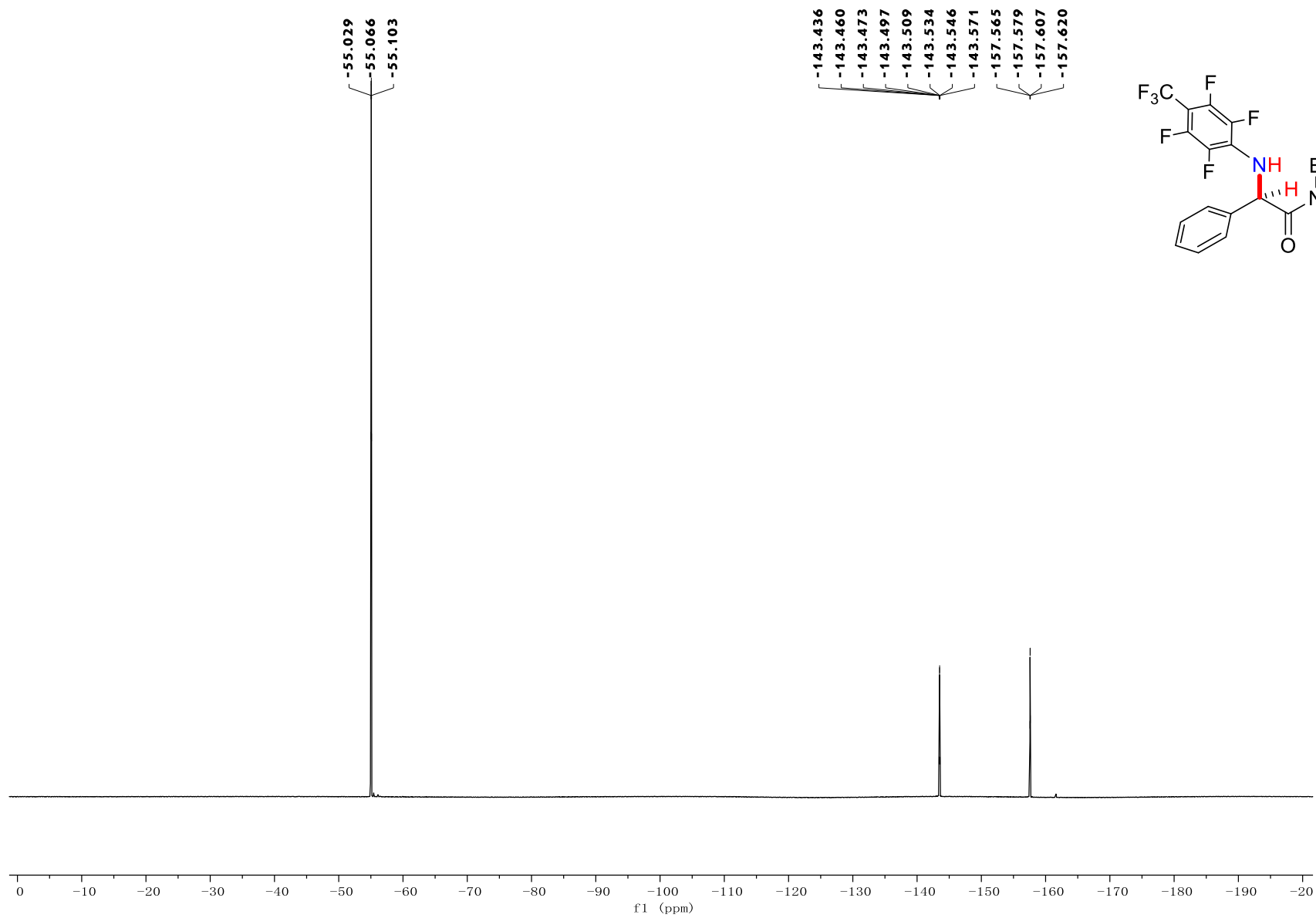
1.116
1.104
1.092
0.950
0.938
0.926



¹³C NMR

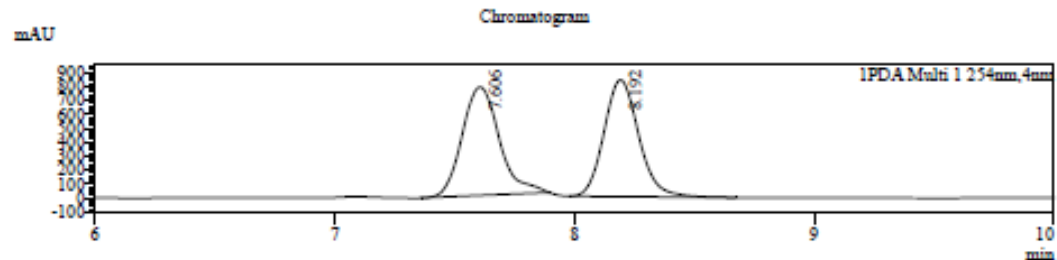


^{19}F NMR

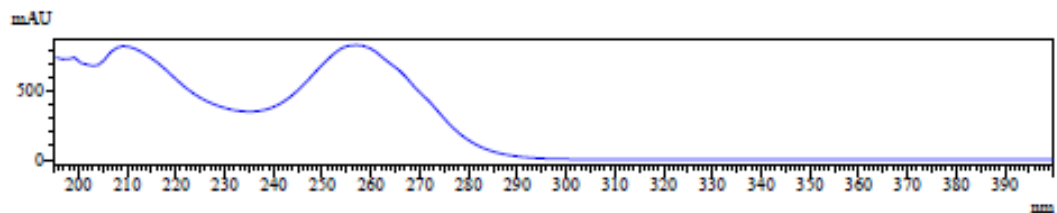


HPLC

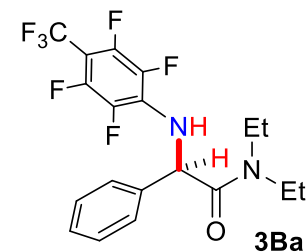
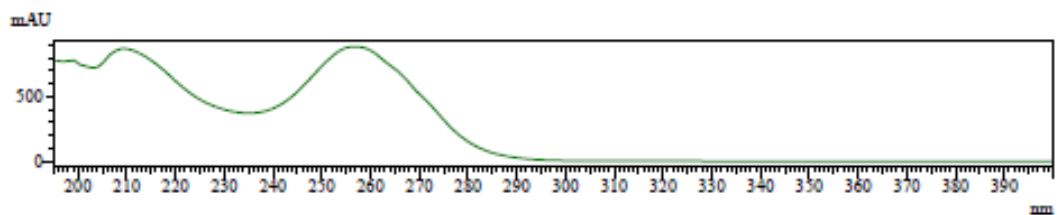
Sample Information
Sample Name : POX-1948-IA-20%-0.5mL
Sample ID : POX-1948-IA-20%-0.5mL
Data File : POX-1948-IA-20%-0.5mL.002.lcd
Method File : POX-20%.0.5 mL-50min.lcm



UV Spectrum
Retention time = 7.606



UV Spectrum
Retention time = 8.192



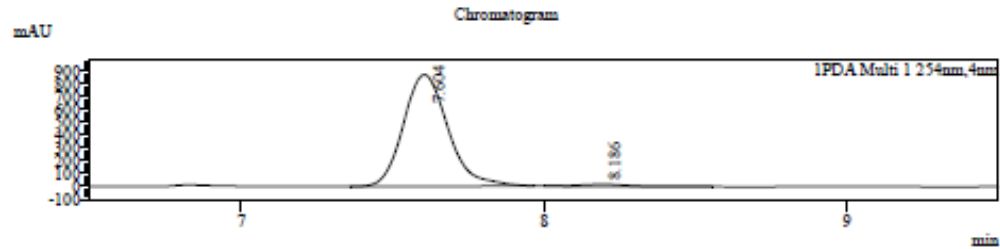
Peak Table

PDA Ch1 254nm

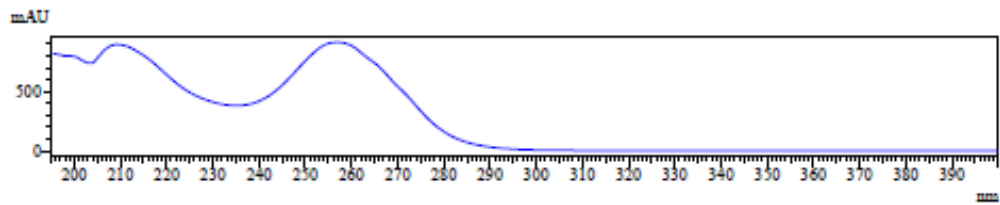
Peak#	Ret. Time	Area	Area%
1	7.606	8402484	49.855
2	8.192	8451195	50.145
Total		16853680	100.000

HPLC

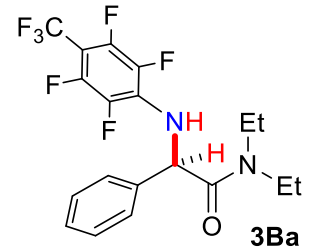
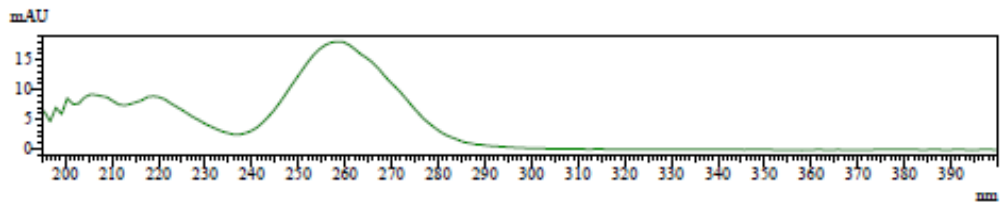
Sample Information
Sample Name : POX-1949-IA-20%-0.5mL
Sample ID : POX-1949-IA-20%-0.5mL
Data File : POX-1949-IA-20%-0.5mL.lcd
Method File : POX-20%.0.5.mL-50min.lcm



UV Spectrum
Retention time = 7.604



UV Spectrum
Retention time = 8.186



Peak Table

Peak#	Ret. Time	Area	Area%
1	7.604	8942684	98.504
2	8.186	135809	1.496
Total		9078493	100.000

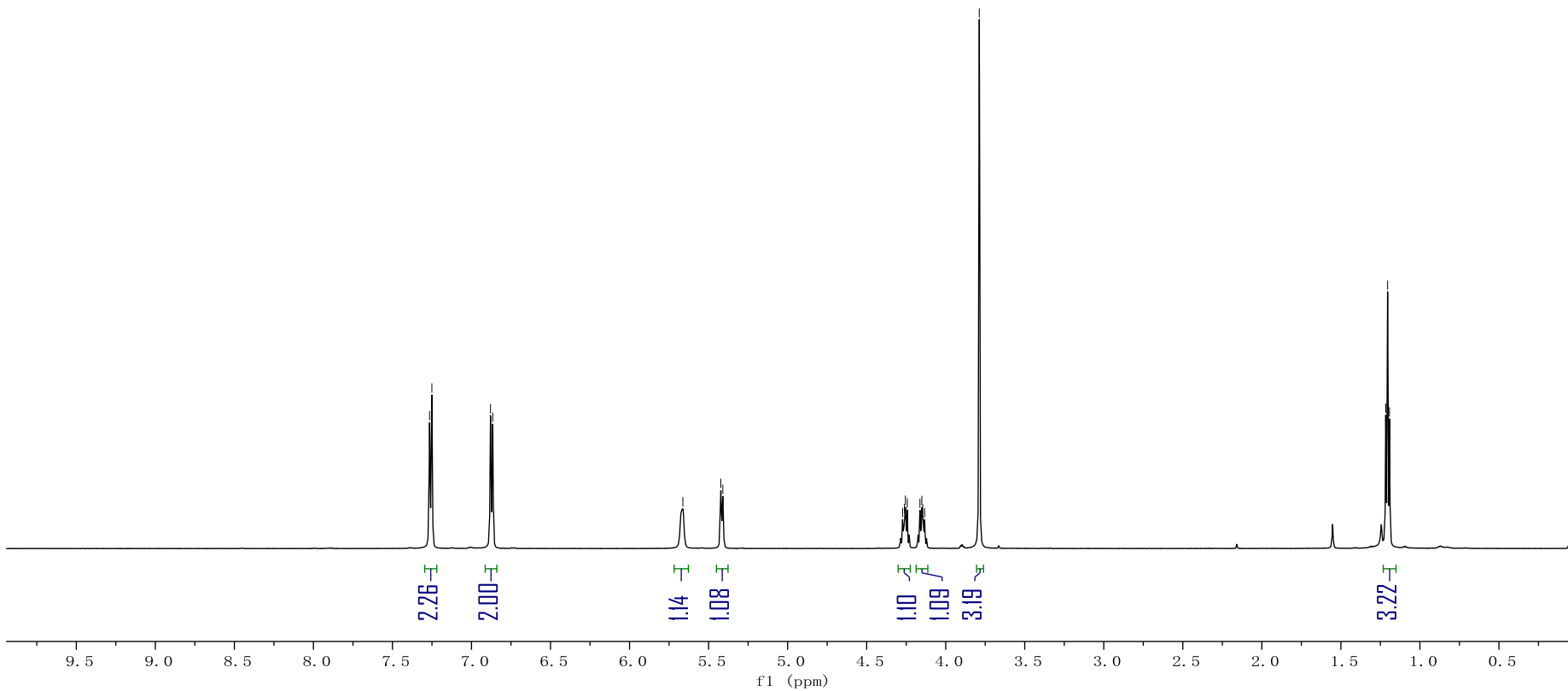
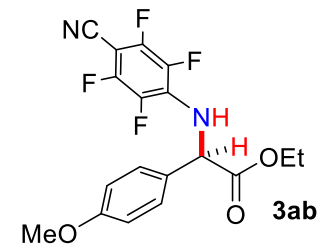
¹H NMR

7.265
7.251
6.880
6.865

5.663
5.422
5.410

4.273
4.261
4.255
4.243
4.163
4.151
4.145
4.133
3.787

1.216
1.204
1.192



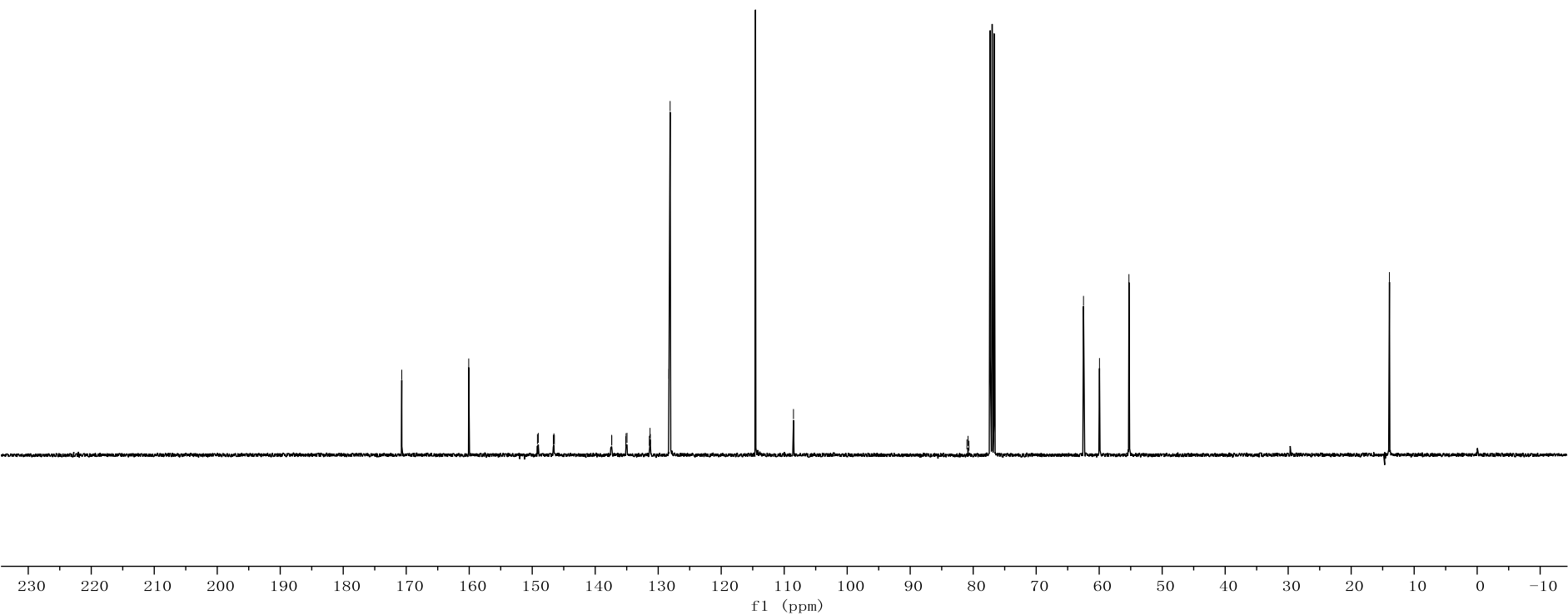
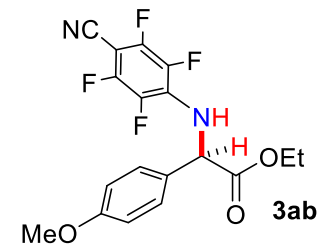
¹³C NMR

170.707
160.080
149.174
149.036
146.652
146.559
146.479
137.386
137.372
135.160
135.099
134.952
131.407
131.295
131.256
128.297
128.108
114.555
108.563
108.527
108.492

80.997
80.821
80.665

62.489
59.953
55.275

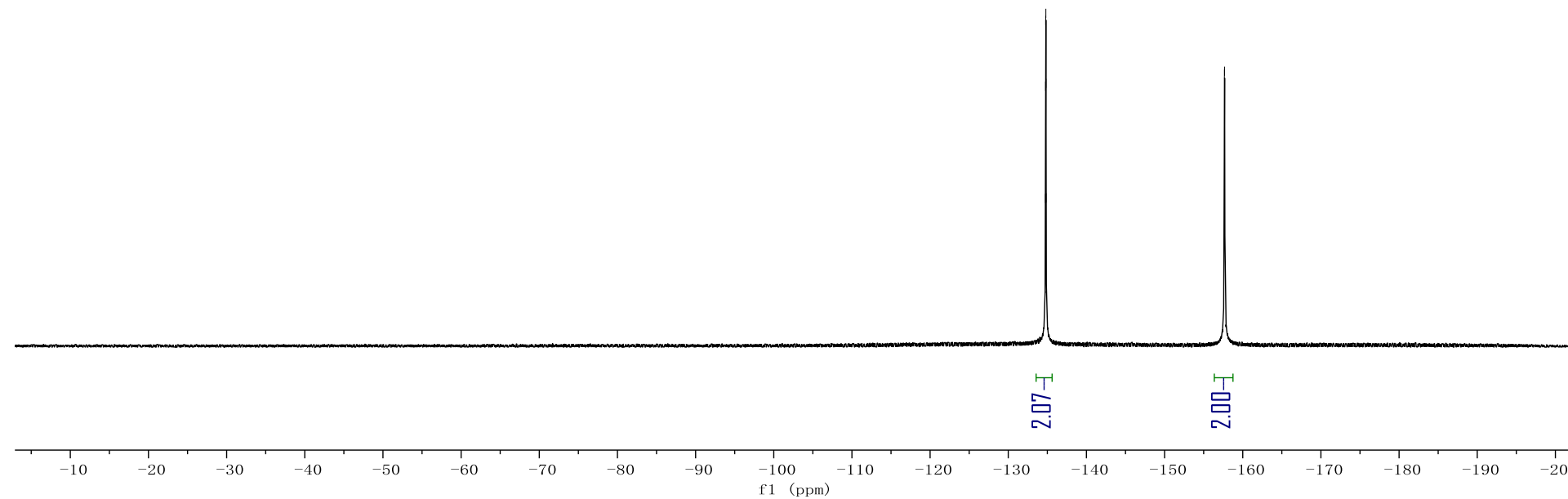
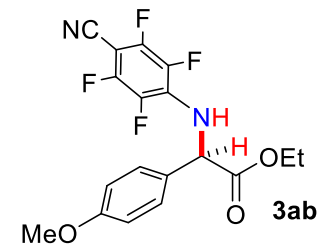
13.927



^{19}F NMR

-134.758
-134.775
-134.786
-134.811
-134.823
-134.840

-157.653
-157.682



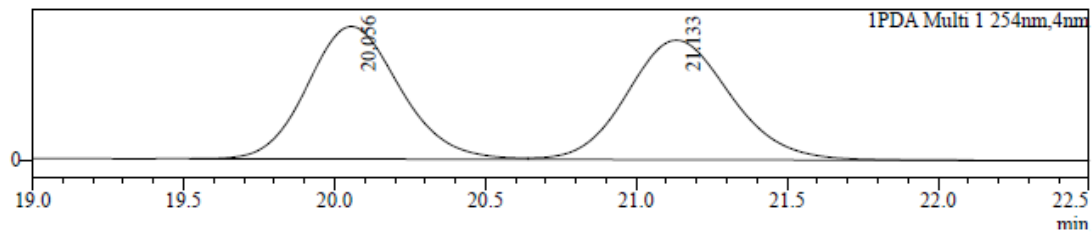
HPLC

Sample Information

Sample Name : P0X-0809-IC10%-0.8MI
Sample ID : P0X-0809-IC10%-0.8MI
Data File : P0X-0809-IC10%-0.8MI.lcd
Method File : P0X-10%-0.8ml.lcm

Chromatogram

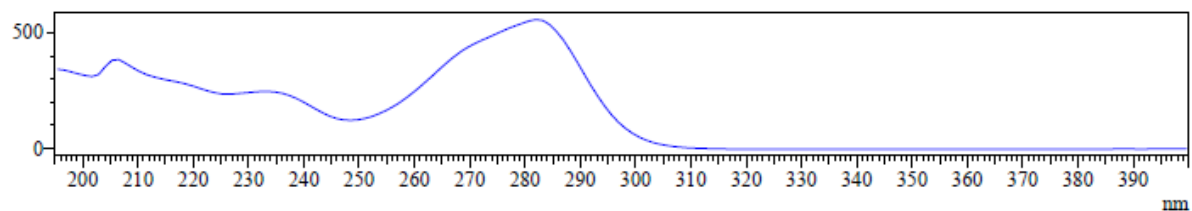
mAU



UV Spectrum

Retention time = 20.056

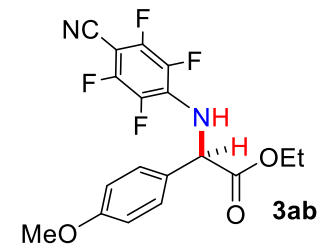
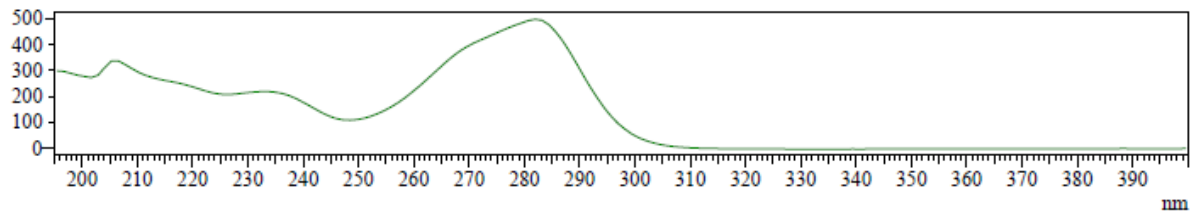
mAU



UV Spectrum

Retention time = 21.133

mAU



Peak Table

PDA Ch1 254nm

Peak#	Ret. Time	Area	Area%
1	20.056	3388030	49.763
2	21.133	3420340	50.237
Total		6808370	100.000

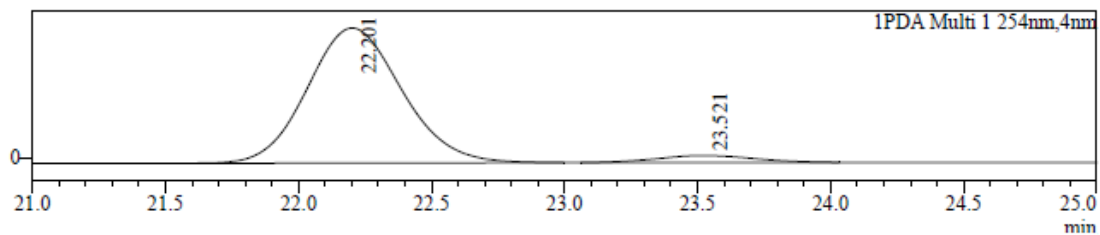
HPLC

Sample Information

Sample Name : P0X-0810-IC10%-0.8MI
Sample ID : P0X-0810-IC10%-0.8MI
Data File : P0X-0810-IC10%-0.8MI.lcd
Method File : P0X-10%-0.8ml.lcm

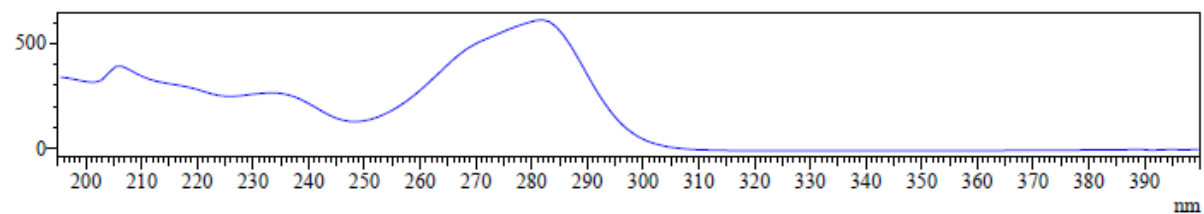
Chromatogram

mAU



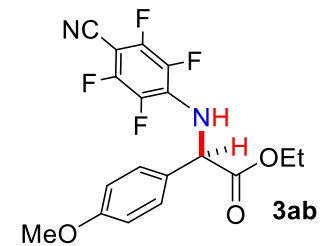
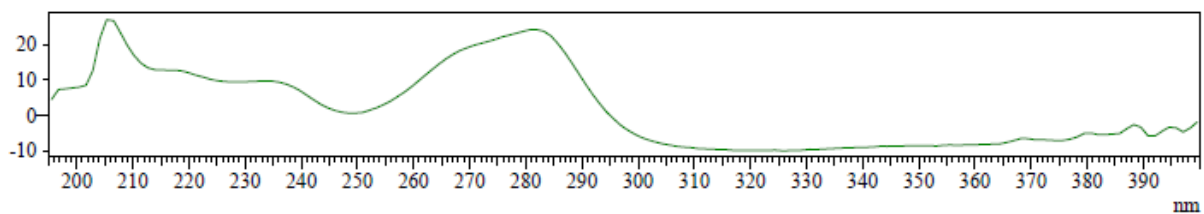
UV Spectrum
Retention time = 22.201

mAU



UV Spectrum
Retention time = 23.521

mAU

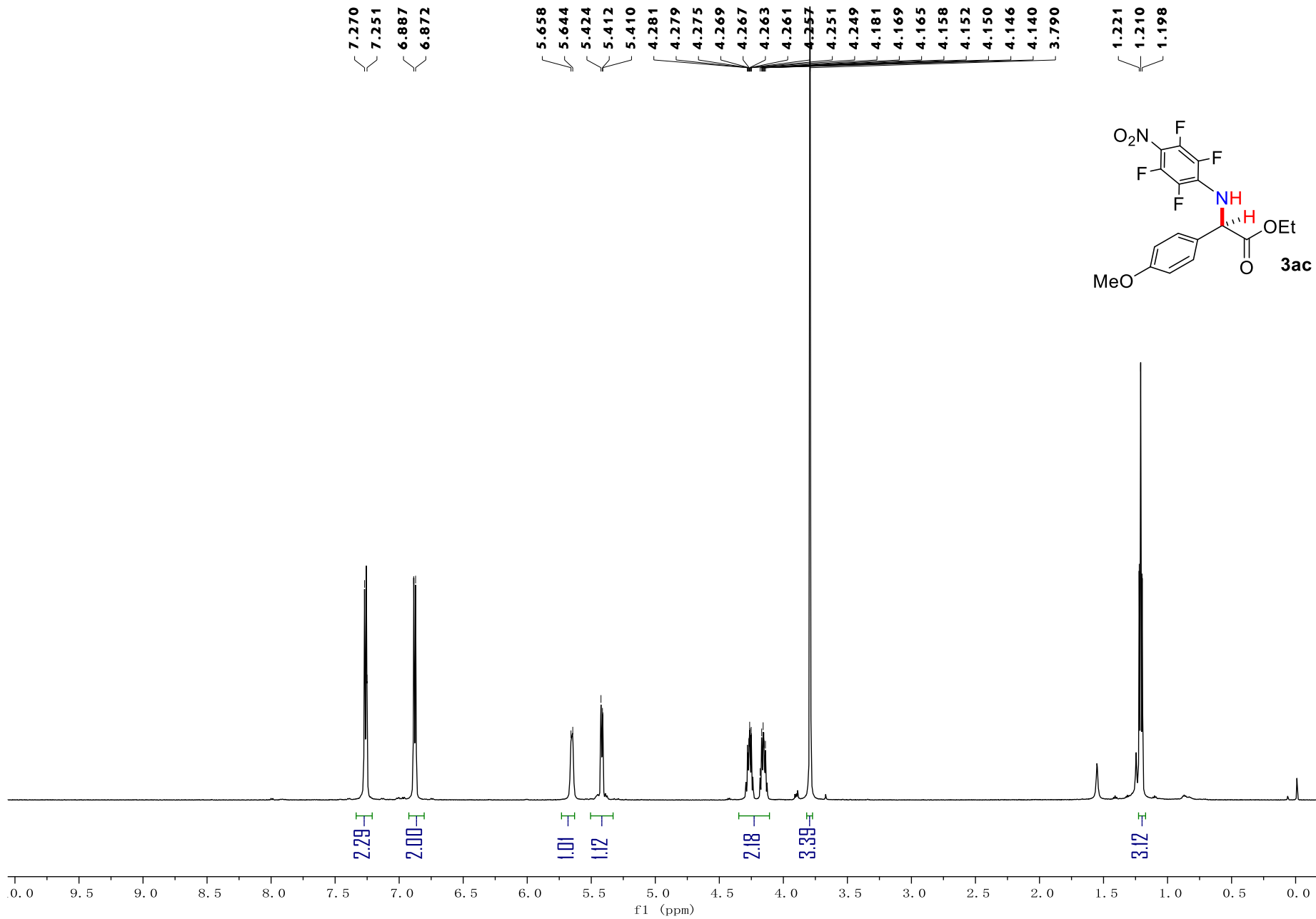


Peak Table

PDA Ch1 254nm

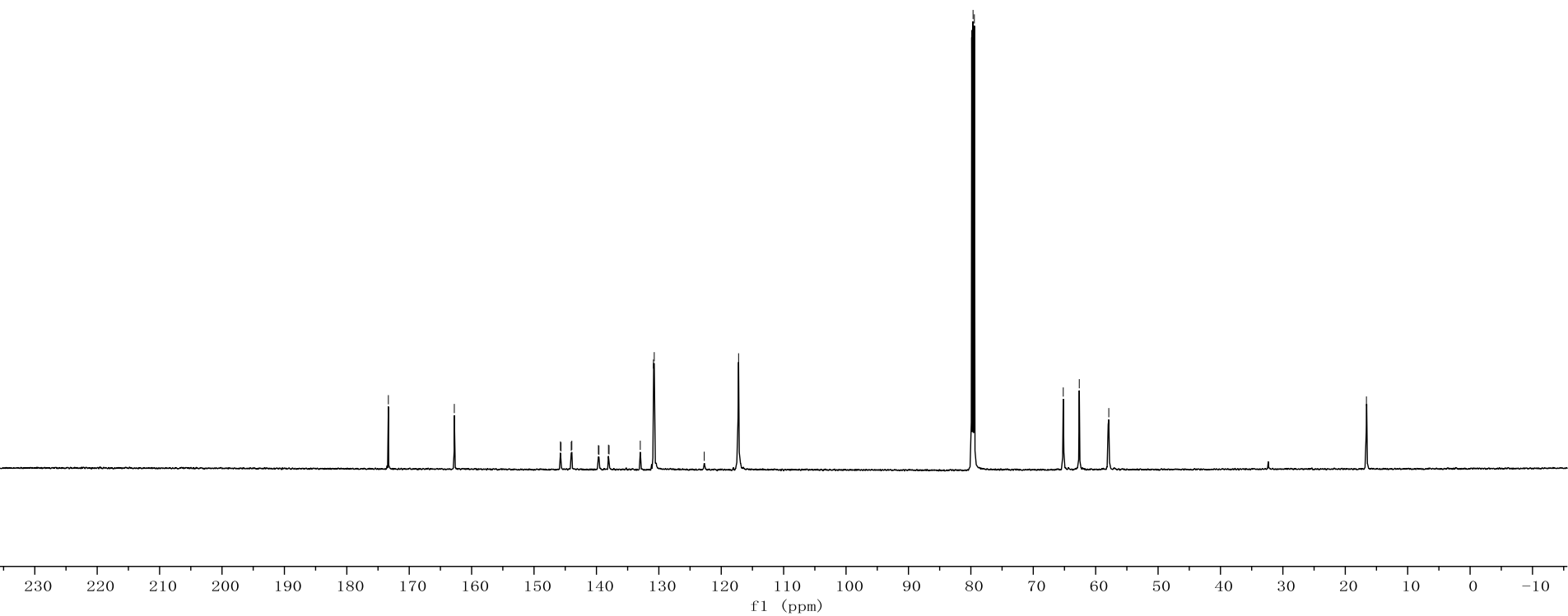
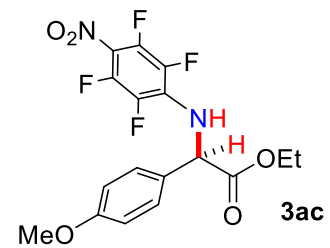
Peak#	Ret. Time	Area	Area%
1	22.201	4481981	95.079
2	23.521	231967	4.921
Total		4713948	100.000

¹H NMR



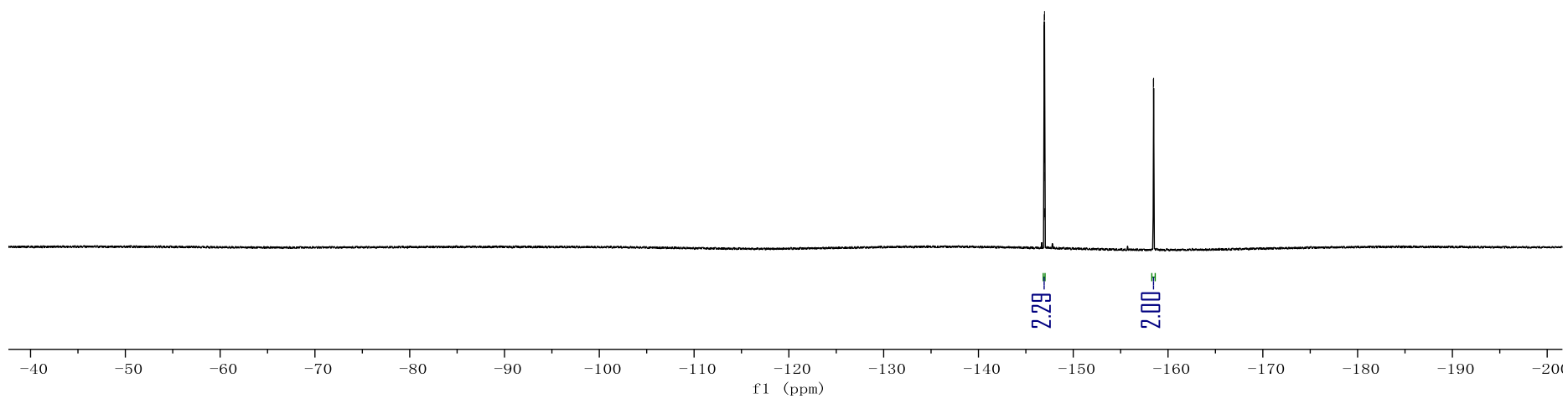
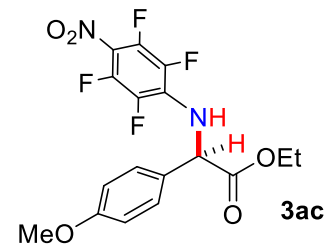
¹³C NMR

173.340
162.775
145.771
145.674
144.039
143.941
139.687
139.596
138.075
137.977
132.973
130.864
130.735
122.724
117.221
79.862
79.649
79.438
65.194
62.627
57.896
16.610



^{19}F NMR

-146.914
-146.929
-146.941
-146.970
-146.981
-146.997
-158.449
-158.480



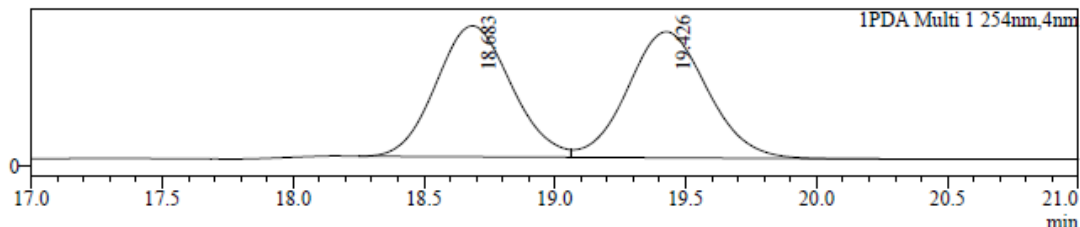
HPLC

Sample Information

Sample Name : P0X-0721-IC-10%-0.8mL
Sample ID : P0X-0721-IC-10%-0.8mL
Data File : P0X-0721-IC-10%-0.8mL.lcd
Method File : P0X-10%-0.8ml.lcm

Chromatogram

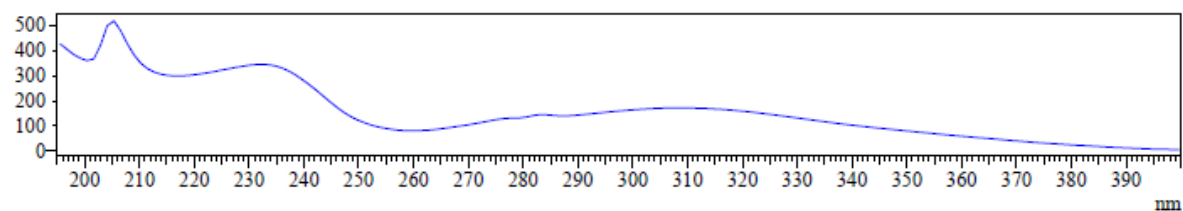
mAU



UV Spectrum

Retention time = 18.683

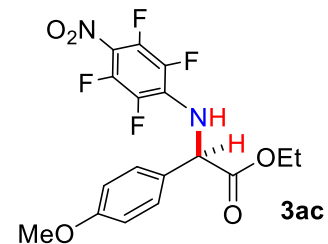
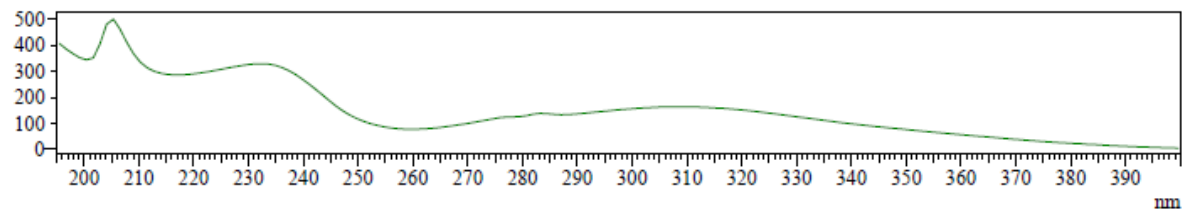
mAU



UV Spectrum

Retention time = 19.426

mAU



Peak Table

PDA Ch1 254nm

Peak#	Ret. Time	Area	Area%
1	18.683	1787988	49.507
2	19.426	1823624	50.493
Total		3611613	100.000

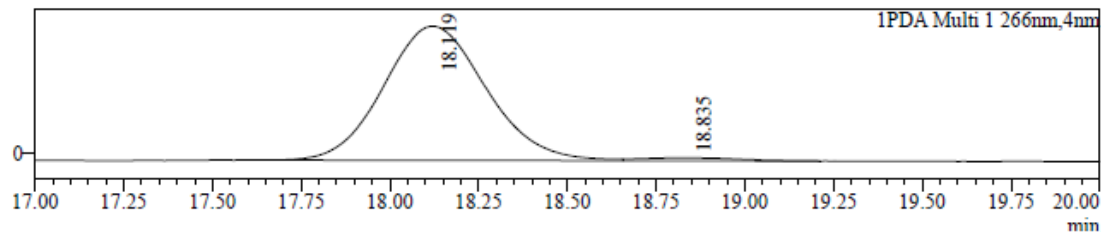
HPLC

Sample Information

Sample Name : P0X-0722-IC-10%-0.8mL
Sample ID : P0X-0722-IC-10%-0.8mL
Data File : P0X-0722-IC-10%-0.8mL.lcd
Method File : P0X-10%-0.8ml.lcm

Chromatogram

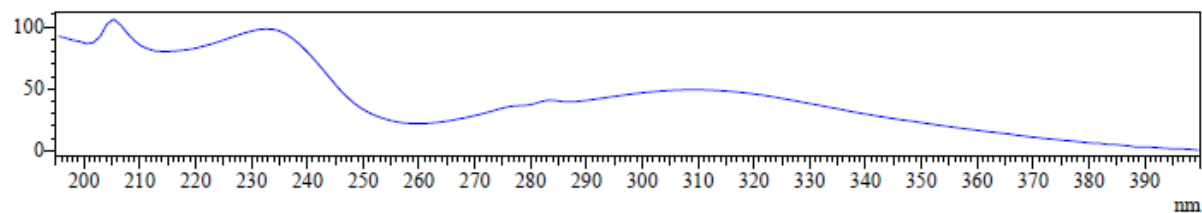
mAU



UV Spectrum

Retention time = 18.119

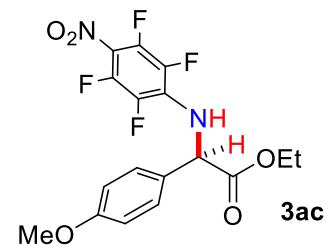
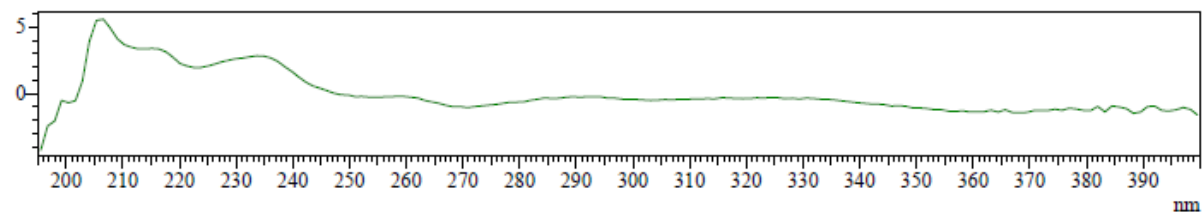
mAU



UV Spectrum

Retention time = 18.835

mAU



Peak Table

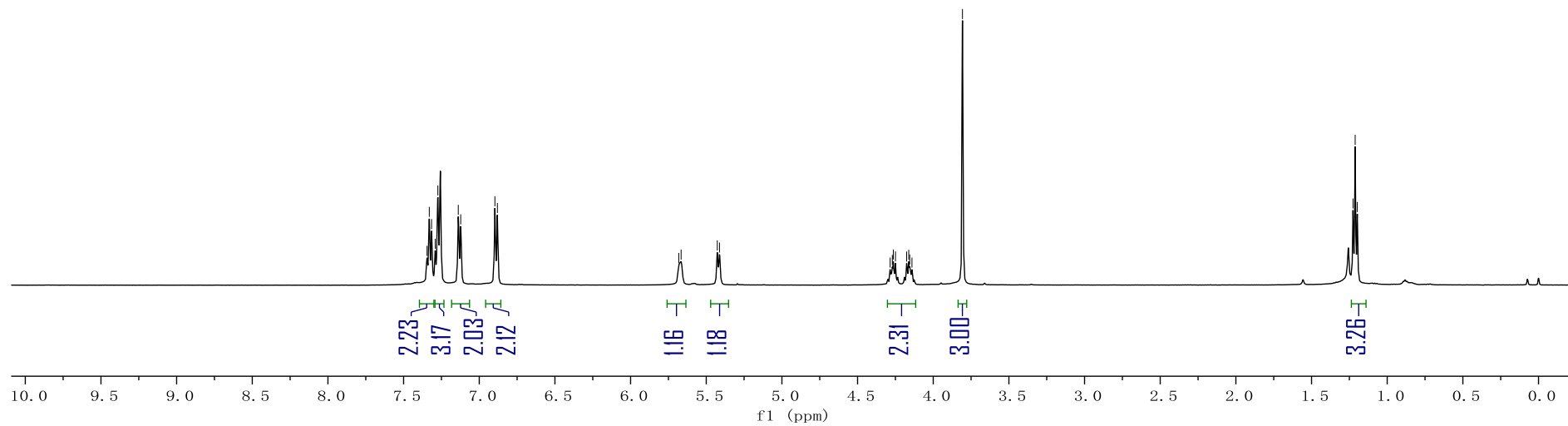
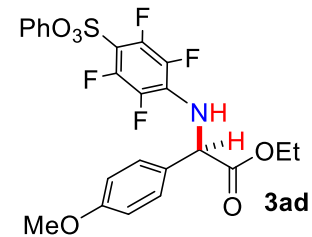
PDA Ch1 266nm

Peak#	Ret. Time	Area	Area%
1	18.119	520416	97.840
2	18.835	11489	2.160
Total		531905	100.000

¹H NMR

7.345
7.330
7.315
7.291
7.274
7.139
7.123
6.897
6.880
5.681
5.666
5.427
5.412
4.285
4.270
4.264
4.250
4.176
4.162
4.155
4.141
3.807

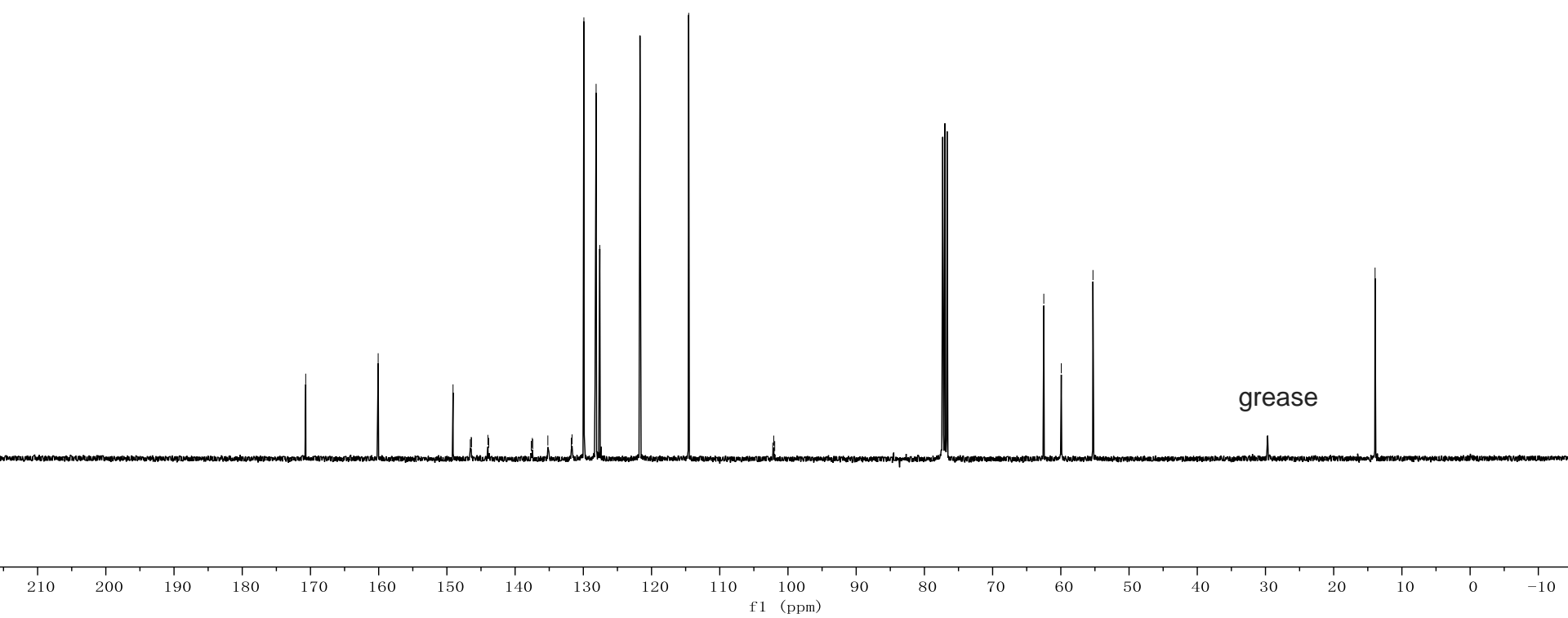
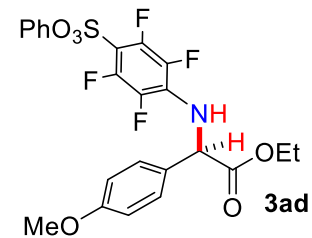
1.226
1.212
1.198



¹³C NMR

170.697
160.075
149.099
146.573
146.440
146.390
143.991
143.865
137.635
137.478
137.421
135.199
131.764
131.653
129.920
128.126
127.588
121.702
114.553
102.230
102.083
101.939
62.495
59.934
55.289

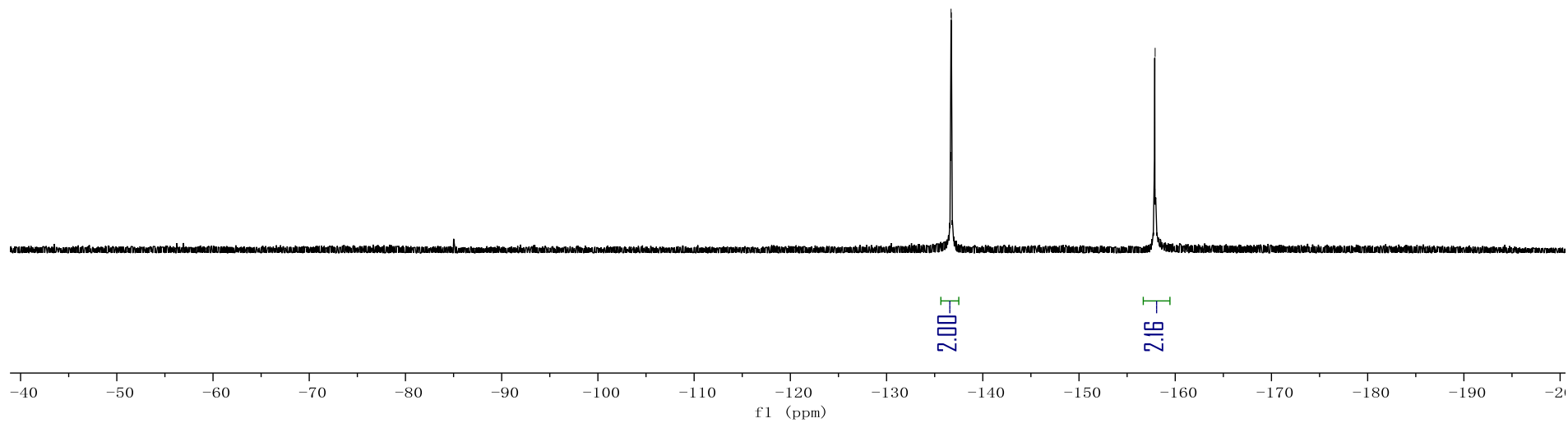
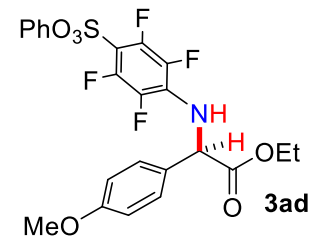
13.933



^{19}F NMR

-136.664
-136.686
-136.702
-136.735
-136.747

-157.891
-157.911



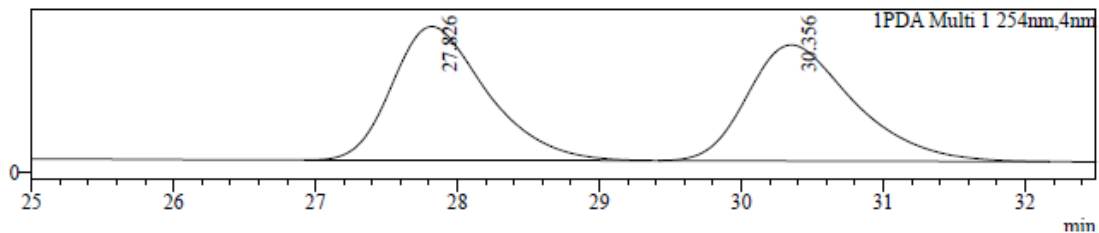
HPLC

Sample Information

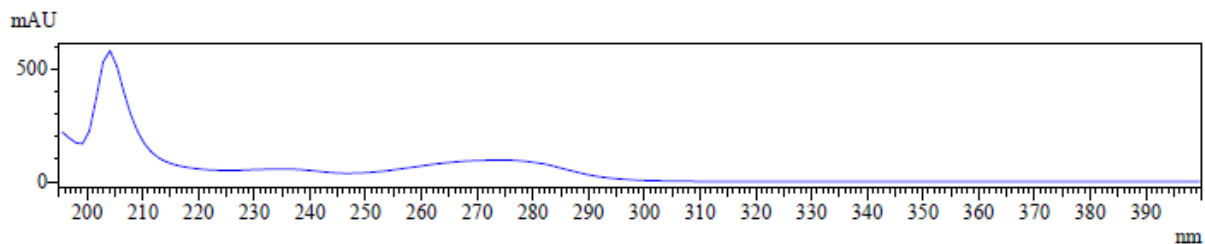
Sample Name : P0X781-ODH-5%-0.8Ml
Sample ID : P0X781-ODH-5%-0.8Ml
Data File : P0X781-ODH-5%-0.8Ml.lcd
Method File : P0X-5.0%-0.8ml.lcm

Chromatogram

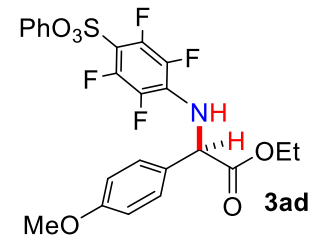
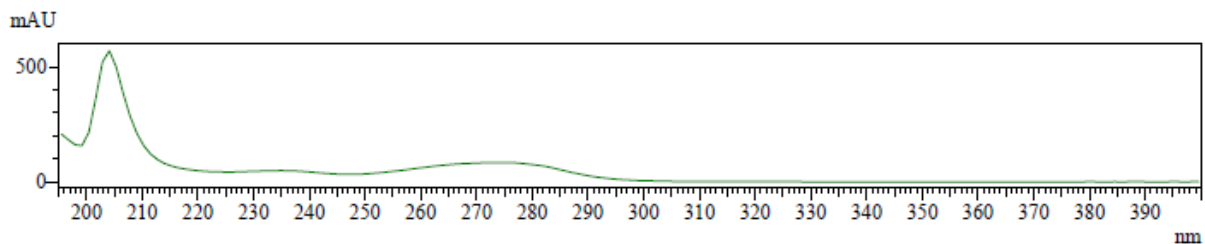
mAU



UV Spectrum
Retention time = 27.826



UV Spectrum
Retention time = 30.356



Peak Table

PDA Ch1 254nm

Peak#	Ret. Time	Area	Area%
1	27.826	2142424	50.621
2	30.356	2089898	49.379
Total		4232322	100.000

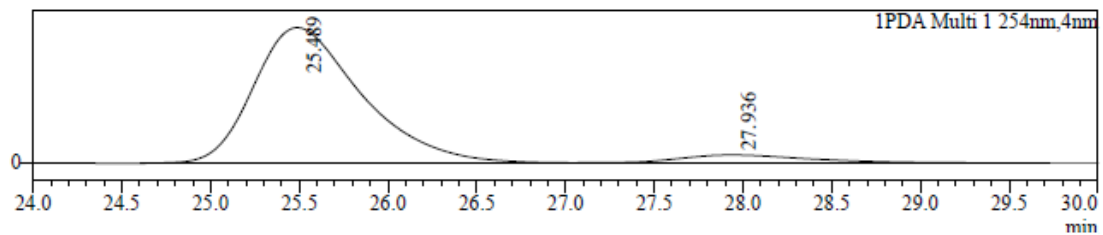
HPLC

Sample Information

Sample Name : P0X788---ODH-5%-0.8Ml
Sample ID : P0X788---ODH-5%-0.8Ml
Data File : P0X788---ODH-5%-0.8Ml.lcd
Method File : P0X-5.0%-0.8ml.lcm

Chromatogram

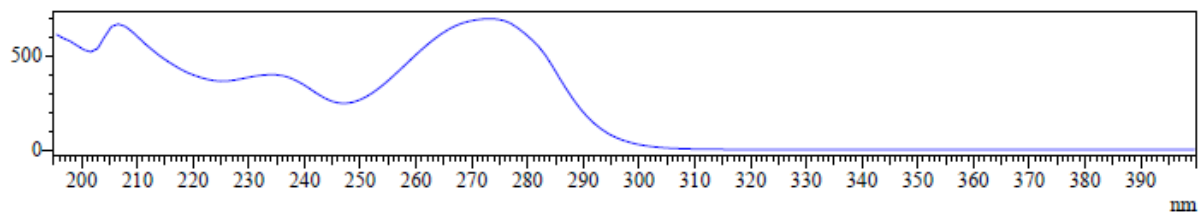
mAU



UV Spectrum

Retention time = 25.489

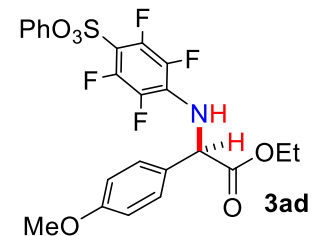
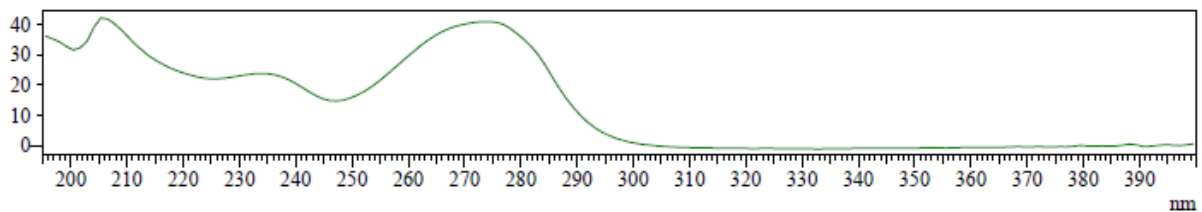
mAU



UV Spectrum

Retention time = 27.936

mAU



Peak Table

PDA Ch1 254nm

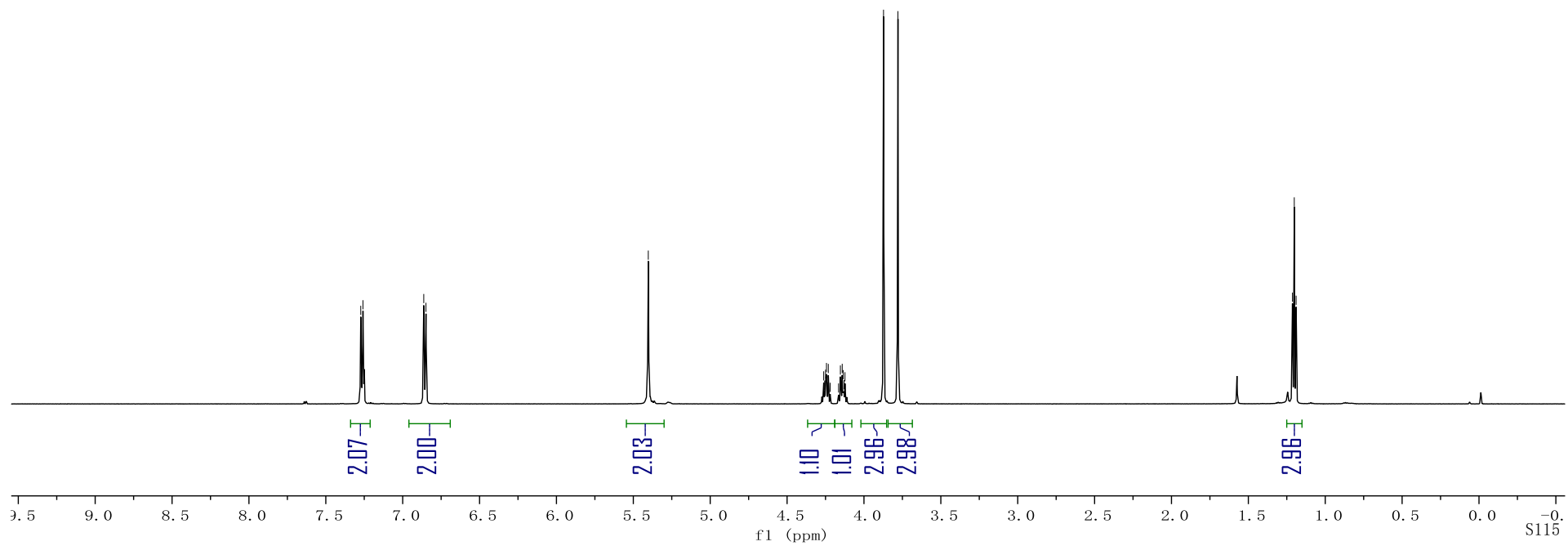
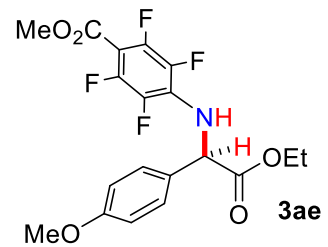
Peak#	Ret. Time	Area	Area%
1	25.489	14766510	93.292
2	27.936	1061720	6.708
Total		15828230	100.000

¹H NMR

7.273
7.269
7.258
6.863
6.849

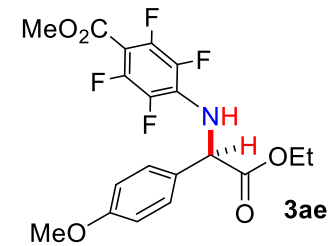
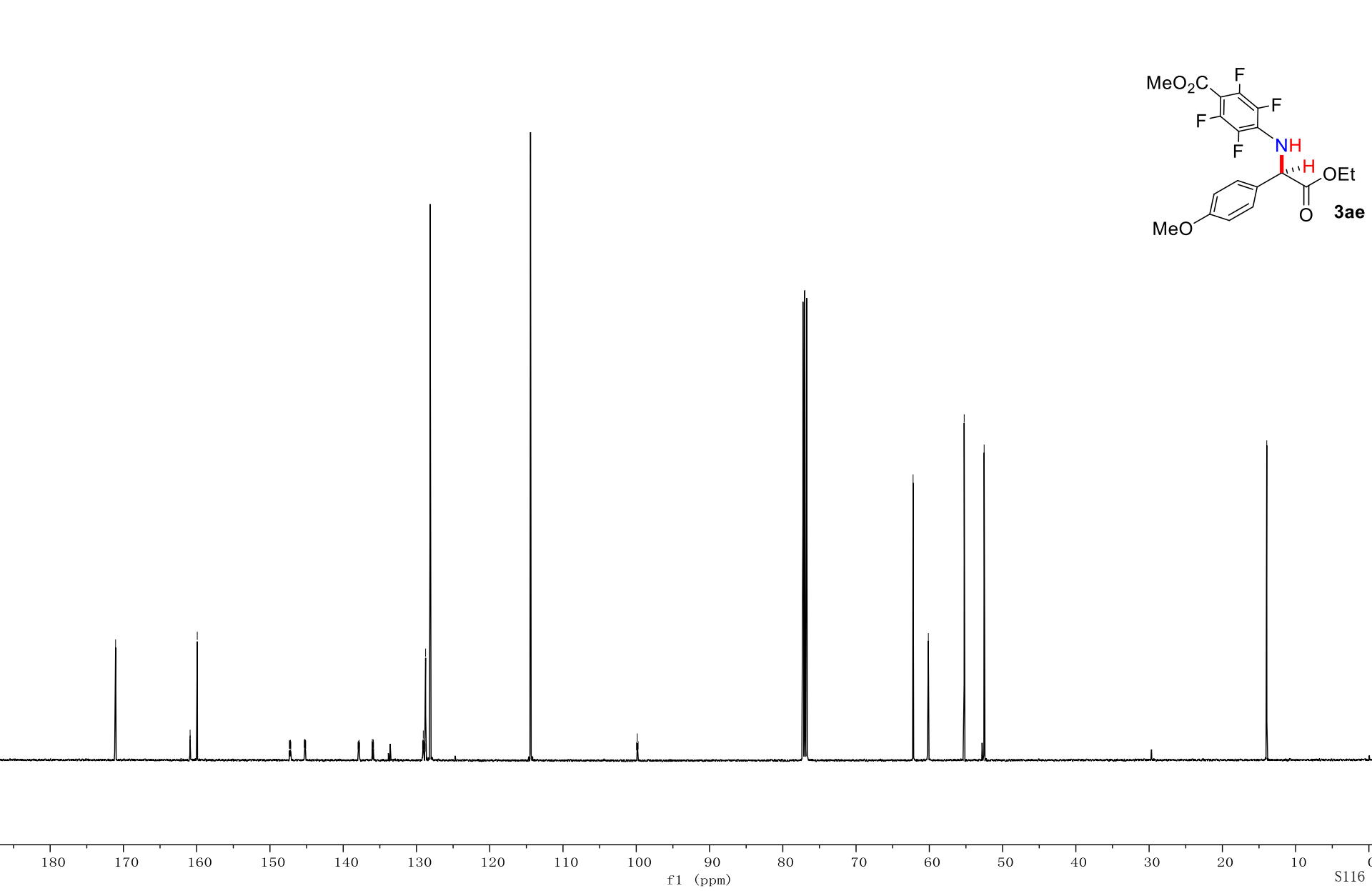
5.403
4.262
4.256
4.250
4.244
4.238
4.232
4.220
4.165
4.153
4.147
4.141
4.135
4.130
4.123
3.873
3.779

1.213
1.201
1.189



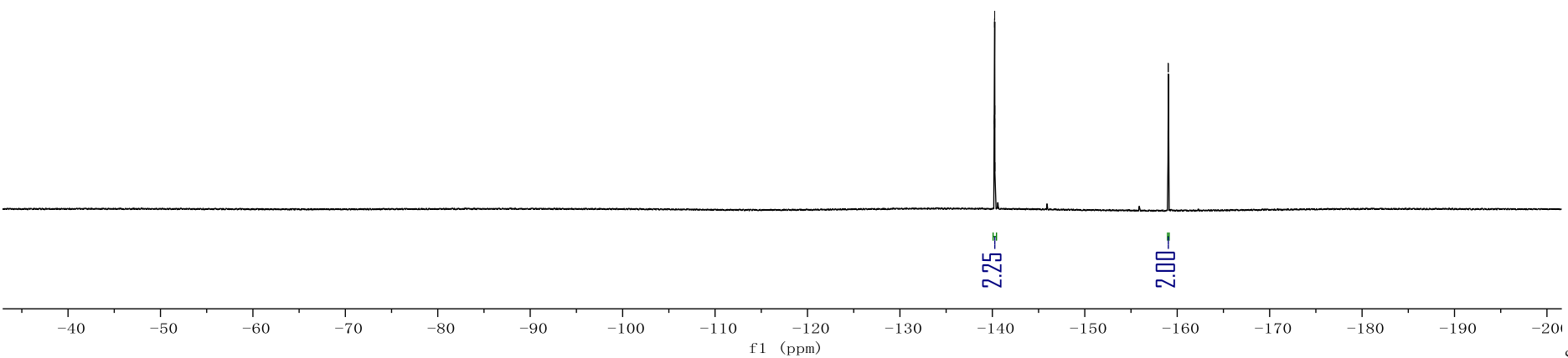
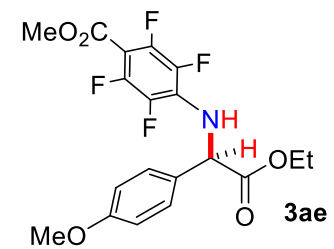
^{13}C NMR

171.066
160.921
160.897
160.873
159.921
147.339
147.312
147.285
147.252
147.233
147.205
147.151
145.310
145.276
145.254
145.226
145.206
145.174
145.146
145.120
137.977
137.942
137.898
137.845
137.809
137.768
136.066
136.024
135.990
135.938
135.902
135.887
135.857
129.163
129.132
129.102
129.071
129.041
128.979
128.817
128.753
128.142
114.435
99.971
99.857
99.744
62.216
60.133
55.240
52.517
13.941



¹⁹F NMR

-140.207
-140.217
-140.222
-140.231
-140.254
-140.263
-140.268
-140.279
-159.004
-159.012
-159.019
-159.042
-159.050
-159.057



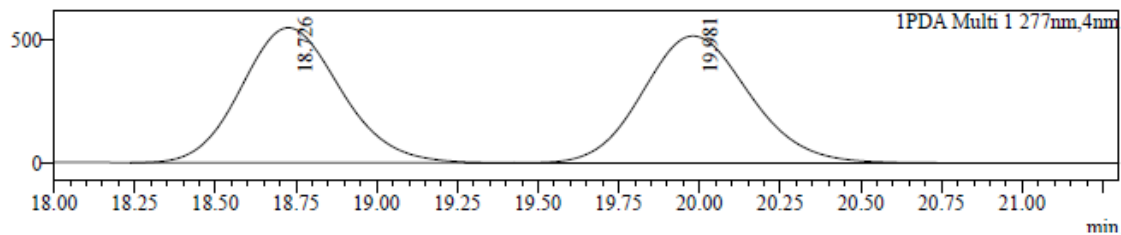
HPLC

Sample Information

Sample Name : P0X-0711-IC-10%-0.8mL
Sample ID : P0X-0711-IC-10%-0.8mL
Data File : P0X-0711-IC-10%-0.8mL.lcd
Method File : YW-10%-0.8ml.lcm

Chromatogram

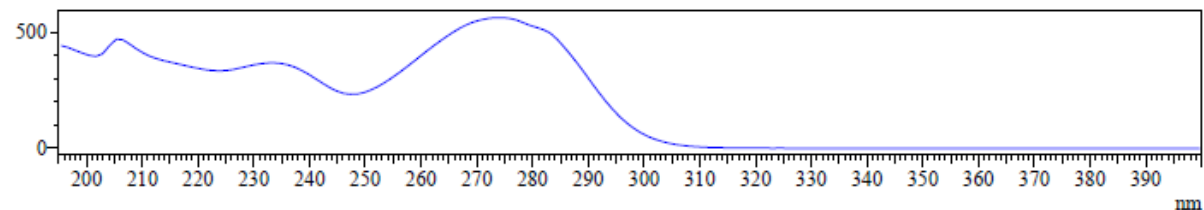
mAU



UV Spectrum

Retention time = 18.726

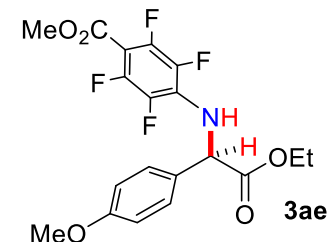
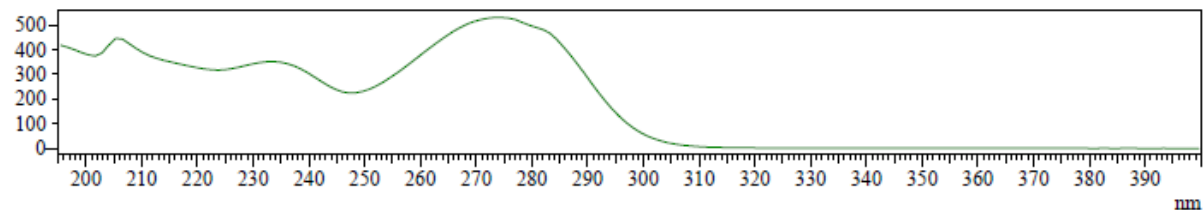
mAU



UV Spectrum

Retention time = 19.981

mAU



Peak Table

PDA Ch1 277nm

Peak#	Ret. Time	Area	Area%
1	18.726	11750787	49.665
2	19.981	11909174	50.335
Total		23659961	100.000

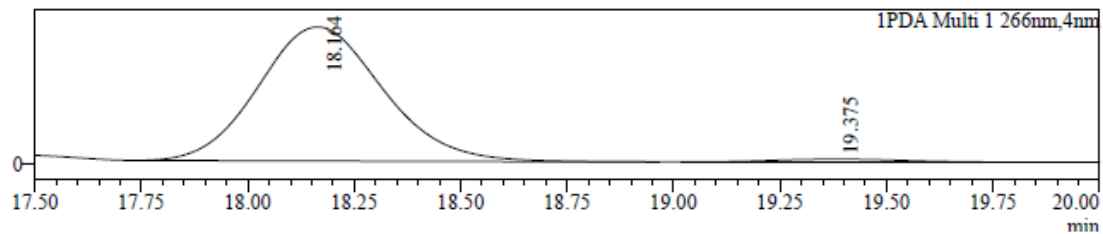
HPLC

Sample Information

Sample Name : P0X-0712IC-10%-0.8mL
Sample ID : P0X-0712IC-10%-0.8mL
Data File : P0X-0712IC-10%-0.8mL.lcd
Method File : P0X-10%-0.8ml.lcm

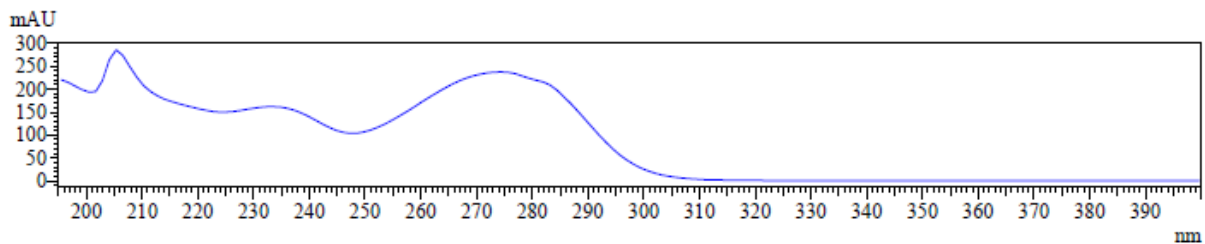
Chromatogram

mAU



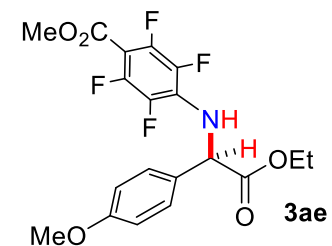
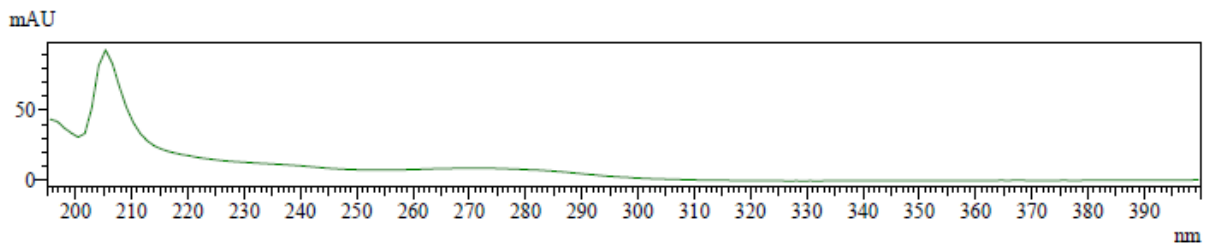
UV Spectrum

Retention time = 18.164



UV Spectrum

Retention time = 19.375

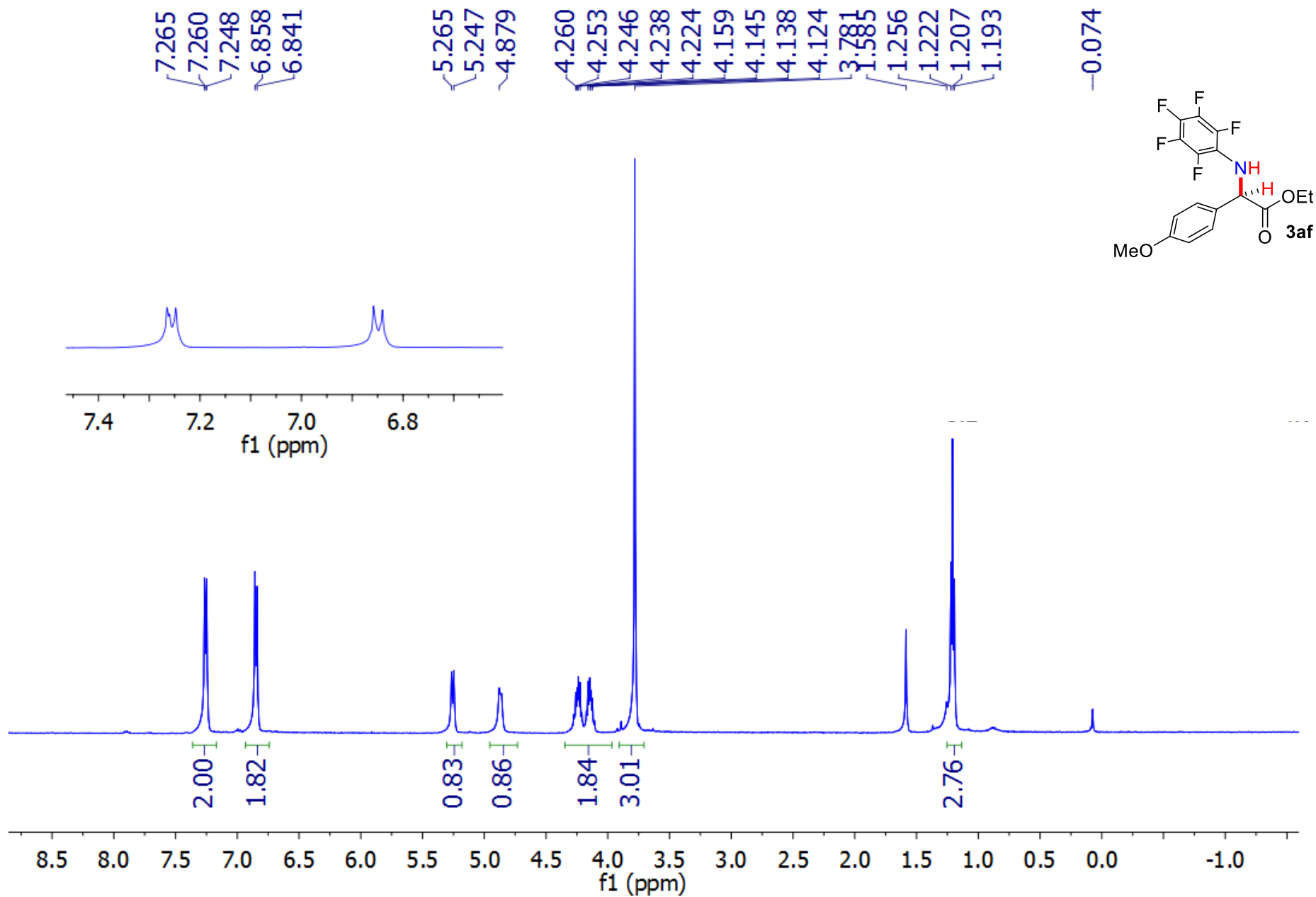


Peak Table

PDA Ch1 266nm

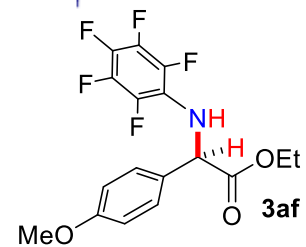
Peak#	Ret. Time	Area	Area%
1	18.164	4159076	97.883
2	19.375	89963	2.117
Total		4249040	100.000

