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Article Title:	The activin receptor IIa ligand trap Sotatercept corrects ineffective erythropoiesis in beta-thalassemia
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Supplementary Item & Number	Supplementary figure legends and Supplementary Figures 1-8
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Supplementary Figure 1	[TITLE OF FIGURE (descriptive title used in legend)]
Supplementary Table 1	[TITLE OF TABLE (descriptive title)]
Supplementary Methods (or Discussion or Data or Note)	[NO DESCRIPTIVE TITLE NECESSARY]
Supplementary Video 1 (or Audio or Spreadsheet)	[CAPTION TO BE POSTED ONLINE (include title and explanation of contents)]

The activin receptor IIa ligand trap Sotatercept corrects

ineffective erythropoiesis in beta-thalassemia

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SUPPLEMENTAL FIGURE LEGENDS

Supplemental figure 1: (a-h) RAP-011 effects on hematological parameters in wild-type mice.

C57BL/6 mice were treated with RAP-011 (10 mg/Kg twice a week) or PBS for 30 days. Evaluation of

(a) red blood cell counts (RBC), (b) hematocrit level, (c) percentage and (d) absolute numbers of

reticulocytes, (e) hemoglobin, (f) mean corpuscular volume (MCV), (g) mean corpuscular hemoglobin

(MCH) and (h) MCH concentration (MCHC). (i-l) RAP-011 effects on bone marrow and spleen

cellularity and erythroblasts numbers in wild-type mice 30 days post-treatment. C57BL/6 mice were

treated with RAP-011 or PBS. (i,j) spleen and bone marrow cellularity. (k,l) Absolute numbers of Ter-

119⁺ cells in spleen and bone marrow. (m-o) RAP-011 effects on erythroblast differentiation in

thalassemic mice. Hbb^{th1/th1} mice were treated 5-30 days with RAP-011 or PBS and cell differentiation

was evaluated by flow cytometry. The absolute numbers of different erythroblast populations (ProE,

proerythroblast; Ery.A, basophilic erythroblast; Ery.B, late basophilic and polychromatic erythroblast;

Ery.C, orthochromatic erythroblast) on (m) spleen and (n) bone marrow are shown. (o) An index of

ineffective erythropoiesis was established by calculating the ratio of Ery.B and Ery.C populations. (p)

Erythroid differentiation in C57BL/6 mice treated twice a week for 30 days with 10 mg/kg RAP-011 or

PBS. Bone marrow and spleen were harvested and erythroblast differentiation was evaluated by

CD71/Ter-119 staining and FSC distribution. The absolute numbers of erythroblast populations are

shown. *p<0.05, N=5 per group. Data represent means ± S.E.M.

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Supplemental figure 2: (a) RAP-011 reduces ineffective erythropoiesis in Hbb^{th1/th1} thalassemic mice. (a) lactate dehydrogenase (LDH) levels in thalassemic mice treated for up to 60 days with 10 mg/kg RAP-011 or PBS. (b-d) RAP-011 effects on bilirubin and LDH levels in wild-type mice. Biochemical analysis of sera from wild-type mice treated for 30 days with 10 mg/kg RAP-011 or PBS: (b) direct bilirubin, (c) total bilirubin and (d) LDH. (e-g) RAP-011 effects on iron homeostasis parameters in wildtype mice. Biochemical analysis of iron homeostasis parameters of sera from wild-type mice treated for 30 days with RAP-011 or PBS: (e) Systemic iron levels, (f) transferrin levels and (g) transferrin saturation. (h-j) RAP-011 effects on GDF15 expression. Hbb^{th1/th1} mice were treated with PBS or RAP-011 for 30 days. Gdf15 mRNA levels measured by qPCR in (h) spleen or (i) bone marrow samples. (j) RAP-011 has no effect on GDF15 signaling. CAGA-Luc/GFP C2C12 cells were treated with GDF15 (100 ng/ml) or with vehicle in the presence of RAP-011 (10 µg/ml) or a specific GDF15 antibody (1 µg/ml). Luciferase activity was normalized to GFP fluorescence; values are expressed as relative luminescence units per relative fluorescence units (RLU/RFU). (k-m) RAP-011 effects on globin chains expression 30 days post-treatment. Hbb^{th1/th1} mice were treated with PBS or RAP-011 for 30 days. Adult (Hbb-b2) (k) or embryonic (Hbb-bh1 and Hbb-Ey) (I,m) globin mRNAs expression was evaluated by qPCR in purified immature (CD71⁺) erythroblasts from wild-type and Hbb^{th1/th1} mice. All data are expressed as the mean ± S.E.M. *P<0.05, **P<0.01, ns=not significant; N=5 per group for each of three independent experiments performed.

