

The Relative Densities of Cytoplasm and Nuclear Compartments Are Robust against Strong Perturbation

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ABSTRACT The cell nucleus is a compartment in which essential processes such as gene transcription and DNA replication occur. Although the large amount of chromatin confined in the finite nuclear space could install the picture of a particularly dense organelle surrounded by less dense cytoplasm, recent studies have begun to report the opposite. However, the generality of this newly emerging, opposite picture has so far not been tested. Here, we used combined optical diffraction tomography and epifluorescence microscopy to systematically quantify the mass densities of cytoplasm, nucleoplasm, and nucleoli of human cell lines, challenged by various perturbations. We found that the nucleoplasm maintains a lower mass density than cytoplasm during cell cycle progression by scaling its volume to match the increase of dry mass during cell growth. At the same time, nucleoli exhibited a significantly higher mass density than the cytoplasm. Moreover, actin and microtubule depolymerization and changing chromatin condensation altered volume, shape, and dry mass of those compartments, whereas the relative distribution of mass densities was generally unchanged. Our findings suggest that the relative mass densities across membrane-bound and membraneless compartments are robustly conserved, likely by different as-of-yet unknown mechanisms, which hints at an underlying functional relevance. This surprising robustness of mass densities contributes to an increasing recognition of the importance of physico-chemical properties in determining cellular characteristics and compartments.

SIGNIFICANCE The cell nucleus has been perceived as a dense compartment, with 2 m of DNA, histones, and other proteins tightly packed into its micron-sized interior, compared with a less dense cytoplasm. Our results from optical diffraction tomography measurements, however, reveal that nucleoplasm is less dense than the cytoplasm, whereas nucleoli exhibit a higher mass density than either. This situation was robust against drastic perturbations of the cytoskeleton and chromatin compaction. This robustness suggests that there might be an underlying functional relevance and yet-unknown mechanisms that maintain these relative mass densities across both membrane-bound and membraneless compartments.

INTRODUCTION

The physical properties of cells and their membrane-bound and membraneless compartments are increasingly becoming the focus of current cell and developmental biological research (1,2). The cell nucleus is a prominent example of a membrane-bound compartment that maintains physical and biochemical conditions distinct from the surrounding cytoplasm. Alterations of nuclear physical properties are associated with various diseases (3). Nuclei also

harbor nucleoli, within which ribosomal subunits are synthesized and assembled for protein translation. Nucleoli are prime examples of membraneless compartments being dynamically maintained by liquid-liquid phase separation (4,5). Because of the high levels of metabolic activity, it might be conceivable that the nucleolus is less dense than the surrounding nucleoplasm, whereas a previous study reported that nucleoli consist of a fibrillar region with higher density than the nucleoplasm surrounded by low-density, sponge-like granular components (6). And, because of the large amount of DNA, histones, and various other types of proteins tightly packed into the finite space of the nucleus, the nucleus has been perceived as a particularly dense organelle compared with the less dense cytoplasm (7–9).

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Recent studies, however, have started to paint a different picture. Using quantitative phase imaging and optical diffraction tomography (ODT), the cell nucleus was reported to have a lower refractive index (RI) than cytoplasm (10–14). Because the RI in most biological materials is linearly proportional to their mass density (15,16), the results indicated that the cell nucleus also has lower mass density than cytoplasm. This finding is supported by other approaches measuring RI, including surface plasmon resonance microscopy (17), transport-of-intensity microscopy (18), and orientation-independent differential interference microscopy (19), but also other indirect approaches to quantify mass density such as Raman microscopy (20). For a generalization of this finding, though, important questions still remain open. In particular, mass density distributions could change during the cell cycle when the genetic material and protein machinery in the nucleus is being duplicated, and more mass added to the nucleus during S/G2 phase. Nuclear volume, and thus density, could depend on the tensional state of the nucleus via the cytoskeleton (21,22). Or, chromatin density might be a direct function of epigenetic markers, such as methylation or acetylation of histone proteins, controlling chromatin condensation (23). Perturbing these aspects permits testing the robustness of the phenomenon.

Here, we systematically investigated the relative densities of nucleoplasm, cytoplasm, and nucleoli in two different adherent mammalian cell types (HeLa-fluorescent-ubiquitination-based-cell-cycle-indicator (FUCCI) and retinal pigment epithelium (RPE)-FUCCI) by quantifying their three-dimensional (3D) RI distributions using combined ODT and epi-fluorescence microscopy. By correlating the RI tomograms of cells with epi-fluorescence images of their nuclei stained with the cell-cycle-dependent FUCCI-fluorescence markers, we quantitatively characterized the RI, mass density, dry mass, and volume of nucleoplasm, nucleoli, and cytoplasm for the different cell cycle phases. We observed that—throughout the cell cycle—nucleoplasm has a lower RI and mass density than cytoplasm. This does not change because the volume of the nucleoplasm and cytoplasm scales with the increase of dry mass because the cell grows and duplicates its genetic material. In contrast, nucleoli inside the nucleus exhibit significantly higher mass density than both nucleoplasm and cytoplasm. This relative mass density distribution between compartments was robust against considerable drug-induced perturbations such as depolymerization of actin and microtubules as well as chromatin condensation and decondensation, even though the shape, density, volume, and dry mass of each compartment of course changed. The robustness of this surprising finding suggests that there might be an underlying functional relevance and that there are as-of-yet unknown active, or passive, mechanisms that maintain relative mass densities across both membrane-bound and membraneless compartments.

MATERIALS AND METHODS

Cell culture and preparation

The stable HeLa-FUCCI and RPE-FUCCI cell lines were kindly provided by the lab of Frank Buchholz (Technische Universität Dresden). FUCCI is a fluorescent ubiquitination-based cell cycle indicator (24). The stable HeLa cell line transfected with GFP:NIFK, which labels nucleolar protein interacting with the FHA domain of pKI-67, was kindly provided by the lab of Anthony Hyman (Max Planck Institute of Molecular Cell Biology and Genetics, Dresden, Germany). All cell lines were cultured under standard conditions at 37°C, 5% CO₂. HeLa cells were cultured in standard Dulbecco's Modified Eagle Medium, high glucose with GlutaMAX medium (61965-026; Thermo Fisher Scientific, Waltham, MA), and RPE cells were cultured in Dulbecco's Modified Eagle Medium/Nutrient Mixture F-12, high glucose with GlutaMAX medium (31331-028; Thermo Fisher Scientific). All culture media were supplemented with 10% FBS and 1% penicillin-streptomycin. The cells were subcultured in a glass-bottom petri dish (FluoroDish; World Precision Instruments, Friedberg, Germany) 1 day before the measurement, and the culture media were exchanged to CO₂-independent medium (18045-088; Thermo Fisher Scientific) before imaging. HeLa-GFP:NIFK cells were stained with Hoechst (1:1000 dilution) for nuclei staining and washed with fresh CO₂-independent medium before imaging.

Drug treatments

To investigate the role of the cytoskeleton on maintaining the RI of the nucleus and cytoplasm, cells were treated with 1 μM cytochalasin D (cytoD) for 30 min or 5 μM nocodazole (noco) for 1 h before imaging to depolymerize actin and microtubules, respectively. To decondense chromatin, cells were incubated with 300 nM trichostatin A (TSA), a histone deacetylase (HDAC) inhibitor, in cell culture medium for 9 h. To condense chromatin, cells were incubated with 100 μM anacardic acid (ANA), a histone acetyltransferase inhibitor, in cell culture medium for 1 h.

Optical setup

The optical setup consisted of a combined ODT and epi-fluorescence microscope (Fig. 1) is as described previously (12). Briefly, ODT employs Mach-Zehnder interferometry to measure the 3D RI distribution of cells (Fig. 1 a). A laser beam ($\lambda = 532$ nm, frequency-doubled Nd-YAG laser; Torus, Laser Quantum, Stockport, UK) was coupled into an optical fiber and divided into two beams by a 2×2 single-mode fiber-optic coupler.

One beam was used as a reference beam and the other beam illuminated the cells on the stage of a custom-made inverted microscope through a tube lens ($f = 175$ mm) and a water-dipping objective lens (numerical aperture (NA) = 1.0, 40 \times ; Carl Zeiss, Oberkochen, Germany). To reconstruct a 3D RI tomogram of cells, the samples were illuminated from 150 different incident angles scanned by a dual-axis galvano-mirror (GVS012/M; Thorlabs, Newton, NJ). The beam diffracted by the sample was collected by a high numerical aperture objective lens (NA = 1.2, 63 \times , water immersion; Carl Zeiss) and a tube lens ($f = 200$ mm). The total magnification was set to be 57 \times . The diffracted beam interfered with the reference beam at an image plane and generated a spatially modulated hologram, which was recorded with a CCD camera (FL3-U3-13Y3M-C; FLIR Systems, Wilsonville, OR). To maintain the temperature of the culture medium in the glass-bottom petri dish, both objective lenses were heated at 37°C by resistive foil heaters (Thorlabs).

Epi-fluorescence imaging was performed using the same optical setup (Fig. 1 b). To excite fluorescence probes in cells, an incoherent beam from a halogen lamp (DC-950; Dolan-Jenner Industries, Boxborough, MA) was coupled into the same beam path using a three-channel dichroic mirror (FF409/493/596-Di01-25 \times 36; Semrock, Rochester, NY). The excitation wavelength was selected by alternating bandpass filters corresponding to Hoechst, monomeric Azami-Green 1, and monomeric Kusabira-Orange 2.