^1H NMR

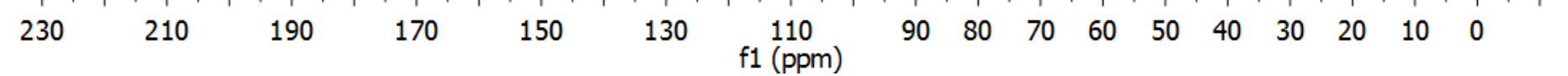
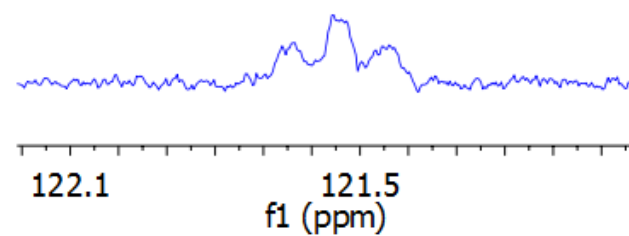
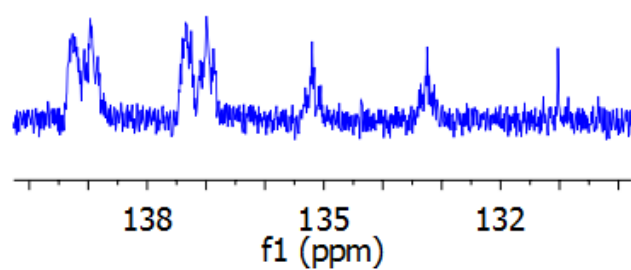


^{13}C NMR

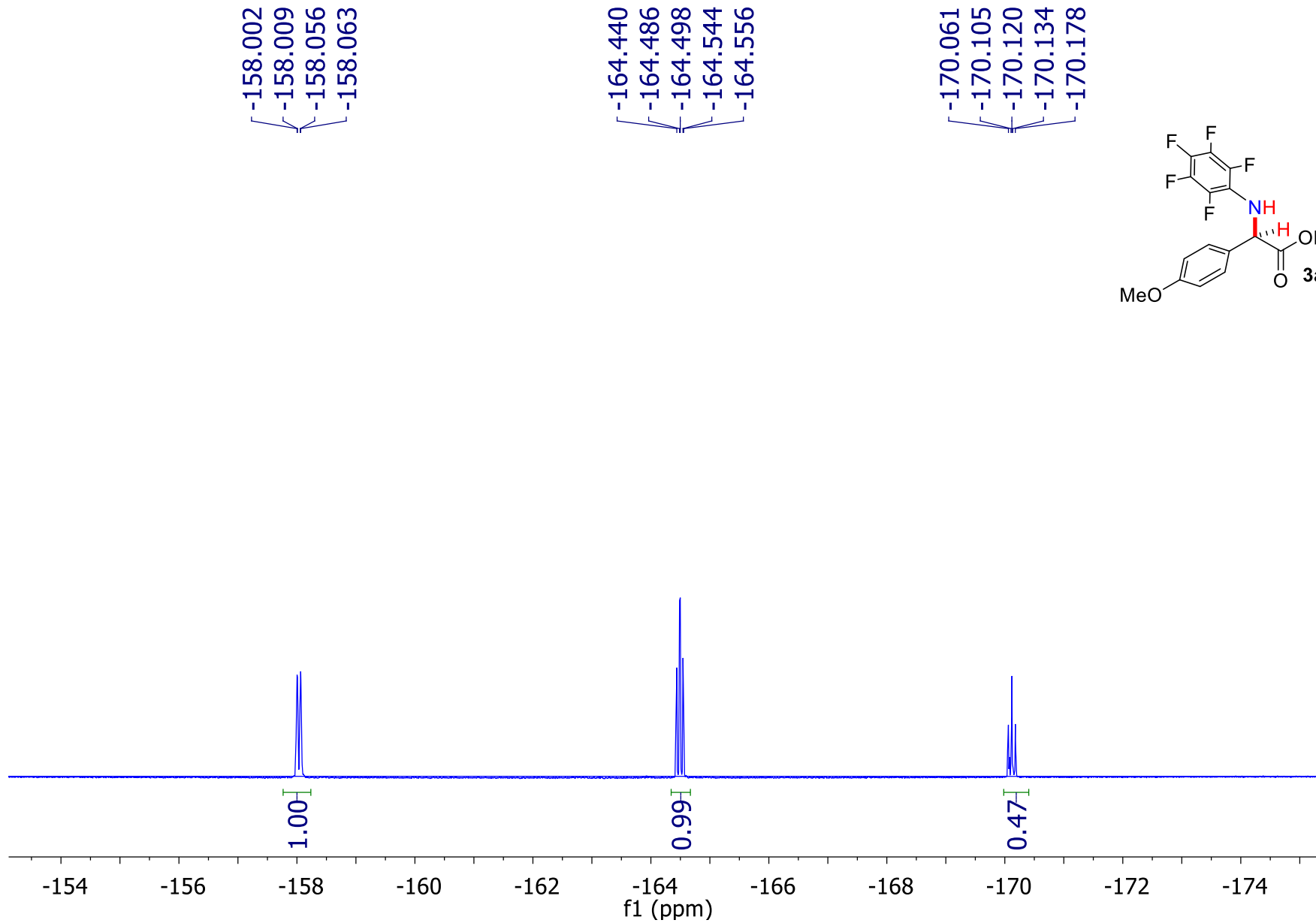
—171.495
—159.814
138.968
137.346
136.991
135.490
135.198
133.245
131.026
128.859
128.188
121.556
114.347
77.254
77.000
76.745
62.013
60.905
60.876
60.846
55.218



—13.960



^{19}F NMR

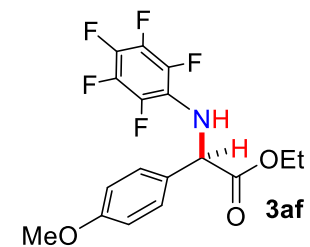
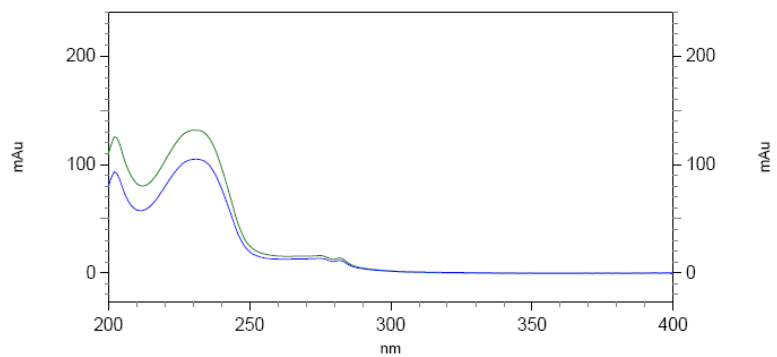
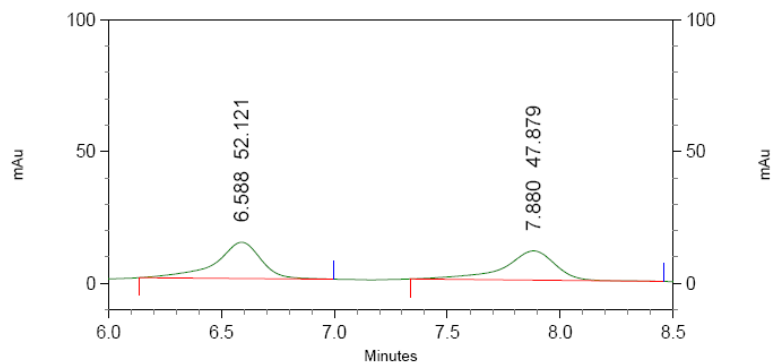


HPLC

JLM-V-213-1-whe1k-1@1ml

C:\EZStart\Projects\Default\Method\SMJ.met

C:\EZStart\Projects\Default\Data\JLM-V-213-1-whe1k-1@1ml



5: 247 nm, 4 nm

Results

Name	Retention Time	Area Percent	Pk #
	6.588	52.121	1
	7.880	47.879	2

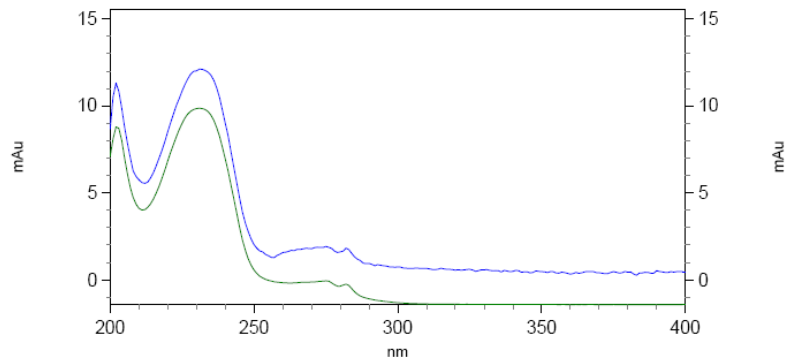
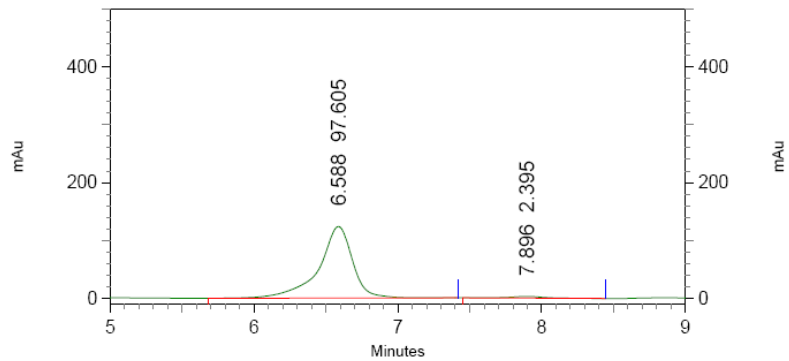
Totals	100.000	
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HPLC

JLM-V-213-2-whelk-1@1ml

C:\EZStart\Projects\Default\Method\SMJ.met

C:\EZStart\Projects\Default\Data\JLM-V-213-2-whelk-1@1ml

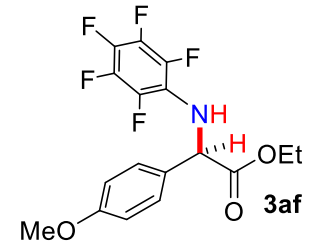


1: 236 nm, 4 nm

Results

Name	Retention Time	Area Percent	Pk #
	6.588	97.605	1
	7.896	2.395	2

Totals	100.000	
--------	---------	--

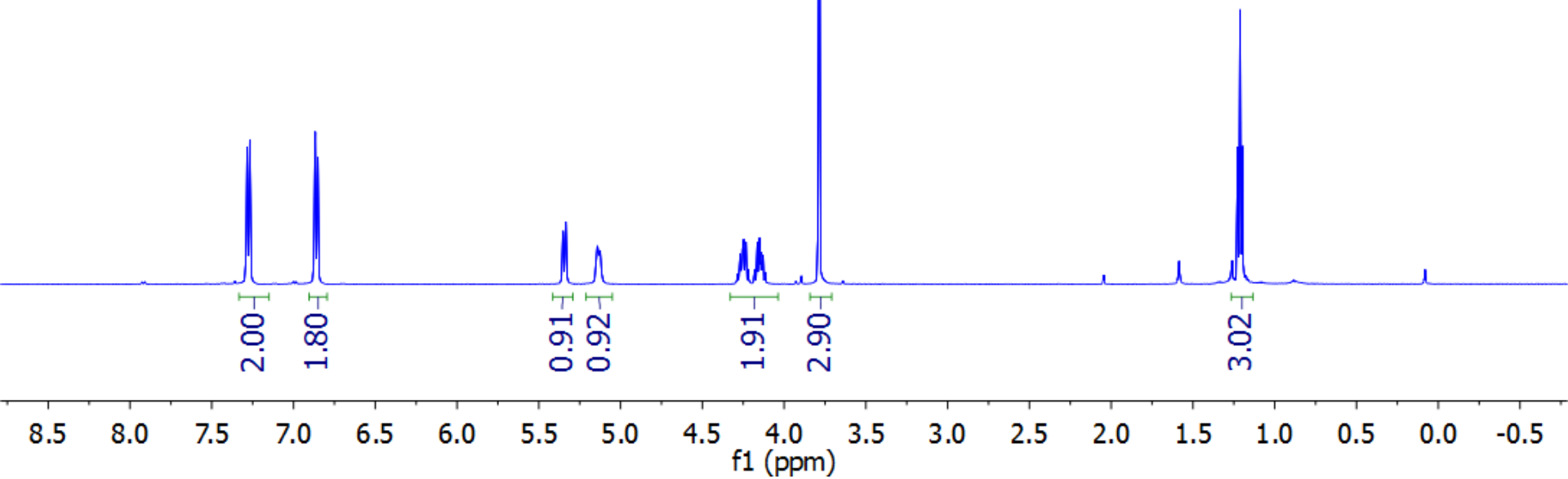
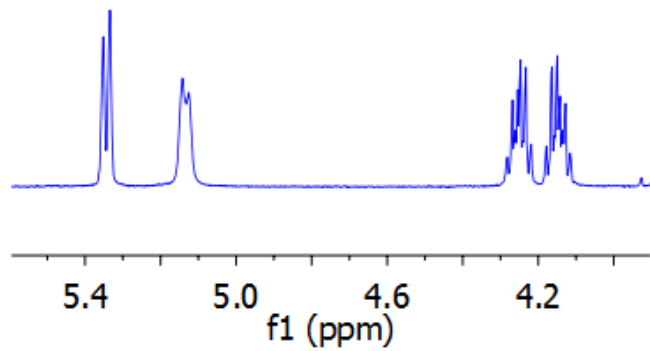
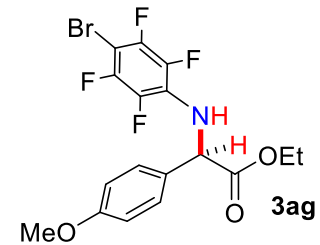


¹H NMR

7.284
7.267
7.260
6.868
6.852

5.351
5.334
5.142
5.125
4.268
4.254
4.247
4.233
4.164
4.150
4.142
4.130
4.128
3.786
3.785

1.225
1.213
1.211
1.197

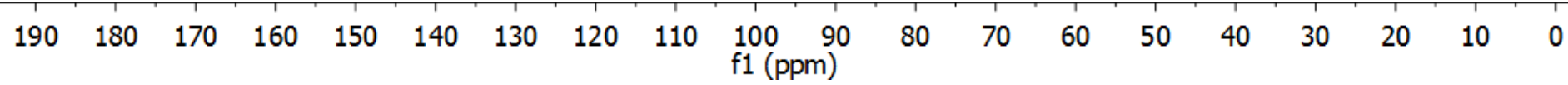
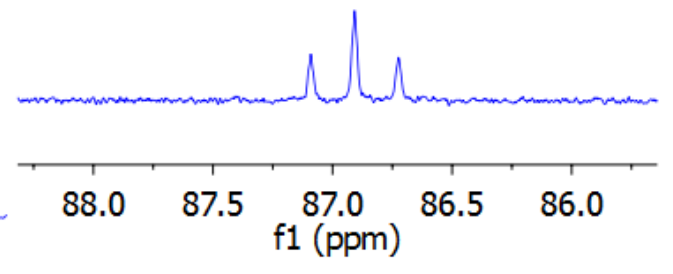
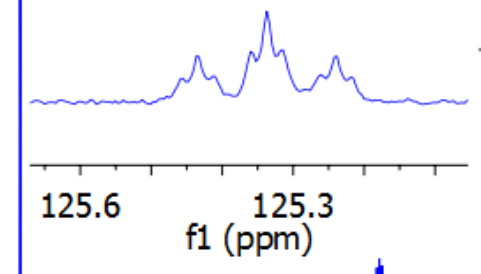
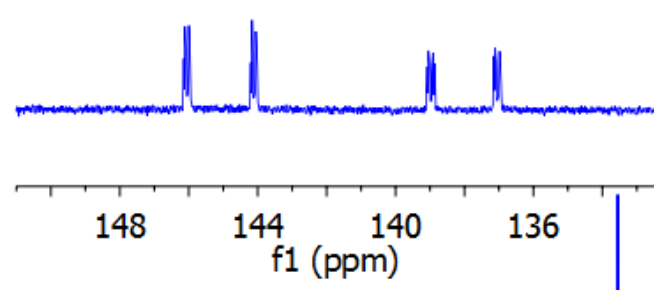
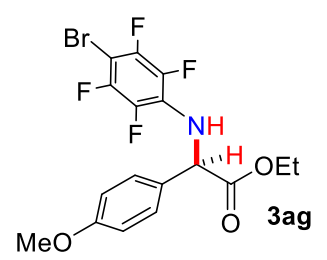


¹³C NMR

171.353
159.840
146.108
145.997
144.164
144.053
137.112
128.911
128.175
125.337
124.372

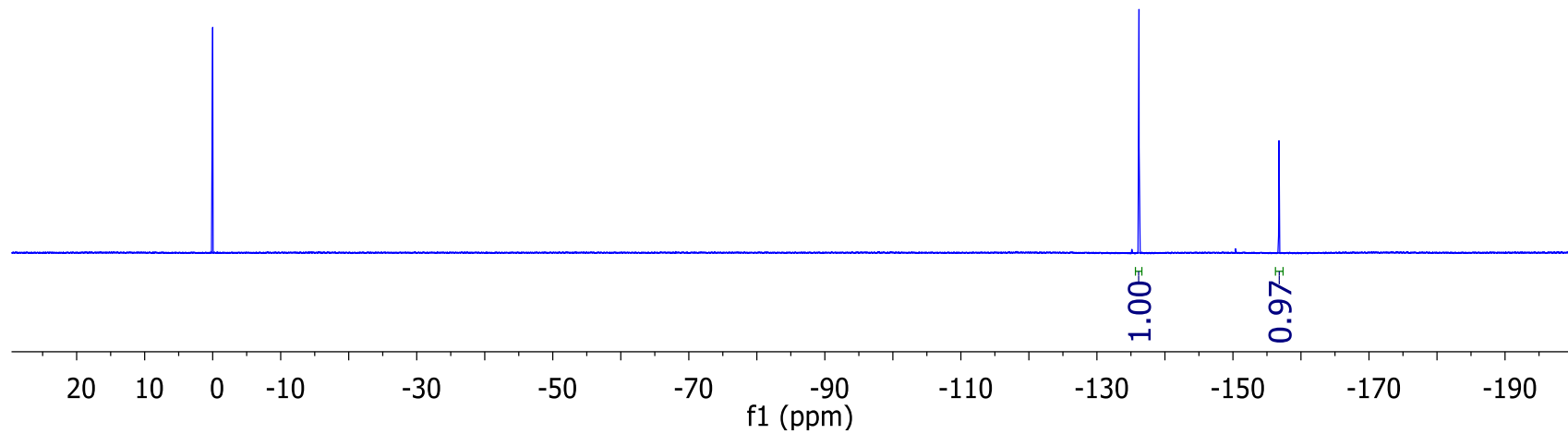
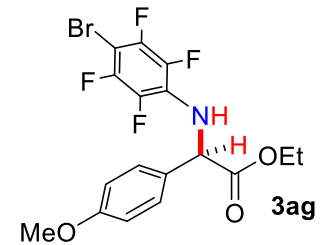
87.091
86.907
86.724
77.255
77.000
76.746
62.072
60.534
60.503
60.471
55.222

13.960



^{19}F NMR

-136.117
-136.129
-136.138
-136.146
-136.188
-136.196
-136.206
-136.218
-156.731
-156.736
-156.780
-156.785

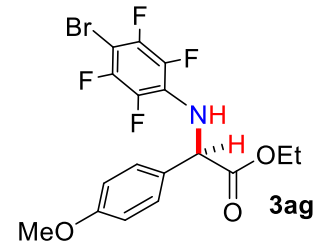
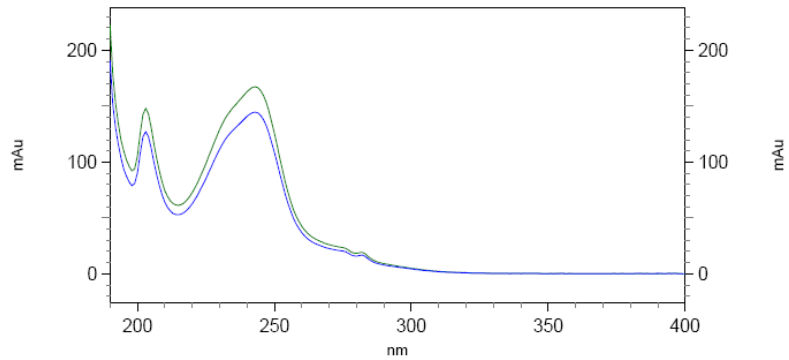
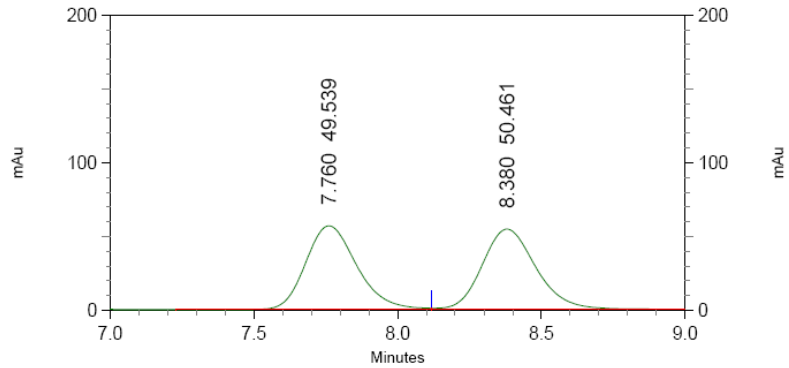


HPLC

JLM-V-206-1-ADH-1%1ML

C:\EZStart\Projects\Default\Method\shifatest_2,5-dimehoxy.met

C:\EZStart\Projects\Default\Data\JLM-V-206-1-ADH1%1ML



7: 251 nm, 4 nm

Results

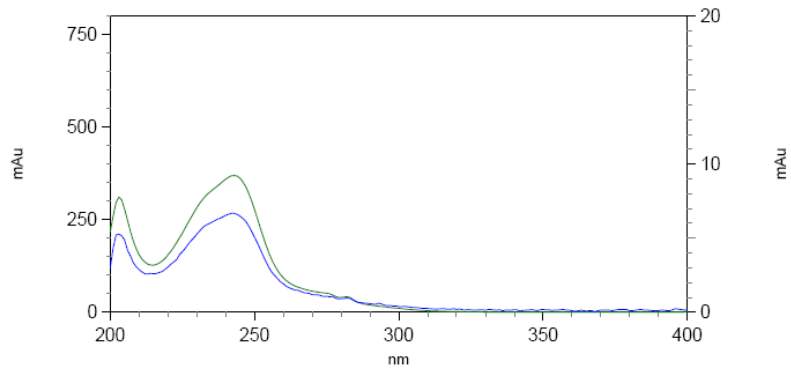
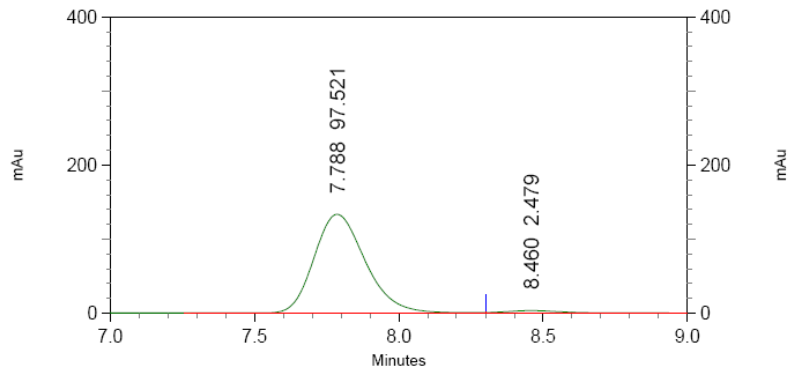
Pk #	Name	Retention Time	Area Percent
1		7.760	49.539
2		8.380	50.461
Totals			100.000

HPLC

JLM-V-206-2-ADH-1%1ML

C:\EZStart\Projects\Default\Method\shifatest_2,5-dimehoxy.met

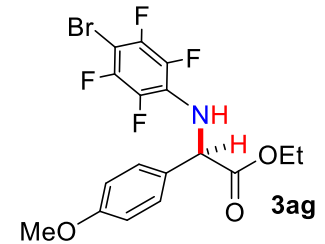
C:\EZStart\Projects\Default\Data\JLM-V-206-2-ADH1%1ML



7: 238 nm, 4 nm

Results

Pk #	Name	Retention Time	Area Percent
1		7.788	97.521
2		8.460	2.479
Totals			100.000

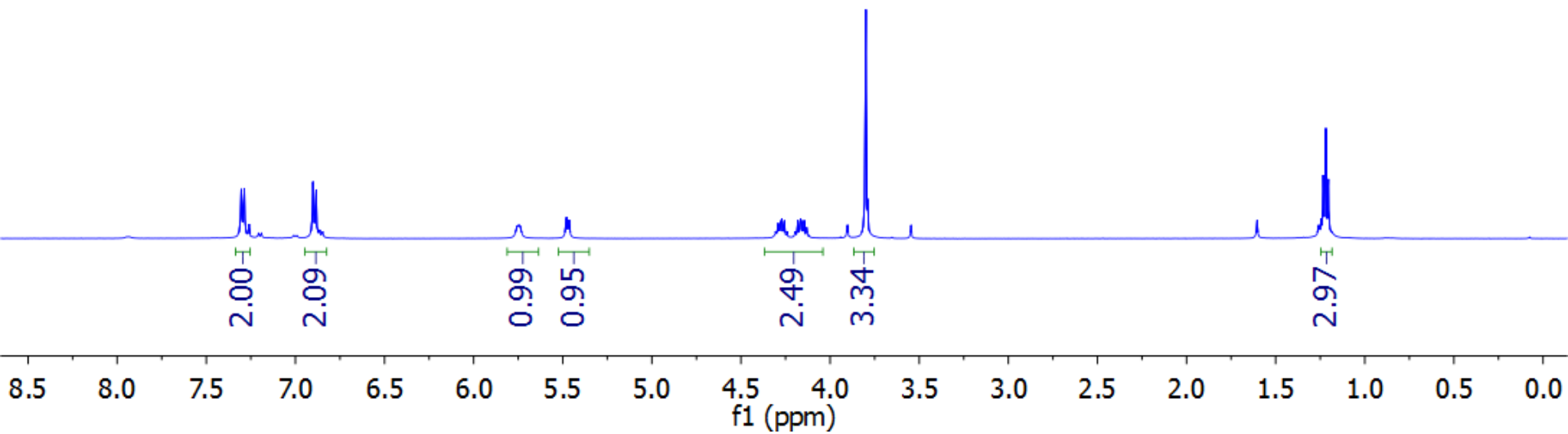
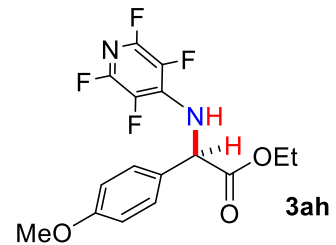
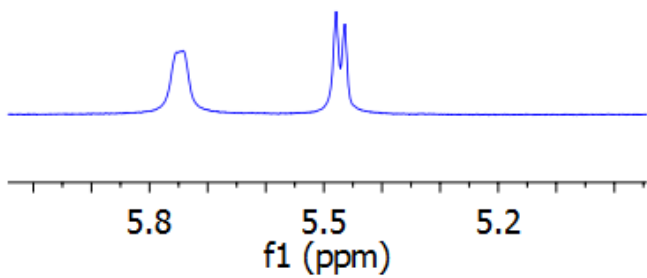


¹H NMR

7.304
7.286
7.260
6.901
6.884

5.742
5.479
5.464
4.292
4.284
4.277
4.270
4.256
4.181
4.166
4.159
4.152
4.145
4.131
3.799

1.233
1.219
1.204



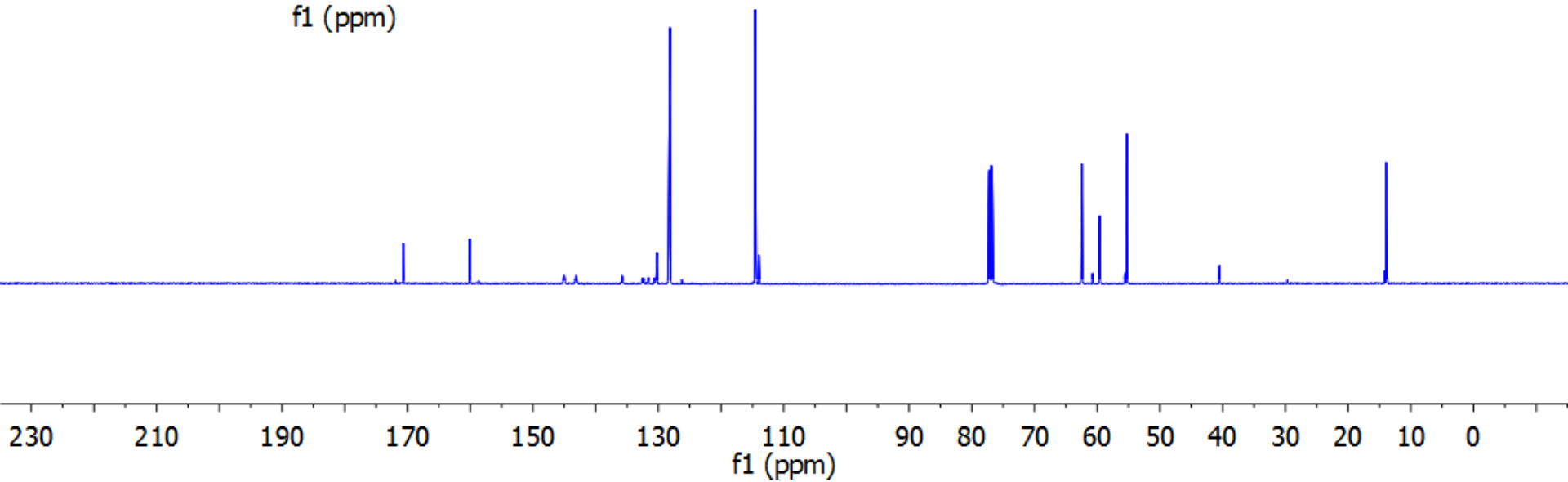
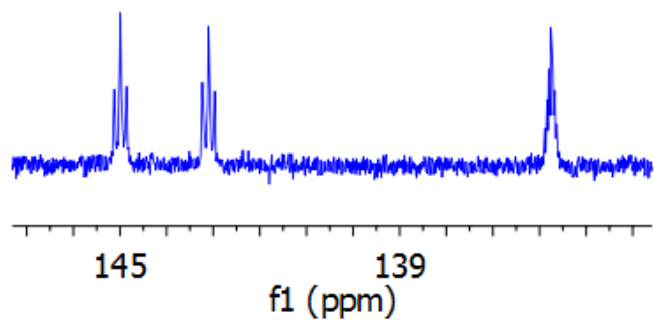
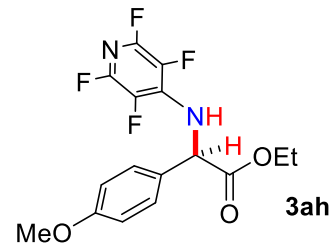
¹³C NMR

-170.688
-160.068
144.994
143.101
135.757
130.215
128.384
128.139
114.539
114.386
113.939

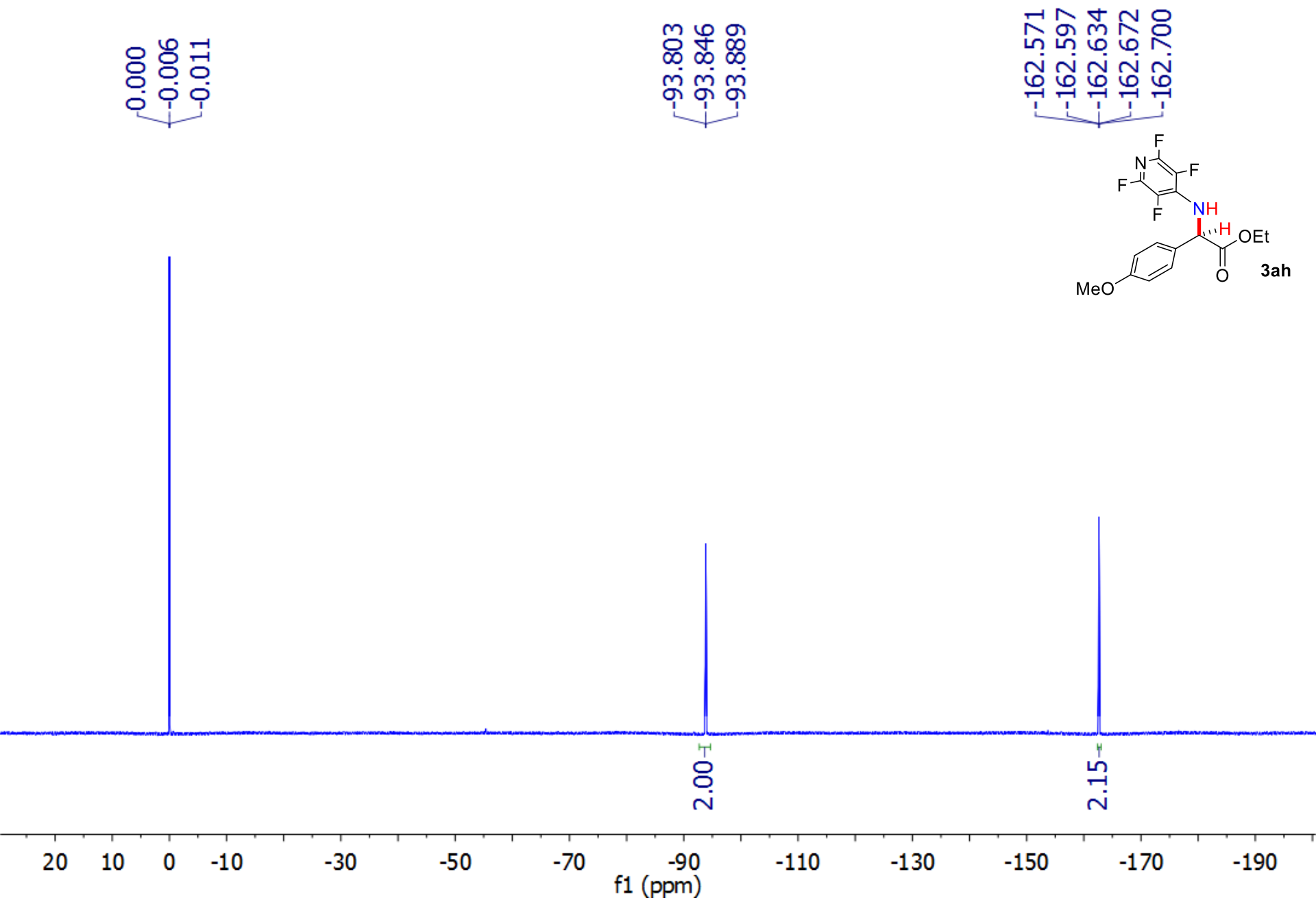
77.254
77.000
76.746

62.448
59.607
59.578
55.275
40.494

14.155
13.929
13.812



^{19}F NMR

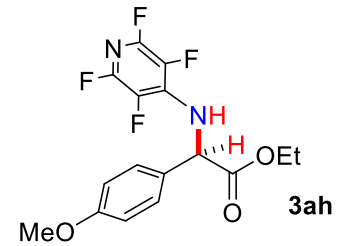
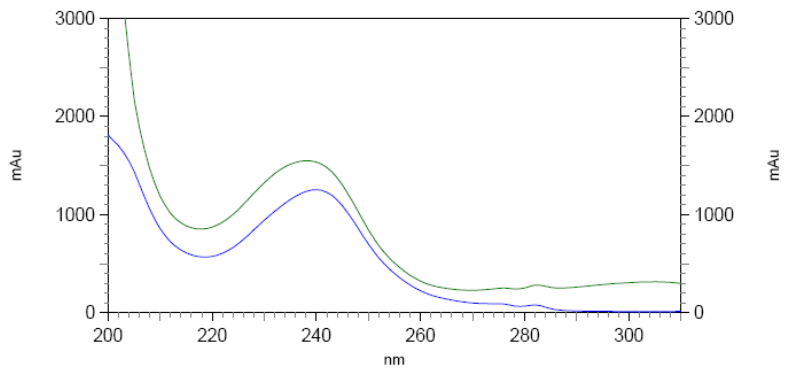
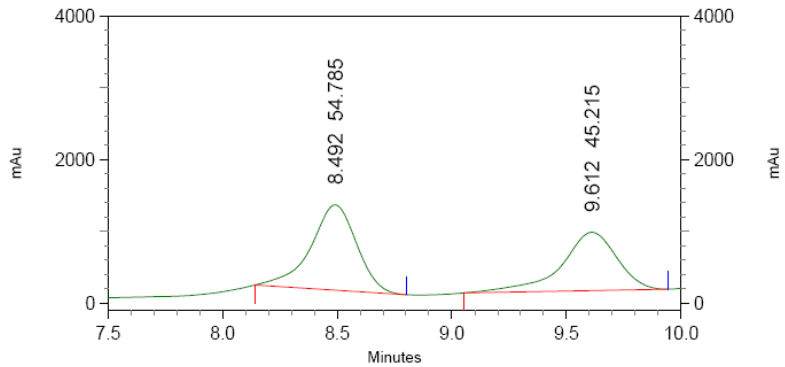


HPLC

JLM-V-174-2-WHELK-3%1ML

C:\EZStart\Projects\Default\Method\XC-5%-ADH1ml.met

E:\JLM-V-174-2-WHELK3%1ML



4: 231 nm, 4 nm

Results

Name	Retention Time	Area Percent	Pk #
	8.492	54.785	1
	9.612	45.215	2

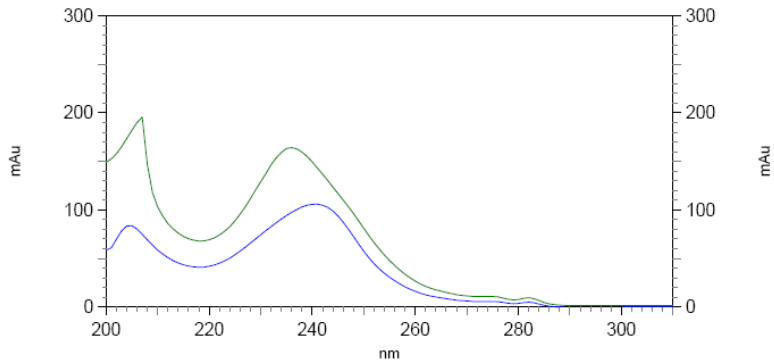
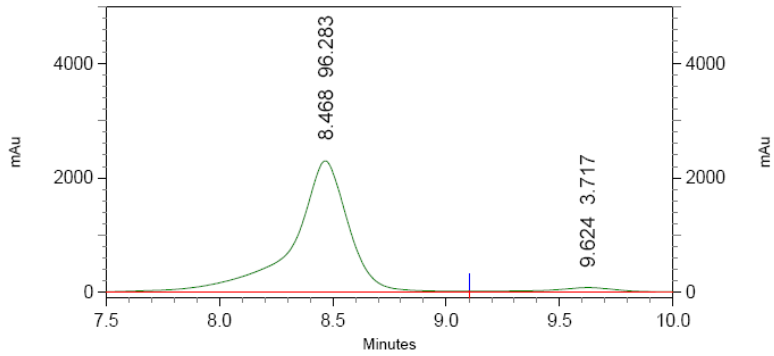
Totals		100.000	
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HPLC

JLM-V-174-1-WHELK-3%1ML

C:\EZStart\Projects\Default\Method\XC-5%-ADH1ml.met

E:\JLM-V-174-1-WHELK3%1ML

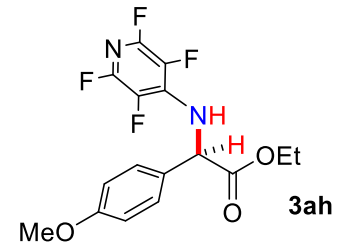


4: 231 nm, 4 nm

Results

Name	Retention Time	Area Percent	Pk #
	8.468	96.283	1
	9.624	3.717	2

Totals	100.000	
--------	---------	--

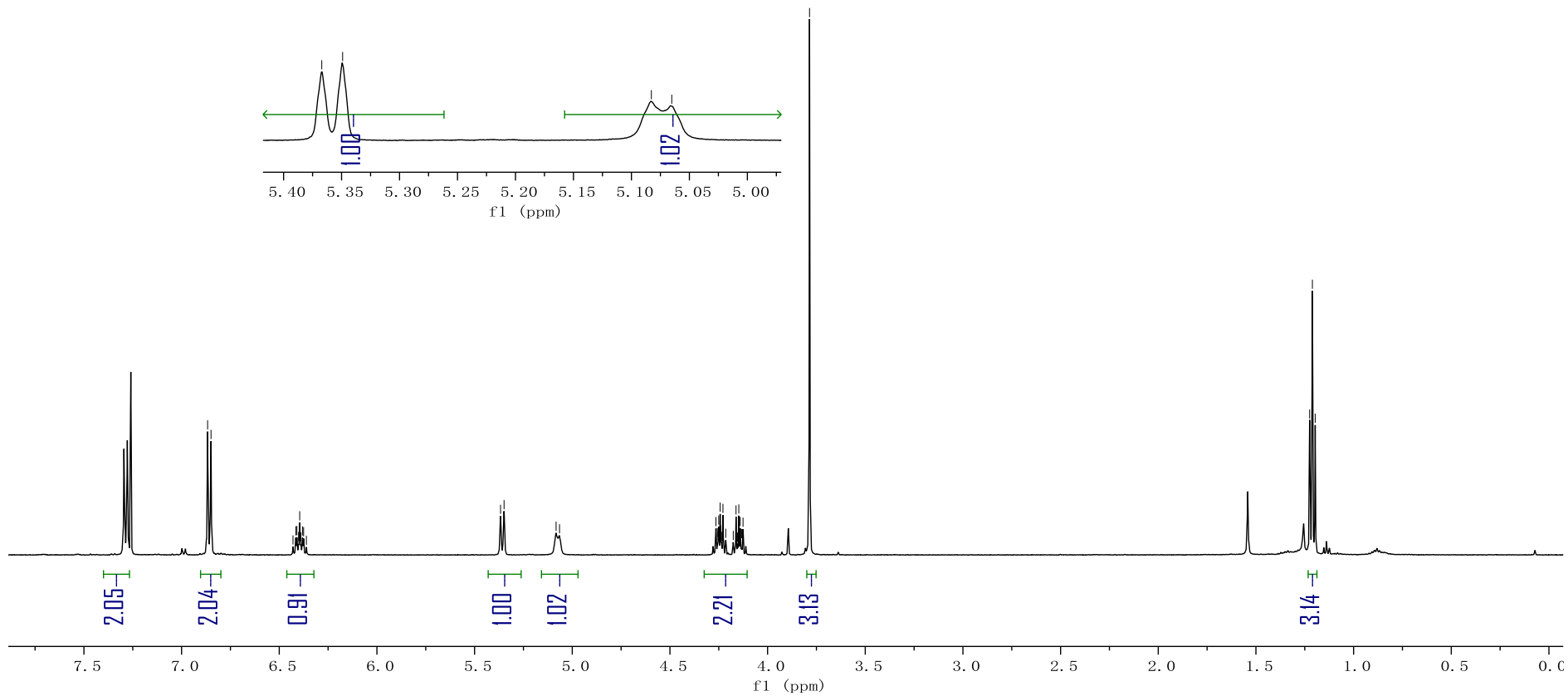
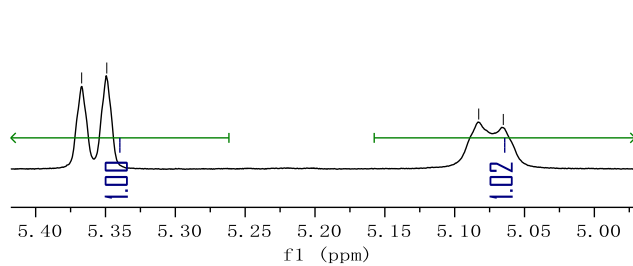
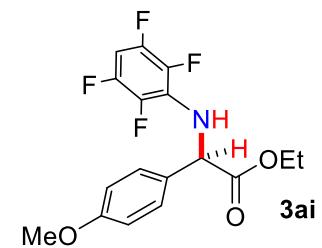


¹H NMR

6.867
6.849
6.430
6.415
6.410
6.401
6.395
6.389
6.381
6.375
6.361
5.367
5.349
5.083
5.065
4.264
4.257
4.254
4.250
4.243
4.240
4.236
4.229
4.214
4.176
4.162
4.154
4.147
4.140
4.133
4.126
3.785

1.225
1.211
1.197

5.367
5.349

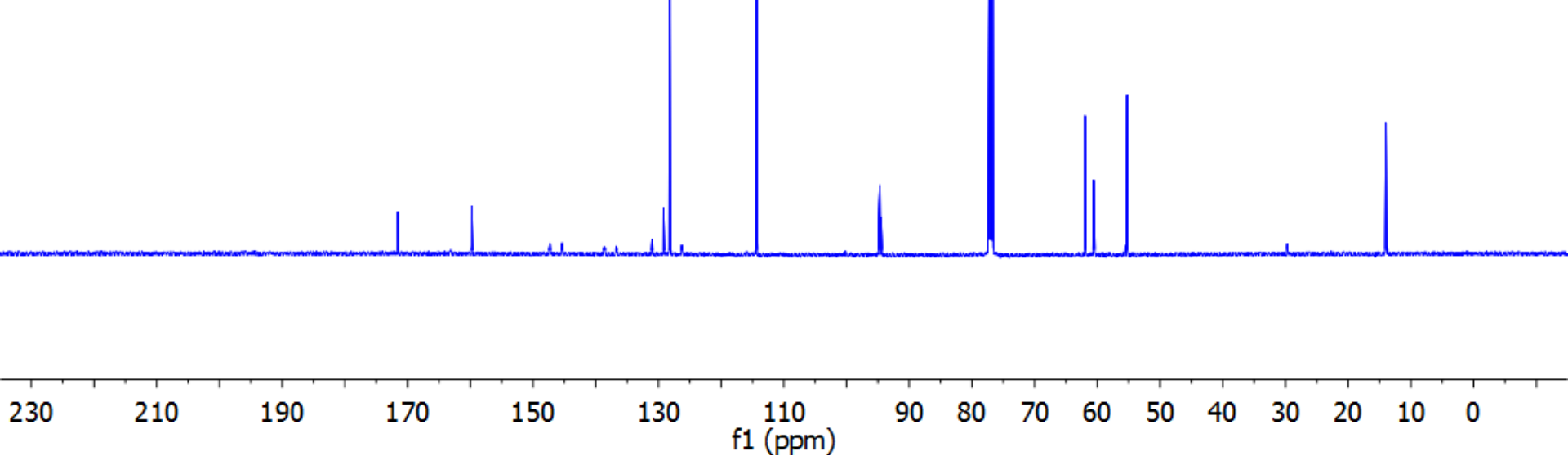
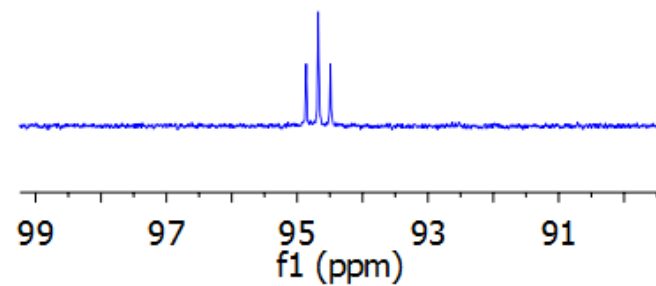
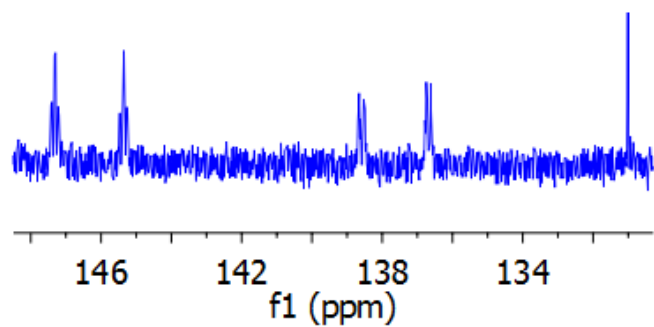
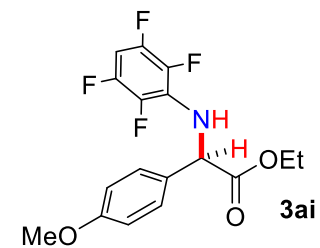


^{13}C NMR

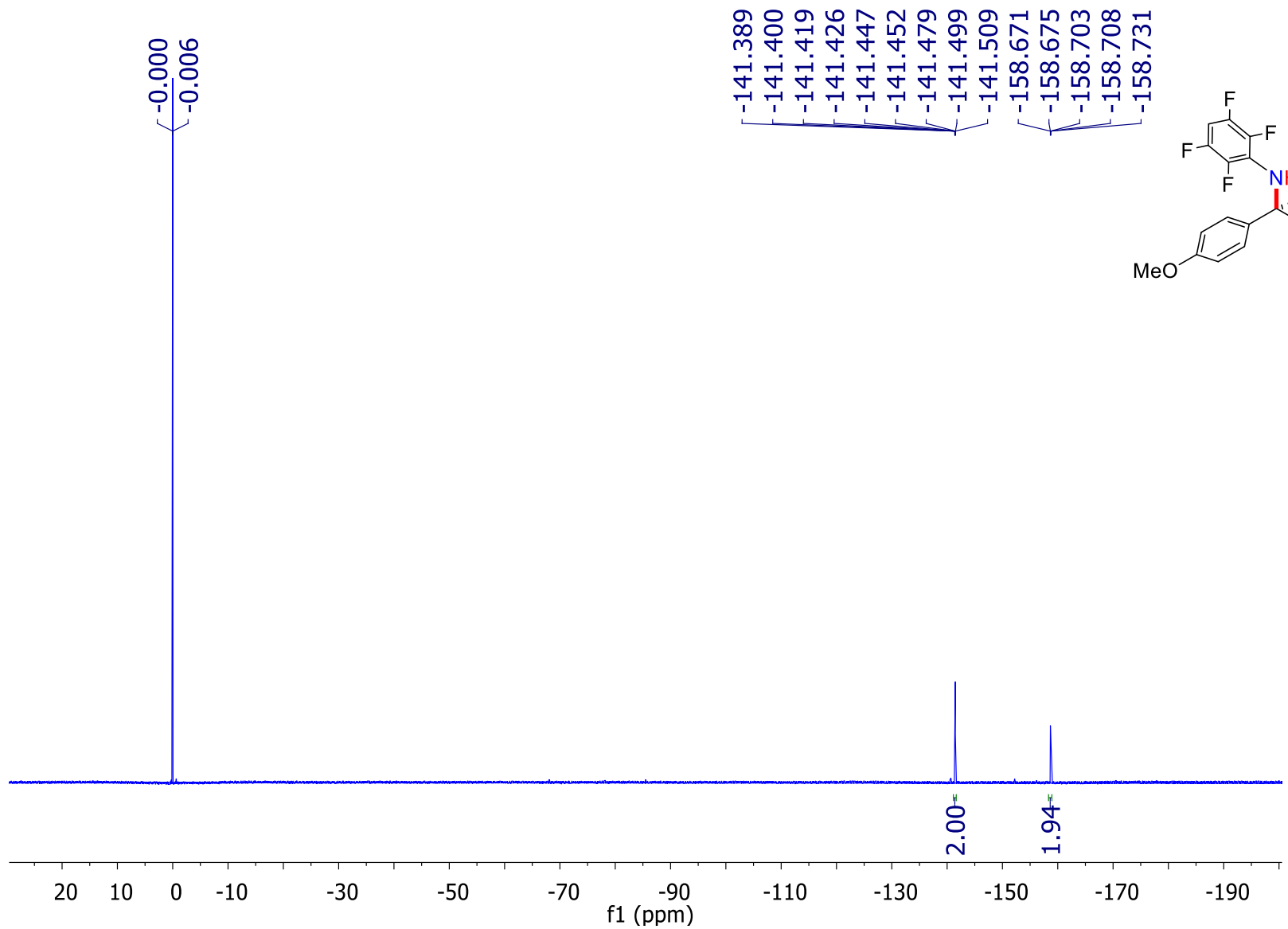
171.539
159.764
147.294
145.350
138.654
136.745
131.009
129.185
128.209
126.281
114.315
94.861
94.676
94.491

77.000
76.746
61.968
60.625
60.592
60.559
55.237

13.982

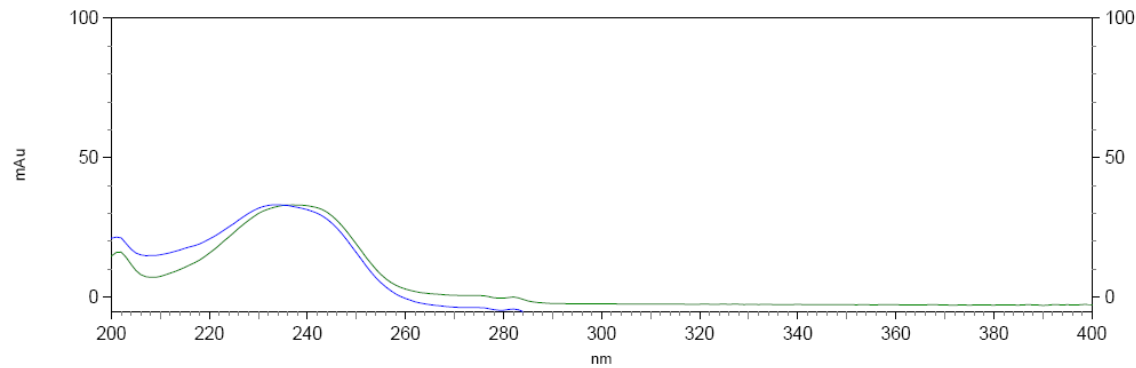
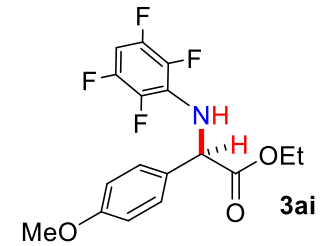
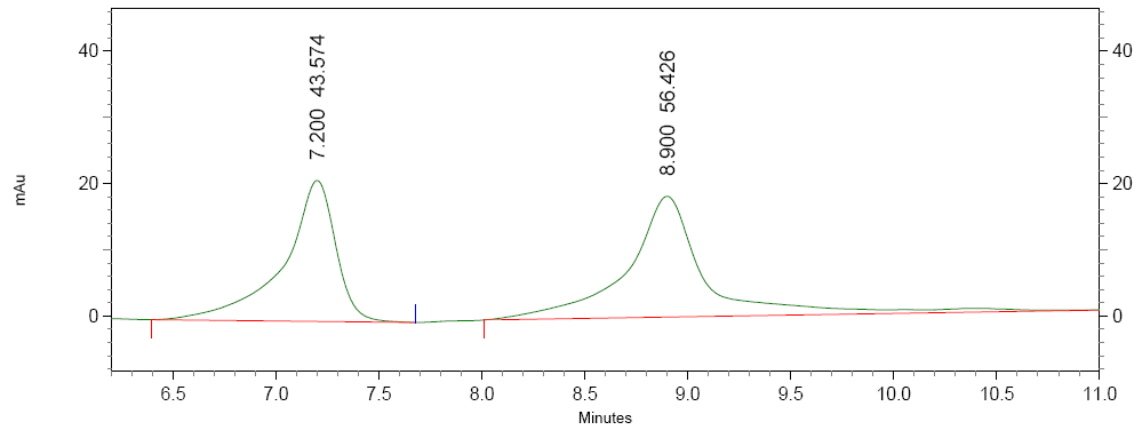


^{19}F NMR



HPLC

JLM-V-227-1-whe1k-1@1ml
C:\EZStart\Projects\Default\Data\JLM-V-227-1-whe1k-1@1ml
C:\EZStart\Projects\Default\Method\CQL-AD-H10%1.0ml60min.met
AD-H column 20%IPA @ 0.8ml/min

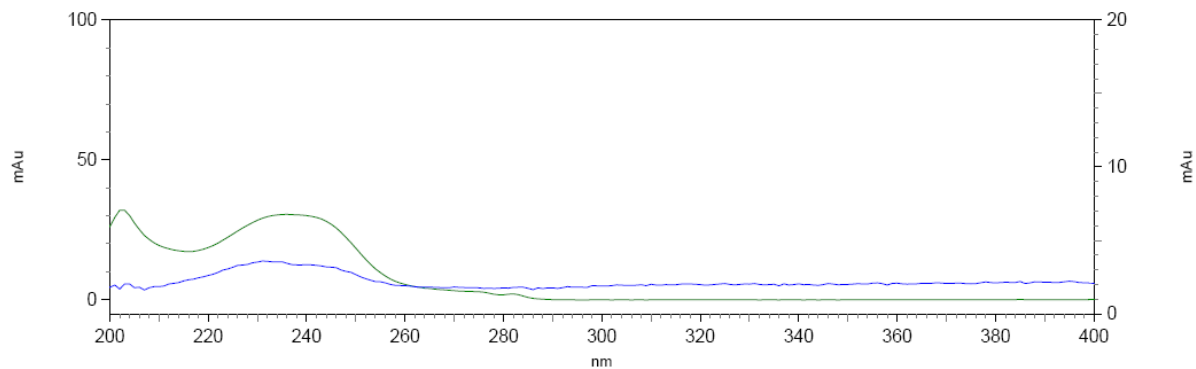
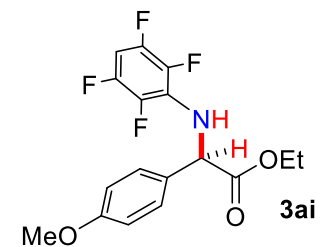
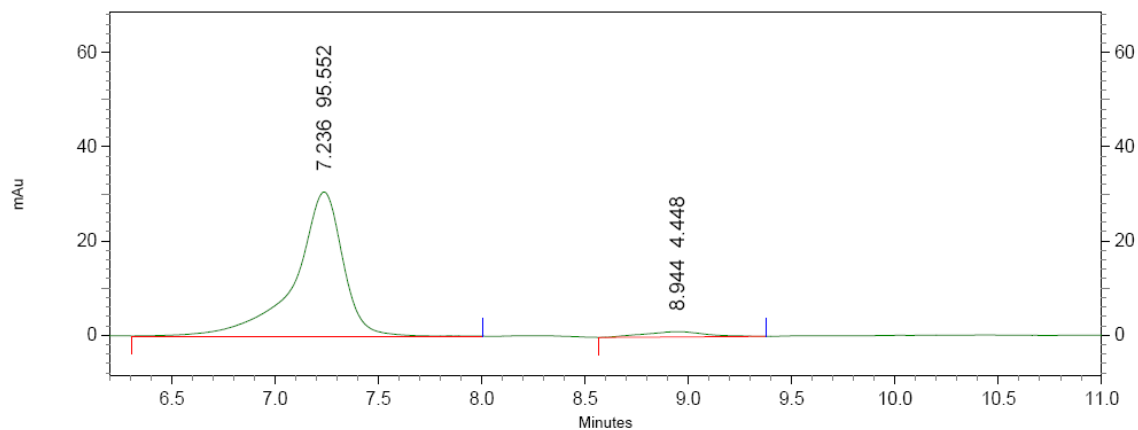


2: 235 nm, 4 nm Results

Pk #	Retention Time	Area Percent
1	7.200	43.574
2	8.900	56.426
Totals		100.000

HPLC

JLM-V-227-2-whelk-1@1ml
C:\EZStart\Projects\Default\Data\JLM-V-227-2-whelk-1@1ml
C:\EZStart\Projects\Default\Method\CQL-AD-H10%1.0ml60min.met
AD-H column 20%IPA @ 0.8ml/min



2: 235 nm, 4 nm Results

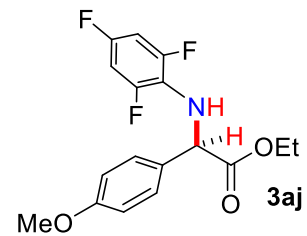
Pk #	Retention Time	Area Percent
1	7.236	95.552
2	8.944	4.448
Totals		100.000

¹H NMR

7.284
7.268
6.846
6.829
6.578
6.561
6.547

5.237
5.220

4.653
4.634
4.253
4.251
4.248
4.245
4.238
4.234
4.231
4.227
4.216
4.212
4.159
4.155
4.145
4.141
4.137
4.133
4.130
4.123
4.119
3.779
1.222
1.208
1.194



2.28

1.96

1.95

1.00

0.93

2.09

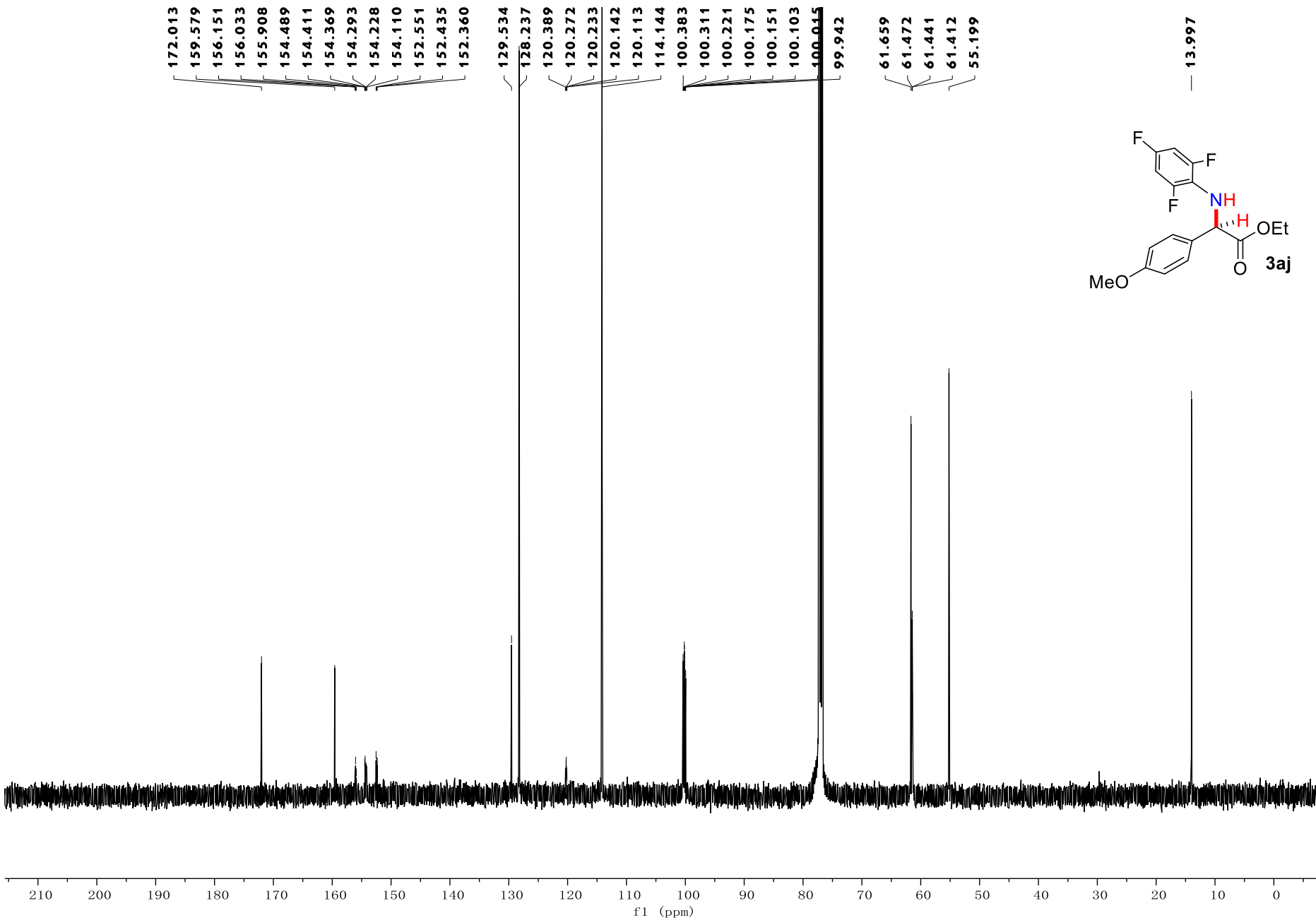
3.18

3.05

8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0

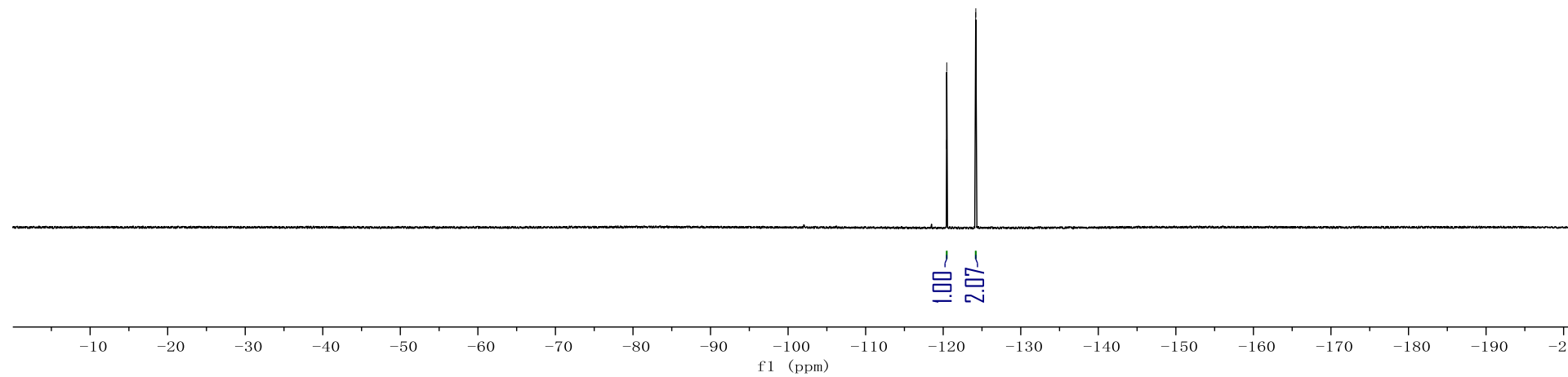
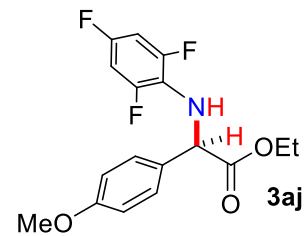
f1 (ppm)

¹³C NMR



^{19}F NMR

-120.440
-120.459
-120.477
-124.188
-124.206

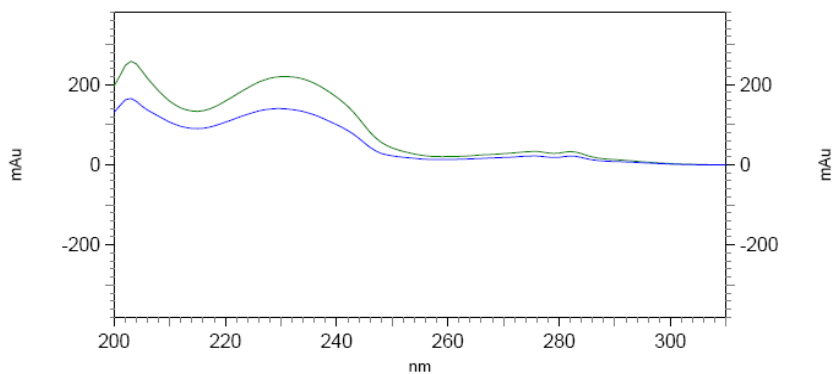
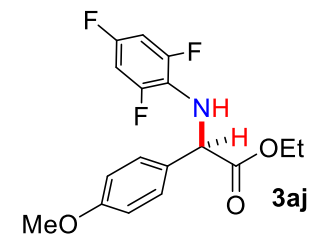
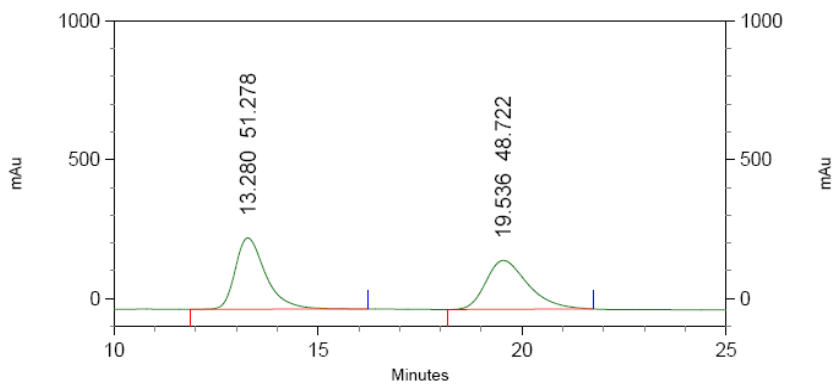


HPLC

JLM-V-205-1-OJH-1%1ML

C:\EZStart\Projects\Default\Method\XC-5%-ADH1ml.met

E:\JLM-V-205-1-OJH-1%1ML



4: 232 nm, 4 nm

Results

Name	Retention Time	Area Percent	Pk #
	13.280	51.278	1
	19.536	48.722	2

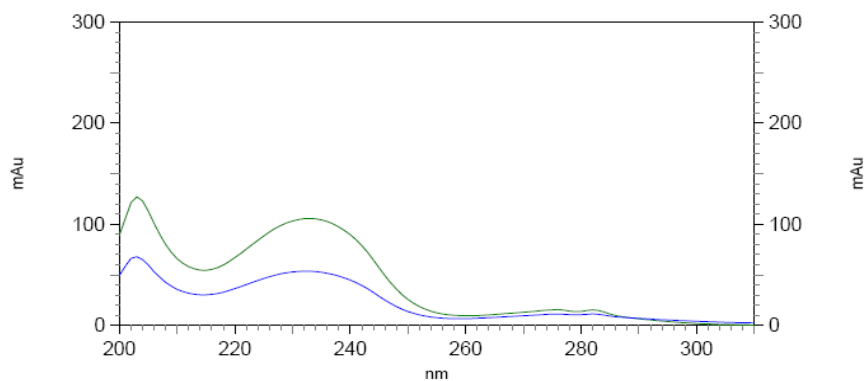
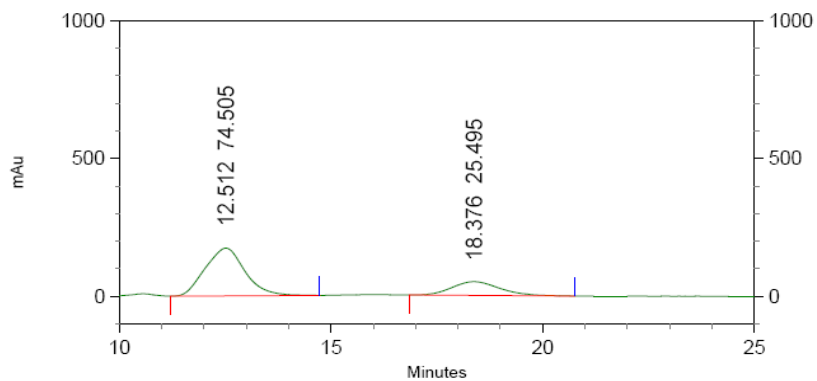
Totals	100.000	
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HPLC

JLM-V-205-2-OJH-1%1ML

C:\EZStart\Projects\Default\Method\XC-5%-ADH1ml.met

E:\JLM-V-205-2-OJH-1%1ML

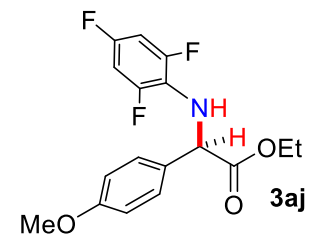


4: 232 nm, 4 nm

Results

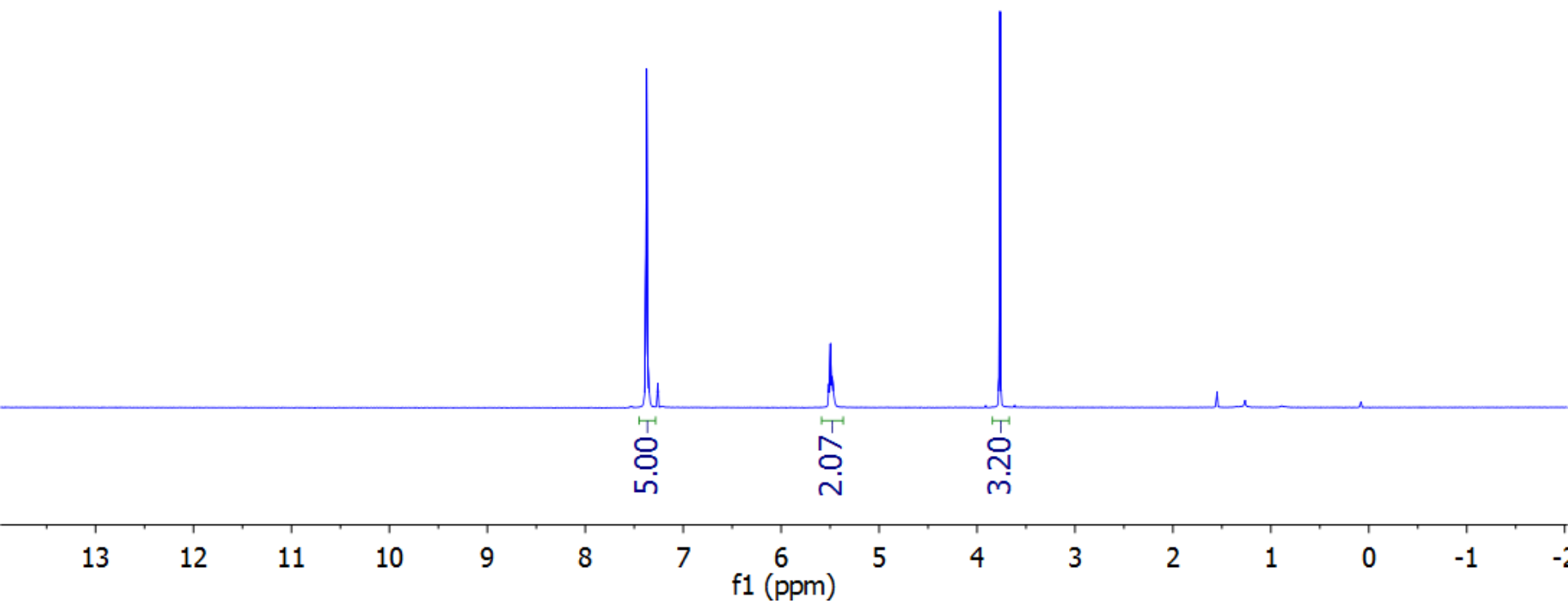
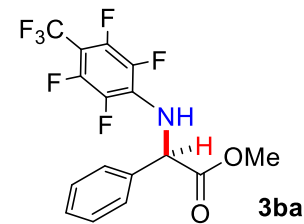
Name	Retention Time	Area Percent	Pk #
	12.512	74.505	1
	18.376	25.495	2

Totals		100.000	
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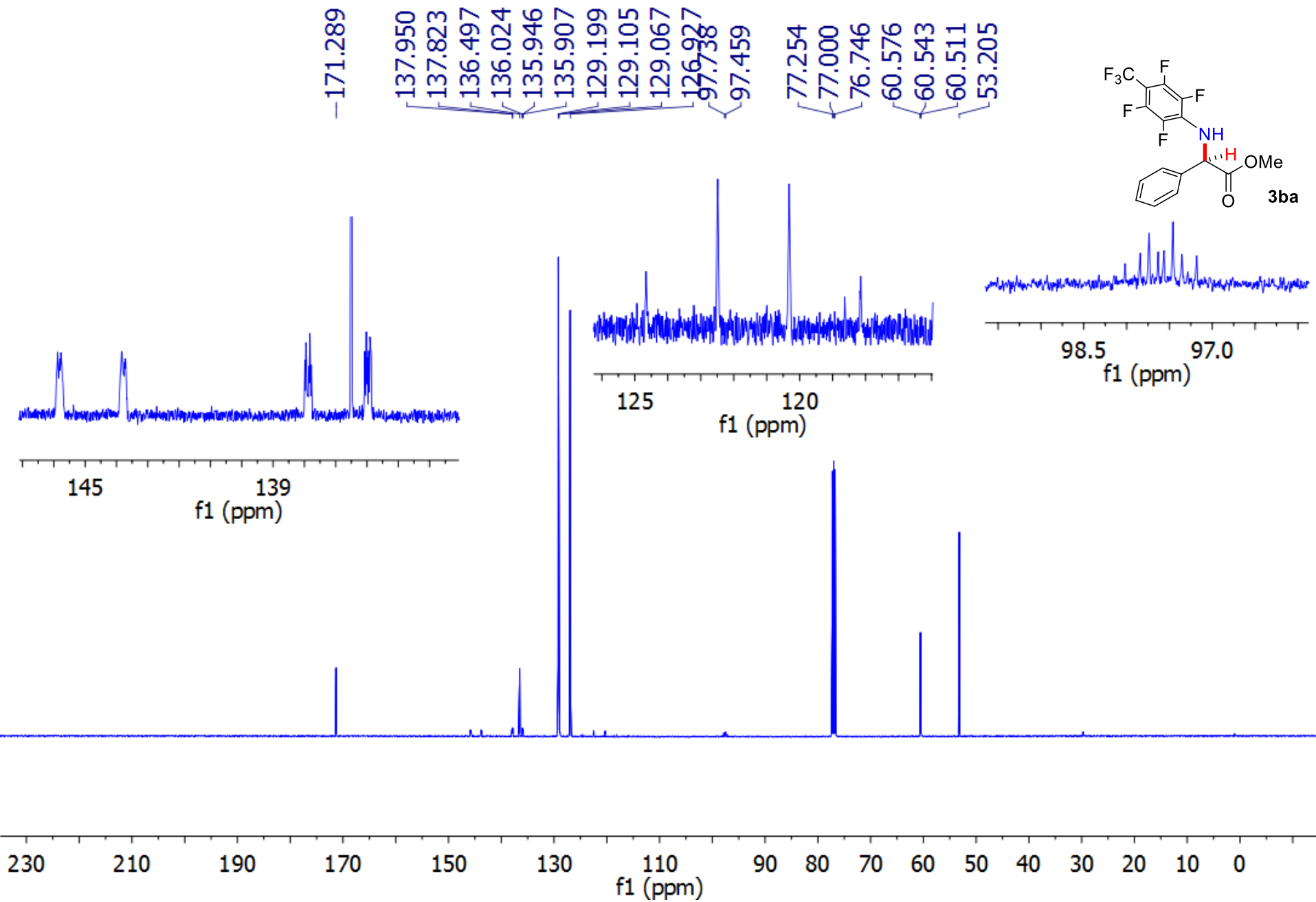


