

Synthesis and Anti-HIV Activity of Conformationally Restricted Bicyclic Hexahydroisobenzofuran Nucleoside Analogs

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General Spectroscopic and Experimental Data

General techniques. All reagents were bought from Aldrich and Acros at highest commercial quality and used without further purification except where noted. Air- and moisture-sensitive liquids and solutions were transferred via syringe or stainless steel cannula. Organic solutions were concentrated by rotary evaporation below 45 °C at approximately 20 mmHg. All non-aqueous reactions were carried out under anhydrous conditions using flame-dried glassware within an argon atmosphere in dry, freshly distilled solvents, unless otherwise noted. THF and CH₂Cl₂ were purified by passage through a bed of activated alumina. Pyridine (Py) was distilled from CaH₂ prior to use. Yields refer to chromatographically and spectroscopically (¹H NMR) homogeneous materials, unless otherwise stated. Reactions were monitored by TLC carried out on 0.25 mm E. Merck silica gel plates (60F-254) using UV light as visualizing agent and 7% ethanolic phosphomolybdic acid, or *p*-anisaldehyde solution and heat as developing agents. E. Merck silica gel (60, particle size 0.040-0.063 mm) was used for flash chromatography. LC-ESI-MS analyses were carried out in a chromatograph with UV detector at 280 nm using a VYDAC C-18 column, Cat. Number: 218TP54, (4.6mm x 250 nm); flow 1 mL/min; r.t.; gradient MeOH/H₂O as eluent. NMR spectra were recorded on Varian Mercury 300, 400 and/or Unity 500 MHz or Bruker AV-400 or 600 MHz instruments and calibrated using residual undeuterated solvent as an internal reference. The following abbreviations were used to explain

the multiplicities: s= singlet; d= doublet, t= triplet, q= quartet, m= multiplet, br= broad. IR spectra were recorded on a Nicolet 320 Avatar FT-IR spectrometer and values are reported in cm^{-1} units. Optical rotations were recorded on a Jasco P-1010 polarimeter and values are reported as follows: $[\alpha]_{\lambda}^T$ (c: g/100mL, solvent). High resolution mass spectra (HRMS) were recorded on a VG 7070 HS mass spectrometer under electron spray ionization (ESI) conditions. X-ray data were recorded on a Bruker SMART APEX 3kW Sealed Tube X-ray diffraction system.

The bicyclic motif of the synthetic hexahydroisobenzofuran nucleosides is abbreviated below as HIBF. The assignment of the NMR data is based on the nucleoside numbering (1'- 5') and the second ring as 6'- 9' positions (Figure 2 of the article).

Synthesis of Lactone 12. It was obtained following a previously described procedure [a] T. P. Brady, S. H. Kim, K. Wen, E. A. Theodorakis, *Angew. Chem. Int. Ed.* **2004**, *43*, 739-742. b) T. P. Brady, S. H. Kim, K. Wen, C. Kim, E. A. Theodorakis, *Chemistry Eur. J.* **2005**, *11*, 7175-7190]. R_f (33% hexane/EtOAc): 0.41; $[\alpha]_{\text{D}}^{20} = 19.4$ (c 3.40, CH_2Cl_2); IR (film): ν 3070, 3043, 2932, 2857, 1775, 1112, 704 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3 , 400 MHz): δ 1.08 (s, 9H, ^tBu), 1.90-2.25 (m, 4H, $\text{H}_6 + \text{H}_9$), 2.61 (m, 1H, H_3), 3.01 (m, 1H, H_2), 3.80 (m, 2H, H_5), 4.16 (m, 1H, H_4), 5.82 (m, 2H, $\text{H}_7 + \text{H}_8$), 7.43 (m, 6H, $\text{H}_o + \text{H}_p$), 7.69 (m, 4H, H_m); $^{13}\text{C NMR}$ (CDCl_3 , 100.6 MHz): δ 19.5 (C^tBu), 22.8, 25.8 ($\text{C}_6 + \text{C}_9$), 27.1 (Me^tBu), 34.3 (C_3), 37.7 (C_2), 64.5 (C_5), 85.2 (C_4), 125.9-135.9 ($\text{C}_7 + \text{C}_8 + \text{C}_m + \text{C}_o + \text{C}_p$), 179.8 ($\text{C}=\text{O}$); MS (ESI^+ , m/z): 407 $[(\text{M}+\text{H})^+]$, 100%; Exact mass found for $\text{C}_{25}\text{H}_{30}\text{O}_3\text{Si}$: 406.1964.

Synthesis of Lactol 13. A solution of **12** (12.8 g, 31.5 mmol) in dry CH_2Cl_2 (150 mL) was cooled to -78°C under argon and treated with DIBAL-H (35 mL, 35 mmol, 1M in CH_2Cl_2) for 1 h. The reaction was quenched with MeOH (10 mL) and the solution was brought to r.t. over a 20 min period. Saturated Rochelle salt was added (stirred for 15 min) and the crude was extracted with EtOAc (4 x 100mL). Lactol **13** was isolated in 92% yield as yellow oil. R_f (33% hexane/EtOAc): 0.29; $[\alpha]_{\text{D}}^{20} = -16.9$ (c 1.9, CH_2Cl_2); $^1\text{H NMR}$ (CDCl_3 , 400 MHz): δ 1.05 (s, 9H, ^tBu), 1.72-1.85 (m, 2H, $\text{H}_6 + \text{H}_9$), 2.18-2.25 (m, 3H, $\text{H}_6 + \text{H}_9 + \text{H}_3$), 2.78 (m, 1H, H_2), 3.08 (m, 1H, OH), 3.62 (m, 1H, H_5), 3.83 (m, 2H, $\text{H}_4 + \text{H}_5$), 5.10 (m, 1H, H_1), 5.65 (m, 2H, $\text{H}_7 + \text{H}_8$), 7.42 (m, 6H, $\text{H}_o + \text{H}_p$), 7.70 (m, 4H, H_m); $^{13}\text{C NMR}$ (CDCl_3 , 100.6 MHz): δ 19.8 (C^tBu), 23.1, 23.5 ($\text{C}_6 + \text{C}_9$), 27.1 (Me^tBu), 32.4 (C_2), 41.9 (C_3), 64.8 (C_5), 83.6 (C_4), 103.0 (C_1), 124.3, 125.2 ($\text{C}_7 + \text{C}_8$), 128.0, 130.0, 130.1, 135.9 ($\text{C}_o + \text{C}_m + \text{C}_p$); MS (ESI^+ , m/z): 409 $[(\text{M}+\text{H})^+]$, 100%; Exact mass found for $\text{C}_{25}\text{H}_{32}\text{O}_3\text{Si}$: 408.2120.

Synthesis of Acetate 14. Lactol **13** (6.65 mmol) was dissolved in Py (40 mL) and Ac_2O (1.8 mL, 19.95 mmol) was added dropwise. The reaction was stirred overnight and then quenched with saturated NaHCO_3 . The solution was extracted with CH_2Cl_2 (3 x 60 mL). The combined organic extracts were dried (Na_2SO_4). The residue was purified by flash chromatography (10% hexane/EtOAc) to obtain **14** (90%). R_f (25% hexane/EtOAc): 0.65; $[\alpha]_{\text{D}}^{20} = 22.8$ (c 0.6, CHCl_3); IR (film): ν 3069, 3025, 2925, 2849, 1744, 1421, 1226, 1099 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 1.06 (s, 9H, ^tBu), 1.92 (m, 2H, H_6 , H_9), 1.97 (m, 3H, CH_3), 2.26 (m, 3H, H_3 , H_6 , H_9), 2.66 (m, 1H, H_2), 3.73 (dd, 1H, H_5 , 2J 11.5 Hz, 3J 4.5 Hz), 3.84 (dd, 1H, H_5 , 2J 11.5 Hz, 3J 4.0 Hz), 3.90 (dt, 1H, H_4 , 3J 10.0, 4.0 Hz), 5.71 (m, 2H, $\text{H}_7 + \text{H}_8$), 5.97 (s, 1H, H_1), 7.41 (m, 6H, $\text{H}_o + \text{H}_p$), 7.71 (m, 4H, H_m); $^{13}\text{C NMR}$ (CDCl_3 , 75.5 MHz): δ 19.5 (C^tBu), 21.7 (Me), 22.6, 23.0 ($\text{C}_6 + \text{C}_9$), 26.9 (3 CH_3 - ^tBu), 33.5 (C_3), 40.2 (C_2), 65.3 (C_5), 84.5 (C_4), 102.5 (C_1), 124.5, 124.7 ($\text{C}_7 + \text{C}_8$), 127.9, 130.0, 133.7, 135.2, 135.9 (C_{arom}), 171.2 ($\text{C}=\text{O}$); MS (ESI^+ , m/z): 468 $[(\text{M}+\text{NH}_4)^+]$, 100%, 473 $[(\text{M}+\text{Na})^+]$, 45, and 489 $[(\text{M}+\text{K})^+]$, 15.

Synthesis of Acetate 15. Compound **14** (3.34 mmol) was dissolved in 60 mL of dry THF and then treated with TBAF (4.3 mL, 4.3 mmol, 1M in THF). The reaction was kept under argon at r.t. for 30 min. The solution was extracted with EtOAc (3 x 30 mL). The combined organic extracts were dried using Na_2SO_4 and then filtered. The residue was purified by flash

chromatography (30% hexane/EtOAc) to give **15** (89%). R_f (EtOAc): 0.74; $[\alpha]_D^{20} = 47.4$ (c 0.9, CHCl₃); IR (film): ν 3019, 2922, 2845, 1728, 1427, 1369, 1233, 932 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.85-1.92 (m, 2H, H₆, H₉), 2.04 (m, 3H, CH₃), 2.24 (m, 1H, H₃), 2.33 (m, 2H, H₆, H₉), 2.62 (m, 1H, H₂), 3.56 (dd, 1H, H₅, ²J 12.0 Hz, ³J 3.2 Hz), 3.83 (dd, 1H, H₅, ²J 12.0 Hz, ³J 2.0 Hz), 3.90 (ddd, 1H, H₄, ³J 10.0, 3.2, 2.0 Hz), 5.70 (m, 2H, H₇+H₈), 5.95 (s, 1H, H₁); ¹³C NMR (CDCl₃, 100.6 MHz): δ 21.7 (Me), 22.4, 22.9 (C₆+C₉), 32.2 (C₃), 40.4 (C₂), 63.1 (C₅), 84.9 (C₄), 102.7 (C₁), 124.2, 124.7 (C₇+C₈), 170.5 (C=O); MS (ESI⁺, m/z): 230 [(M+NH₄)⁺, 50%], 235 [(M+Na)⁺, 75] and 251 [(M+K)⁺, 20].

Synthesis of Diacetate 16. A solution of **17** (31 mmol) in Py (100 mL) was treated with Ac₂O (12 mL, 124 mmol). The reaction was stirred overnight and quenched with saturated NaHCO₃. The solution was extracted with CH₂Cl₂ (4 x 200 mL). The combined organic extracts were dried using Na₂SO₄ and then filtered. The residue was purified by flash chromatography (15% hexane/EtOAc) to afford **16** (93%). R_f (50% EtOAc/hexane): 0.53; $[\alpha]_D^{20} = -21.8$ (c 1.9, CHCl₃); IR (film): ν 3030, 2918, 2838, 1737, 1641, 1440, 1239, 933 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz): δ 1.87 (m, 2H, H₆, H₉), 2.04 (s, 3H, CH₃), 2.09 (s, 3H, CH₃), 2.12-2.46 (m, 4H, H₂, H₃, H₆, H₉), 4.01 (m, 1H, H₅), 4.08 (m, 1H, H₄), 4.30 (dd, 1H, H₅, ²J=11.5 Hz, ³J=2.5 Hz), 5.68 (m, 2H, H₇+H₈), 5.96 (s, 1H, H₁); ¹³C NMR (CDCl₃, 100.6 MHz): δ 21.1 (Me), 21.6 (Me), 22.4, 22.8 (C₆+C₉), 34.2 (C₃), 40.0 (C₂), 66.4 (C₅), 81.7 (C₄), 102.7 (C₁), 124.0, 124.7 (C₇+C₈), 170.1, 170.2 (C=O); MS (ESI⁺, m/z): 272 [(M+NH₄)⁺, 20%] and 277 [(M+Na)⁺, 100].

Synthesis of Diol 17. Lactol **13** (31.5 mmol) was dissolved in THF (100 mL) under argon and then treated with TBAF (37.8 mL, 37.8 mmol, 1M in THF) at r.t. for 25 min. The solution was quenched with Brine and extracted with EtOAc (3 x 100mL). The residue was chromatographed using a gradient 30-70% EtOAc/hexane to give **17** (96%). R_f (EtOAc): 0.44; $[\alpha]_D^{20} = -16.9$ (c 1.9, CH₂Cl₂); ¹H NMR (CDCl₃, 100.6 MHz): δ 1.80 (m, 2H, H₆+H₉), 2.08-2.35 (m, 3H, H₆+H₉+H₃), 2.65 (m, 1H, H₂), 3.51 (m, 2H, H₄+OH), 3.83 (d, 1H, H₅, ²J 13.2 Hz), 3.87 (d, 1H, H₅, ²J 13.2 Hz), 4.83 (br s, 1H, OH), 5.12 (s, 1H, H₁), 5.65 (m, 2H, H₇+H₈); ¹³C NMR (CDCl₃, 75.5 MHz): δ 22.9, 23.5 (C₆+C₉), 31.9 (C₃), 41.5 (C₂), 62.6 (C₅), 84.0 (C₄), 103.0 (C₁), 124.3, 125.2 (C₇+C₈); MS (ESI⁺, m/z): 193 [(M+Na)⁺, 55%].

Synthesis of Diacetate 19. A solution of **13** (31.5 mmol) in THF (100mL) was treated with TBAOH (47.3 mmol, 33% in H₂O) at r.t. for 6 h. Then, solvents were removed in vacuo. The orange residue was treated in Py (100 mL) with Ac₂O (12 mL, 124 mmol). The reaction was stirred overnight and then quenched with saturated NaHCO₃. The solution was extracted with CH₂Cl₂ (4 x 200 mL). The combined organic extracts were dried using Na₂SO₄, and then filtered. The residue was purified by flash chromatography (20% hexane/EtOAc) to give **19** (70%). R_f (50% EtOAc/hexane): 0.55; IR (film): ν 3026, 2931, 2839, 1734, 1633, 1233, 936 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.60-2.11 (m, 12H, 2CH₃, 2H₆, 2H₉, H₂, H₃), 3.68 (d, 1H, H₅, ²J 13.0 Hz), 3.97 (dd, 1H, H₅, ²J 13.0 Hz, ³J 1.2 Hz), 4.68 (s, 1H, H₄), 5.28 (d, 1H, H₁, ³J 8.8 Hz), 5.54 (m, 2H, H₇+H₈); ¹³C NMR (CDCl₃, 100.6 MHz): δ 20.8 (Me), 20.9 (Me), 25.8, 26.0 (C₆+C₉), 34.9 (C₃), 37.0 (C₂), 68.1 (C₅), 68.3 (C₄), 97.0 (C₁), 124.5, 125.6 (C₇+C₈), 169.5, 170.6 (C=O); MS (ESI⁺, m/z): 272 [(M+NH₄)⁺, 80%] and 277 [(M+Na)⁺, 100].

General procedure for condensation of diacetate 16 with heterocyclic bases. To a stirred solution of the different bases (1.81 mmol) in dry MeCN (5.0 mL), BSA was added (1.5 mL, 6.338 mmol). The reaction mixture was heated at 70 °C for 1.5 h. To the resulting clear mixture was added at r.t. a solution of diacetate **16** (0.9 mmol) in dry MeCN (3 mL), followed by DBU (0.434 mL, 2.899 mmol). This solution was cooled to 0 °C and then TMSOTf was added dropwise (1.05 mL, 5.798 mmol). The resulting mixtures were stirred for 2-3 h. Each mixture was poured into an ice-cool saturated NaHCO₃ solution and extracted with CH₂Cl₂ (3 x 25 mL). Residues were purified by silica gel column chromatography to give **20** and **21** using a gradient 33-100% EtOAc/hexane.

HIBF- α / β -Uracil (20a+21a). R_f (EtOAc): 0.55; $[\alpha]_D^{20} = -23.4$ (c 0.7, CHCl₃); **IR** (KBr): ν 3200, 3032, 2922, 2844, 1743, 1686, 1461, 1271, 1237, 1042, 705 cm⁻¹; **¹H NMR** (CDCl₃, 500 MHz): δ 2.00 (m, 2H), 2.12 (s, 3H, CH₃), 2.18 (s, 3H, CH₃), 2.20-2.58 (m, 8H), 2.83 (m, 1H, H₂), 4.05-4.45 (m, 6H, H₄+H₅), 5.68 (d, 1H, H₁, ³J 2.0 Hz), 5.70-5.76 (m, 6H, 2H₅+2H₇+2H₈), 5.89 (d, 1H, H₁, ³J 4.0 Hz), 7.82 (d, 1H, H₆, ³J 8.3 Hz), 7.64 (d, 1H, H₆, ³J 8.3 Hz), 8.45 (br s, 1H, NH), 8.55 (br s, 1H, NH); **¹³C NMR** (CDCl₃, 75.5 MHz): δ 19.7 (C₆ or C₉), 21.08 (Me), 21.12 (Me), 22.7, 22.9, 23.6 (C₆+C₉), 33.6 (C₃), 36.3 (C₃), 37.3 (C₂), 41.6 (C₂), 63.6 (C₅), 64.8 (C₅), 80.2 (C₄), 82.4 (C₄), 90.0 (C₁), 91.6 (C₁), 101.1 (C₅), 101.4 (C₅), 123.8, 124.4, 124.7, 125.0 (2C₇+2C₈), 140.0 (C₆), 140.4 (C₆), 150.6 (C₂), 150.9 (C₂), 164.2 (C₄), 164.5 (C₄), 170.7 (C=O), 171.1 (C=O); **MS** (ESI⁺, m/z): 307 [(M+H)⁺, 70%], 324 [(M+NH₄)⁺, 80], and 329 [(M+Na)⁺, 100]; LC-ESI-MS (UV 280 nm): Coupling Ratio β/α 1.1:1 t_R (min)= 17.46, 17.91; Exact mass found for C₁₅H₁₈N₂O₅: 306.1206.