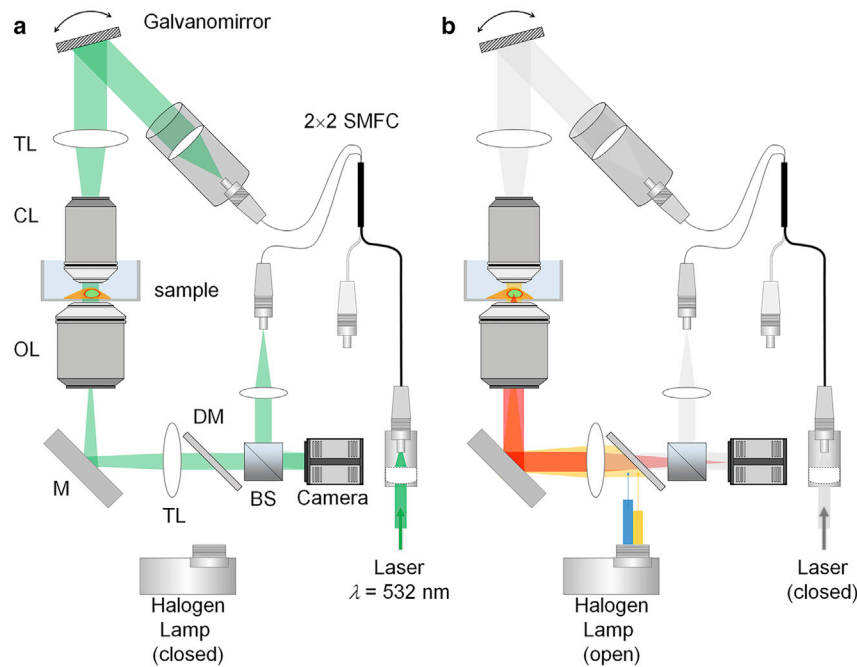


FIGURE 1 Experimental setup. (a) Shown is the optical setup for optical diffraction tomography (ODT). (b) Shown is the same optical setup when used for epi-fluorescence microscopy. To see this figure in color, go online.

Tomogram reconstruction and quantitative analysis

The complex optical fields of light scattered by the samples were retrieved from the recorded holograms by applying a Fourier-transform-based field retrieval algorithm (25). The 3D RI distribution of the samples was reconstructed from the retrieved complex optical fields via the Fourier diffraction theorem (26,27). A more detailed description of tomogram reconstruction can be found elsewhere (28,29). The spatial resolution of ODT is 121 nm (lateral) and 444 nm (axial), which was determined by the NAs of the objective lens and condenser lens (30). The precision of the ODT was characterized by measuring the standard error (SE) of RI tomograms of the same field of view with repetitive measurements (Fig. S1). We measured a time series of 10 RI tomograms of silica beads immersed in 0.7 M sucrose solution ($n = 1.3665$ at $\lambda = 532$ nm) in the same field of view with the tomogram acquisition rate of 1 Hz. We calculated the standard error of the time series of 10 RI tomograms and averaged the standard error within an individual silica bead. The averaged standard error of RI was calculated as 4.15×10^{-5} , which can be converted to the error of the absolute mass density in cells as 0.22 mg/mL.

From the reconstructed tomograms (Fig. 2 a), Otsu's thresholding method (31) was used to segment the regions occupied by cells from background and the watershed algorithm to identify individual cells from segmented RI tomograms. Then, epi-fluorescence images of nuclei stained with the FUCCI marker (Fig. 2 b) were correlated with the RI tomograms to segment nuclei inside cells. We also segmented the periphery of nuclei, i.e., a perinuclear cytoplasm where the endoplasmic reticulum and Golgi apparatus are dominant, by dilating the binary nuclei masks by $2\text{-}\mu\text{m}$ thickness. Inside the 3D RI distribution of nuclei, regions having higher RI-values than surrounding nucleoplasm were segmented by Otsu's thresholding method and identified as nucleoli. We confirmed that nucleoli had higher RI than nucleoplasm by correlating the RI tomograms of HeLa cells with epi-fluorescence images in which nuclei and nucleoli were stained with Hoechst and GFP, respectively. The number of voxels in the region of nucleoli segmented by this RI-based method had 99.5% true positive rate (sensitivity) compared with the epi-fluorescence images of nucleoli, and the nucleoplasm could be segmented with 88.9% sensitivity (see Fig. S2). Finally, we quantified the 3D RI distribution of cytoplasm (n_c), perinuclear cytoplasm (n_{pc}), nucleoplasm (n_{np}), and nucleoli (n_n) separately from the segmented tomogram (Fig. 2, c-f).

The mass density of each compartment was directly calculated from the mean RI-value because the RI-value in biological samples, $n(x, y, z)$, is linearly proportional to the mass density of the material, $\rho(x, y, z)$, as $n(x, y, z) = n_m + \alpha\rho(x, y, z)$, where n_m is the RI-value of the surrounding medium, and α is an RI increment (dn/dc). The RI of the medium was measured using an Abbe refractometer (2WJ; Arcarda, Frankfurt, Germany) to be $n_m = 1.3370 \pm 0.00025$ at $\lambda = 532$ nm. The RI increment used was $\alpha = 0.190$ mL/g for the protein and nucleic acid (32,33). In addition, the volume of the compartments was extracted by counting the number of voxels in the corresponding binary mask, and the dry mass of the compartments was calculated by integrating the mass density inside the corresponding binary mask. The sphericity of the compartments, Ψ , was calculated as the ratio between surface area, A , and volume of the mask, V , as $\Psi = [\pi^{1/3}(6V)^{2/3}]/A$. All tomogram acquisition and data analysis were performed using custom-written MATLAB scripts (R2017b; The MathWorks, Natick, MA), which are available in GitHub (<https://github.com/OpticalDiffractionTomography/NucleiAnalysis>). The datasets analyzed are available on Figshare (<https://doi.org/10.6084/m9.figshare.12649592>).

Statistical analysis

We tested the normality of the RI distributions and the RI ratios using the Shapiro-Wilk test, which revealed that most distributions were normal. We report means and standard error of means for the data passing the normality test and means when the data did not pass the normality test. Statistical significance between data groups was determined using either the two-tailed Student's t -test or the two-tailed Mann-Whitney U test when the data did not pass the normality test. The shown asterisks indicate the statistical significance as $*p < 0.01$, $**p < 0.001$, and $***p < 0.0001$, respectively.

RESULTS

Relative compartment RI and mass densities are independent of cell cycle and cell types

To investigate the effect of the cell cycle on the RI distribution of cytoplasm, nucleoplasm, and nucleoli, we correlated

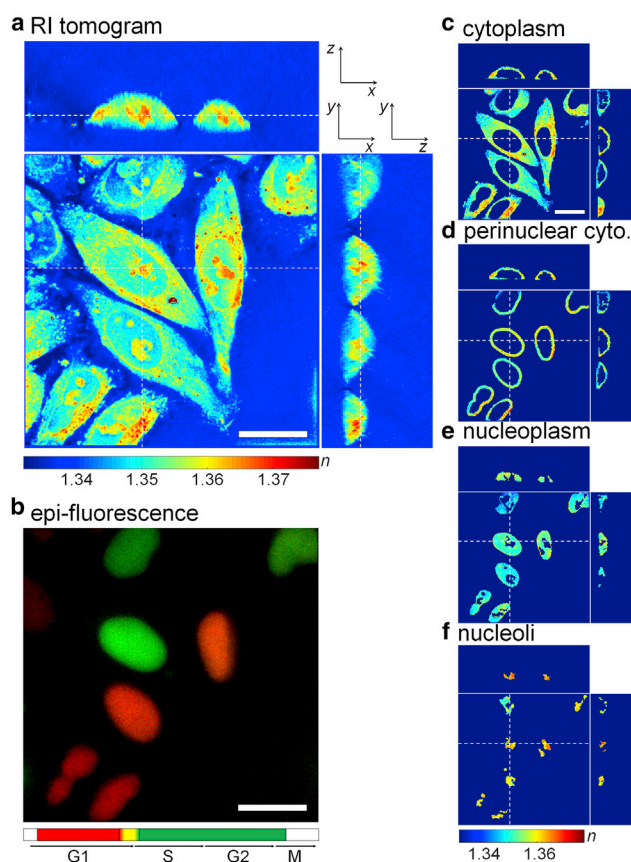


FIGURE 2 Segmentation of cellular and nuclear compartments from a refractive index (RI) tomogram and epi-fluorescence map. (a) Shown are the cross-sectional slices of an RI tomogram of HeLa-FUCCI cells in the x - y , x - z , and y - z planes. The color scale quantifies the RI, n . (b) Shown is the epi-fluorescence map of the same field of view as in (a). Red color identifies cells in the G1 phase of the cell cycle, yellow in early S, and green in S/G2. (c–f) Shown are the RI maps segmented for (c) cytoplasm, (d) perinuclear cytoplasm, (e) nucleoplasm, and (f) nucleoli. Dashed lines indicate the corresponding x - z and y - z planes. Scale bars, 20 μm . To see this figure in color, go online.

the reconstructed RI tomograms of two adherent cell lines, HeLa-FUCCI and RPE-FUCCI cells, with the respective FUCCI-fluorescence images. We grouped each cell into the G1, early S, and S/G2 phases of the cell cycle ($N = 557$, 505, and 483 for HeLa cells, and $N = 122$, 92, and 128 for RPE cells in the G1, early S, and S/G2 phases, respectively) by the average intensity of their epi-fluorescence images in the mAG1 and mKO2 channels (see Fig. S3 for details). As shown in the typical examples in Fig. 3, *a*, *b*, *d*, and *e*, the cell nucleoplasm of both HeLa and RPE cells had lower RI-values compared with the cytoplasm in all phases of the cell cycle. Staining with FUCCI dyes did not affect the RI of the compartments because non-labeled HeLa cells also showed similar RI distribution (Fig. S4).