^1H NMR

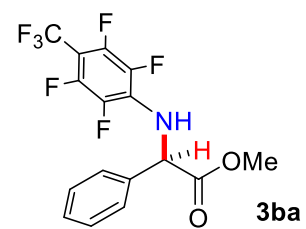
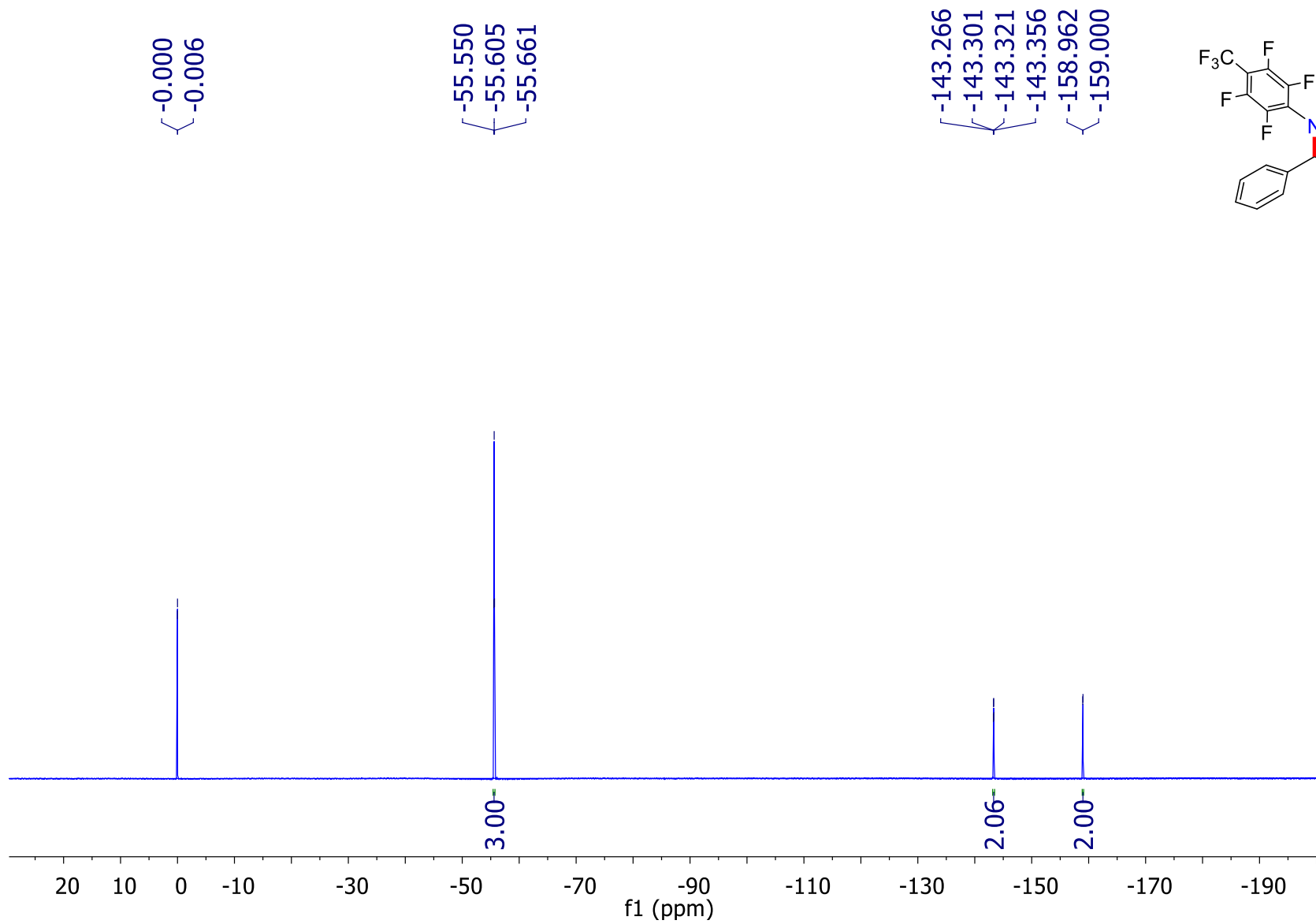
7.374
7.350
7.260
5.511
5.495
5.478
-3.765



^{13}C NMR

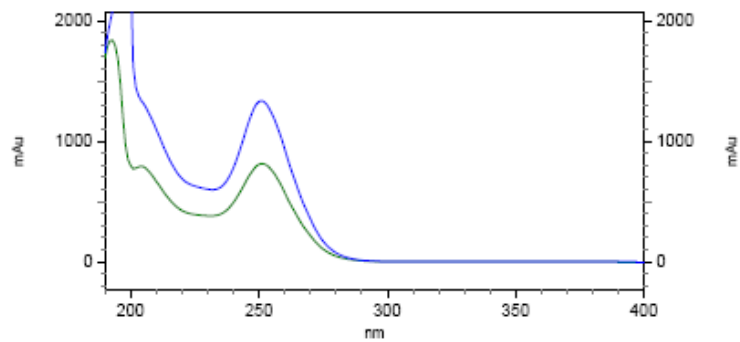
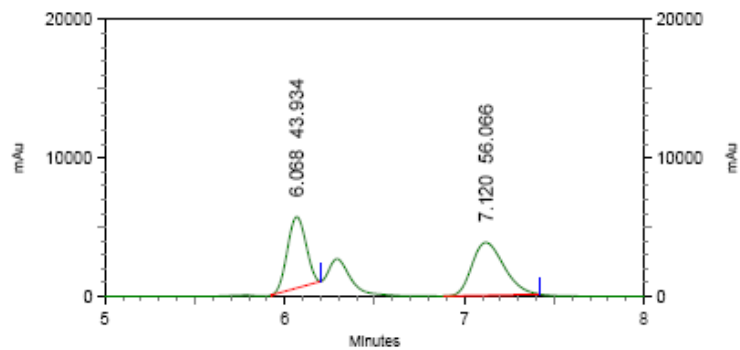
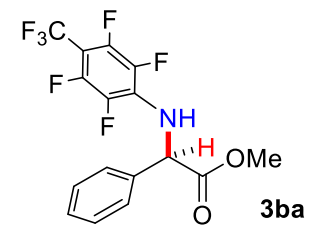


^{19}F NMR



HPLC

JLM-II-198-1b-ODH1%1ML
 C:\EZStart\Projects\Default\Method\JLM-ODH-0.2%-0.7ml.met
 C:\EZStart\Projects\Default\Data\JLM-II-198-1b-ODH1%1ML



3: 254 nm, 4 nm

Results

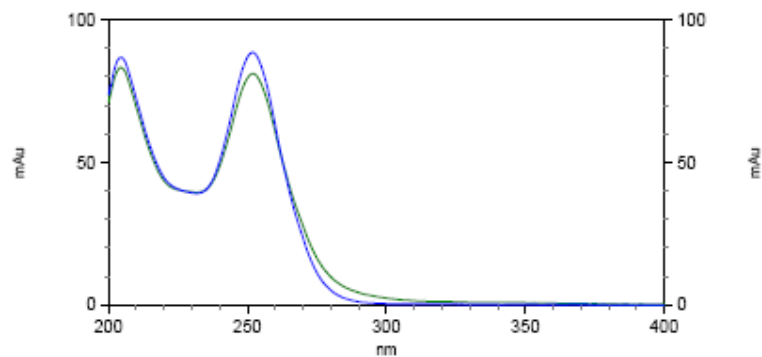
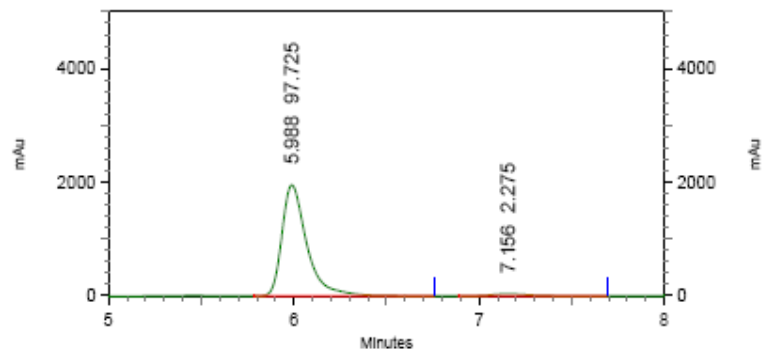
Pk #	Name	Retention Time	Area Percent
1		6.068	43.934
2		7.120	56.066
Totals			100.000

HPLC

JLM-V-167-1b-ODH1%1ML

C:\EZStart\Projects\Default\Method\JLM-ODH-0.2%-0.7ml.met

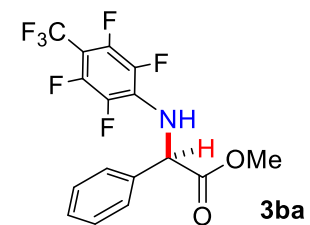
C:\EZStart\Projects\Default\Data\JLM-V-167-1b-ODH1%1ML



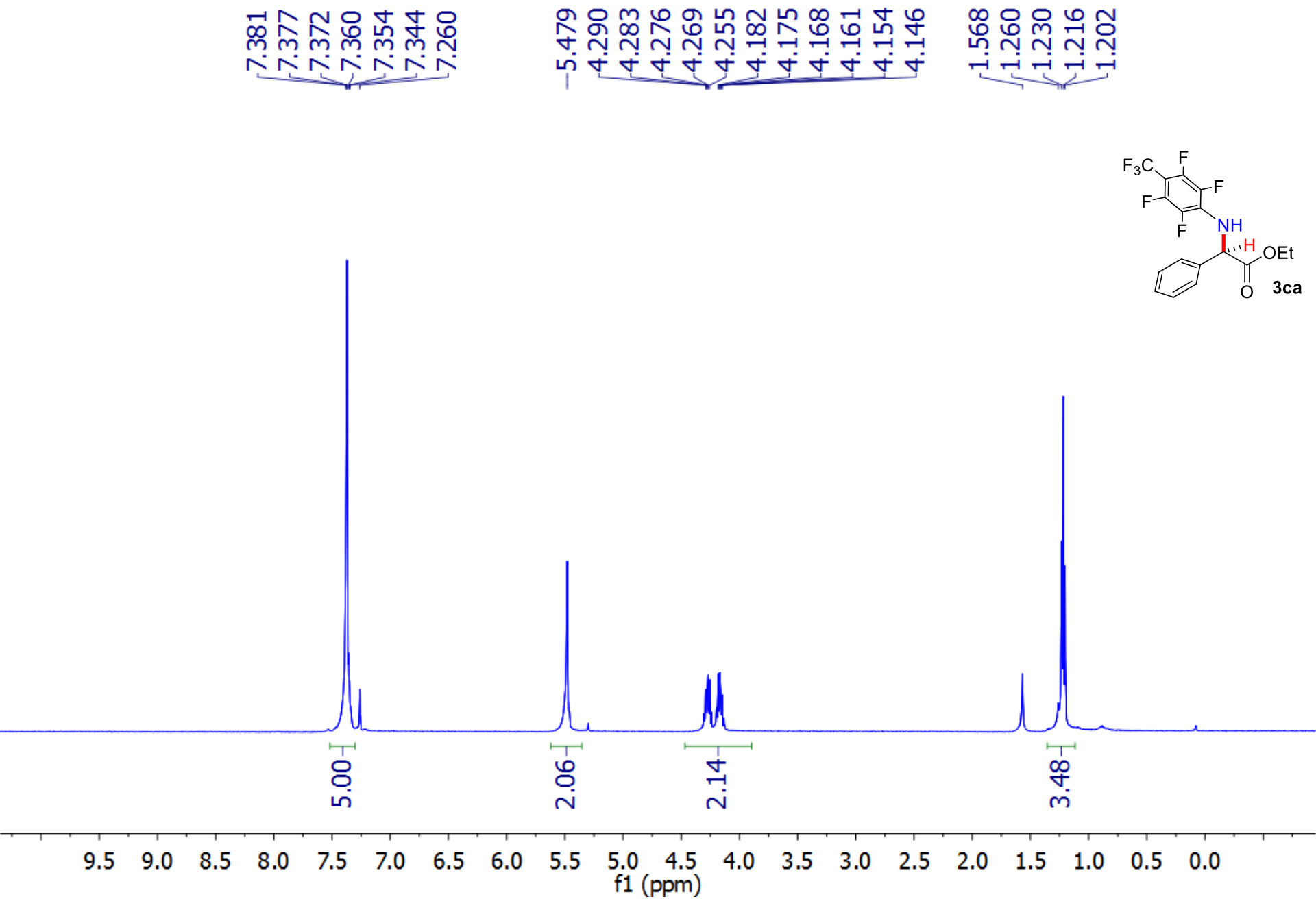
3: 275 nm, 4 nm

Results

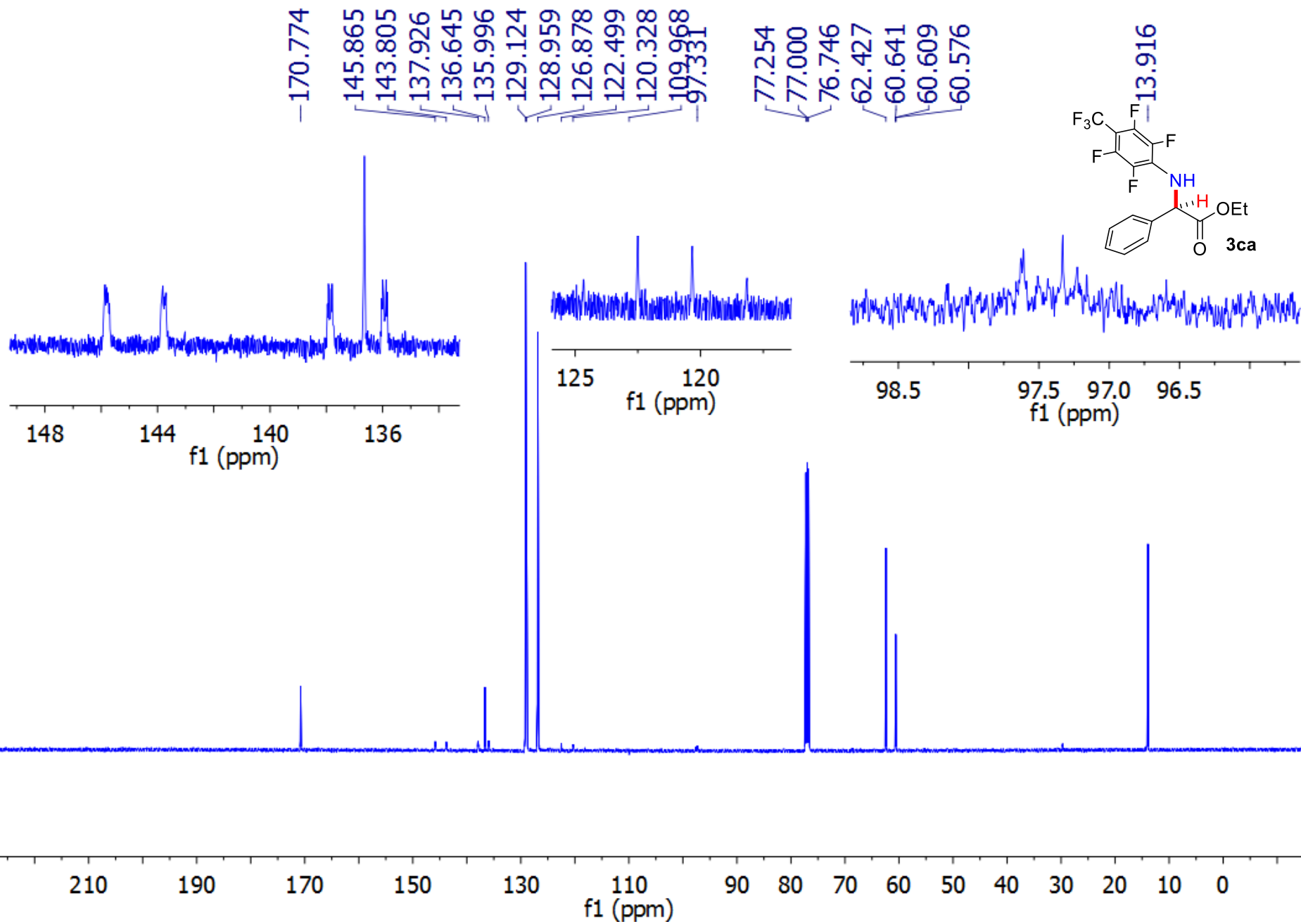
Pk #	Name	Retention Time	Area Percent
1		1.800	0.000
2		5.988	97.725
3		7.156	2.275
Totals			100.000



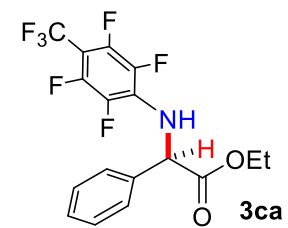
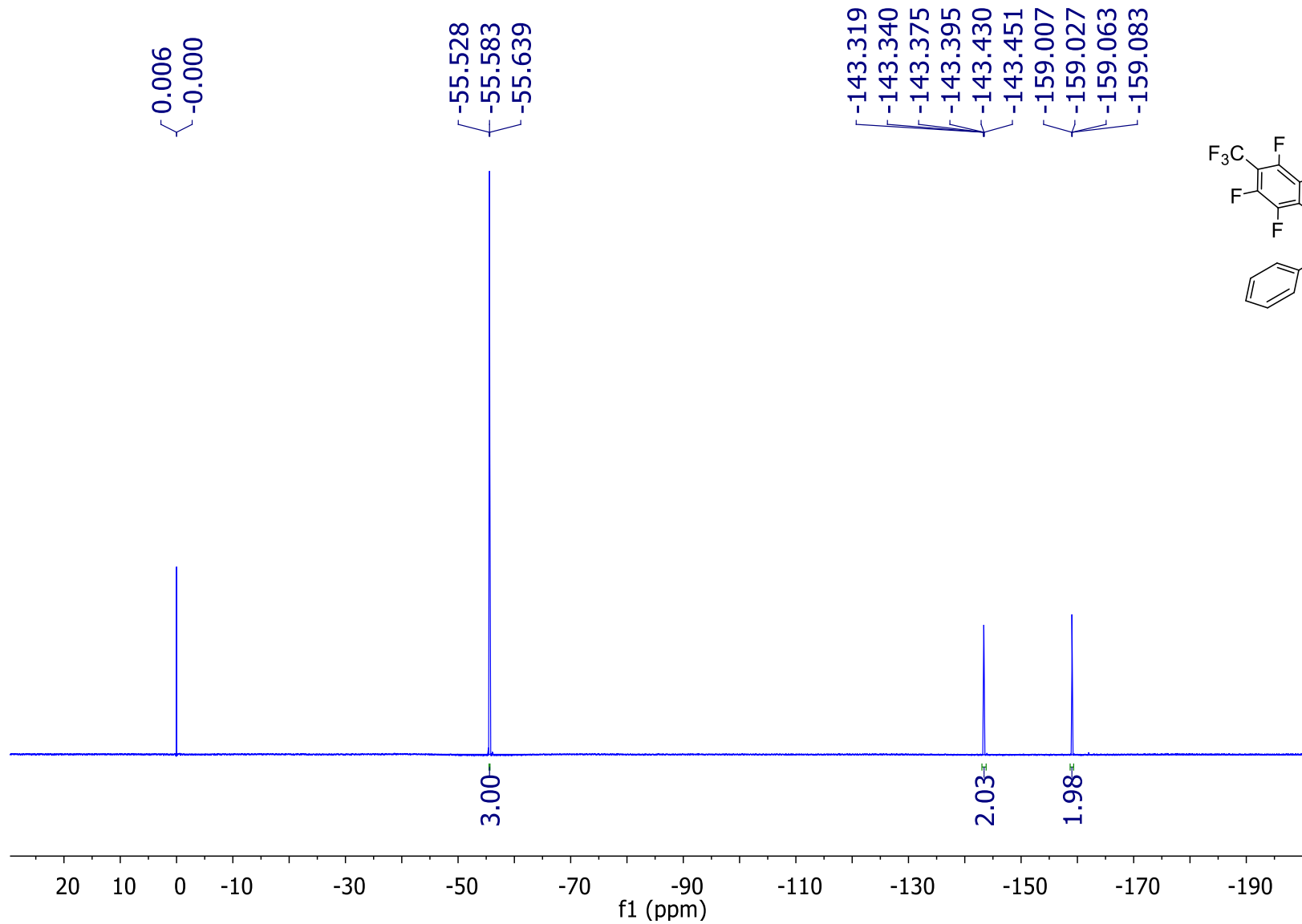
^1H NMR



^{13}C NMR

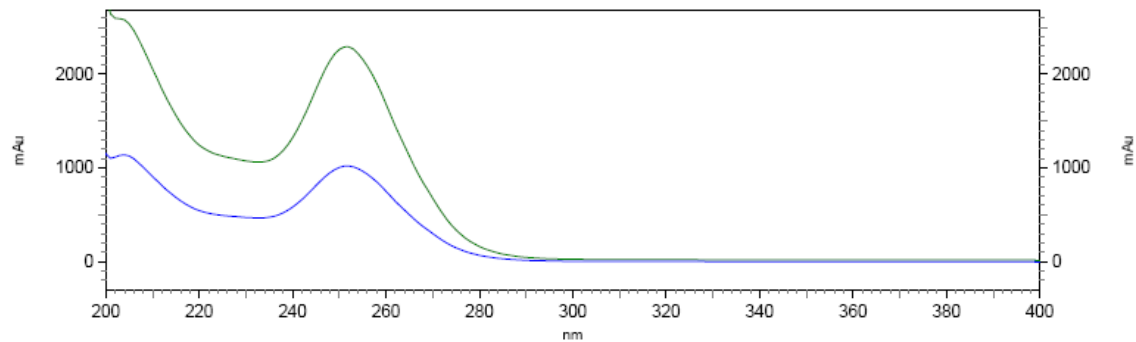
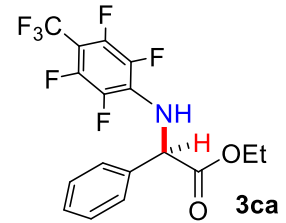
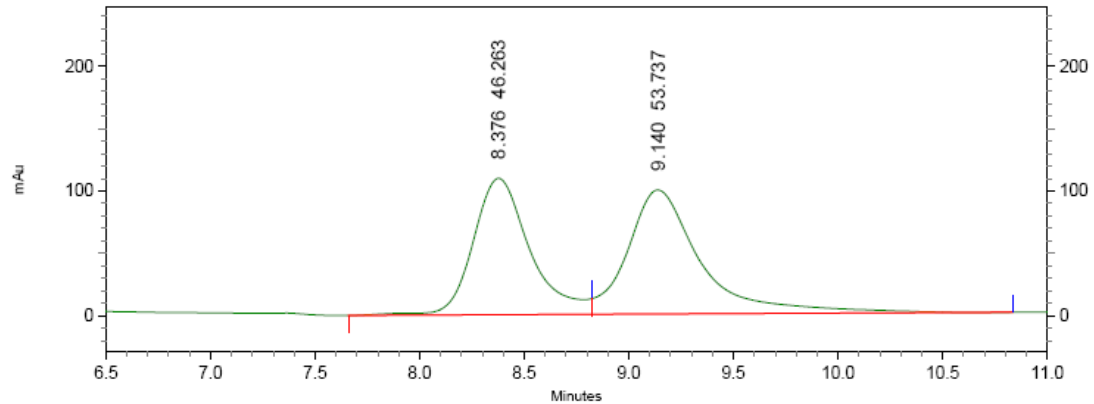


^{19}F NMR



HPLC

JLM-V-193-1-ODH-3%0.5ML
C:\EZStart\Projects\Default\Data\JLM-V-193-1-ODH-3%0.5ml
C:\EZStart\Projects\Default\Method\CQL-AD-H10%1.0ml60min.met
AD-H column 20%IPA @ 0.8ml/min

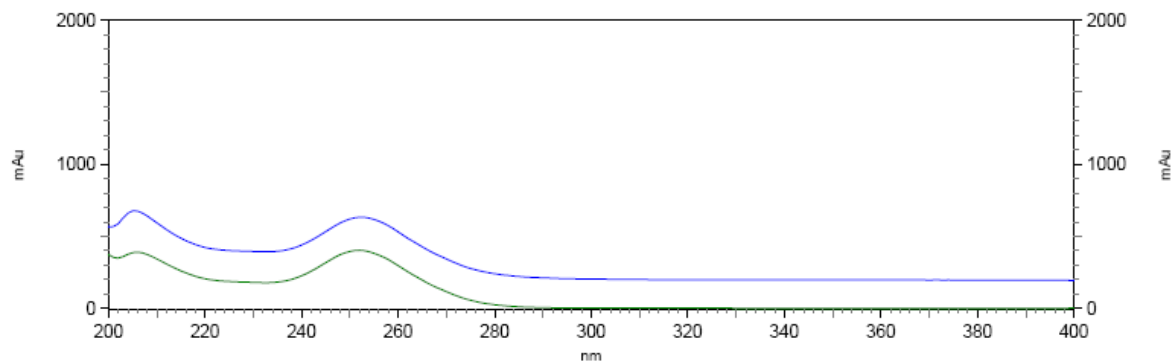
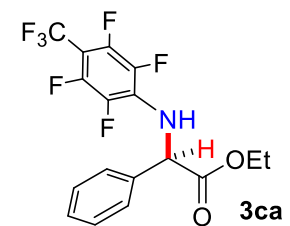
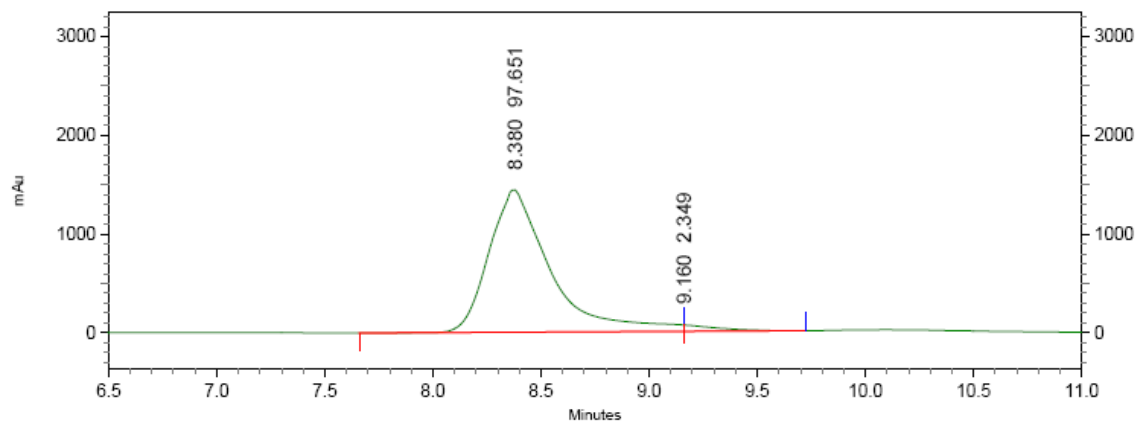


3: 273 nm, 4 nm Results

Pk #	Retention Time	Area Percent
1	8.376	46.263
2	9.140	53.737
Totals		100.000

HPLC

JLM-V-193-2-ODH-3%0.5ML
C:\EZStart\Projects\Default\Data\JLM-V-193-2-ODH-3%0.5ml
C:\EZStart\Projects\Default\Method\CQL-AD-H10%1.0ml60min.met
AD-H column 20%IPA @ 0.8ml/min

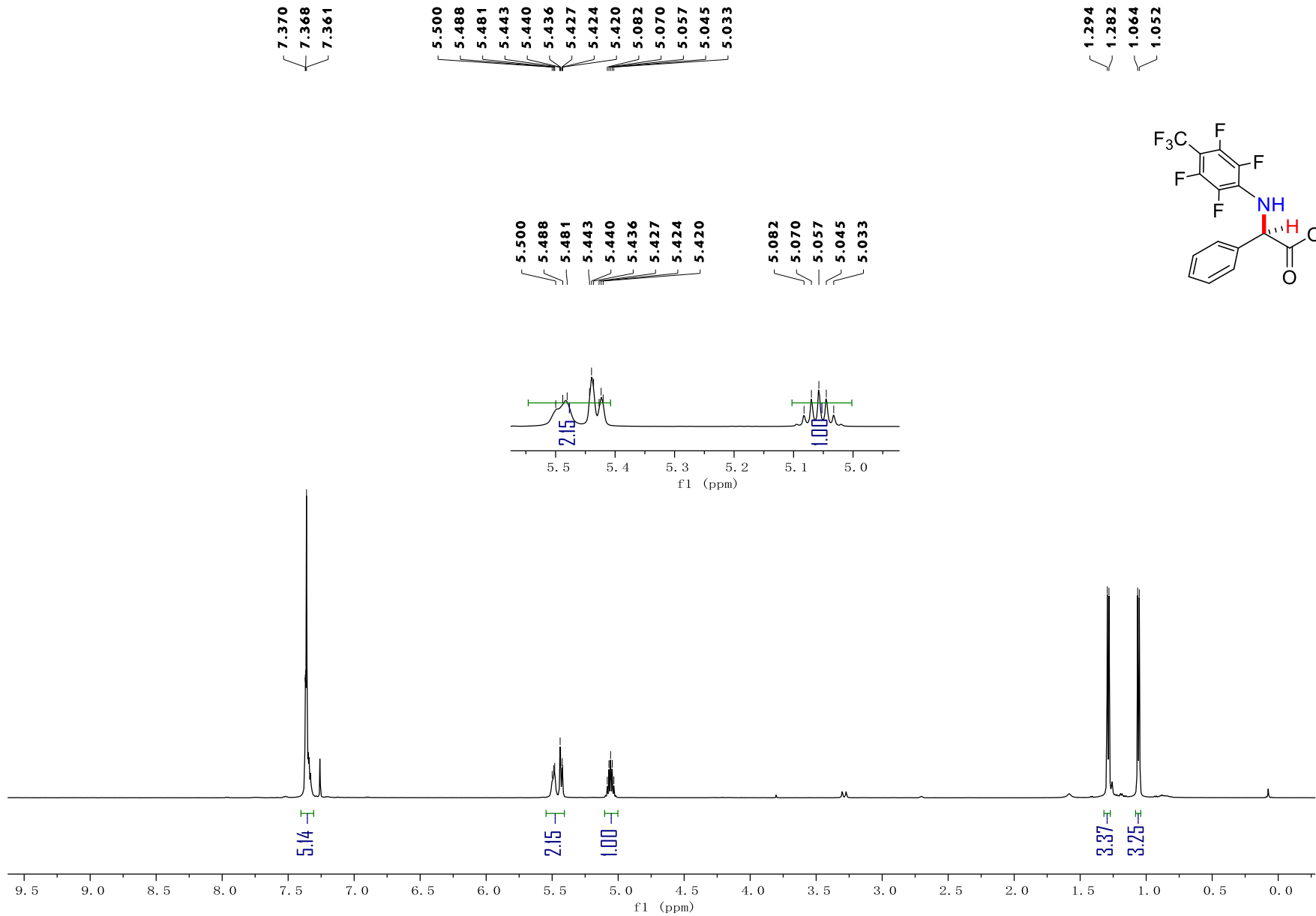


3: 255 nm, 4 nm Results

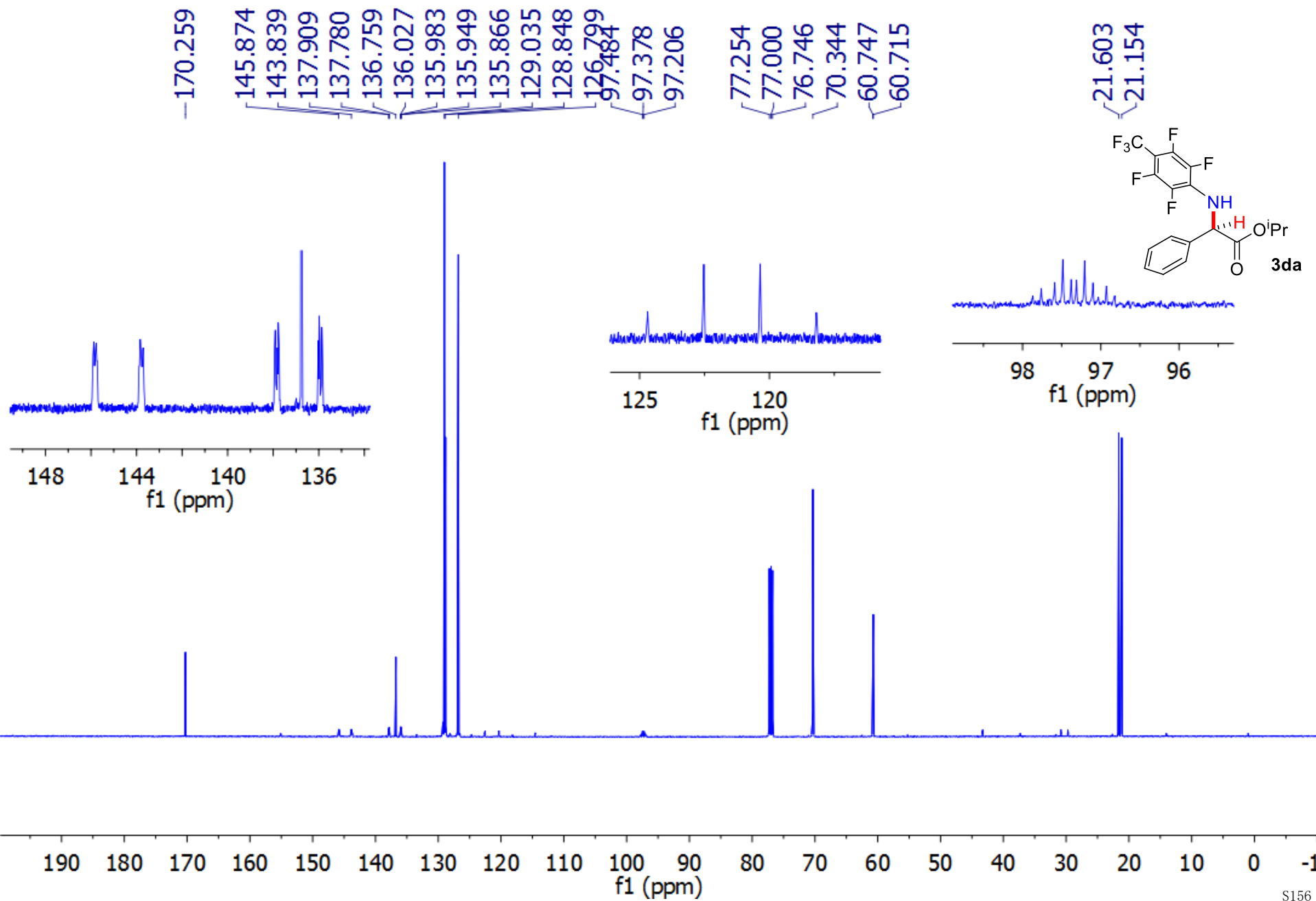
Pk #	Retention Time	Area Percent
1	8.380	97.651
2	9.160	2.349

Totals		100.000
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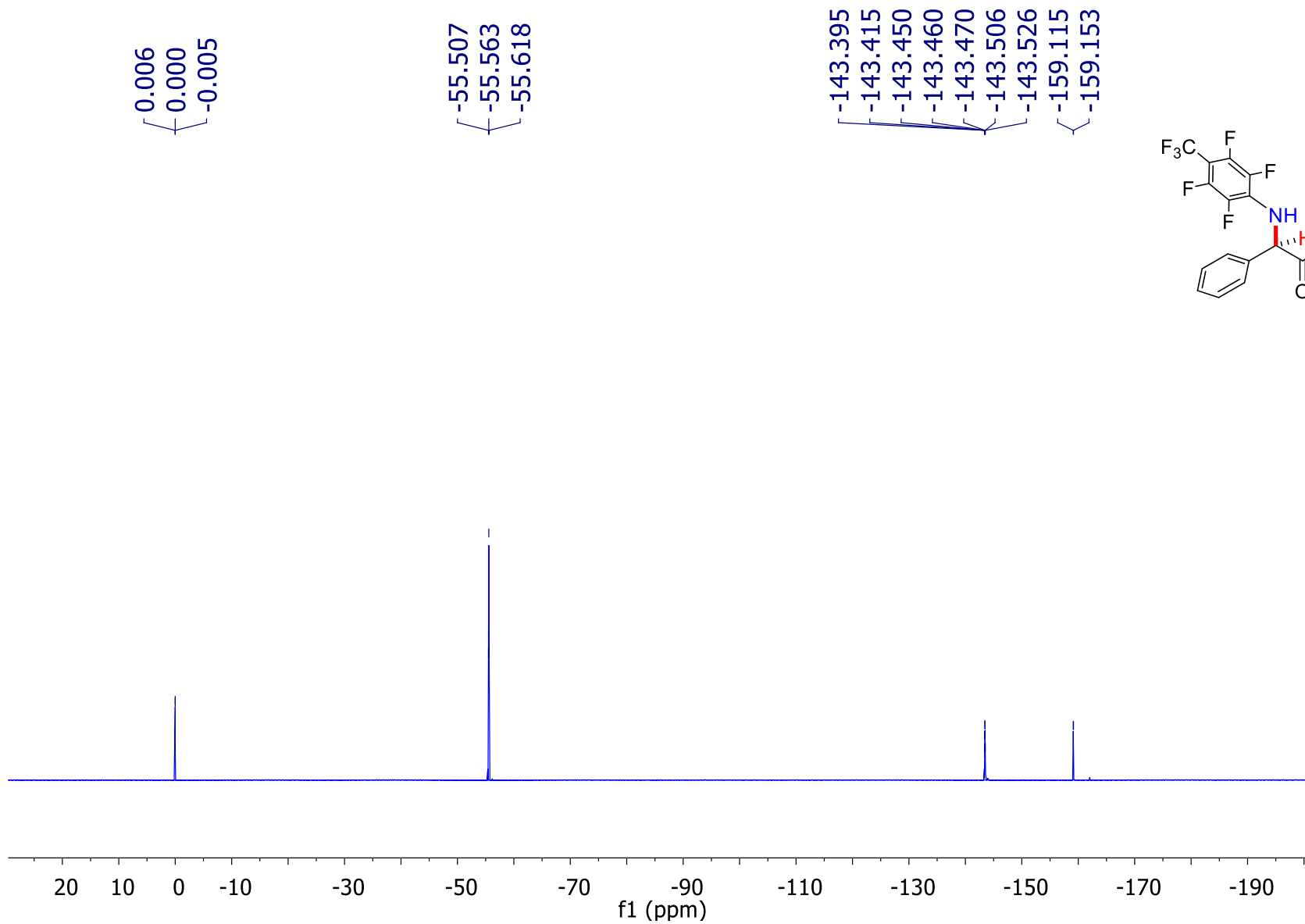
^1H NMR



¹³C NMR



^{19}F NMR

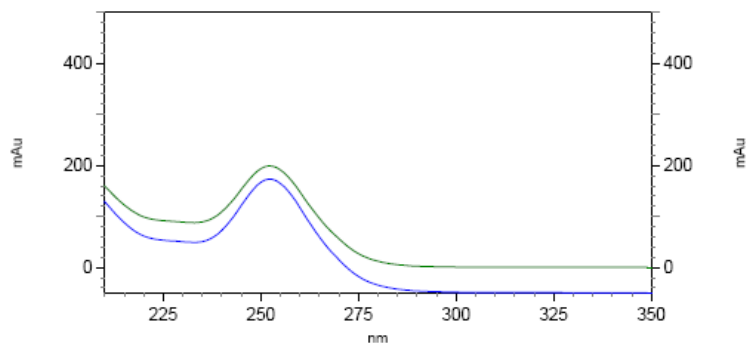
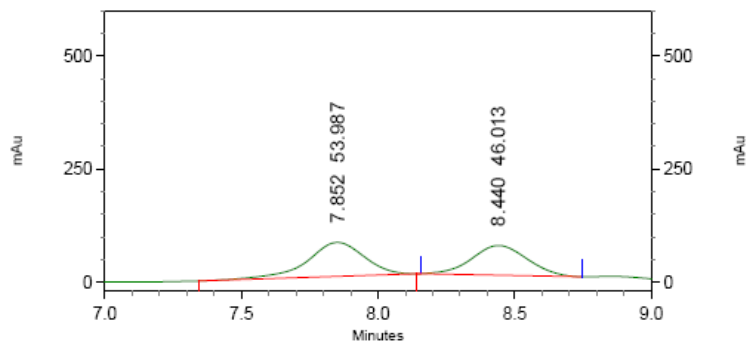


HPLC

JLM-V-168-1A-whe1k-0%1ML

C:\EZStart\Projects\Default\Method\JLM-ODH-0%-0.7ml.met

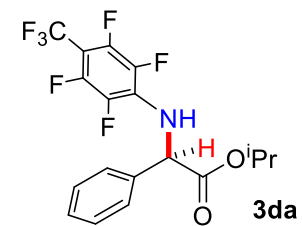
C:\EZStart\Projects\Default\Data\JLM-V-168-1A-whe1k-0%1ML



3: 259 nm, 4 nm

Results

Pk #	Name	Retention Time	Area Percent
1		7.852	53.987
2		8.440	46.013
Totals			100.000

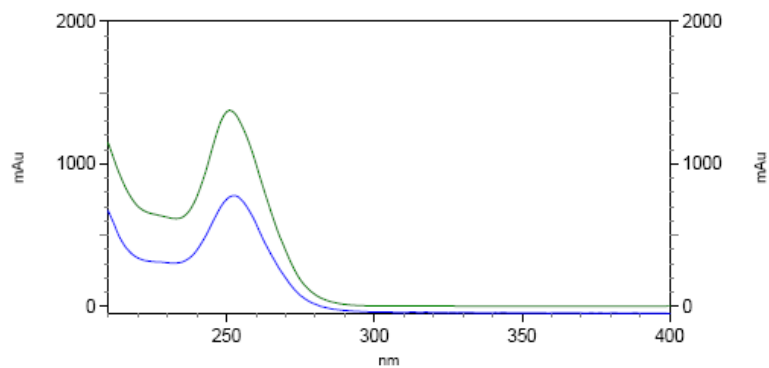
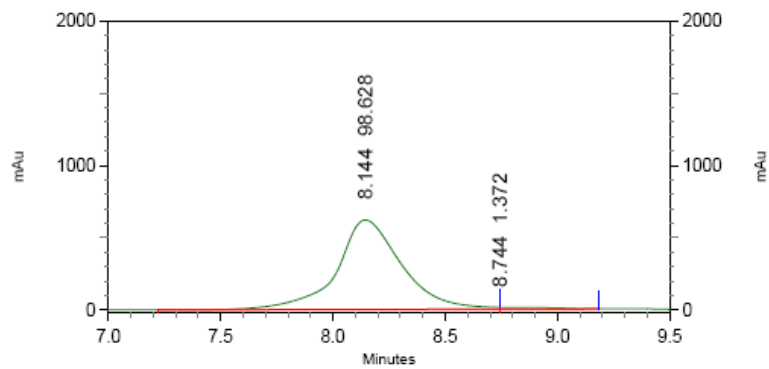


HPLC

JLM-V-173-wheIk-0%1ML

C:\EZStart\Projects\Default\Method\JLM-ODH-0%-0.7ml.met

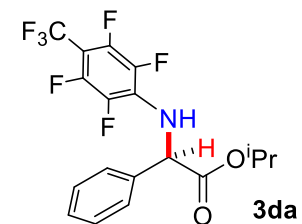
C:\EZStart\Projects\Default\Data\JLM-V-173-wheIk-0%1ML



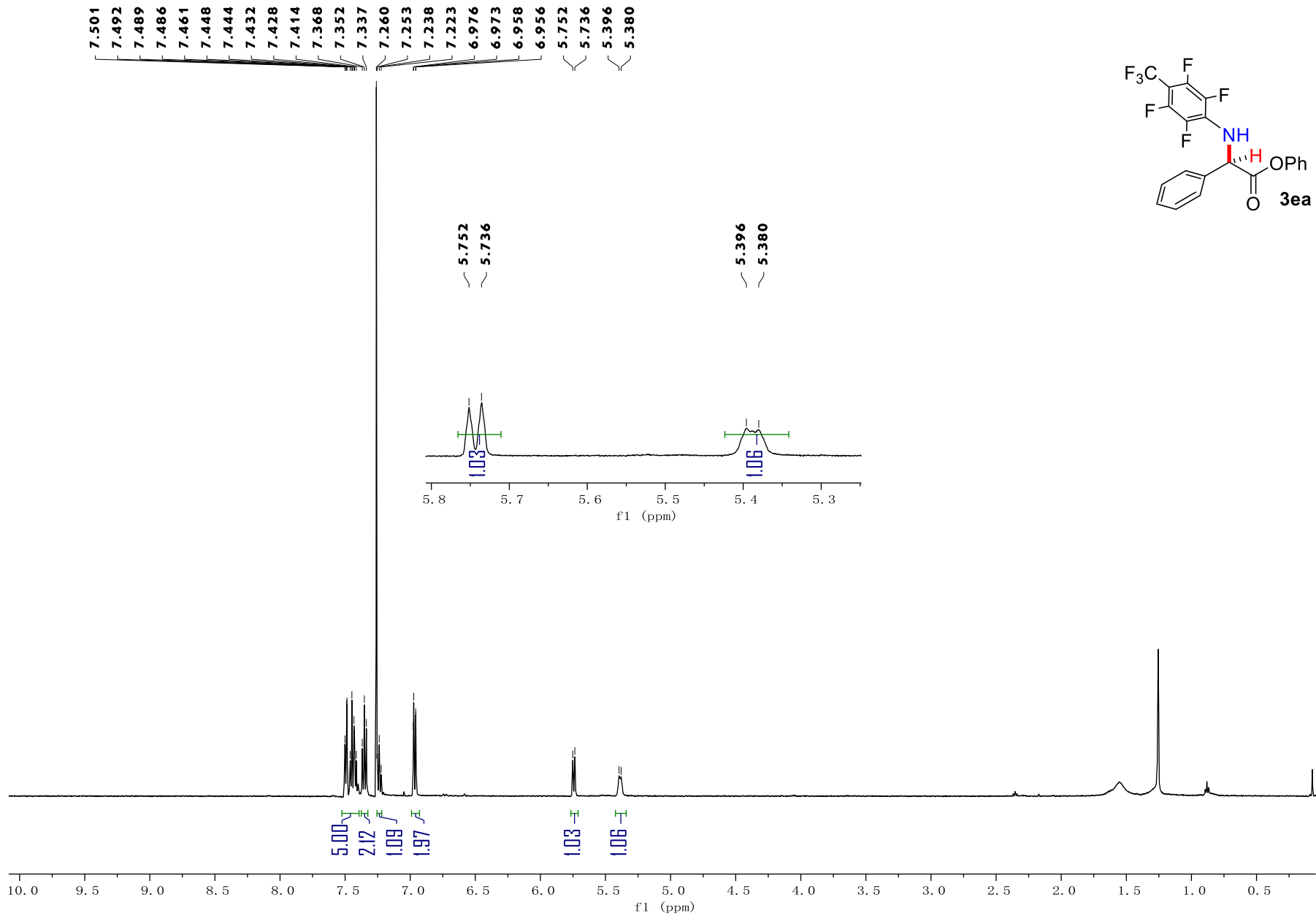
3: 258 nm, 4 nm

Results

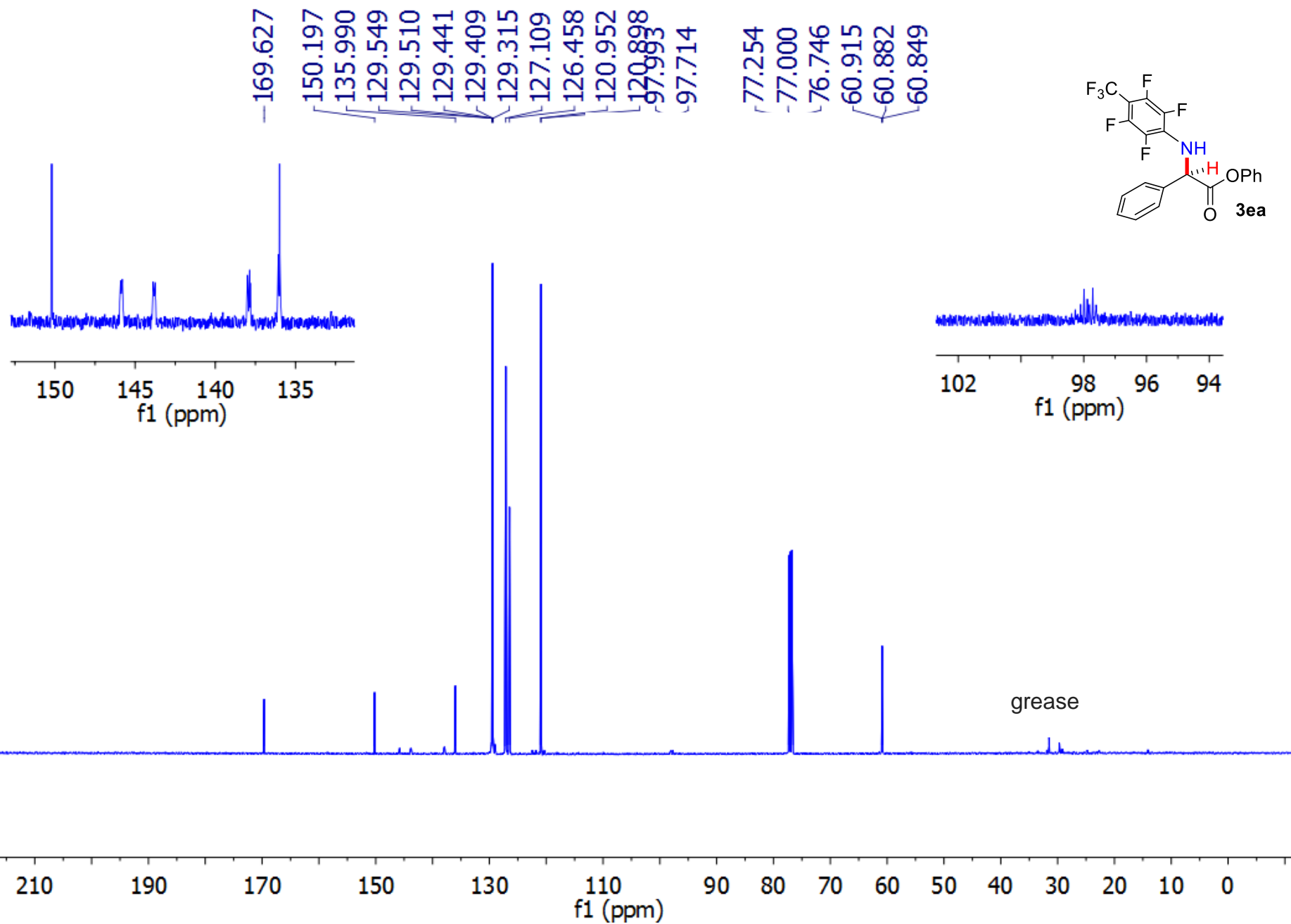
Pk #	Name	Retention Time	Area Percent
1		8.144	98.628
2		8.744	1.372
Totals			100.000



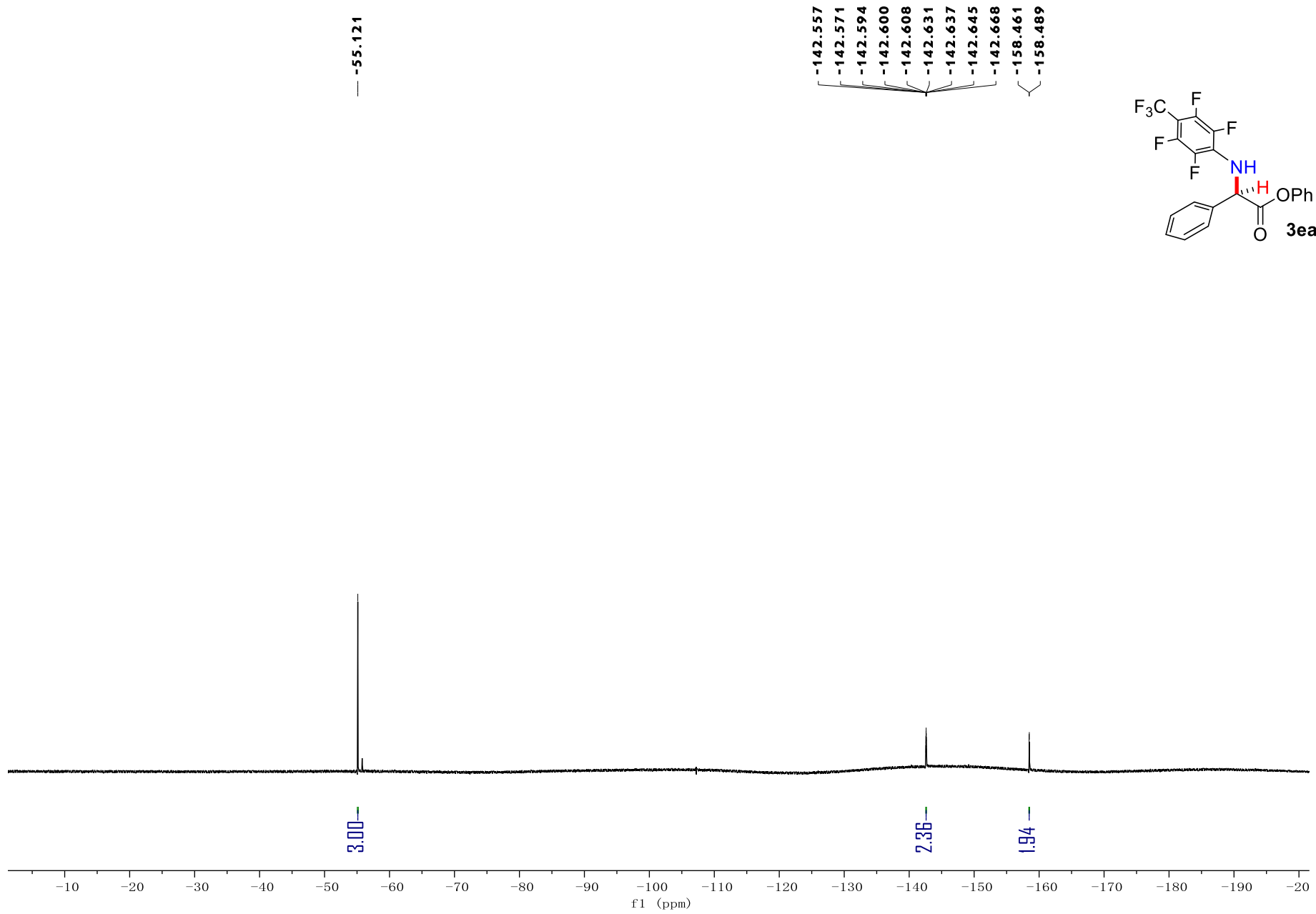
¹H NMR



^{13}C NMR



^{19}F NMR

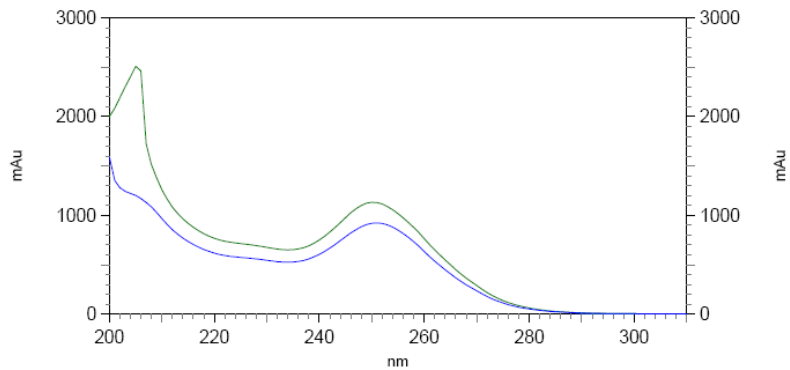
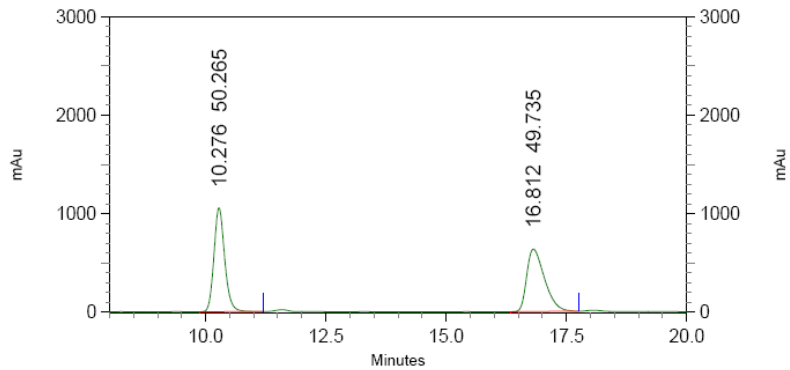


HPLC

JLM-II-243-1-ADH-1%0.7ML

C:\EZStart\Projects\Default\Method\XC-5%-ADH1ml.met

C:\EZStart\Projects\Default\Data\JLM-II-243-1-ADH-1%0.7ML

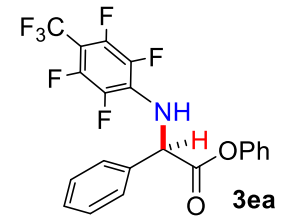


4: 241 nm, 4 nm

Results

Name	Retention Time	Area Percent	Pk #
	10.276	50.265	1
	16.812	49.735	2

Totals		100.000	
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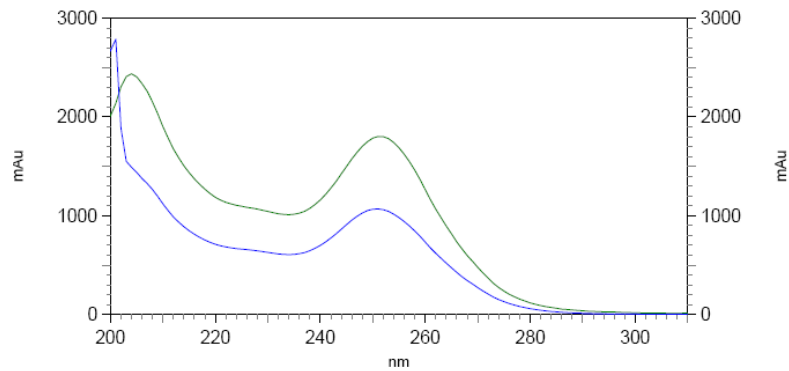
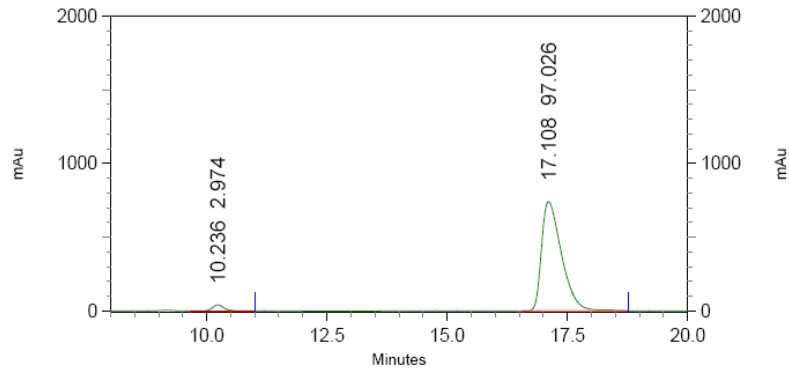
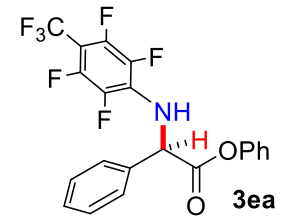


HPLC

JLM-V-199-1A-ADH-1%0.7ML

C:\EZStart\Projects\Default\Method\XC-5%-ADH1ml.met

C:\EZStart\Projects\Default\Data\JLM-V-199-1A-ADH-1%0.7ML



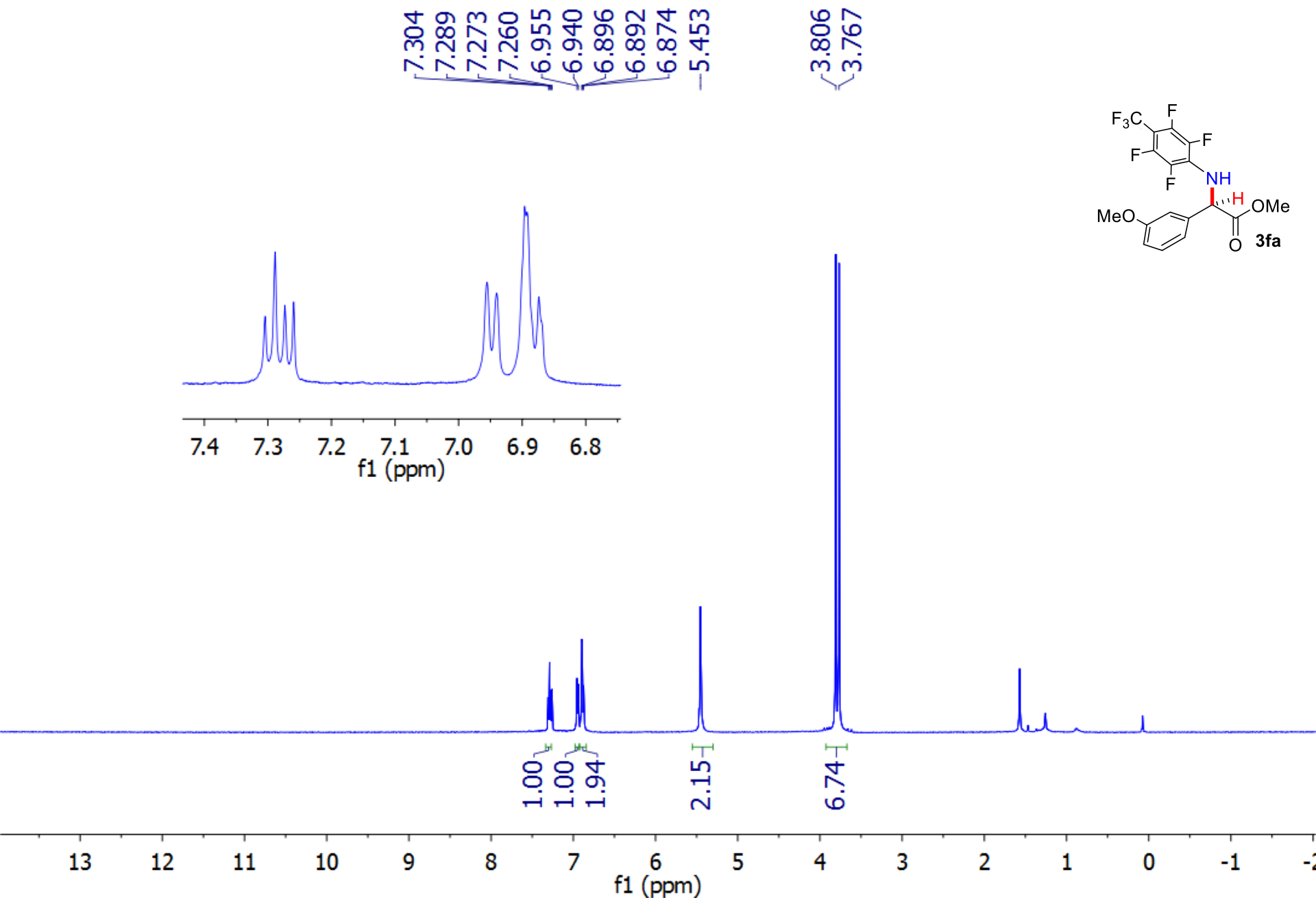
4: 241 nm, 4 nm

Results

Name	Retention Time	Area Percent	Pk #
	10.236	2.974	1
	17.108	97.026	2

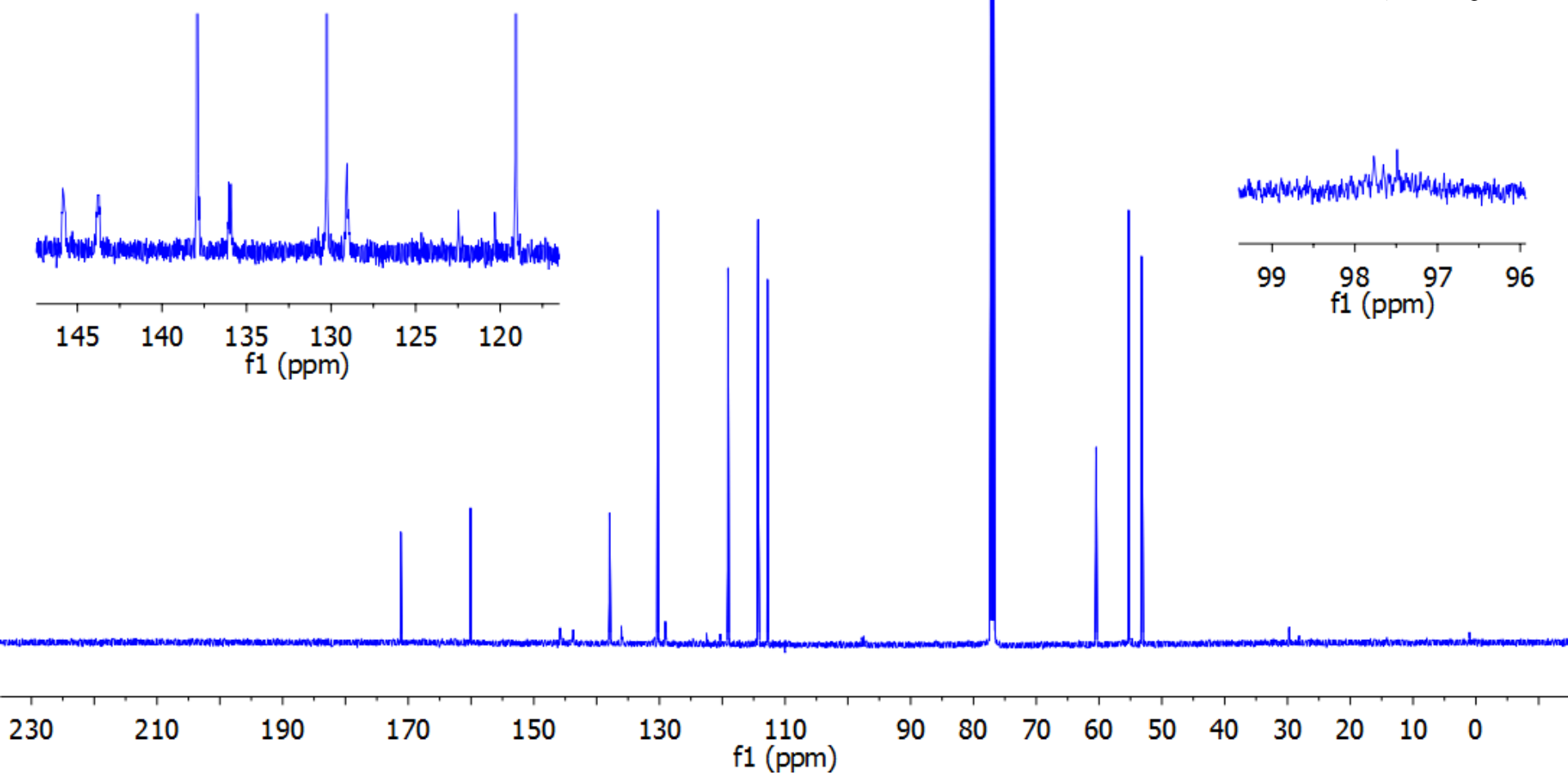
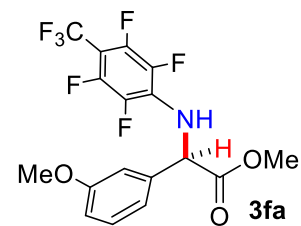
Totals	100.000	
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¹H NMR

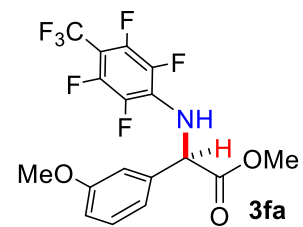
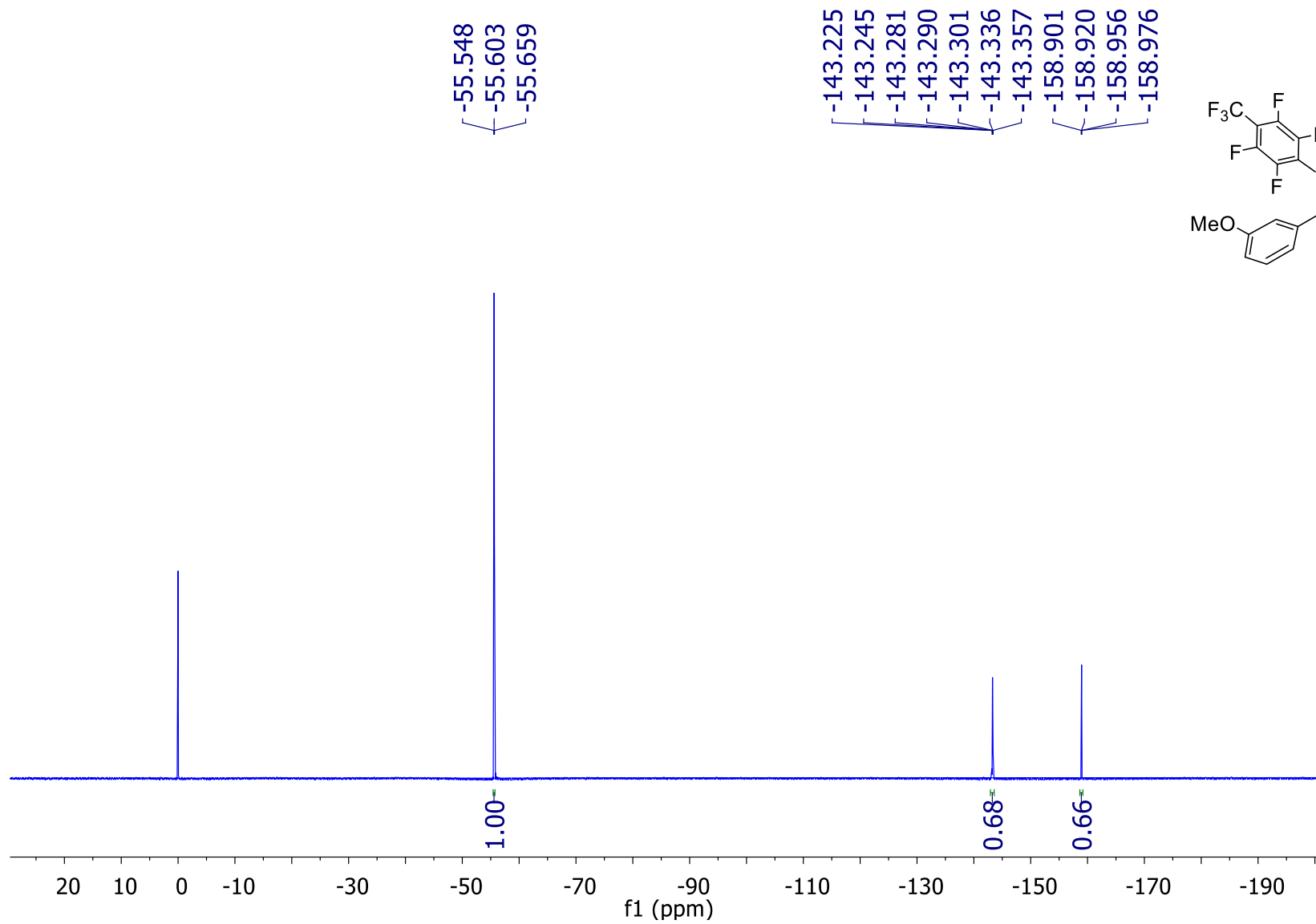


¹³C NMR

-171.183
-160.091
145.867
137.934
136.049
135.928
130.263
129.060
119.085
114.275
112.791
97.732
97.487
77.254
77.000
76.746
60.506
60.473
60.440
55.298
53.236



^{19}F NMR

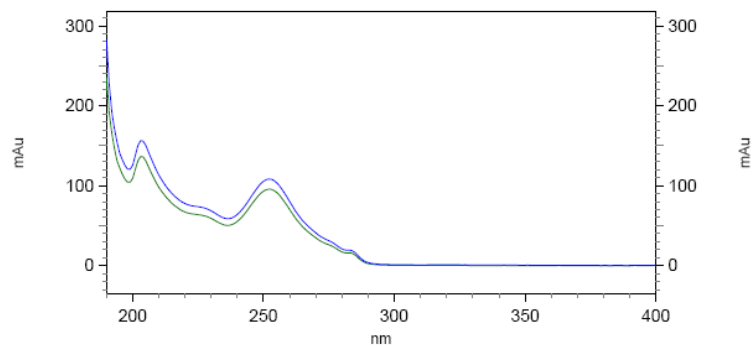
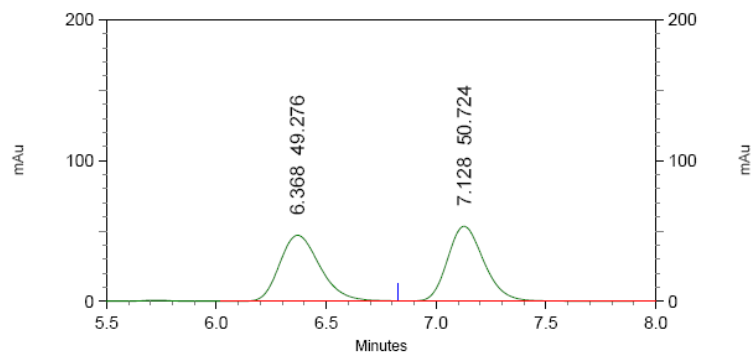


HPLC

JLM-V-209-1-ADH-1%1ML

C:\EZStart\Projects\Default\Method\shifatest_2,5-dimehoxy.met

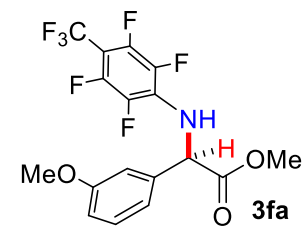
C:\EZStart\Projects\Default\Data\JLM-V-209-1-ADH1%1ML



7: 251 nm, 4 nm

Results

Pk #	Name	Retention Time	Area Percent
1		6.368	49.276
2		7.128	50.724
Totals			100.000

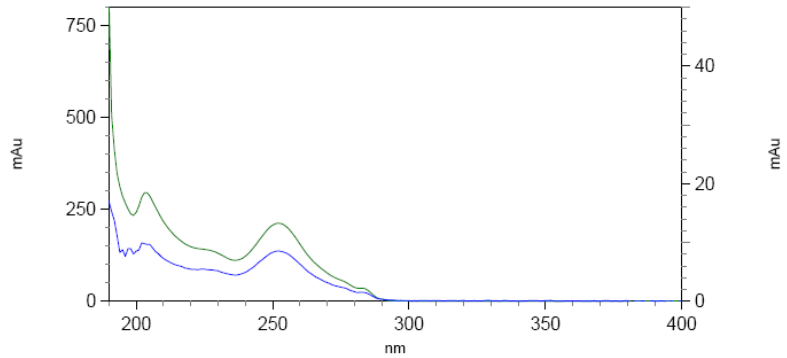
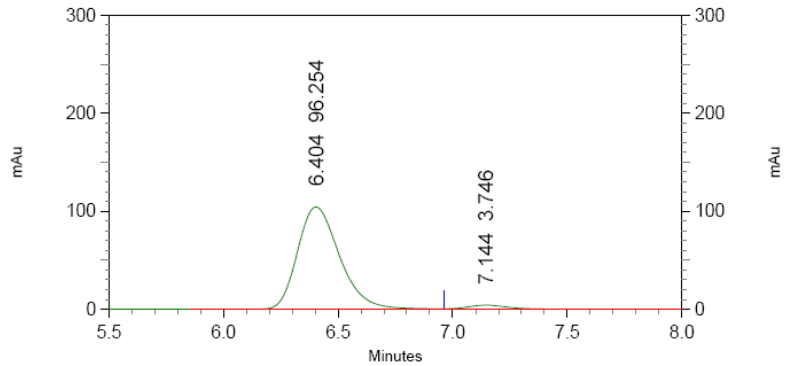


HPLC

JLM-V-209-2-ADH-1%1ML

C:\EZStart\Projects\Default\Method\shifatest_2,5-dimehoxy.met

C:\EZStart\Projects\Default\Data\JLM-V-209-2-ADH1%1ML

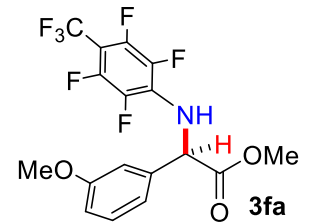


7: 251 nm, 4 nm

Results

Pk #	Name	Retention Time	Area Percent
1		6.404	96.254
2		7.144	3.746

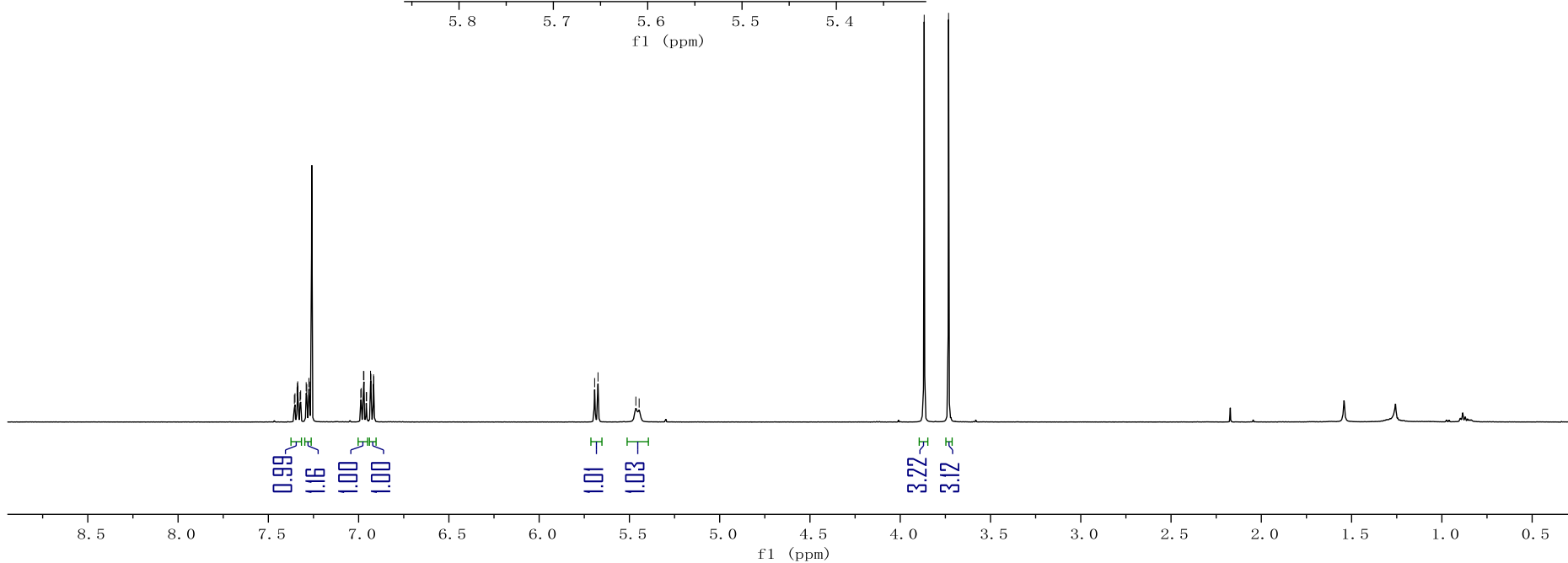
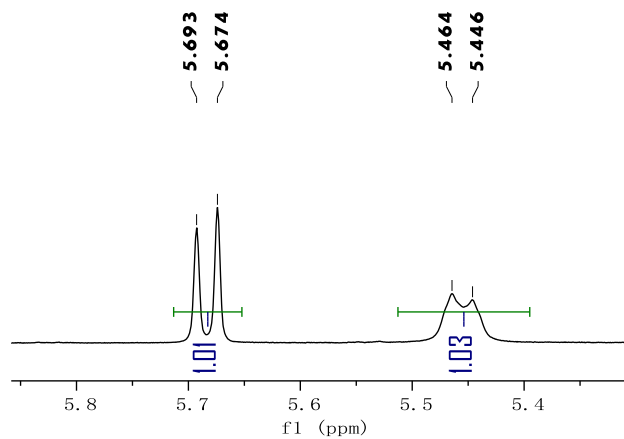
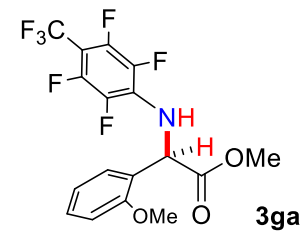
Totals	Area Percent
	100.000