HIBF- α / β -Thymine (20b+21b). R_f (50% Hexane/EtOAc): 0.22; $[\alpha]_D^{20} = -41.26$ (c 1.0, CHCl₃); **IR** (KBr): ν 3197, 3032, 2926, 1744, 1684, 1662, 1473, 1271, 1237, 1044 cm⁻¹; **¹H NMR** (CDCl₃, 400 MHz): δ 1.67 (m, 2H), 2.00 (m, 6H, CH₃), 2.14-2.60 (m, 15H, 2CH₃, H₂, 2H₃, 6(H₆+H₉)), 2.85 (m, 1H, H₂), 4.11 (m, 3H, H₄+2H₅), 4.31 (m, 1H, H₅), 4.39 (m, 1H, H₅), 4.39 (ddd, 1H, H₄, ²J 11.5 Hz, ³J 3.0, 2.5 Hz), 5.68 (m, 5H, H₁+2H₇+2H₈), 5.89 (m, 1H, H₁), 7.46 (q, 1H, ⁴J 1.6 Hz), 7.56 (q, 1H, ⁴J 0.8 Hz), 9.08 (brs, 1H, NH), 9.20 (brs, 1H, NH); **¹³C NMR** (CDCl₃, 100.6 MHz): δ 12.9, 13.0 (Me), 19.8 (CH₂), 21.1, 21.2 (Me), 22.9 (CH₂), 23.1 (CH₂), 23.6 (CH₂), 34.2 (C₃), 36.3 (C₃), 37.3 (C₂), 41.6 (C₂), 64.0 (C₅), 64.8 (C₅), 80.1 (C₄), 82.4 (C₄), 89.9 (C₁), 91.1 (C₁), 109.5 (C₅), 110.0 (C₅), 123.9, 124.4, 124.6, 125.0 (2C₇+2C₈), 135.7 (C₆), 136.1 (C₆), 150.6 (C₂), 150.9 (C₂), 164.4 (C₄), 164.5 (C₄), 170.8 (C=O), 171.1 (C=O); **MS** (ESI⁺, m/z): 321 [(M+H)⁺, 100%], 338 [(M+NH₄)⁺, 90], and 343 [(M+Na)⁺, 95]; LC-ESI-MS (UV 280 nm): Coupling Ratio β/α 1.2:1 t_R (min)= 19.11, 19.29; Exact mass found for C₁₆H₂₀N₂O₅: 320.1371.

HIBF- β -5-Fluorouracil (20c). R_f (50% Hexane/EtOAc): 0.29; $[\alpha]_D^{20} = +10.0$ (c 1.3, CHCl₃); **IR** (KBr): ν 3189, 3075, 3044, 2950, 2857, 1718, 1661, 1469, 1262, 1230, 1093, 1064 cm⁻¹; **¹H NMR** (CDCl₃, 500 MHz): δ 1.97-2.54 (m, 9H, H₂+H₃+ 2H₆+2H₉+CH₃), 4.10 (m, 1H, H₄), 4.36 (dd, 1H, H₅, ²J 13.3 Hz, ³J 2.0 Hz), 4.48 (dd, 1H, H₅, ²J 13.3 Hz, ³J 3.5 Hz), 5.61 (s, 1H, H₁), 5.72 (m, 2H, H₇+H₈), 8.14 (d, 1H, H₆, ³J 6.5 Hz), 8.52 (br s, 1H, NH); **¹³C NMR** (CDCl₃, 100.6 MHz): δ 20.9 (Me), 22.3, 23.6 (C₆+C₉), 32.7 (C₃), 41.8 (C₂), 62.6 (C₅), 82.2 (C₄), 91.5 (C₁), 124.3, 124.5, 124.8 (C₇+C₈+C₆), 140.2 (d, C₅, ¹J_{C,F} 234 Hz), 149.3 (C₂), 157.5 (d, C₄, ²J_{C,F} 26 Hz), 170.7 (C=O); **¹⁹F NMR** (CDCl₃, 282 MHz): δ 167.1; **MS** (ESI⁺, m/z): 325 [(M+H)⁺, 30%], 342 [(M+NH₄)⁺, 60], and 347 [(M+Na)⁺, 100]; LC-ESI-MS (UV 280 nm): t_R (min)= 19.31; Exact mass found for C₁₅H₁₇FN₂O₅: 324.1122.

HIBF- β -5-Bromovinyluracil (20d). R_f (50% Hexane/EtOAc): 0.41; $[\alpha]_D^{20} = -22.3$ (c 0.6, CHCl₃); **IR** (KBr): ν 3424, 2926, 2874, 1793, 1747, 1700, 1472, 1368, 1280, 1166, 1109, 1041 cm⁻¹; **¹H NMR** (CDCl₃, 400 MHz): δ 2.05 (m, 2H, H₆+H₉), 2.16 (s, 3H, CH₃), 2.18-2.58 (m, 4H, H₆, H₉, H₃, H₂), 4.14 (m, 1H, H₄), 4.29 (dd, 1H, H₅, ²J 12.8 Hz, ³J 2.0 Hz), 4.58 (dd, 1H, H₅, ²J 12.8 Hz, ³J 4.8 Hz), 5.64 (s, 1H, H₁), 5.73 (m, 2H, H₇+H₈), 6.73 (d, 1H, H₇, ³J 13.6 Hz), 7.40 (d, 1H, H₈, ³J 13.6 Hz), 7.85 (s, 1H, H₆), 9.49 (br s, 1H, NH); **¹³C NMR** (CDCl₃, 100.6 MHz): δ 21.0 (CH₃), 22.3 (CH₂), 23.4 (CH₂), 33.1 (C₃), 41.4 (C₂), 63.2 (C₅), 82.5 (C₄), 91.7 (C₁), 109.7 (C₅), 110.2 (C₈), 124.1, 124.4, (C₇+C₈), 128.5 (C₇), 137.7 (C₆), 149.3 (C₂), 161.8 (C₄), 170.6 (C=O); **MS** (ESI⁺, m/z): 433 [(M⁷⁹Br+Na)⁺, 100%], 435 [(M⁸¹Br+Na)⁺, 97]; Exact mass found for C₁₇H₁₉BrN₂O₅: 410.0872.

HIBF- α / β -Cytosine (20e+21e). R_f (EtOAc): 0.24; $[\alpha]_D^{20} = -25.8$ (c 0.4, MeOH); **¹H NMR** (MeOH-*d*₄, 400 MHz): δ 1.90-2.80 (m, 23H, 2H₃+H₂+ 4H₆+ 4H₉+ 4CH₃), 3.00 (m, 1H, H₂), 4.10-4.50 (m, 6H, 2H₄+4H₅), 5.60 (m, 4H, 2H₇+2H₈), 5.69 (s, 1H, H₁), 5.99 (d, 1H, H₁, ³J 4.4 Hz), 7.48 (d, 1H, H₅, ³J 7.4 Hz), 7.54 (d, 1H, H₅, ³J 7.4 Hz), 8.04 (d, 1H, H₆, ³J 7.4 Hz),

8.30 (d, 1H, H₆, ³J 7.4 Hz), 9.78 (brs, 1H, NH), 9.86 (brs, 1H, NH); ¹³C NMR (MeOH-*d*₄, 100.6 MHz): δ 19.6 (C_{6'} or C_{9'}), 21.0 (Me), 21.1 (Me), 22.0, 22.8, 24.0 (C_{6'}+C_{9'}), 24.96 (Me), 24.97 (Me), 32.5 (C_{3'}), 36.2 (C_{3'}), 36.5 (C_{2'}), 41.2 (C_{2'}), 63.0 (C_{5'}), 64.7 (C_{5'}), 80.5 (C_{4'}), 82.5 (C_{4'}), 91.0 (C_{1'}), 92.9 (C_{1'}), 96.4 (C₅), 96.7 (C₅), 123.2, 123.9, 124.6, 124.7 (C₇+C₈), 144.5 (C₆), 145.0 (C₆), 156.40 (C₂), 155.44 (C₂), 162.9 (C₄), 163.0 (C₄), 170.6, 171.1, 171.3, 171.5 (C=O); MS (ESI⁺, *m/z*): 370 [(M+Na)⁺, 100%]; LC-ESI-MS (UV 280 nm): Coupling Ratio β/α 1.2:1 t_R(min)= 19.09, 19.63; Exact mass found for C₁₇H₂₁N₃O₅: 347.1480.

HIBF-α/β-6-Chloropurine (20f+21f). R_f (50% hexane/EtOAc): 0.37; [α]_D²⁰= -25.8 (c 1.0, CHCl₃); IR (KBr): ν 3389, 2942, 2890, 1708, 1633, 1489, 1245, 1193, 1094, 920 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 1.95-2.99 (m, 18H, 2CH₃+2H₂+2H₃+4H₆+4H₉), 4.10-4.35 (m, 6H, H₄+H₅), 5.54 (m, 2H, H₇+H₈), 5.70 (m, 2H, H₇+H₈), 5.95 (d, 1H, H₁, ³J 2.8 Hz), 6.30 (m, 1H, H₁), 8.35 (s, 1H, H₈), 8.48 (s, 1H, H₈), 8.62 (s, 1H, H₂), 8.64 (s, 1H, H₂); ¹³C NMR (CDCl₃, 100.6 MHz): δ 19.8 (C_{6'} or C_{9'}), 21.1 (2Me), 22.7, 23.3, 23.7 (3C, C_{6'}+C_{9'}), 33.7 (C_{3'}), 36.6 (C_{3'}), 38.1 (C_{2'}), 40.7 (C_{2'}), 63.5 (C_{5'}), 64.8 (C_{5'}), 80.5 (C_{4'}), 82.6 (C_{4'}), 89.5 (C_{1'}), 91.0 (C_{1'}), 123.6, 124.1, 124.3, 124.6 (C₇+C₈), 132.2 (C₅), 132.6 (C₅), 143.5 (C₈), 143.9 (C₈), 151.0, 151.1, 151.8 (C₂+C₄), 151.9 (C₆), 152.0 (C₆), 170.8, 171.0 (C=O); MS (ESI⁺, *m/z*): 371 [(M³⁵Cl+Na)⁺, 100], 373 [(M³⁷Cl+Na)⁺, 30]; LC-ESI-MS (UV 280 nm): Coupling Ratio β/α 1.5:1 t_R(min)= 21.19, 21.43; Exact mass found for C₁₆H₁₇ClN₄O₃: 348.0982.

HIBF-α/β-2,6-Dichloropurine (20g+21g). The two anomers were separated. **20g:** R_f (50% hexane/EtOAc): 0.52; [α]_D²⁰= -7.3 (c 0.3, CHCl₃); IR (KBr): ν 3370, 2940, 2879, 1728, 1627, 1480, 1239, 1190, 1076, cm⁻¹; ¹H NMR (CDCl₃, 400 MHz): δ 2.05-2.13 (m, 5H, H₆, H₉, CH₃), 2.40-2.63 (m, 3H, H₃, H₆, H₉), 3.00 (m, 1H, H₂), 4.21(dt, 1H, H₄, ³J 9.3, 3.4 Hz), 4.42 (m, 2H, H₅), 5.78 (m, 2H, H₇+H₈), 5.97 (d, 1H, H₁, ³J 1.7 Hz), 8.55 (s, 1H, H₈); ¹³C NMR (CDCl₃, 100.6 MHz): δ 20.9 (CH₃), 22.4 (C_{9'}), 23.5 (C_{6'}), 33.5 (C_{3'}), 40.7 (C_{2'}), 63.1 (C_{5'}), 82.5 (C_{4'}), 91.0 (C_{1'}), 123.9, 124.3 (C₇+C₈), 131.6 (C₅), 144.4 (C₈), 151.7 (C₄), 152.0 (C₂), 152.8 (C₆), 170.5 (C=O); MS (ESI⁺, *m/z*): 405 [(M³⁵Cl³⁵Cl+Na)⁺, 100%], 407 [(M³⁵Cl³⁷Cl+Na)⁺, 80], and 409 [(M³⁷Cl³⁷Cl+Na)⁺, 10%]; LC-ESI-MS (UV 280 nm): Ratio β/α 2.4:1 t_R(min)= 23.62, 24.27; Exact mass found for C₁₆H₁₆Cl₂N₄O₃: 382.0600.

21g: R_f (50% hexane/EtOAc): 0.52; [α]_D²⁰= -40.8 (c 1.3, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 1.53 (m, 1H, H₆), 1.56 (m, 1H, H₆), 2.08 (m, 1H, H₉), 2.15 (s, 3H, CH₃), 2.46 (m, 1H, H₉), 2.66 (m, 1H, H₃), 3.02 (m, 1H, H₂), 4.19 (dd, 1H, H₅, ³J 12.1, 5.3 Hz), 4.30 (m, 1H, H₄), 4.42 (dd, 1H, H₅, ³J 12.1, 2.4 Hz), 5.65 (m, 2H, H₇+H₈), 6.33 (d, 1H, H₁, ³J 4.3 Hz), 8.39 (s, 1H, H₈); ¹³C NMR (CDCl₃, 100.6 MHz): δ 19.6 (CH₂), 20.9 (CH₃), 23.1 (CH₂), 36.4 (C_{3'}), 37.9 (C_{2'}), 64.4 (C_{5'}), 80.5 (C_{4'}), 89.3 (C_{1'}), 123.3, 124.2 (C₇+C₈), 131.2 (C₅), 144.0 (C₈), 151.7 (C₄), 151.9 (C₂), 152.9 (C₆), 170.8 (C=O); MS (ESI⁺, *m/z*): 405 [(M³⁵Cl³⁵Cl+Na)⁺, 100%], 407 [(M³⁵Cl³⁷Cl+Na)⁺, 80], and 409 [(M³⁷Cl³⁷Cl+Na)⁺, 10]; LC-ESI-MS (UV 280 nm): Coupling Ratio β/α 2.4:1 t_R(min)= 23.62, 24.27; Exact mass found for C₁₆H₁₆Cl₂N₄O₃: 382.0651.

General protocols for deprotection of HIBF nucleoside derivatives 20-21. Method A (deprotection of 5'-OAc group). Nucleoside derivatives **20a-f+21a-f** (0.1mmol) were dissolved in a 2M NH₃/MeOH solution (1.5 mL) and stirred at r.t. overnight. Residues were purified by silica gel column chromatography using a gradient 33-100% EtOAc/hexane. **Method B.** Nucleoside derivatives **20f+21f, 20g** and **21g** (0.1mmol) were treated with a NH₃/MeOH saturated solution (1.5 mL) and stirred at 110 °C in a Parr reactor for 36 h. Residues were purified by silica gel column chromatography (gradient 50-100% EtOAc/hexane) obtaining **22i, 23i** (95-97%), **22j-22m, 23j-23m** (quantitative combined yield). **Method C.** Nucleoside derivatives **20f** and **20g** (0.11 mmol) were treated with a 2M NH₃/MeOH solution (1.8 mL) and stirred at 90-100 °C in a Parr reactor for 30 h. Residues were purified by silica gel column chromatography using a gradient 50-100% EtOAc/hexane.

HIBF-β-Uracil (22a). From **20a** using Method A. R_f (EtOAc): 0.40; [α]_D²⁰= +0.7 (c 0.3,

CHCl₃); ¹H NMR (CDCl₃, 500 MHz): δ 1.99 (m, 2H, H_{6'}+H_{9'}), 2.34 (m, 1H, H_{3'}), 2.47 (m, 2H, H_{6'}+H_{9'}), 2.53 (m, 1H, H_{2'}), 3.80 (dt, 1H, H_{5'}, ²J 12 Hz, ³J 3.5 Hz), 3.96 (dt, 1H, H_{4'}, ³J 9.0 Hz, ³J = 3.5 Hz), 4.07 (ddd, 1H, H_{5'}, ²J 12.0 Hz, ³J 3.5 Hz, ⁴J 2.5 Hz), 5.70 (m, 3H, H_{7'}+H_{8'}+H_{1'}), 8.05 (d, 1H, H_{6'}, ³J 8.0 Hz), 8.36 (br s, 1H, NH); ¹³C NMR (CDCl₃, 100.6 MHz): δ 23.1 (C_{6'}), 23.7 (C_{9'}), 32.6 (C_{3'}), 41.6 (C_{2'}), 62.0 (C_{5'}), 85.0 (C_{4'}), 91.2 (C_{1'}), 101.4 (C₅), 124.7, 124.8 (C_{7'}+C_{8'}), 141.0 (C₆), 150.5 (C₂), 163.5 (C₄); MS (ESI⁺, *m/z*): 287 [(M+Na)⁺, 100%]; Exact mass found for C₁₃H₁₅N₂O₄: 264.1104.

HIBF-β-Thymine (22b). From **20b** using Method A. R_f (50% hexane/EtOAc): 0.16; [α]_D²⁰ = -13.6 (c 0.3, CHCl₃); ¹H NMR (CDCl₃, 500 MHz): δ 1.91 (d, 3H, Me, ⁴J 1.0 Hz), 2.00 (m, 2H, H_{6'}+H_{9'}), 2.00 (m, 4H, H_{6'}+H_{9'}+H_{3'}+H_{2'}), 3.81 (dt, 1H, H_{5'}, ²J 12 Hz, ³J 4.0 Hz), 3.94 (dt, 1H, H_{4'}, ³J 9.0 Hz, ³J 4.0 Hz), 4.06 (ddd, 1H, H_{5'}, ³J 9.0 Hz, ³J 4.0 Hz, ⁴J 2.5 Hz), 5.73 (m, 3H, H_{7'}, H_{8'}, H_{1'}), 7.79 (d, 1H, H_{6'}, ⁴J 1.0 Hz), 8.50 (brs, 1H, NH); ¹³C NMR (CDCl₃, 100.6 MHz): δ 12.9 (Me), 23.5, 23.7 (C_{6'}+C_{9'}), 33.2 (C_{3'}), 41.5 (C_{2'}), 62.5 (C_{5'}), 85.0 (C_{4'}), 91.0 (C_{1'}), 110.1 (C₅), 125.0, 125.1 (C_{7'}+C_{8'}), 136.8 (C₆), 150.6 (C₂), 163.9 (C₄); MS (ESI⁺, *m/z*): 301 [(M+Na)⁺, 100%]; Exact mass found for C₁₄H₁₈N₂O₄: 278.1257.