For a more quantitative analysis, we calculated the mean RI-values, and the mean mass densities, of cytoplasm, peri-

nuclear cytoplasm (as a part of the cytoplasm), nucleoplasm, and nucleoli from the segmented RI tomograms of individual cells (see Materials and Methods; Fig. S5). We found that the nucleoplasm of HeLa cells was 7.3 ± 0.6 , 5.8 ± 0.6 , and $5.7 \pm 0.6\%$ (mean \pm SE) less dense than the cytoplasm at the G1, early S, and S/G2 phases, respectively (Fig. 3 *c*). The absolute mass density differences of 6.0 ± 0.5 , 4.9 ± 0.5 , and 4.7 ± 0.5 mg/mL were statistically significant (see Fig. S5, *a* and *b*). The immediate periphery of the nucleus, i.e., the perinuclear cytoplasm, where the membrane-rich endoplasmic reticulum and Golgi apparatus are dominant, had an even higher RI than other cytoplasmic regions (Fig. S5, *a* and *b*). Hence, the relative difference in mass density between cell nucleoplasm and the perinuclear cytoplasm was even more pronounced (10.8 ± 0.5 , 10.6 ± 0.4 , and $10.3 \pm 0.4\%$ at the G1, early S, and S/G2 phases, respectively; Fig. 3 *c*).

The RPE cells, as a noncancerous cell line, showed similar results because nucleoplasm was 10.5 ± 2.9 , 8.7 ± 3.9 , and $13.3 \pm 2.7\%$ less dense than cytoplasm and 18.8 ± 2.0 , 16.4 ± 2.3 , and $19.6 \pm 1.6\%$ less dense than the perinuclear cytoplasm at the G1, early S, and S/G2 phases, respectively (Fig. 3 *f*). The absolute mass density differences between nucleoplasm and cytoplasm (4.0 ± 1.2 , 3.7 ± 1.8 , and 5.2 ± 1.1 mg/mL in the three respective cell cycle phases) was again statistically significant (see Fig. S5, *c* and *d*). The lower density of the nucleoplasm compared with the surrounding cytoplasm, thus, seems to be cell type independent.

In this study, we used the same RI increment, $\alpha = 0.190$ mL/g, to calculate the mass density of each compartment from measured RI, which is valid for protein and nucleic acid (32,33). However, because the RI increment of phospholipid is lower as $\alpha = 0.135$ – 0.138 mL/g (34,35), the mass density of such membrane-rich perinuclear cytoplasm may be underestimated and have even higher mass density. Nonetheless, nucleoplasm is still less dense than cytoplasm without the perinuclear area under this consideration (the absolute mass density differences of 3.9 ± 1.0 , 3.4 ± 1.0 , and 3.8 ± 0.9 mg/mL for HeLa cells and 2.8 ± 1.7 , 1.8 ± 2.1 , and 3.8 ± 1.5 mg/mL for RPE cells in the three respective cell cycle phases, Fig. S5).

During the cell cycle, cells are growing and duplicating genetic material, which may increase the dry mass, volume, and/or mass density of the nucleus and cytoplasm and could therefore alter our findings. However, both the volume and dry mass of both nucleoplasm and cytoplasm in HeLa and RPE cells increased twofold during the cell cycle so that the mass density of both compartments was maintained at about the same level (see Fig. S6). This result, obtained with two different attached cell lines, is consistent with our previous study in suspended cells (36) and implies that the volume, or the mass density, of the nucleoplasm and cytoplasm is well controlled in response to the increase of cellular content during the cell cycle.

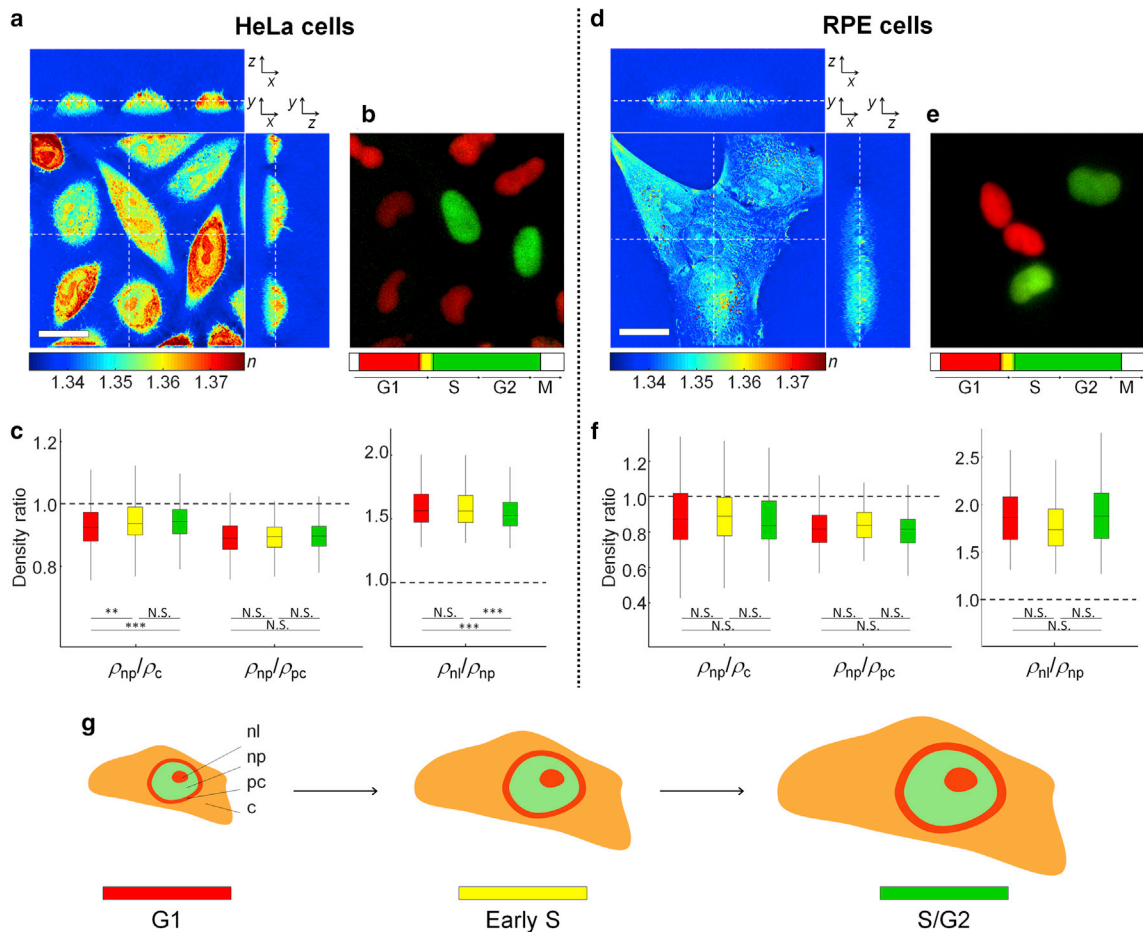


FIGURE 3 RI and mass density distributions in HeLa-FUCCI (*a–c*) and RPE-FUCCI (*d–f*) cells as a function of the cell cycle. (*a* and *d*) Shown are cross-sectional slices of 3D RI tomograms of cells in the x - y , x - z , and y - z planes. The dashed lines indicate the corresponding x - z and y - z planes. The color scale quantifies the RI, n . Scale bars, 20 μm . (*b–e*) Shown are the corresponding FUCCI-fluorescence images of nuclei in different cell cycle phases. Red color identifies cells in the G1 phase of the cell cycle, yellow in early S, and green in S/G2. (*c–f*) Given is the ratio of the mass density between nucleoplasm/cytoplasm (ρ_{np}/ρ_c), nucleoplasm/perinuclear cytoplasm (ρ_{np}/ρ_{pc}), and nucleoli/nucleoplasm (ρ_{nl}/ρ_{np}). The dashed lines indicate equal mass density between compartments. The colors indicate cell cycle phases as in (*b*) and (*e*). The numbers of cells measured are $N = 557$, 505, and 483 for HeLa cells and $N = 122$, 92, and 128 for RPE cells in the G1, early S, and S/G2 phases, respectively. (*g*) Given is a schematic indicating the four segmented regions and how they scale in size to maintain the same relative mass densities throughout the cell cycle. To see this figure in color, go online.

Interestingly, the nucleoli inside the nucleoplasm, where ribosomal subunits are synthesized for protein translation, had significantly higher mean RI and density than the nucleoplasm (58% in HeLa cells and 92% in RPE cells) and the cytoplasm (48% in HeLa cells and 71% in RPE cells) (Fig. 3, *c* and *f*; Fig. S5, *b* and *d*). Here, their relative density ratio distribution exhibited the non-normal distribution. Moreover, the averaged mass density of the entire nucleus, including nucleoplasm and nucleoli, is similar to that of cytoplasm and perinuclear cytoplasm in HeLa cells or even higher than cytoplasm in RPE cells (Fig. S7). Recently, several reports have described the nucleolus as a membraneless organelle, which was proposed to assemble by liquid-liquid phase separation (4,5). Phase separation is a density transition in which two phases—one dilute and one dense phase—stably coexist. Our findings suggest that cells may

redistribute the mass within the nucleus by phase separation and formation of the nucleolus and that this generates the relative mass density difference between nucleoplasm and cytoplasm. In addition, our results demonstrate the narrow distributions of the mass density ratio, and difference, between nucleoli and nucleoplasm, which suggests that nucleoli formation and ribosome biogenesis are precisely regulated in these membraneless organelles during the cell cycle.

Changes due to cytoskeletal perturbations do not abolish relative mass density differences

In the previous section, we have confirmed that nucleoplasm has lower mass density than cytoplasm. We found the same pattern in various cell types, both suspended (13,36) and

attached, and throughout the cell cycle. Apparently, in physiological conditions, the pattern is robustly conserved. But can we identify the origin of this pattern by specific perturbations?