¹H NMR

7.341
7.339
7.337
7.336
7.324
7.321
7.291
7.287
7.275
7.272
6.988
6.986
6.973
6.971
6.958
6.956
6.934
6.932
6.917
5.693
5.691
5.693
5.674
5.674
5.464
5.446

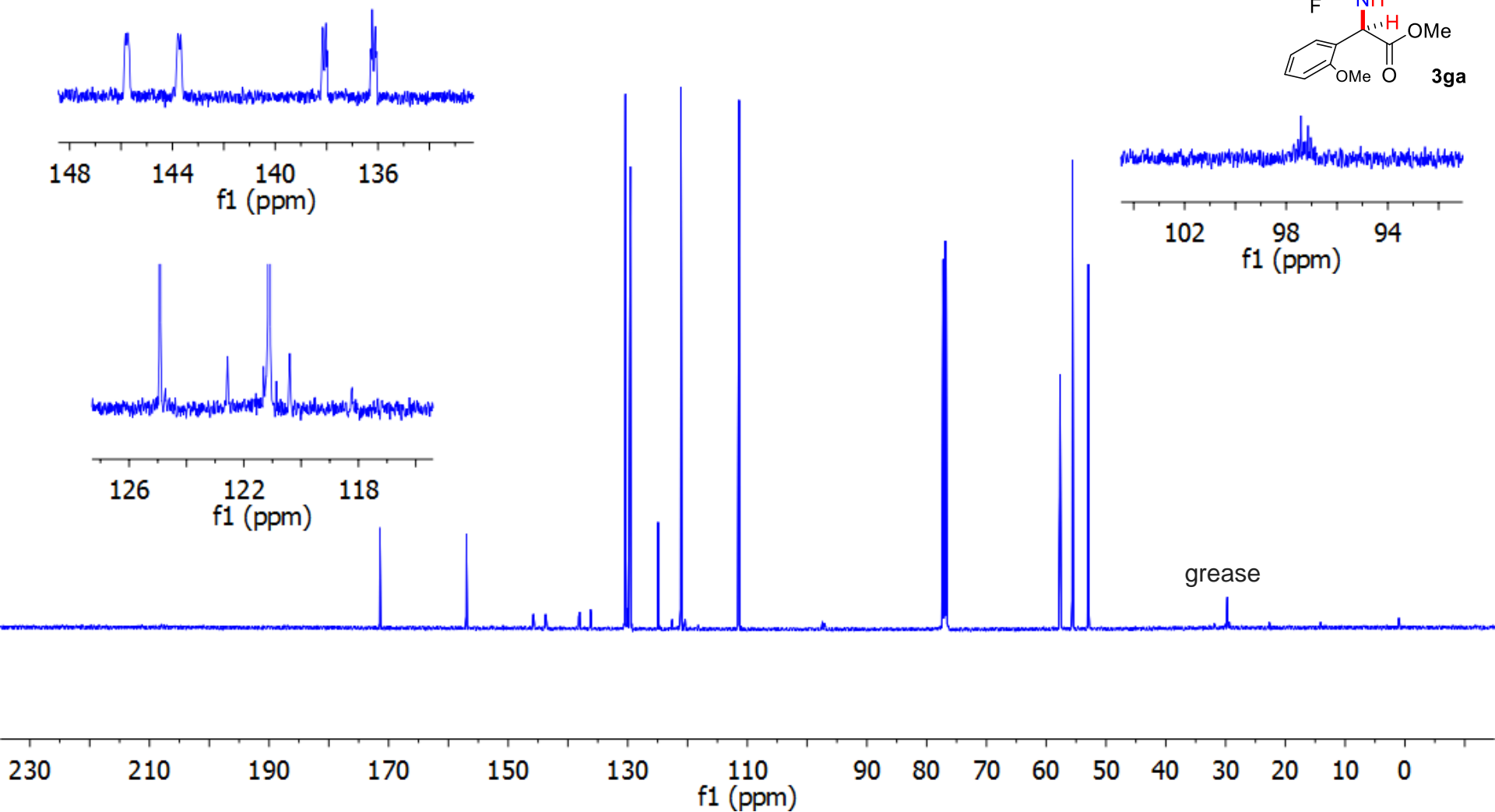
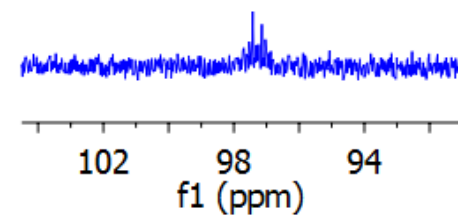
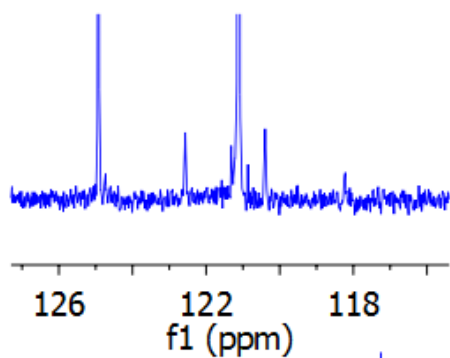
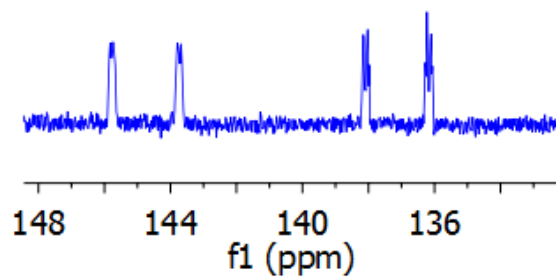
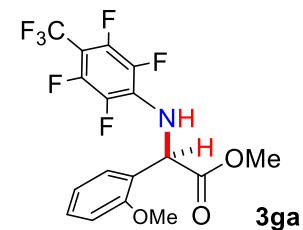
3.867
3.732



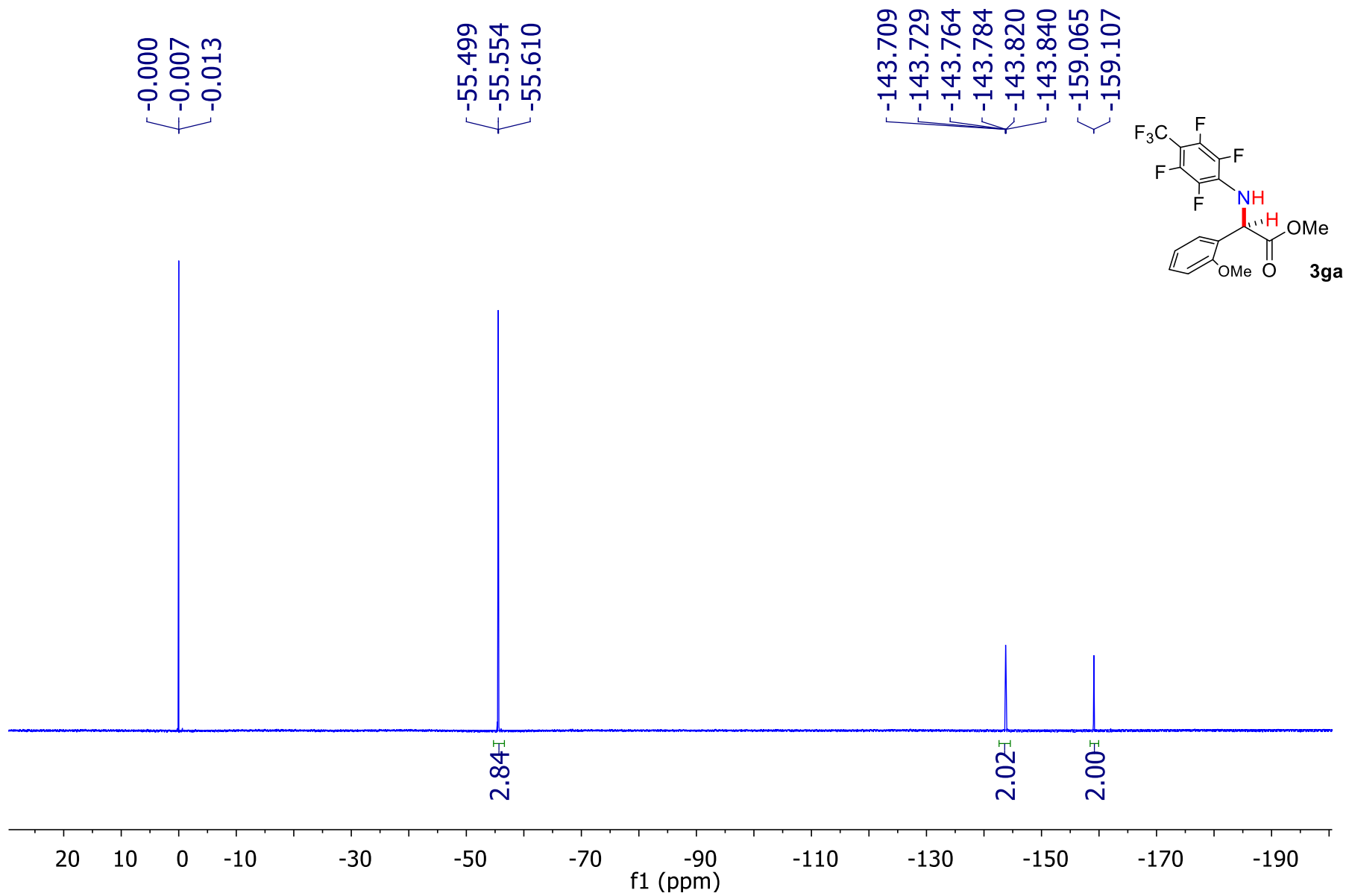
^{13}C NMR

171.450
156.989
145.744
138.154
138.026
136.228
136.099
130.366
129.568
124.921
121.115
111.381
97.419

77.254
77.000
76.746
57.733
57.697
57.662
55.622
52.927

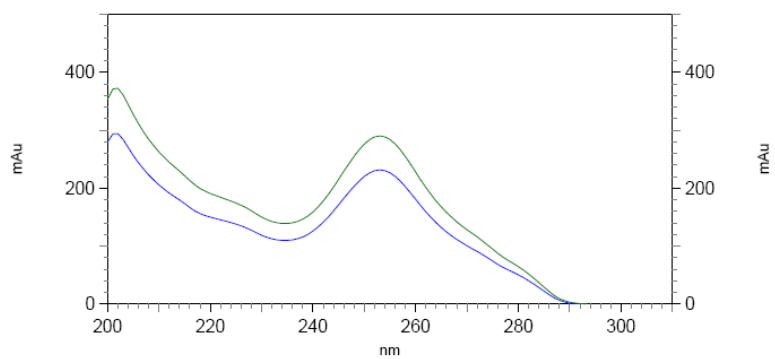
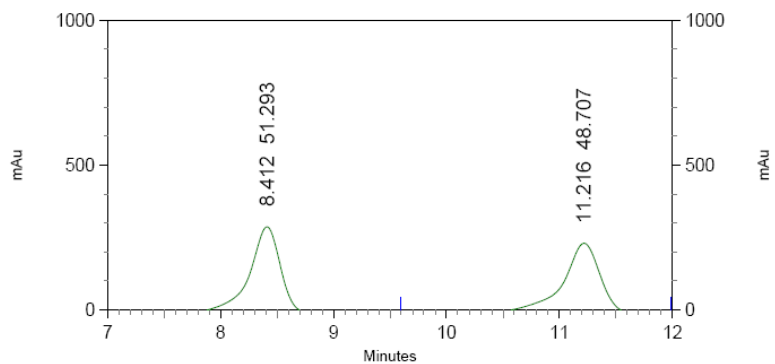


^{19}F NMR



HPLC

JLM-II-207-1-Whelk-0.5@-1mL
C:\EZStart\Projects\Default\Method\XC-5%-ADH1ml.met
E:\JLM-II-207-1-Whelk-0.5 @1ml

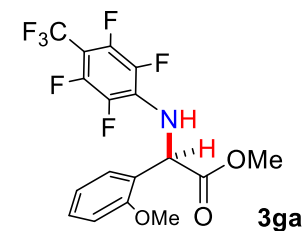


4: 254 nm, 4 nm

Results

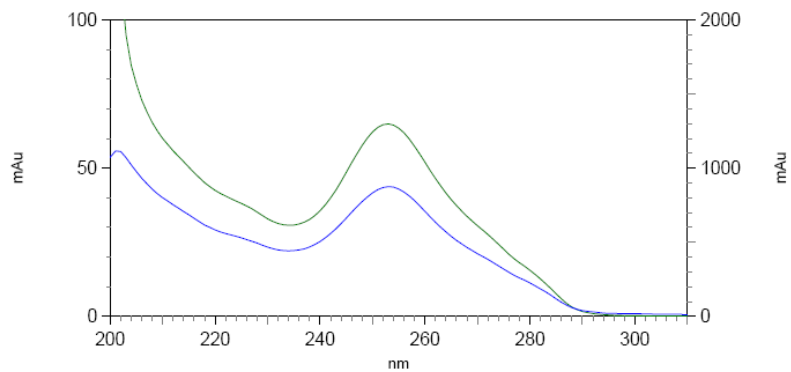
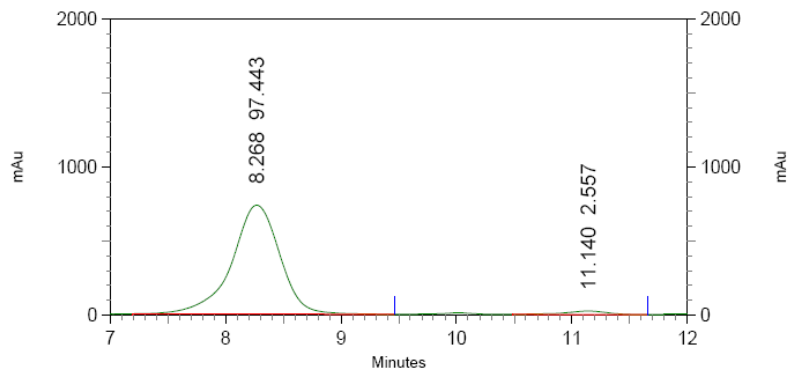
Name	Retention Time	Area Percent	Pk #
	8.412	51.293	1
	11.216	48.707	2

Totals	100.000
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HPLC

JLM-V-198-1-WHELK-0.5@-1mL
 C:\EZStart\Projects\Default\Method\XC-5%-ADH1ml.met
 E:\JLM-V-198-1-WHELK0.5@1ml

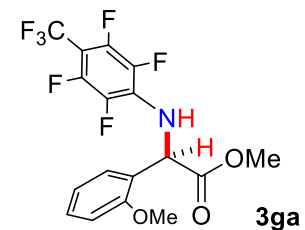


4: 284 nm, 4 nm

Results

Name	Retention Time	Area Percent	Pk #
	8.268	97.443	1
	11.140	2.557	2

Totals		100.000	
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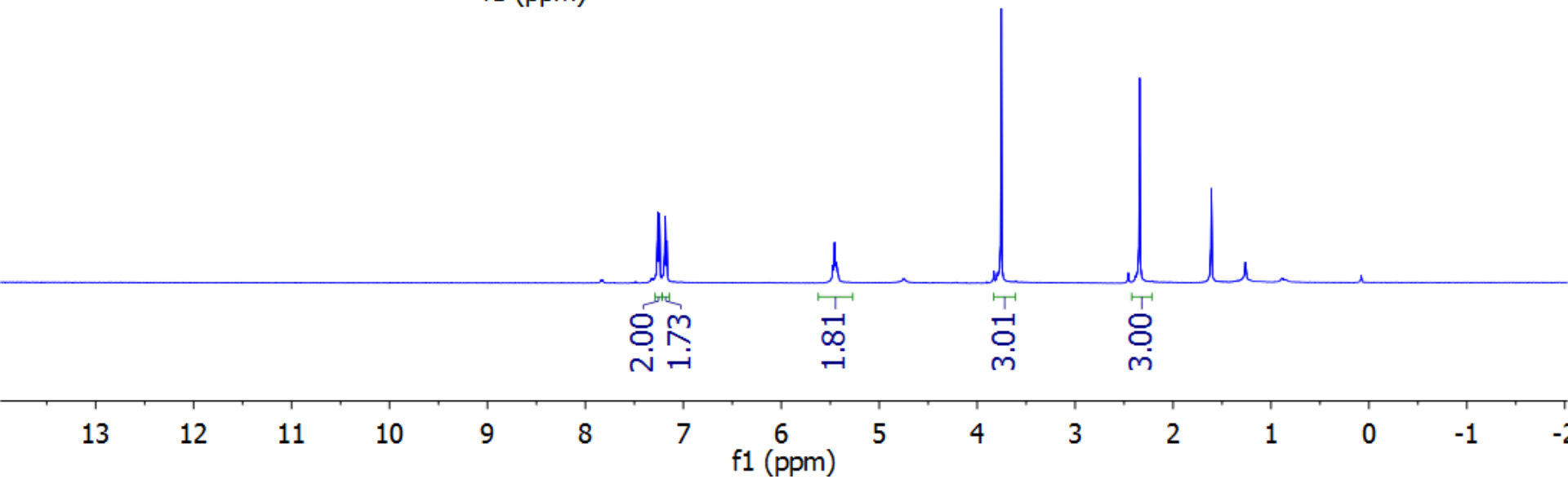
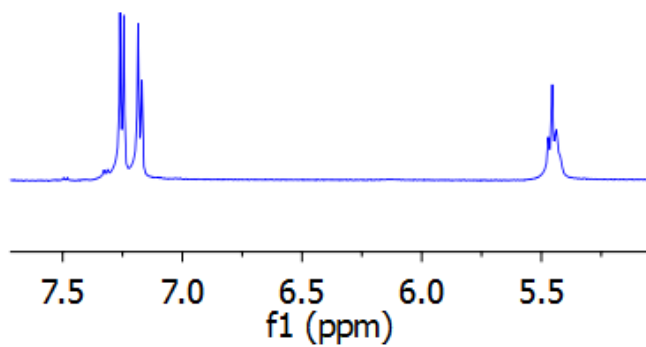
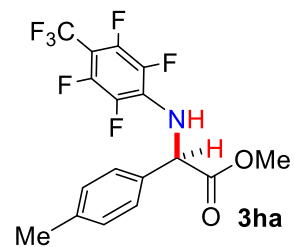
¹H NMR

7.260
7.244
7.185
7.169

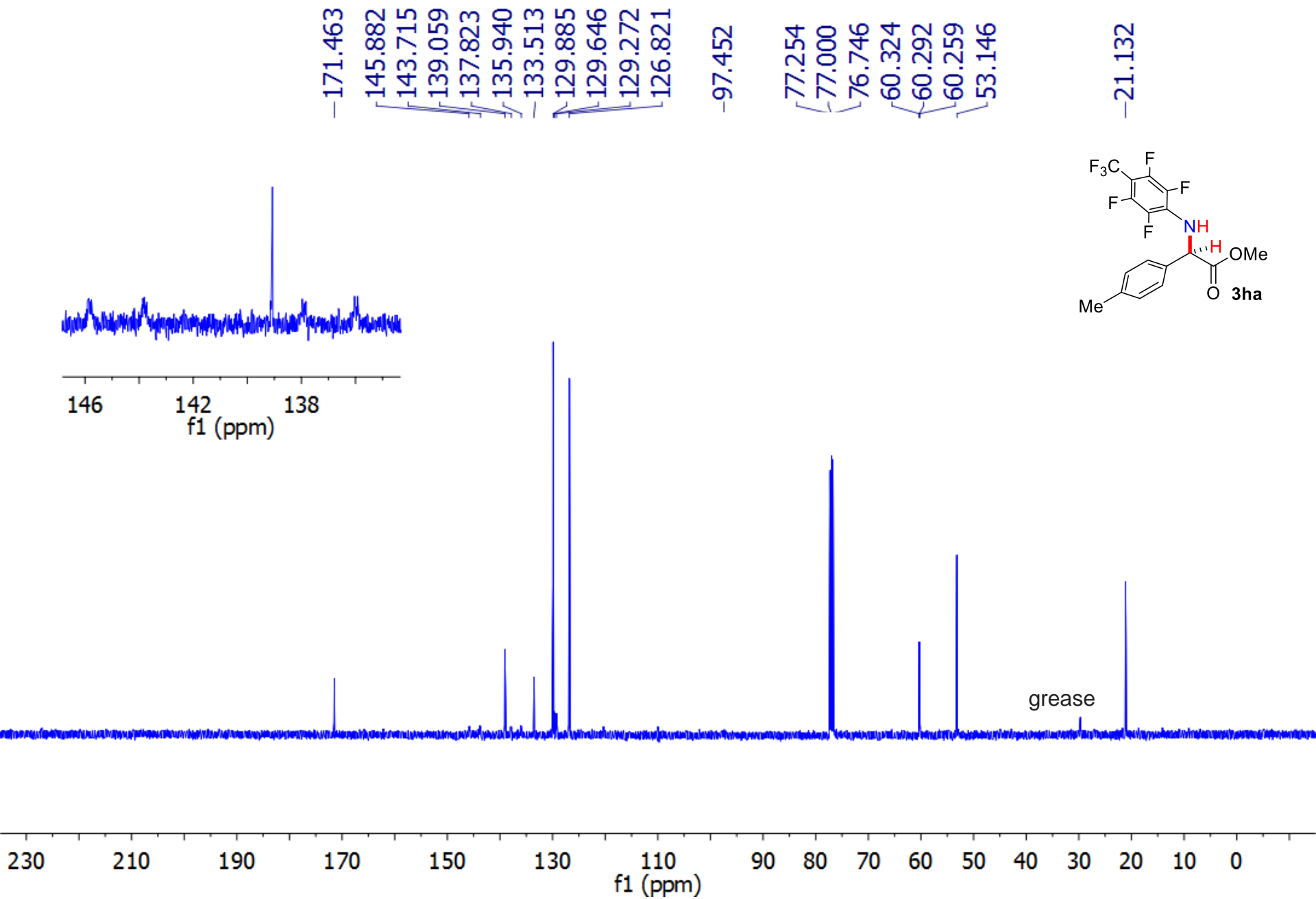
5.455
5.438

-3.755

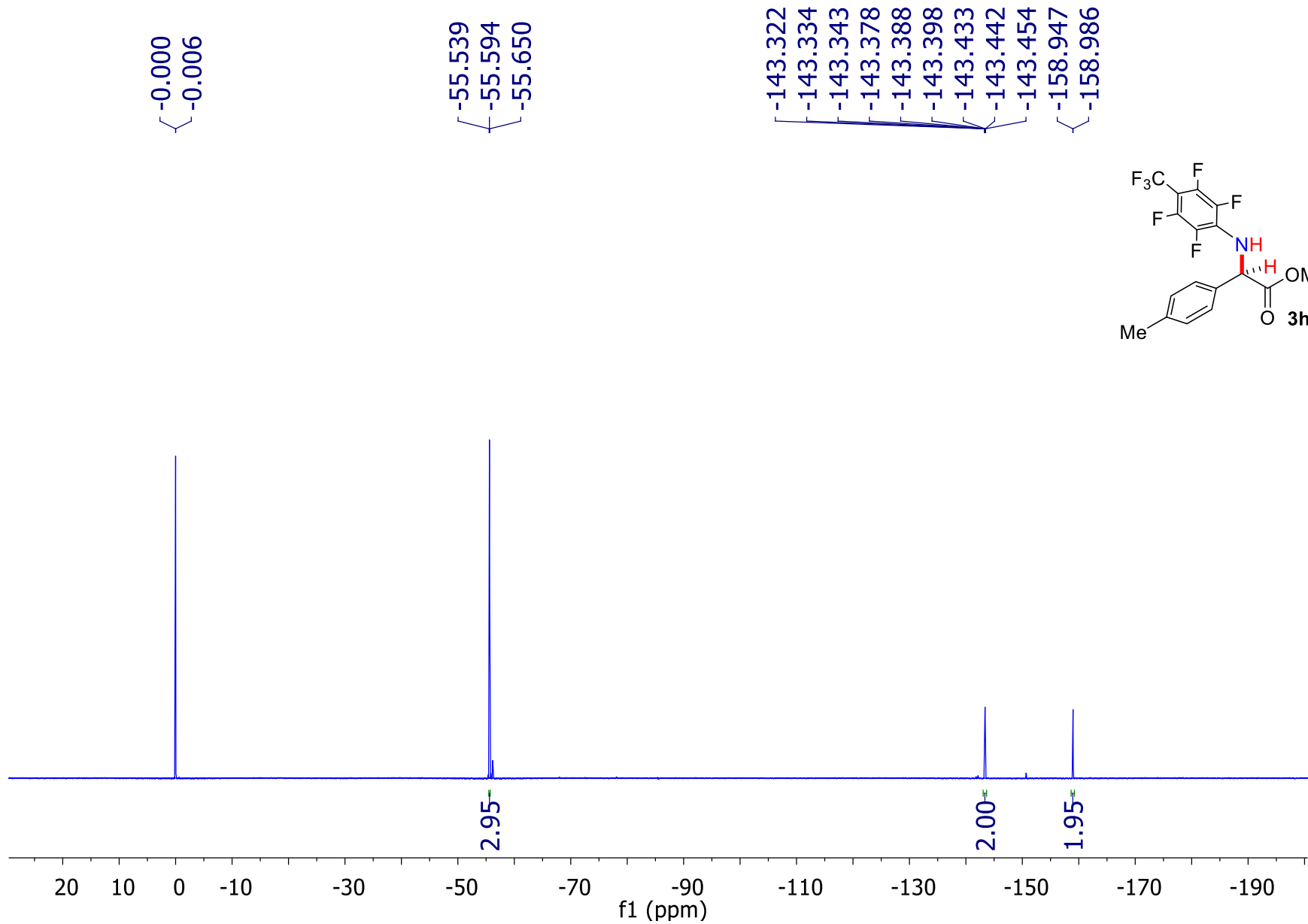
-2.341



^{13}C NMR

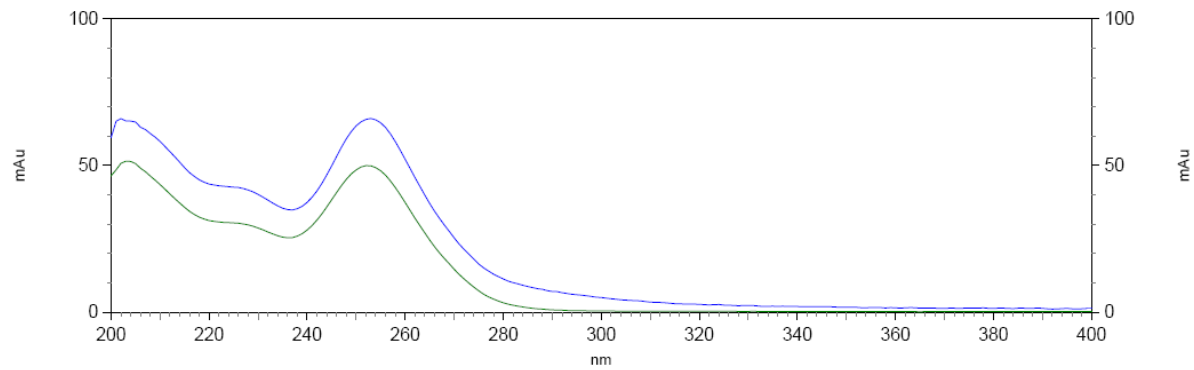
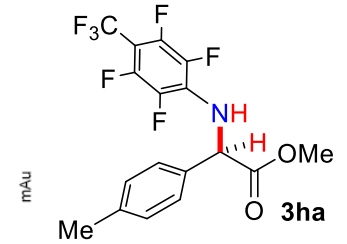
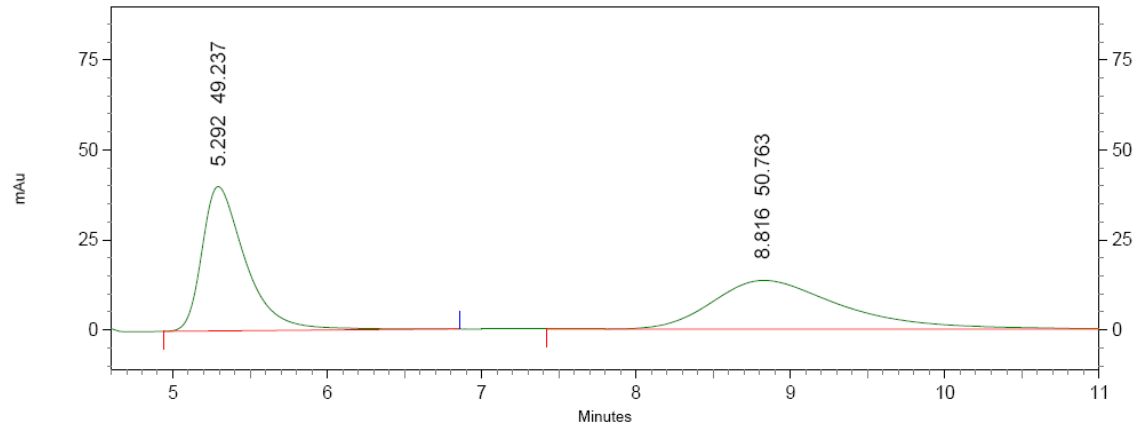


^{19}F NMR



HPLC

JLM-V-222-1a-OJH-1%1ML
C:\EZStart\Projects\Default\Data\JLM-V-222-1a-OJH-1%1ML
C:\EZStart\Projects\Default\Method\CQL-AD-H10%1.0ml60min.met
AD-H column 20%IPA @ 0.8ml/min

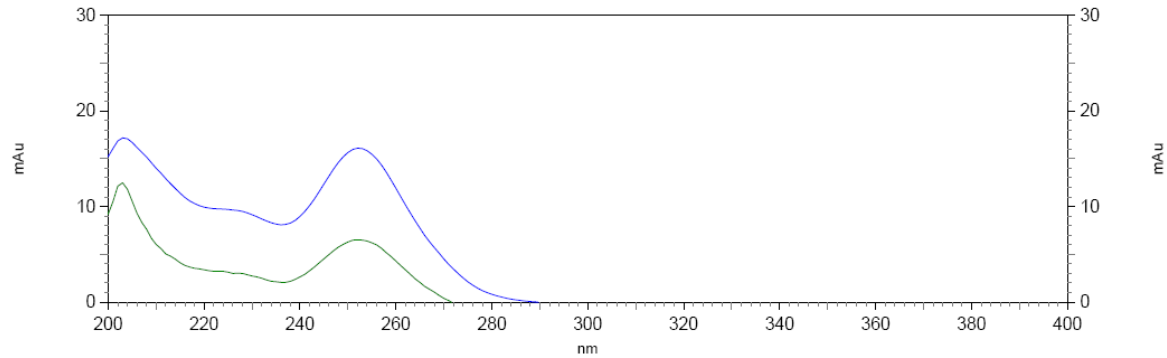
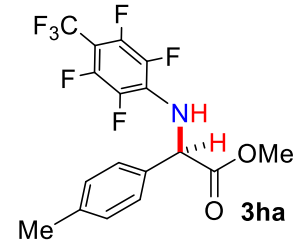
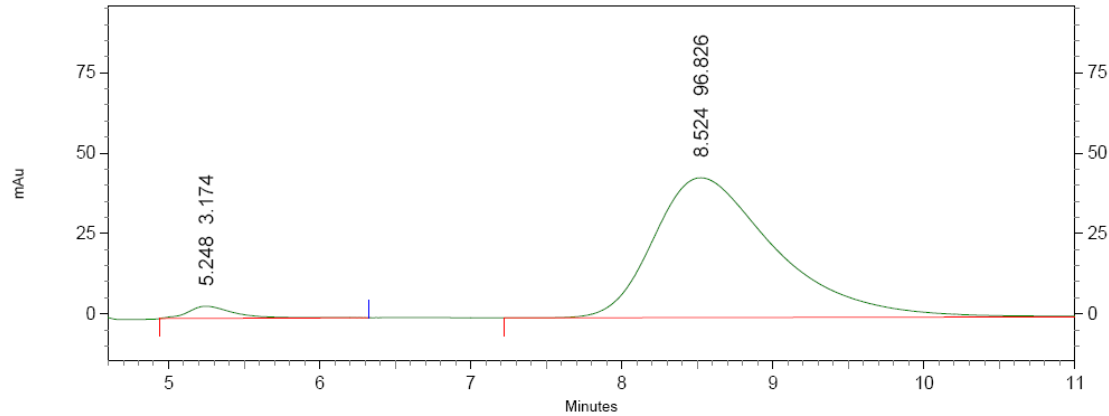


4: 259 nm, 4 nm Results

Pk #	Retention Time	Area Percent
1	5.292	49.237
2	8.816	50.763
Totals		100.000

HPLC

JLM-V-222-2-OJH-1%1ML
C:\EZStart\Projects\Default\Data\JLM-V-222-2-OJH-1%1ML
C:\EZStart\Projects\Default\Method\CQL-AD-H10%1.0ml60min.met
AD-H column 20%IPA @ 0.8ml/min

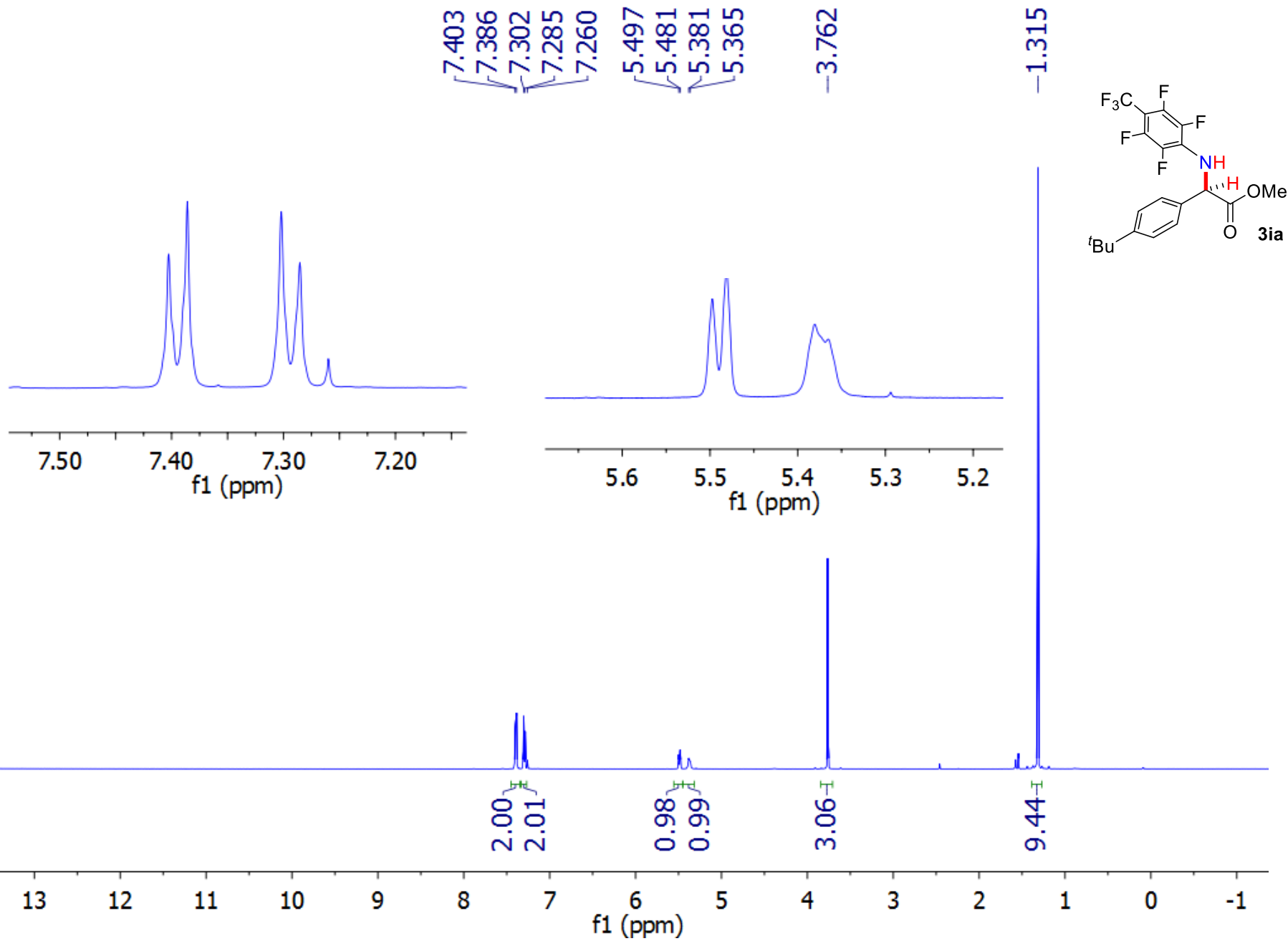


4: 259 nm, 4 nm Results

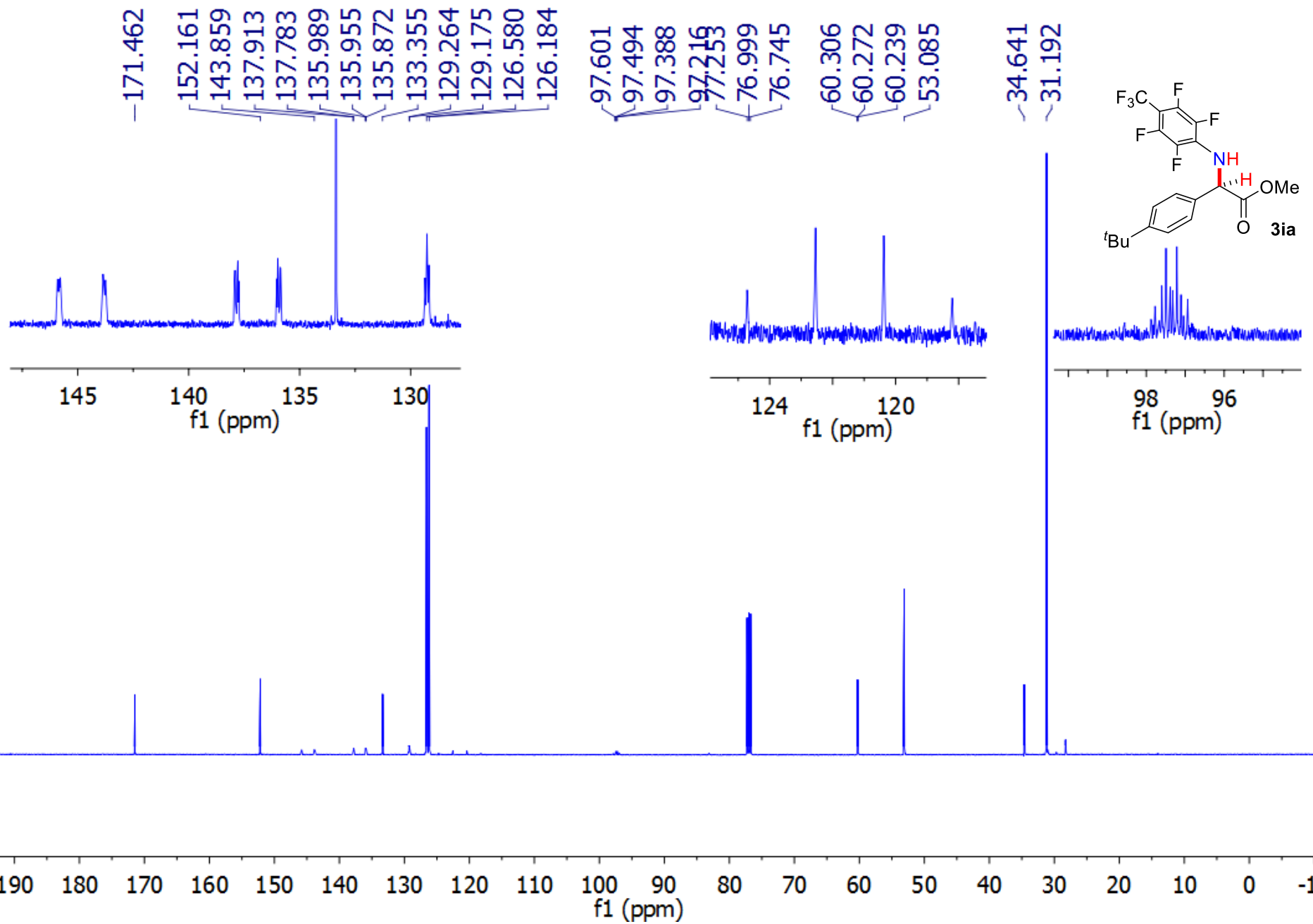
Pk #	Retention Time	Area Percent
1	5.248	3.174
2	8.524	96.826

Totals	100.000
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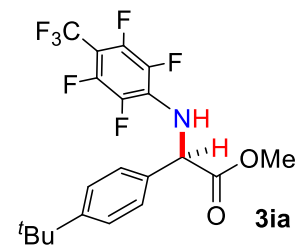
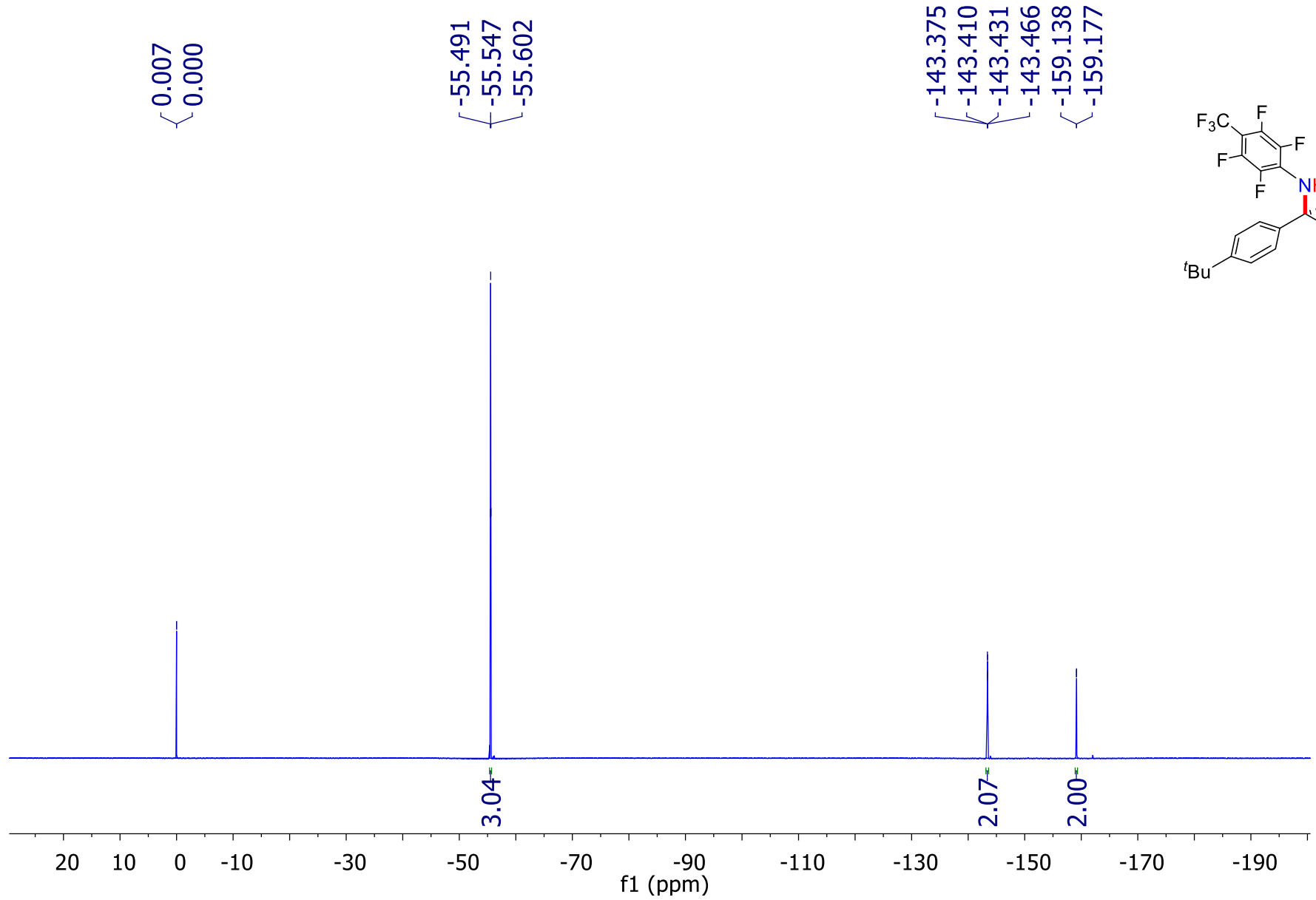
^1H NMR



^{13}C NMR



¹⁹F NMR

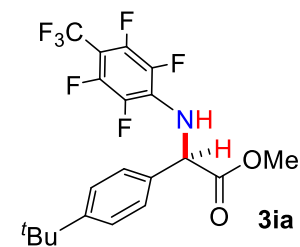
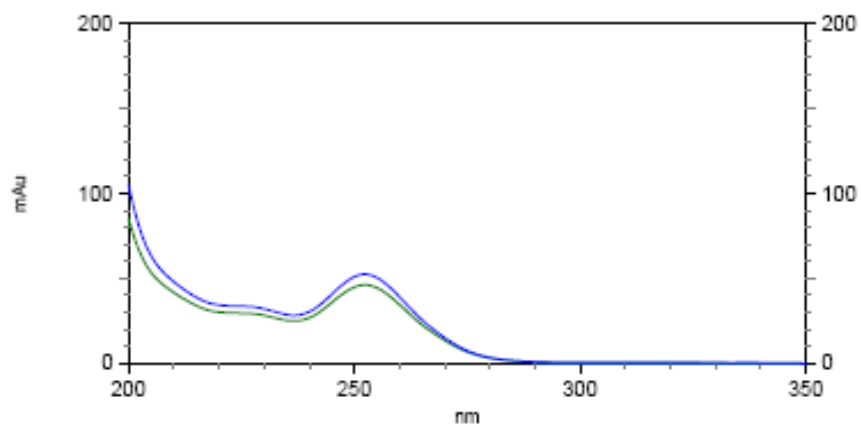
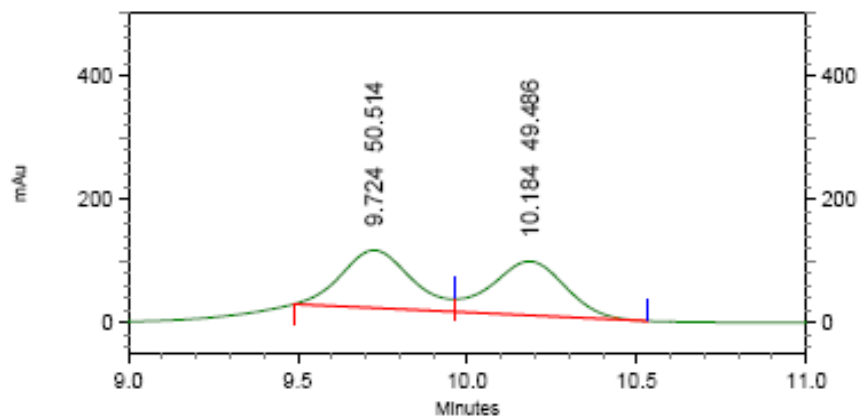


HPLC

JLM-V-194-1C-whehk-0%0.8ML

C:\EZStart\Projects\Default\Method\LK-20%-0.8-90min.met

C:\EZStart\Projects\Default\Data\JLM-V-194-1C-whehk-0%0.8ML



2: 252 nm, 4 nm

Results

Name	Retention Time	Area Percent	Pk #
	9.724	50.514	1
	10.184	49.486	2

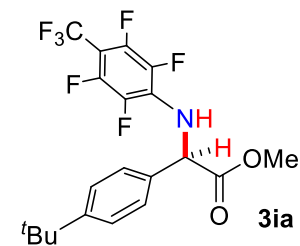
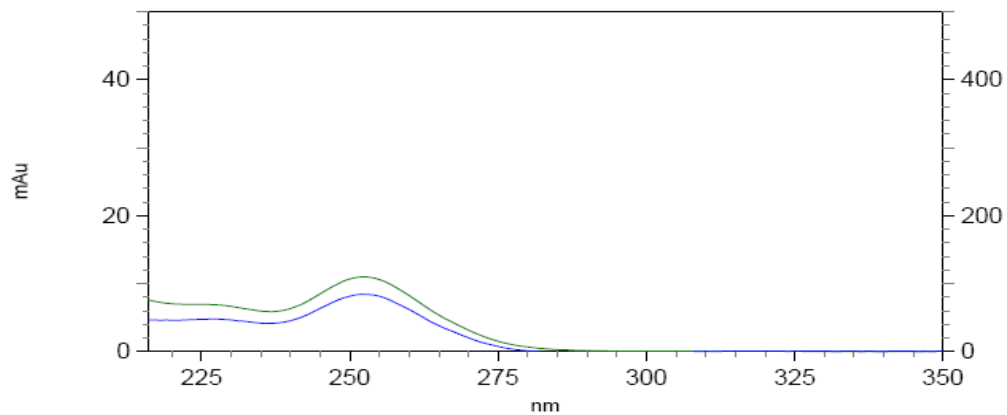
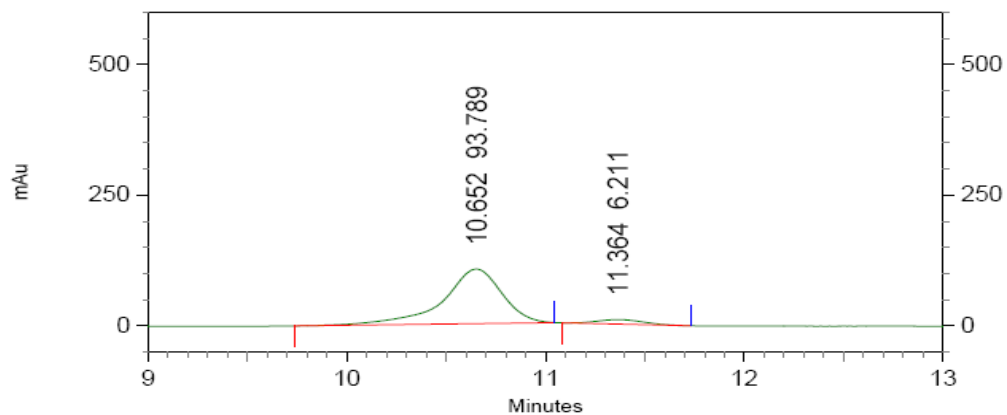
Totals		100.000	
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HPLC

JLM-V-194-2-whelk-0%0.8ML

C:\EZStart\Projects\Default\Method\LK-20%-0.8-90min.met

C:\EZStart\Projects\Default\Data\JLM-V-194-2-whelk-0%0.8ML



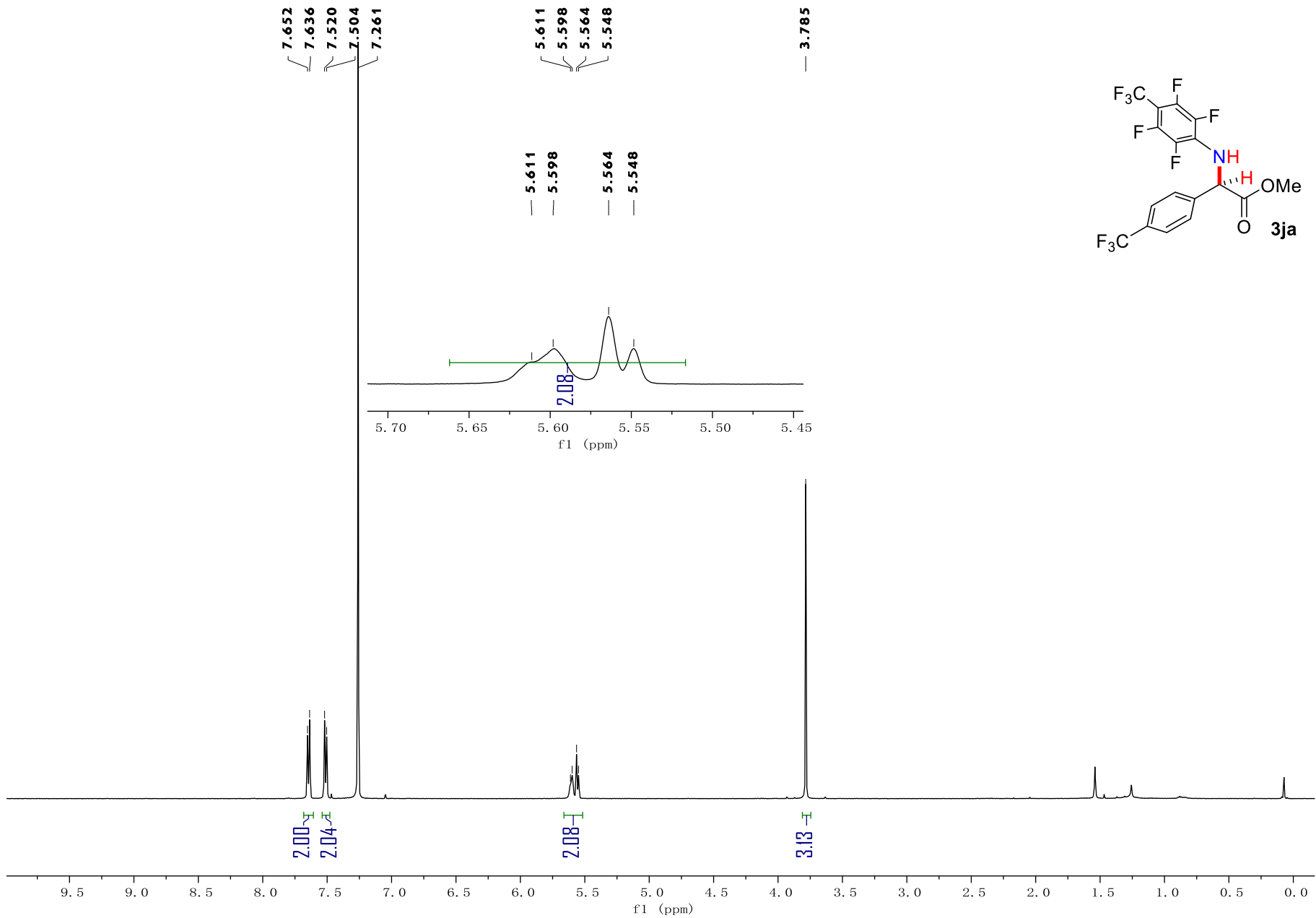
2: 253 nm, 4 nm

Results

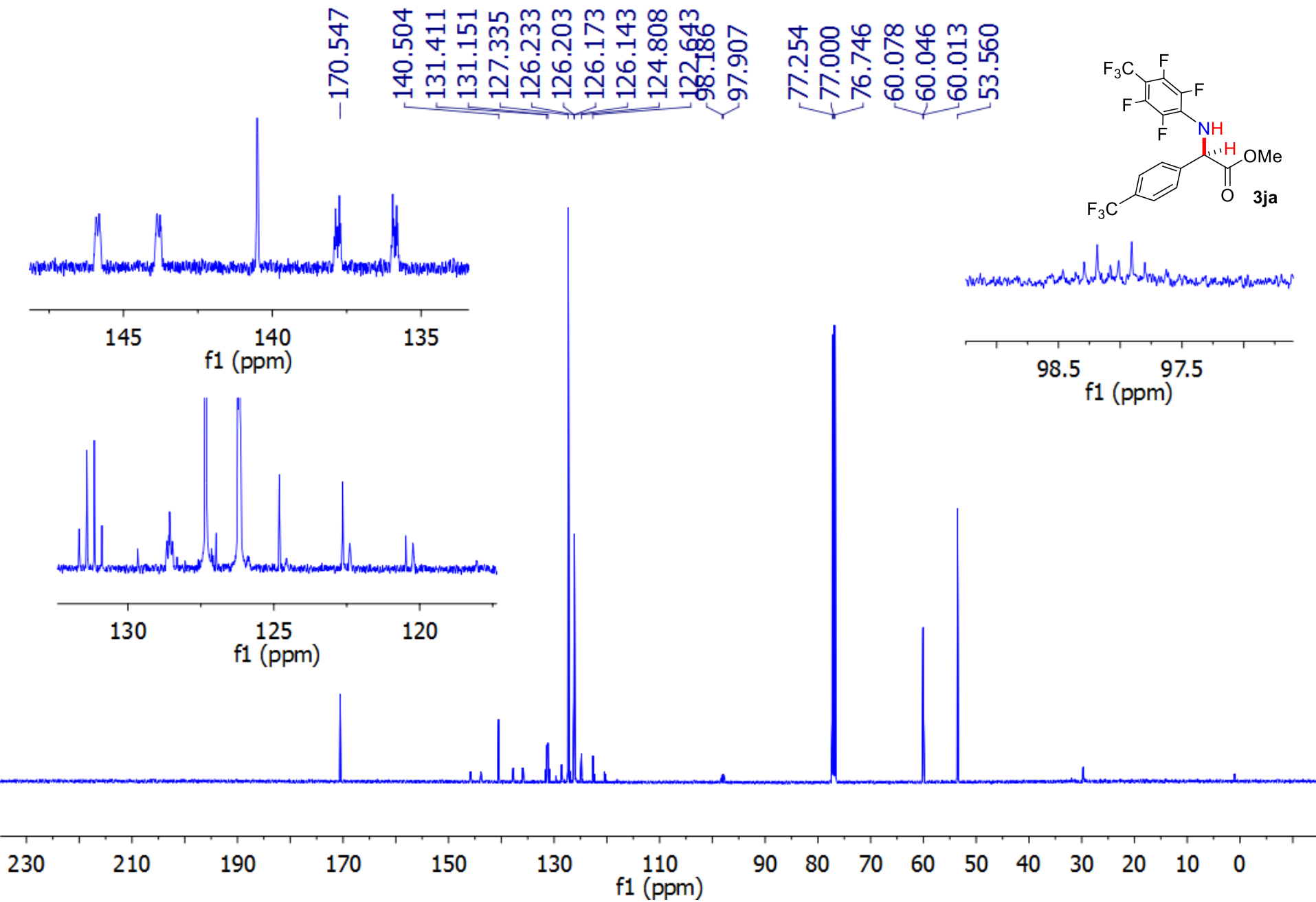
Name	Retention Time	Area Percent	Pk #
	10.652	93.789	1
	11.364	6.211	2

Totals		100.000	
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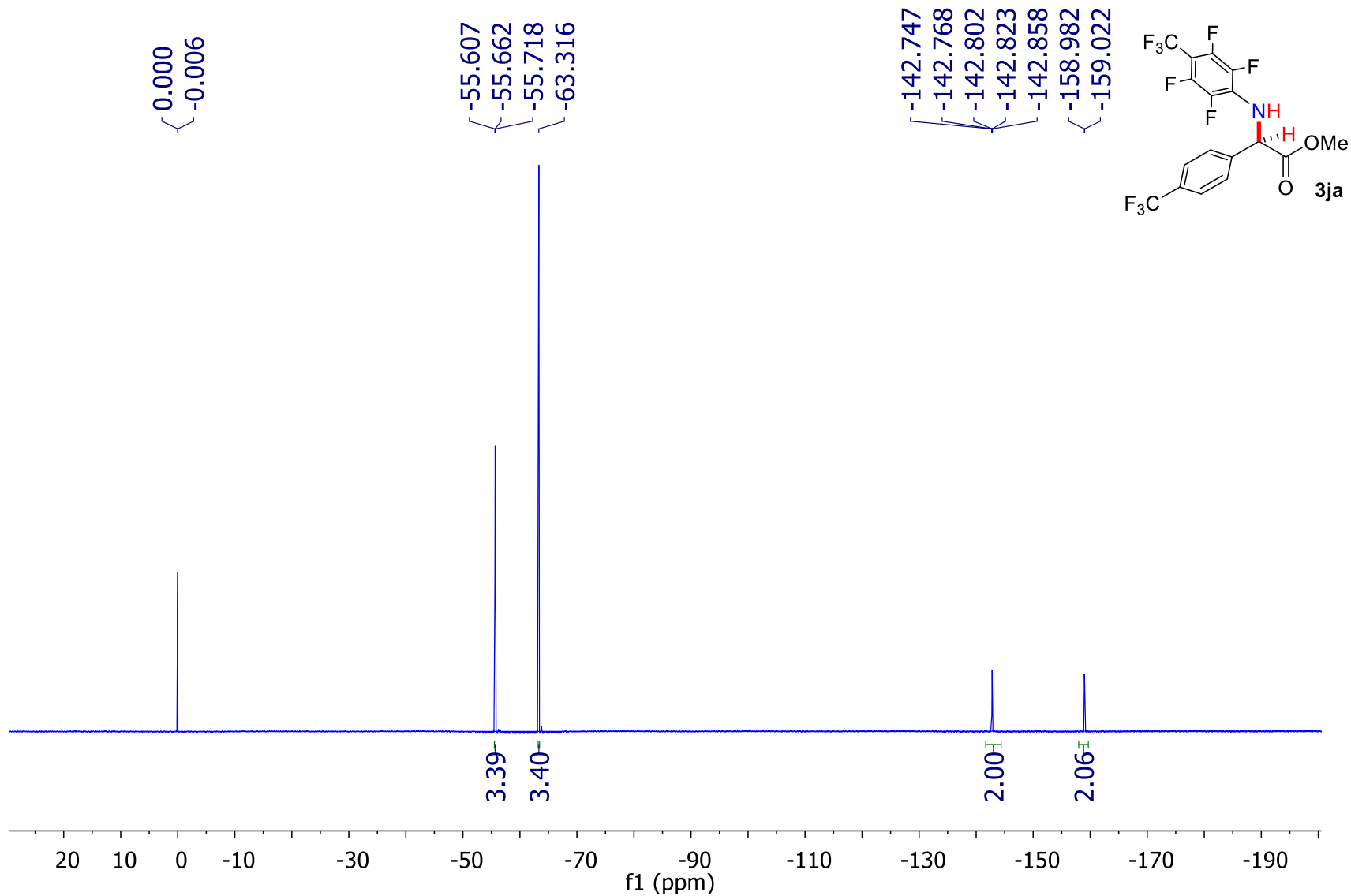
^1H NMR



^{13}C NMR



^{19}F NMR

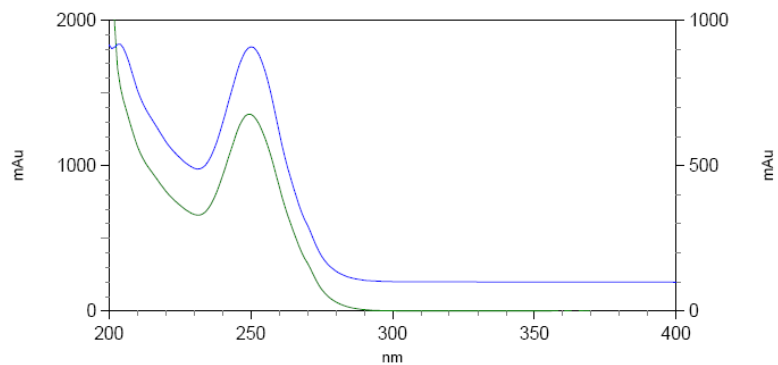
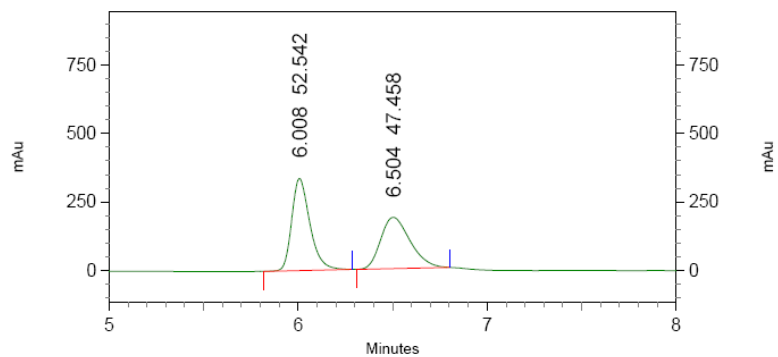


HPLC

JLM-V-195-1-ADH-1%1ML

C:\EZStart\Projects\Default\Method\JTL-3%-ADH1ml.met

C:\EZStart\Projects\Default\Data\JLM-V-195-1-ADH1%1ML

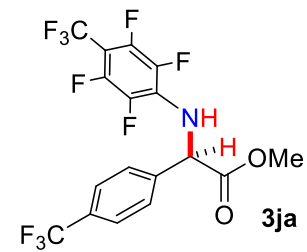


3: 220 nm, 4 nm

Results

Name	Retention Time	Area Percent	Pk #
	6.008	52.542	1
	6.504	47.458	2

Totals		100.000	
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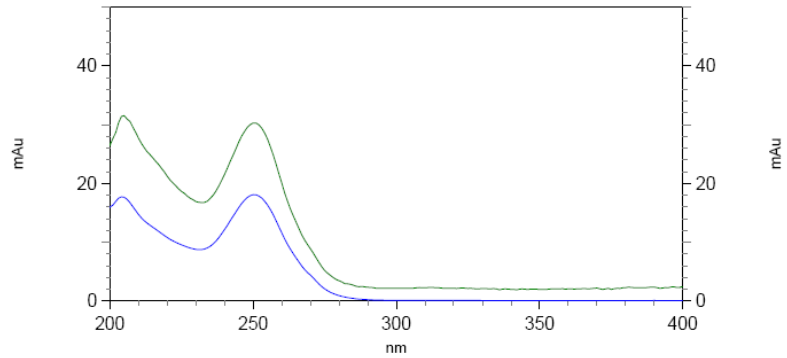
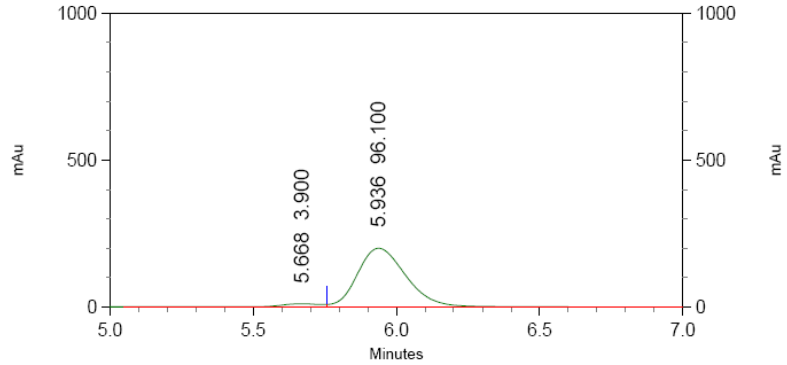


HPLC

JLM-V-204-1A-ADH-1%1ML

C:\EZStart\Projects\Default\Method\shifatest_2,5-dimehoxy.met

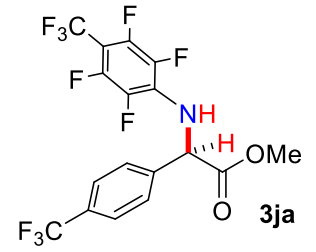
C:\EZStart\Projects\Default\Data\JLM-V-204-1A-ADH1%1ML



7: 251 nm, 4 nm

Results

Pk #	Name	Retention Time	Area Percent
1		5.668	3.900
2		5.936	96.100
Totals			100.000

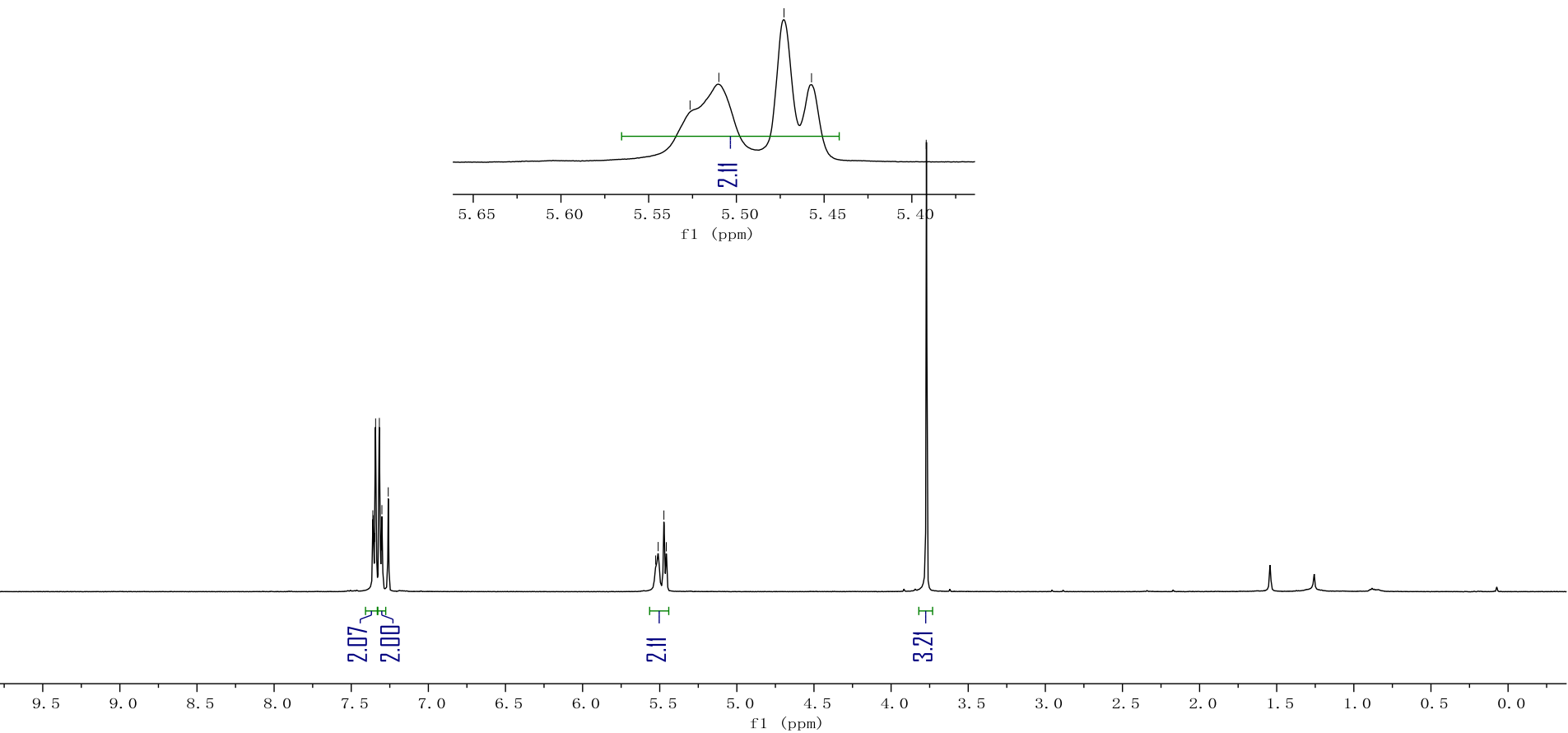
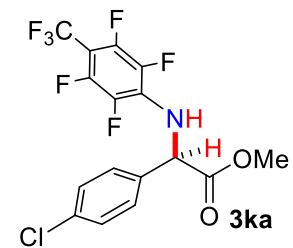


¹H NMR

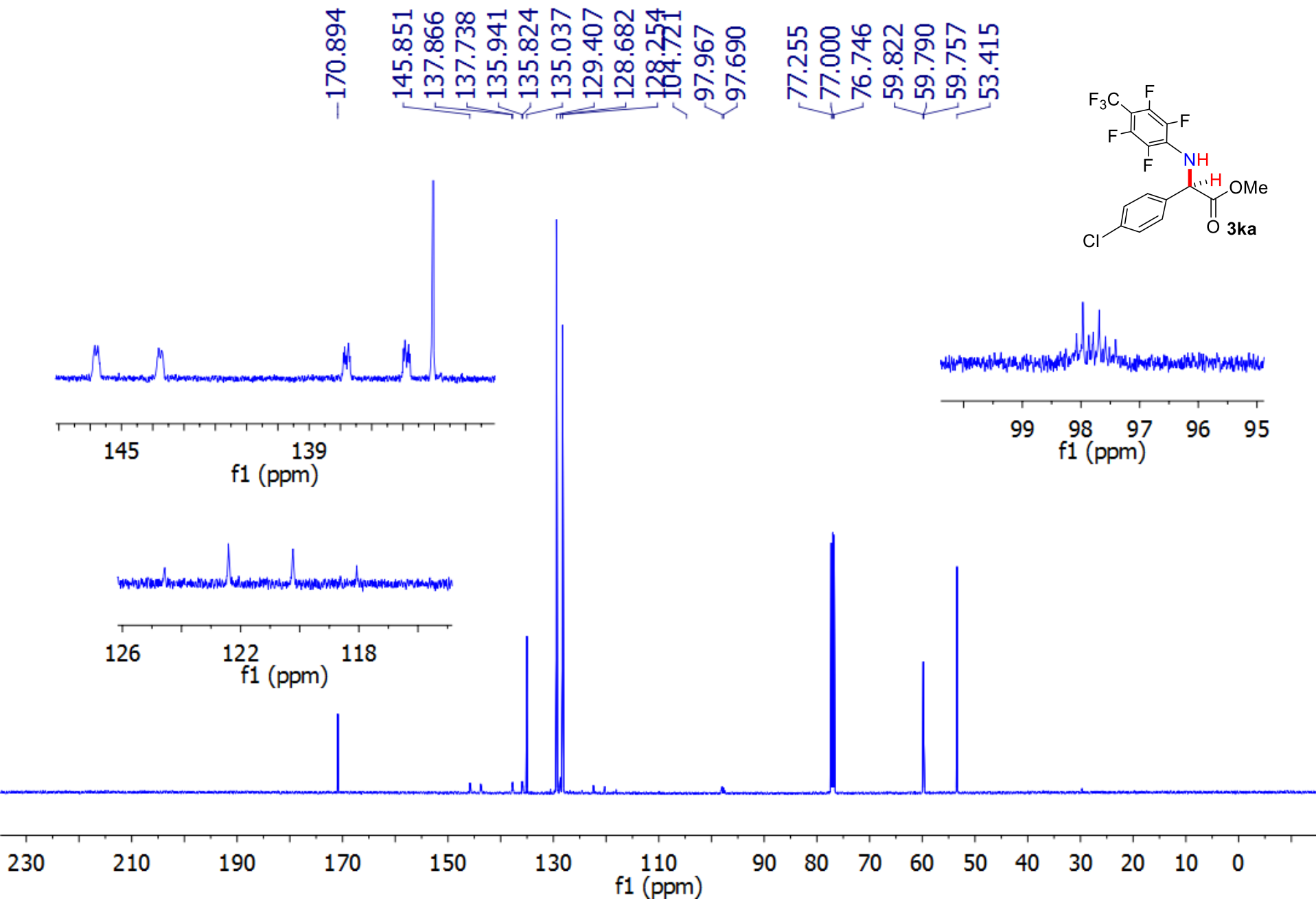
7.360
7.357
7.348
7.342
7.318
7.305
7.301
7.260

5.526
5.510
5.473
5.457

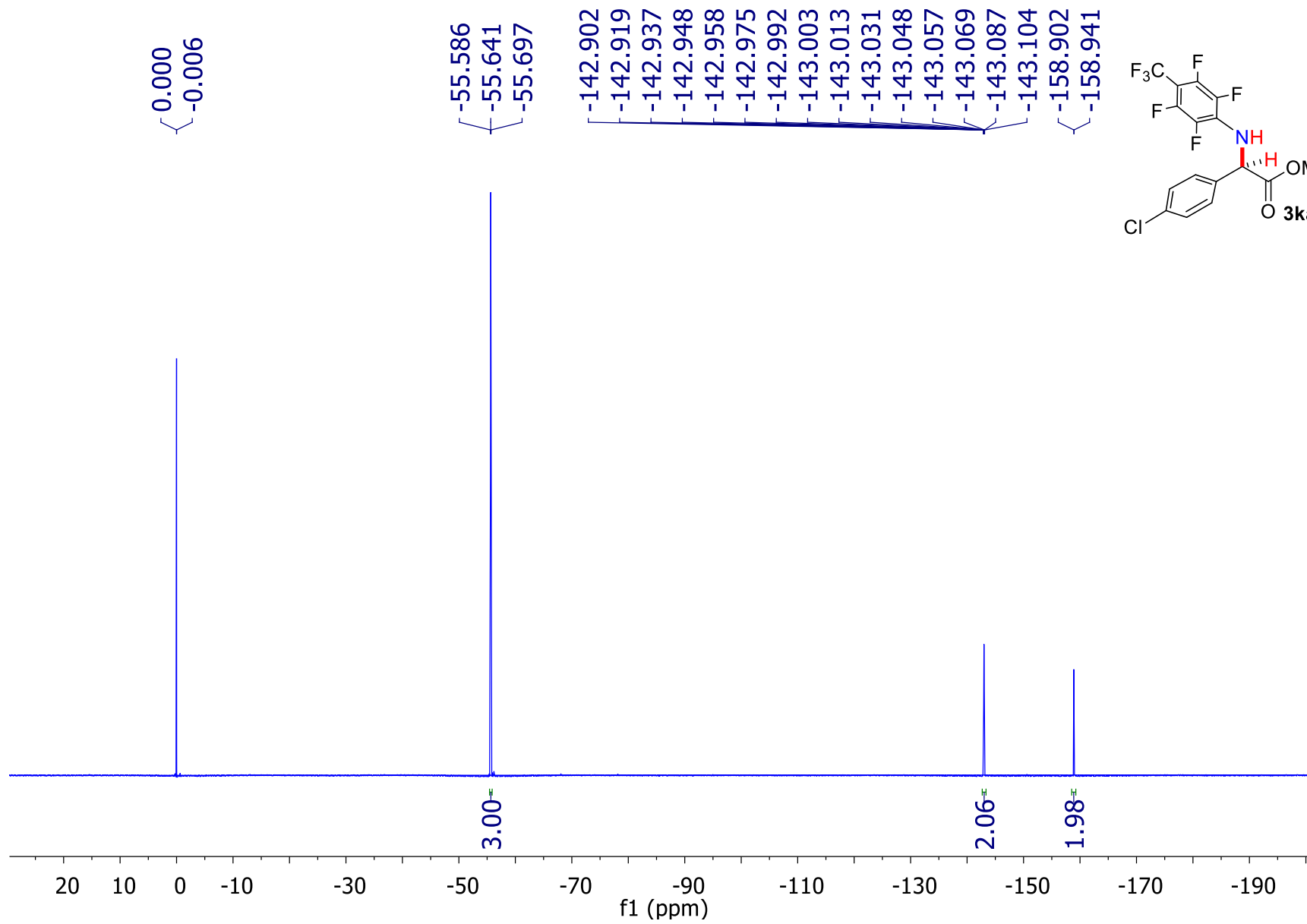
3.772



^{13}C NMR

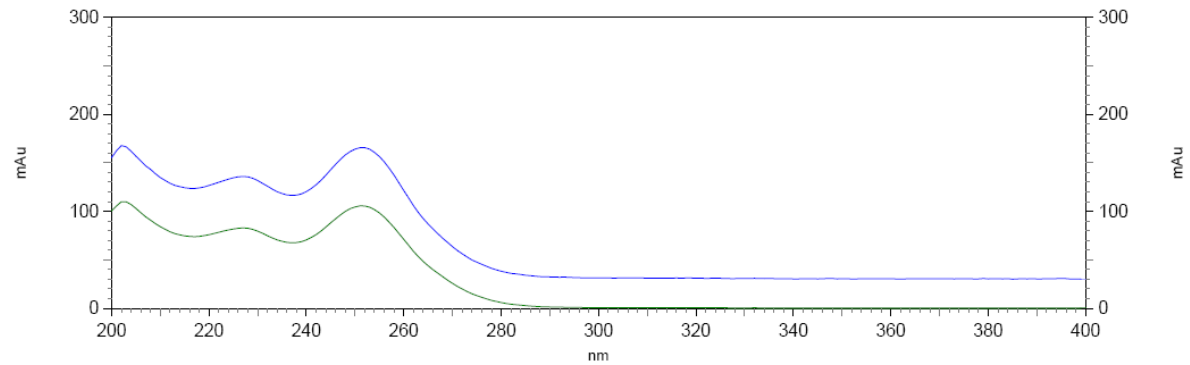
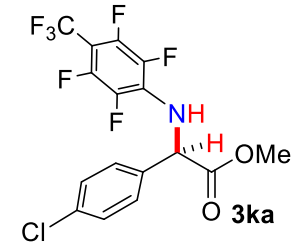
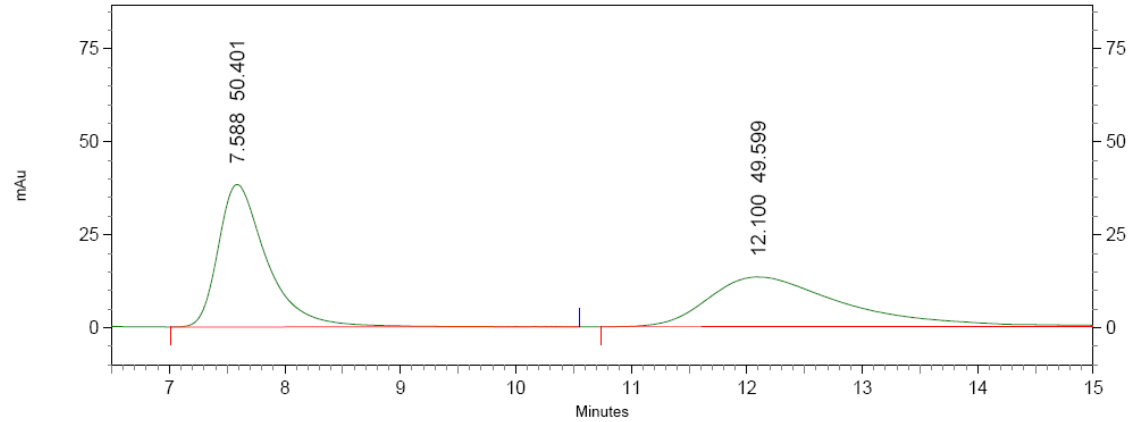