HIBF-β-5-Fluorouracil (22c). From **20c** using Method A. R_f (50% hexane/EtOAc): 0.28; [α]_D²⁰ = +28.6 (c 0.8, CHCl₃); IR (KBr): ν 3508, 3448, 3156, 3040, 2925, 2851, 1682, 1652, 1266, 1093, 1066, 1026 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz): δ 1.98 (m, 2H, H_{6'}, H_{9'}), 2.26-2.53 (m, 4H, H_{6'}, H_{9'}, H_{3'}, H_{2'}), 3.83 (dd, 1H, H_{5'}, ²J 12.5 Hz, ³J 2.5 Hz), 3.95 (dt, 1H, H_{4'}, ³J 8.5 Hz, ³J 2.5 Hz), 4.13 (dd, 1H, H_{5'}, ²J 12.5 Hz, ³J 2.5 Hz), 5.68 (m, 1H, H_{1'}), 5.78 (m, 2H, H_{7'}+H_{8'}), 8.42 (d, 1H, H_{6'}, ³J 7.0 Hz), 8.50 (br s, 1H, NH); ¹³C NMR (CDCl₃, 75.5 MHz): δ 22.9, 23.7 (C_{6'}+C_{9'}), 32.0 (C_{3'}), 41.7 (C_{2'}), 61.5 (C_{5'}), 85.0 (C_{4'}), 91.2 (C_{1'}), 124.6, 124.7 (C_{7'}+C_{8'}), 125.8 (d, C₆, ²J_{C,F} 139.8 Hz), 140.2 (d, C₅, ¹J_{C,F} 930 Hz), 149.1 (C₂), 157.2 (d, C₄, ²J_{C,F} 105.9 Hz); ¹⁹F NMR (CDCl₃, 282 MHz): δ 167.0; MS (ESI⁺, *m/z*): 305 [(M+Na)⁺, 100%]; Exact mass found for C₁₃H₁₅FN₂O₄: 282.1014.

HIBF-β-5-Bromovinyluracil (22d). From **20d** using Method A. R_f (50% hexane/EtOAc): 0.32; [α]_D²⁰ = -24.8 (c 0.6, CHCl₃); ¹H NMR (CDCl₃, 500 MHz): δ 1.99 (m, 2H, H_{6'}, H_{9'}), 2.30-2.52 (m, 4H, H_{6'}, H_{9'}, H_{2'}, H_{3'}), 3.85 (d, 1H, H_{5'}, ²J 12.9 Hz), 3.99 (m, 1H, H_{4'}), 4.17 (d, 1H, H_{5'}, ²J 12.9 Hz), 5.73 (m, 3H, H_{7'}+H_{8'}+H_{1'}), 6.70 (d, 1H, H_{7'}, ³J 13.6 Hz), 7.32 (d, 1H, H_{8'}, ³J 13.6 Hz), 8.33 (s, 1H, H_{6'}), 8.52 (br s, 1H, NH); ¹³C NMR (CDCl₃, 100.6 MHz): δ 22.5, 23.6 (C_{6'}+C_{9'}), 31.6 (C_{3'}), 41.6 (C_{2'}), 61.1 (C_{5'}), 84.7 (C_{4'}), 91.1 (C_{1'}), 108.8 (C₈), 110.1 (C₅), 124.3, 124.4 (C_{7'}+C_{8'}), 128.6 (C₇), 138.8 (C₆), 149.2 (C₂), 161.3 (C₄); MS (ESI⁺, *m/z*): 369 [(M⁷⁹Br+H)⁺, 100%] and 371 [(M⁸¹Br+H)⁺, 96]; Exact mass found for C₁₅H₁₇BrN₂O₄: 368.0395.

HIBF-β-6-Chloropurine (22f). From **20f** using Method A. R_f (EtOAc): 0.50; [α]_D²⁰ = -37.2 (c 1.3, CHCl₃); ¹H NMR (CDCl₃, 600 MHz): δ 1.95, 2.11 (m, 2H, H_{6'}+H_{9'}), 2.43 (m, 2H, H_{6'}+H_{9'}), 2.75 (m, 1H, H_{3'}), 3.26 (m, 1H, H_{2'}), 3.82 (dd, 1H, H_{5'}, ²J 12.2 Hz, ³J 4.8 Hz), 4.10 (m, 2H, H_{5'}+H_{4'}), 5.80 (m, 2H, H_{7'}+H_{8'}), 5.94 (d, 1H, H_{1'}, ³J 6.0 Hz), 8.46 (s, 1H, H₈), 8.78 (s, 1H, H₂); ¹³C NMR (CDCl₃, 150.9 MHz): δ 22.8, 24.7 (C_{6'}+C_{9'}), 34.0 (C_{3'}), 40.4 (C_{2'}), 63.5 (C_{5'}), 87.2 (C_{4'}), 91.5 (C_{1'}), 124.0, 125.0 (C_{7'}+C_{8'}), 133.1 (C₅), 144.7 (C₈), 150.7 (C₄), 151.6 (C₂), 151.7 (C₆); MS (ESI⁺, *m/z*): 329 [(M³⁵Cl+Na)⁺, 100%] and 331 [(M³⁷Cl+Na)⁺, 20]; Exact mass found for C₁₄H₁₅ClN₄O₂: 306.0682.

HIBF-β-6-Aminopurine (22i). From **20f** using Method B. R_f (5% CH₂Cl₂/MeOH): 0.20; [α]_D²⁰ = -19.6 (c 0.8, DMSO); ¹H NMR (CDCl₃, 400 MHz): δ 1.95-2.12 (m, 2H, H_{6'}+H_{9'}), 2.37-2.49 (m, 2H, H_{6'}+H_{9'}), 2.67 (m, 1H, H_{3'}), 3.42 (m, 1H, H_{2'}), 3.76 (dd, 1H, H_{5'}, ²J 12.4 Hz, ³J 3.6 Hz), 4.06 (m, 2H, H_{4'}+H_{5'}), 5.79 (m, 3H, H_{7'}+H_{8'}, H_{1'}), 5.90 (m, 2H, NH₂), 7.90 (s, 1H, H₈), 8.35 (s, 1H, H₂); ¹³C NMR (CDCl₃, 100.6 MHz): δ 22.8, 25.2 (C_{6'}+C_{9'}), 34.4 (C_{3'}), 39.8 (C_{2'}), 64.7 (C_{5'}), 88.8 (C_{4'}), 91.5 (C_{1'}), 121.1 (C₅), 124.9, 125.6 (C_{7'}+C_{8'}), 140.9 (C₈), 148.6 (C₄), 152.2 (C₂), 155.9 (C₆); MS (ESI⁺, *m/z*): 288 [(M+H)⁺, 100%] and 310 [(M+Na)⁺, 60]; Exact

mass found for C₁₄H₁₇N₅O₂: 287.0713.

HIBF- β -6-Amino-2-chloropurine (22j). From **20g** using Method B. R_f (10% EtOAc/MeOH): 0.44; $[\alpha]_D^{20} = -8.5$ (c 0.3, DMSO); $^1\text{H NMR}$ (CD₂Cl₂, 400 MHz): δ 2.05 (m, 2H, H_{6'}+H_{9'}), 2.42 (m, 2H, H_{6'}+H_{9'}), 2.72 (m, 1H, H_{3'}), 3.16 (m, 1H, H_{2'}), 3.77 (dd, 1H, H_{5'}, 2J 13.2 Hz, 3J 3.6 Hz), 3.77 (dd, 1H, H_{5'}, 2J 13.2 Hz, 3J 3.6 Hz), 4.02 (m, 2H, H_{4'}+H_{5'}), 5.79 (m, 2H, H_{7'}+H_{8'}), 5.83 (d, 1H, H_{1'}, 3J 6.0 Hz), 6.54 (brs, 2H, NH₂), 8.08 (s, 1H, H₈); $^{13}\text{C NMR}$ (CD₂Cl₂, 100.6 MHz): δ 23.4, 25.5 (C_{6'}+C_{9'}), 35.4 (C_{3'}), 40.8 (C_{2'}), 63.8 (C_{5'}), 87.7 (C_{4'}), 91.2 (C_{1'}), 119.8 (C₅), 124.6, 125.5 (C_{7'}+C_{8'}), 140.9 (C₈), 150.4 (C₄), 154.9 (C₂), 156.9 (C₆); MS (ESI⁺, m/z): 322 [(M³⁵Cl+H)⁺, 100%], 324 [(M³⁷Cl+H)⁺, 20], 344 [(M³⁵Cl+Na)⁺, 80], and 346 [(M³⁷Cl+Na)⁺, 10]; C₁₄H₁₆ClN₅O₂.

HIBF- β -6-Methoxypurine (22k). From **20f** using Method C. R_f (EtOAc): 0.39; $^1\text{H NMR}$ (CDCl₃, 500 MHz): δ 1.83 (m, 1H, H_{6'} or H_{9'}), 2.08 (m, 1H, H_{6'} or H_{9'}), 2.33, 2.42 (2m, 2H, H_{6'}, H_{9'}), 2.74 (m, 1H, H_{3'}), 3.45 (m, 1H, H_{2'}), 3.75 (m, 1H, H_{5'}), 4.08 (m, 2H, H_{5'}+H_{4'}), 4.19 (s, 3H, OMe), 5.50 (m, 1H, OH), 5.76 (m, 3H, H_{1'}, H_{7'}, H_{8'}), 8.05 (s, 1H, H₈), 8.55 (s, 1H, H₂); MS (ESI⁺, m/z): 325 [(M+Na)⁺, 100%]; Exact mass found for C₁₅H₁₈N₄O₃: 302.0774.

HIBF- β -2-Chloro-6-methoxypurine (22l). From **20g** using Method C. R_f (EtOAc): 0.44; $^1\text{H NMR}$ (CDCl₃, 600 MHz): δ 2.01 (m, 2H, H_{6'}+H_{9'}), 2.39 (m, 2H, H_{6'}+H_{9'}), 2.72 (m, 1H, H_{3'}), 3.23 (m, 1H, H_{2'}), 3.80 (m, 1H, H_{5'}), 4.04 (m, 2H, H_{4'}+H_{5'}), 5.79 (m, 3H, H_{1'}, H_{7'}, H_{8'}), 8.12 (s, 1H, H₈); MS (ESI⁺, m/z): 359 [(M³⁵Cl+Na)⁺, 100%] and 361 [(M³⁷Cl+Na)⁺, 20]; Exact mass found for C₁₅H₁₇ClN₄O₃: 336.1102.

HIBF- β -2,6-Diaminopurine (22m). From **20g** using Method B. R_f (10% EtOAc/MeOH): 0.17; $[\alpha]_D^{20} = -19.1$ (c 2.5, DMSO); $^1\text{H NMR}$ (MeOH-*d*₄, 400 MHz): δ 2.27 (m, 2H, H_{6'}+H_{9'}), 2.57 (m, 2H, H_{6'}+H_{9'}), 2.78 (m, 1H, H_{3'}), 3.14 (m, 1H, H_{2'}), 3.90 (dd, 1H, H_{5'}, 2J 13.2 Hz, 3J 3.6 Hz), 4.10 (m, 2H, H_{4'}+H_{5'}), 5.92 (m, 2H, H_{7'}+H_{8'}), 5.97 (d, 1H, H_{1'}, 3J 4.8 Hz), 8.31 (s, 1H, H₈); $^{13}\text{C NMR}$ (MeOH-*d*₄, 100.6 MHz): δ 23.0 (C_{6'}), 24.1 (C_{9'}), 34.3 (C_{3'}), 40.6 (C_{2'}), 62.7 (C_{5'}), 86.3 (C_{4'}), 90.0 (C_{1'}), 113.8 (C₅), 124.4, 125.0 (C_{7'}+C_{8'}), 137.4 (C₈), 151.1 (C₄), 156.6 (C₂), 160.6 (C₆); MS (ESI⁺, m/z): 303 [(M+H)⁺, 100%]; C₁₄H₁₈N₆O₂.

HIBF- α -Thymine (23b). From **21b** using Method A. R_f (50% hexane/EtOAc): 0.10; $[\alpha]_D^{20} = -34.5$ (c 0.3, CHCl₃); $^1\text{H NMR}$ (CDCl₃, 500 MHz): δ 1.61-1.82 (m, 2H, H_{6'}, H_{9'}), 1.94 (s, 3H, Me), 2.36 (m, 2H, H_{6'}, H_{9'}), 2.62 (m, 1H, H_{3'}), 2.85 (m, 1H, H_{2'}), 3.81 (m, 1H, H_{4'}), 3.92 (ddd, 1H, H_{5'}, 2J 12 Hz, 3J 6.0 Hz, 4J 3.0 Hz), 4.06 (m, 1H, H_{5'}), 5.69 (m, 2H, H_{7'}+H_{8'}), 5.89 (d, 1H, H_{1'}, 3J 4.0 Hz), 7.45 (d, 1H, H₆, 4J 1.0 Hz), 8.19 (br s, 1H, NH); $^{13}\text{C NMR}$ (CDCl₃, 100.6 MHz): δ 12.9 (Me), 19.9, 23.0 (C_{6'}+C_{9'}), 35.1 (C_{3'}), 37.6 (C_{2'}), 63.2 (C_{5'}), 83.0 (C_{4'}), 89.9 (C_{1'}), 109.3 (C₅), 123.8, 124.6 (C_{7'}+C_{8'}), 136.2 (C₆), 150.1 (C₂), 163.9 (C₄); MS (ESI⁺, m/z): 301 [(M+Na)⁺, 100%]; Exact mass found for C₁₄H₁₈N₂O₄: 278.1260.

HIBF- α -5-Bromovinyluracil (23d). From **21d** using Method A. R_f (50% hexane/EtOAc): 0.14; $^1\text{H NMR}$ (CDCl₃, 600 MHz): δ 1.66 (m, 1H, H_{6'} or H_{9'}), 1.82 (m, 1H, H_{9'} or H_{6'}), 2.01, 2.38 (2m, 2H, H_{6'}, H_{9'}), 2.67 (m, 1H, H_{3'}), 2.89 (m, 1H, H_{2'}), 3.69 (m, 1H, H_{4'}), 3.95 (d, 1H, H_{5'}, 2J 12.0 Hz), 4.08 (d, 1H, H_{5'}, 2J 12.0 Hz), 5.70 (m, 2H, H_{7'}+H_{8'}), 5.92 (d, 1H, H_{1'}, 3J 4.0 Hz), 6.73 (d, 1H, H₇, 3J 13.8 Hz), 7.43 (d, 1H, H₈, 3J 13.8 Hz), 7.64 (s, 1H, H₆), 8.12 (br s, 1H, NH); MS (ESI⁺, m/z): 369 [(M⁷⁹Br+H)⁺, 100] and 371 [(M⁸¹Br+H)⁺, 96]; Exact mass found for C₁₅H₁₇BrN₂O₄: 368.0874.

HIBF- α -6-Chloropurine (23f). From **21f** using Method A. R_f (EtOAc): 0.40; $[\alpha]_D^{20} = -9.7$ (c 0.8, CHCl₃); $^1\text{H NMR}$ (CDCl₃, 600 MHz): δ 1.50-1.75 (m, 2H, H_{6'}+H_{9'}), 2.00, 2.48 (m, 2H, H_{6'}+H_{9'}), 2.83 (m, 1H, H_{3'}), 3.06 (m, 1H, H_{2'}), 3.75 (dd, 1H, H_{5'}, 2J 12.6 Hz, 3J 4.2 Hz), 4.02 (dd, 1H, H_{5'}, 2J 12.6 Hz, 3J 2.4 Hz), 4.23 (dt, 1H, H_{4'}, 3J 10.2 Hz, 3J 3 Hz), 5.67 (m, 2H, H_{7'}+H_{8'}), 6.40 (d, 1H, H_{1'}, 3J 4.2 Hz), 8.43 (s, 1H, H₈), 8.74 (s, 1H, H₂); $^{13}\text{C NMR}$ (CDCl₃, 150.9 MHz): δ 19.7, 23.1 (C_{6'}+C_{9'}), 35.2 (C_{3'}), 38.2 (C_{2'}), 62.8 (C_{5'}), 83.2 (C_{4'}), 89.5 (C_{1'}), 123.5, 124.3

(C₇+C₈), 132.1 (C₅), 143.3 (C₈), 150.7 (C₄), 152.0 (C₂), 155.5 (C₆); MS (ESI⁺, *m/z*): 329 [(M³⁵Cl+Na)⁺, 100%] and 331 [(M³⁷Cl+Na)⁺, 30]; Exact mass found for C₁₄H₁₅ClN₄O₂: 306.0548.

HIBF- α -6-Aminopurine (23i). From **21f** using Method B. R_f (5% CH₂Cl₂/MeOH): 0.09; [α]_D²⁰ = -9.6 (c 2.5, MeOH); ¹H NMR (MeOH-*d*₄, 400 MHz): δ 1.50, 1.85 (2m, 2H, H₆+H₉), 2.10, 2.42 (2m, 2H, H₆+H₉), 2.75 (m, 1H, H₃), 2.90 (m, 1H, H₂), 3.75 (dd, 1H, H₅, ²J 12.2 Hz, ³J 4.8 Hz), 3.95 (m, 1H, H₅, ²J 12.2 Hz, ³J 2.8 Hz), 4.16 (m, 1H, H₄), 5.74 (m, 2H, H₇+H₈), 6.39 (d, 1H, H₁, ³J 4.4 Hz), 8.23 (s, 1H, H₈), 8.33 (s, 1H, H₂); ¹³C NMR (MeOH-*d*₄, 100.6 MHz): δ 19.4, 22.9 (C₆+C₉), 35.5 (C₃), 38.3 (C₂), 62.1 (C₅), 83.5 (C₄), 89.0 (C₁), 119.0 (C₅), 123.4, 124.2 (C₇+C₈), 139.0 (C₈), 148.2 (C₄), 152.4 (C₂), 155.9 (C₆); MS (ESI⁺, *m/z*): 288 [(M+H)⁺, 100%] and 310 [(M+Na)⁺, 60]; Exact mass found for C₁₄H₁₇N₅O₂: 287.0772.