There is evidence that the nucleus is under tension from the cytoskeleton, which in turn is anchored to the cell membrane (21,37). Is it thus possible to reduce the volume of the nucleus, enclosing a relatively fixed amount of solid material, by perturbing the tensional balance between cell surface and nucleus to increase its density? cytoD inhibits actin polymerization and destabilizes the actin cortex, whereas noco depolymerizes microtubules and, at the same time, activates Rho signaling and increases actomyosin contractility (38). Visual inspection of the reconstructed RI tomograms (Fig. 4, *a–c*) showed that HeLa-FUCCI cells treated with cytochalasin D exhibited dramatic morphological changes at the cell boundary (Fig. 4 *b*). Cells treated with nocodazole seemed to be larger and exhibited lower RI-values in the cytoplasm (Fig. 4 *c*).

Quantitatively, cells treated with cytochalasin D had higher mean RI-values in all subcellular compartments compared with controls, whereas nocodazole-treated cells had lower mean RI-values in all compartments (Fig. 4 *d*). Particularly, the mean RI-values of cytoplasm and nucleoplasm under the cytoD treatment were 1.3571 ± 0.0002 and 1.3563 ± 0.0002 (mean \pm SE, $N = 973$) and thus

higher than controls (1.3538 ± 0.0002 and 1.3528 ± 0.0002 for cytoplasm and nucleoplasm, $N = 1565$). As shown in Fig. 4 *e*, this increase of mean RI-values in the cytoD-treated cells resulted from different physical phenomena. The mean RI-value and mass density of nucleoplasm increased as the volume occupied by nucleoplasm decreased $11.9 \pm 2.4\%$ compared with controls, whereas its dry mass only increased by $5.6 \pm 2.7\%$. In contrast, the mean mass density of cytoplasm increased because of the $19.4 \pm 1.9\%$ increase of cytoplasmic dry mass, whereas its volume remained the same ($1.3 \pm 1.7\%$ increases). It is also worth noting that cytoD-treated cells became more spherical as the sphericity of cells increased by $5.0 \pm 0.6\%$, whereas the nucleus maintained the same sphericity ($0.6 \pm 2.0\%$ increases). The sphericity of the entire cell can be increased by the detachment from the substrate because of depolymerized actin filaments at the cell boundary, whereas constant nuclear sphericity during the shrinkage of a nucleus implies that the nuclear volume shrinks uniformly in all directions. These percentage changes are averaged ratios for each cell cycle phase, each of which shows the same change (see Fig. S8).

Nuclear shrinkage by the cytochalasin D treatment is as expected because cytochalasin D dissociates the actomyosin cytoskeleton, which normally exerts outward tension on the nucleus (39–41). The treatment, then, increased the mass

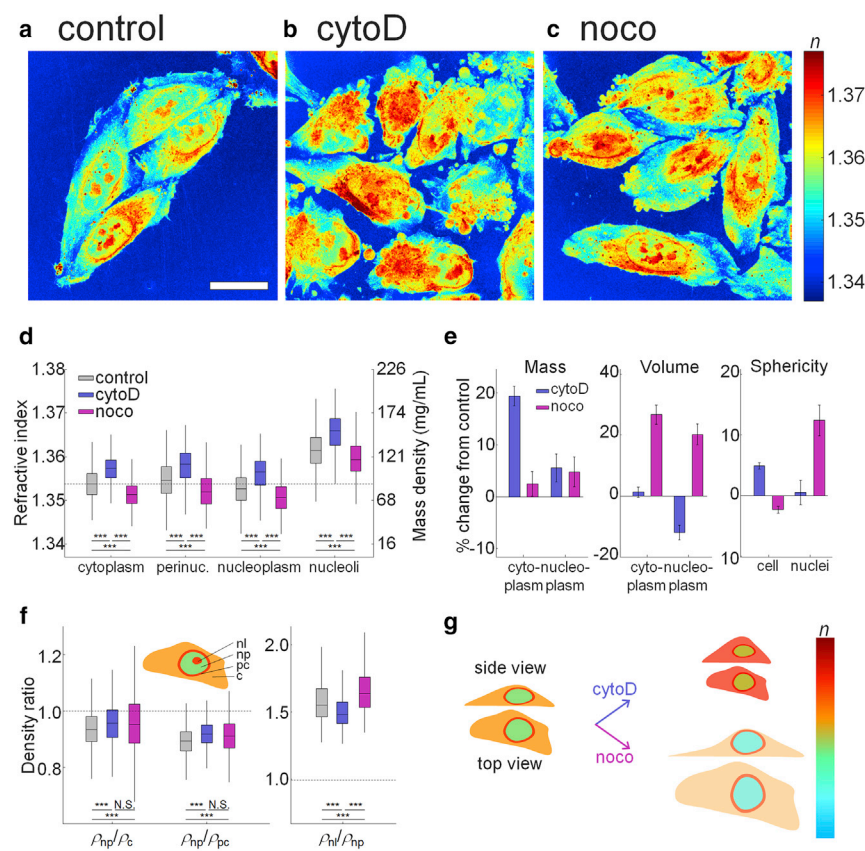


FIGURE 4 Changes in RI distribution of HeLa-FUCCI cells under cytoskeletal perturbation. (*a–c*) Shown is the maximal projection of RI tomograms of HeLa-FUCCI cells in (*a*) control, (*b*) cytoD, and (*c*) noco treatment. Scale bars, 20 μ m. (*d*) Given are the mean RI-values of cytoplasm, perinuclear cytoplasm, nucleoplasm, and nucleoli in individual cells under cytoskeletal perturbation. The dashed line indicates the mean RI-value of the cytoplasm in control cells. (*e*) Shown are the relative changes of the mass, volume, and sphericity of the cytoplasm and nuclei under cytoD and noco treatment compared with control. (*f*) Shown is the mass density ratio between nucleoplasm/cytoplasm (ρ_{np}/ρ_c), nucleoplasm/perinuclear cytoplasm (ρ_{np}/ρ_{pc}), and nucleoli/nucleoplasm (ρ_{nl}/ρ_{np}) at the same phases of the cell cycle. The dashed lines indicate equal mass density of the compartments. The numbers of cells measured are $N = 1565$, 973, and 717 for control, cytoD, and noco treatments, respectively. (*g*) Given is a schematic indicating the effect of the cytoD and noco treatment on the volume, RI, and shape of cells. To see this figure in color, go online.

density of nucleoplasm because of the reduced volume at constant dry mass enclosed. At the same time, the mass density of cytoplasm increased because of the increase of its dry mass at about constant volume. Even though there are previous reports that cytochalasin D treatment promotes the synthesis of actin (42) and other proteins (43), it is unclear whether this is the actual mechanism because 30 min of treatment time seems too short to account for a 20% mass increase. The actomyosin network can also form a perinuclear actin cap and exert compressive forces on the nucleus in the axial direction, in particular on nuclei in stromal cells (44,45). In this study, a compressive force on the nucleus by the actomyosin network is probably not relevant because the perinuclear actin cap is absent in HeLa cells (46–48). Taken together, we found that nucleoplasm still had lower mass density than cytoplasm and perinuclear cytoplasm (4.6 ± 0.5 and $8.0 \pm 0.3\%$) (Fig. 4 f).

This robustness of the relative mass density between compartments was also observed in cells treated with nocodazole, whereas the mass RI-values of cytoplasm and nucleoplasm decreased as 1.3515 ± 0.0002 and 1.3509 ± 0.0003 (mean \pm SE, $N = 717$), respectively (1.3538 ± 0.0002 and 1.3528 ± 0.0002 for cytoplasm and nucleoplasm in controls). The decreases of mean RI-value and the mass density of cytoplasm and nucleoplasm mostly originated from their volume expansion (26.7 ± 3.1 and $20.2 \pm 3.4\%$ in cytoplasm and nucleoplasm, respectively), whereas their dry mass only slightly increased as 2.5 ± 2.3 and $4.8 \pm 2.9\%$ (Fig. 4 e). The nuclear volume expansion and increased nuclear sphericity in response to the nocodazole treatment can be rationalized based on previous studies because the microtubules are depolymerized by the nocodazole treatment. Microtubules normally confine the nucleus in the direction perpendicular to the cell culture substrate (41,49,50). The effect can be also observed from the increased sphericity of the nucleus increased by $12.4 \pm 2.5\%$, whereas the cell sphericity slightly decreased by $2.3 \pm 0.6\%$ (Fig. 4 e). The volume increase of the nucleus at a constant dry mass of the nucleoplasm then led to a decrease of the nucleoplasm mass density. Interestingly, at the same time, the mass density of cytoplasm also decreased because of the increase of its volume as nocodazole inhibits the normal regulatory volume decrease (49), which in sum again conserved the relative mass density difference between nucleoplasm and cytoplasm. Quantitatively, the nucleoplasm of the nocodazole-treated cells was still $4.3 \pm 0.8\%$ less dense than the cytoplasm and $8.5 \pm 0.5\%$ less dense than perinuclear cytoplasm (Fig. 4 f).

Our finding shows that the relative mass density between nucleoplasm and cytoplasm is still maintained against the perturbation of cytoskeletal integrity, which is graphically summarized in Fig. 4 g. We also revealed that the mass density difference across the nuclear membrane of cytochalasin-D- and nocodazole-treated cells was 8.8 ± 0.4 and 7.1 ± 0.4 mg/mL, which are slightly lower than that of control cells

as 10.2 ± 0.3 mg/mL. It may imply that cytoskeletal perturbation may induce a subtle force imbalance between the osmotic pressure gradient across the nuclear membrane and the force exerted by the cytoskeleton (50–52).

Perturbing chromatin condensation still does not overturn relative mass density differences

If perturbing the cytoskeleton and the tensional balance between nucleus and cell boundary does not alter relative mass densities, what about changing chromatin compaction directly? After all, chromatin compaction is linked to transcriptional activity and controlled by epigenetic marks, which can be specifically altered using drugs.

We treated HeLa-FUCCI cells with TSA or ANA and measured their RI tomograms. TSA decondenses chromatin by inhibition of HDACs (23), whereas ANA condenses chromatin by inhibition of histone acetyltransferases (53). As shown in Fig. 5, a–c for the visual inspection and in Fig. 5 d and Fig. S9 for the quantitative analysis, the TSA treatment decreased the RI of each cellular compartment, whereas the ANA treatment induced the opposite effect.