¹⁹F NMR



HPLC

JLM-V-223-1-OJH-1%1ML
 C:\EZStart\Projects\Default\Data\JLM-V-223-1-OJH-1%1ML
 C:\EZStart\Projects\Default\Method\CQL-AD-H10%1.0ml60min.met
 AD-H column 20%IPA @ 0.8ml/min



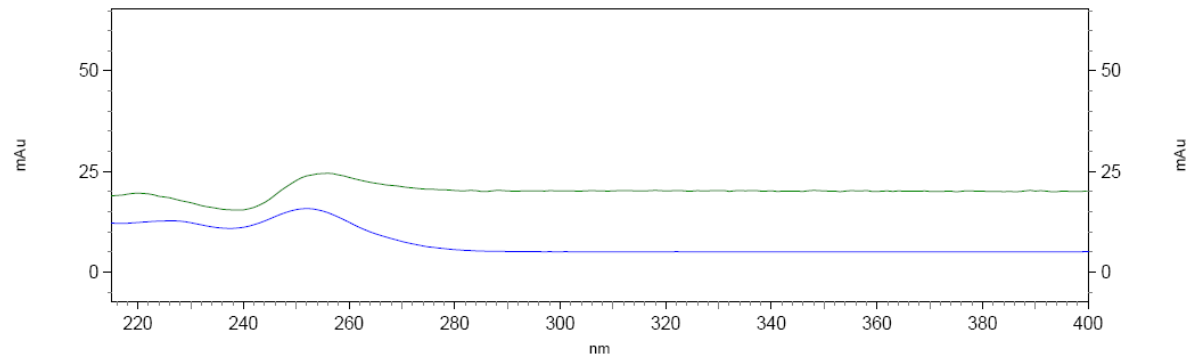
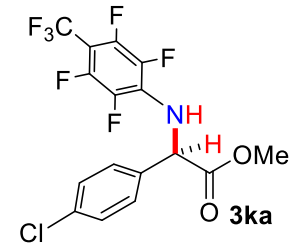
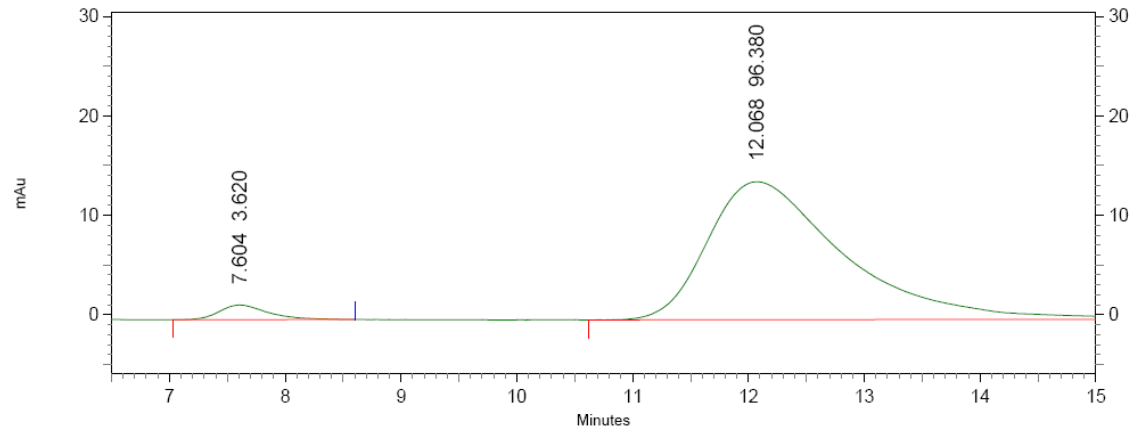
4: 259 nm, 4 nm Results

Pk #	Retention Time	Area Percent
1	7.588	50.401
2	12.100	49.599

Totals		100.000
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HPLC

JLM-V-223-2-OJH-1%1ML
C:\EZStart\Projects\Default\Data\JLM-V-223-2-OJH-1%1ML
C:\EZStart\Projects\Default\Method\CQL-AD-H10%1.0ml60min.met
AD-H column 20%IPA @ 0.8ml/min



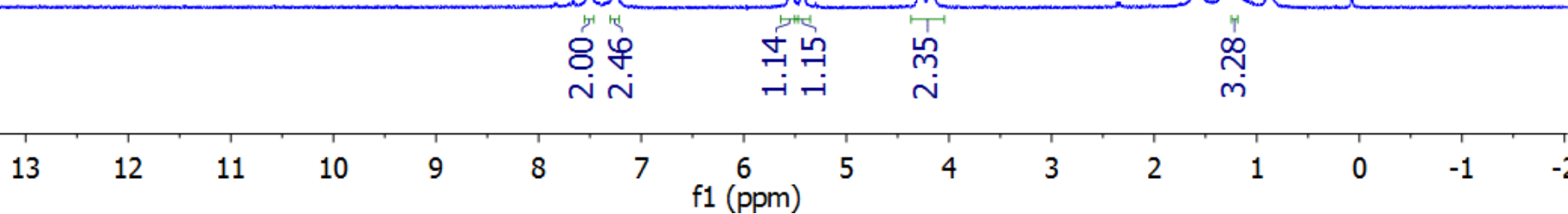
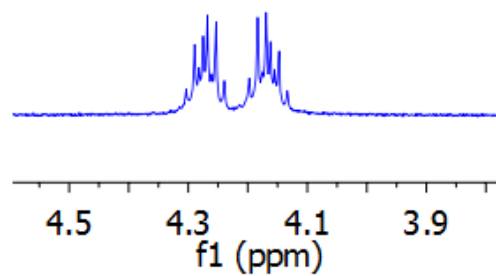
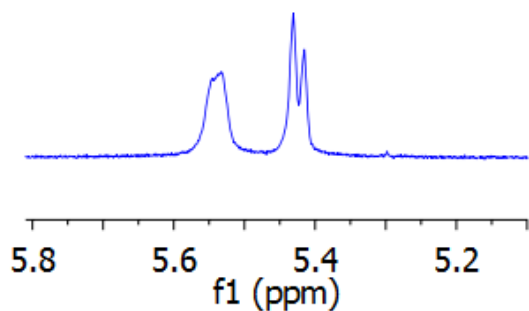
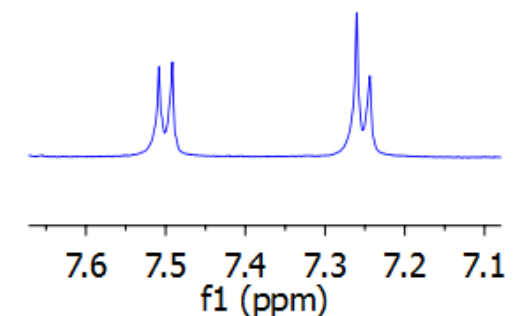
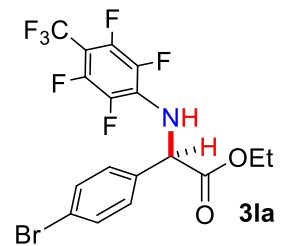
4: 259 nm, 4 nm Results

Pk #	Retention Time	Area Percent
1	7.604	3.620
2	12.068	96.380

Totals	100.000
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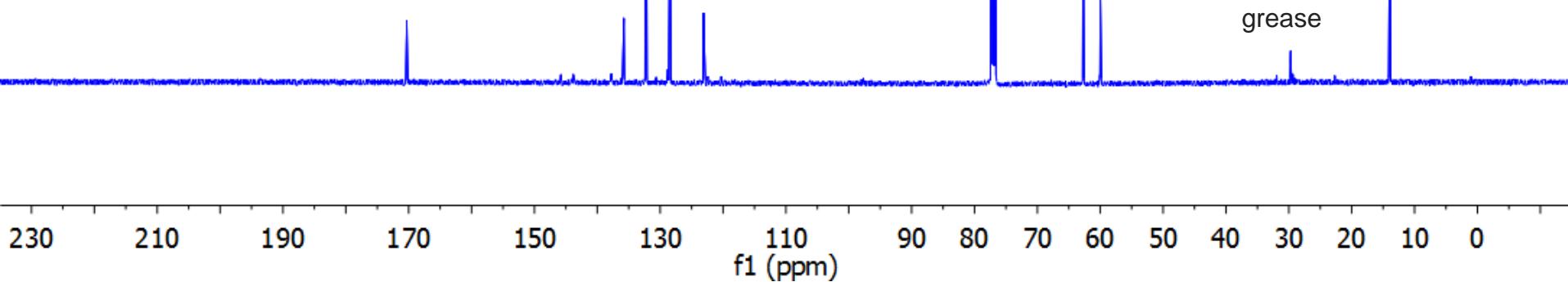
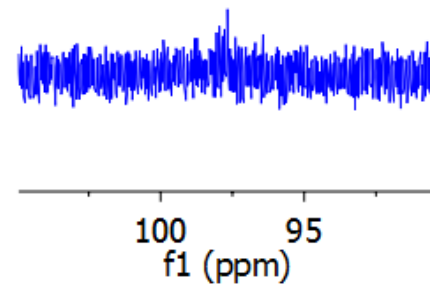
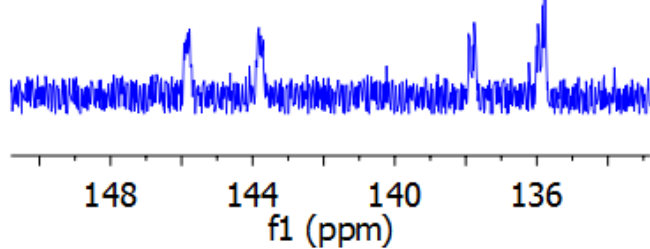
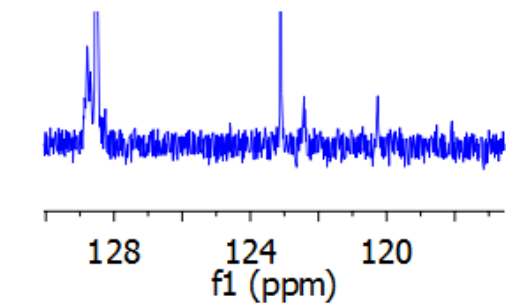
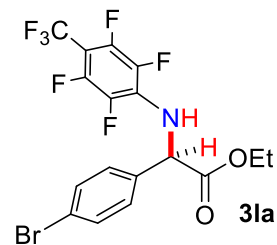
¹H NMR

7.509
7.492
7.260
7.244
5.533
5.431
5.415
4.289
4.275
4.268
4.253
4.183
4.169
4.162
4.147
1.236
1.222
1.208

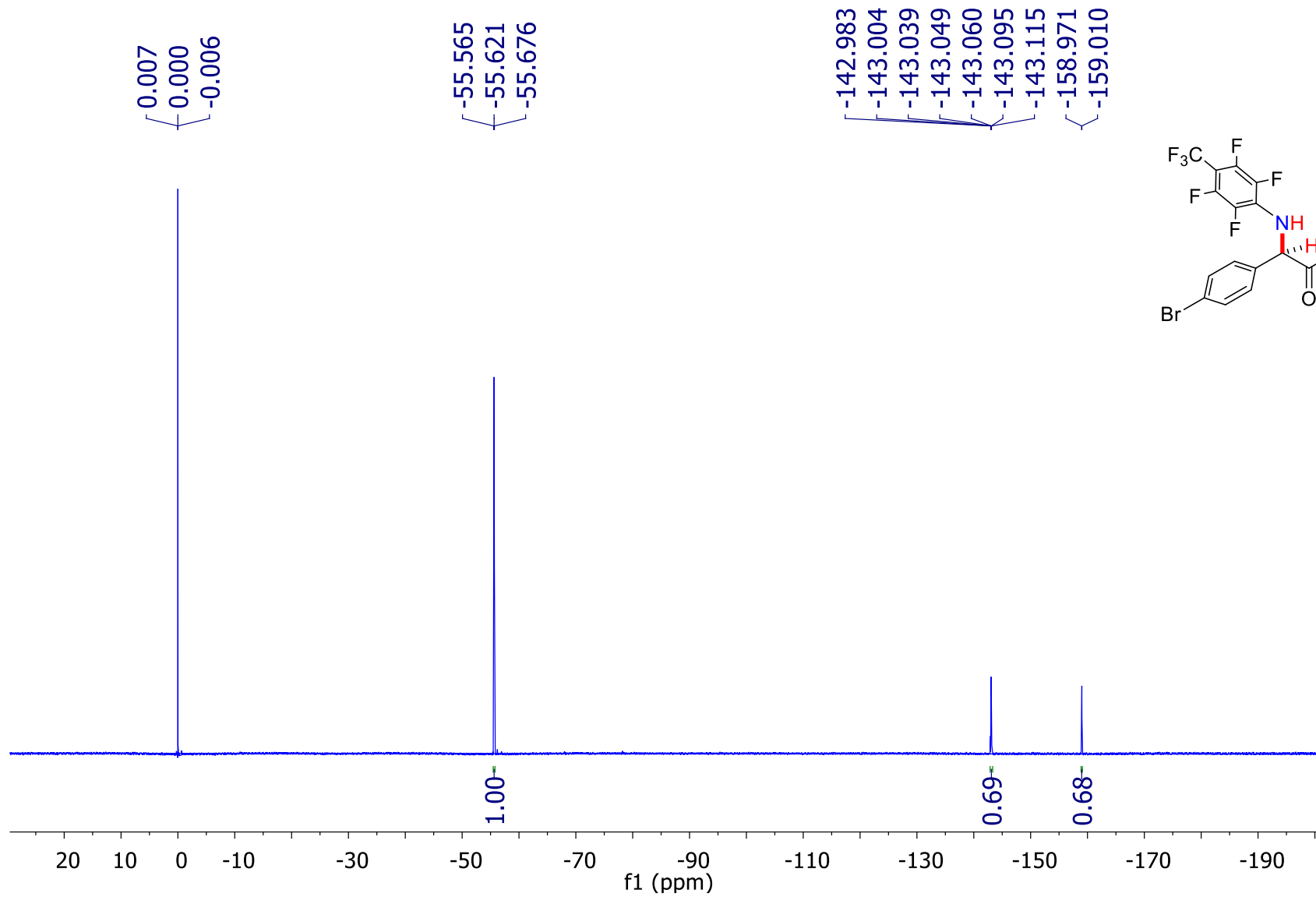


^{13}C NMR

170.299
145.775
145.614
143.824
139.602
137.764
135.780
132.313
128.514
123.109
122.456
120.251
97.673
77.254
77.000
76.746
62.719
60.011
59.979
59.947
13.924

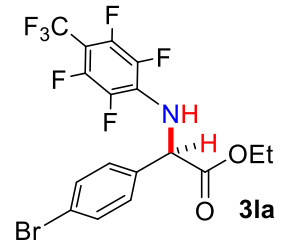
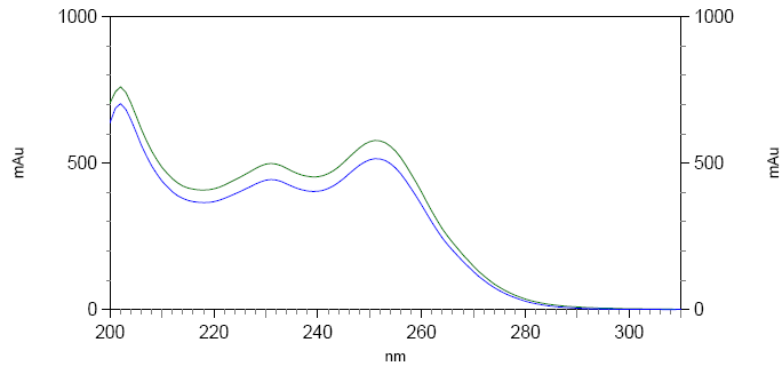
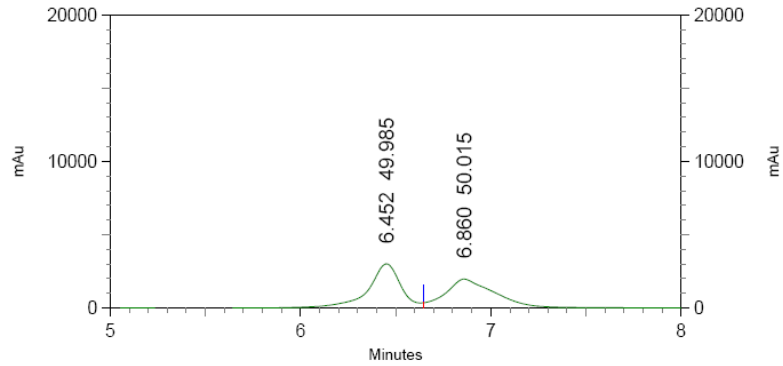


^{19}F NMR



HPLC

JLM-III-1-1-Whelk-0.5@-1mL
C:\EZStart\Projects\Default\Method\XC-5%-ADH1ml.met
E:\JLM-III-1-1-Whelk-0.5 @1ml



4: 254 nm, 4 nm

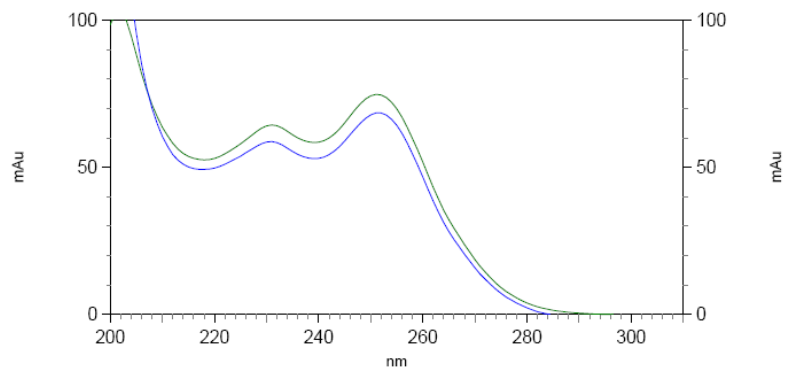
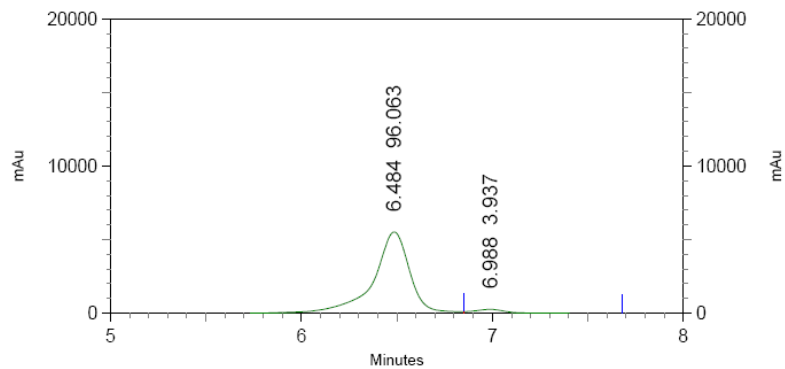
Results

Name	Retention Time	Area Percent	Pk #
	6.452	49.985	1
	6.860	50.015	2

Totals		100.000	
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HPLC

JLM-V-137-1-WHELK-0.5@-1mL
C:\EZStart\Projects\Default\Method\XC-5%-ADH1ml.met
E:\JLM-V-137-1-WHELK0.5@1ml

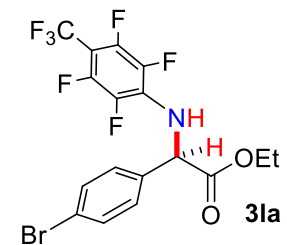


4: 254 nm, 4 nm

Results

Name	Retention Time	Area Percent	Pk #
	6.484	96.063	1
	6.988	3.937	2

Totals	100.000	
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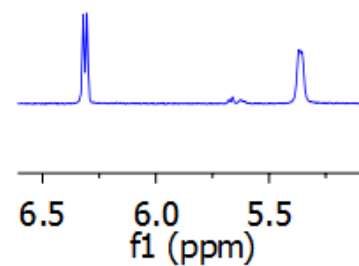
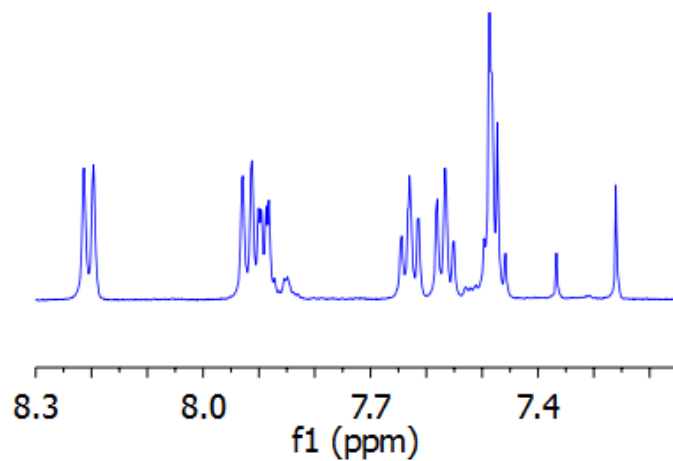
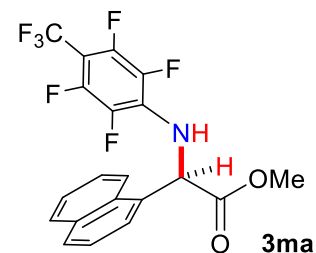


¹H NMR

8.214
8.197
7.929
7.913
7.901
7.887
7.882
7.630
7.580
7.566
7.487
7.482
7.473
7.260
6.320
6.304

5.369
5.355

-3.751



1.05
2.24
2.03
2.17

1.04

1.02

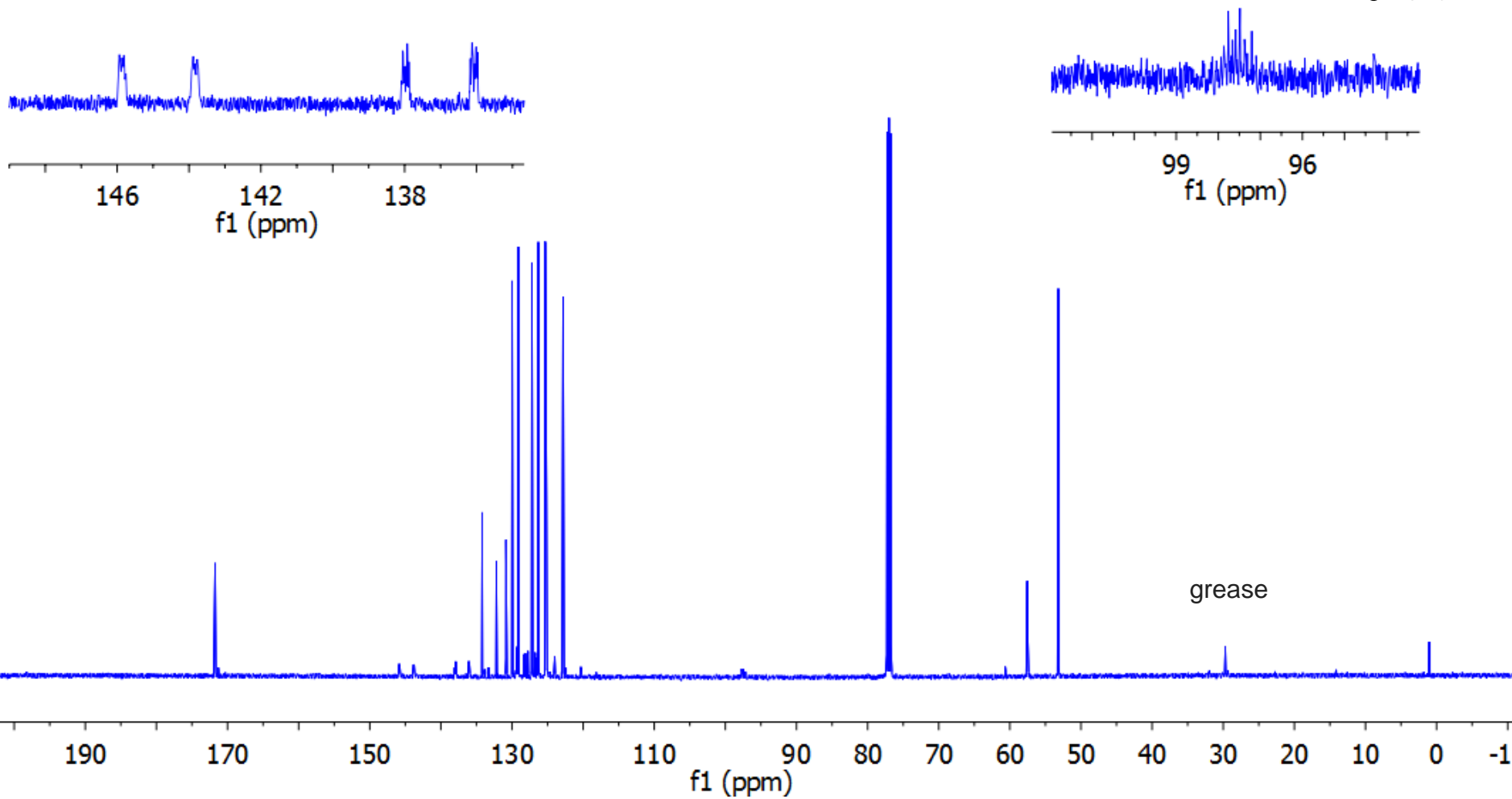
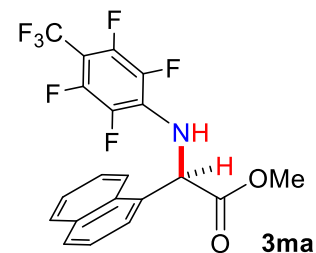
3.33

10.0 9.0 8.0 7.0 6.0 5.0 4.0 3.0 2.0 1.0 0.0 -1.0

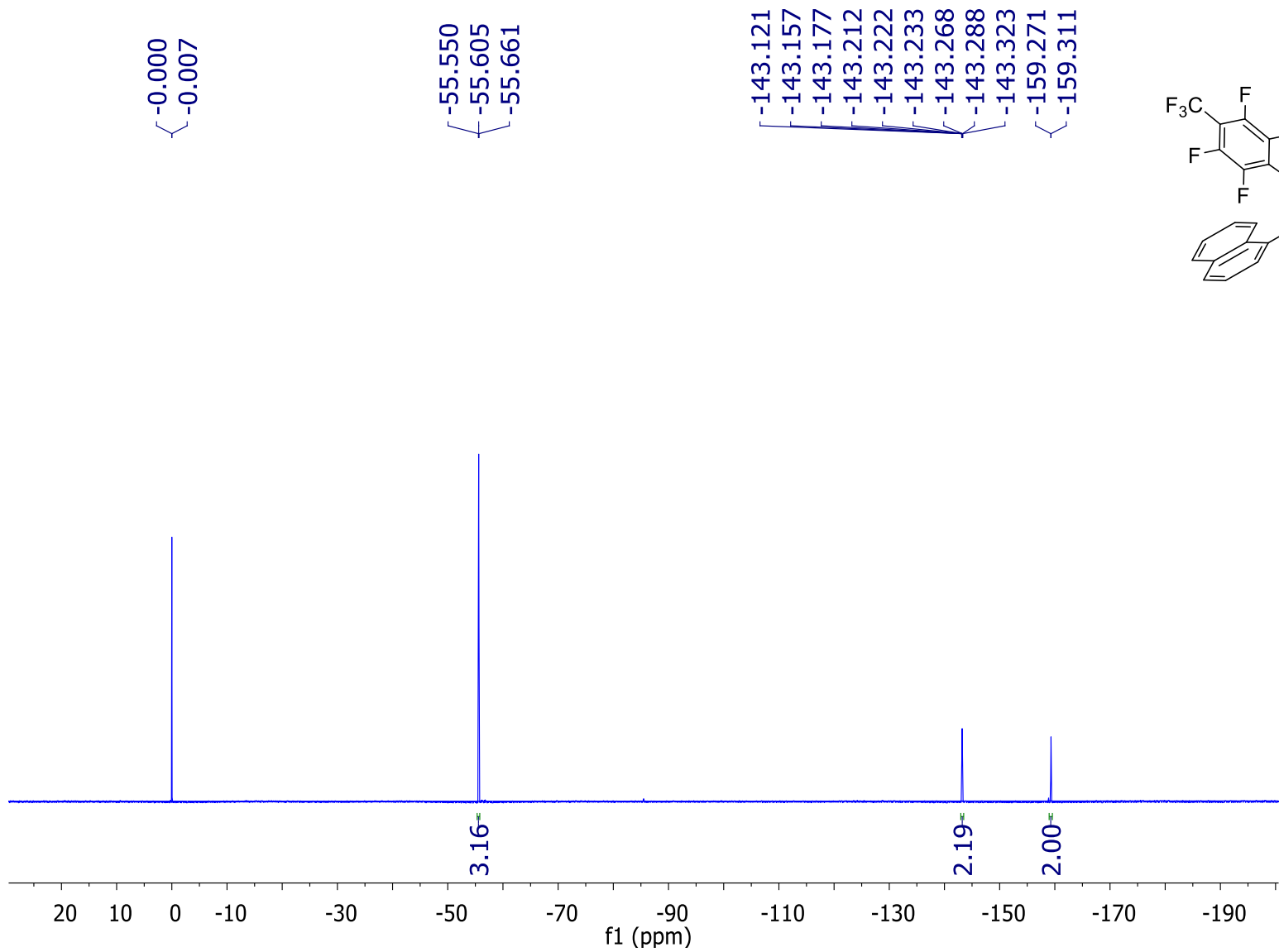
f1 (ppm)

¹³C NMR

171.749
145.919
136.107
134.191
132.201
130.839
129.983
129.119
127.203
126.301
125.333
125.146
122.817
120.317
97.462
77.254
77.000
76.746
57.557
53.187

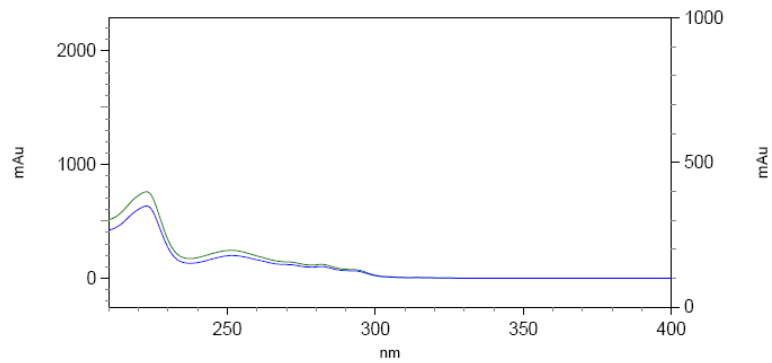
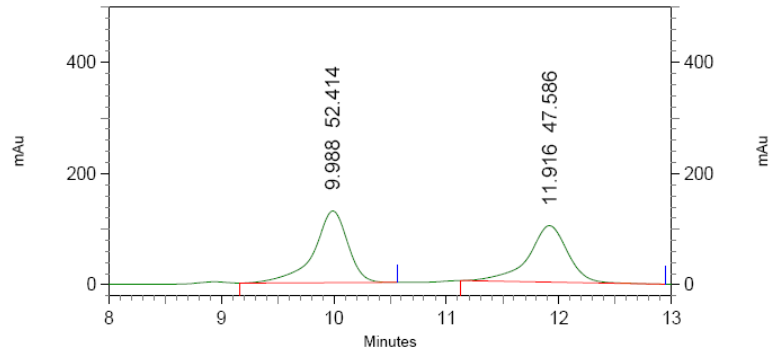
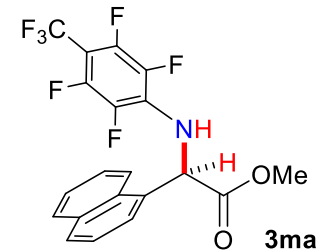


^{19}F NMR



HPLC

JLM-II-218-1-WHELK-0.5@1ML
C:\EZStart\Projects\Default\Method\JLM-ODH-0%-0.7ml.met
C:\Documents and Settings\zhang\Desktop\Jin-Lim\HPLC-data\JLM-II-218-1-WHELK-0.5@1ML.dat



3: 249 nm, 4 nm
Results

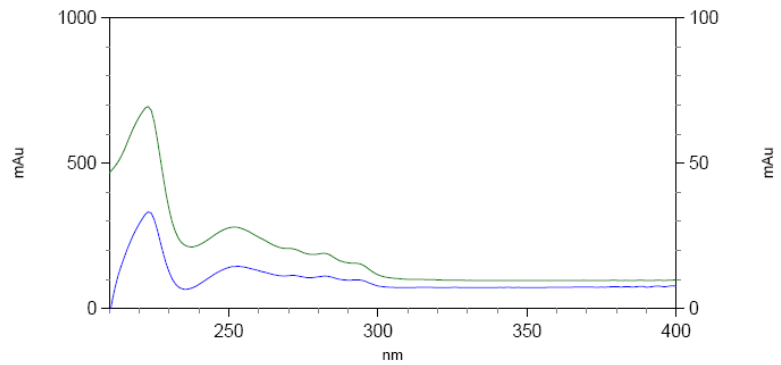
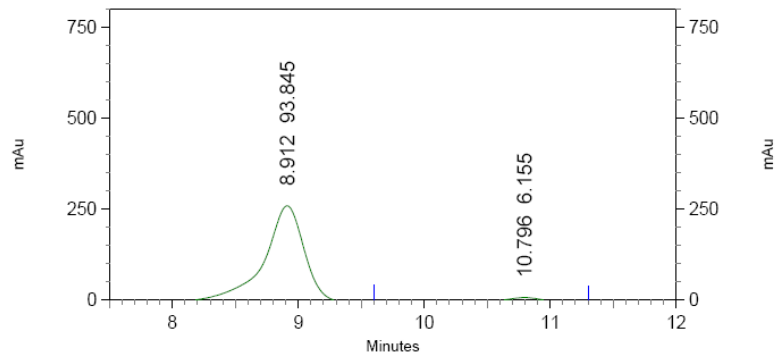
Pk #	Name	Retention Time	Area Percent
1		9.988	52.414
2		11.916	47.586
Totals			100.000

HPLC

JLM-V-167-2-WHELK-0.5@-1mL

C:\EZStart\Projects\Default\Method\Joey-ODH-20%-0.8mL.met

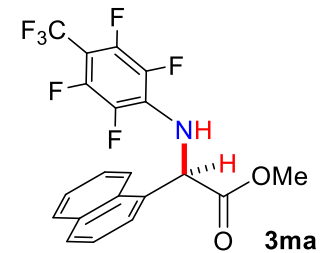
C:\EZStart\Projects\Default\Data\JLM-V-167-2-WHELK0.5@1ml



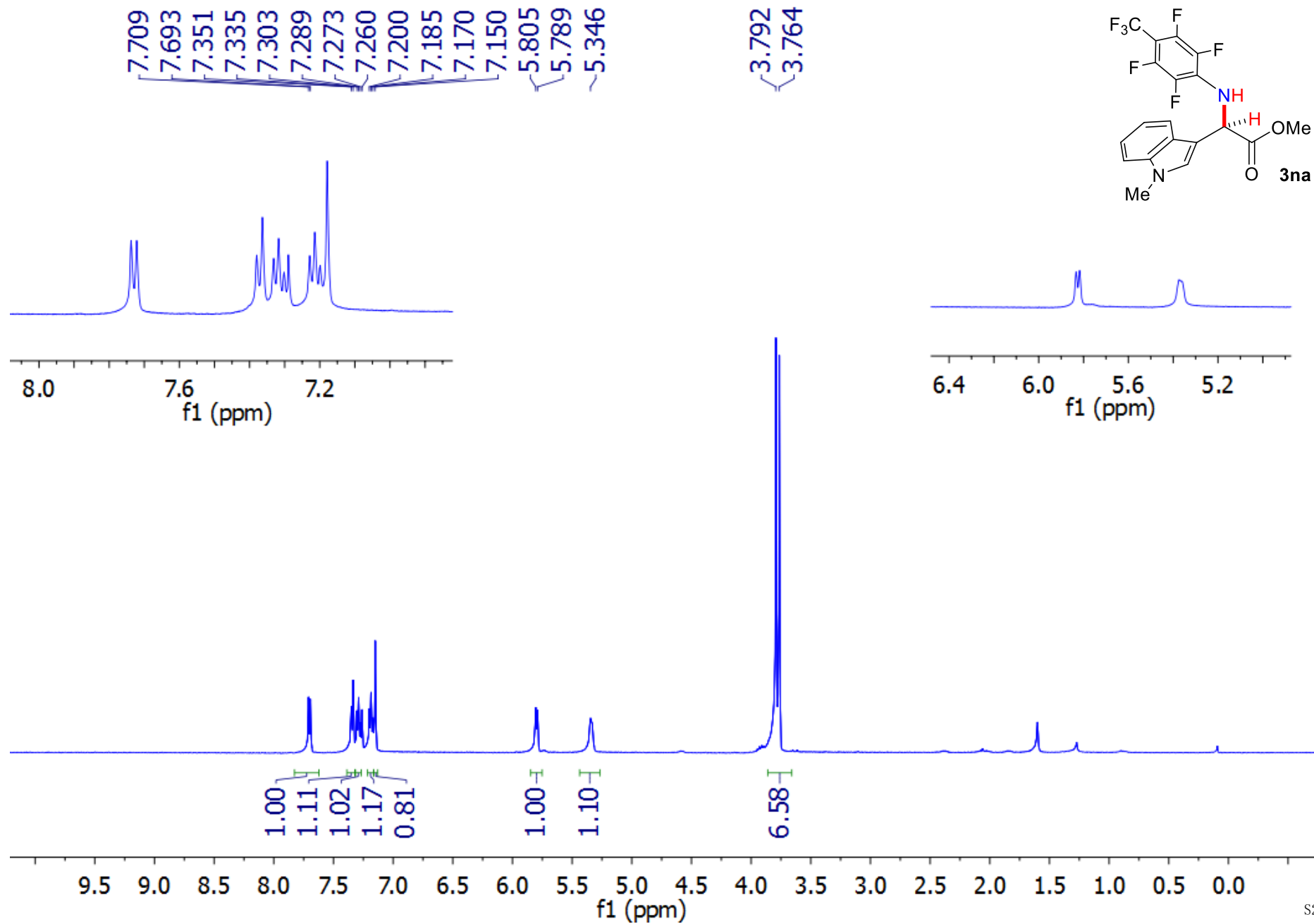
2: 216 nm, 4 nm

Results

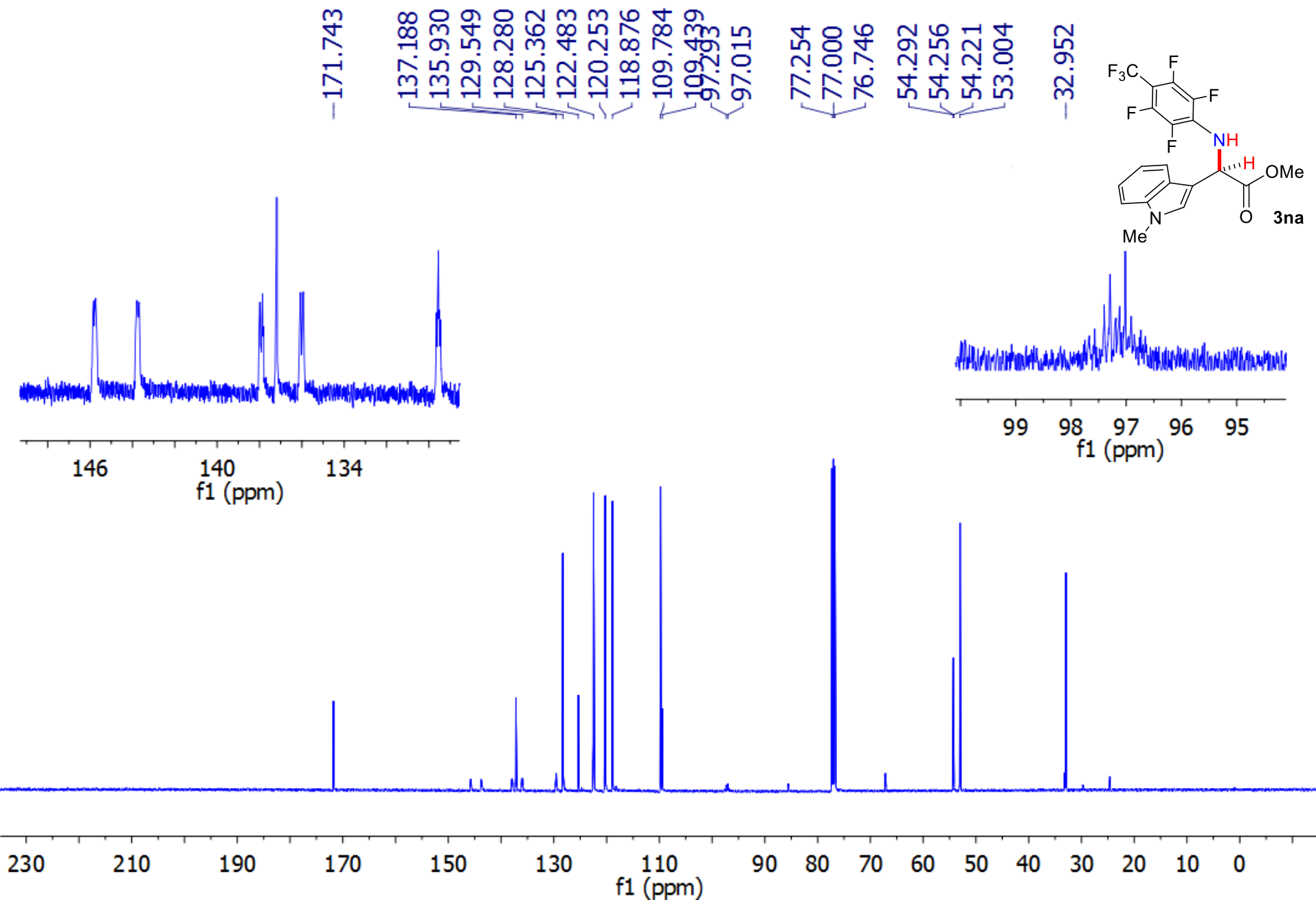
Pk #	Name	Retention Time	Area Percent
1		8.912	93.845
2		10.796	6.155
Totals			100.000



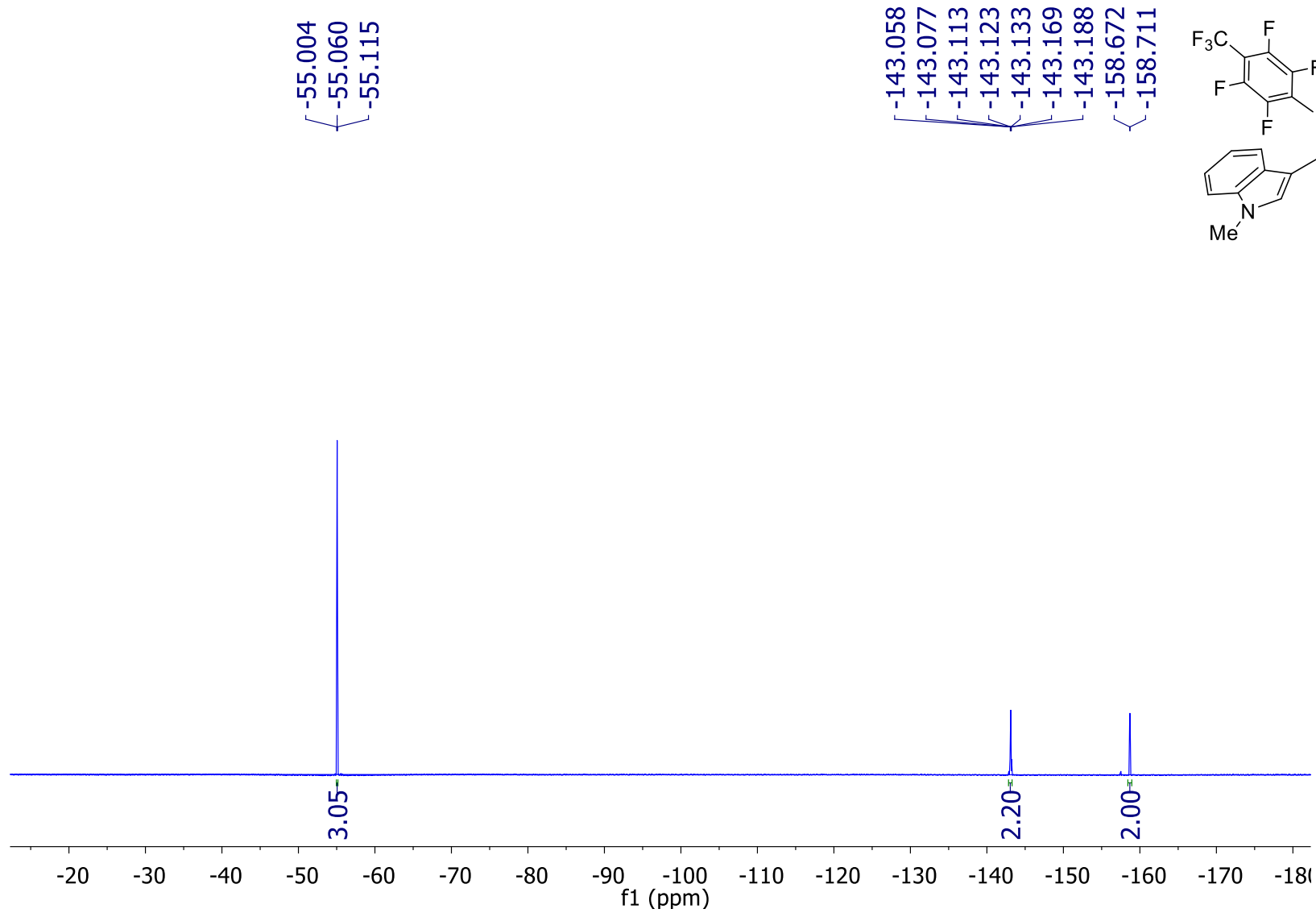
^1H NMR



^{13}C NMR



^{19}F NMR

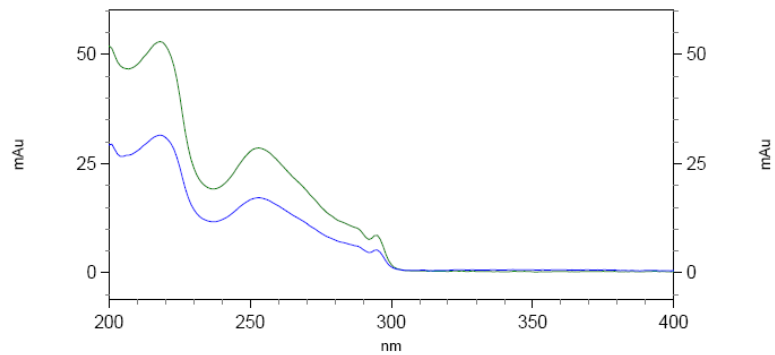
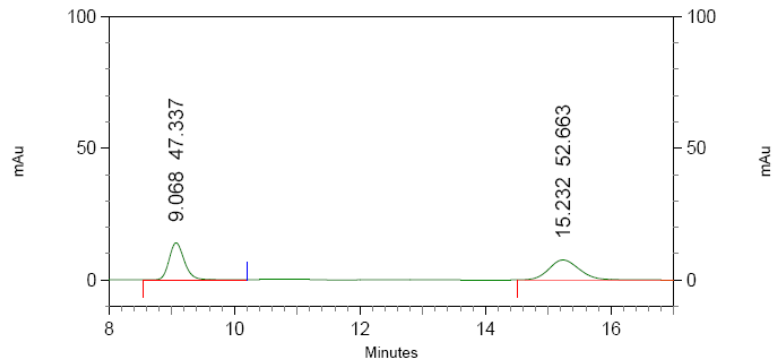


HPLC

JLM-V-264-1-ADH-1%1ML

E:\SMJ 0% 1mL ADH.met

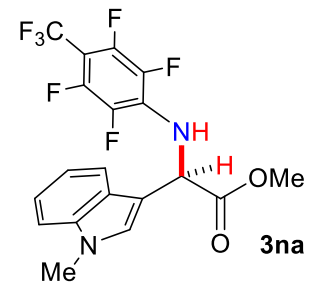
C:\EZStart\Projects\Default\Data\JLM-V-264-1-ADH1%1ML



1: 260 nm, 4 nm

Results

Name	Retention Time	Area Percent	Pk #
	9.068	47.337	1
	15.232	52.663	2
Totals		100.000	

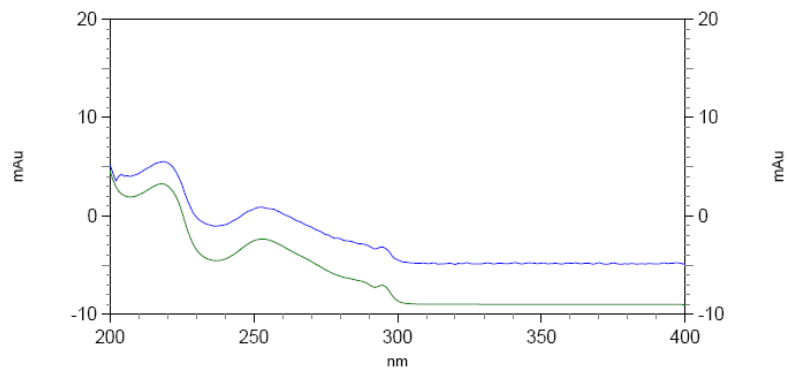
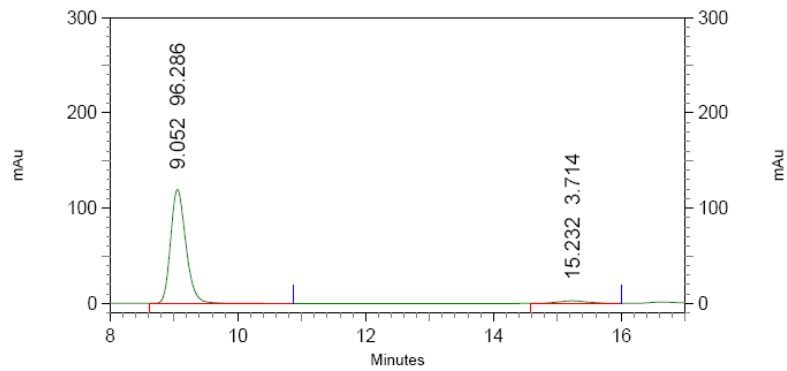


HPLC

JLM-V-264-2-ADH-1%1ML

E:\SMJ 0% 1mL ADH.met

C:\EZStart\Projects\Default\Data\JLM-V-264-2-ADH1%1ML

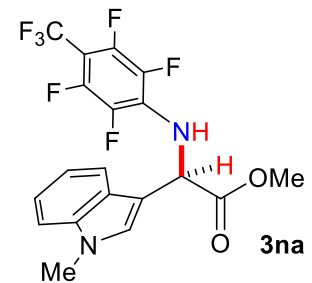


1: 260 nm, 4 nm

Results

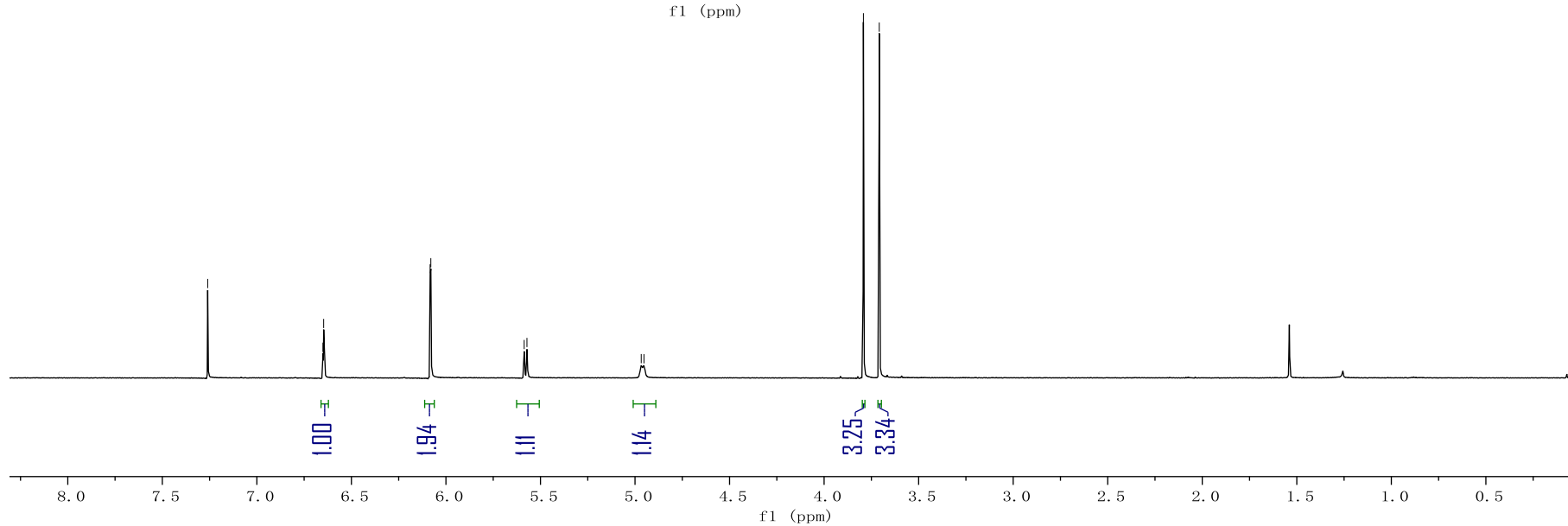
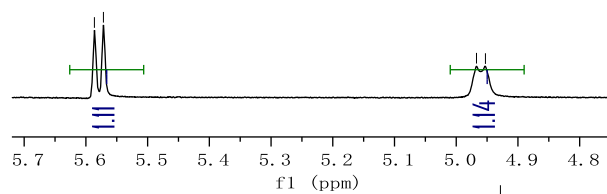
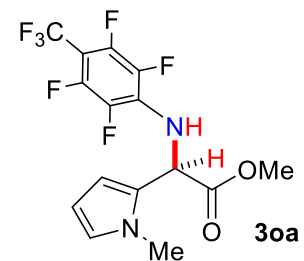
Name	Retention Time	Area Percent	Pk #
	9.052	96.286	1
	15.232	3.714	2

Totals	100.000	
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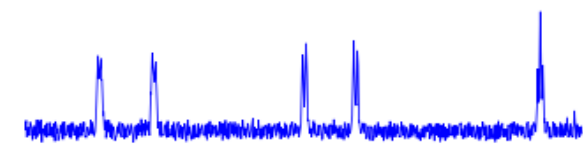
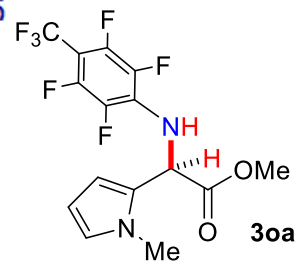
^1H NMR

7.260
6.650
6.647
6.643
6.084
6.080
5.586
5.572
4.967
4.953
3.792
3.708

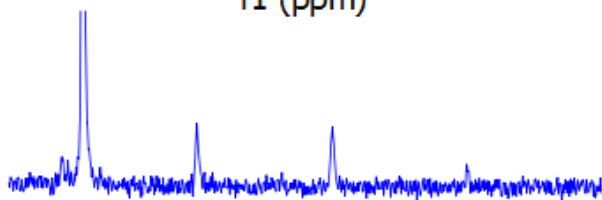


¹³C NMR

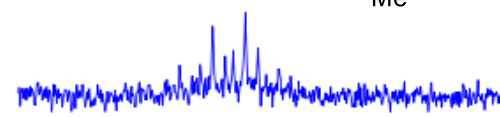
170.554
145.883
143.849
138.216
138.088
136.292
136.164
129.302
126.634
124.298
122.471
120.296
108.135
107.531
77.254
77.000
76.746
53.754
53.716
53.679
53.130
33.916



145
140
135
130
f1 (ppm)



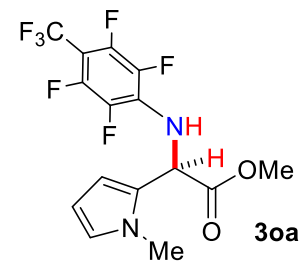
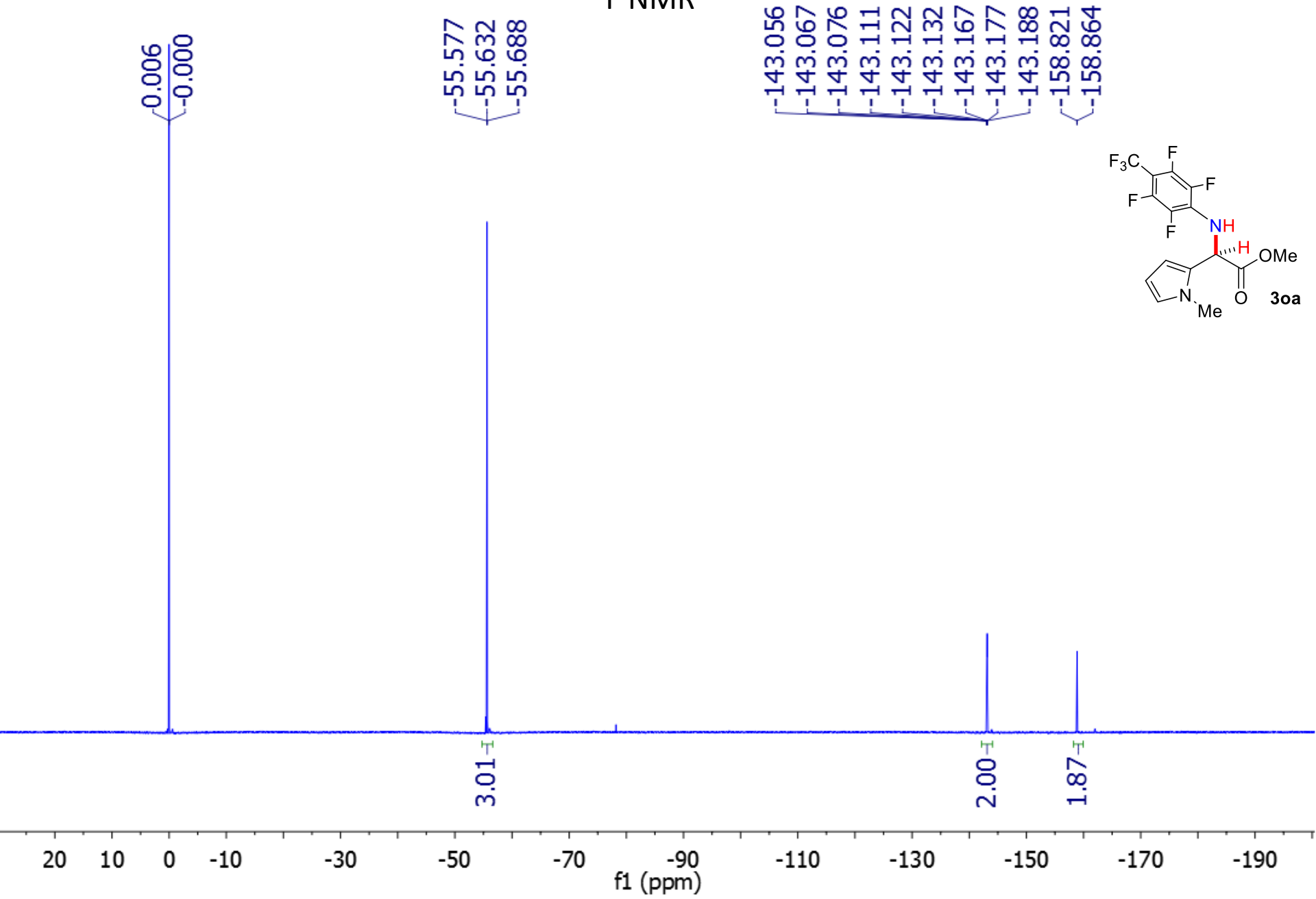
124
121
118
f1 (ppm)



99
98
97
96
f1 (ppm)

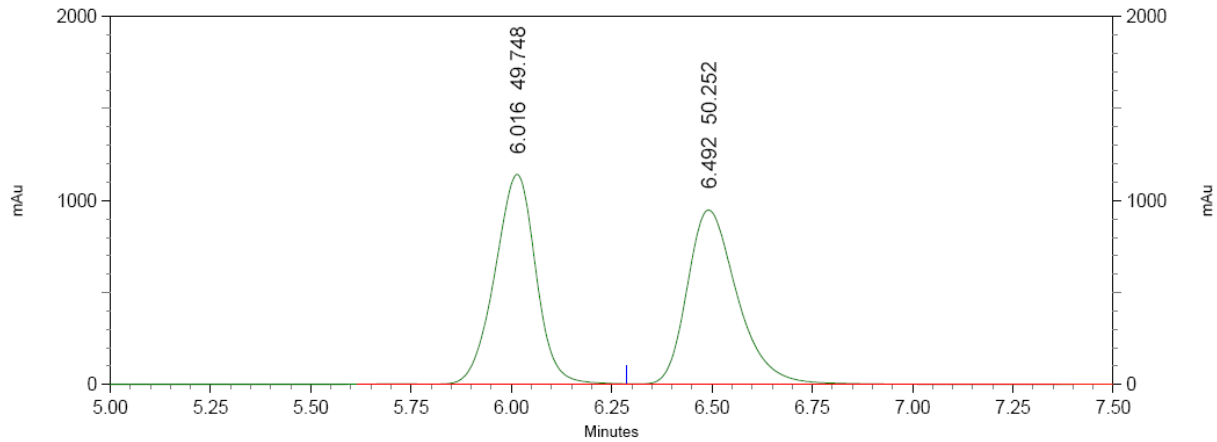
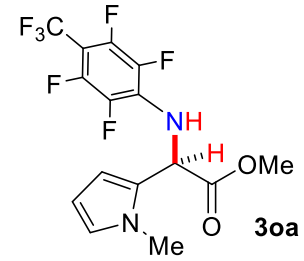
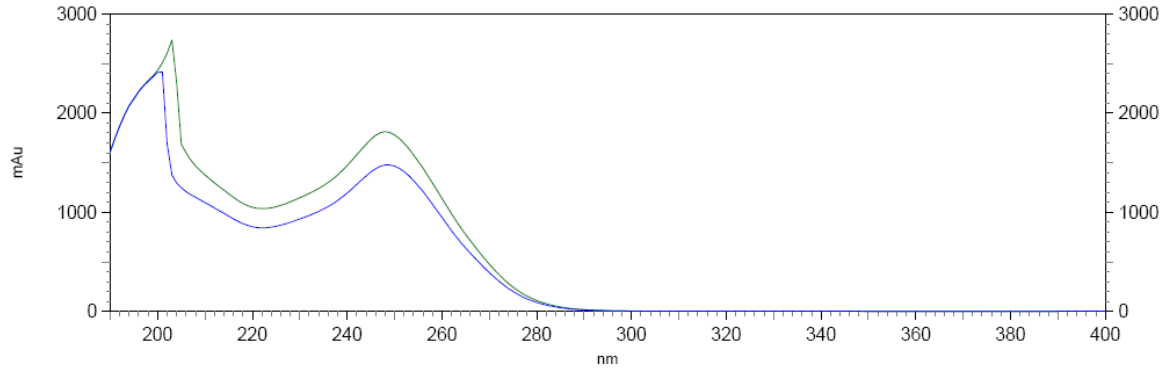
230 210 190 170 150 130 110 90 80 70 60 50 40 30 20 10 0
f1 (ppm)

¹⁹F NMR



HPLC

JLM-V-294-1-ADH-1%1ML
C:\EZStart\Projects\Default\Data\JLM-V-294-1-ADH1%1ML
C:\EZStart\Projects\Default\Method\YC-1%-OJH1ml.met
AD-H column 20%IPA @ 0.8ml/min



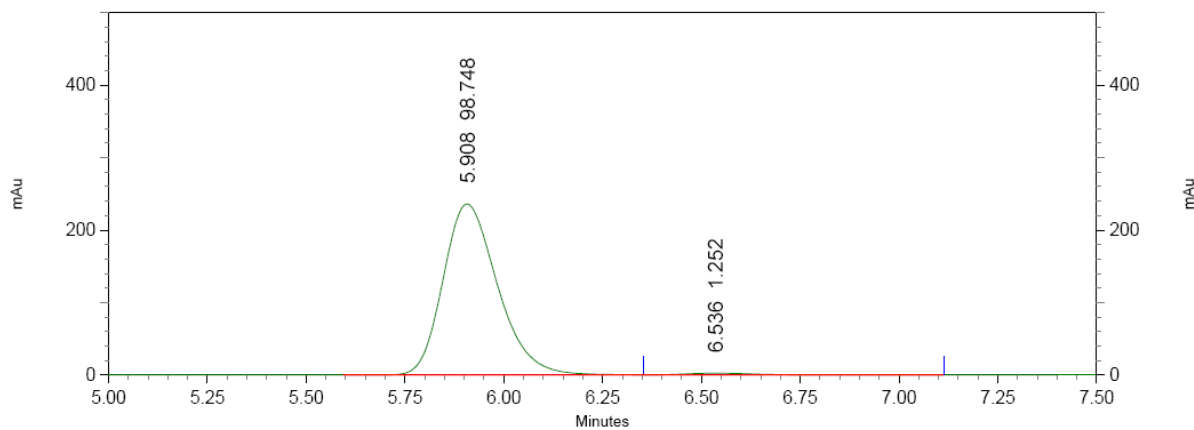
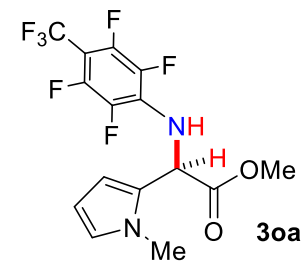
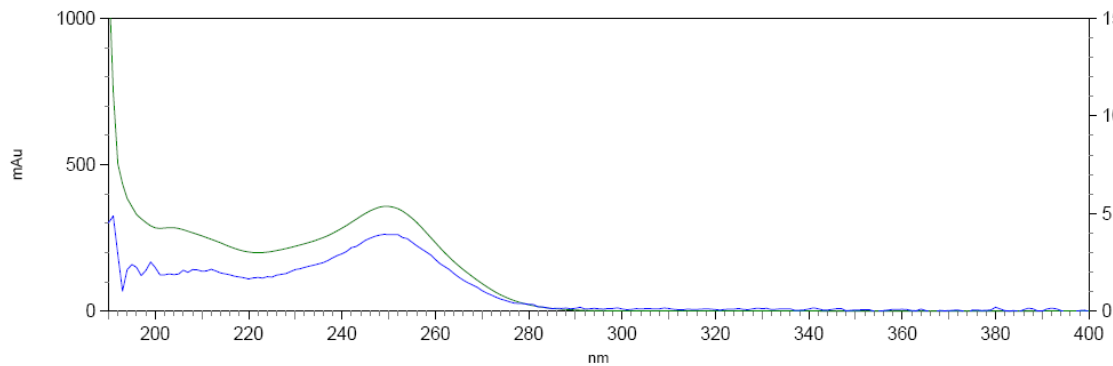
1: 260 nm, 4 nm Results

Pk #	Retention Time	Area Percent
1	6.016	49.748
2	6.492	50.252

Totals	100.000
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HPLC

JLM-V-294-2a-ADH-1%1ML
C:\EZStart\Projects\Default\Data\JLM-V-294-2a-ADH1%1ML
C:\EZStart\Projects\Default\Method\YC-1%-OJH1ml.met
AD-H column 20%IPA @ 0.8ml/min

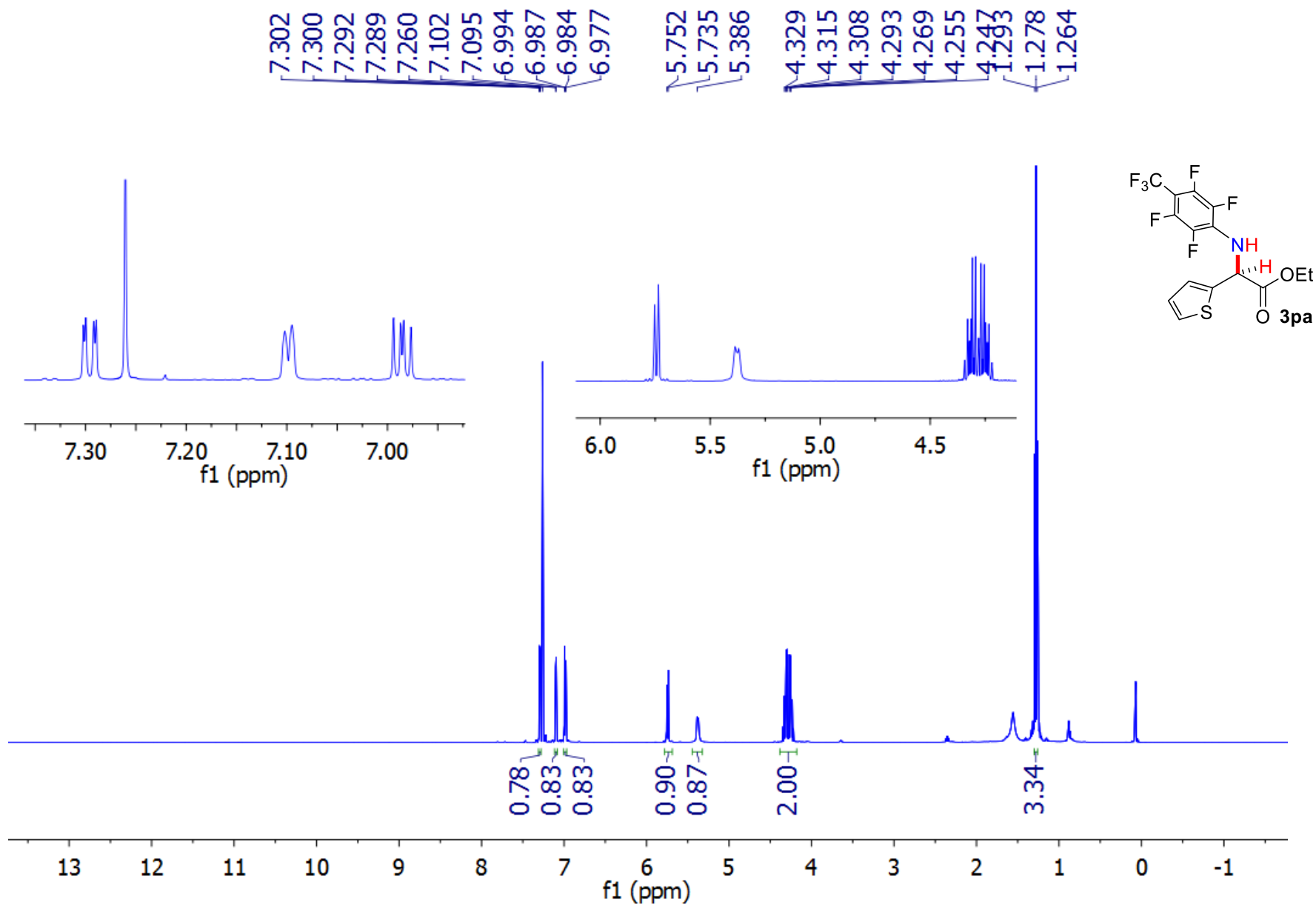


1: 260 nm, 4 nm Results

Pk #	Retention Time	Area Percent
1	5.908	98.748
2	6.536	1.252

Totals	Area Percent
	100.000

^1H NMR



¹³C NMR

169.849

145.892

143.724

139.184

138.029

136.108

128.906

127.241

126.458

126.377

122.464

97.981

77.269

77.015

76.761

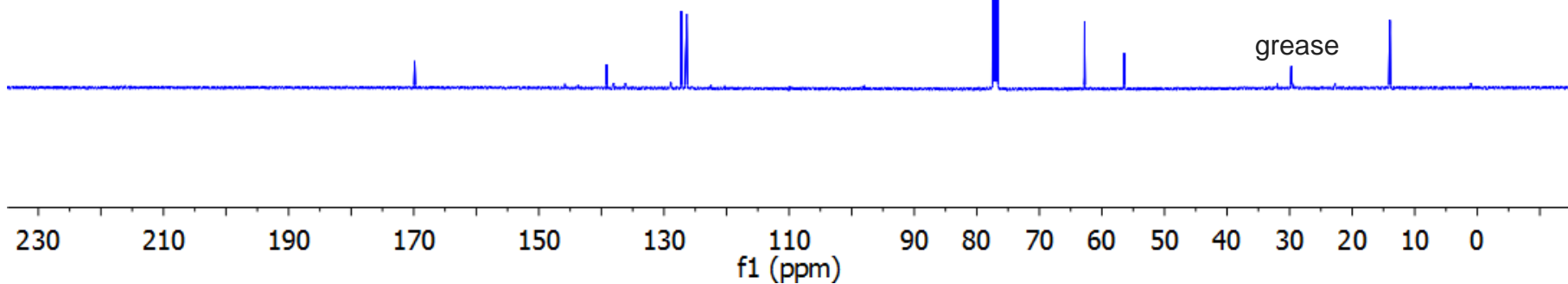
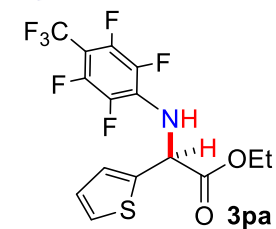
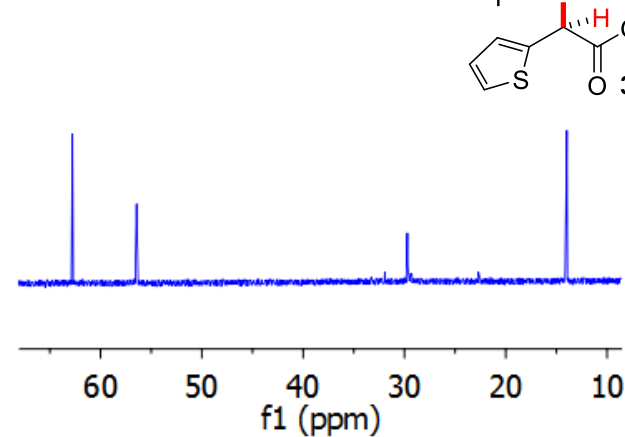
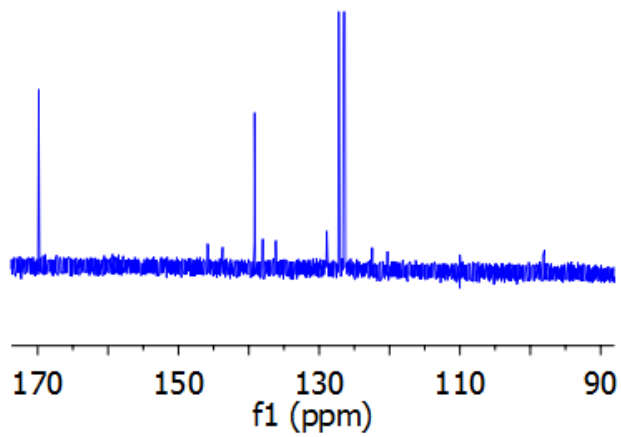
62.776

56.479

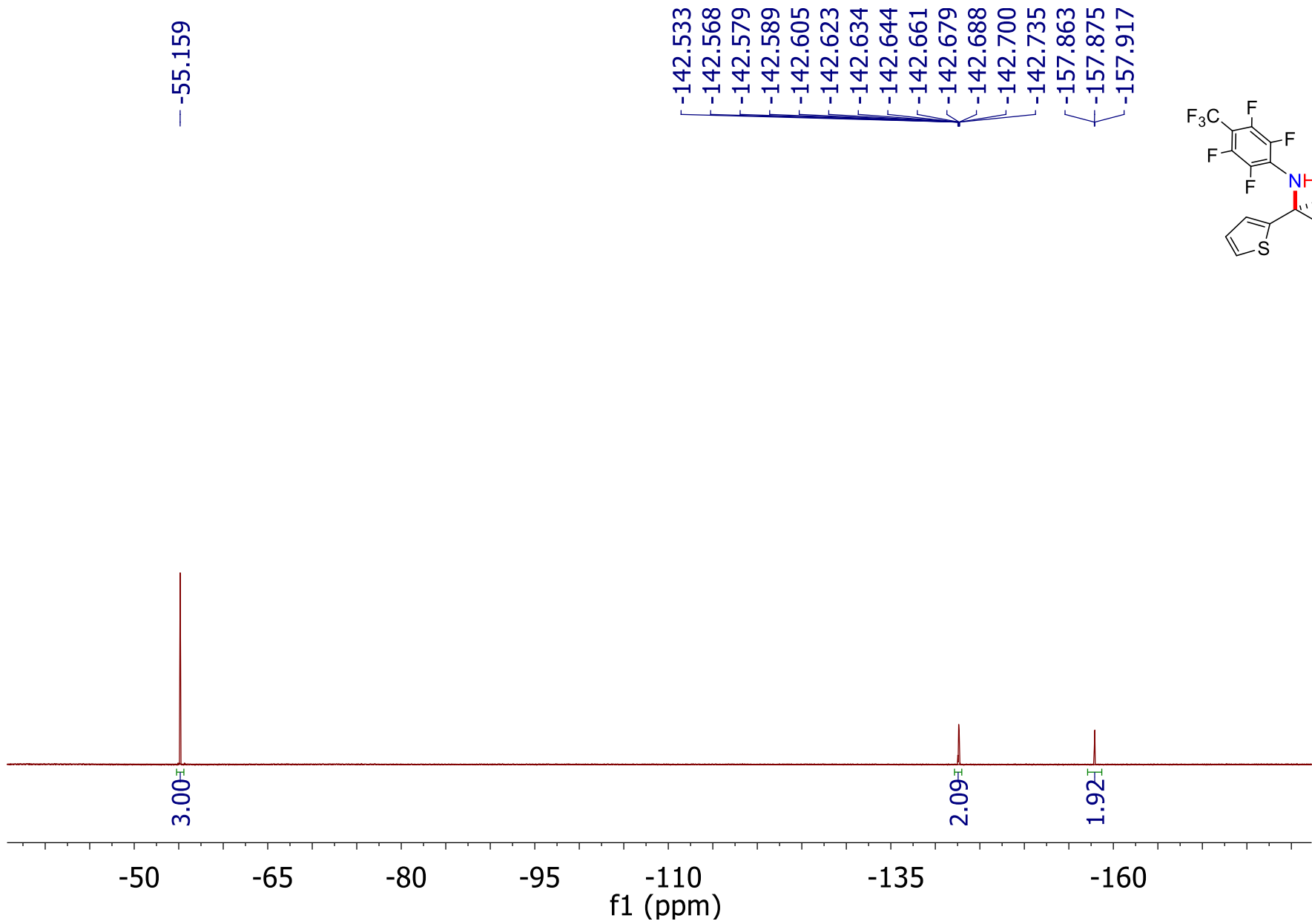
56.442

56.405

13.971



^{19}F NMR

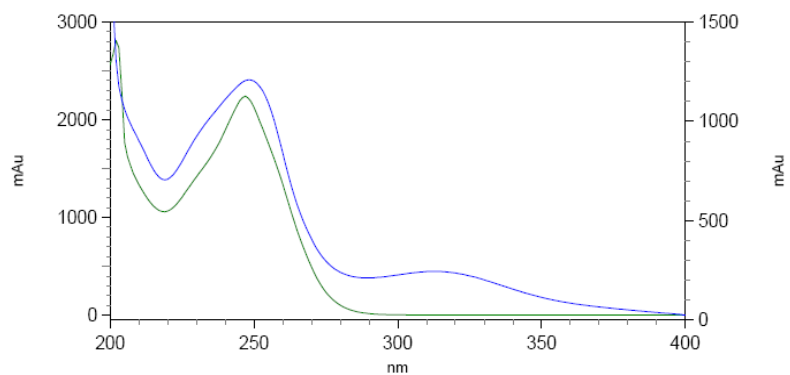
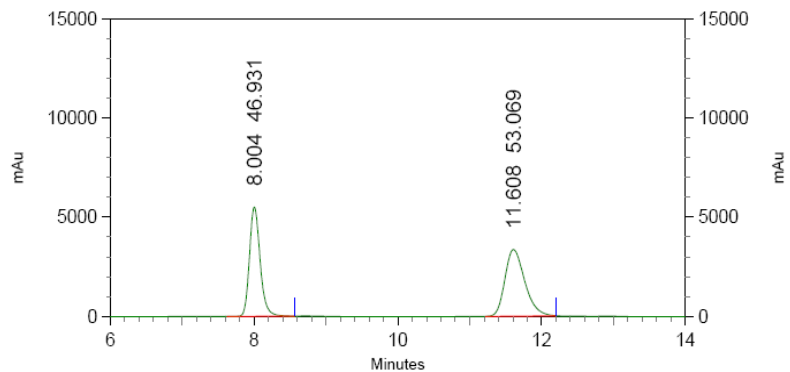
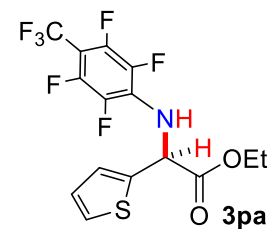