Supplemental figure 3: (a-b) retrospective analysis of dataset GSE34125 showing the expression levels of several TGF- β superfamily. Expression of TGF- β superfamily (a) ligands and (b) receptors on CD71⁺Ter-119⁺FSC^{high} matched erythroblasts from E14.5 fetal livers of wild-type and Hbb^{Th3/Th3} embryos. Data were obtained at Gene Expression Omnibus (http://www.ncbi.nlm.nih.gov/geo) and analyzed using GEO2R software. (c-f) TGF- β superfamily gene expression analysis in spleen and bone marrow samples from wild-type and Hbb^{th1/th1} mice. mRNA levels of (c) Acvr2a, (d) Acvr2b, (e) Acvr1b

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(ALK4) and (**f**) Tgfbr1 (ALK5) were evaluated by qPCR in spleen and bone marrow purified erythroblasts derived from wild-type and thalassemic mice. (**g-i**) *Gene expression analysis from spleen and bone marrow-derived cultured erythroblasts from wild-type and Hbb*^{th1/th1} mice. (**g**) Acvr2a, (**h**) Acvr2b and (**i**) Acvr1b (ALK4) mRNA levels evaluated by qPCR. All data are expressed as the mean \pm S.E.M. *P<0.05, **P<0.01, ***P<0.005; ns = not significant; N=5 per group for each independent experiment.

Supplemental figure 4: mRNA levels for TGF-6 family members in spleen and bone marrow erythroblast samples of wild-type and Hbb^{th1/th1} mice. (a) Inha (activin A), (b) Inhb (activin B) and (c) Gdf15 mRNA levels in spleen and bone marrow erythroblasts from wild-type and thalassemic mice. (d) RAP-011 effects on Actlla ligands expression. Thalassemic or wild-type erythroblasts were either cultured in the presence of RAP-011 (10 µg/ml) or PBS for 48h. mRNA levels for activin A (Inha), activin B (Inhb), GDF11 and ActRIIa (Acvr2a) evaluated by qPCR. (e-g) RAP-011 traps GDF11. (e) Immunoblotting analysis of bone marrow from thalassemic mice treated with PBS and RAP-011 show a significant decrease of GDF11 protein levels. Recombinant protein was used as a positive control. (f) Immunohistochemical analysis of GDF11 pro-peptide and mature form in bone marrow from wildtype and thalassemic mice. (g) Intracellular flow cytometric analysis of splenic erythroblasts treated with 10 µg/ml RAP-011 or PBS for 48 hours and stained using specific antibodies against activin A, activin B and GDF11 (proform and mature). Quantification of the staining is shown in the bar graph and expressed as mean fluorescence intensity (MFI). Right panel demonstrates a representative FACS histogram where gray lines indicate isotype antibody control, black and red represent PBS or RAP-011 treated erythroblasts (respectively). Data represent means ± S.E.M. All data are expressed as the mean ± S.E.M.*P<0.05, ns=not significant; N=5 per group for each independent experiment.