HIBF- α -6-Amino-2-chloropurine (23j). From **21g** using Method B. R_f (6.6% EtOAc/MeOH): 0.37; [α]_D²⁰ = -25.9 (c 1.0, DMSO); ¹H NMR (MeOH-*d*₄, 400 MHz): δ 1.73 (m, 1H, H₆), 2.02 (m, 1H, H₉), 2.26 (m, 1H, H₆), 2.58 (m, 1H, H₉), 2.91 (m, 1H, H₃), 3.07 (m, 1H, H₂), 3.85 (dd, 1H, H₅, ²J 12.2 Hz, ³J 4.6 Hz), 4.05 (dd, 1H, H₅, ²J 12.2 Hz, ³J 2.8 Hz), 4.35 (ddd, 1H, H₄, ³J 9.5 Hz, 4.6 Hz, 2.8 Hz), 5.79 (m, 2H, H₇+H₈), 6.42 (d, 1H, H₁, ³J 4.4 Hz), 8.42 (s, 1H, H₈); ¹³C NMR (MeOH-*d*₄, 100.6 MHz): δ 19.8 (C₆), 23.3 (C₉), 35.9 (C₃), 38.7 (C₂), 62.5 (C₅), 84.0 (C₄), 89.3 (C₁), 118.2 (C₅), 123.8, 124.6, (C₇+C₈), 139.7 (C₈), 150.0 (C₄), 154.3 (C₂), 157.0 (C₆); MS (ESI⁺, *m/z*): 322 [(M³⁵Cl+H)⁺, 100%] and 324 [(M³⁷Cl+H)⁺, 20]; C₁₄H₁₆ClN₅O₂.

HIBF- α -2,6-Diaminopurine (23m). From **21g** using Method B. R_f (10% EtOAc/MeOH): 0.12; [α]_D²⁰ = -59.8 (c 0.8, MeOH); ¹H NMR (MeOH-*d*₄, 600 MHz): δ 1.72 (m, 1H, H₆), 2.04 (m, 1H, H₆), 2.26 (m, 1H, H₉), 2.58 (m, 1H, H₉), 2.89 (m, 1H, H₃), 3.06 (m, 1H, H₂), 3.84 (dd, 1H, H₅, ²J 12.0 Hz, ³J 4.8 Hz), 4.03 (dd, 1H, H₅, ²J 12.0 Hz, ³J 3.0 Hz), 4.29 (m, 1H, H₄), 5.81 (m, 2H, H₇+H₈), 6.32 (d, 1H, H₁, ³J 4.8 Hz), 8.12 (s, 1H, H₈); ¹³C NMR (MeOH-*d*₄, 150.9 MHz): δ 19.4 (C₆), 22.9 (C₉), 35.5 (C₃), 38.1 (C₂), 62.1 (C₅), 83.2 (C₄), 88.5 (C₁), 113.0 (C₅), 123.6, 124.1 (C₇+C₈), 136.0 (C₈), 150.5 (C₄), 156.1 (C₂), 160.4 (C₆); MS (ESI⁺, *m/z*): 303 [(M+H)⁺, 100%]; C₁₄H₁₈N₆O₂.

HIBF- α / β -Uracil (24a). From **20a+21a** using Method A. R_f (EtOAc): 0.40 β , 0.36 α ; ¹H NMR (CDCl₃, 500 MHz): δ 1.80-2.89 (m, 12H, 4H₆, 4H₉, 2H₂, 2H₃), 3.48 (d, 1H, OH, ³J 3.5 Hz), 3.64 (m, 1H), 3.79 (dt, 1H, H₅, ²J 12.0 Hz, ³J 3.0 Hz), 3.92 (m, 1H), 3.96 (dt, 1H, H₃, ³J 9.0 Hz, ³J 3.0 Hz), 4.07 (m, 2H), 5.70 (m, 7H, 2H₇+2H₈+H₁+2H₅), 5.90 (d, 1H, H₁, ³J 4.5 Hz), 7.65 (d, 1H, H₆, ³J 8.0 Hz), 8.05 (d, 1H, H₆, ³J 8.5 Hz), 8.60 (br s, 1H, NH), 8.73 (br s, 1H, NH); ¹³C NMR (CDCl₃, 100.6 MHz): δ 19.8, 23.0, 23.1, 23.7 (C₆+C₉), 32.7 (C₃), 35.1 (C₃), 37.6 (C₂), 41.6 (C₂), 62.0 (C₅), 63.0 (C₅), 83.1 (C₄), 85.0 (C₄), 90.1 (C₁), 91.2 (C₁), 101.0 (C₅), 101.4 (C₅), 123.7, 124.6, 124.7, 124.8 (2C₇+2C₈), 140.4 (C₆), 141.0 (C₆), 150.3 (C₂), 150.7 (C₂), 163.6 (C₄), 163.7 (C₄); MS (ESI⁺, *m/z*): 287 [(M+Na)⁺, 100%].

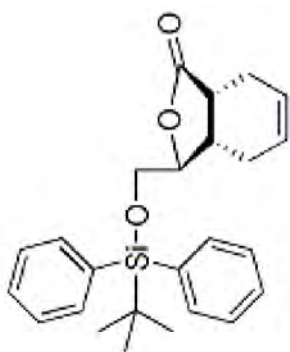
HIBF- α / β -Cytosine (24h). From **20h+21h** using Method A. R_f (EtOAc/MeOH 10%): 0.20; [α]_D²⁰ = -28.6 (c 0.7, MeOH); ¹H NMR (MeOH-*d*₄, 500 MHz): δ 1.90-2.84 (m, 12H, 2H₆+2H₉+2H₃+2H₂), 3.62 (m, 1H), 3.74 (m, 1H), 3.82 (m, 1H), 3.90 (m, 1H), 3.94 (m, 1H), 4.00 (m, 1H), 5.66 (m, 1H, H₁), 5.70 (m, 3H, H₇+H₈+H₅), 5.86 (m, 4H, H₁+H₇+H₈+H₅), 7.86 (d, 1H, H₆, ³J 8.0 Hz), 8.33 (d, 1H, H₆, ³J 7.0 Hz); ¹³C NMR (MeOH-*d*₄, 100.6 MHz): δ 20.7, 23.6, 23.9, 24.8 (C₆+C₉), 33.3 (C₃), 36.6 (C₃), 38.8 (C₂), 43.0 (C₂), 61.8 (C₅), 63.5 (C₅), 84.3 (C₄), 86.4 (C₄), 91.5 (C₁), 92.7 (C₁), 94.6 (C₅), 94.8 (C₅), 124.8, 125.5, 125.6, 125.7 (C₇+C₈), 142.7 (C₆), 142.9 (C₆), 158.2 (C₂), 158.4 (C₂), 167.7 (C₄), 167.8 (C₄); MS (ESI⁺, *m/z*): 264 [(M+H)⁺, 70%] and 286 [(M+Na)⁺, 100]; Exact mass found for C₁₃H₁₇N₃O₃: 263.1318.

Synthesis of HIBF- β -2,6-Diaminopurine (20m). A solution of **20n** (30 mg, 0.076 mmol) in

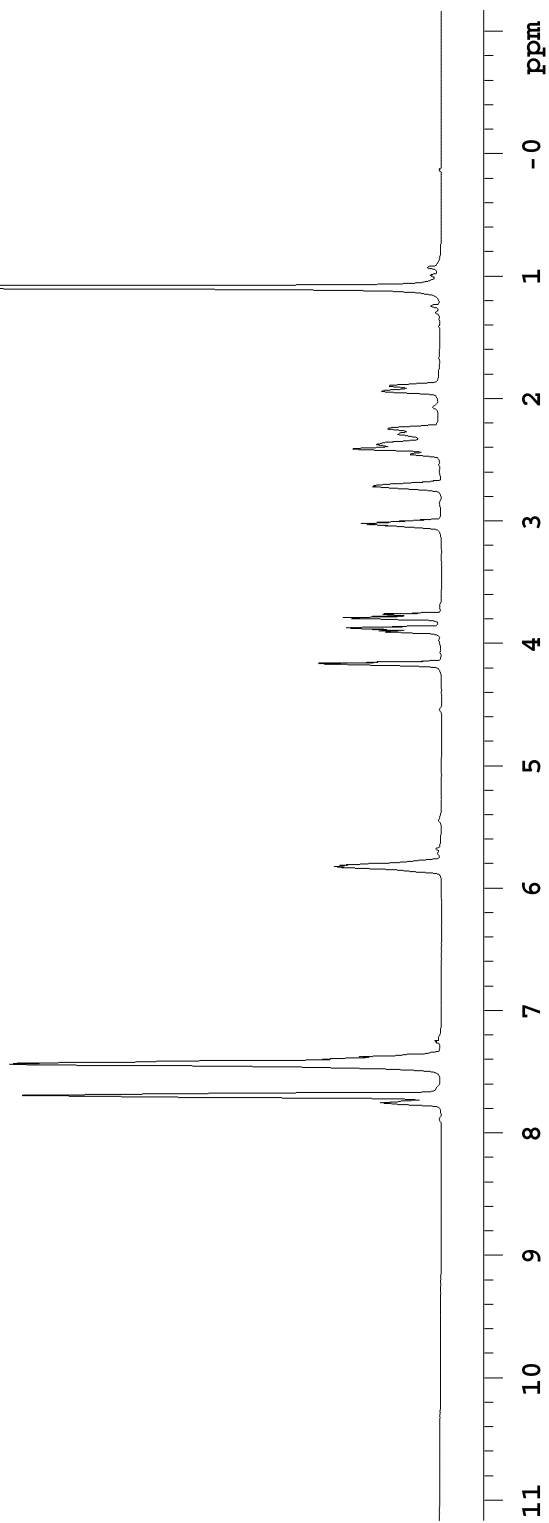
a mixture THF/H₂O was treated with Me₃P (15.4 μL, 0.174 mmol). Immediately, N₂ gave off and the solution became yellowish. The reaction was stirred for 26 h and then solvents were removed. The residue was purified by silica gel chromatography (2-10% MeOH/EtOAc) to give **20m** in 73% yield. R_f (20% EtOAc/MeOH): 0.49; $^1\text{H NMR}$ (CDCl₃, 500 MHz): δ 2.06 (m, 2H, H_{6'}+H_{9'}), 2.09 (s, 3H, CH₃), 2.34-2.47 (m, 3H, H_{3'}, H_{6'}, H_{9'}), 3.04 (m, 1H, H_{2'}), 4.13 (m, 1H, H_{4'}), 4.40 (m, 2H, H_{5'}), 4.72 (brs, 2H, NH₂), 5.40 (br s, 2H, NH₂), 5.75 (m, 2H, H_{7'}+H_{8'}), 5.78 (d, H_{1'}, ^3J 3.5 Hz), 7.82 (s, 1H, H₈); MS (ESI⁺, m/z): 367 [(M+Na)⁺, 100%]; C₁₆H₂₀N₆O₃.

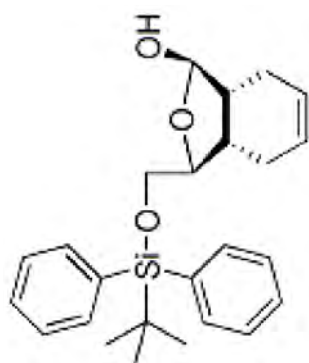
Synthesis of HIBF-β-2,6-Diazidopurine (20n). A solution of **20g** (0.262 mmol, 100 mg) in DMF (1.8 mL) was treated with NaN₃ (1.048 mmol, 68 mg) at 70 °C for 4 h. Solvent was removed and the residue obtained was purified by silica gel flash chromatography (gradient 30-50% EtOAc/hexane) to give **20n** in 76% yield. R_f (50% hexane/EtOAc): 0.47; $^1\text{H NMR}$ (CDCl₃, 400 MHz): δ 2.10 (m, 2H, H_{6'}+H_{9'}), 2.12 (s, 3H, CH₃), 2.35-2.48 (m, 3H, H_{3'}, H_{6'}, H_{9'}), 2.98 (m, 1H, H_{2'}), 4.16 (m, 1H, H_{4'}), 4.49 (m, 2H, H_{5'}), 5.74 (m, 2H, H_{7'}+H_{8'}), 5.92 (s, 1H, H_{1'}), 8.29 (s, 1H, H₈); MS (ESI⁺, m/z): 419 [(M+Na)⁺, 100%]; C₁₆H₂₀N₁₀O₃.

Synthesis of HIBF-β-Hypoxanthine 22o. Nucleoside **22i** (20 mg, 0.070 mmol) was dissolved in a phosphate buffer pH=7/ 3% DMSO (400 μL) and treated with 1 mg of ADA dissolved in that buffer (100 μL). The residue was purified by flash chromatography (gradient 5%-20% MeOH/EtOAc) to give **22o** (50%). R_f (10% EtOAc/MeOH): 0.10; $^1\text{H NMR}$ (MeOH-*d*₄, 600 MHz): δ 2.25 (m, 2H, H_{6'}+H_{9'}), 2.65 (m, 2H, H_{6'}+H_{9'}), 2.83 (m, 1H, H_{3'}), 3.06 (m, 1H, H_{2'}), 3.94 (dd, 1H, H_{5'}, ^2J 12.3 Hz, ^3J 3.6 Hz), 4.12 (dd, 1H, H_{5'}, ^2J 12.3 Hz, ^3J 3.0 Hz), 4.17 (dt, 1H, H_{4'}, ^3J 8.4 Hz, 3.6 Hz), 5.96 (m, 2H, H_{7'}+H_{8'}), 6.17 (d, 1H, H_{1'}, ^3J 3.6 Hz), 8.23 (s, 1H, H₈), 8.73 (s, 1H, H₂); $^{13}\text{C NMR}$ (MeOH-*d*₄, 150.9 MHz): δ 22.9, 23.0 (C_{6'}+C_{9'}), 33.1 (C_{3'}), 41.3 (C_{2'}), 61.4 (C_{5'}), 85.7 (C_{4'}), 90.1 (C_{1'}), 123.8 (C_{7'} or C_{8'}), 124.4 (C₅), 123.9 (C_{8'} or C_{7'}), 139.1 (C₈), 145.2 (C₄), 148.0 (C₂), 157.5 (C₆); MS (ESI⁺, m/z): 289 [(M+H)⁺, 100%] and 311 [(M+Na)⁺, 60].



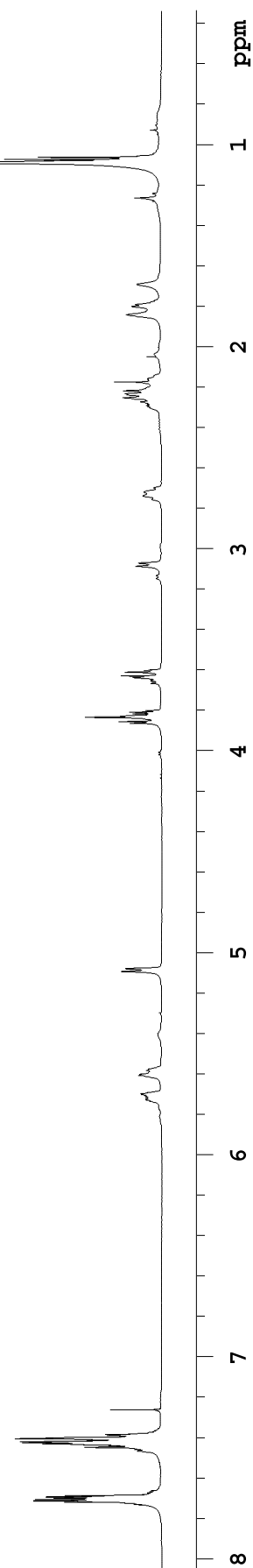
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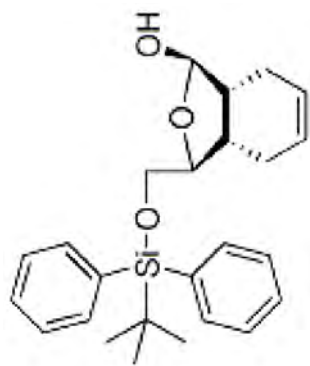




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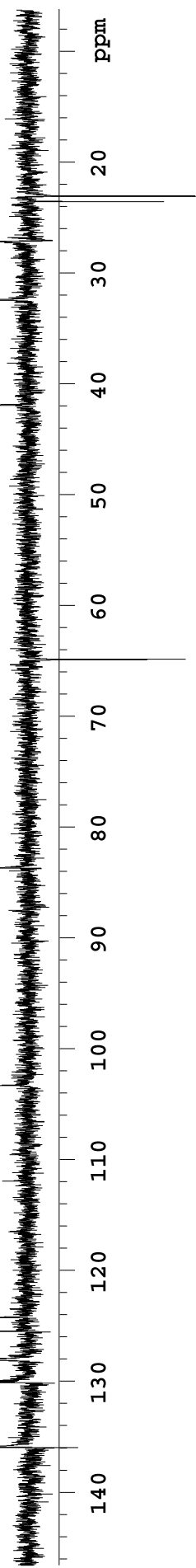
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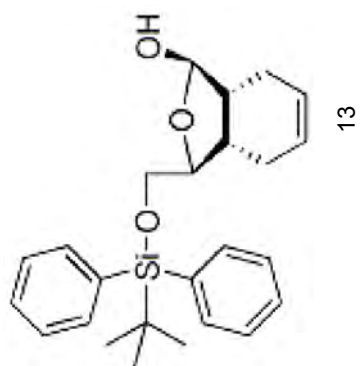