HPLC

JLM-V-144-1a-ODH1%0.7ML

C:\EZStart\Projects\Default\Method\JLM-ODH-0.2%-0.7ml.met

C:\EZStart\Projects\Default\Data\JLM-V-144-1a-ODH1%0.7ML



3: 254 nm, 4 nm

Results

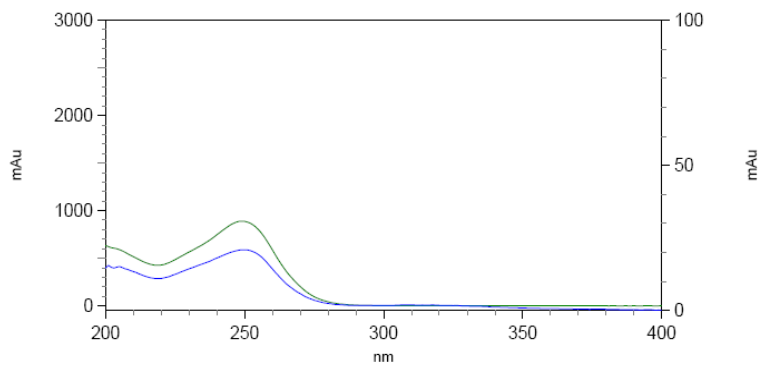
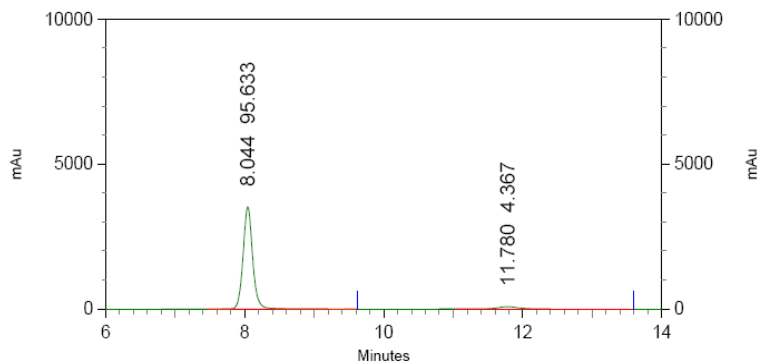
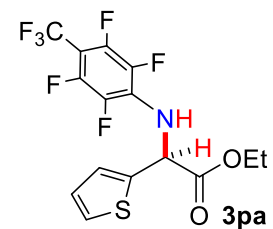
Pk #	Name	Retention Time	Area Percent
1		8.004	46.931
2		11.608	53.069
Totals			100.000

HPLC

JLM-V-144-2a-ODH1%0.7ML

C:\EZStart\Projects\Default\Method\JLM-ODH-0.2%-0.7ml.met

C:\EZStart\Projects\Default\Data\JLM-V-144-2a-ODH1%0.7ML



4: 247 nm, 4 nm

Results

Pk #	Name	Retention Time	Area Percent
1		8.044	95.633
2		11.780	4.367

Totals			100.000
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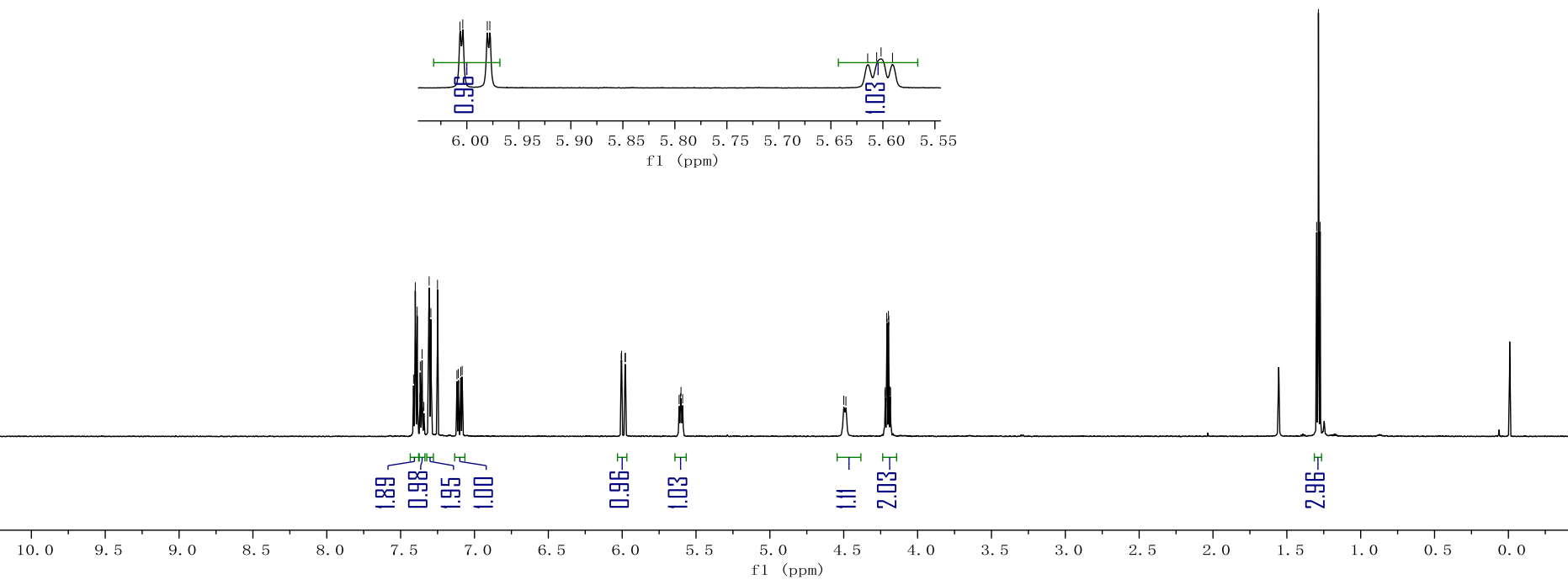
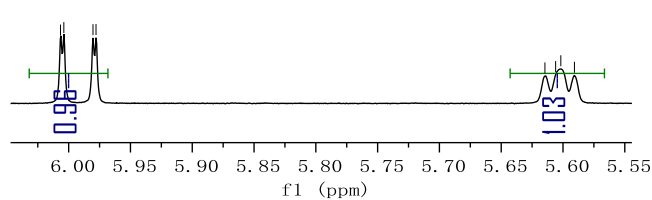
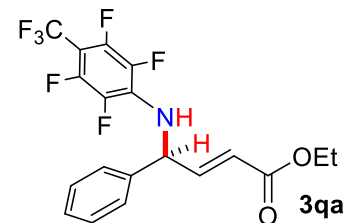
¹H NMR

7.401
7.398
7.388
7.367
7.355
7.355
7.310
7.307
7.298
7.295
7.250
7.119
7.109
7.093
7.083
6.007
6.004
5.980
5.978
5.615
5.606
5.602
5.591
4.500
4.485
4.221
4.219
4.209
4.207
4.197
4.195
4.185
4.183

1.298
1.286
1.274

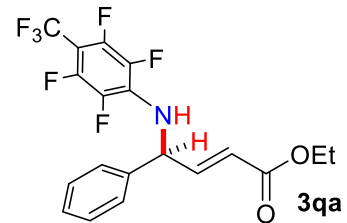
6.007
6.004
5.980
5.978

5.615
5.606
5.602
5.591



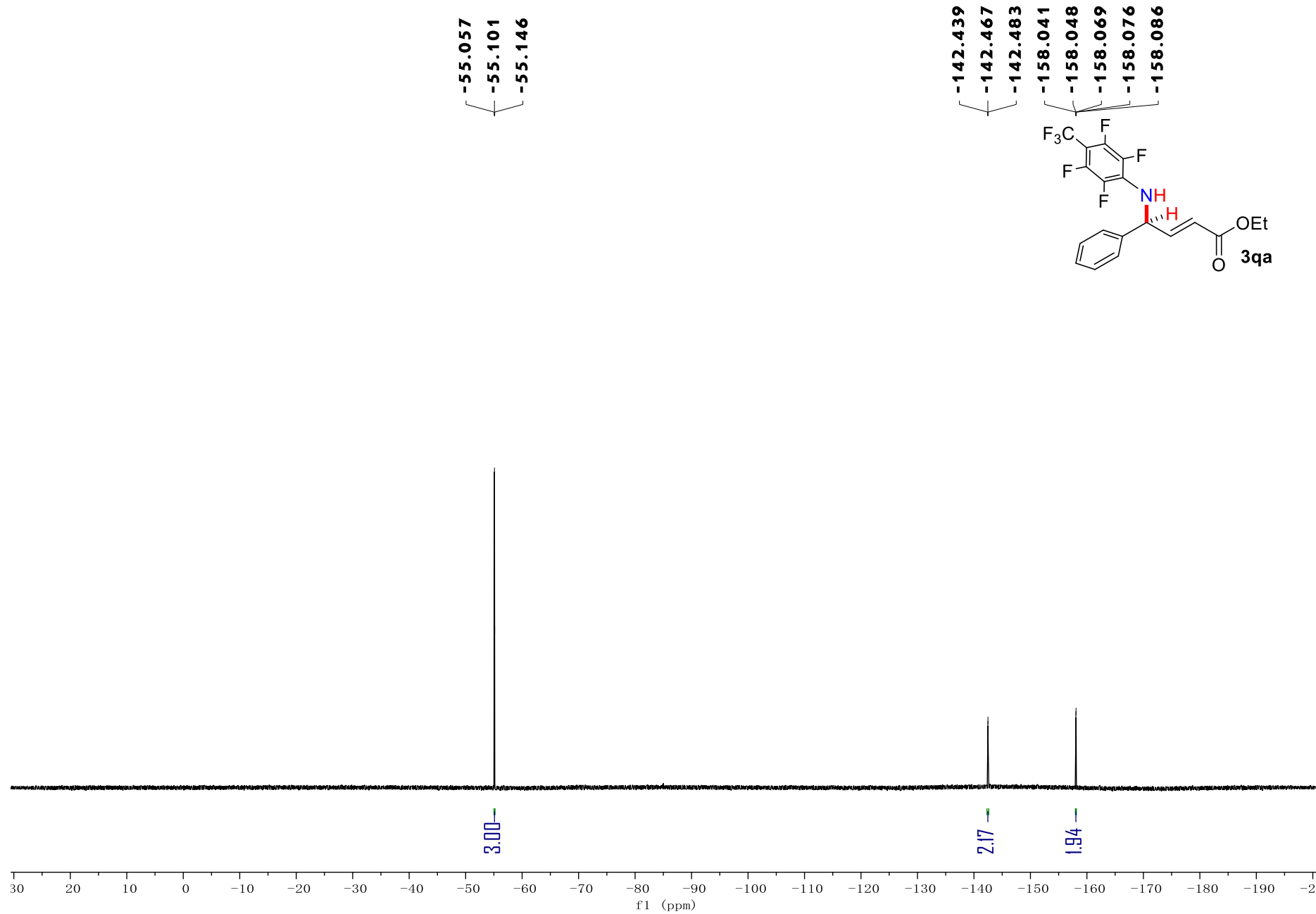
¹³C NMR

165.644
146.029
145.755
145.649
144.001
143.952
143.936
143.851
138.639
137.639
137.605
137.501
137.467
136.041
136.007
135.977
135.938
135.905
135.872
129.447
129.384
129.361
129.288
128.782
126.889
124.017
122.604
122.207
120.397
118.586
97.856
97.768
97.679
97.623
97.535
97.444
60.775
59.447
59.416
59.386
14.083



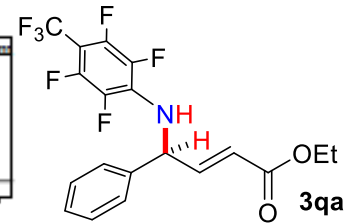
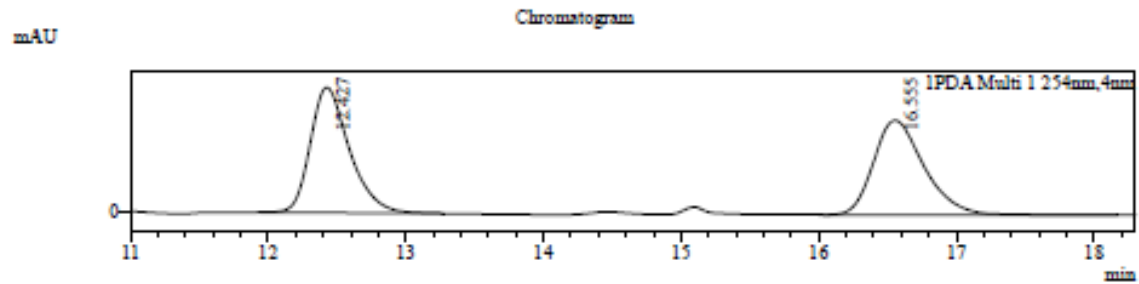
200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10
f1 (ppm)

^{19}F NMR

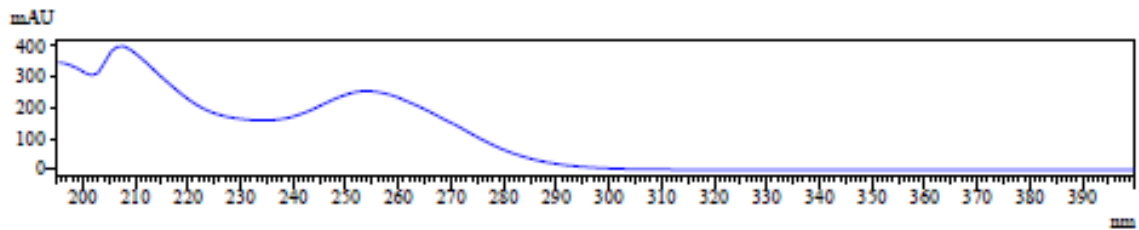


HPLC

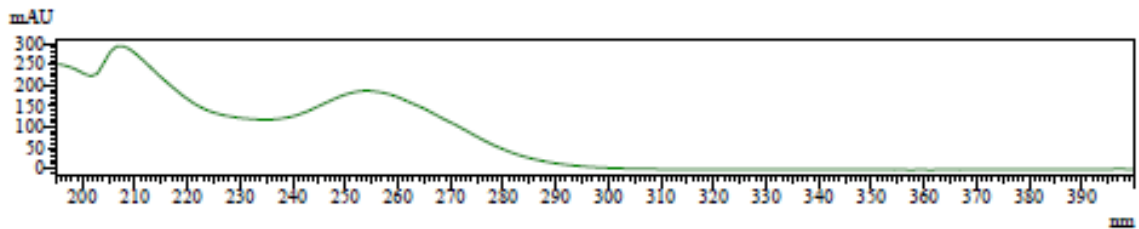
Sample Information
Sample Name : P0X403-ODH-5%0.8mL
Sample ID : P0X403-ODH-5%0.8mL
Data File : P0X403-ODH-5%0.8mL.lcd
Method File : XXW-5%.0.8.mL.lcm



UV Spectrum
Retention time = 12.427



UV Spectrum
Retention time = 16.555



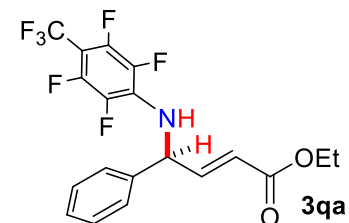
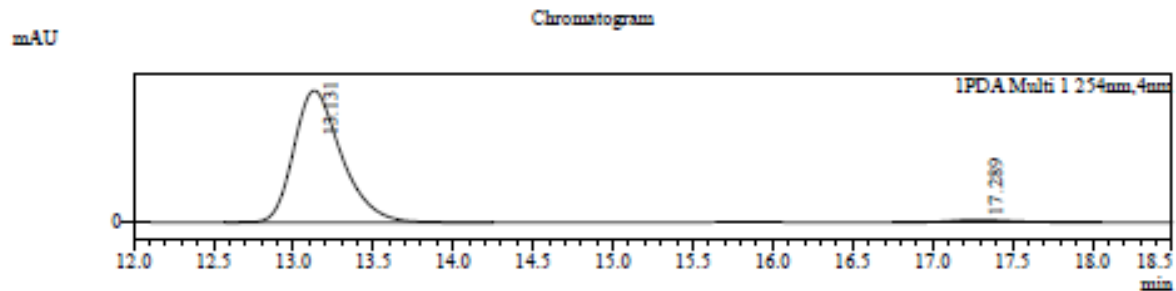
Peak Table

PDA Ch1 254nm

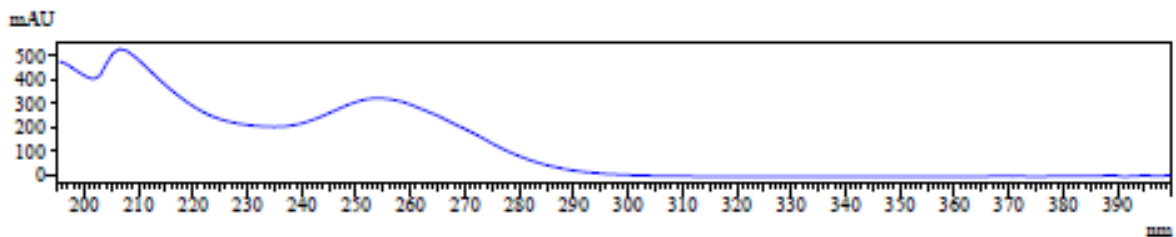
Peak#	Ret. Time	Area	Area%
1	12.427	4973148	50.583
2	16.555	4858545	49.417
Total		9831693	100.000

HPLC

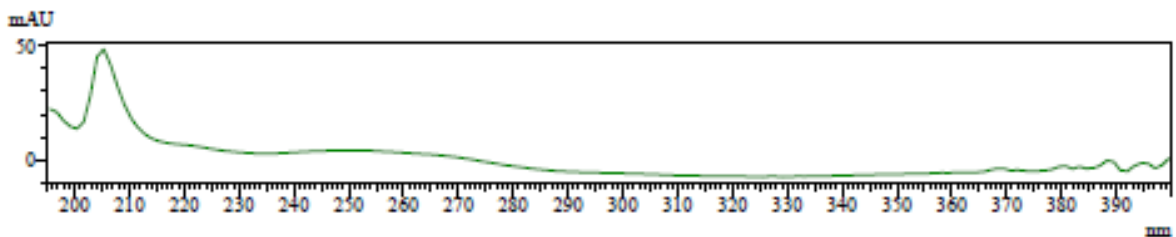
Sample Information
 Sample Name : P0X413-ODH-5%0.8mL
 Sample ID : P0X413-ODH-5%0.8mL
 Data File : P0X413-ODH-5%0.8mL.lcd
 Method File : P0X-5%.0.8.mL-15Min.lcm



UV Spectrum
 Retention time = 13.131



UV Spectrum
 Retention time = 17.289



Peak Table

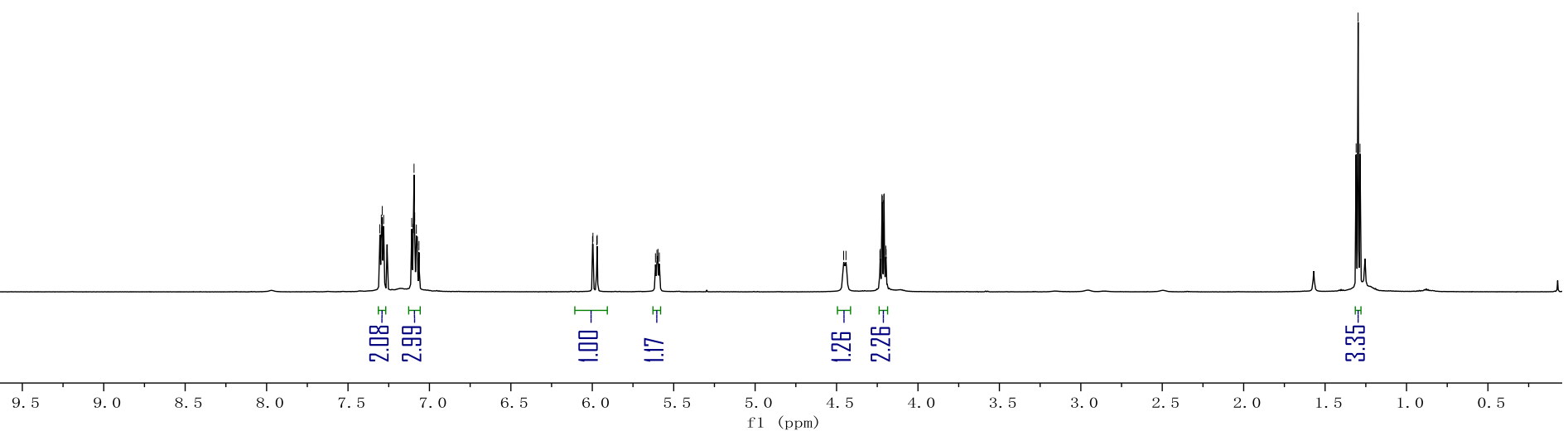
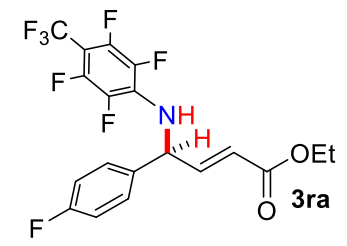
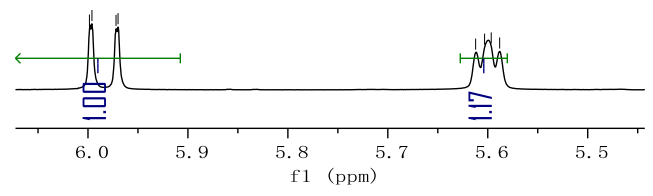
Peak#	Ret. Time	Area	Area%
1	13.131	6677753	97.205
2	17.289	191990	2.795
Total		6869744	100.000

¹H NMR

7.304
7.296
7.290
7.281
7.109
7.100
7.095
7.090
7.080
7.074
7.064
5.999
5.996
5.972
5.970
5.612
5.603
5.597
5.588
4.457
4.442
4.234
4.231
4.228
4.221
4.219
4.210
4.208
4.197
4.196

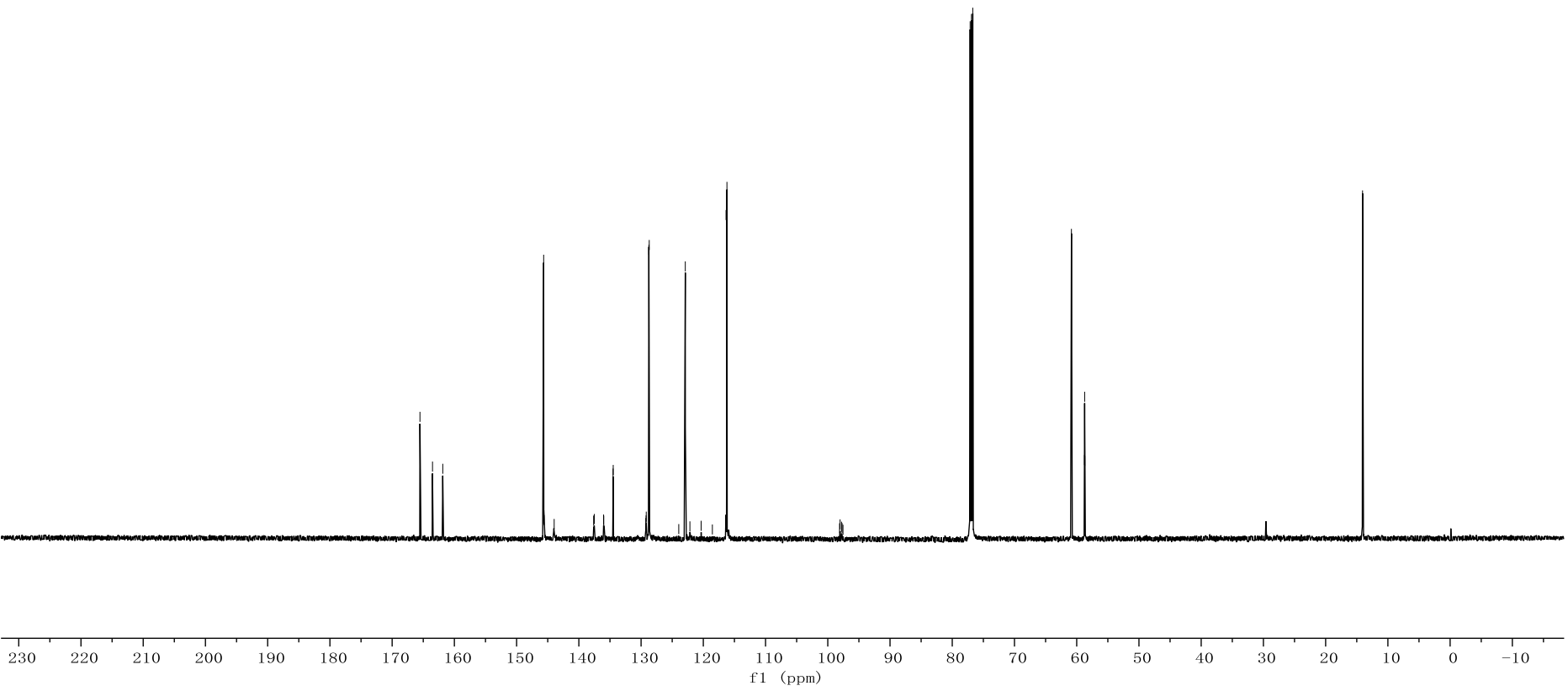
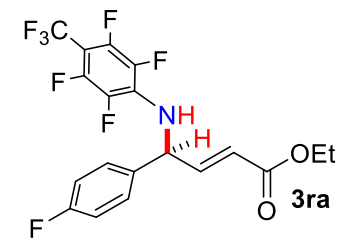
1.309
1.298
1.286

5.999
5.996
5.972
5.970
5.612
5.603
5.597
5.588

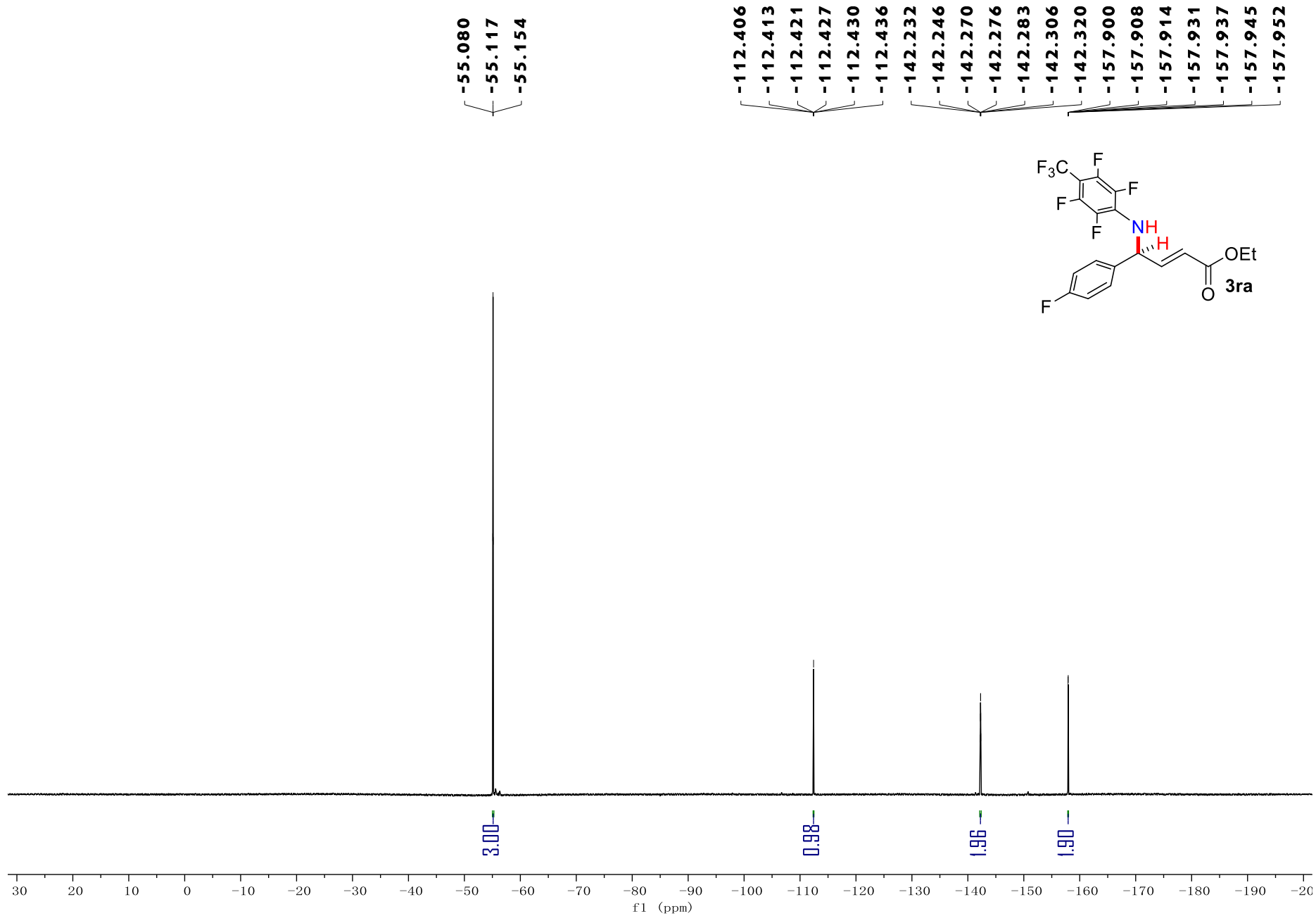


¹³C NMR

165.529
163.513
161.865
145.709
145.653
145.608
145.585
143.964
137.624
137.516
136.025
135.996
134.500
134.477
129.242
129.192
129.168
128.747
128.690
123.946
122.895
122.156
120.351
118.544
116.348
116.203
98.109
98.021
97.789
97.700
97.557
77.125
76.912
76.701
60.849
58.754
58.724
58.693
14.071



¹⁹F NMR



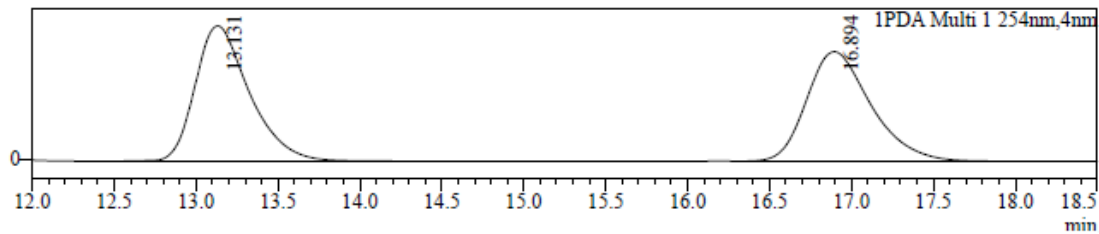
HPLC

Sample information

Sample Name : P0X-0575-ODH-5%0.8mL
Sample ID : P0X-0575-ODH-5%0.8mL
Data File : P0X-0575-ODH-5%0.8mL.lcd
Method File : XXW-5%.0.8.mL.lcm

Chromatogram

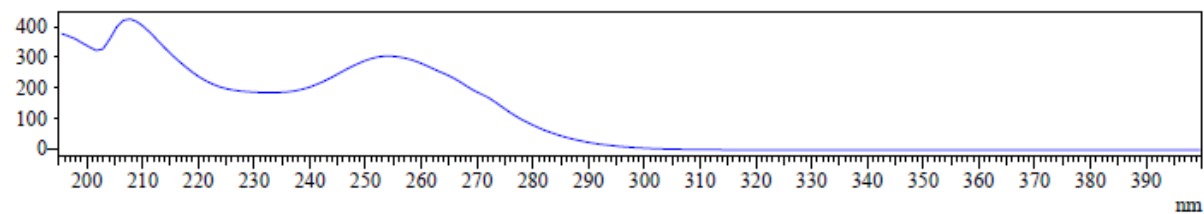
mAU



UV Spectrum

Retention time = 13.131

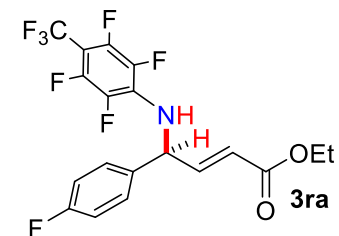
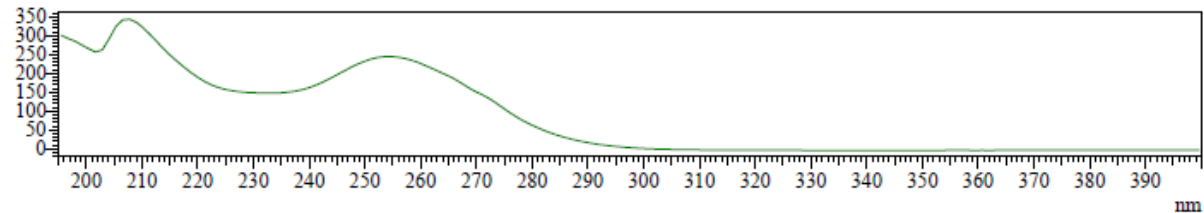
mAU



UV Spectrum

Retention time = 16.894

mAU



Peak Table

PDA Ch1 254nm

Peak#	Ret. Time	Area	Area%
1	13.131	7006663	51.058
2	16.894	6716174	48.942
Total		13722837	100.000

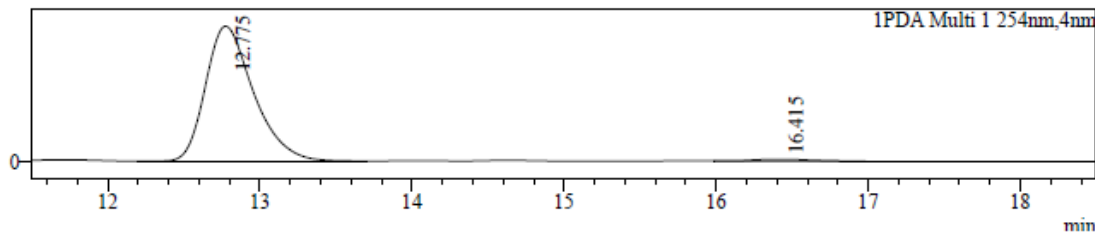
HPLC

Sample Information

Sample Name : P0X-0576-ODH-5%0.8mL
Sample ID : P0X-0576-ODH-5%0.8mL
Data File : P0X-0576-ODH-5%0.8mL.lcd
Method File : XXW-5%0.8.mL.lcm

Chromatogram

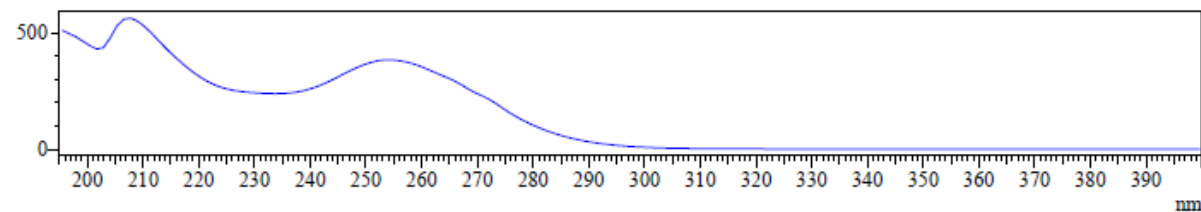
mAU



UV Spectrum

Retention time = 12.775

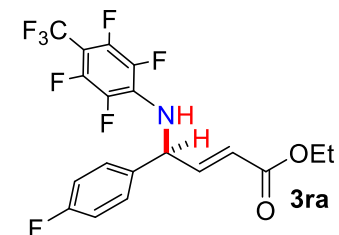
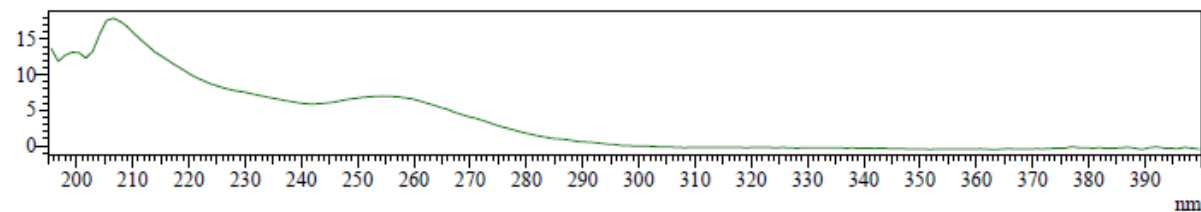
mAU



UV Spectrum

Retention time = 16.415

mAU



Peak Table

PDA Ch1 254nm

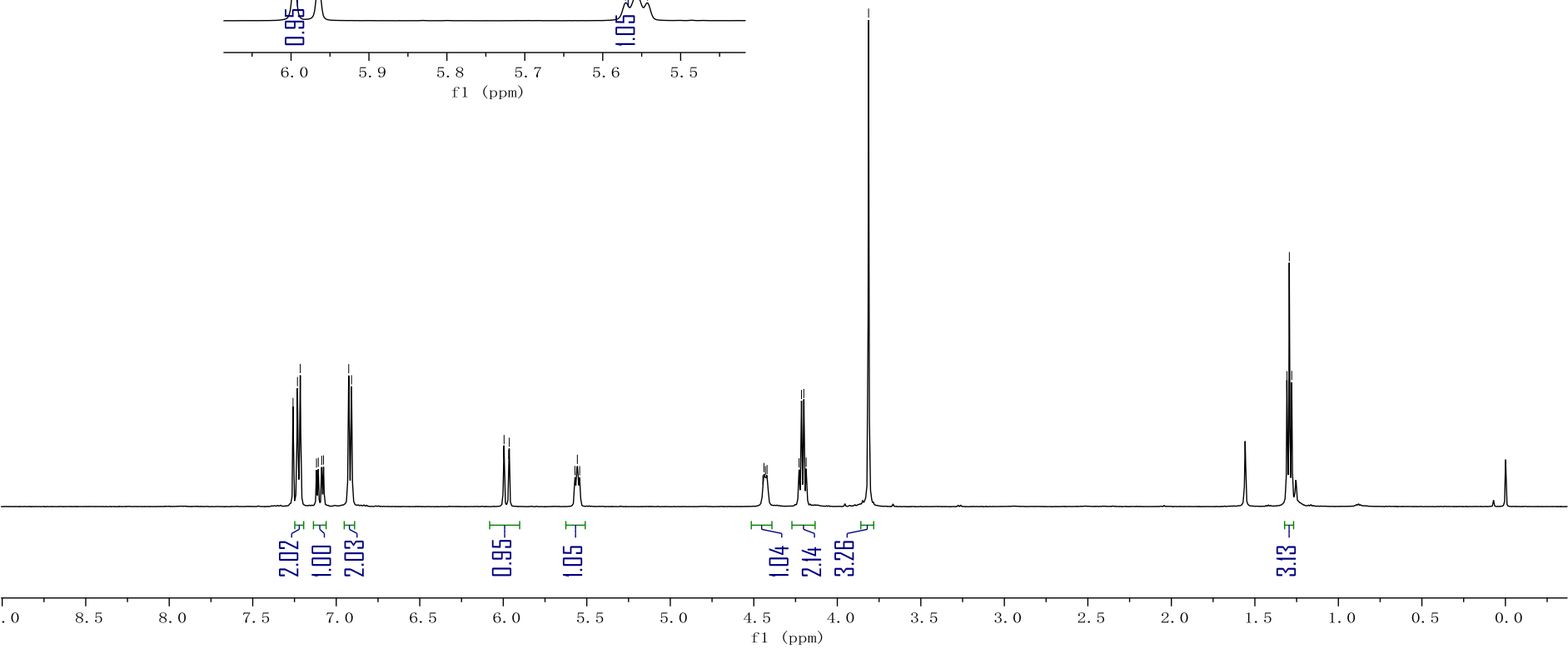
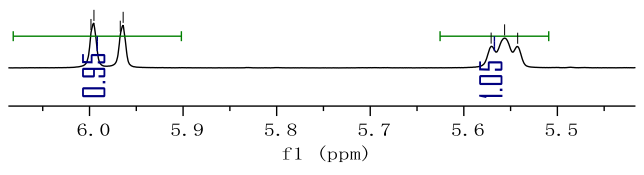
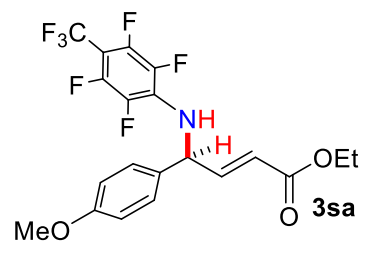
Peak#	Ret. Time	Area	Area%
1	12.775	8307811	98.288
2	16.415	144733	1.712
Total		8452543	100.000

¹H NMR

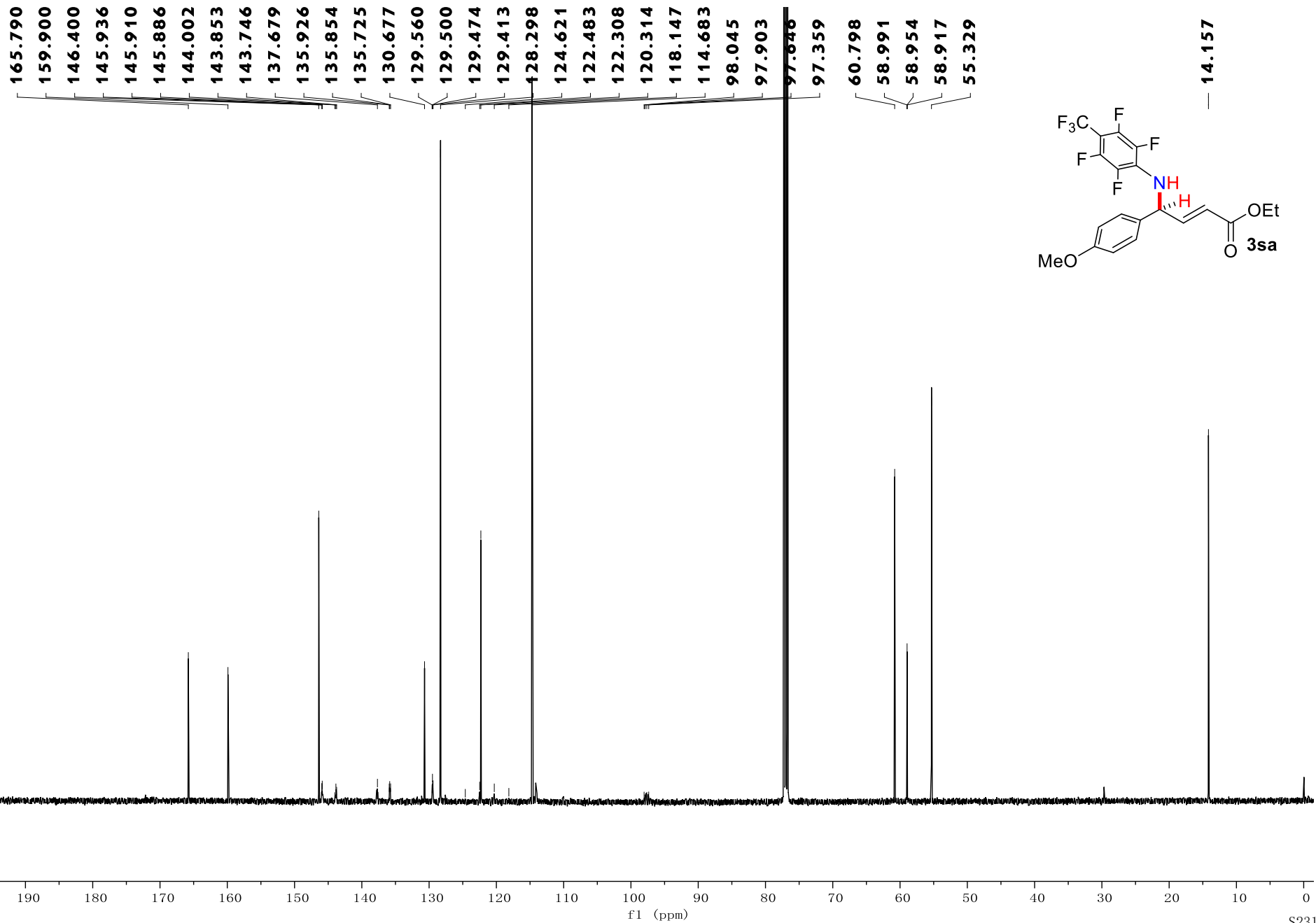
7.259
7.232
7.216
7.118
7.107
7.087
7.076
6.925
6.909
5.998
5.995
5.967
5.964
5.571
5.556
5.542
4.446
4.439
4.430
4.421
4.229
4.215
4.201
4.186
3.813

1.307
1.294
1.279

5.998
5.995
5.967
5.964
5.571
5.556
5.542



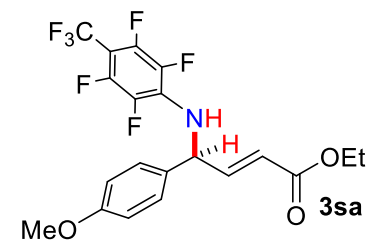
¹³C NMR



¹⁹F NMR

-55.039
-55.084
-55.128

-142.503
-142.517
-142.532
-142.547
-142.562
-142.576
-142.591
-142.607
-142.620
-142.634
-142.665
-142.681
-158.086
-158.094
-158.104
-158.111
-158.131
-158.138
-158.148
-158.156



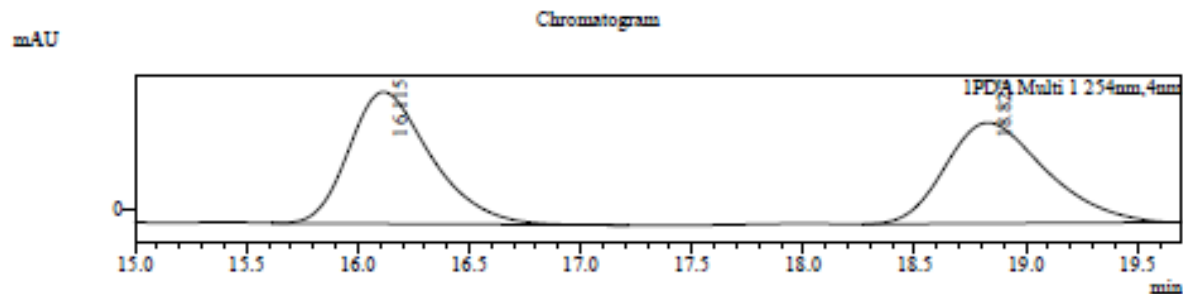
3.00

2.03

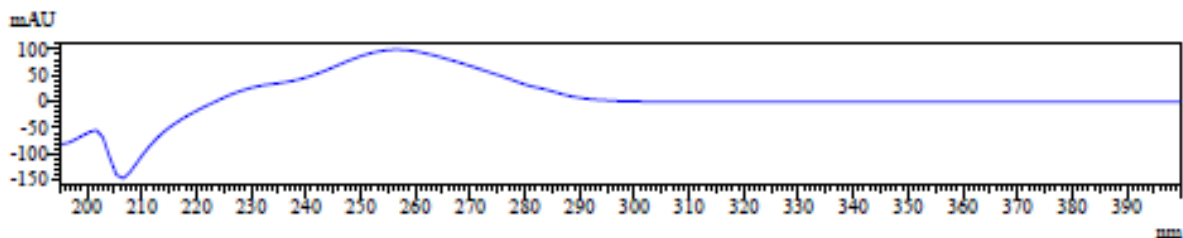
1.97

HPLC

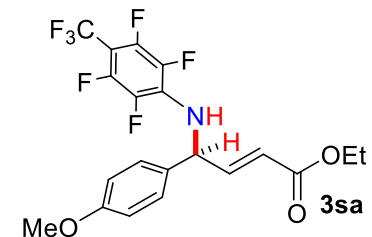
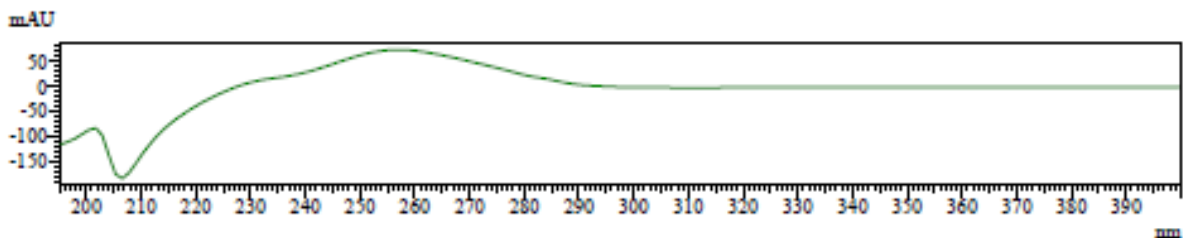
Sample Information
 Sample Name : P0X-0548-3-ODH-5%0.8mL
 Sample ID : P0X-0548-3-ODH-5%0.8mL
 Data File : P0X-0548-3-ODH-5%0.8mL.lcd
 Method File : XXW-5%0.8.mL.lcm



UV Spectrum
 Retention time = 16.115



UV Spectrum
 Retention time = 18.827



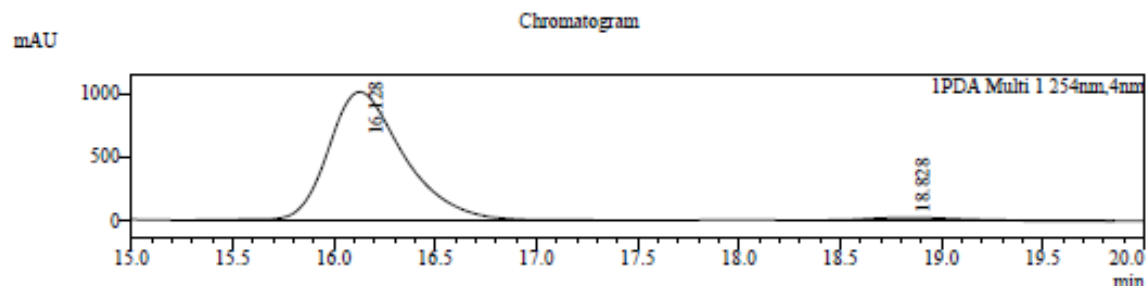
Peak Table

PDA Ch1 254nm

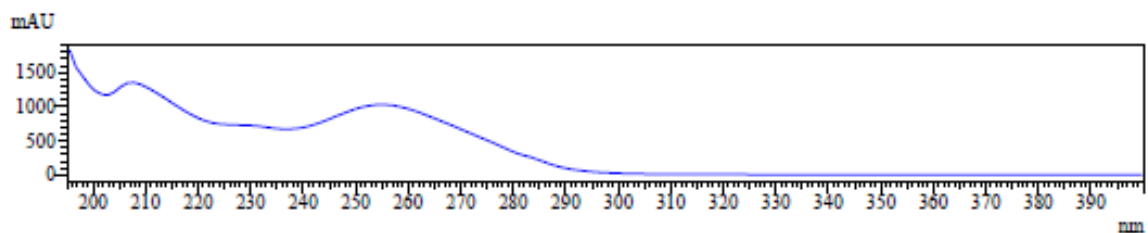
Peak#	Ret. Time	Area	Area%
1	16.115	2740801	51.715
2	18.827	2559026	48.285
Total		5299827	100.000

HPLC

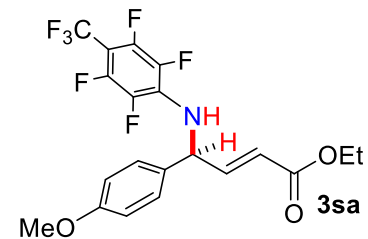
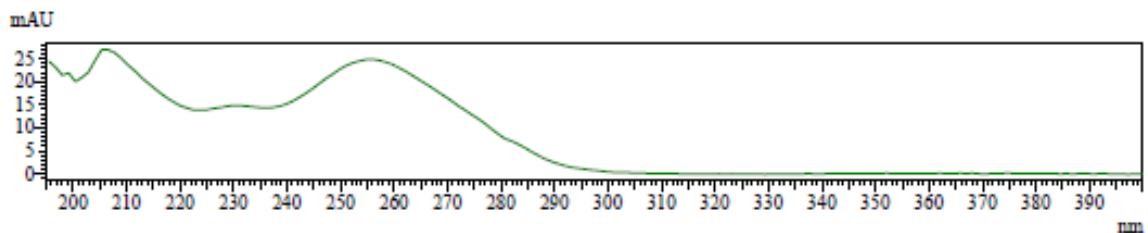
Sample Information
 Sample Name : P0X-0582-ODH-5%0.8mL
 Sample ID : P0X-0582-ODH-5%0.8mL
 Data File : P0X-0582-ODH-5%0.8mL.lcd
 Method File : P0X-5.0%-0.8ml.lcm



UV Spectrum
 Retention time = 16.128



UV Spectrum
 Retention time = 18.828



Peak Table

Peak#	Ret. Time	Area	Area%
1	16.128	26349173	97.394
2	18.828	704902	2.606
Total		27054075	100.000

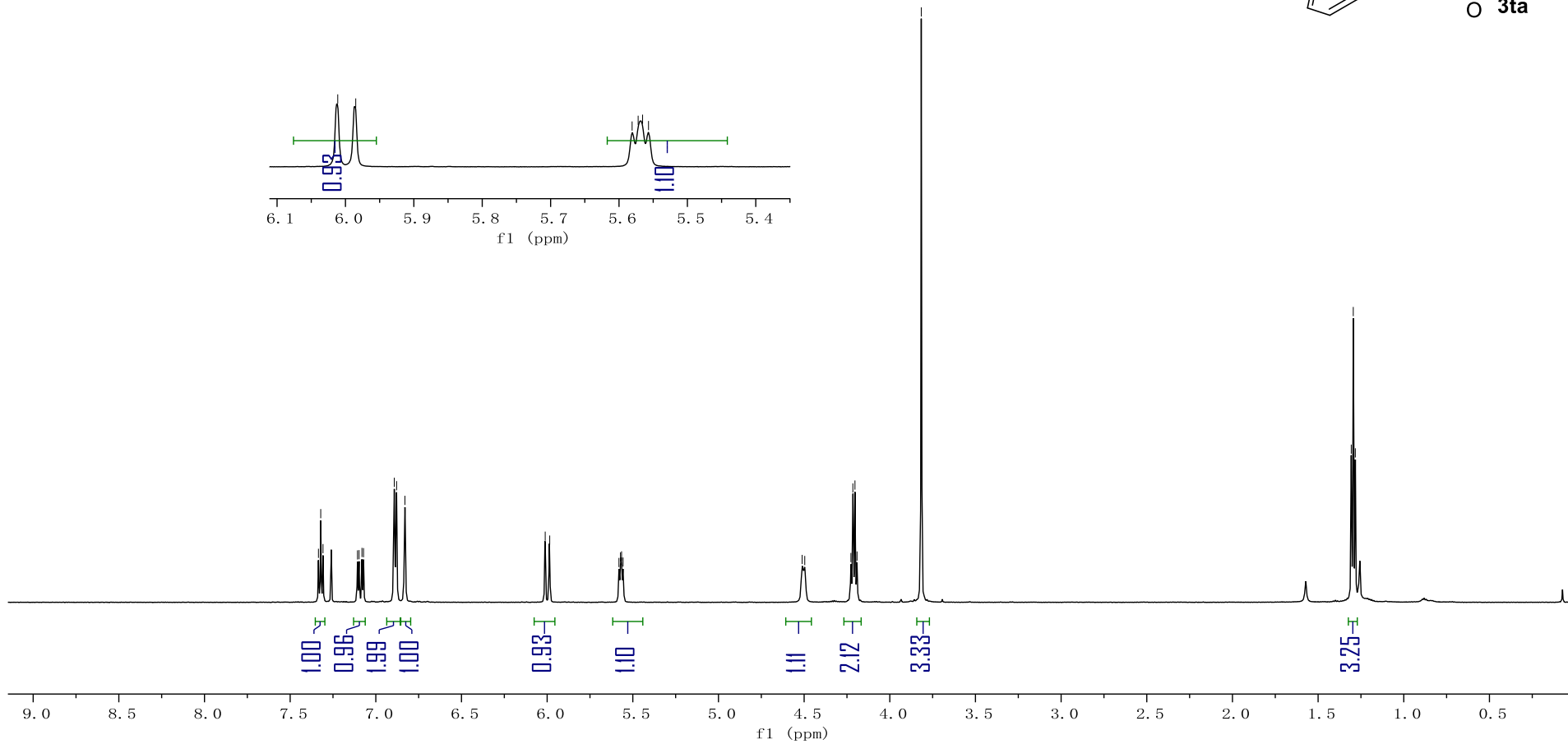
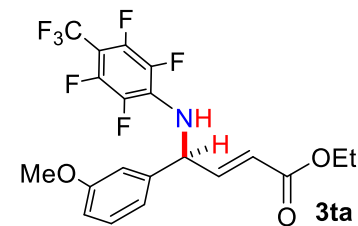
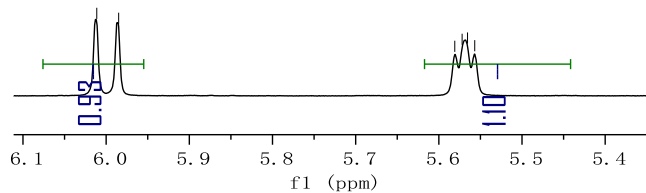
¹H NMR

7.335
7.322
7.308
7.107
7.098
7.081
7.072
6.892
6.879
6.830
6.011
5.985
5.581
5.572
5.566
5.557
4.512
4.496
4.227
4.216
4.203
4.191
3.816

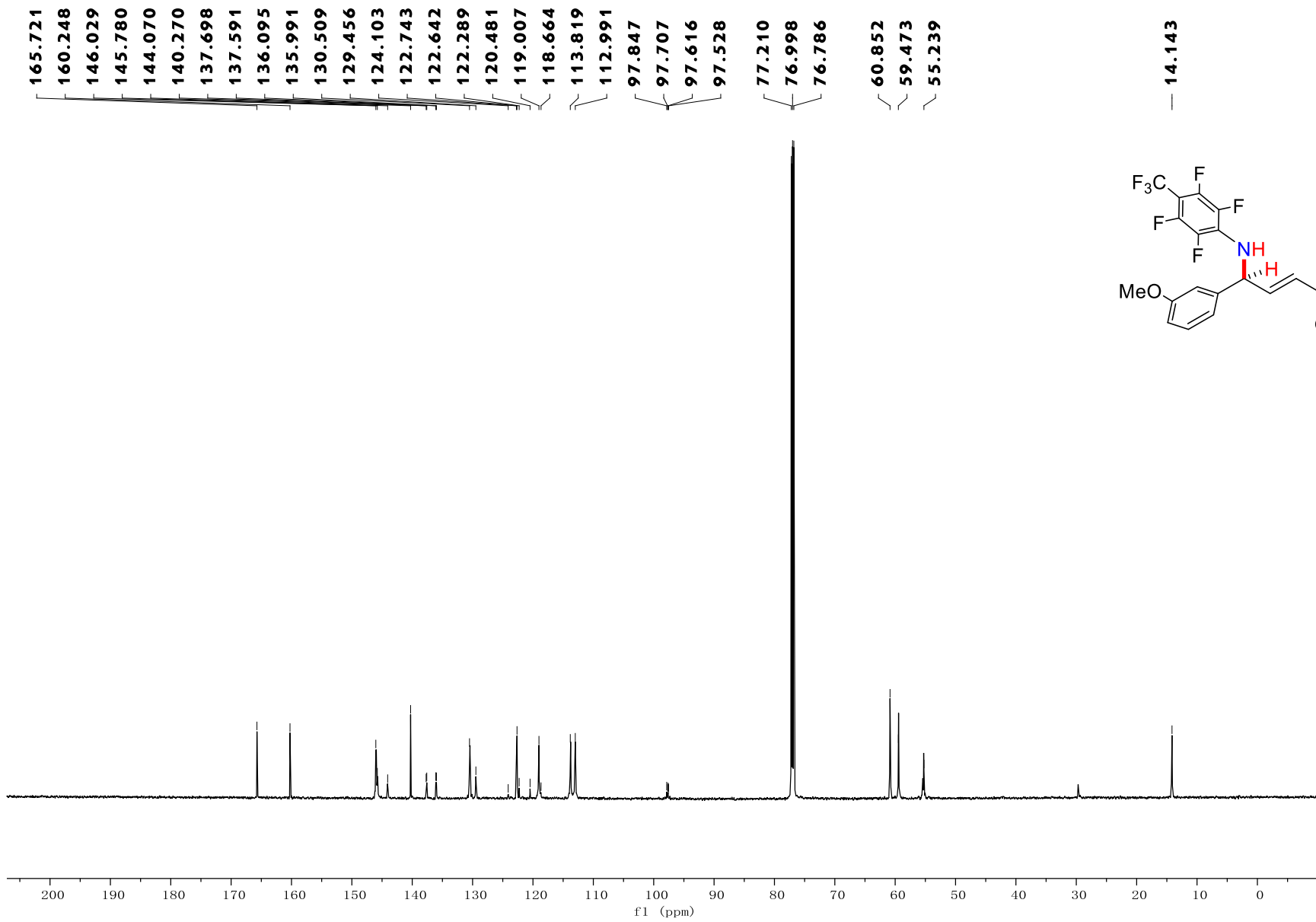
1.307
1.295
1.283

6.011
5.985

5.581
5.572
5.566
5.557



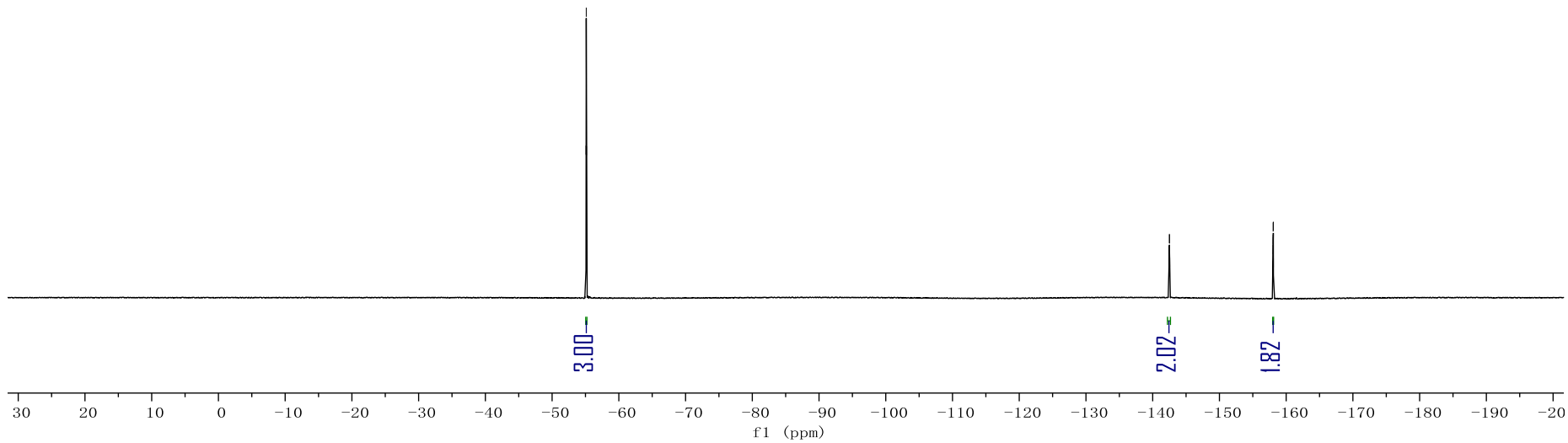
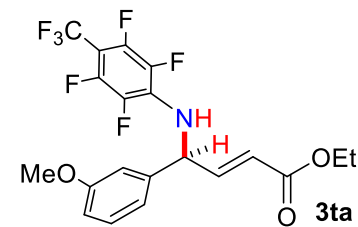
¹³C NMR



¹⁹F NMR

-55.080
-55.117
-55.154

-142.464
-142.471
-142.477
-142.501
-142.508
-142.514
-142.538
-142.544
-142.552
-158.031
-158.038
-158.045
-158.051
-158.068
-158.074
-158.082
-158.089



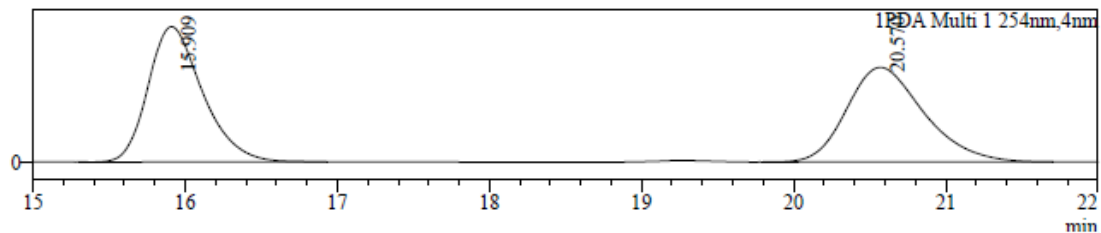
HPLC

Sample Information

Sample Name : POX-0731ODH-5%-0.8mL
Sample ID : POX-0731ODH-5%-0.8mL
Data File : POX-0731ODH-5%-0.8mL.lcd
Method File : POX-5.0%-0.8ml.lcm

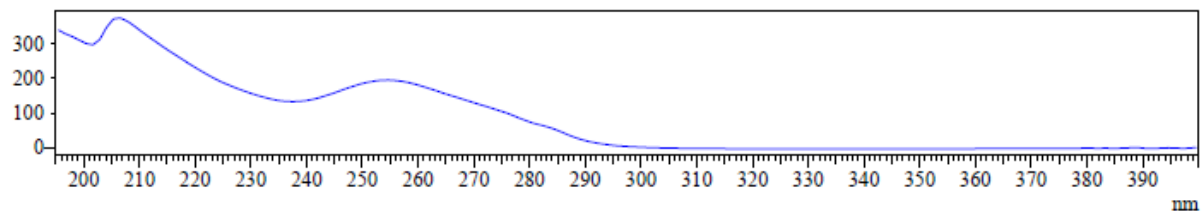
Chromatogram

mAU



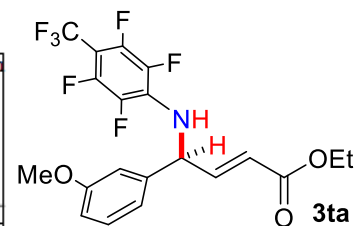
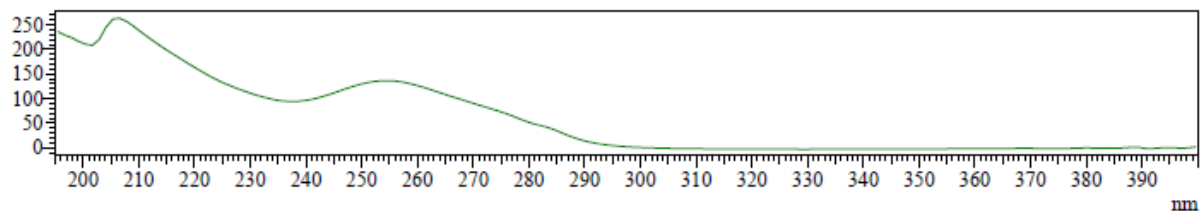
UV Spectrum
Retention time = 15.909

mAU



UV Spectrum
Retention time = 20.570

mAU



Peak Table

PDA Ch1 254nm

Peak#	Ret. Time	Area	Area%
1	15.909	4776937	50.343
2	20.570	4711898	49.657
Total		9488835	100.000

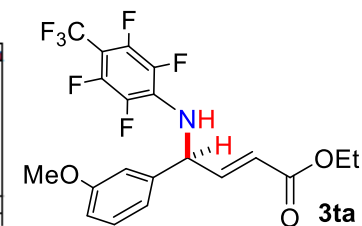
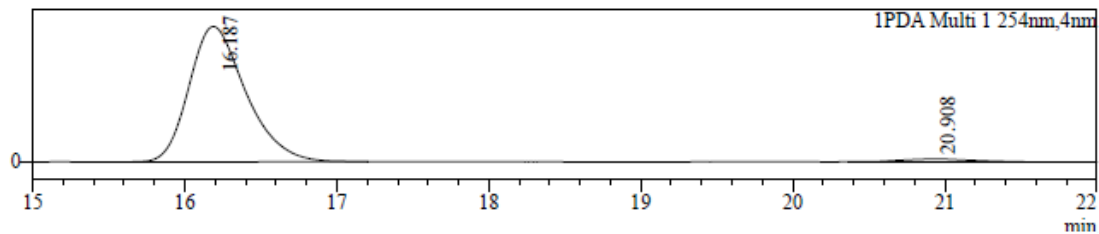
HPLC

Sample Information

Sample Name : P0X-0732-ODH-5%-0.8mL
Sample ID : P0X-0732-ODH-5%-0.8mL
Data File : P0X-0732-ODH-5%-0.8mL.lcd
Method File : P0X-5.0%-0.8ml.lcm

Chromatogram

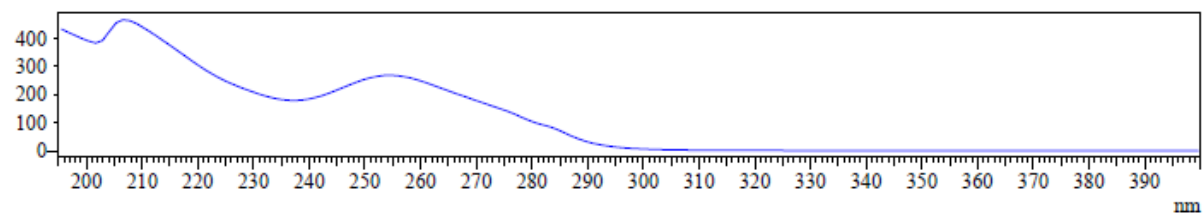
mAU



UV Spectrum

Retention time = 16.187

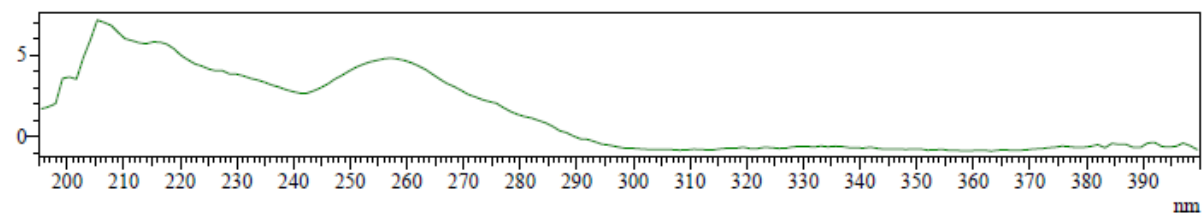
mAU



UV Spectrum

Retention time = 20.908

mAU



Peak Table

PDA Ch1 254nm

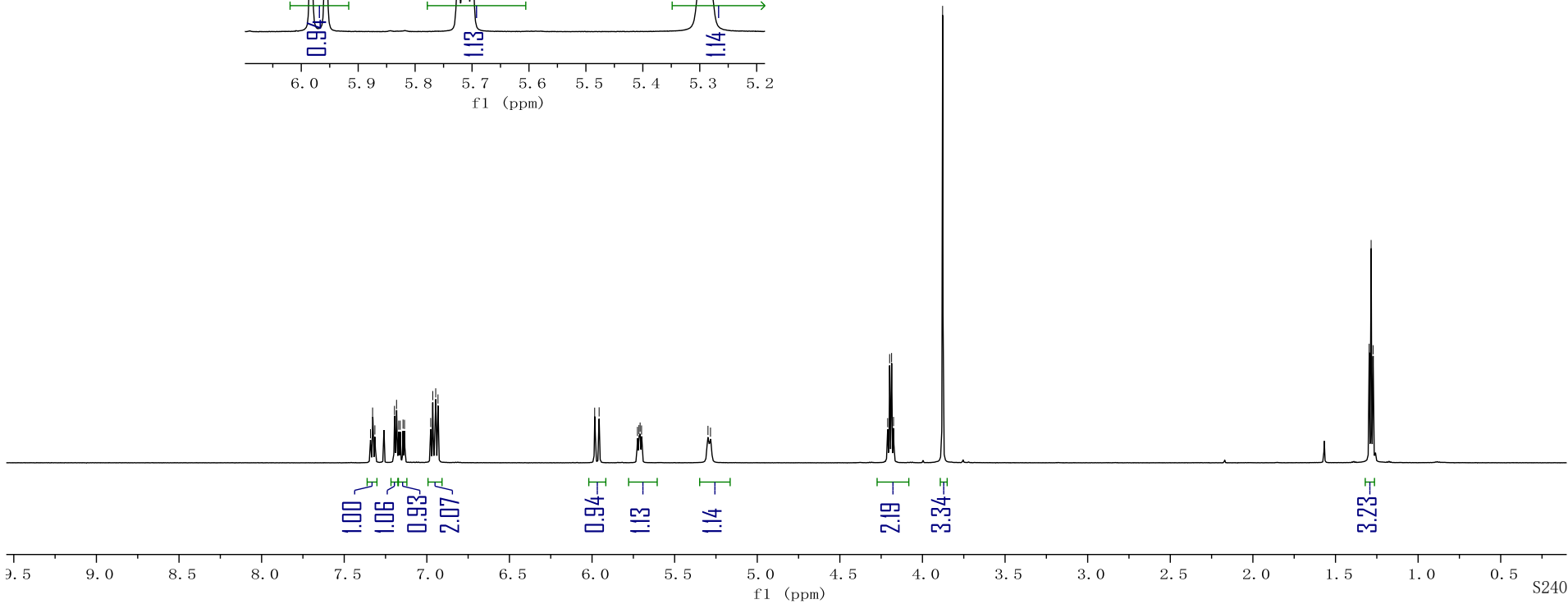
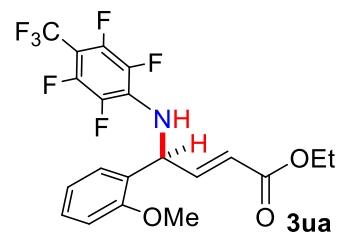
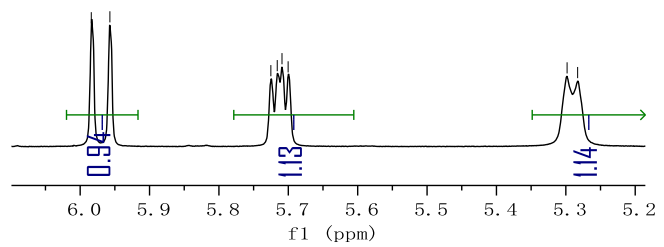
Peak#	Ret. Time	Area	Area%
1	16.187	6937205	97.368
2	20.908	187500	2.632
Total		7124705	100.000

¹H NMR

7.341
7.328
7.314
7.196
7.183
7.170
7.161
7.144
7.135
6.976
6.964
6.948
6.946
6.932
5.984
5.957
5.725
5.716
5.709
5.700
5.298
5.283
4.211
4.199
4.187
4.175
3.878

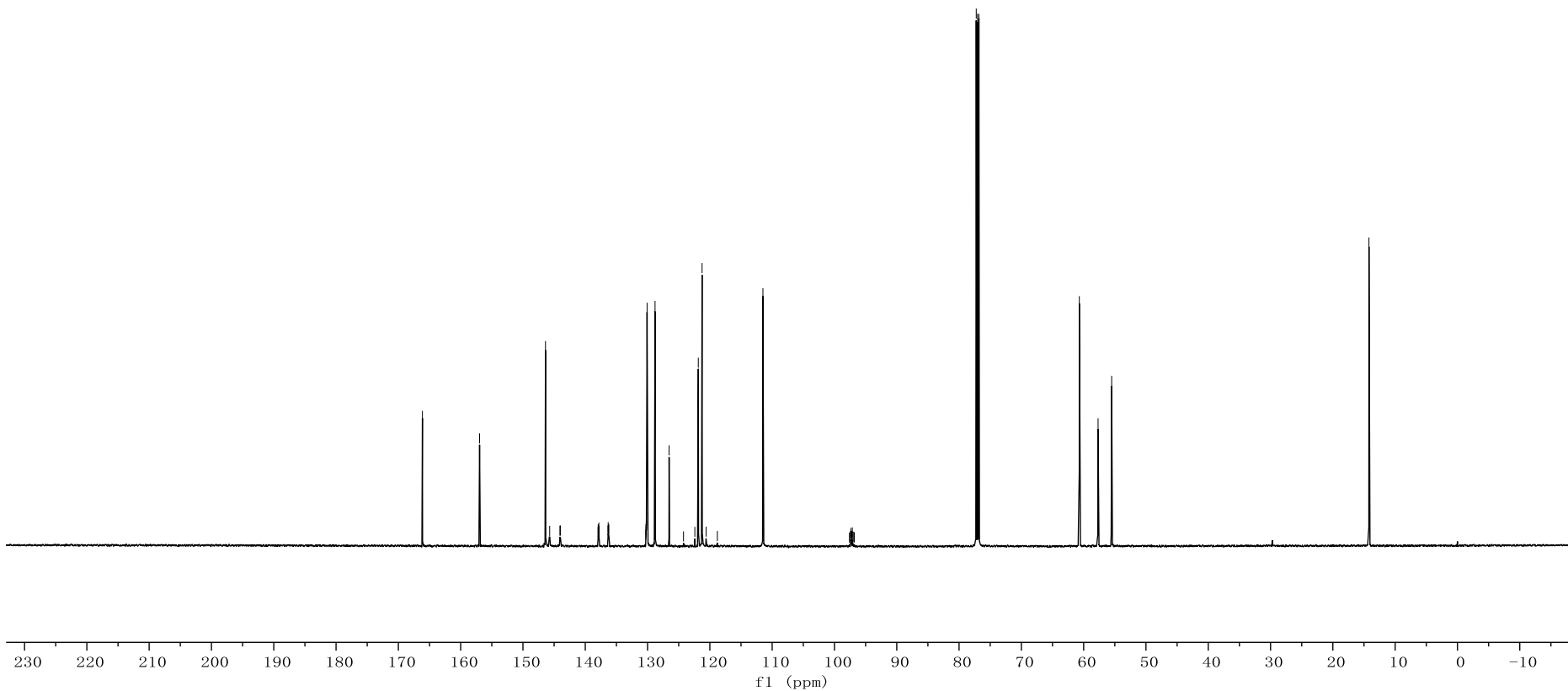
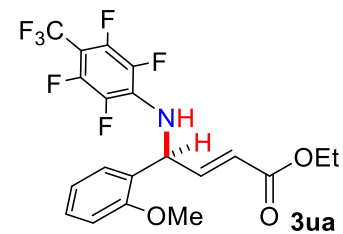
5.984
5.957
5.725
5.716
5.709
5.700
5.298
5.283