Quantitatively, the mean RI-values of cytoplasm and nucleoplasm treated by TSA were lower than controls as 1.3516 ± 0.0004 and 1.3500 ± 0.0004 (1.3538 ± 0.0002 and 1.3528 ± 0.0002 for cytoplasm and nucleoplasm in controls). The decrease of RI and mass density of nucleoplasm in TSA-treated cells was mostly contributed from the significant swelling of nucleoplasm ($28.0 \pm 6.4\%$), whereas its dry mass exhibited subtle changes ($-2.7 \pm 3.9\%$) (Fig. 5 e). We also found that the dry mass of cytoplasm increased significantly by $23.2 \pm 3.8\%$, whereas the extensive volume increase of cytoplasm by $54.0 \pm 5.5\%$ lowered the RI-value of cytoplasm. The cellular and nuclear sphericity also decreased by 5.9 ± 0.9 and $5.0 \pm 3.0\%$ under the TSA treatment, which could be the result of increased volume at constant cell height. These results indicate that increased histone acetylation increases nuclear volume (54,55) and also suggest that the increase of euchromatin in nucleoplasm under the TSA condition enhances the protein synthesis in cytoplasm and is accompanied by an increase of cytoplasmic dry mass. This view is supported by previous findings reporting that TSA treatment increases mRNA expression by twofold (55). In contrast, the mean RI-values of cytoplasm and nucleoplasm under the ANA treatment were higher than in control cells as 1.3567 ± 0.0002 and 1.3560 ± 0.0002 , respectively. The increases of cytoplasmic and nuclear RI under the ANA treatment are mainly due to increases in dry mass as 23.1 ± 2.2 and $17.7 \pm 2.9\%$, respectively, whereas their volume increases were insignificant (8.0 ± 2.0 and $1.9 \pm 2.6\%$), as shown in Fig. 5 e. It can be speculated that perturbing chromatin condensation not only affects the density of nucleoplasm but also alters gene expression and protein synthesis. Moreover, HDACs and histone acetyltransferases can lead to the acetylation/deacetylation of nonhistone proteins such

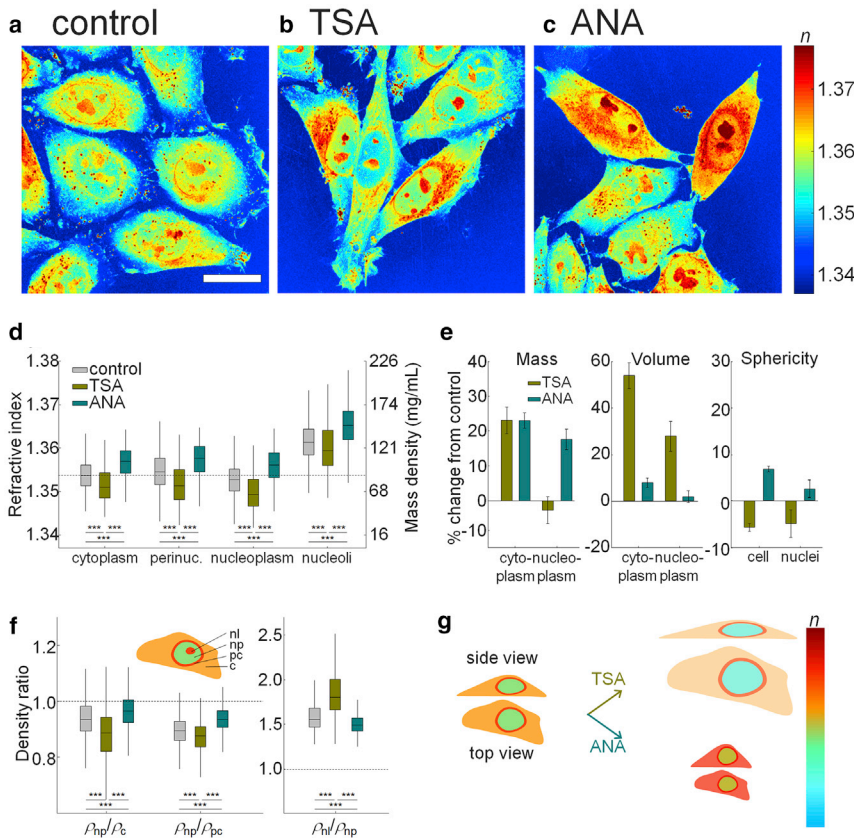


FIGURE 5 Changes in RI distribution of HeLa-FUCCI cells under chromatin condensation. (a–c) Shown is the maximal projection of RI tomograms of HeLa-FUCCI cells in (a) control, (b) TSA, and (c) ANA treatment. Scale bars, 20 μm . (d) Given are the mean RI-values of cytoplasm, perinuclear cytoplasm, nucleoplasm, and nucleoli in individual cells under chromatin condensation. The dashed line indicates the mean RI-value of cytoplasm in control cells. (e) Given are the relative changes of the mass, volume, and sphericity of the cytoplasm and nuclei under TSA and ANA treatment compared with control. (f) Shown is the mass density ratio between nucleoplasm/cytoplasm (ρ_{np}/ρ_c), nucleoplasm/perinuclear cytoplasm (ρ_{np}/ρ_{pc}), and nucleoli/nucleoplasm (ρ_n/ρ_{np}) at the same phases of the cell cycle. The dashed lines indicate equal mass density of the compartments. The numbers of cells measured are $N = 1565, 437,$ and 924 for control, TSA, and ANA treatments, respectively. (g) Given is a schematic indicating the effect of the TSA and ANA treatment on the volume, RI, and shape of cells. To see this figure in color, go online.

as microtubules (56–58). Thus, the inhibition of those histone-modifying enzymes can modify the stability of microtubules for regulating cell volume, which may also indirectly affect the volume of cytoplasm and nucleoplasm.

Interestingly, despite the direct effects on chromatin condensation, nucleoplasm was still less dense than cytoplasm (Fig. 5 f). Under the TSA treatment, nucleoplasm was $11.8 \pm 0.9\%$ less dense than cytoplasm, which was more pronounced than in control cells ($6.3 \pm 0.4\%$). In contrast, nucleoplasm under the ANA treatment was $3.9 \pm 0.9\%$ less dense than cytoplasm. In addition, nucleoplasm under the TSA and ANA treatment was 12.7 ± 0.6 and $6.6 \pm 0.3\%$ less dense than perinuclear cytoplasm, respectively, which were equivalent to 10.0 ± 0.5 and 7.0 ± 0.3 mg/mL of mass density difference across the nuclear membrane. Our finding is graphically summarized in Fig. 5 g.

DISCUSSION

In this study, we investigated the mass densities of nucleoplasm, cytoplasm, and nucleoli of adherent cells in various conditions, including cell cycle progression and perturbations of cytoskeleton polymerization and chromatin condensation. We employed combined ODT and epi-fluorescence microscopy to measure 3D RI distributions of cells from which the mass density, volume, and dry mass of nucleoplasm, nucleoli, and cytoplasm were quantitatively characterized.

We found that nucleoplasm was less dense than cytoplasm in the majority of populations of adherent HeLa (82.6%) and RPE (76.5%) cells. This relative mass density difference was maintained during the cell cycle by scaling the volume of both compartments with the increase of dry mass of genetic and cellular materials. Moreover, this relative mass density difference was robust against drug-induced perturbations, including depolymerization of actin and microtubules as well as chromatin condensation and decondensation.

This ODT provides RI tomograms of individual cells and subcellular organelles with a spatial resolution of 121 nm (lateral) and 444 nm (axial), which are determined by the NAs of the objective lens and condenser lens (30). Spatial resolution is an important aspect of light microscopy techniques. At present, many techniques are published that push the boundary of spatial resolution way beyond what was thought possible because of the diffraction limit (down to nanometers) (59). However, all these techniques rely on fluorescence labels and cannot directly assess the physical properties of the sample. Also, ODT has better resolution than a typical brightfield or phase contrast microscope because the sample is illuminated from many different angles, effectively increasing the NA and thus spatial resolution (27,30). Another measure of quality in

any physical measurement is the precision of the measurement. In our ODT setup, the systematic errors arise from phase measurement, fast Fourier transform, and a missing cone artifact for tomogram reconstruction. Altogether, the precision of our RI measurement comes down to 4.15×10^{-5} , which is sufficient to pick up the differences between the various regions of the cell characterized in the study.

The regulation of volume and dry mass in cell growth is essential for maintaining homeostasis in biological samples, and our finding confirms that the mass density of both nucleoplasm and cytoplasm are maintained during cell cycle progression. Whether this is caused by an active or passive mechanism is unclear at this point and remains to be determined. The result is in agreement with previous studies reporting that the buoyant density of nuclei is independent of the cell cycle (60,61). At present, we do not know what the functional importance of this particular mass density distribution is, but the mere fact that it is surprisingly stable against perturbations suggests that there is one. Moreover, we revealed that the relative mass density difference between nucleoplasm and nucleoli is also maintained during the cell cycle. This finding may imply that the chemical potential for nucleoli formation is maintained during the cell cycle, which governs formation of nucleoli by liquid-liquid phase separation (1). Interestingly, the averaged mass density of the nucleus, including nucleoplasm and nucleoli, is comparable to that of cytoplasm and perinuclear cytoplasm. Hence, the result may suggest that the redistribution of mass inside the nucleus is caused by the formation of nucleoli, thus reducing the mass density of nucleoplasm. This could be further investigated by measuring the mass density of nucleoplasm and nucleoli whose material properties can be modulated by an optogenetic gelation strategy (62). Recent studies have shown that the cell growth rate is size dependent and exhibits exponential growth (63–65), and the size of nucleoli scales with that of cells, whereas nuclei grow slower in the development of *C. elegans* embryo (66). We expect that time-resolved ODT measurements of nuclei and nucleoli during cell cycle progression can reveal the effect of the mass density of both compartments on the growth rate during the cell cycle. It will also be interesting to look specifically at the beginning of mitosis, when nucleoli dissolve, or at cells in developing embryos when there are no nucleoli yet.