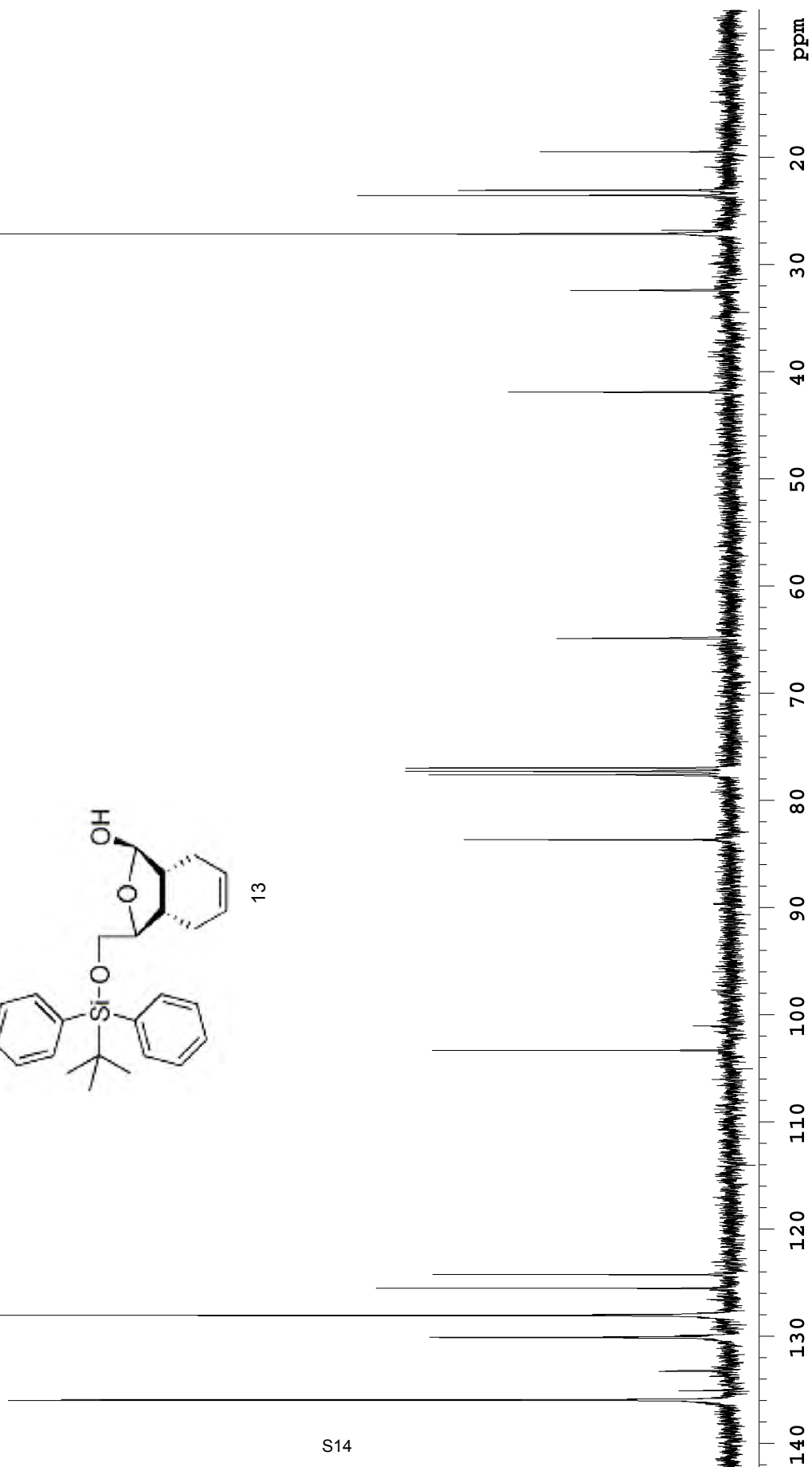
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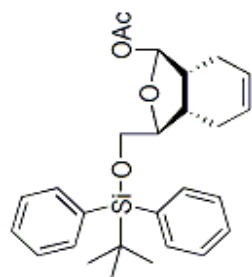
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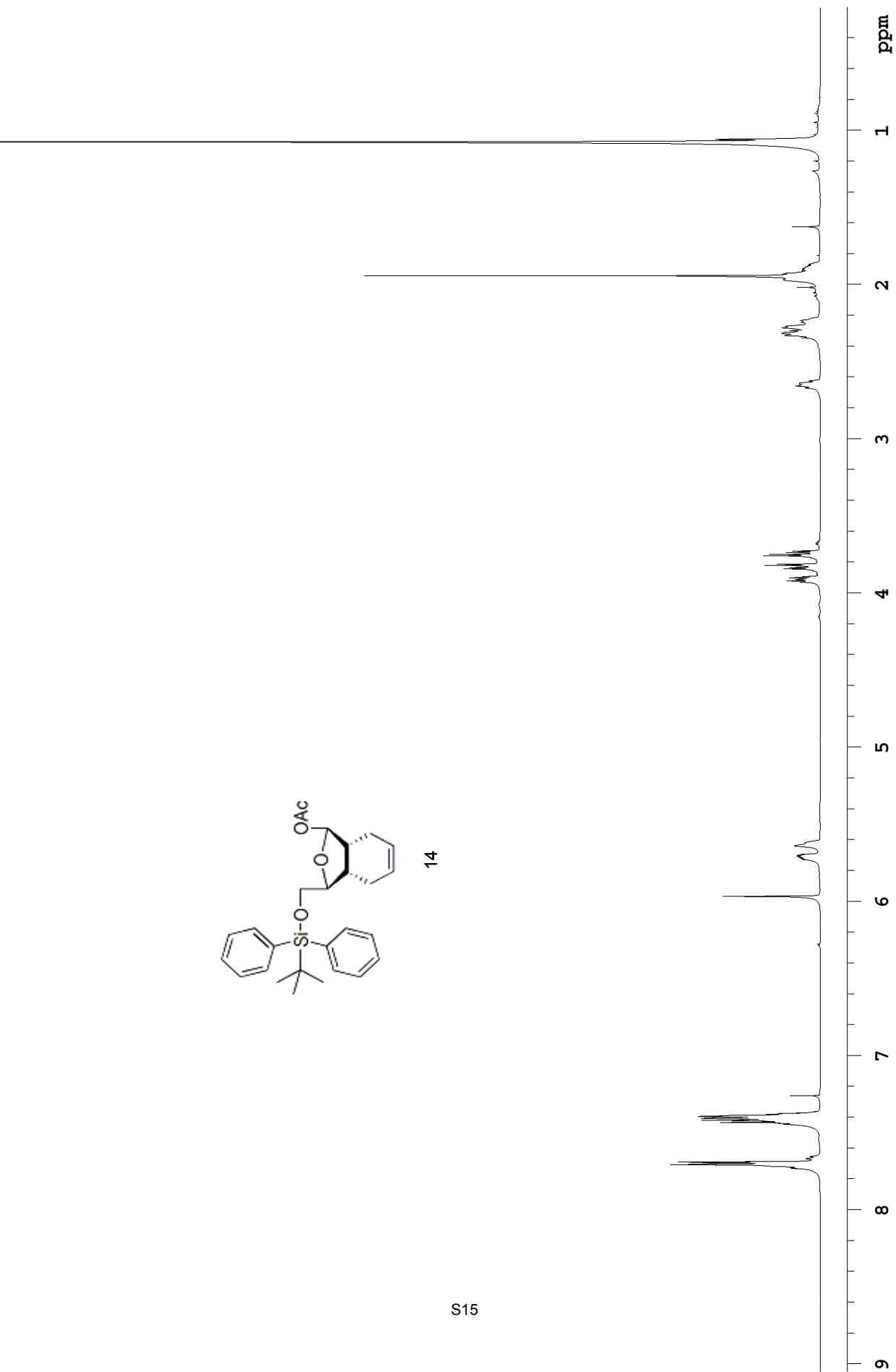


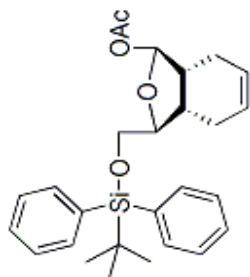
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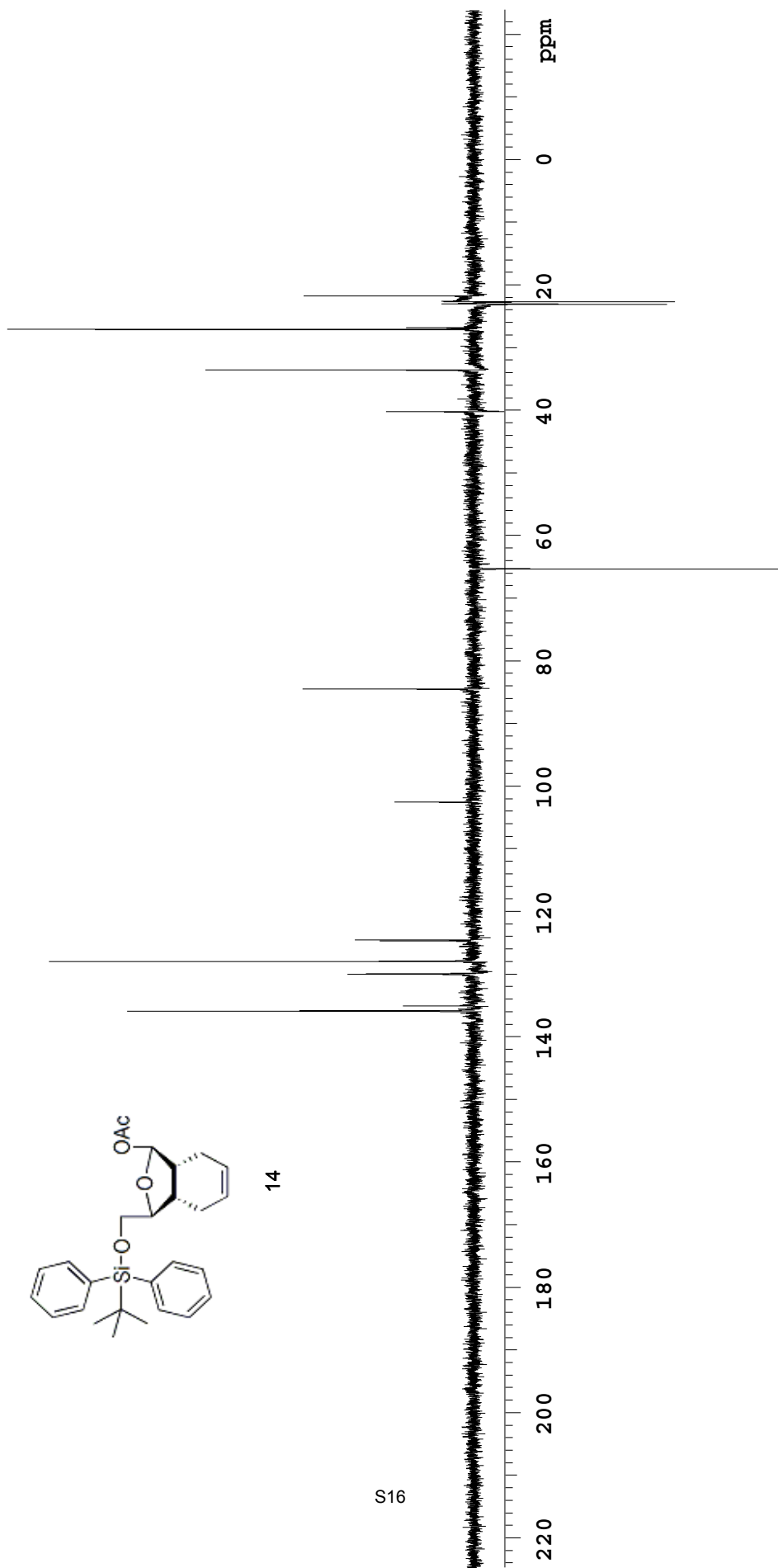


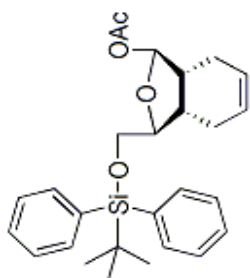
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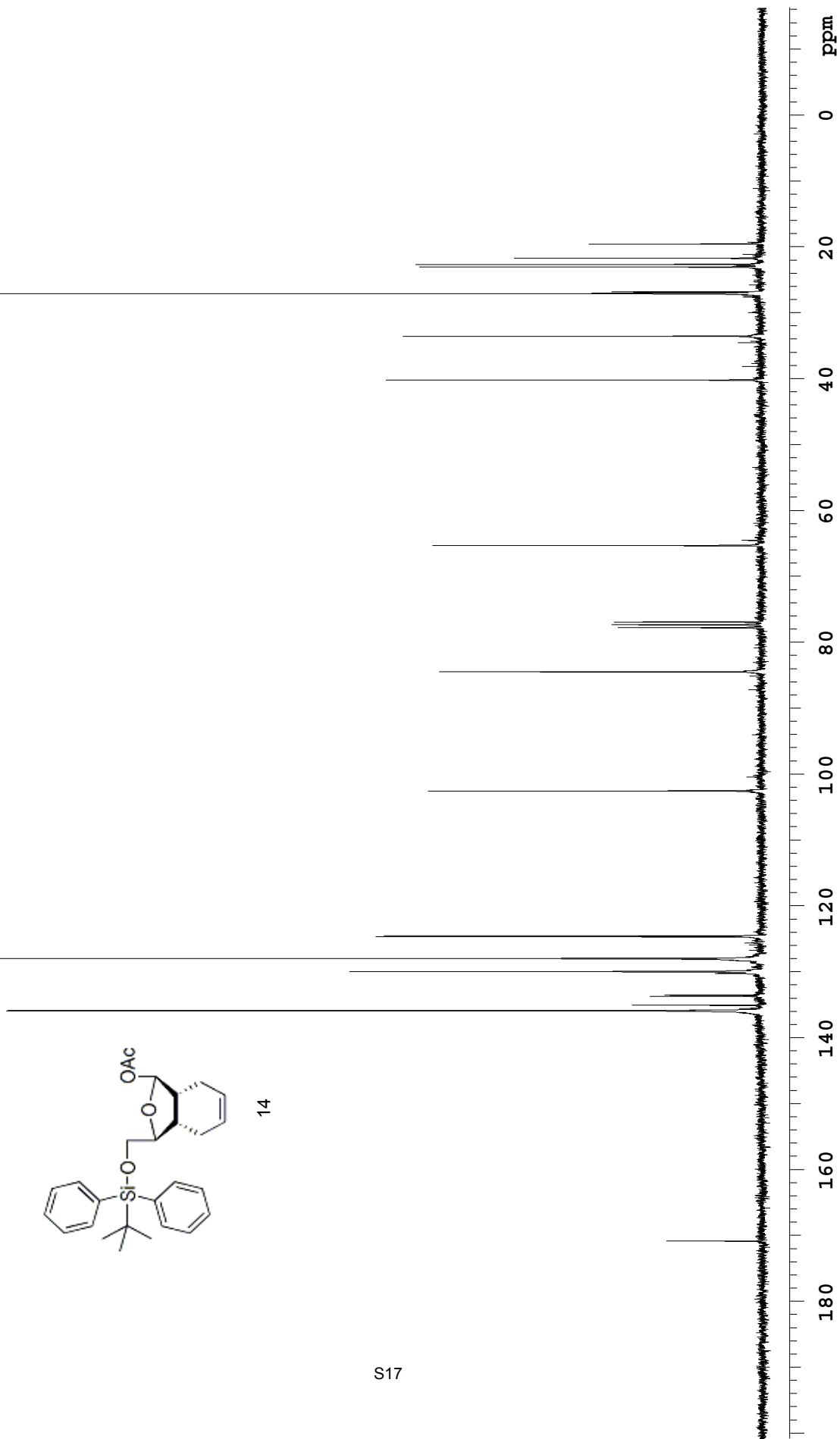


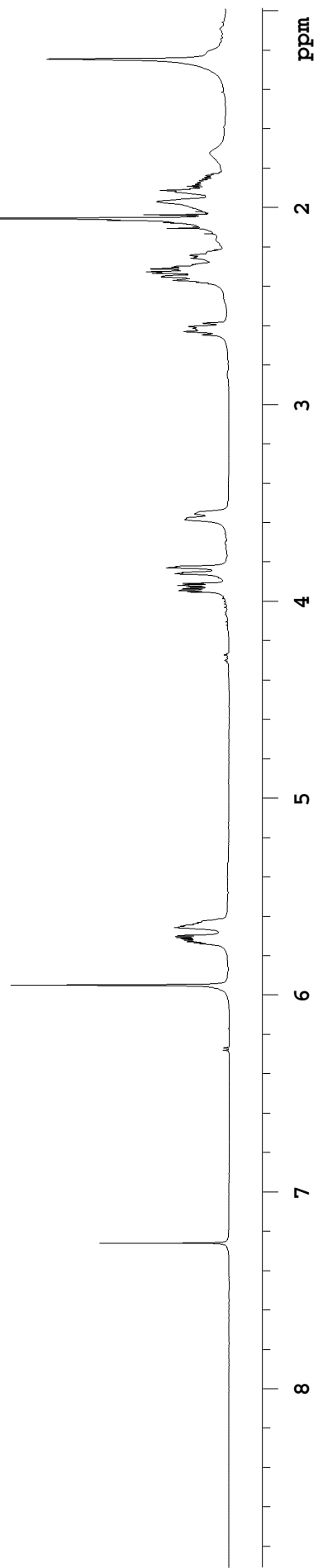
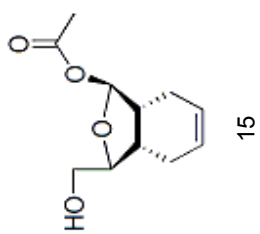
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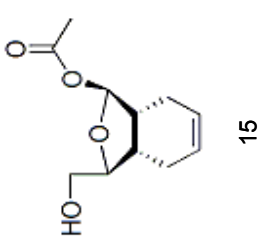
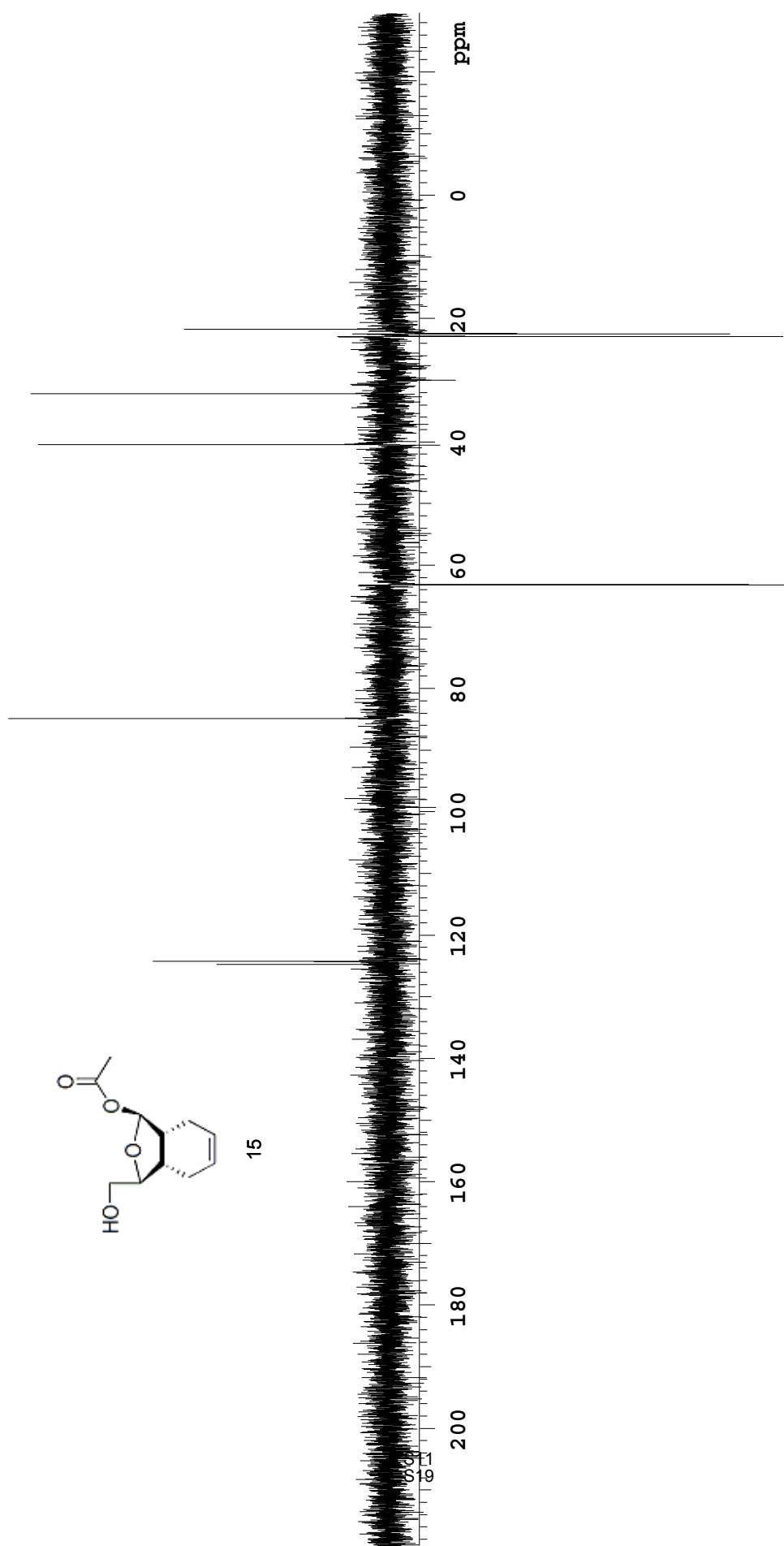