1.296
1.284
1.272

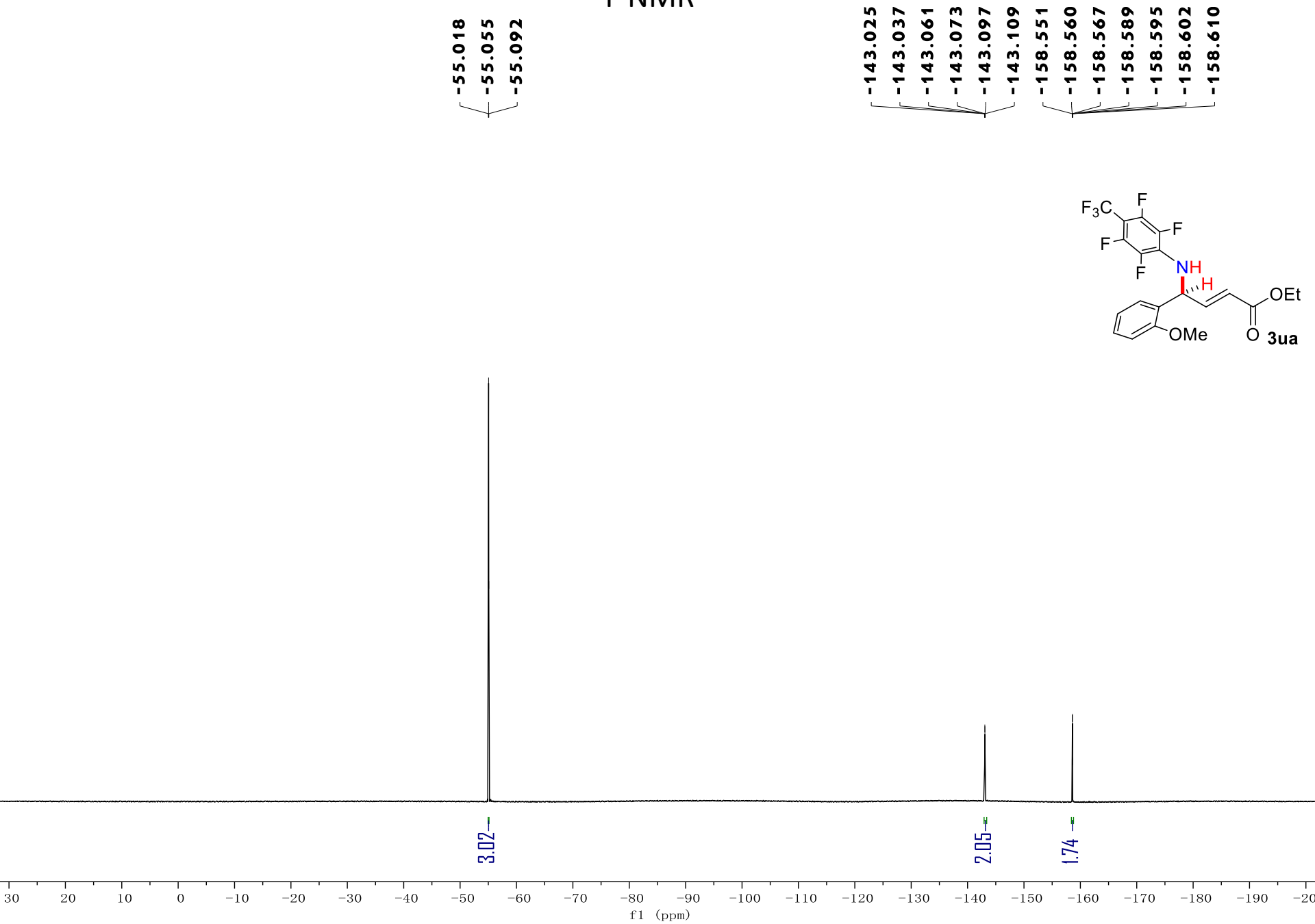


¹³C NMR

166.138
156.963
146.383
145.710
144.074
143.979
137.954
137.922
137.886
137.849
137.816
137.782
136.357
136.321
136.292
136.254
136.224
130.283
130.257
130.233
130.209
130.187
130.164
130.077
128.796
126.542
122.414
121.855
121.249
120.608
118.798
111.484
97.453
97.364
97.221
97.131
77.243
77.031
76.818
60.697
57.719
57.685
57.651
55.505
14.208



¹⁹F NMR



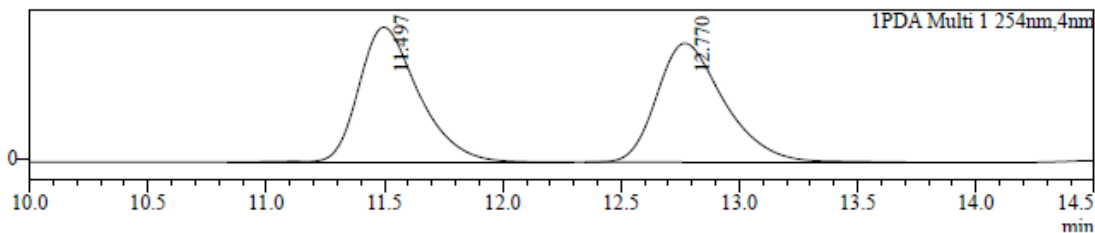
HPLC

Sample Information

Sample Name : P0X-0676-ODH-5%-0.8mL
Sample ID : P0X-0676-ODH-5%-0.8mL
Data File : P0X-0676-ODH-5%-0.8mL.lcd
Method File : P0X-5.0%-0.8ml.lcm

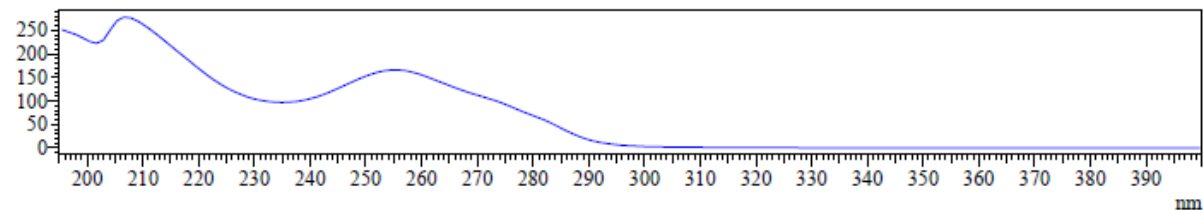
Chromatogram

mAU



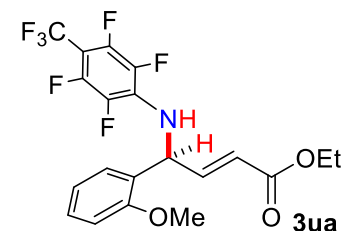
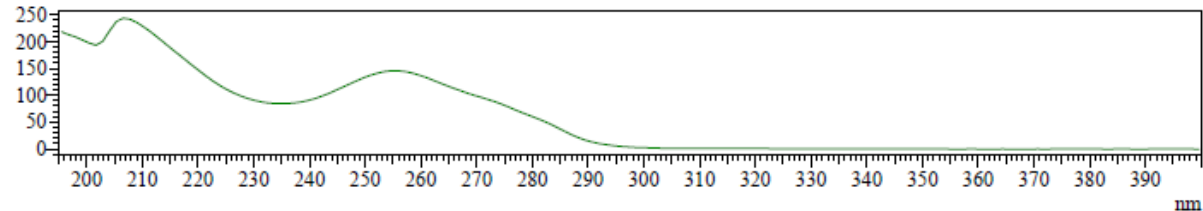
UV Spectrum
Retention time = 11.497

mAU



UV Spectrum
Retention time = 12.770

mAU



Peak Table

PDA Ch1 254nm

Peak#	Ret. Time	Area	Area%
1	11.497	2956116	50.097
2	12.770	2944648	49.903
Total		5900764	100.000

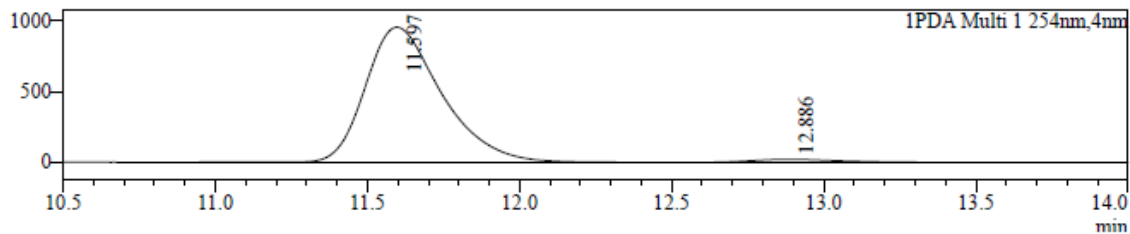
HPLC

Sample Information

Sample Name : P0X-0677ODH-5%-0.8mL
Sample ID : P0X-0677ODH-5%-0.8mL
Data File : P0X-0677-ODH-5%-0.8mL.lcd
Method File : P0X-5.0%-0.8ml.lcm

Chromatogram

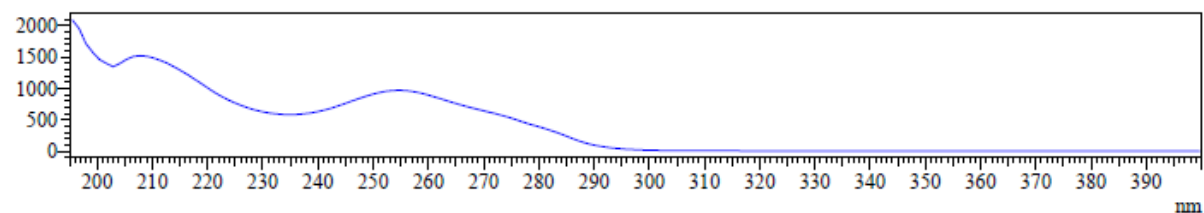
mAU



UV Spectrum

Retention time = 11.597

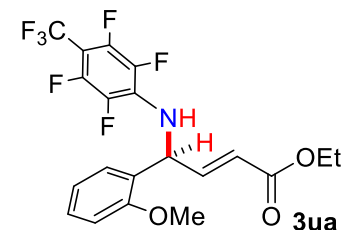
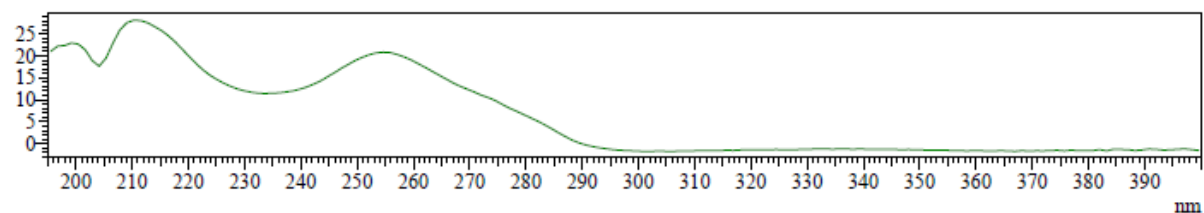
mAU



UV Spectrum

Retention time = 12.886

mAU

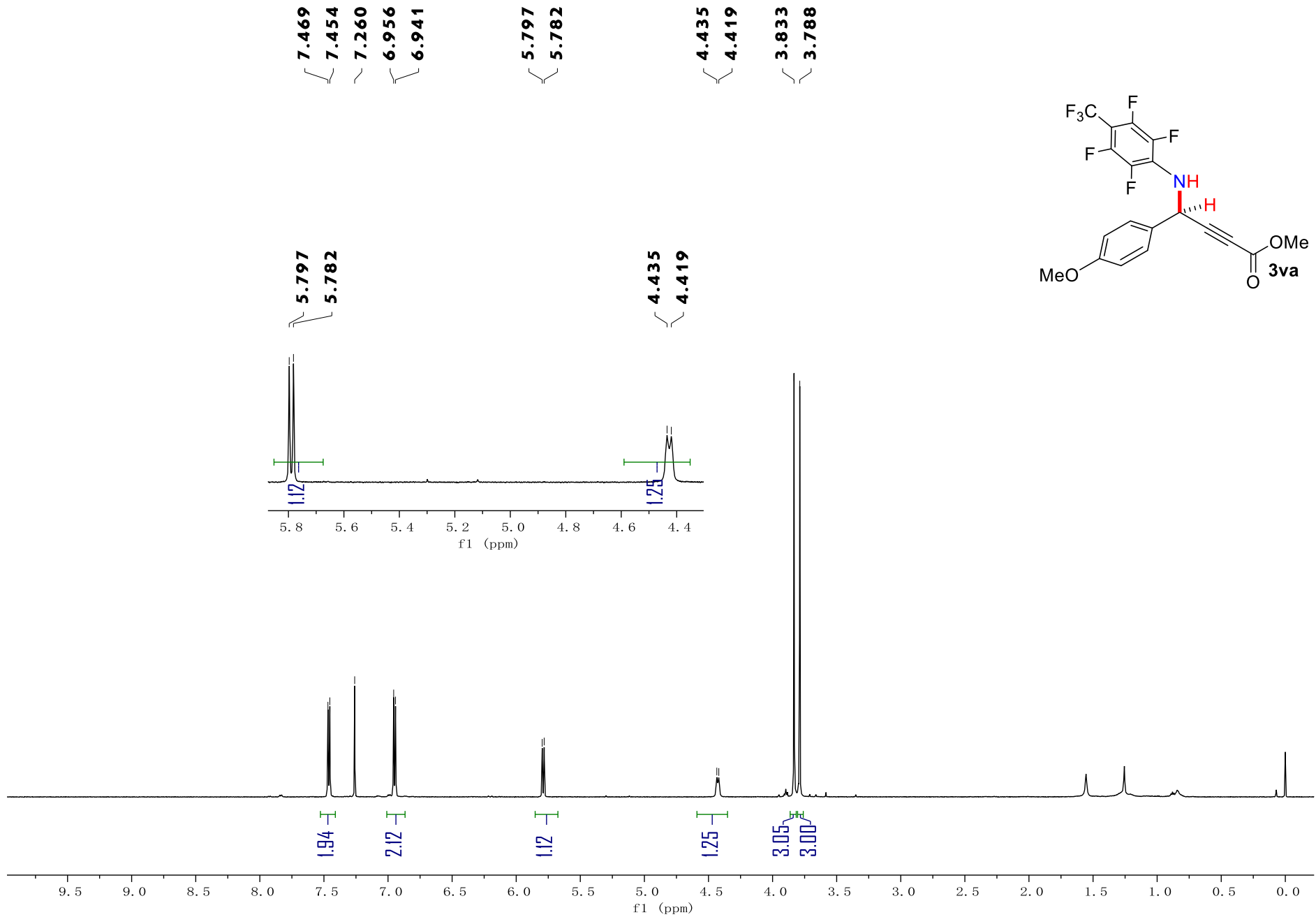


Peak Table

PDA Ch1 254nm

Peak#	Ret. Time	Area	Area%
1	11.597	16688022	97.412
2	12.886	443332	2.588
Total		17131354	100.000

^1H NMR

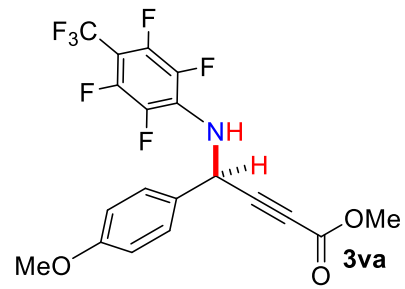


^{13}C NMR

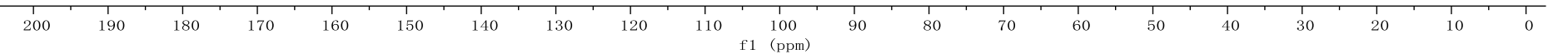
163.019
156.014
148.353
148.325
148.272
146.673
146.574
140.972
140.868
139.368
139.302
131.495
131.419
131.364
131.338
131.163
130.844
126.645
124.853
123.053
117.257
101.764
101.534
101.380

87.019
80.197
79.853
79.841
79.430
79.429

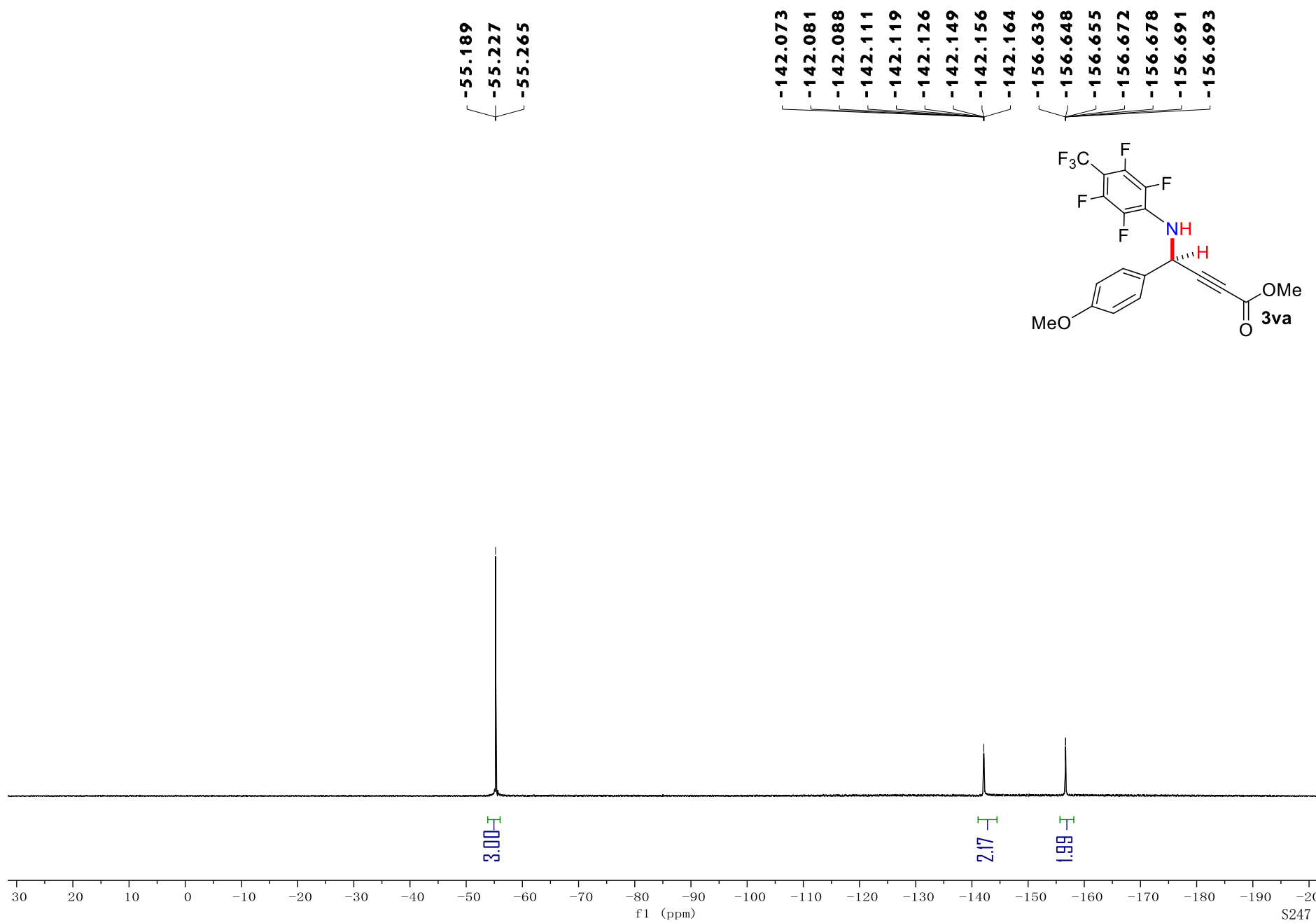
58.083
55.511
53.025



grease

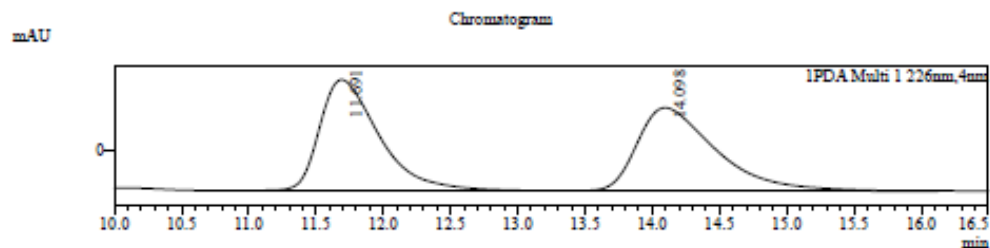


^{19}F NMR

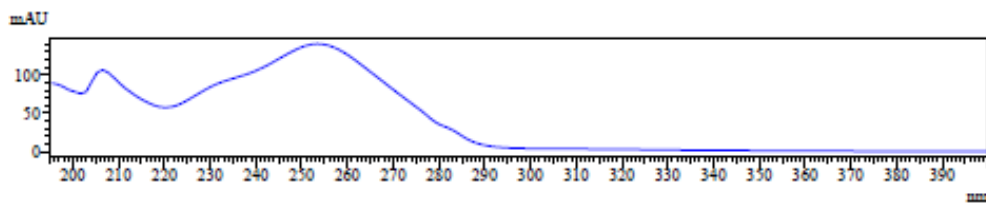


HPLC

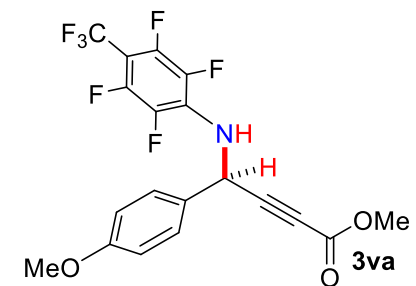
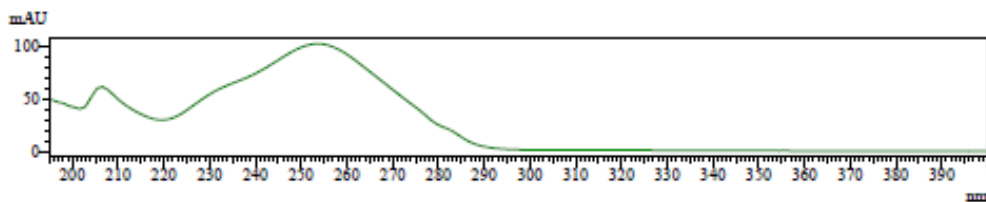
Sample Information
Sample Name : P0X-0587-ASH-10%0.8mL
Sample ID : P0X-0587-ASH-10%0.8mL
Data File : P0X-0587-ASH-10%0.8mL.lcd
Method File : YW-10%-0.8ml.lcm



UV Spectrum
Retention time = 11.691



UV Spectrum
Retention time = 14.098

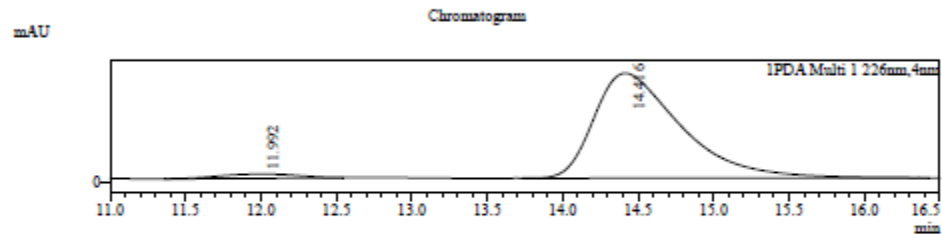


Peak Table

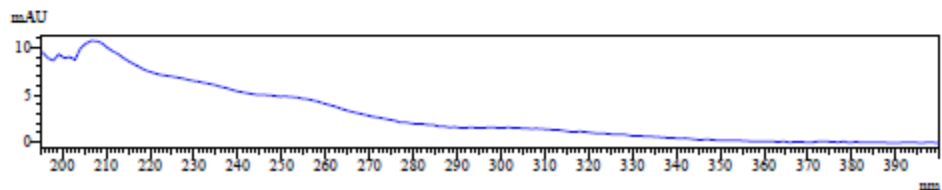
Peak#	Ret. Time	Area	Area%
1	11.691	3451401	50.530
2	14.098	3379031	49.470
Total		6830432	100.000

HPLC

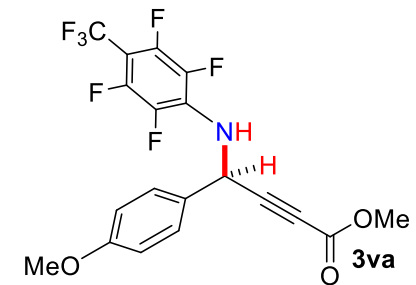
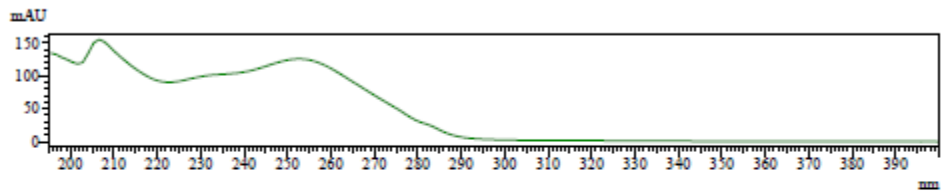
Sample Information
Sample Name : POX-0588-ASH-10%0.8mL
Sample ID : POX-0588-ASH-10%0.8mL
Data File : POX-0588-ASH-10%0.8mL.lcd
Method File : YW-10%-0.8ml.lcm



UV Spectrum
Retention time = 11.992



UV Spectrum
Retention time = 14.416



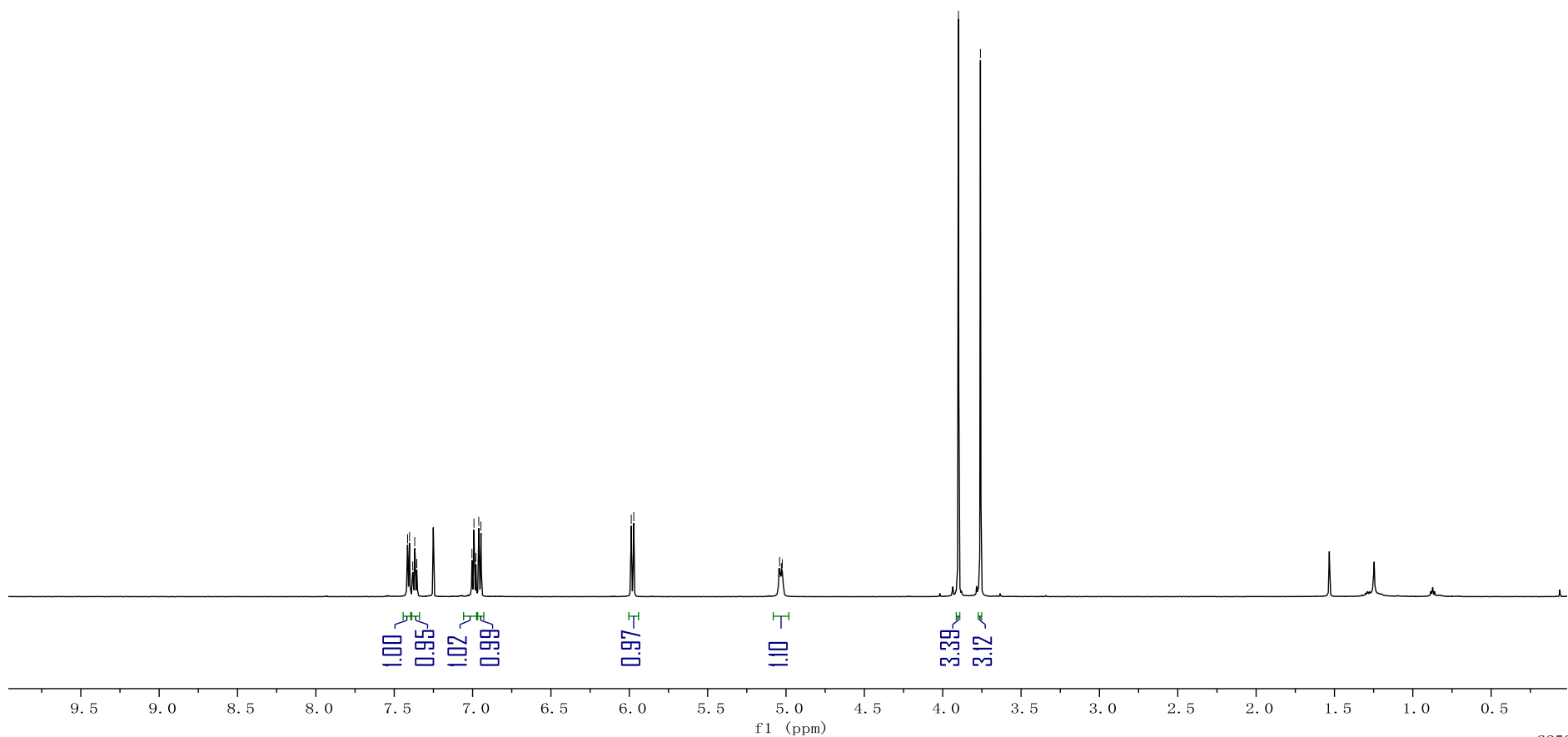
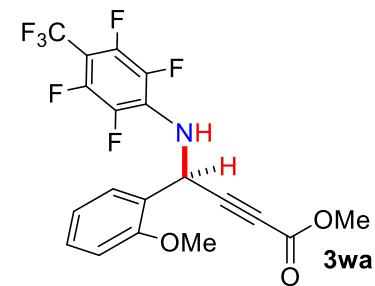
Peak Table

PDA Ch1 226nm

Peak#	Ret. Time	Area	Area%
1	11.992	131952	3.505
2	14.416	3632225	96.495
Total		3764177	100.000

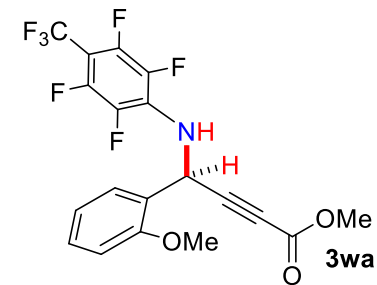
¹H NMR

7.415
7.402
7.382
7.369
7.356
7.004
6.991
6.979
6.960
6.947
5.988
5.972
5.047
5.041
5.030
5.024
3.900
3.759



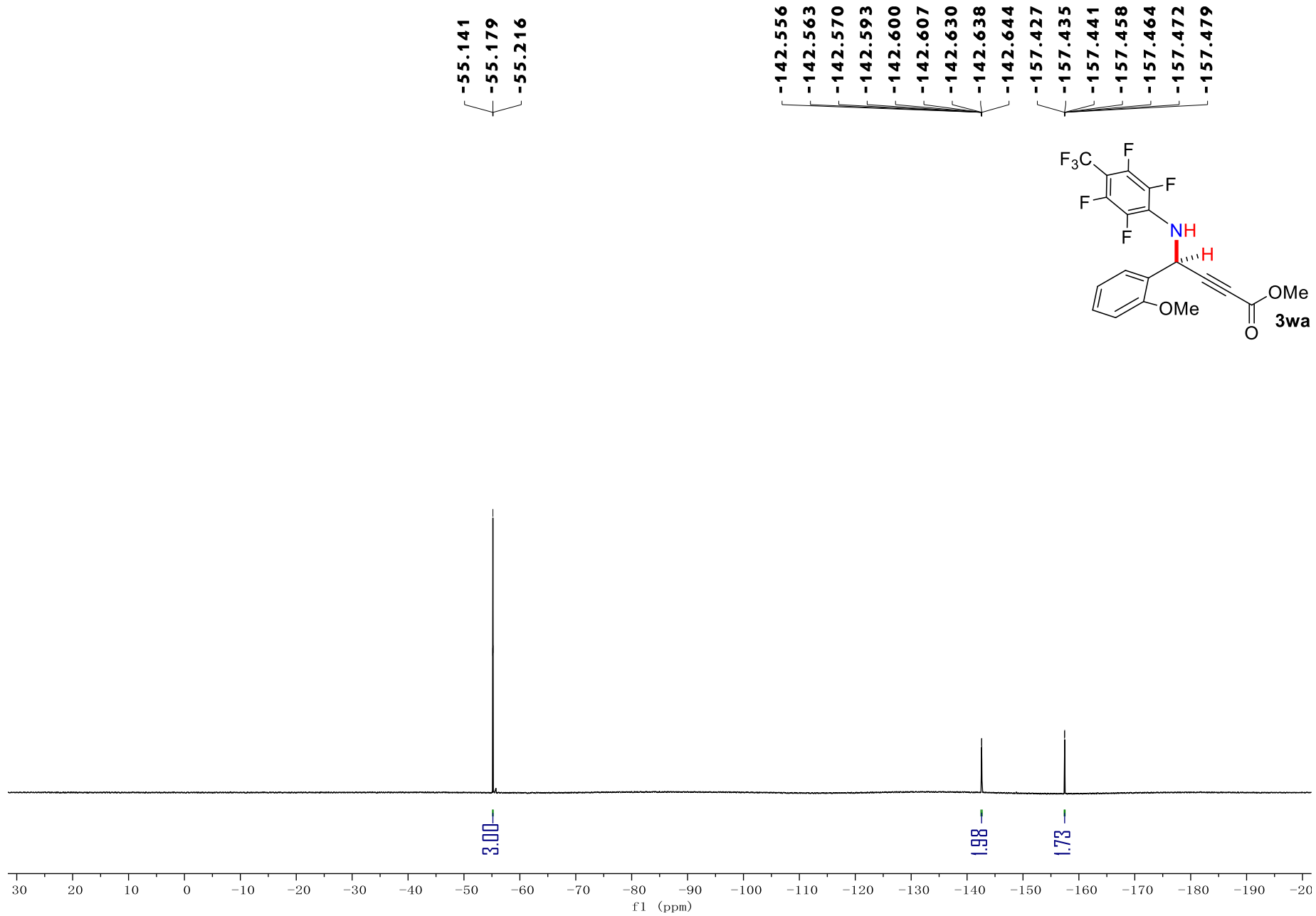
¹³C NMR

156.920
153.579
145.631
143.931
138.272
138.164
136.666
136.561
130.813
129.281
129.254
129.181
129.105
129.081
128.701
124.482
124.086
122.278
121.081
120.465
111.519
98.566
98.566
98.335
98.117
84.891
77.212
77.000
76.789
76.080
55.671
52.781
47.466



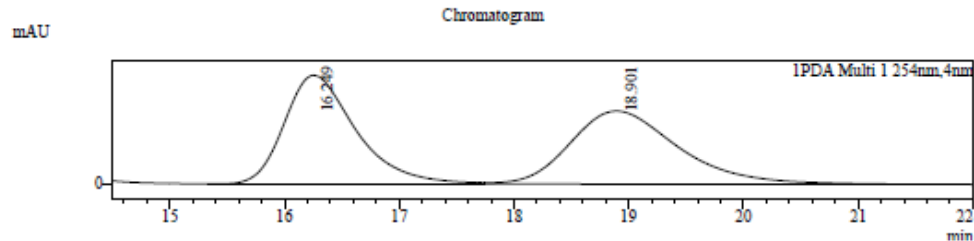
200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10
f1 (ppm)

¹⁹F NMR

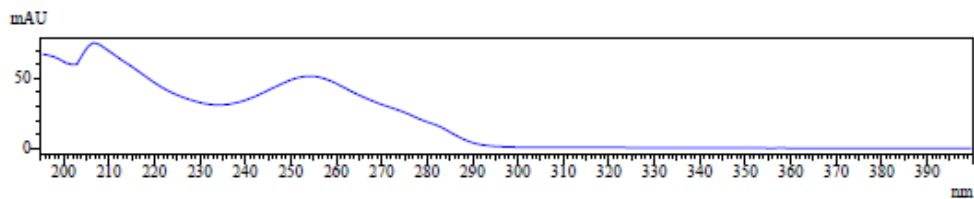


HPLC

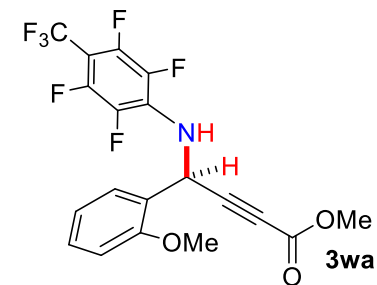
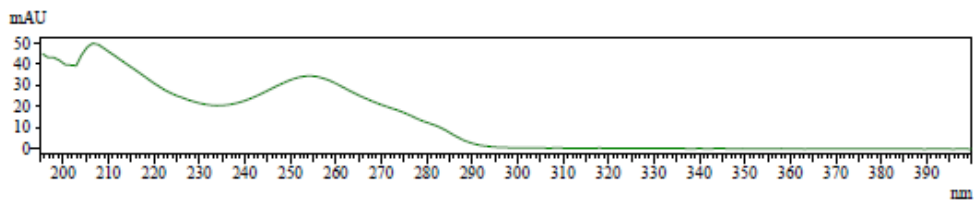
Sample Information
 Sample Name : P0X-0602-OJH10%-0.8mL
 Sample ID : P0X-0602-OJH10%-0.8mL
 Data File : P0X-0602-OJH10%-0.8mL.lcd
 Method File : YW-10%-0.8mL.lcm



UV Spectrum
 Retention time = 16.249



UV Spectrum
 Retention time = 18.901



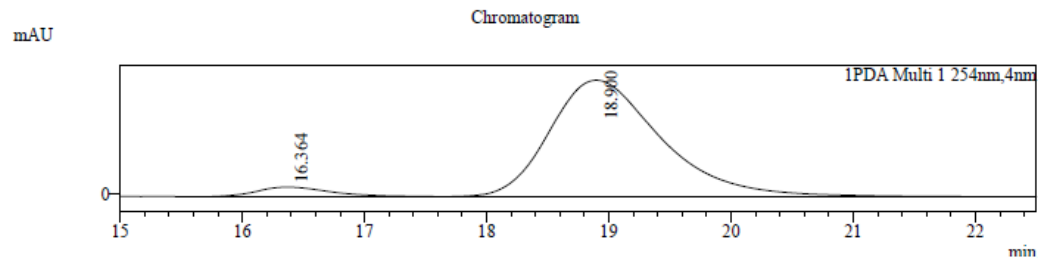
Peak Table

PDA Ch1 254nm

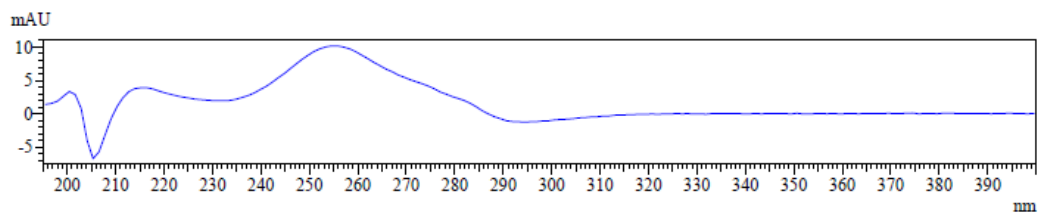
Peak#	Ret. Time	Area	Area%
1	16.249	2206862	49.676
2	18.901	2235673	50.324
Total		4442536	100.000

HPLC

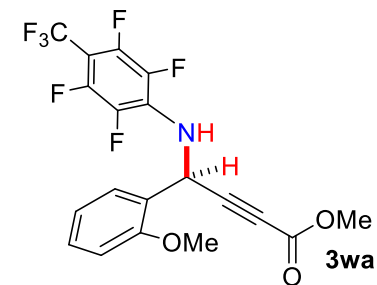
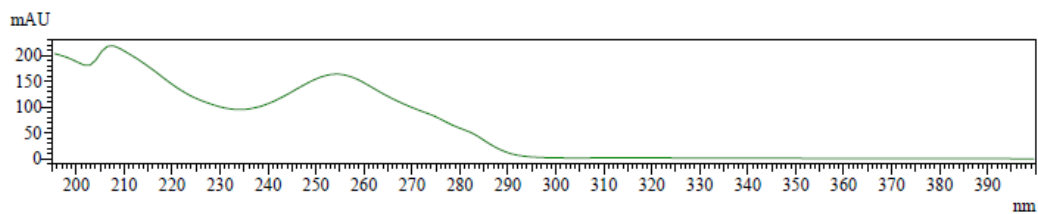
Sample Information
Sample Name : P0X-0603-OJH-10%-0.8mL
Sample ID : P0X-0603-OJH-10%-0.8mL
Data File : P0X-0603-OJH-10%-0.8mL.lcd
Method File : YW-10%-0.8ml.lcm



UV Spectrum
Retention time = 16.364



UV Spectrum
Retention time = 18.900



Peak Table

PDA Ch1 254nm

Peak#	Ret. Time	Area	Area%
1	16.364	582768	5.218
2	18.900	10586414	94.782
Total		11169182	100.000

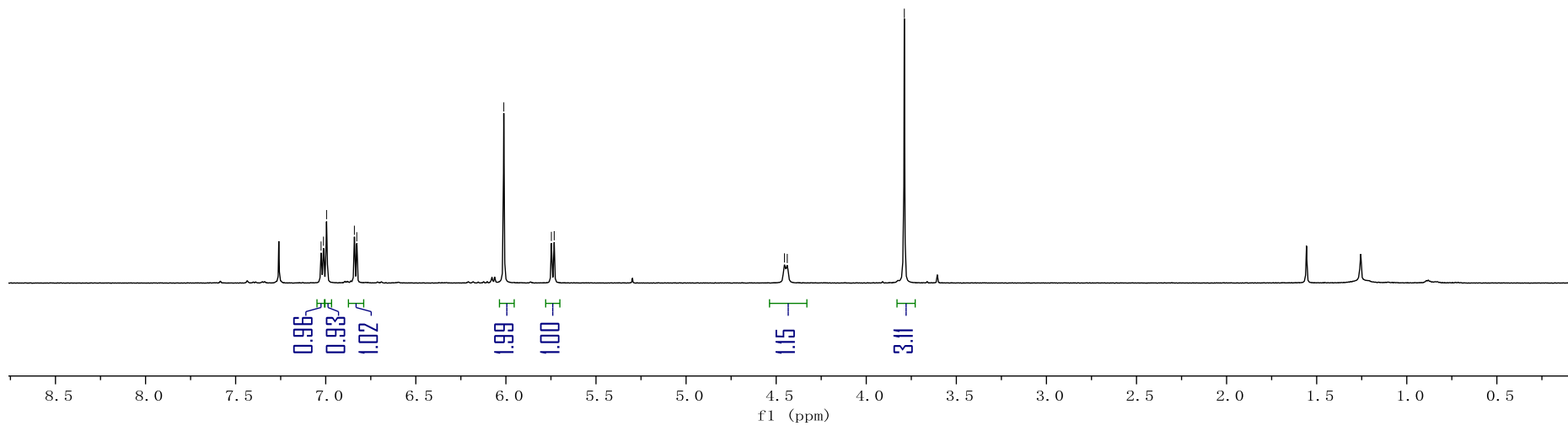
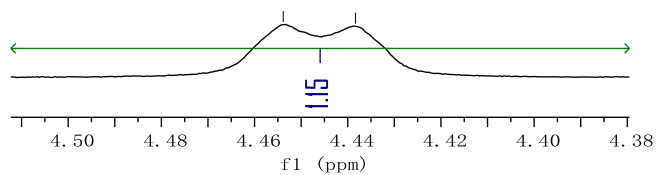
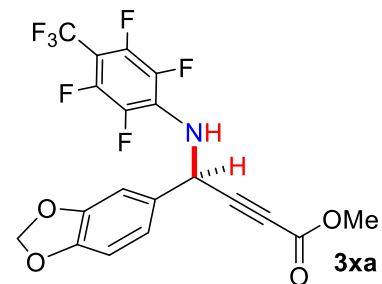
^1H NMR

7.026
7.012
6.995
6.841
6.828

6.012
5.748
5.732

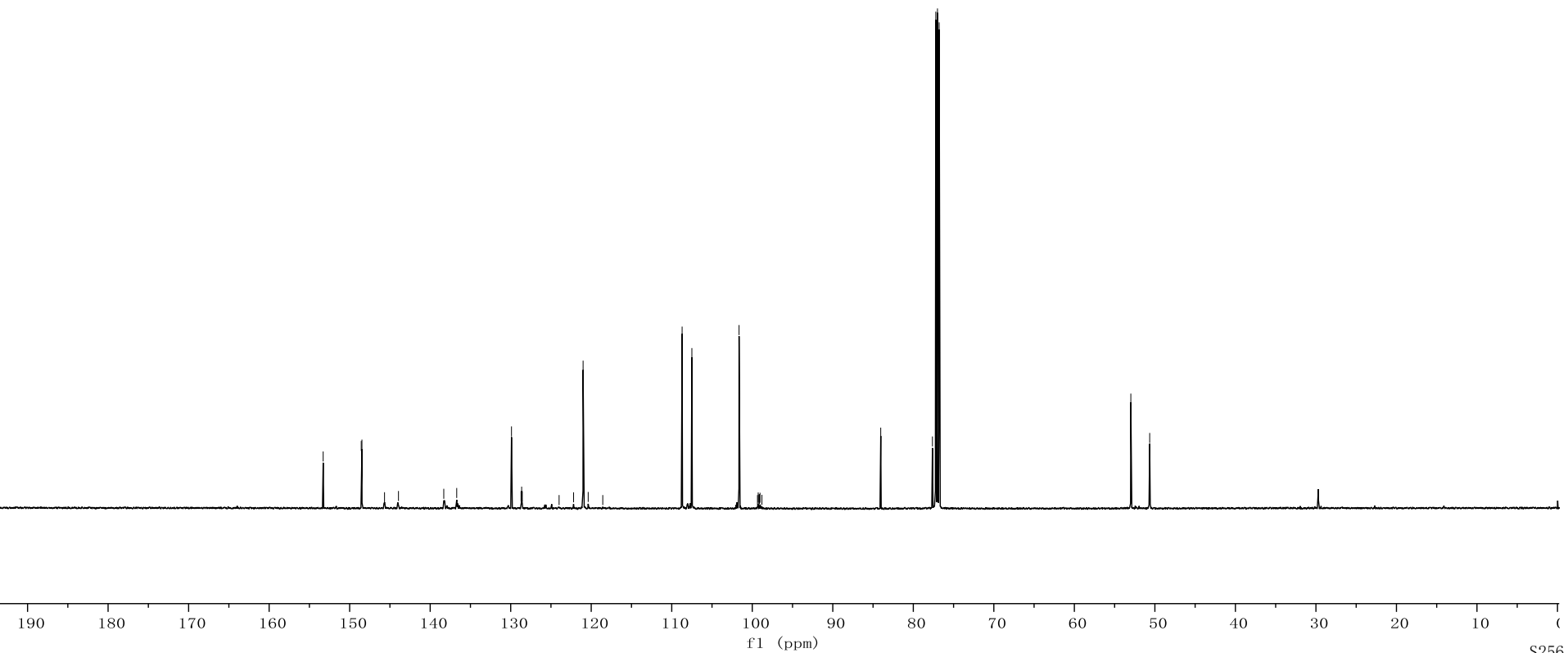
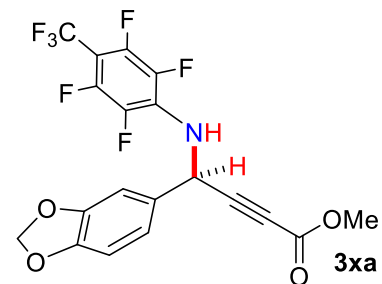
4.454
4.438

3.789



^{13}C NMR

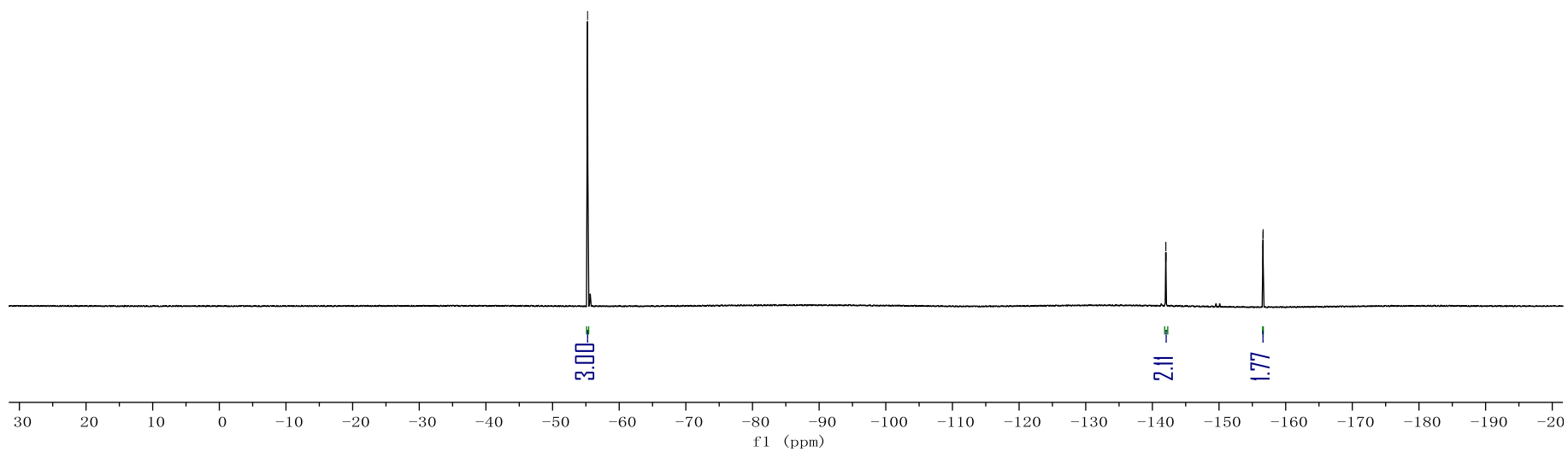
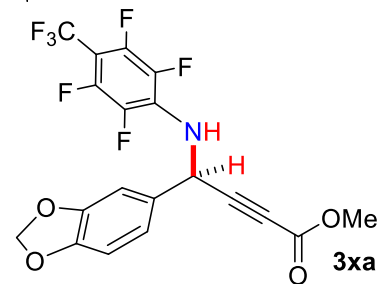
153.289
148.563
148.476
145.670
143.937
138.311
136.705
129.902
128.715
128.660
128.638
128.615
128.562
123.990
122.190
121.009
120.378
118.567
108.711
108.708
107.500
101.641
99.353
99.264
99.176
99.118
99.033
98.800
84.046
77.625
77.211
77.000
76.788
52.987
50.644



^{19}F NMR

-55.215
-55.252
-55.289

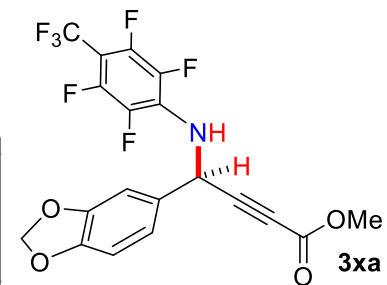
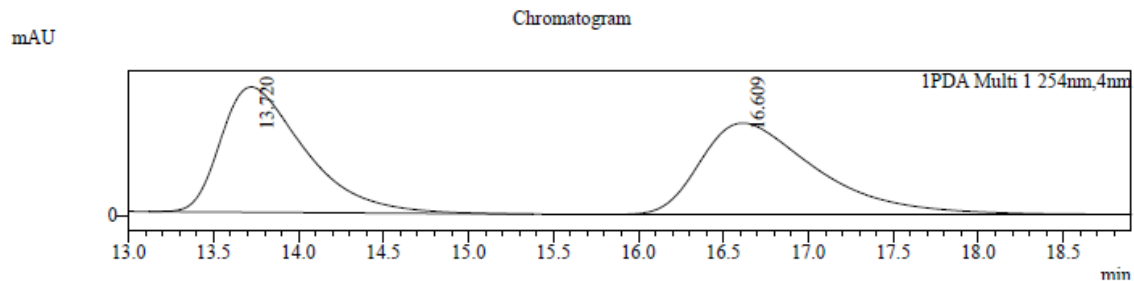
-141.987
-142.010
-142.025
-142.048
-156.586
-156.592
-156.609
-156.615



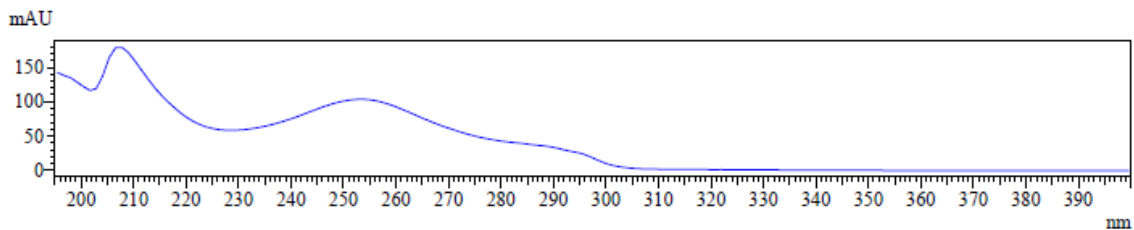
HPLC

Sample Information

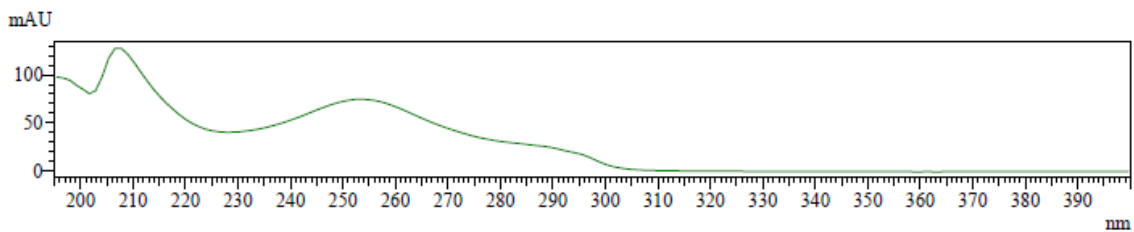
Sample Name : P0X-0627-ASH-10%-0.8mL
Sample ID : P0X-0627-ASH-10%-0.8mL
Data File : P0X-0627-ASH-10%-0.8mL.lcd
Method File : P0X-10%-0.8ml.lcm



UV Spectrum
Retention time = 13.720



UV Spectrum
Retention time = 16.609



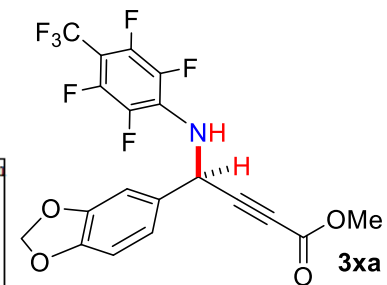
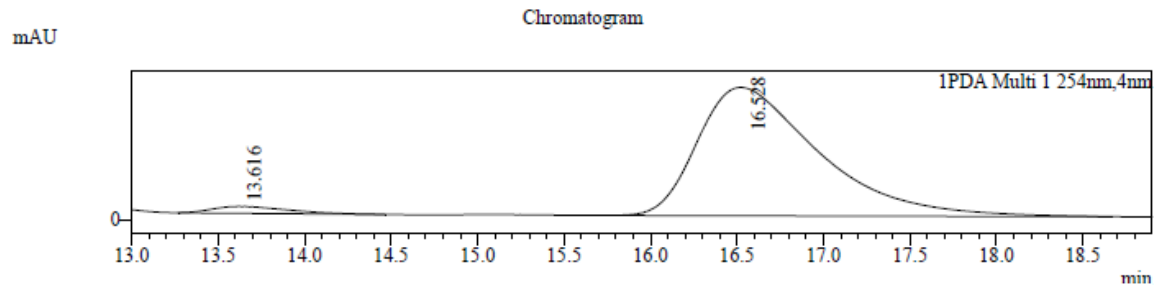
Peak Table

PDA Ch1 254nm

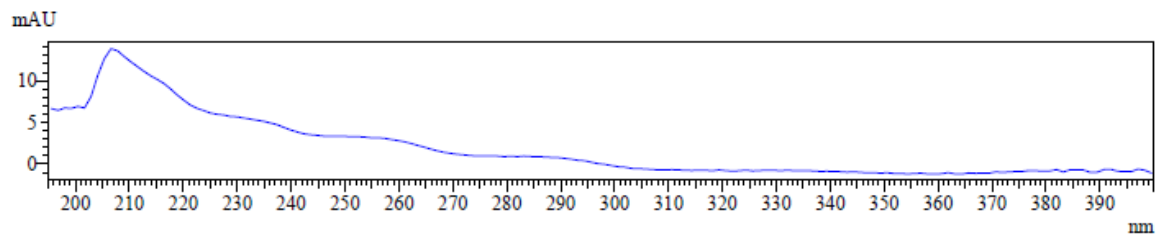
Peak#	Ret. Time	Area	Area%
1	13.720	3433019	49.735
2	16.609	3469604	50.265
Total		6902624	100.000

HPLC

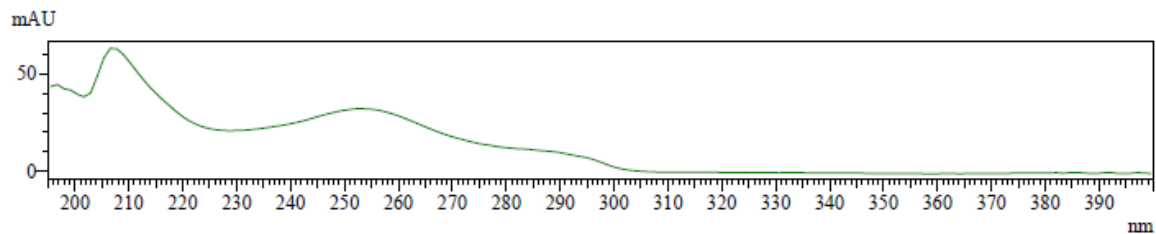
Sample Information
Sample Name : POX-0628X-ASH-10%-0.8mL
Sample ID : POX-0628X-ASH-10%-0.8mL
Data File : POX-0628X-ASH-10%-0.8mL.lcd
Method File : POX-10%-0.8ml.lcm



UV Spectrum
Retention time = 13.616



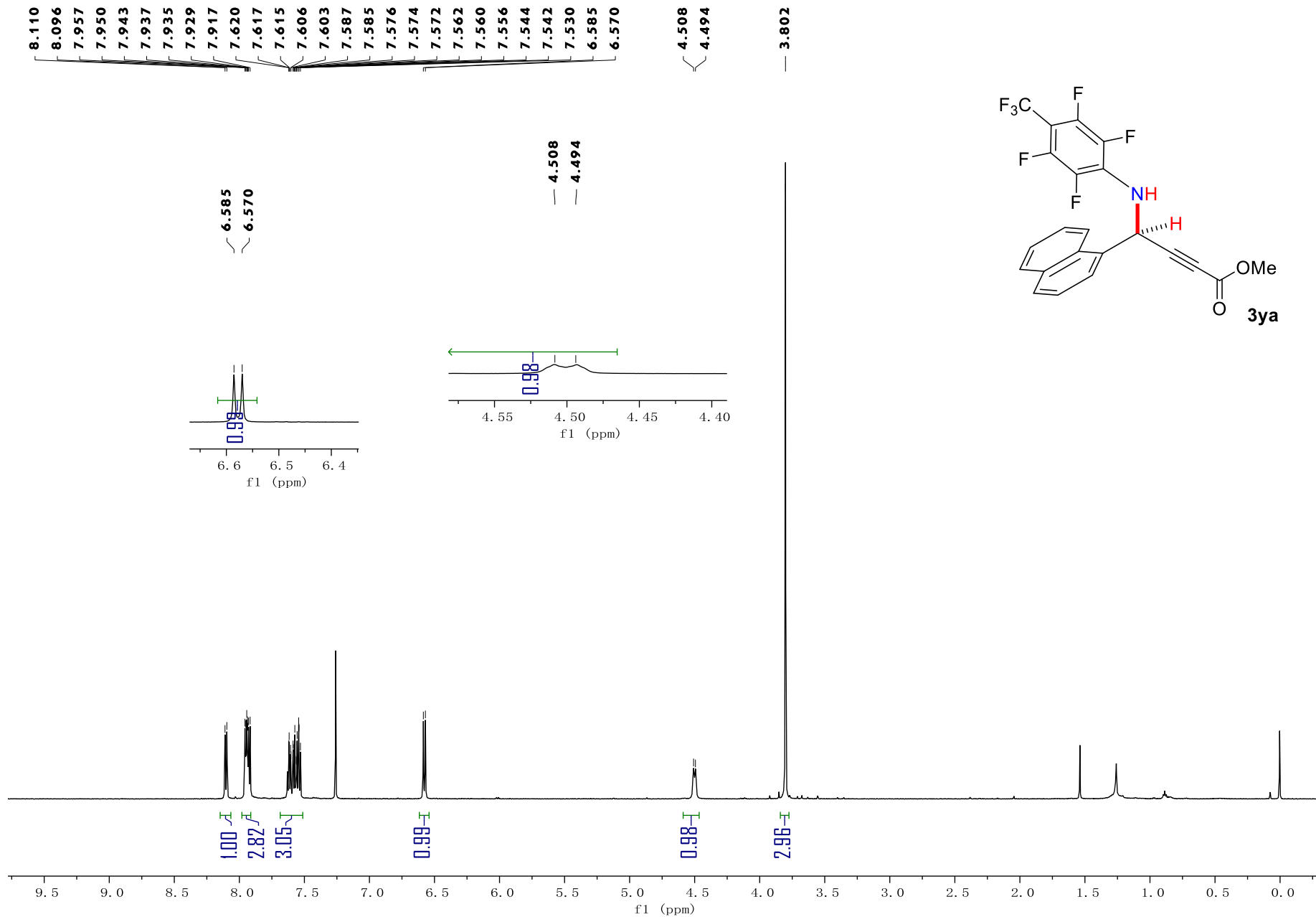
UV Spectrum
Retention time = 16.528



Peak Table

Peak#	Ret. Time	Area	Area%
1	13.616	49819	3.221
2	16.528	1496756	96.779
Total		1546574	100.000

^1H NMR



¹³C NMR

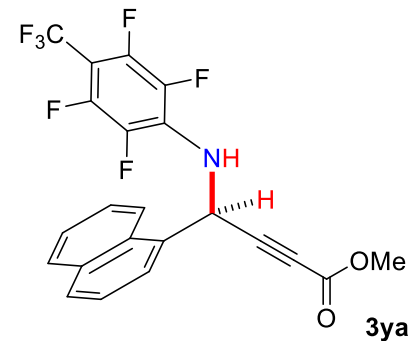
155.983
148.404
146.665
141.116
139.405
136.790
133.681
133.318
132.893
131.838
131.549
131.474
131.397
130.044
129.139
128.645
127.953
125.307
124.864
123.051
102.000
101.768
101.679

86.652

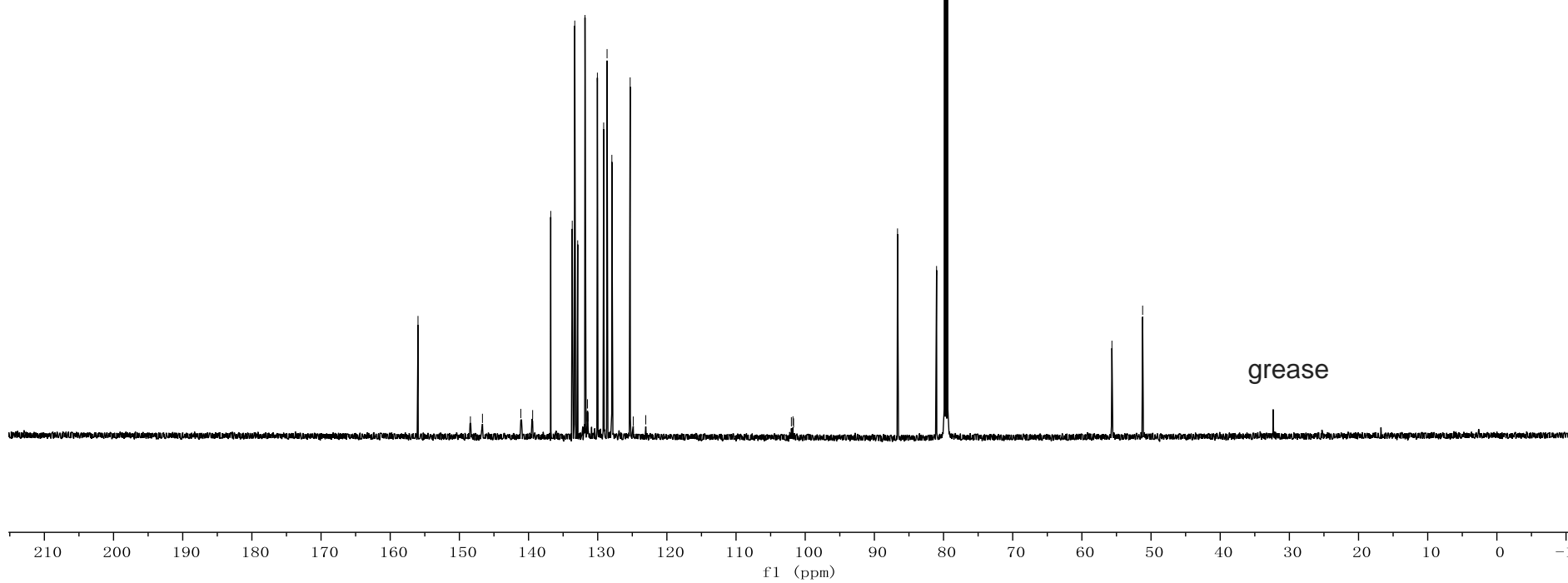
81.007

55.654

51.215



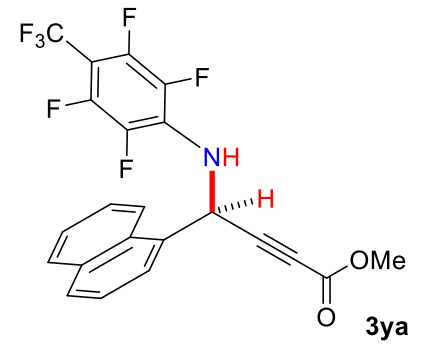
grease



¹⁹F NMR

-55.183
-55.220
-55.257

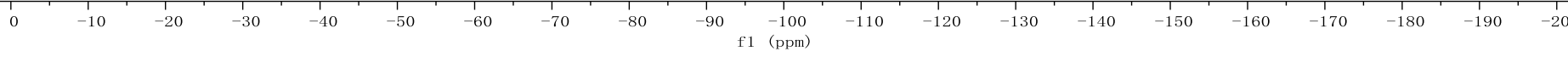
-141.862
-141.885
-141.900
-141.923
-156.785
-156.793
-156.799
-156.816
-156.822
-156.830



3.00

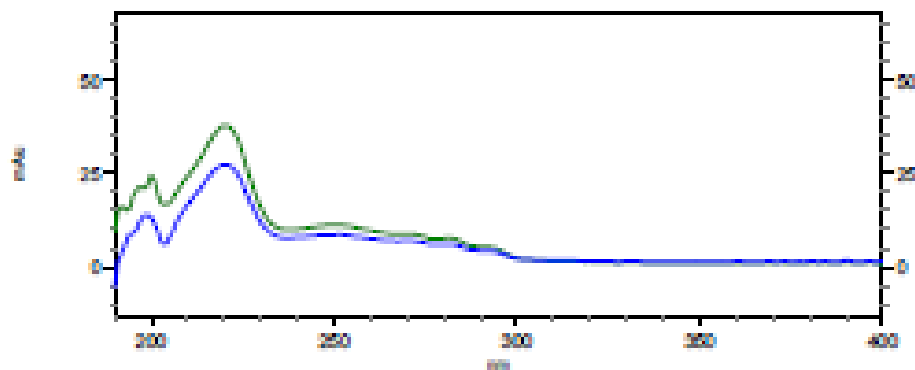
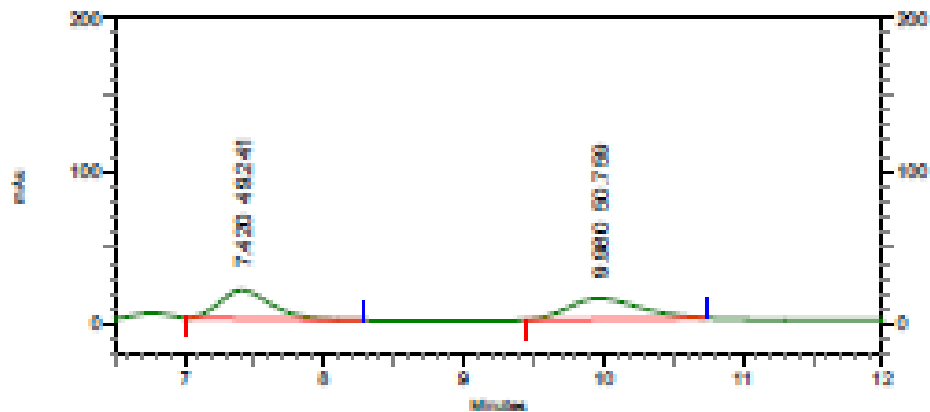
2.27

1.74



HPLC

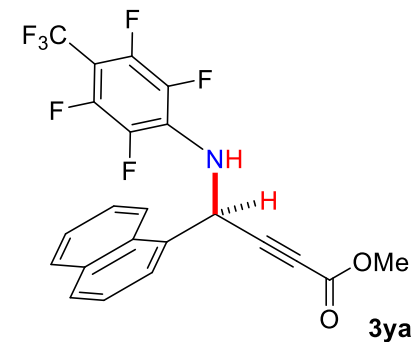
C:\ESStart\Projects\Default\Data\F0X0891ASH-56-0.8mL
C:\Documents and Settings\zhang\Desktop\WCL\Method.net



4: 250 nm, 4
nm Results

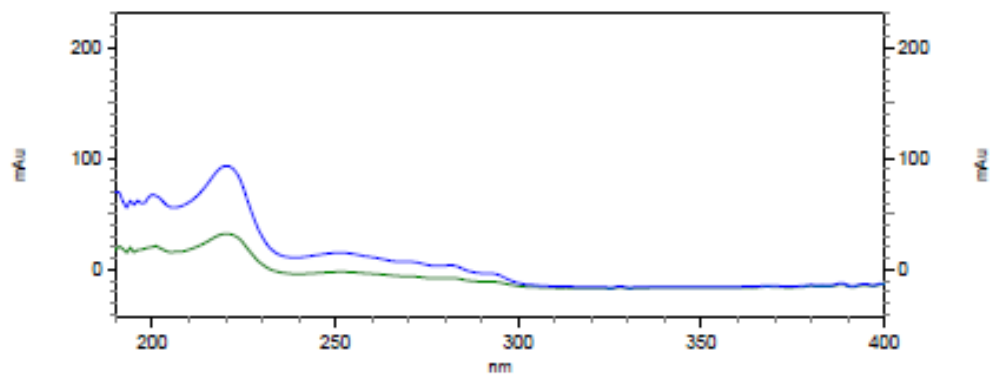
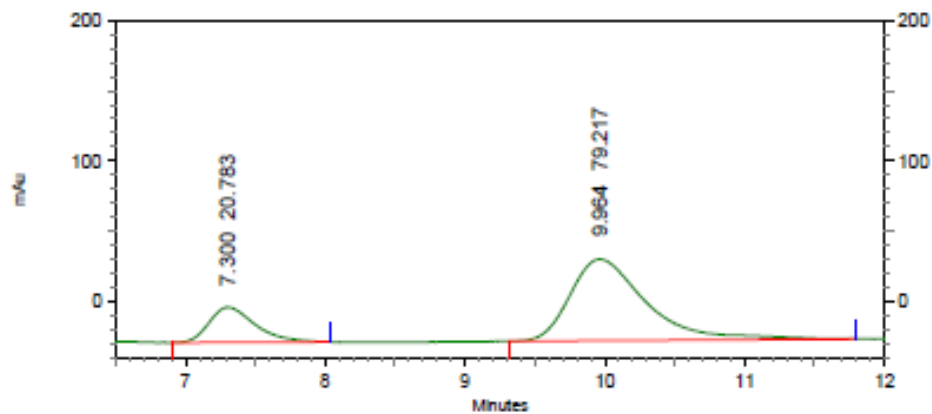
PK #	Retention Time	Area Percent
1	7.420	49.241
2	9.980	50.759

Totals	100.000
--------	---------



HPLC

C:\EZStart\Projects\Default\Data\POX0608ASH-5%-0.8mL
C:\Documents and Settings\zhang\Desktop\WCL\Method.met

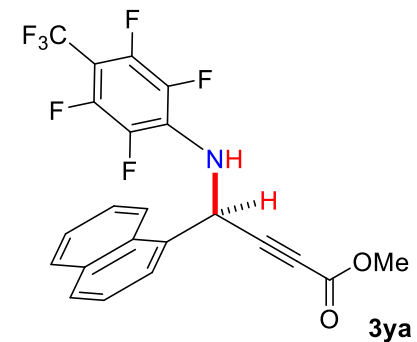


4: 250 nm, 4

nm Results

Pk #	Retention Time	Area Percent
1	7.300	20.783
2	9.964	79.217

Totals	100.000
--------	---------

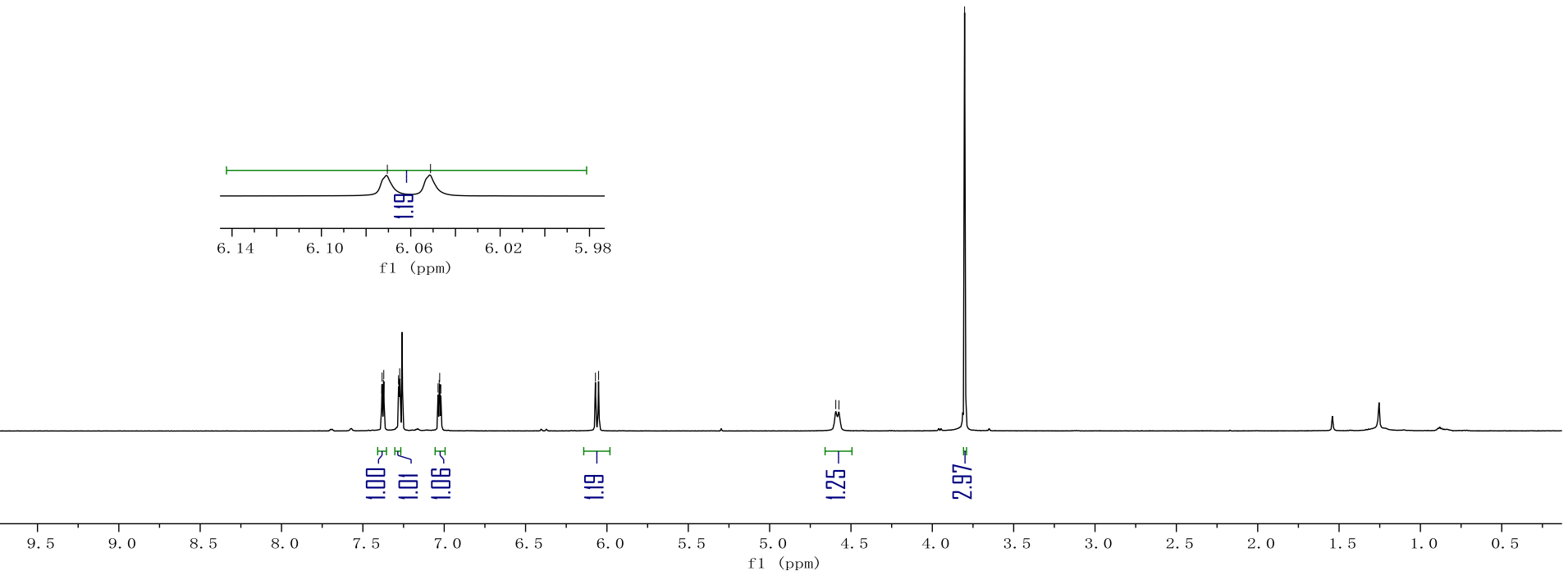
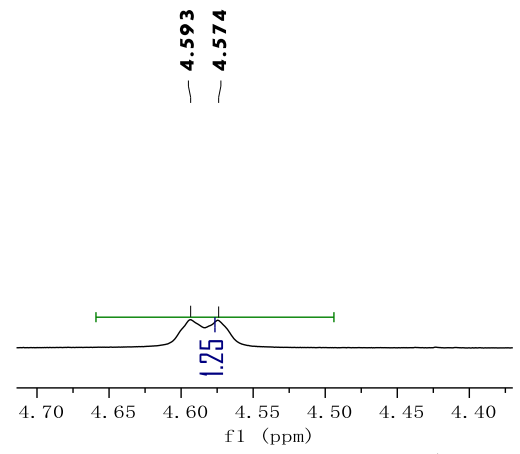
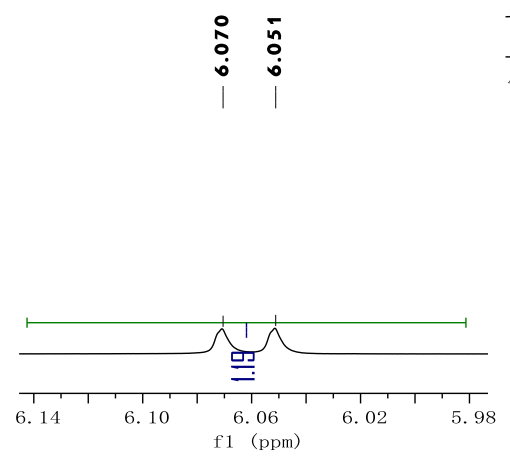
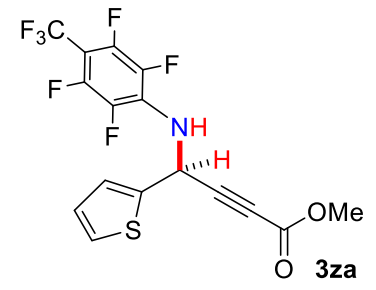


¹H NMR

7.384
7.382
7.374
7.372
7.281
7.279
7.274
7.271
7.038
7.031
7.028
7.021
6.070
6.051