We also discovered that nucleoplasm was still less dense than cytoplasm and perinuclear cytoplasm even when cytoskeleton polymerization or chromatin condensation was perturbed by drug treatments. In addition, we revealed that the mass density difference between nucleoplasm and cytoplasm exhibited subtle changes on the order of few milligrams per milliliter against the drug-induced perturbations. Perhaps these slight alterations of mass density difference result from mechanical force imbalances on the nuclear membrane in response to perturbations of cytoskeletal polymerization and chromatin condensation. On

nuclear membrane, an osmotic pressure gradient generated by the mass density difference between nucleoplasm and perinuclear cytoplasm is mechanically balanced with pressure exerted by the cytoskeleton. The cytoskeletal pressure consists of a compressive (inward) component contributed by microtubules and a tensile (outward) pressure by the actomyosin cytoskeleton. There is also an additional elastic component from the mechanical stiffness of the nuclear envelope (21,50–52,67,68). The osmotic pressure in solution, Π , is determined by the Van't Hoff equation as $\Pi = \rho RT/M$, where ρ is the mass density of solutes, R is the gas constant ($8.314 \text{ J} \cdot \text{mol}^{-1} \cdot \text{K}^{-1}$), T is the temperature of the solution, and M is the molecular weight of solutes. Hence, we can imagine that the slight mass density changes in two compartments alter their volume fraction (ρ/M) difference across the nuclear membrane under various perturbations, which can modify the osmotic pressure difference on the nuclear envelope, adding to the pressure exerted by the cytoskeleton. One way to decode the underlying biophysical principle of how and why nucleoplasm maintains a lower mass density than the surrounding cytoplasm could be to manipulate the osmotic pressure in the nucleoplasm and cytoplasm experimentally (69,70) and determining the changes of the mass density of the compartments.

The nucleus is a very interesting organelle from a materials perspective. It combines a highly packed, topologically constrained, and highly charged polymeric material in its bulk with an internally and externally interconnected envelope and is subject to many active processes (71). Some of its internal structure also derives from liquid-liquid phase separation. Thus, it does not have to be surprising that density is not linearly related to stiffness. A simple gedankenexperiment might help to illustrate the point: cross-linked and non-cross-linked polymer gels have the same density but clearly different elasticity and viscosity. The nucleus has even been shown to behave like an auxetic material with a negative Poisson ratio (72). So, our finding of a low mass density is not in conflict with the fact that it generally has a high elastic modulus. Moreover, the nucleus mechanically interacts with the surrounding cytoplasm and exterior environment via the cytoskeleton. The effects of the nuclear and cytoplasmic mass densities on their mechanical properties, and vice versa, is still unexplored primarily because of the lack of practical techniques to measure local mechanical properties inside cells. There are emerging noninvasive optical microscopic techniques to probe the mechanical properties of biological samples including Brillouin microscopy (52,73,74), fluorescence lifetime imaging (75,76), and time-lapse quantitative phase microscopy (77). In the future, combining ODT with such other microscopic modalities can reveal the interaction between the mass density and mechanical properties of nucleus and cytoplasm.

To conclude, we have combined ODT and epi-fluorescence microscopy to investigate the mass density of nucleoplasm, cytoplasm, and nucleoli of adherent cells. We revealed that

nucleoplasm is less dense than cytoplasm, whereas nucleoli have significantly higher mass density than nucleoplasm. Moreover, the relative mass density difference between the compartments was robustly maintained during cell cycle progression and upon perturbations of cytoskeletal integrity and chromatin condensation. Biological cells are increasingly understood as physical objects amenable to physical modeling, and the mass density of a particular region or organelle in a cell might appear as a fundamental physical parameter in such physical models. Thus, providing quantitative values for such physical parameters has a value in itself because it can then be incorporated in the physical models. Although the physiological relevance of the phenomenon of lower mass density of the nucleoplasm versus the cytoplasm needs further investigation, our findings based on the quantitative characterization may have implications for understanding cell homeostasis during the cell cycle, the mechanical interaction between nucleus and cytoplasm via the cytoskeleton, and nuclear volume regulation. Further studies are also required to identify whether the robustness of the relative mass densities is due to active regulatory mechanisms by the cell or whether they are an emerging result of basic physico-chemical properties of the cell.

SUPPORTING MATERIAL

Supporting Material can be found online at <https://doi.org/10.1016/j.bpj.2020.08.044>.

AUTHOR CONTRIBUTIONS

K.K. realized the combined optical setup for ODT and epi-fluorescence microscopy and conducted the measurements. K.K. and J.G. designed the experiments, interpreted the results, and wrote the manuscript.

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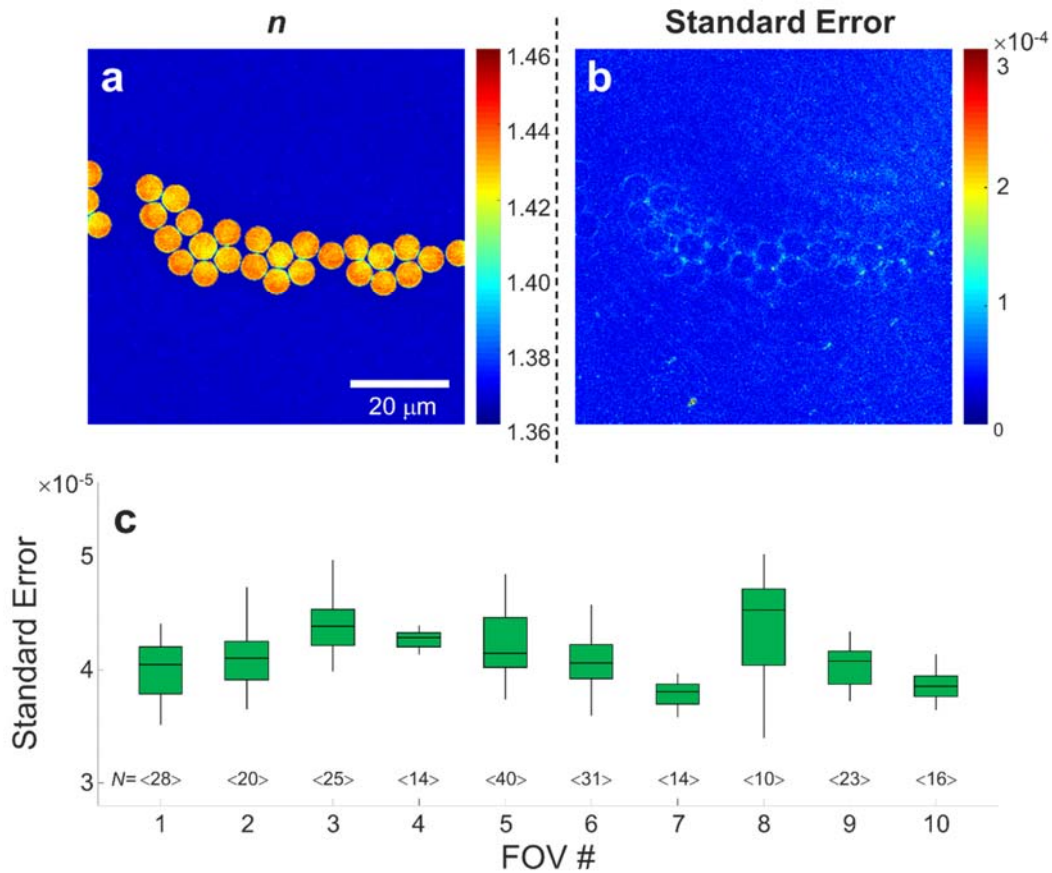
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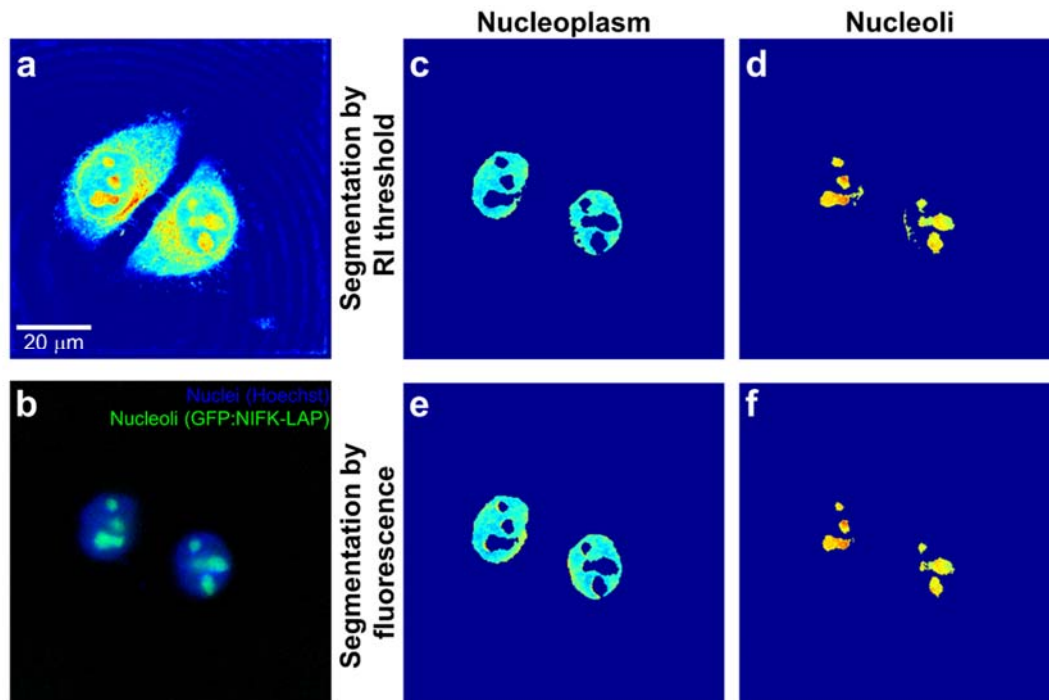
Supplemental Information