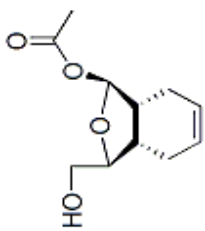


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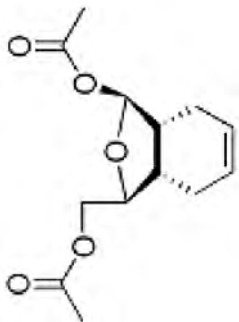




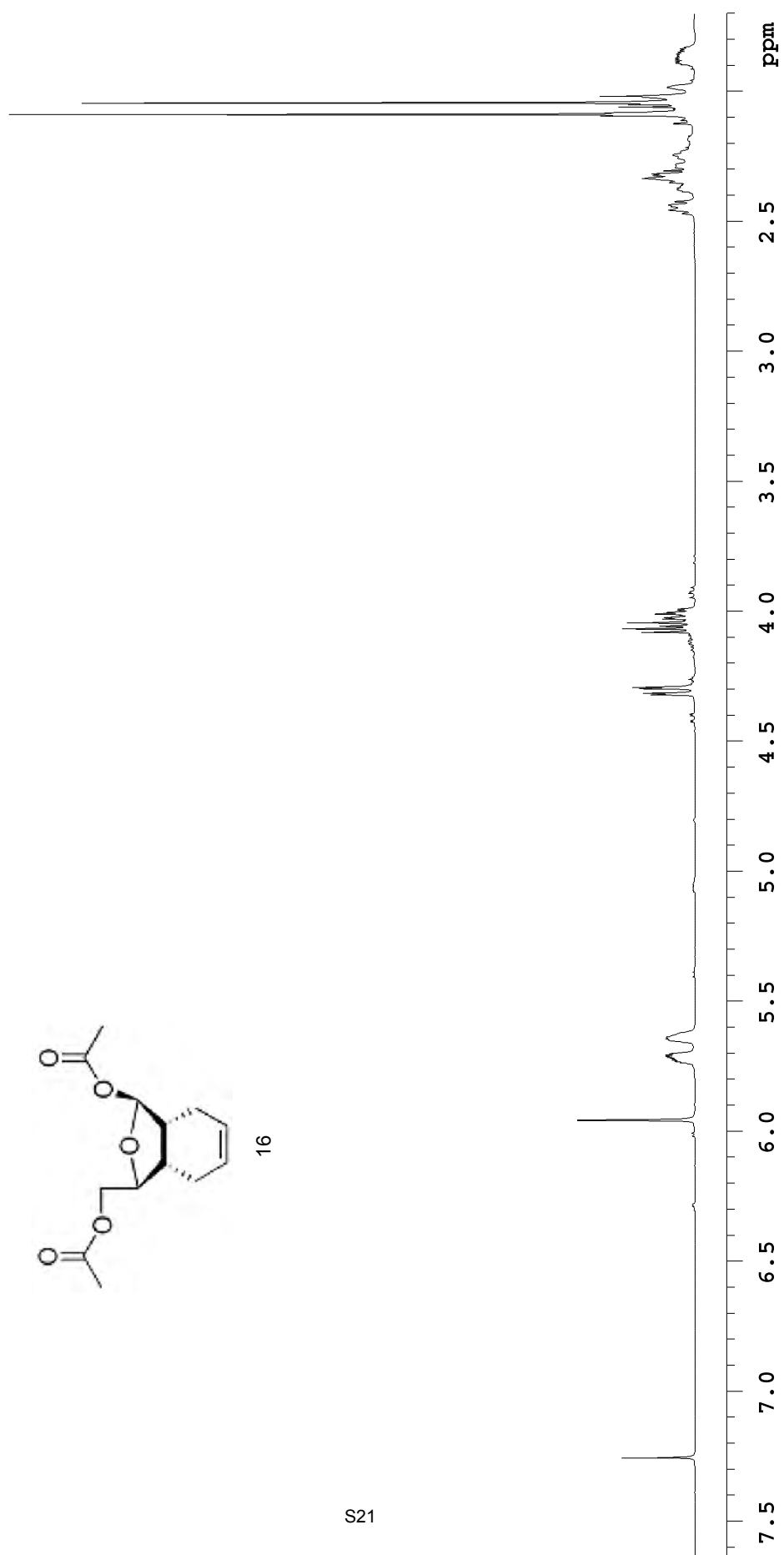
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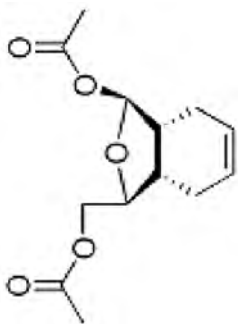
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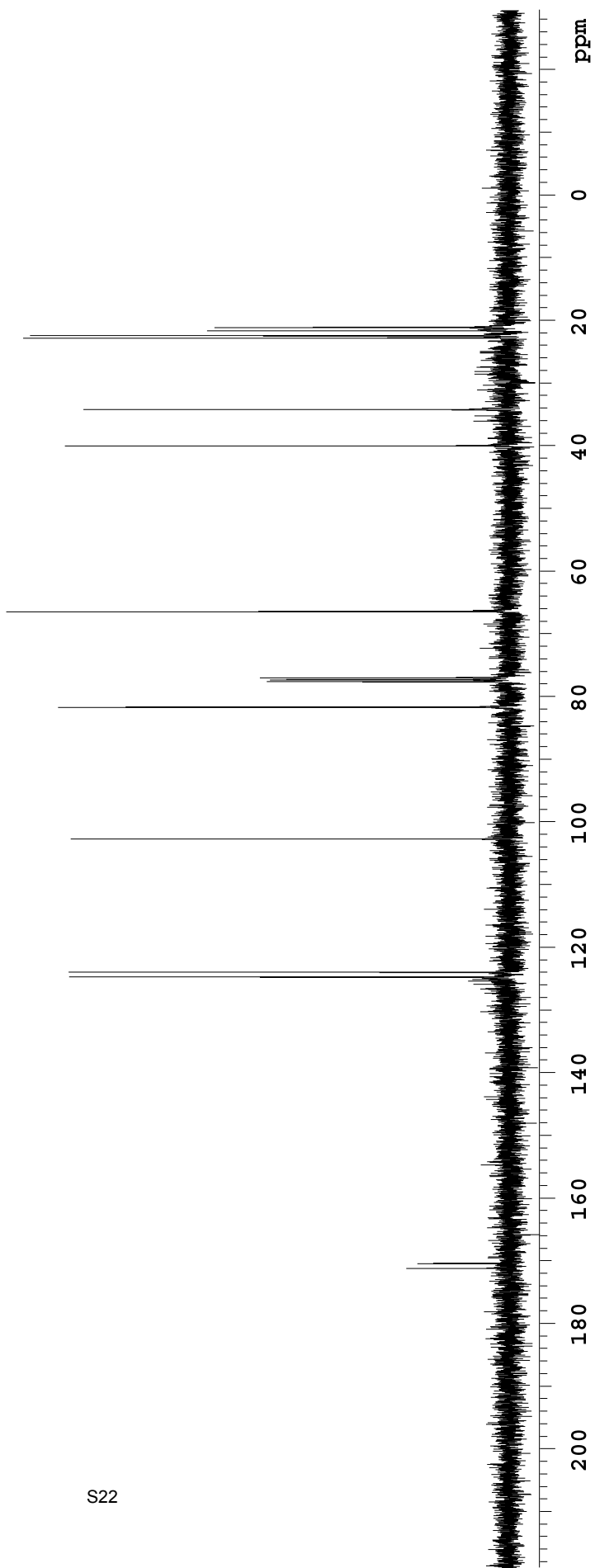


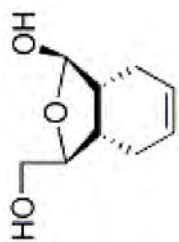
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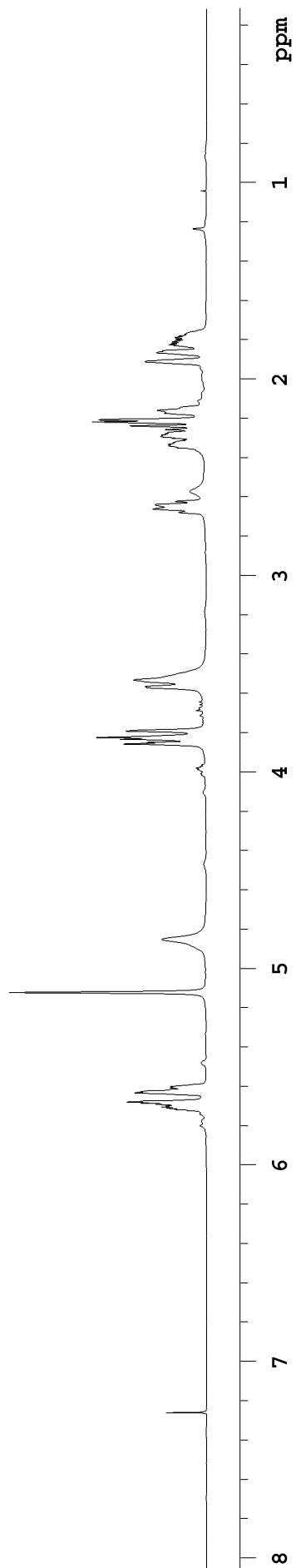


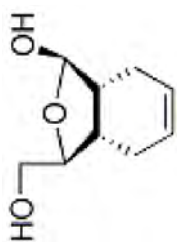
16



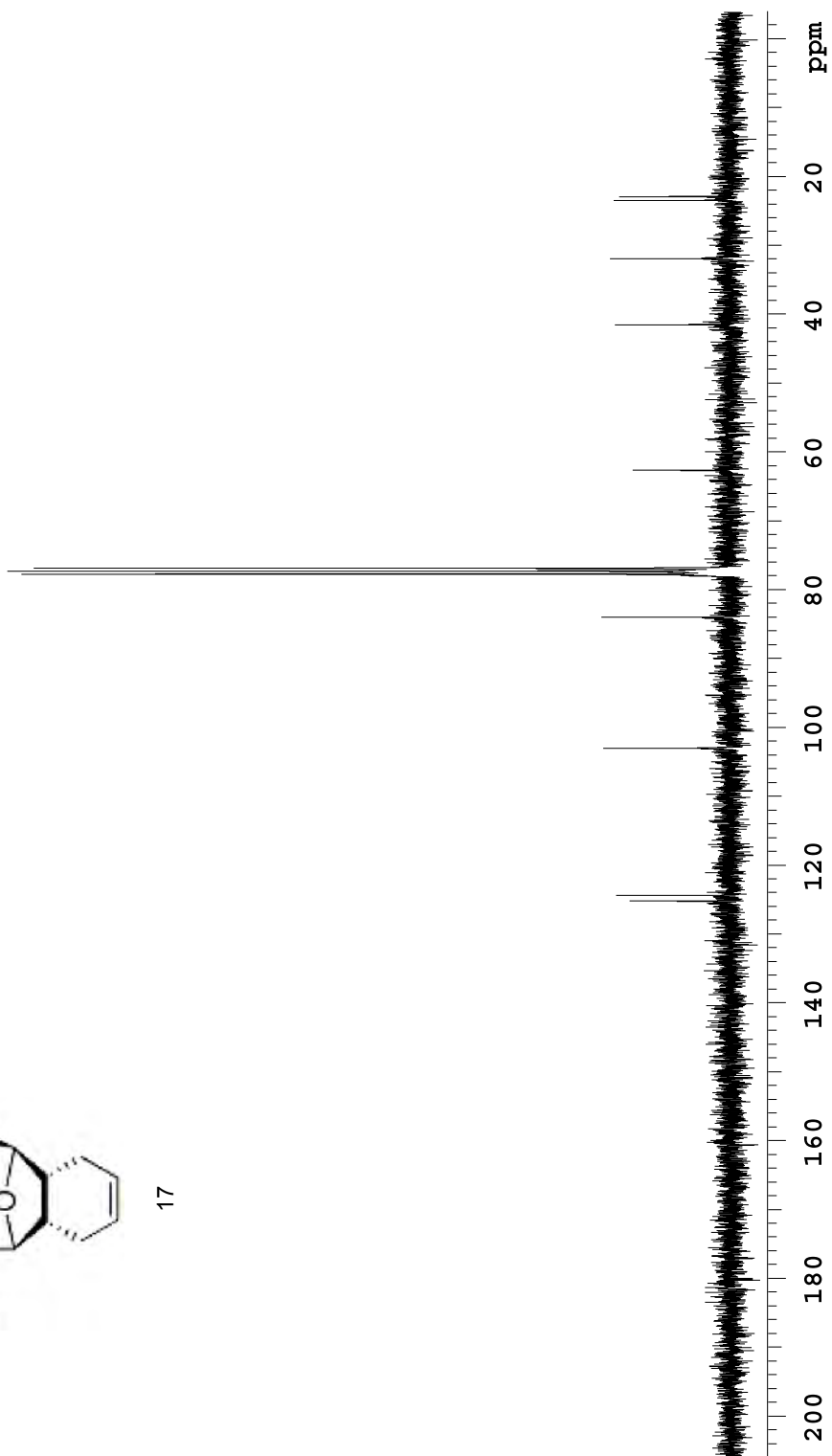


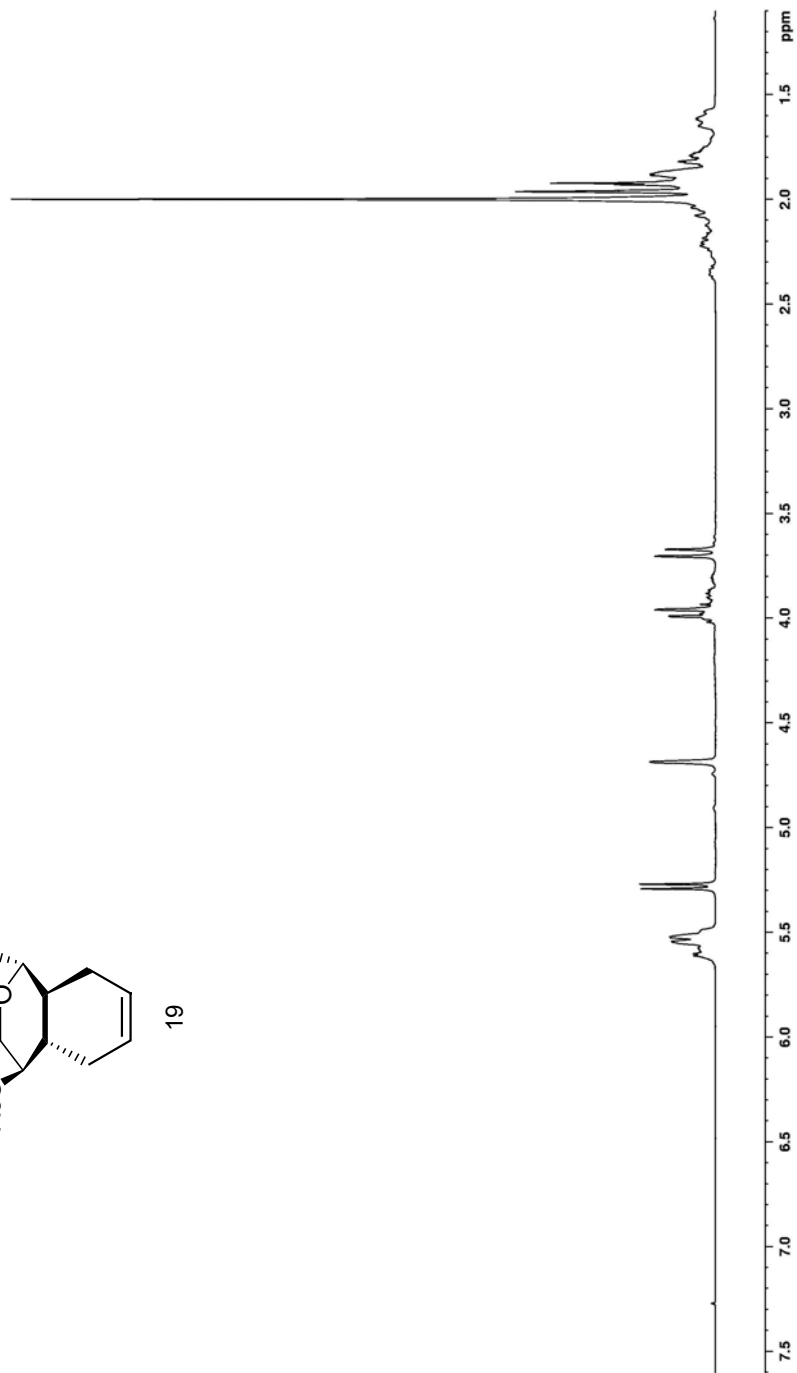
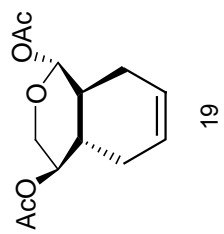
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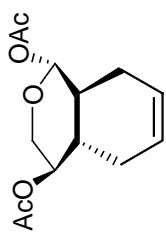