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Supplemental figure 5: GDF11 expression of thalassemic mice and thalassemia subjects. (a) Confocal micrograph showing a representative section of spleen white pulp (WP) and red pulp (RP) from Hbb^{th1/th1} mice. GDF11 protein expression is shown in green, CD71 in red and nuclei (DAPI) are shown in blue. (b) GDF11 in overexpressed in GPA⁺ cells from thalassemic subjects. Confocal micrograph showing a representative cytospin of cultured CD36⁺ erythroblasts from thalassemic subjects and cord blood-derived erythroblasts from healthy donors. GDF11 protein expression is shown in green. GPA⁺ erythroblasts are marked in red and nuclei (DAPI) are shown in blue. (c,d) Flow cytometric quantification of Fas and FasL-positive CD71 Ter-119 populations harvested from (c) spleen or (d) bone marrow of Hbb^{th1/th1} mice treated twice weekly with 10 mg/kg RAP-011 or PBS. (e) GDF11 inactivation promotes spontaneous apoptosis of erythroblasts. Cultured erythroblasts from thalassemic mice were transfected with siRNA against a scrambled control (gray), Inhb (blue) or Gdf11 (red). The histograms represent annexin V binding. (f-h) Recombinant GDF11 activates the Smad2/3 but not the Smad1/5 pathway. (f) Growing doses of GDF11 (0-100 ng/ml) induces Smad2/3 signaling in C2C12 cells as demonstrated by CAGA-Luc/GFP but not Smad 1/5 signaling as demonstrated by BRE-Luc/GFP reporter vectors. BMP2 (20 ng/ml) and TGF-β1 (5 ng/ml) were used as positive controls. (g) The Alk4/Alk5 inhibitors SB505124 and SB431542 abrogate the signaling induced by GDF11 in C2C12 cells. (h) RAP-011 blocks GDF11 and GDF8 Smad2/3 signaling. CAGA-Luc/GFP C2C12 cells were treated with GDF11 (100 ng/ml), GDF8 (100 ng/ml) or vehicle in the presence of RAP-011 (10 μg/ml) or specific neutralizing antibodies (1 μg/ml). Luciferase activity was normalized to GFP fluorescence; values are expressed as relative luminescence units per relative fluorescence units (RLU/RFU). All data are expressed as the mean ± S.E.M. *P<0.05, **P<0.01, ns=not significant; N=3 per group for each of three independent experiments performed.

Supplemental figure 6: (a-b) *GDF11 neutralization induces terminal maturation of erythroblasts* (a) Erythroblast cultures derived from the spleens of thalassemic mice were treated with neutralizing

antibodies against GDF11 or GDF8 for 48h. Erythroblast differentiation was evaluated by flow cytometry through CD71/Ter-119 staining and FSC distribution into immature (Ter-119⁺/CD71⁺) and mature (Ter-119⁺/CD71⁻) erythroblasts. The percentages of different erythroblast populations (immature/mature) are shown. (b) Morphology of erythroblasts treated with neutralizing antibodies against GDF11 or GDF8 for 48h. (c-d) *Recombinant GDF11 induces Smad signaling in thalassemic but not on wild-type erythroblasts*. (c) spleen and bone marrow-derived erythroblast culture from wild-type and Hbb^{th1/th1} mice under increasing concentrations of recombinant GDF11 (0-500 ng/ml) were assayed for Smad2 phosphorylation (red) and CD71 (blue) staining by confocal immunofluorescence. (d) Western blot analysis on bone marrow-derived erythroblasts.

Supplemental figure 7: (a-b) *Recombinant GDF11 promotes thalassemic erythroblasts terminal maturation arrest.* (a) Erythroblast differentiation was evaluated in samples treated with recombinant GDF11 (0-500 ng/ml) for 7 days by flow cytometry through CD71/Ter-119 staining into immature (Ter-119⁺/CD71⁺) and mature (Ter-119⁺/CD71⁻) erythroblasts. (d) Percentages of mature and immature erythroblast populations are shown. (c,d) Effect of recombinant GDF11 on cell proliferation and survival. Spleen and bone marrow-derived erythroblast culture from wild-type and Hbb^{th1}/th1 mice under increasing concentrations of recombinant GDF11 (0-500 ng/ml) were assayed for (c) ATP content by a luminescence-based assay and (d) cell counts. All data are expressed as the mean ± S.E.M.*P<0.05, ns=not significant; N=5 per group for each independent experiment.

Supplemental figure 8: A model illustrating the proposed role of ActRIIa signaling in ineffective erythropoiesis of thalassemia. Increased GDF11 levels induce Smad2 activation leading to (i) early erythroblasts accumulation through the impairment of Fas/Fas-L expression and to (ii) terminal maturation arrest by promoting oxidative stress (left side of figure). RAP-011, an ActRIIa trap, sequesters GDF11 (and potentially other ActRIIa ligands) impairing ActRIIa activation and Smad2/3

phosphorylation. This results in reduced α -globin aggregates/oxidative stress therefore promoting terminal erythroid maturation and in activation of Fas/Fas-L pathway resulting in apoptosis of early erythroblasts (right side of figure). Under these conditions, reversion of ineffective erythropoiesis results in improvement of anemia of thalassemia.

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