**The Relative Densities of Cytoplasm and Nuclear Compartments Are
Robust against Strong Perturbation**

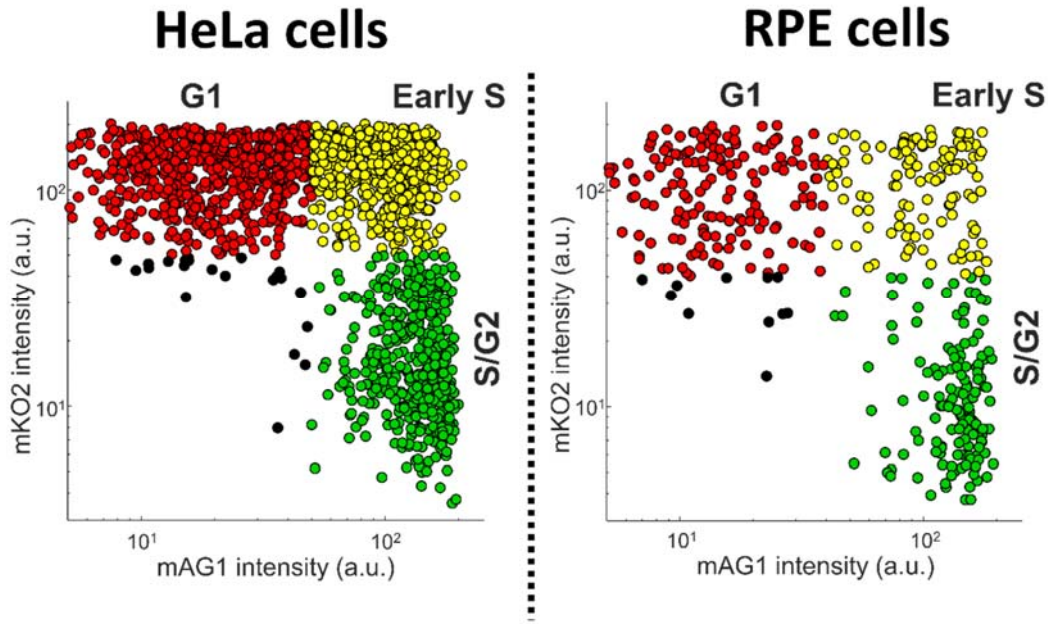
Kyoohyun Kim and Jochen Guck



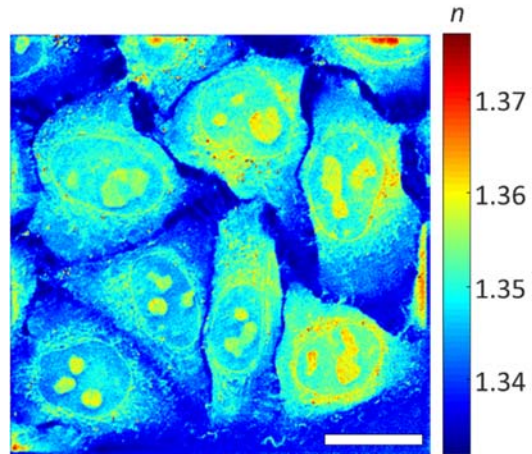
Supplementary Figure 1. Precision of ODT measurements. The cross-sectional slice of (a) an RI tomogram of silica beads immersed in 0.7 M sucrose solution and (b) the standard error of a time series of 10 RI tomograms of the same silica beads. (c) The averaged standard error of a time series of 10 RI tomograms within individual silica bead in each field-of-view (FOV). The number of silica beads in an individual FOV is indicated as N . The averaged standard error of RI is calculated as 4.15×10^{-5} .



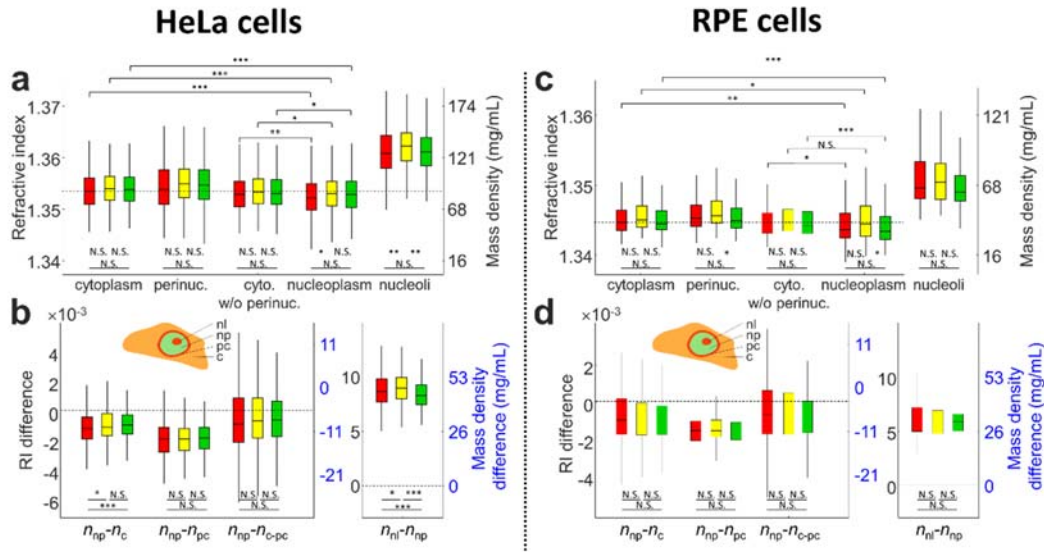
Supplementary Figure 2. Confirmation of the segmentation of nucleoli based on the RI threshold. (a-b) The measured (a) RI tomogram and (b) epi-fluorescence image of HeLa cells which nuclei and nucleoli are stained with Hoechst and GFP, respectively. (c-d) The cross-sectional slices of the RI tomogram of (c) nucleoplasm and (d) nucleoli segmented by the present method based on the RI threshold. (e-f) The cross-sectional slices of the RI tomogram of (e) nucleoplasm and (f) nucleoli segmented by the fluorescence intensity.



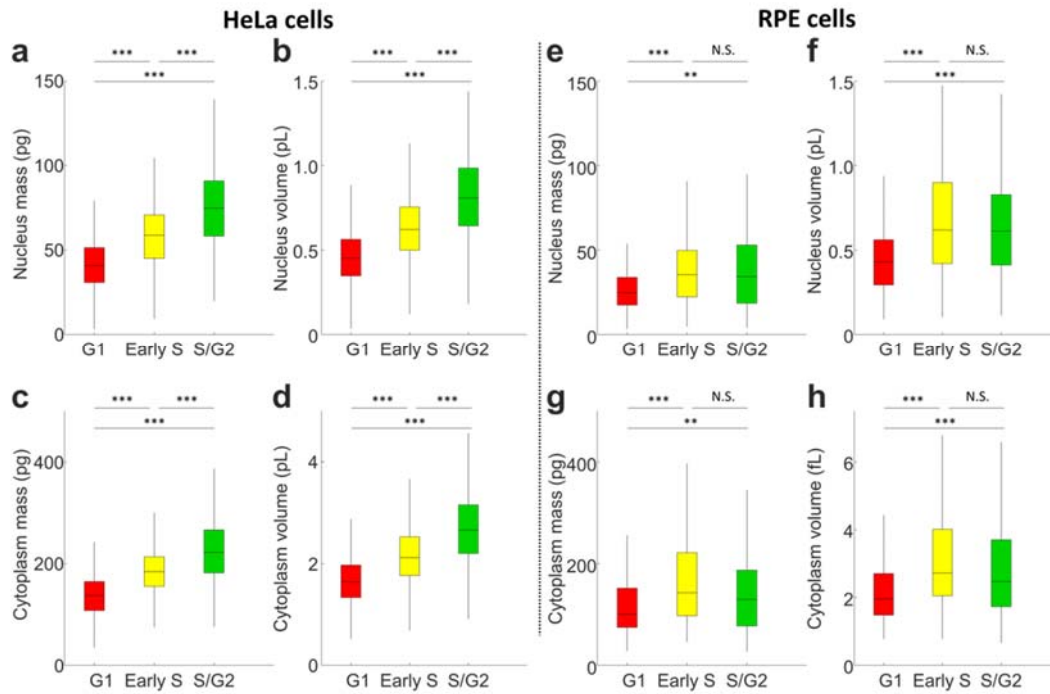
Supplementary Figure 3. Determination of cell cycle phases of individual cells by FUCCI cell-cycle marker. The two-channel epi-fluorescence microscopy measured the intensity of fluorescence signals in FUCCI-stained nucleus in cells (mAG1 and mKO2), and the cell cycle phase of cells was determined in the two-color scatter plot as G1 (high mKO2 signals), early S (high mKO2 and mAG1 signals), and S/G2 (high mAG1 signals) phases. The black dots are determined as either cells in mitosis or cells not expressing fluorescence signals, and discarded in statistical analysis.