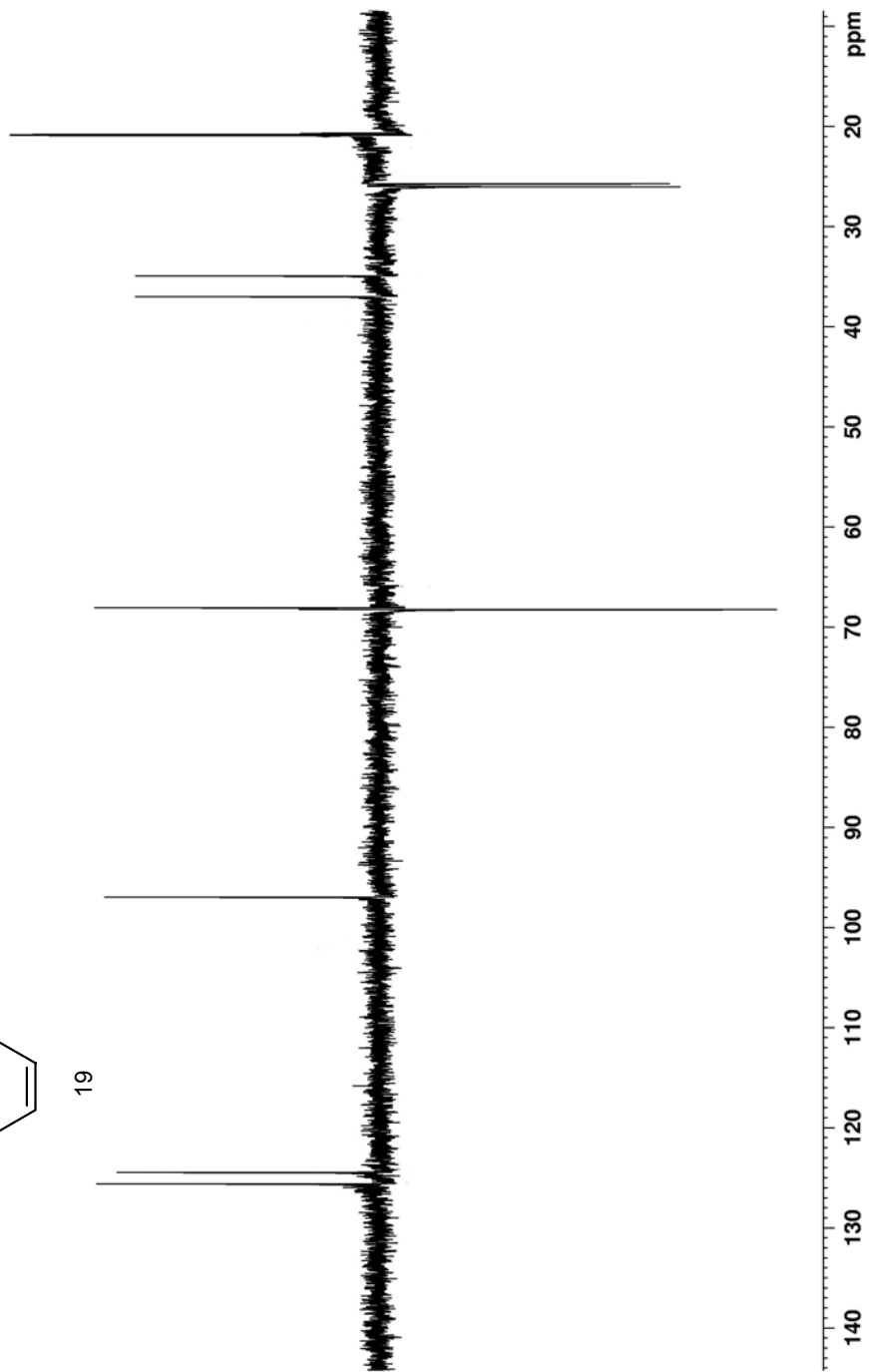
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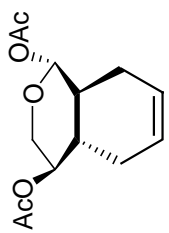




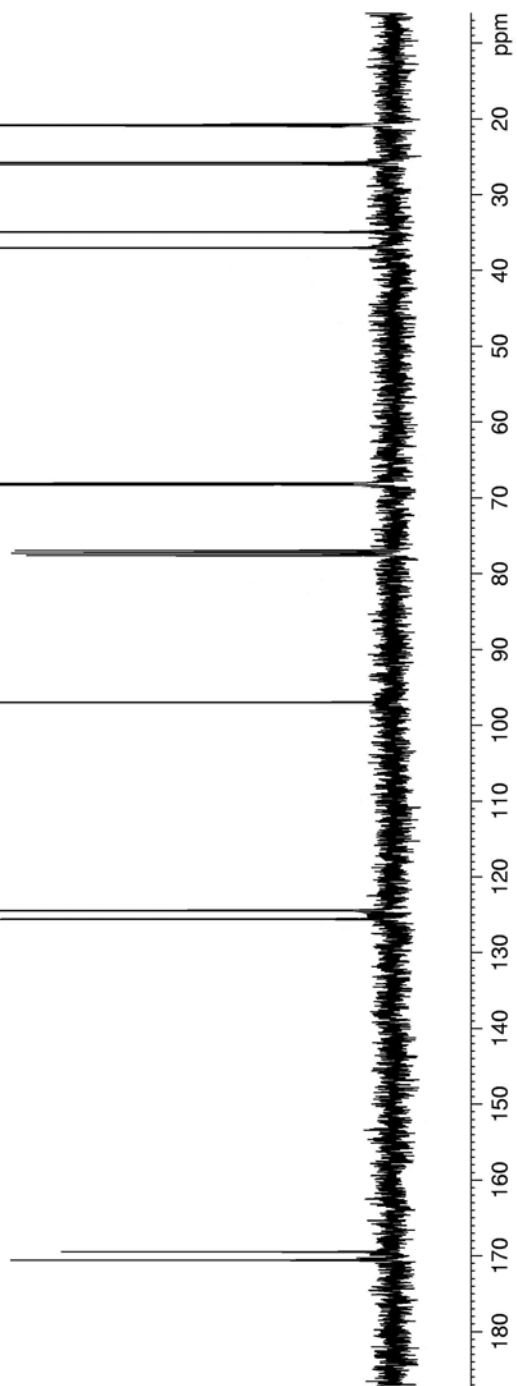


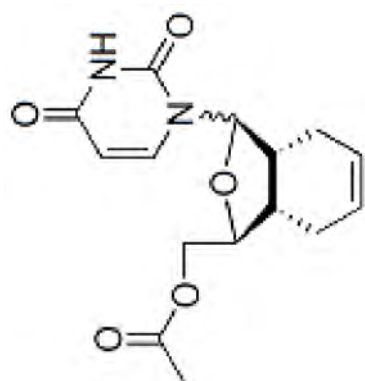
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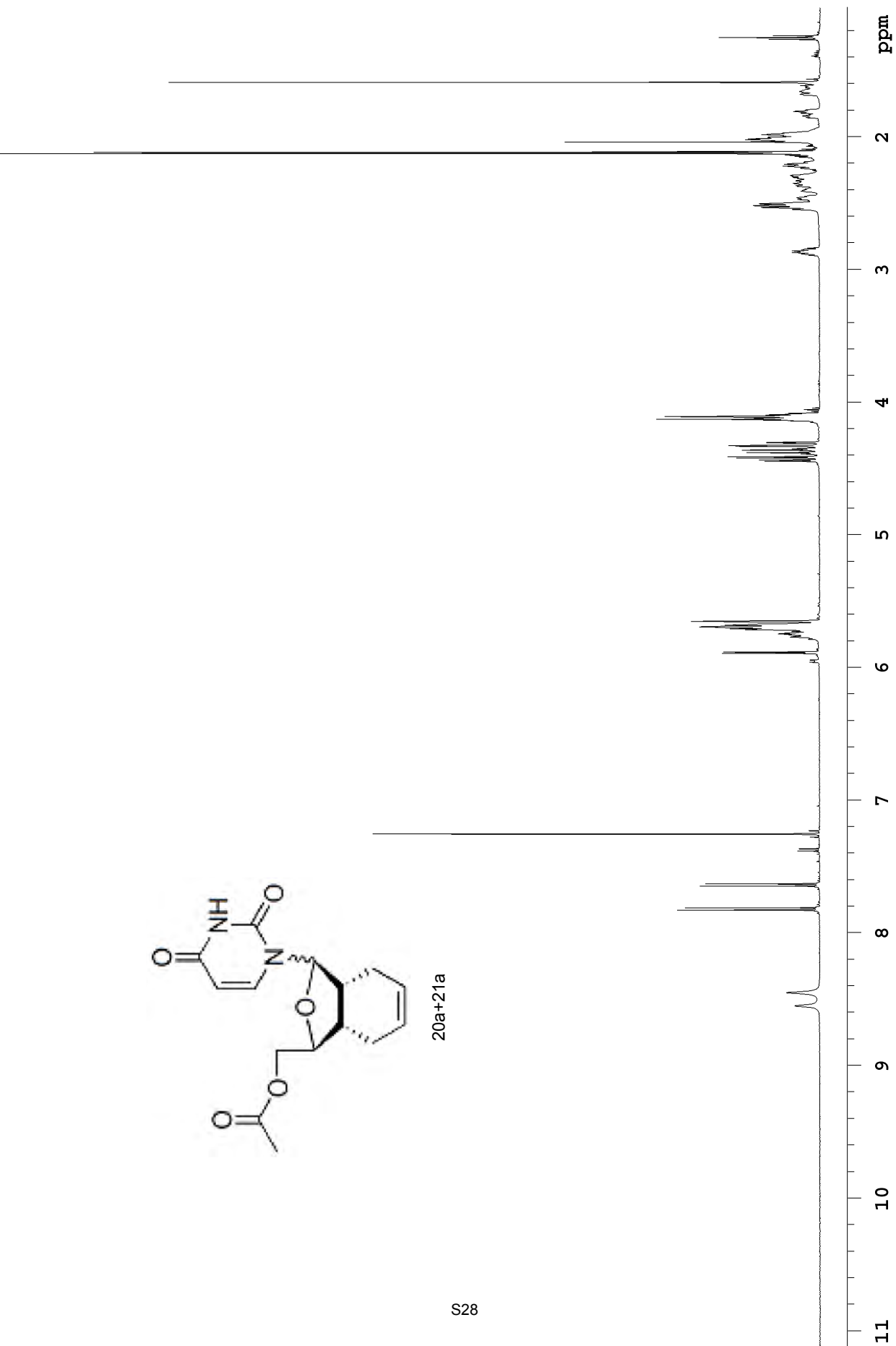


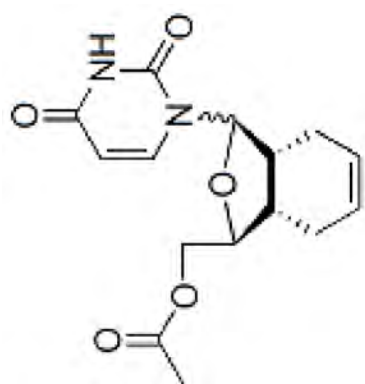
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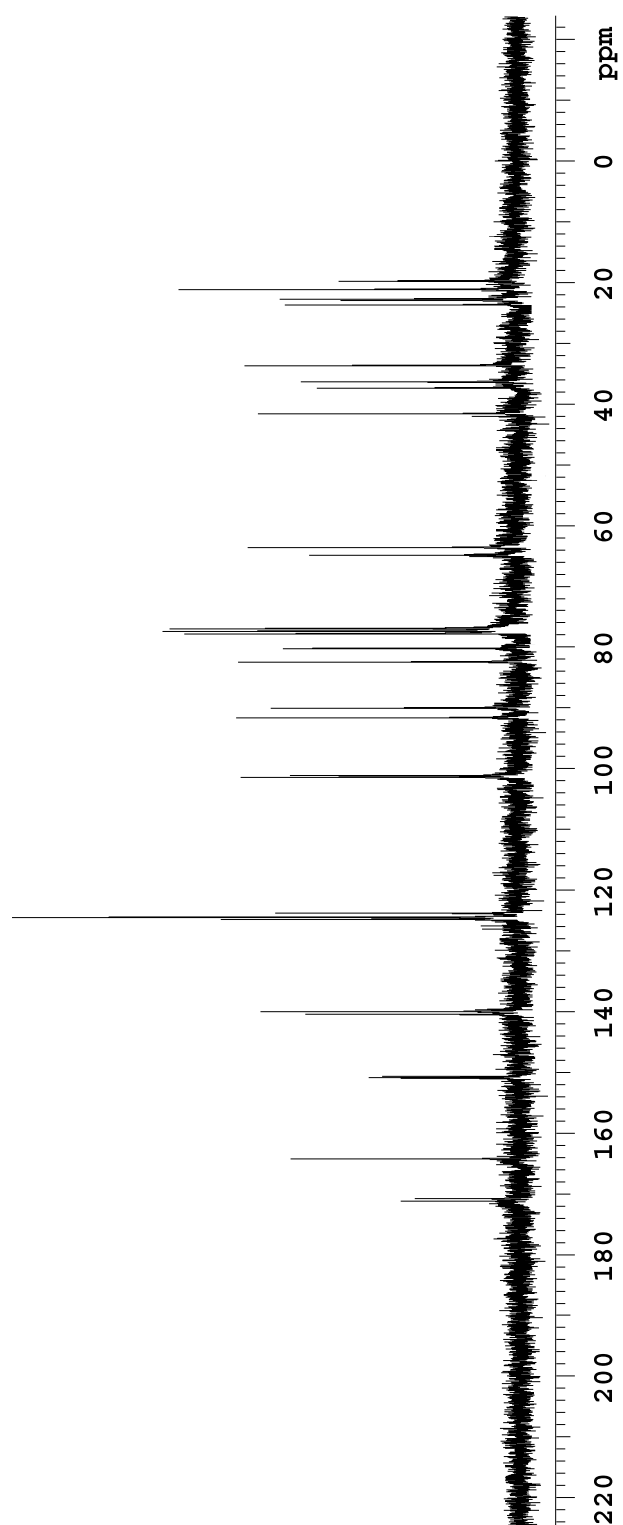


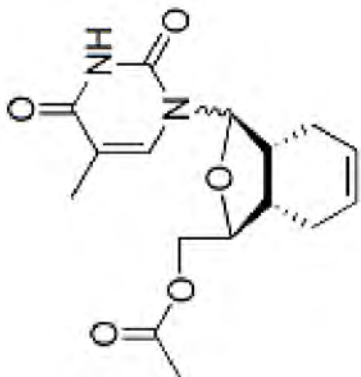
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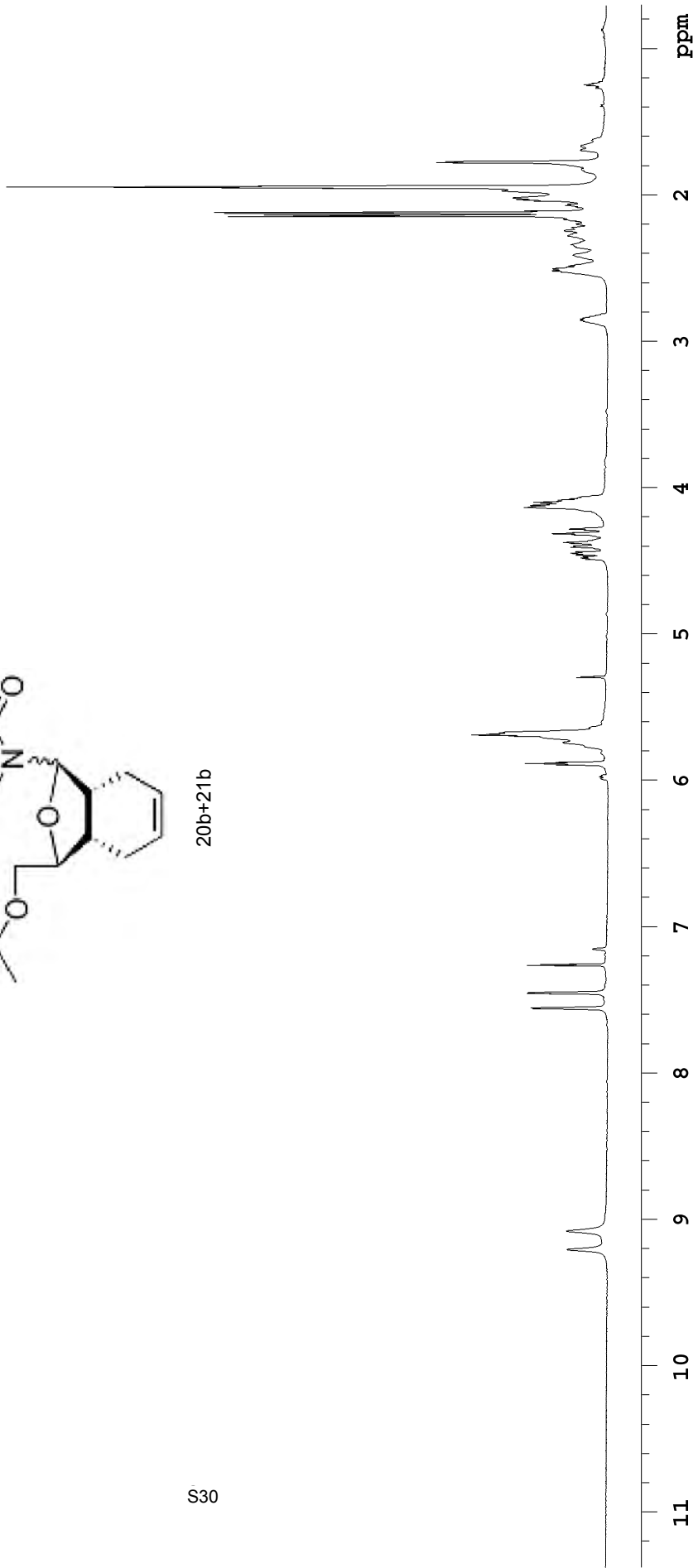