4.593
4.574

3.802



¹³C NMR

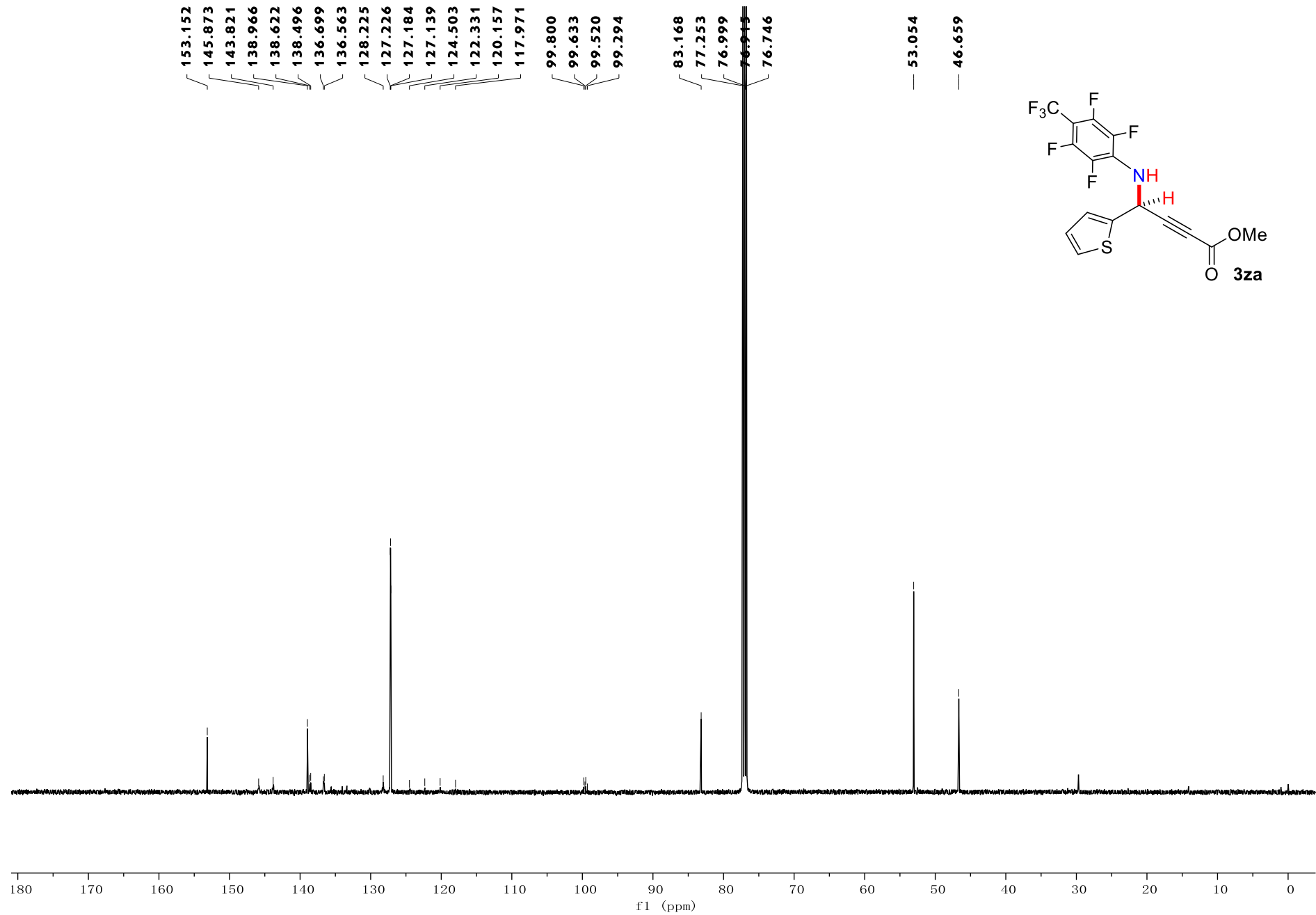
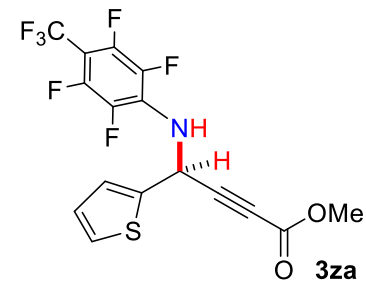
153.152
145.873
143.821
138.966
138.622
138.496
136.699
136.563
128.225
127.226
127.184
127.139
124.503
122.331
120.157
117.971

99.800
99.633
99.520
99.294

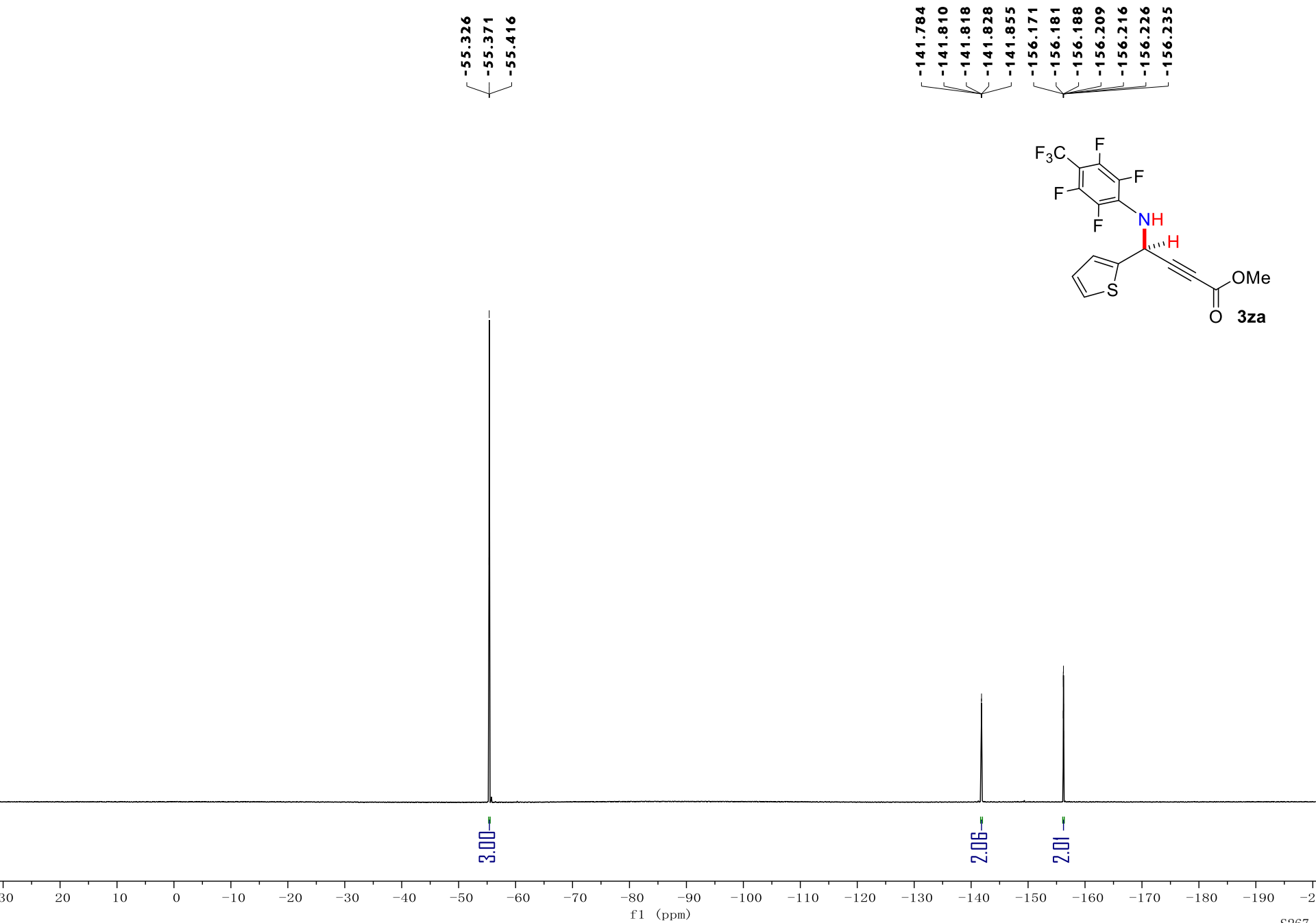
83.168
77.253
76.999
76.915
76.746

53.054

46.659



^{19}F NMR



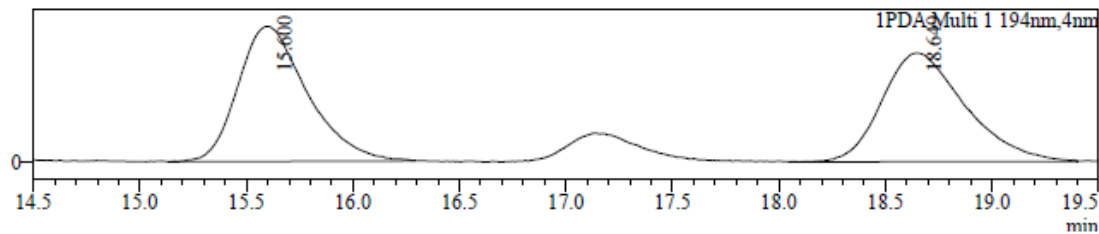
HPLC

Sample Information

Sample Name : POX0768-ODH-5%-0.8
Sample ID : POX0768-ODH-5%-0.8
Data File : POX0768-ODH-5%-0.8.lcd
Method File : POX-5.0%-0.8ml.lcm

Chromatogram

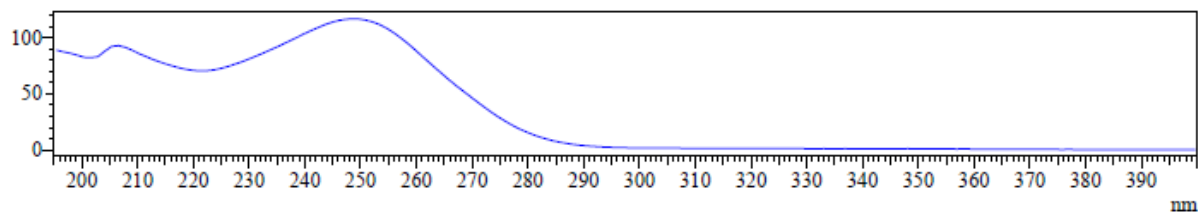
mAU



UV Spectrum

Retention time = 15.600

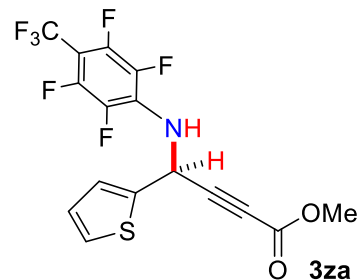
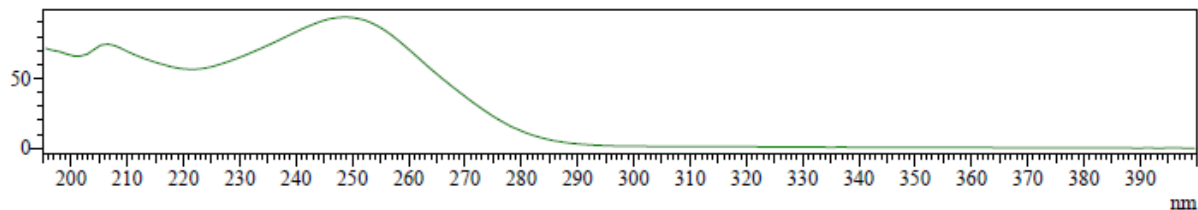
mAU



UV Spectrum

Retention time = 18.649

mAU



Peak Table

PDA Ch1 194nm

Peak#	Ret. Time	Area	Area%
1	15.600	2006994	50.817
2	18.649	1942426	49.183
Total		3949419	100.000

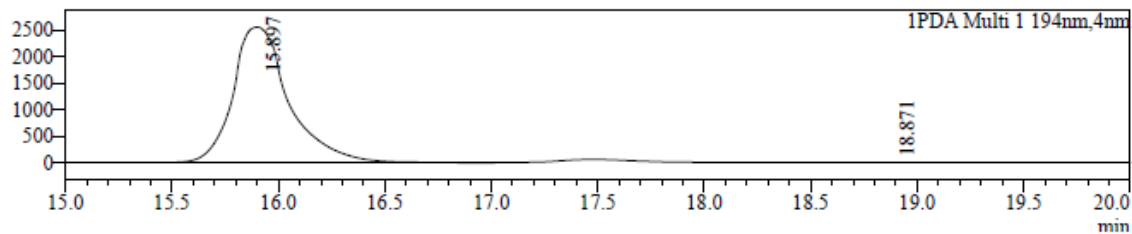
HPLC

Sample Information

Sample Name : P0X-0769-ODH-5%-0.8
Sample ID : P0X-0769-ODH-5%-0.8
Data File : P0X-0769-ODH-5%-0.8.lcd
Method File : P0X-5.0%-0.8ml.lcm

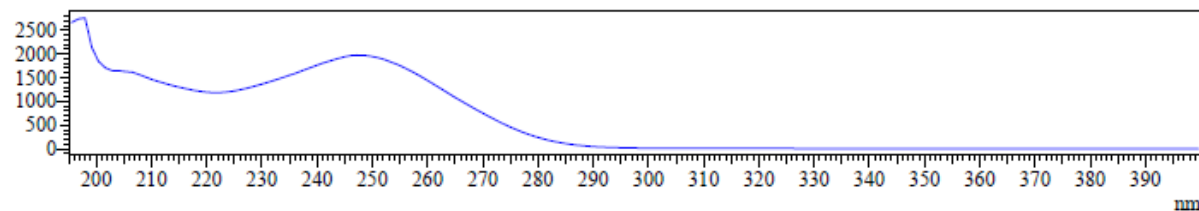
Chromatogram

mAU



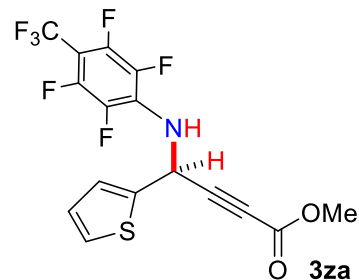
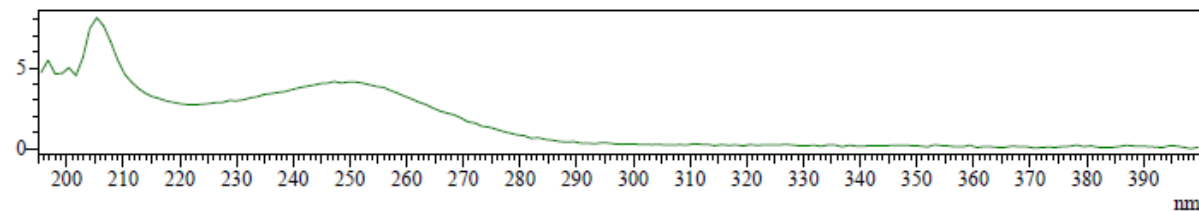
UV Spectrum
Retention time = 15.897

mAU



UV Spectrum
Retention time = 18.871

mAU

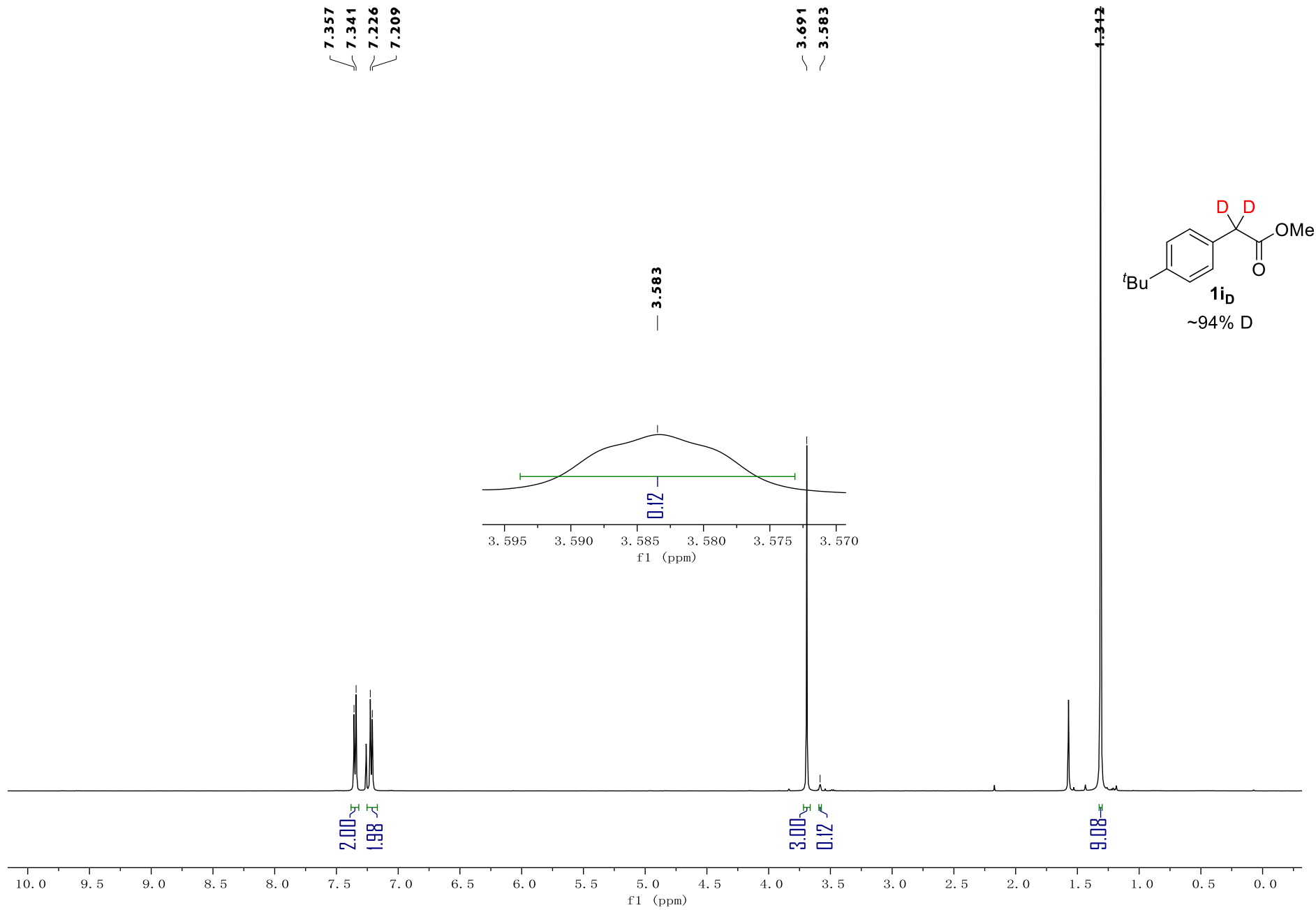


Peak Table

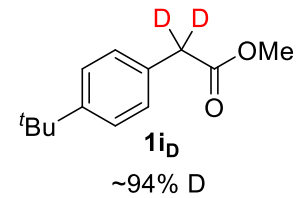
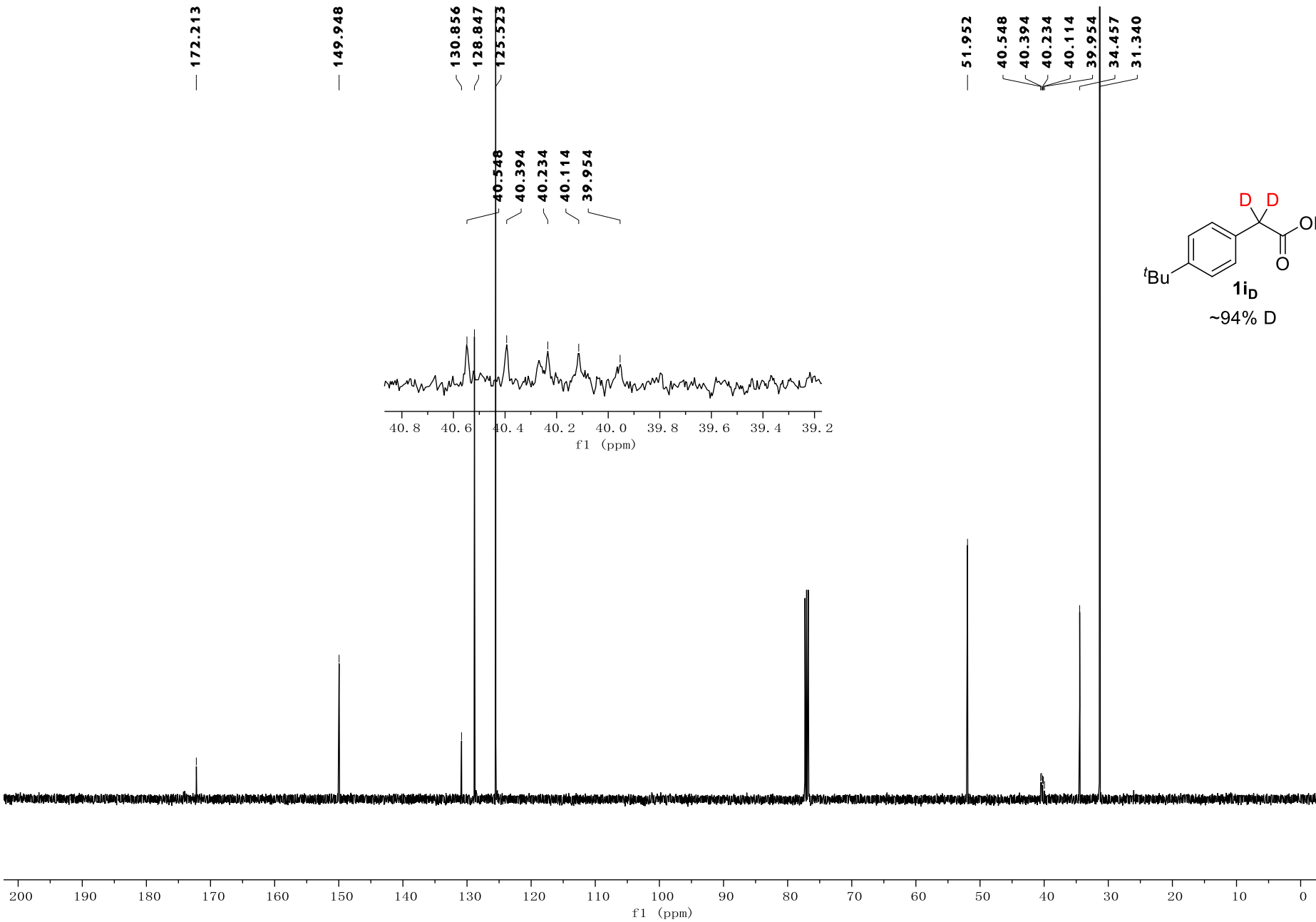
PDA Ch1 194nm

Peak#	Ret. Time	Area	Area%
1	15.897	46609958	99.862
2	18.871	64216	0.138
Total		46674174	100.000

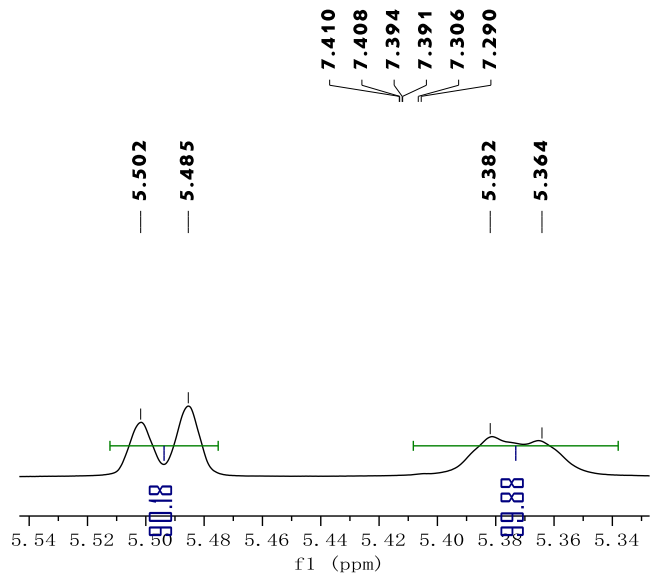
^1H NMR



^{13}C NMR



¹H NMR

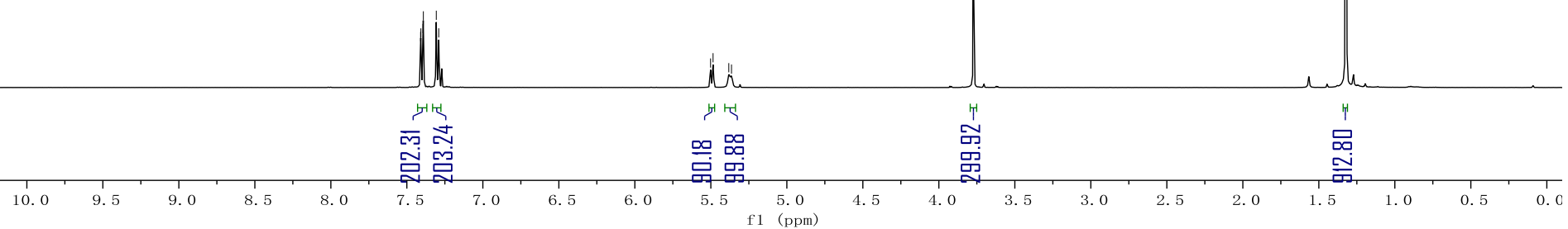
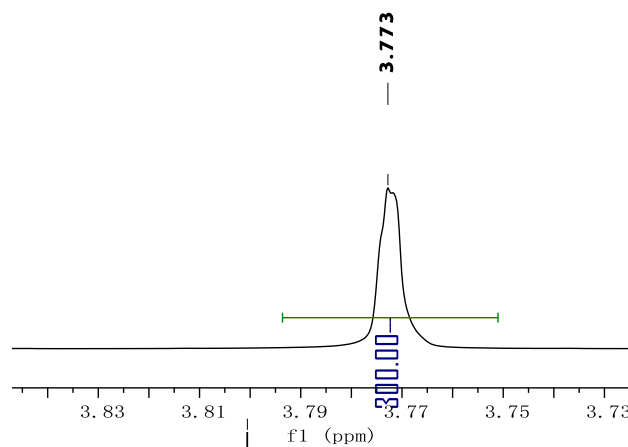
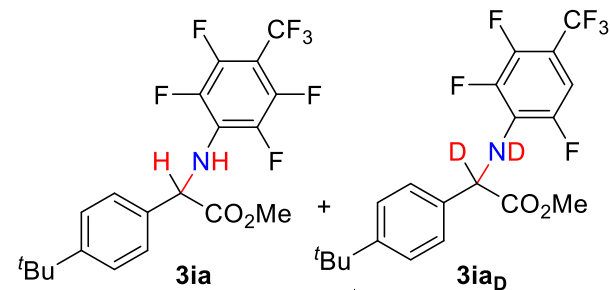


$$\text{KIE} = \frac{90.18}{9.82} \times \frac{94}{106} = 8.1$$

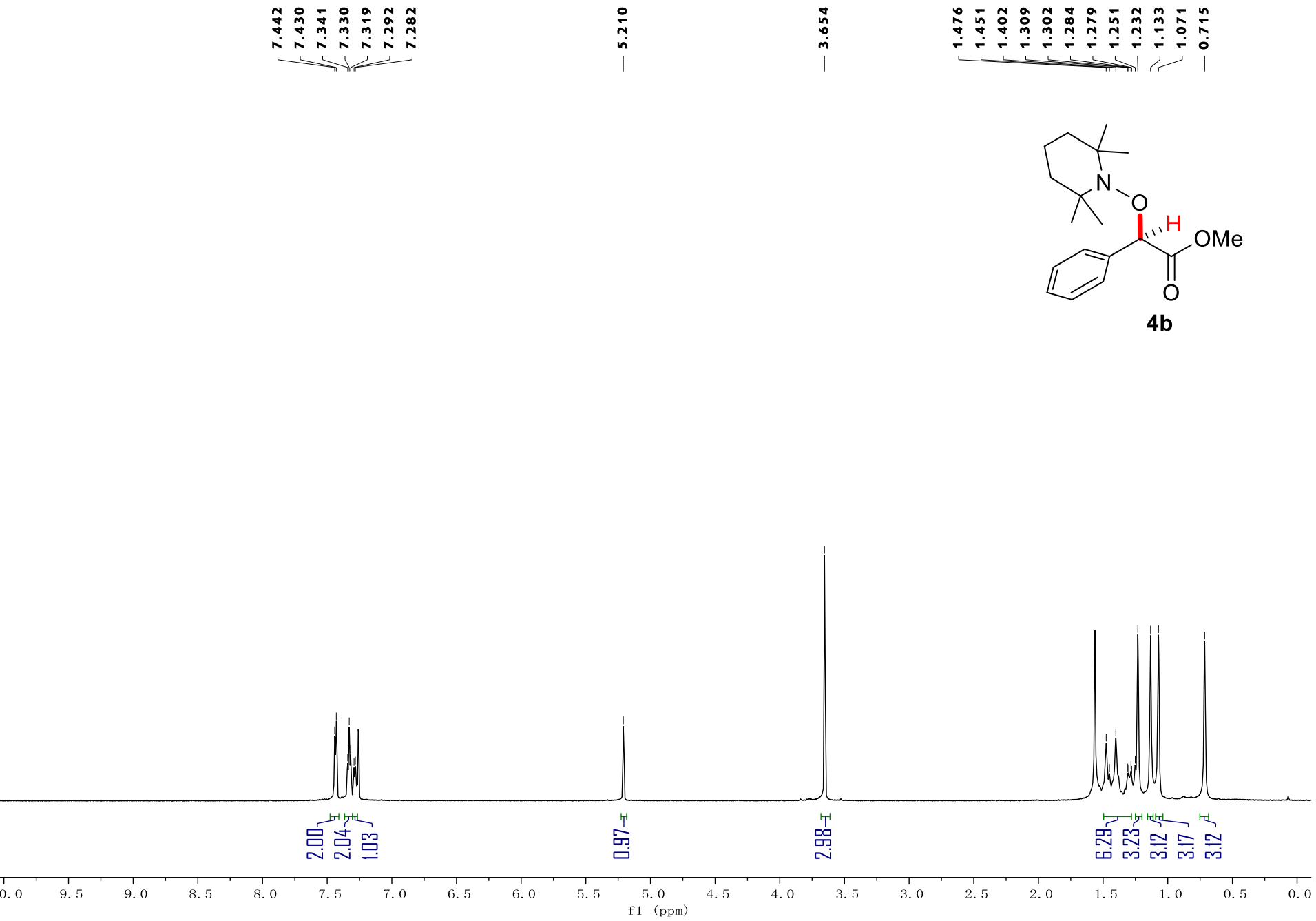
5.502
5.485
5.382
5.364

3.773

1.322



¹H NMR



¹³C NMR

— 172.445

— 138.144

128.312

127.928

126.866

— 88.570

— 59.888

— 51.824

40.214

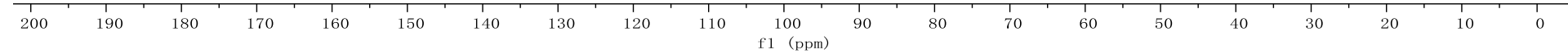
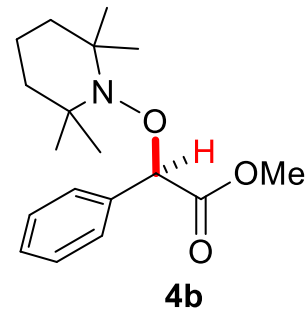
40.079

33.531

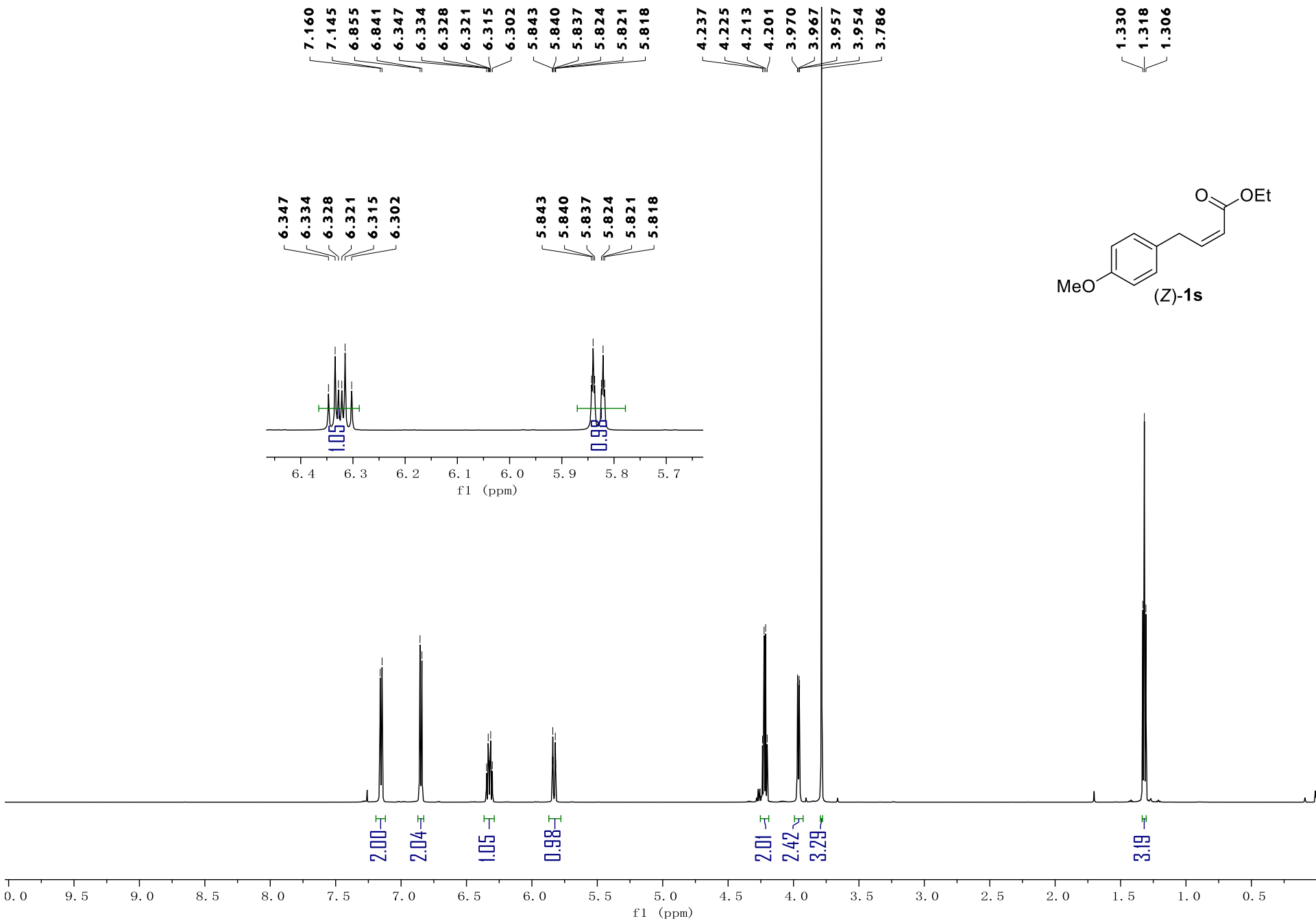
32.836

— 20.133

— 17.069



¹H NMR



¹³C NMR

— 166.345

— 158.113

— 148.309

— 131.437

— 129.490

— 129.480

— 119.517

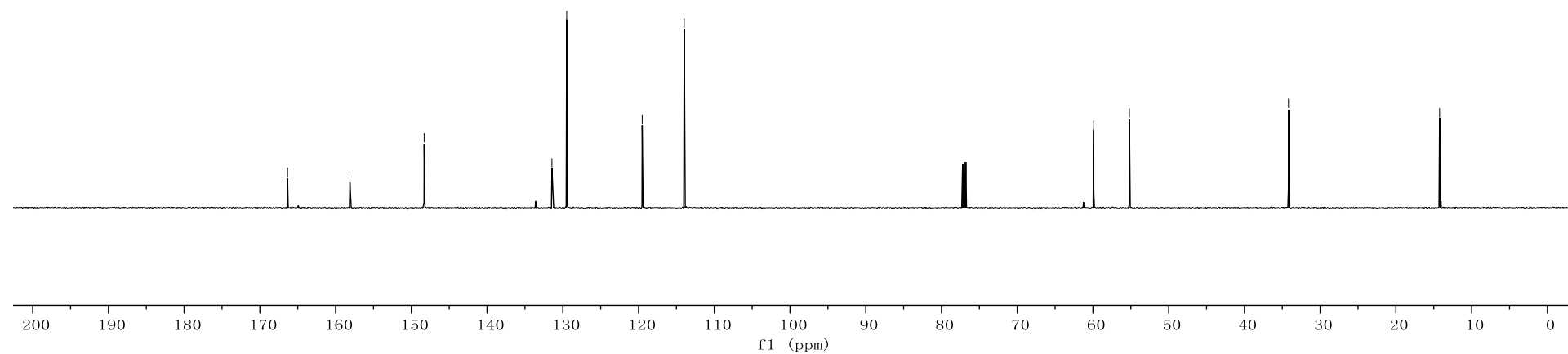
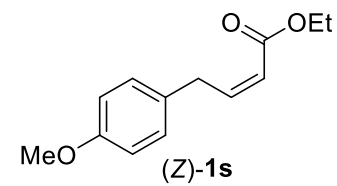
— 113.972

— 59.902

— 55.187

— 34.184

— 14.231

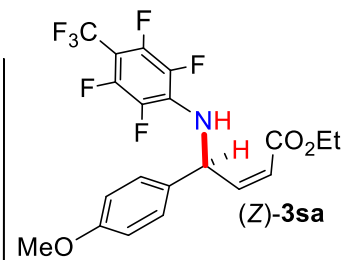


¹H NMR

7.366
7.349
6.925
6.907
6.665
6.647
6.629
6.332
6.314
6.309
6.291
5.937
5.935
5.914
5.912

4.652
4.635
4.255
4.248
4.241
4.237
4.234
4.227
4.223
4.219
4.215
4.208
4.205
4.201
4.194
4.187
3.813

1.320
1.306
1.291



2.01

2.00

0.98

1.00

0.95

0.97

2.04

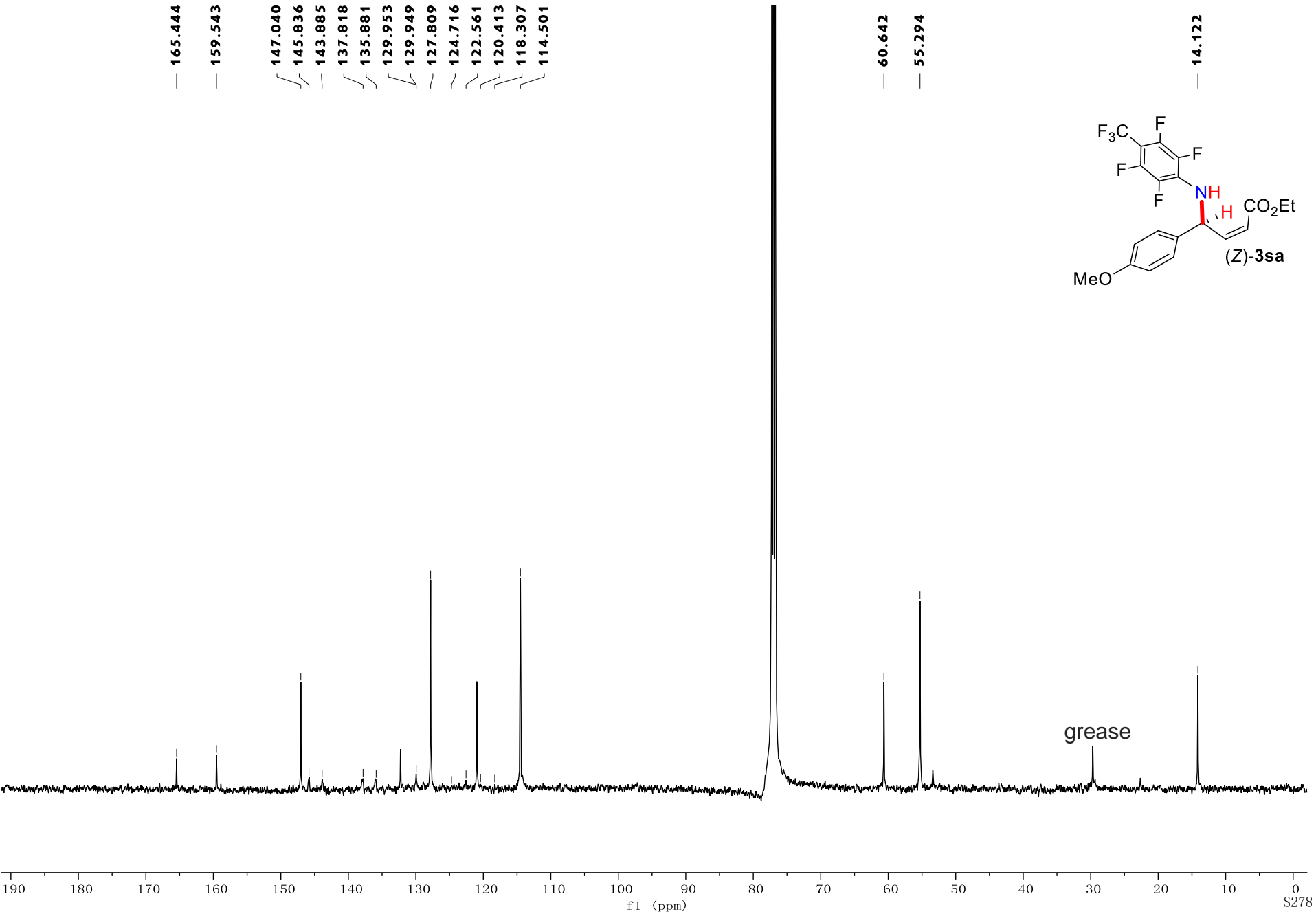
3.09

3.37

f1 (ppm)

S277

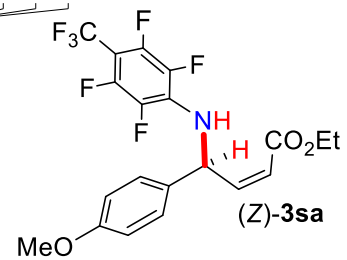
^{13}C NMR



^{19}F NMR

-54.985
-55.030
-55.074

-142.927
-142.936
-142.944
-142.972
-142.978
-142.988
-143.016
-143.023
-143.033
-157.961
-157.976
-158.004
-158.011
-158.020
-158.028



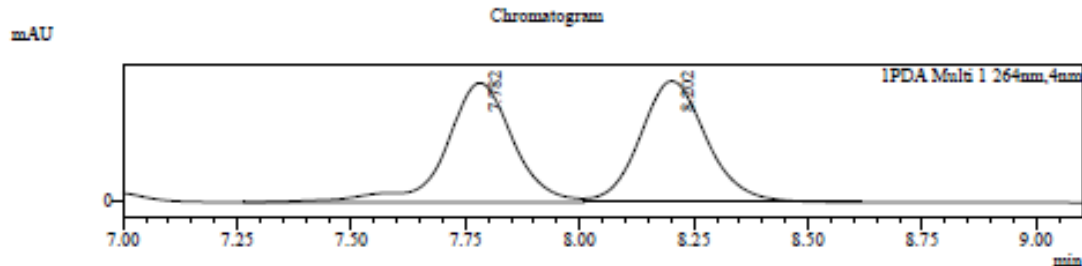
2.94

2.00

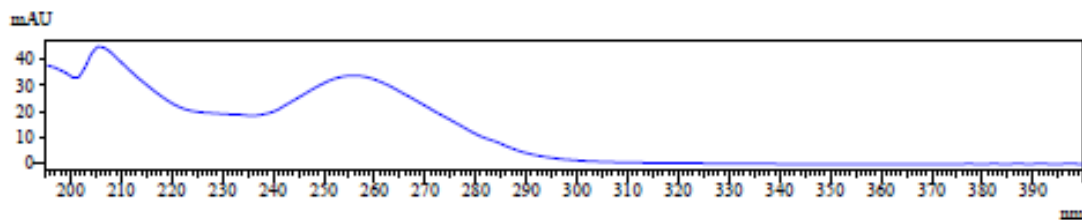
1.90

HPLC

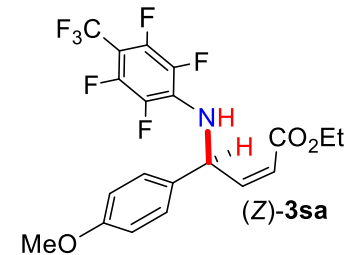
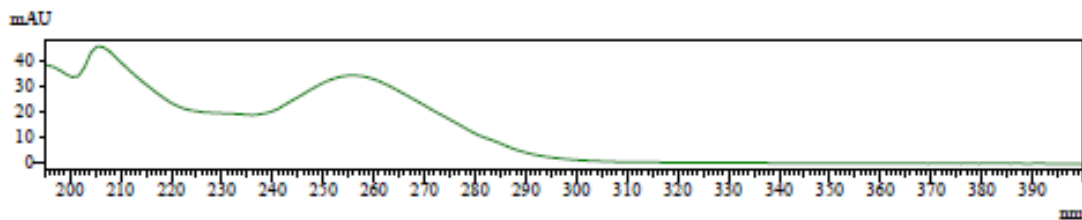
Sample Information
 Sample Name : POX-0548-2-IA5%-0.8mL.lcd
 Sample ID : POX-0548-2-IA5%-0.8mL.lcd
 Data File : POX-0548-2-IA5%-0.8mL.lcd
 Method File : POX-5%.0.8.mL-30min.lcm



UV Spectrum
 Retention time = 7.782



UV Spectrum
 Retention time = 8.202



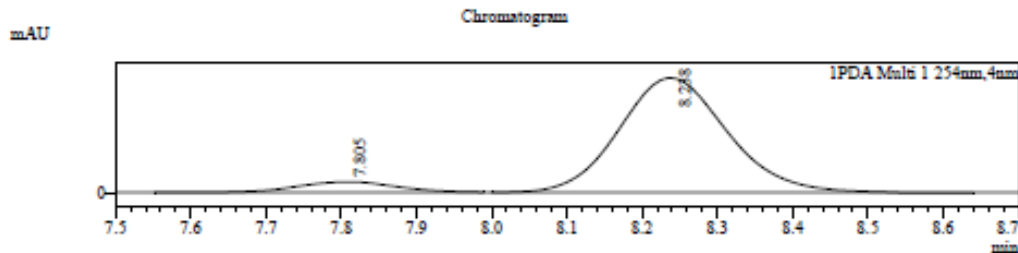
Peak Table

PDA Ch1 264nm

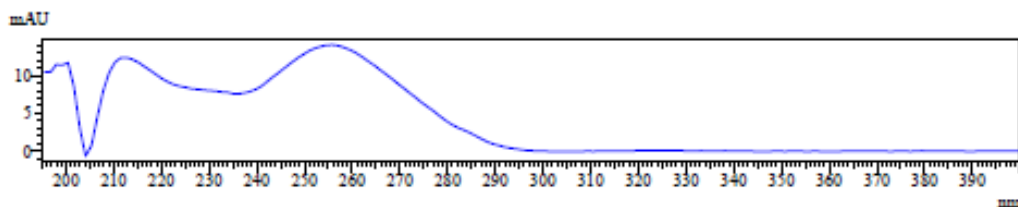
Peak#	Ret. Time	Area	Area%
1	7.782	297484	49.962
2	8.202	297940	50.038
Total		595424	100.000

HPLC

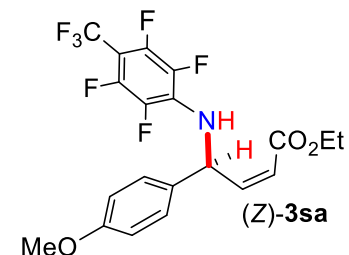
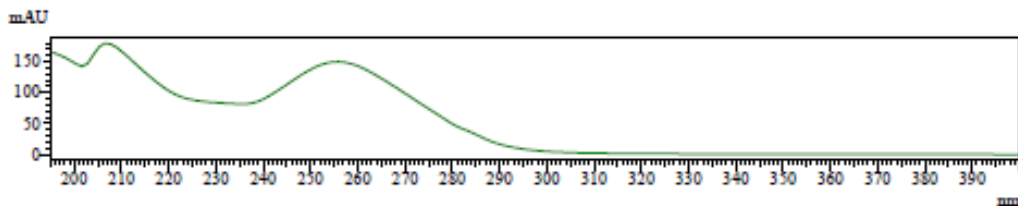
Sample Information
 Sample Name : POX-1252-2-IA-5%-0.8mL.lcd
 Sample ID : POX-1252-2-IA-5%-0.8mL.lcd
 Data File : POX-1252-2-IA-5%-0.8mL.lcd
 Method File : POX-5%.0.8.mL.-30min.lcm



UV Spectrum
 Retention time = 7.805



UV Spectrum
 Retention time = 8.238

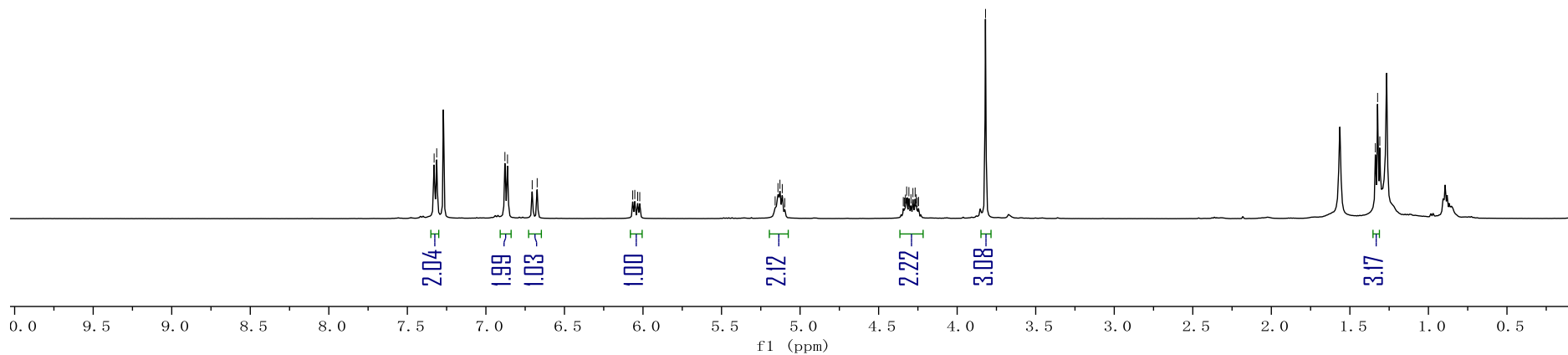
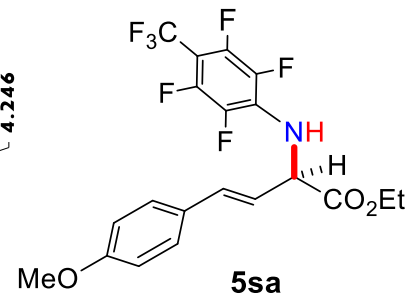
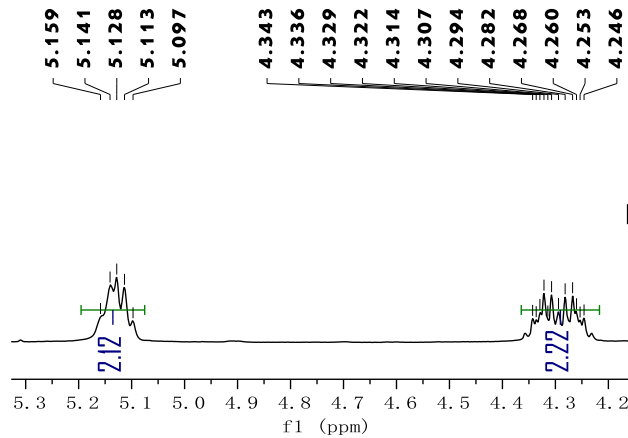
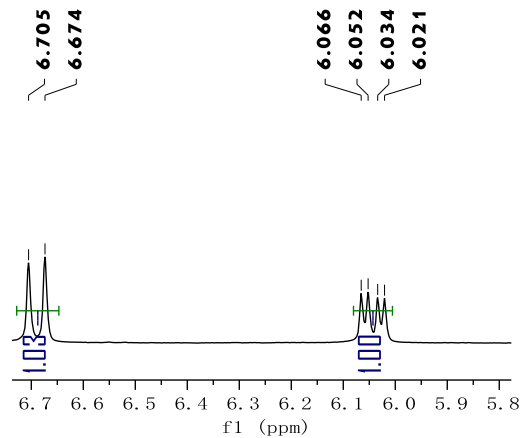


Peak Table

PDA Ch1 254nm

Peak#	Ret. Time	Area	Area%
1	7.805	122218	7.666
2	8.238	1471999	92.334
Total		1594217	100.000

¹H NMR

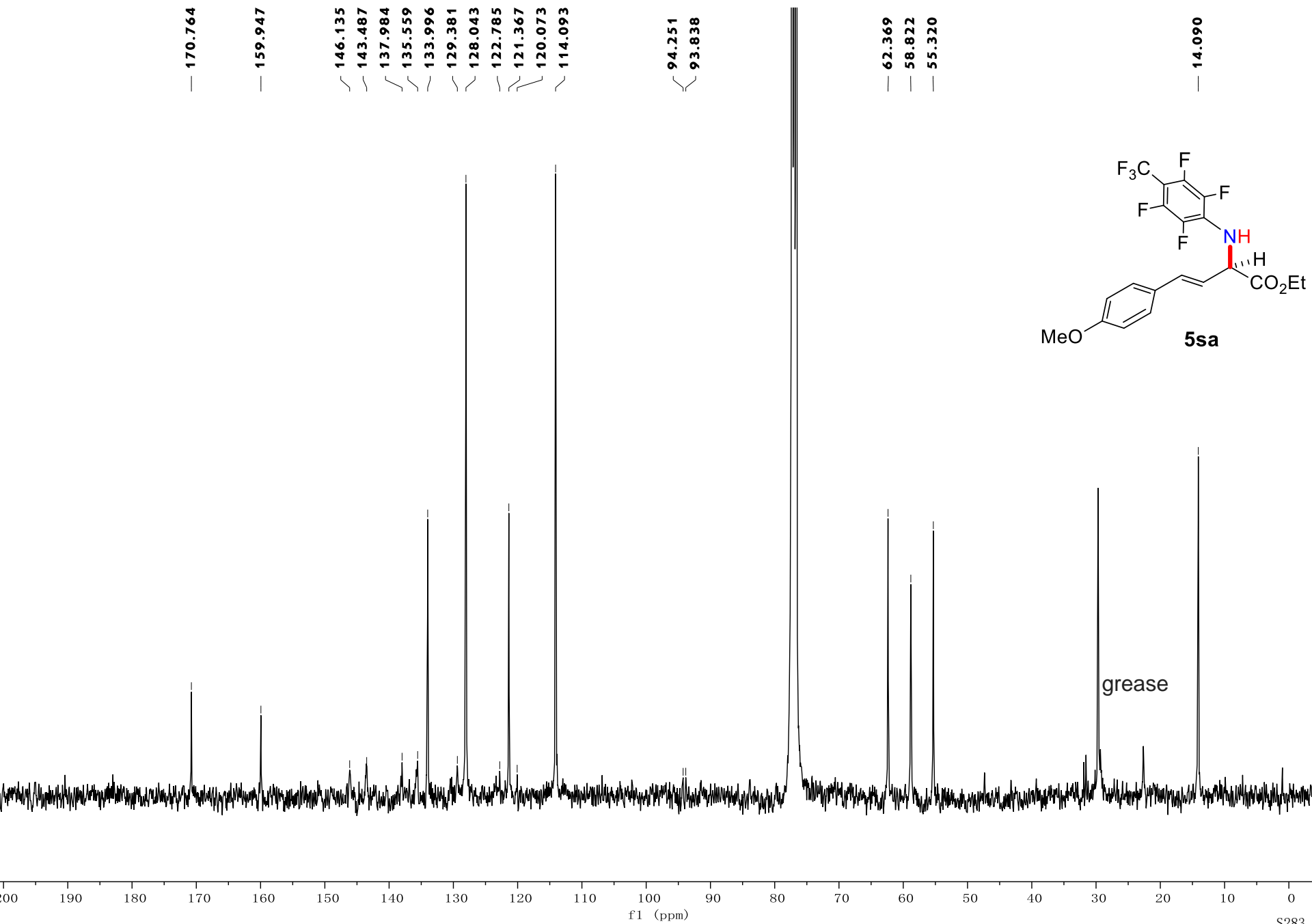


7.330
7.313
6.879
6.862
6.705
6.674
6.066
6.052
6.034
6.021

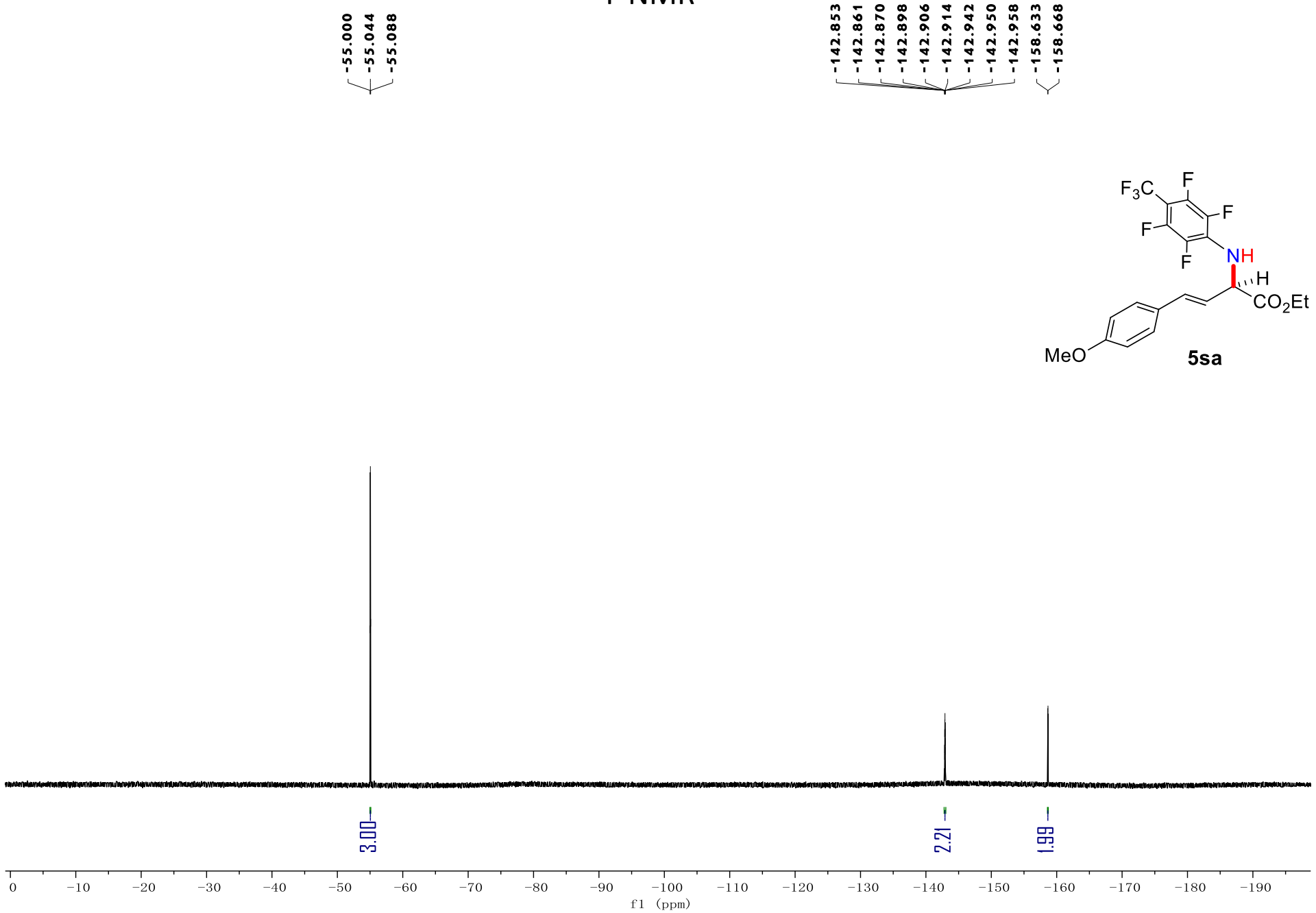
5.159
5.141
5.128
5.113
5.097
4.343
4.336
4.329
4.322
4.314
4.307
4.294
4.282
4.268
4.260
4.253
4.246
3.820

1.338
1.324
1.309

¹³C NMR

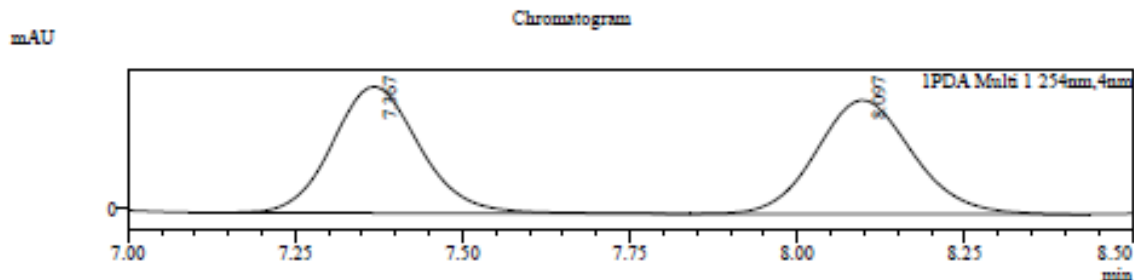


¹⁹F NMR

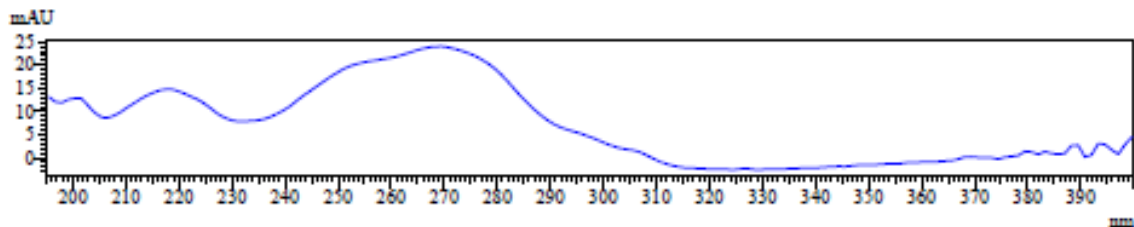


HPLC

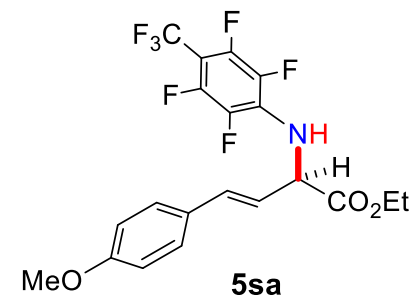
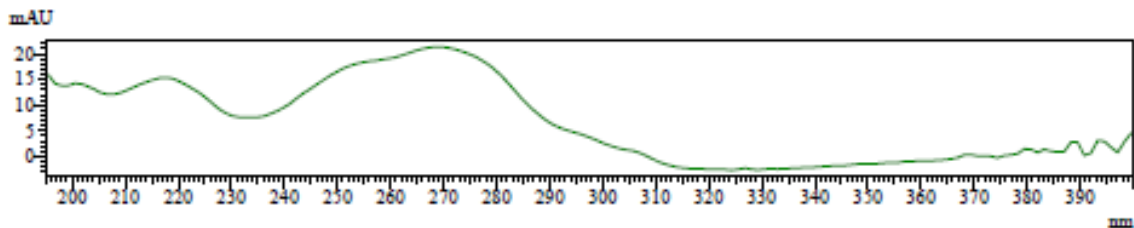
Sample Information
Sample Name : P0X-548-4-IA5%0.8mL.lcd
Sample ID : P0X-548-4-IA5%0.8mL.lcd
Data File : P0X-548-4-IA5%0.8mL.lcd
Method File : P0X-5% 0.8.mL-30min.lcm



UV Spectrum
Retention time = 7.367



UV Spectrum
Retention time = 8.097

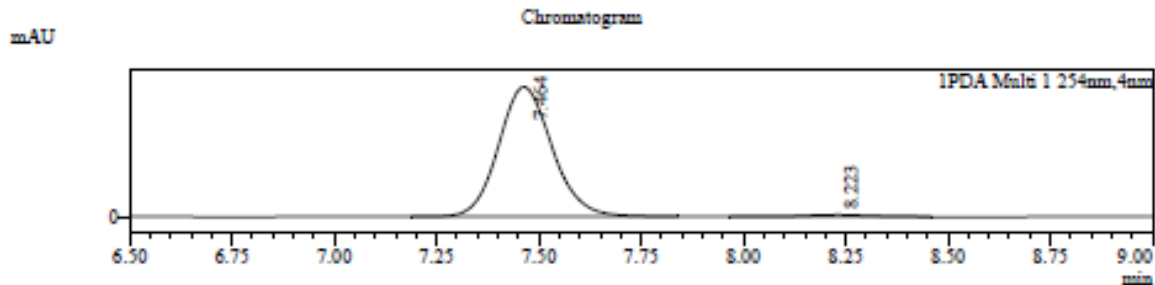


Peak Table

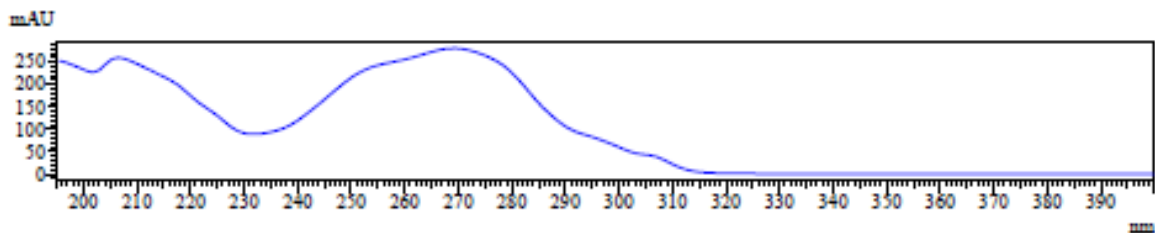
Peak#	Ret. Time	Area	Area%
1	7.367	191108	50.186
2	8.097	189691	49.814
Total		380799	100.000

HPLC

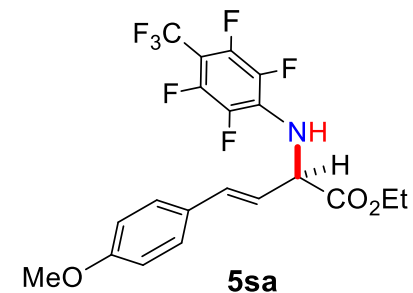
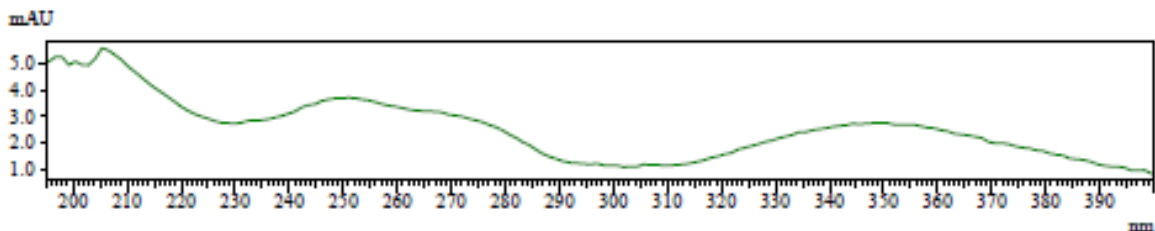
Sample Information
 Sample Name : POX-1252-4-IA-5%-0.8mL.lcd
 Sample ID : POX-1252-4-IA-5%-0.8mL.lcd
 Data File : POX-1252-4-IA-5%-0.8mL.lcd
 Method File : POX-5%.0.8.mL-30min.lcm



UV Spectrum
 Retention time = 7.464



UV Spectrum
 Retention time = 8.223



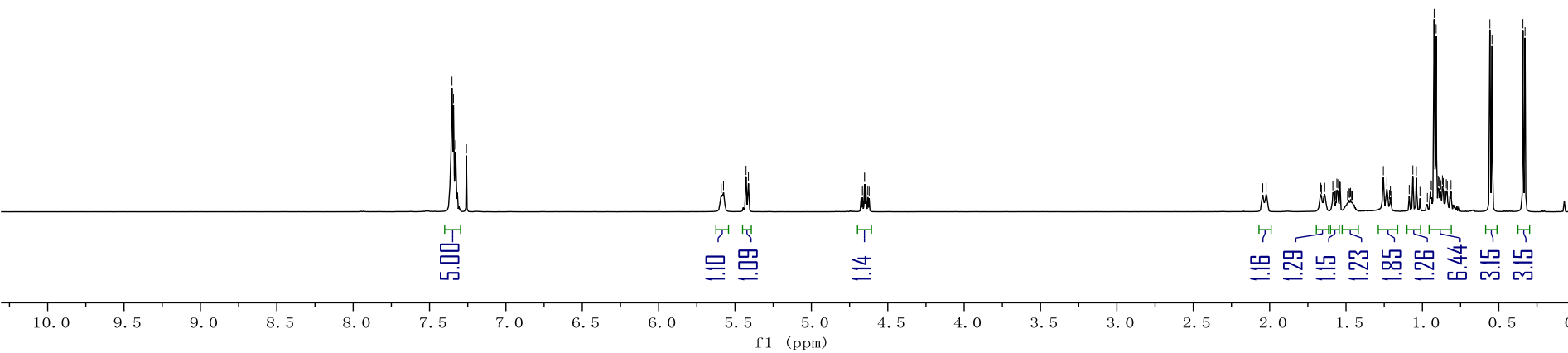
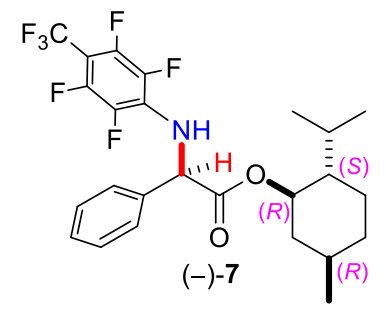
Peak Table

PDA Ch1 254nm

Peak#	Ret. Time	Area	Area%
1	7.464	2170314	98.575
2	8.223	31383	1.425
Total		2201697	100.000

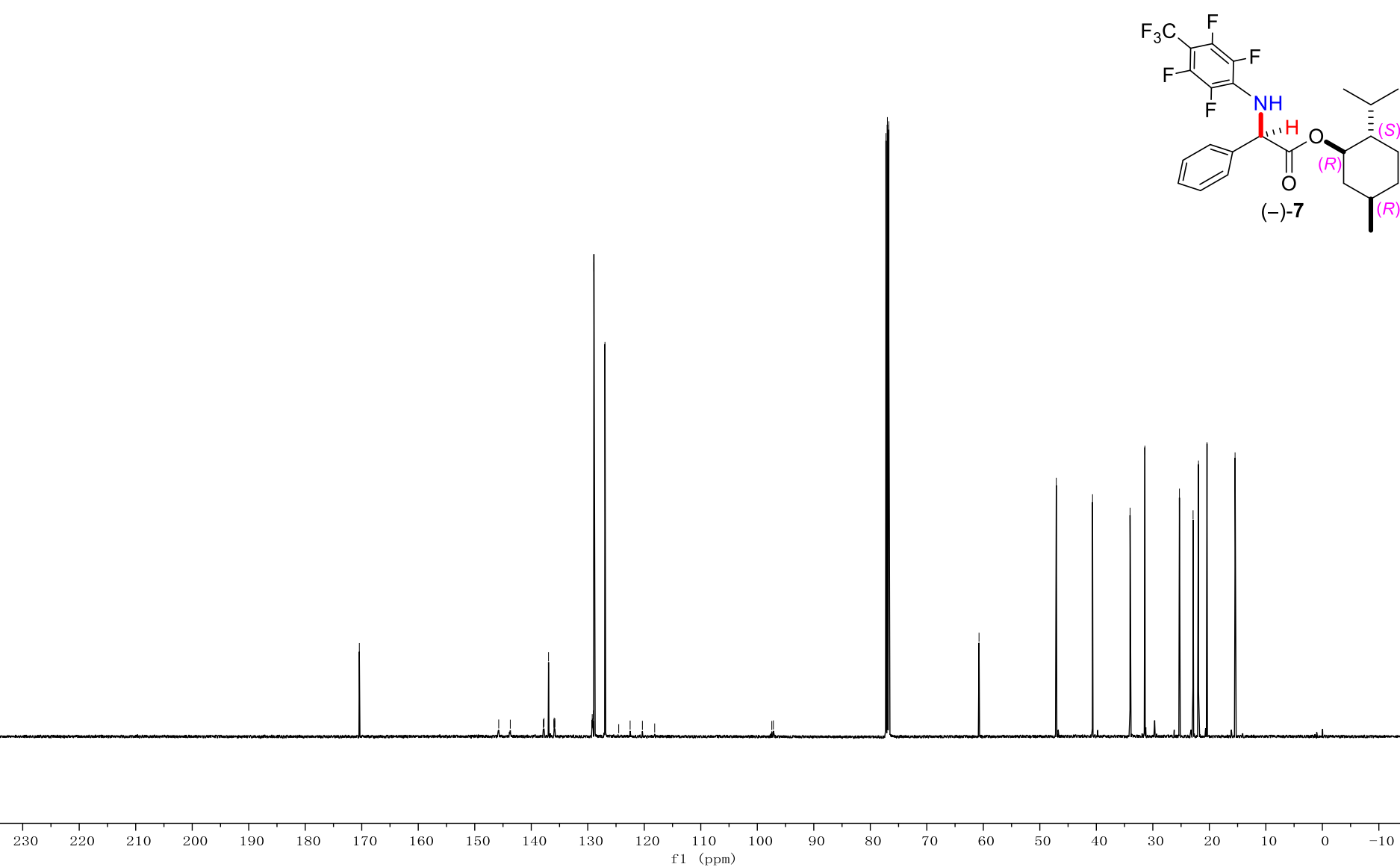
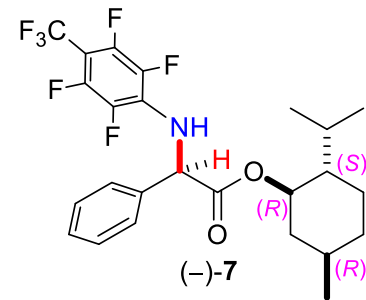
¹H NMR

7.353
7.348
7.343
7.338
7.334
7.329
7.258
5.590
5.575
5.428
5.412
4.652
4.644
2.045
2.023
1.665
1.661
1.640
1.587
1.580
1.560
1.554
1.256
1.232
1.086
1.062
1.040
0.948
0.941
0.923
0.910
0.897
0.891
0.884
0.878
0.869
0.863
0.844
0.837
0.820
0.813
0.558
0.544
0.342
0.328



¹³C NMR

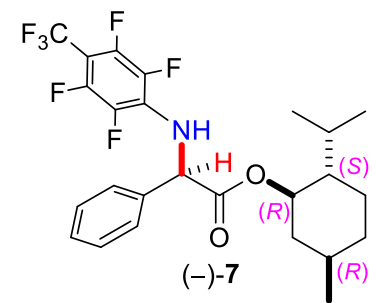
- 170.429
- 145.759
- 143.710
- 137.870
- 137.744
- 136.946
- 135.994
- 135.948
- 135.913
- 135.871
- 135.819
- 129.300
- 129.212
- 129.178
- 129.120
- 129.030
- 128.913
- 128.885
- 126.960
- 124.523
- 122.503
- 120.333
- 118.162
- 97.431
- 97.153
- 77.230
- 76.976
- 76.721
- 60.760
- 47.085
- 40.671
- 34.029
- 31.405
- 25.293
- 22.877
- 21.917
- 20.412
- 15.447



¹⁹F NMR

-55.033
-55.077
-55.122

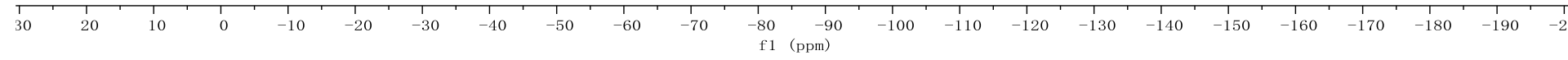
-142.928
-142.941
-142.957
-142.972
-143.001
-143.017
-143.046
-143.061
-143.077
-143.090
-143.105
-158.654
-158.686



3.00

2.07

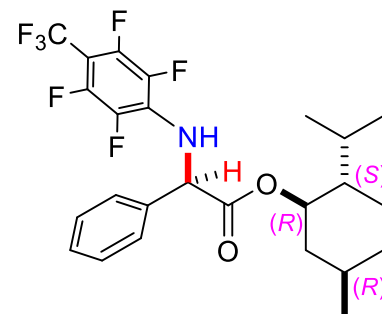
2.04



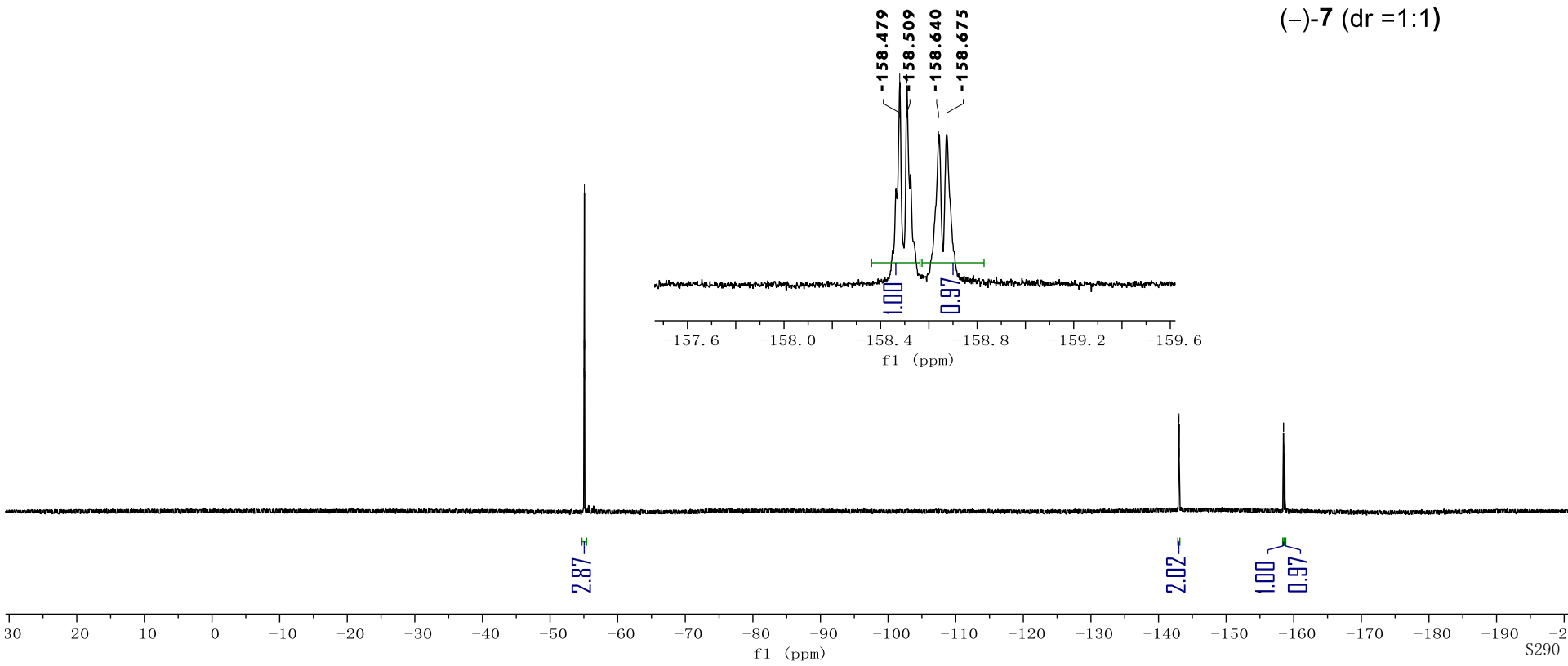
¹⁹F NMR

-55.027
-55.035
-55.072
-55.080
-55.116
-55.124

-142.989
-143.005
-143.016
-143.033
-143.061
-158.479
-158.509
-158.640
-158.675



(-)-7 (dr=1:1)



¹⁹F NMR