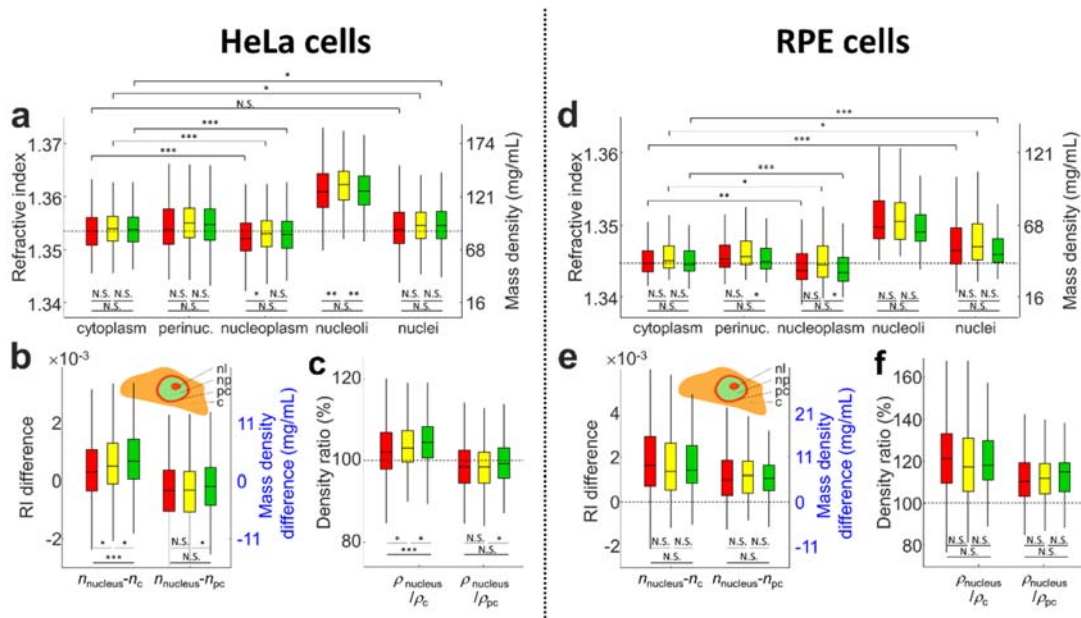
Supplementary Figure 4. Refractive index (RI) and mass density distributions in non-labeled HeLa cells. The maximum projection of an RI tomogram of non-labeled HeLa cells in the direction proportional to the substrate. The scale bar is 20 μm .



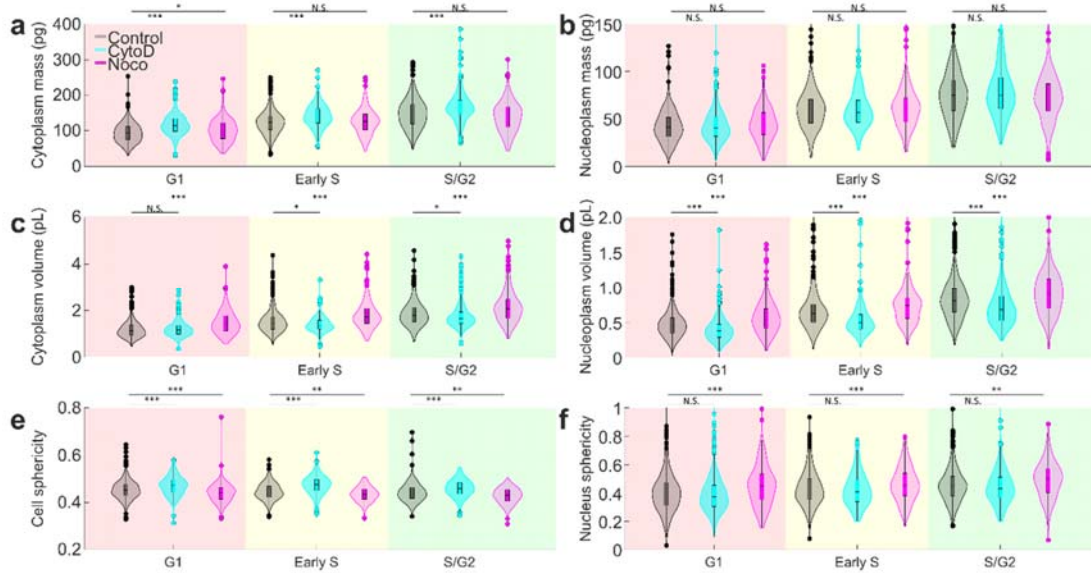
Supplementary Figure 5. Refractive index (RI) and mass density distributions in HeLa-FUCCI (a-b) and RPE-FUCCI (c-d) cells as function of the cell cycle. (a, c) The mean RI values of cytoplasm, perinuclear cytoplasm, cytoplasm excluding perinuclear region, nucleoplasm, and nucleoli. The dashed line indicates the mean RI value of cytoplasm at the G1 phase in the cell cycle. (b, d) The difference of the mean RI values between nucleoplasm and cytoplasm ($n_{np}-n_c$), nucleoplasm and perinuclear cytoplasm ($n_{np}-n_{pc}$), nucleoplasm and cytoplasm excluding perinuclear region ($n_{np}-n_{c-pc}$), and nucleoli and nucleoplasm ($n_{nl}-n_{np}$). The dashed lines indicate the equal mass density of the compartments. The numbers of cells measured are $N = 557, 505,$ and 483 for HeLa cells, and $N = 122, 92,$ and 128 for RPE cells in the G1, early S, and S/G2 phases, respectively.



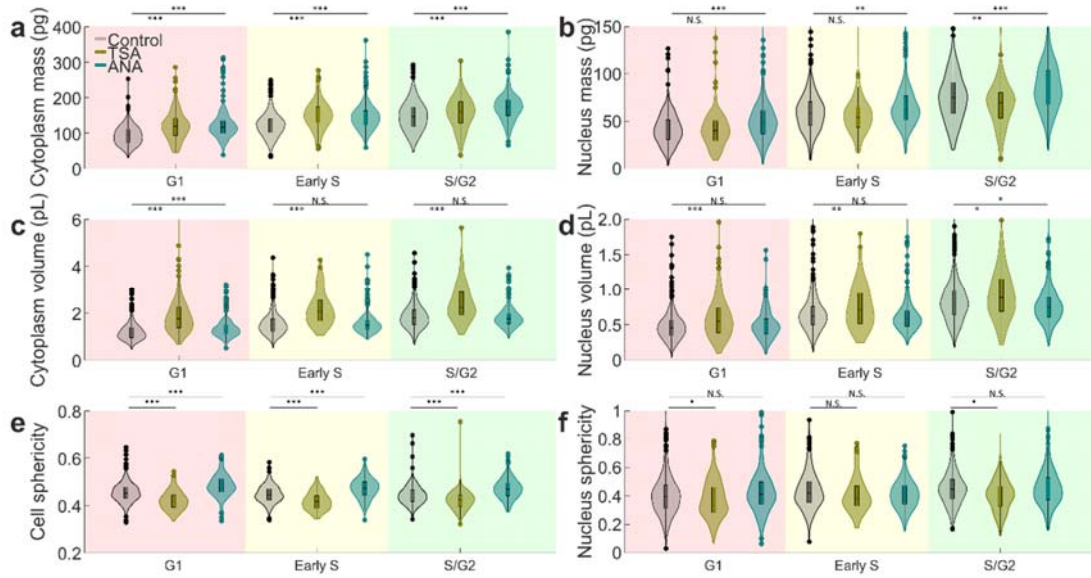
Supplementary Figure 6. Mass and volume distribution of HeLa-FUCCI (a-d) and RPE-FUCCI (e-h) cells as function of the cell cycle. (a, e) The mass and (b, f) volume of nucleus, and (c, g) the mass and (d, h) volume of cytoplasm in HeLa-FUCCI and RPE-FUCCI cells, respectively. The numbers of cells measured are $N = 557$, 505 , and 483 for HeLa cells, and $N = 122$, 92 , and 128 for RPE cells in the G1, early S, and S/G2 phases, respectively.



Supplementary Figure 7. Refractive index (RI) and mass density distributions in HeLa-FUCCI (a-c) and RPE-FUCCI (d-f) cells as function of the cell cycle. (a, c) The mean RI values of cytoplasm, perinuclear cytoplasm, nucleoplasm, nucleoli, and nuclei including nucleoplasm and nucleoli. The dashed line indicates the mean RI value of cytoplasm at the G1 phase in the cell cycle. (b, d) The difference of the mean RI values between nucleus and cytoplasm ($n_{np}-n_c$) and nucleus and perinuclear cytoplasm ($n_{np}-n_{pc}$). The dashed lines indicate the equal mass density of the compartments. (c, f) The ratio of the mass density between nucleus and cytoplasm ($\rho_{nucleus}/\rho_c$) and nucleus and perinuclear cytoplasm ($\rho_{nucleus}/\rho_{pc}$). The dashed lines indicate equal mass density between compartments. The numbers of cells measured are $N = 557, 505,$ and 483 for HeLa cells, and $N = 122, 92,$ and 128 for RPE cells in the G1, early S, and S/G2 phases, respectively.



Supplementary Figure 8. Changes in mass, volume, and sphericity of HeLa-FUCCI cells under cytoskeletal perturbation. (a-d) The mass and volume of nucleus and cytoplasm in HeLa-FUCCI cells in control (gray), cytoD (cyan) and noco (magenta) treatment at various cell cycle phases. (e-f) The sphericity of whole cell (e) and nucleus at various cell cycle phases. The cell cycle phases are indicated as shaded region in red (G1), yellow (Early S) and green (S/G2). The numbers of cells measured are $N = 1,565, 973,$ and 717 for control, cytoD, and nocodazole treatments, respectively.



Supplementary Figure 9. Changes in mass, volume, and sphericity of HeLa-FUCCI cells under chromosome condensation. (a-d) The mass and volume of nucleus and cytoplasm in HeLa-FUCCI cells in control (gray), TSA (dark yellow) and ANA (dark green) treatment at various cell cycle phases. (e-f) The sphericity of whole cell (e) and nucleus at various cell cycle phases. The cell cycle phases are indicated as shaded region in red (G1), yellow (Early S) and green (S/G2). The numbers of cells measured are $N = 1,565, 437,$ and 924 for control, TSA, and ANA treatments, respectively.