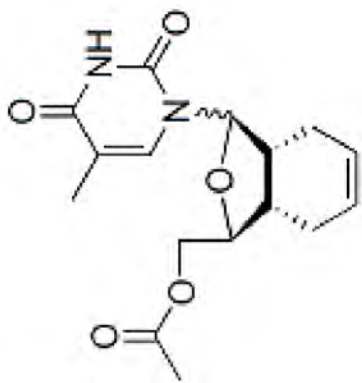
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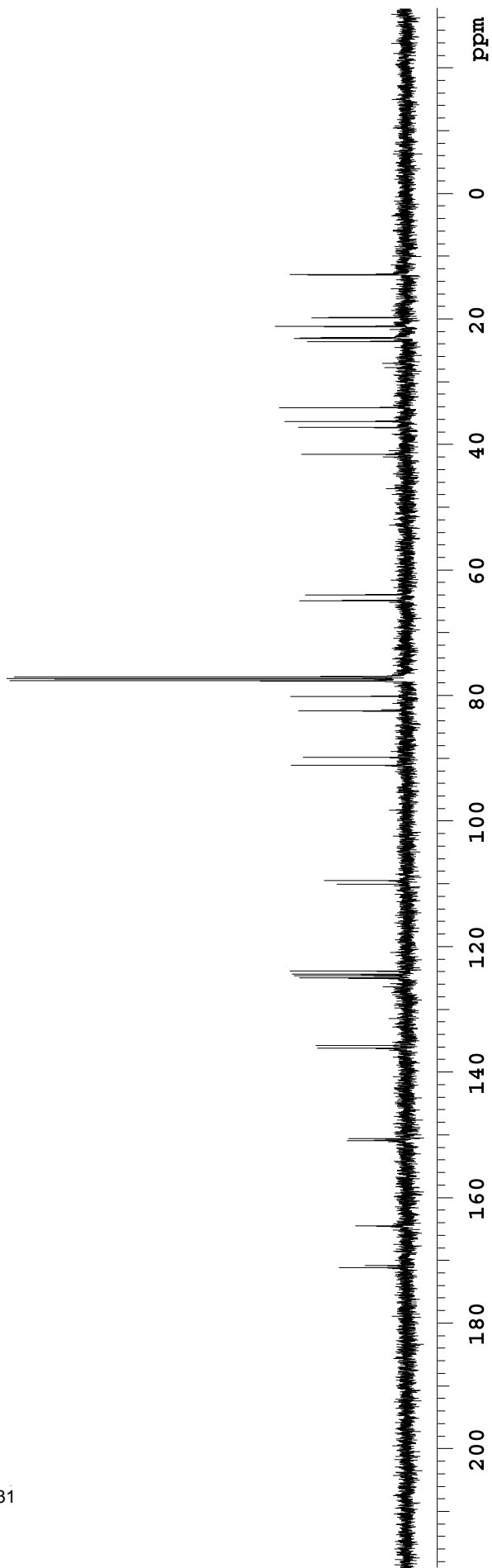


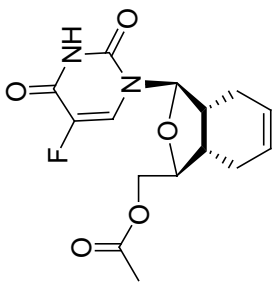
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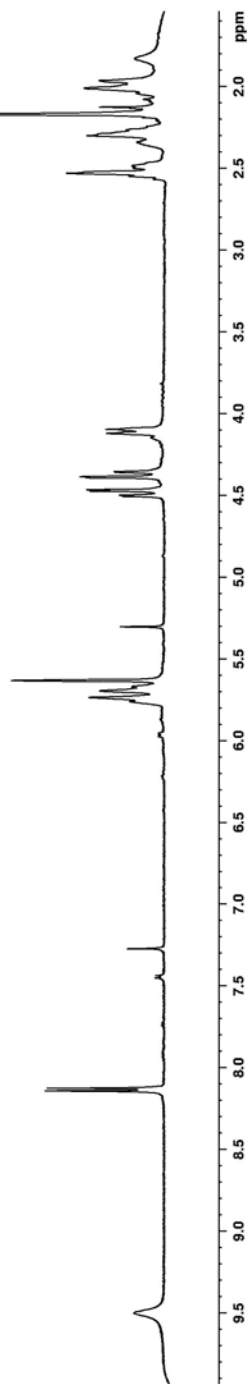


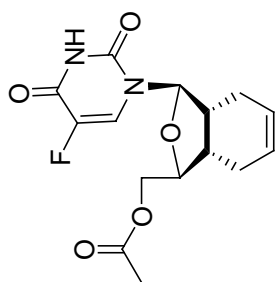
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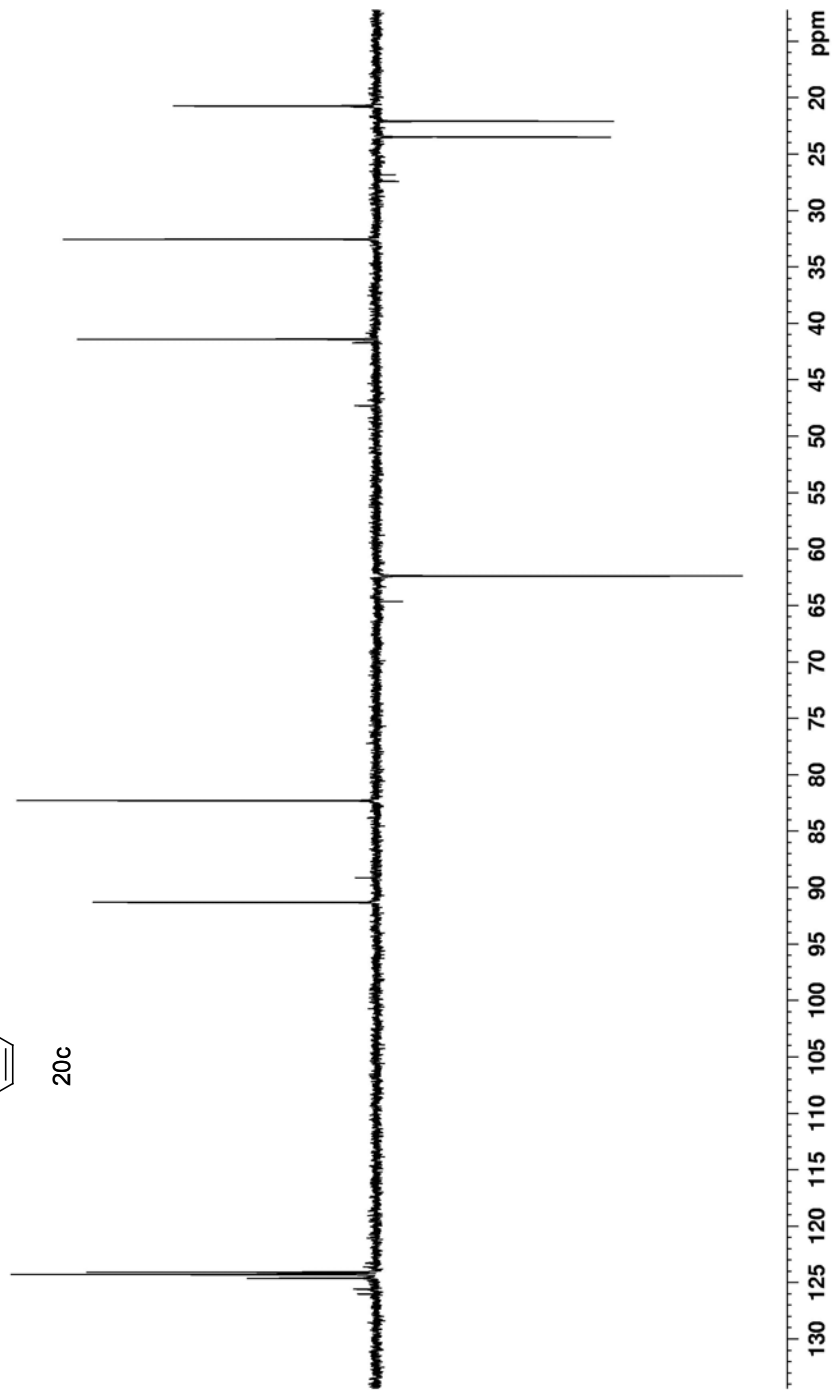


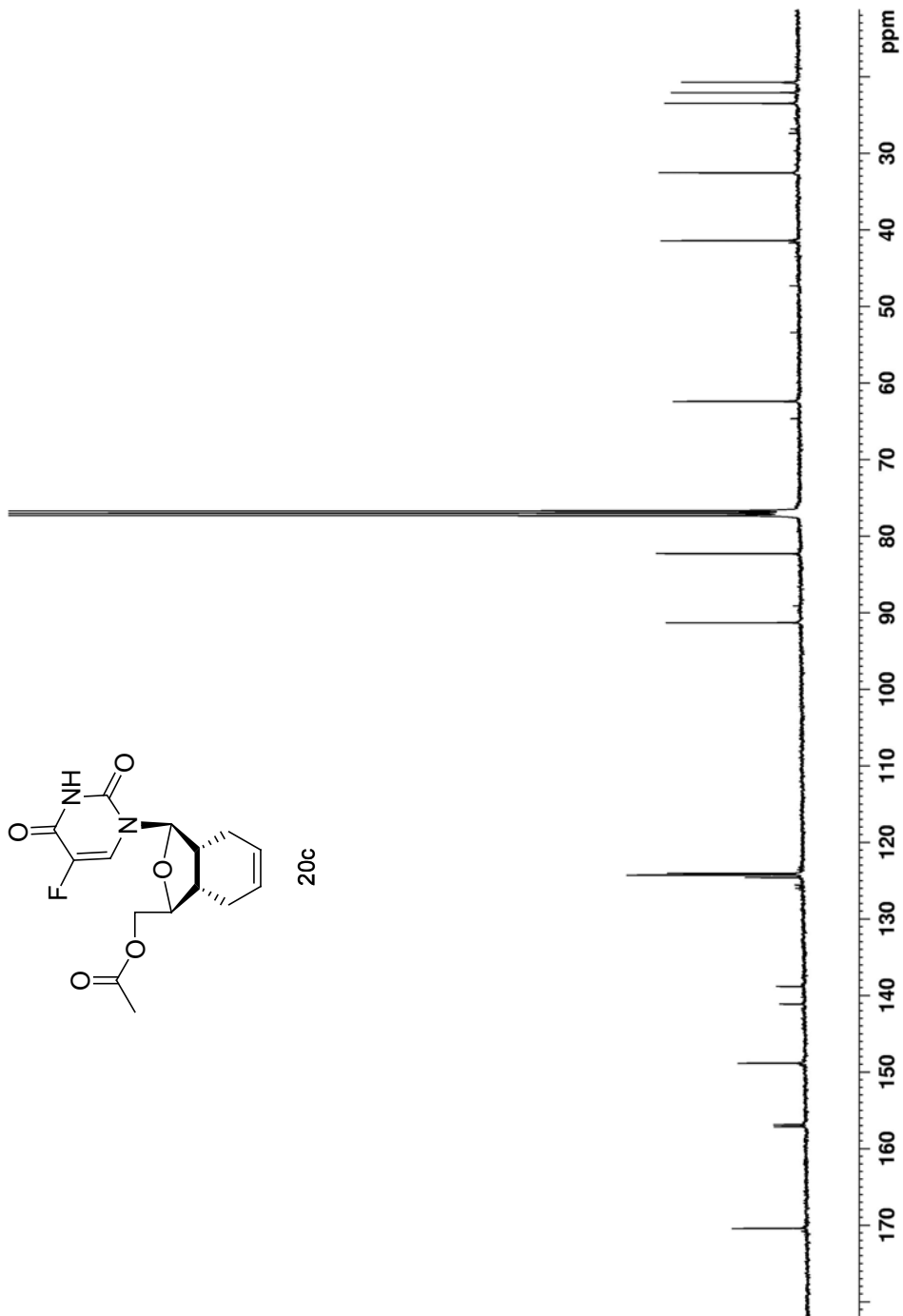
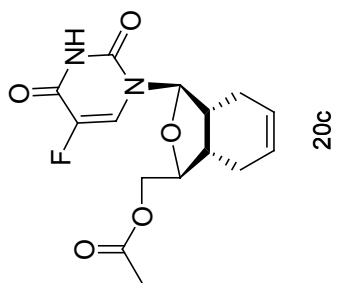
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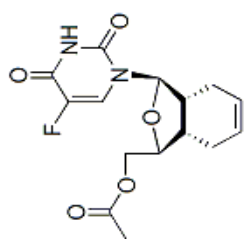




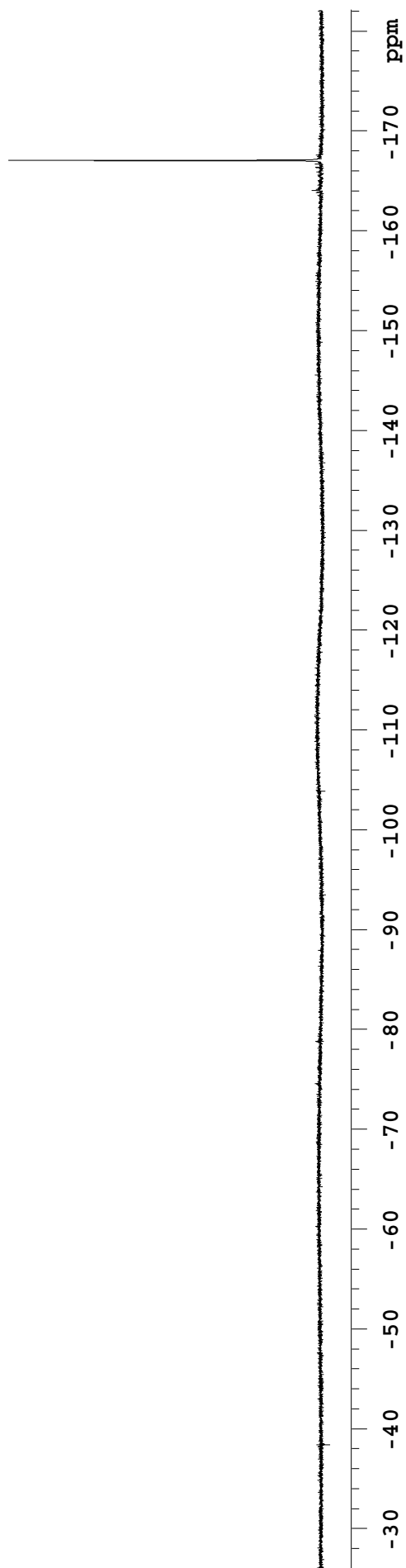
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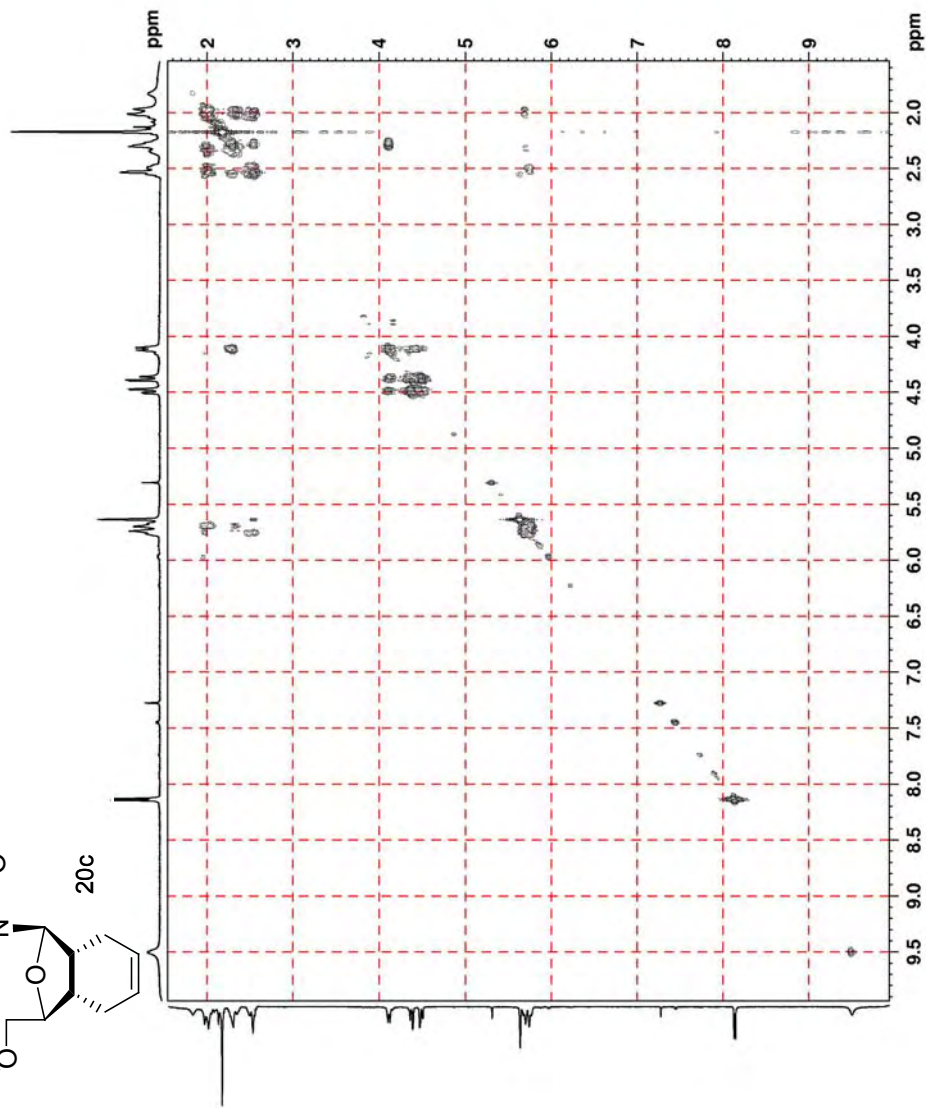
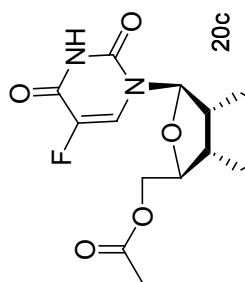




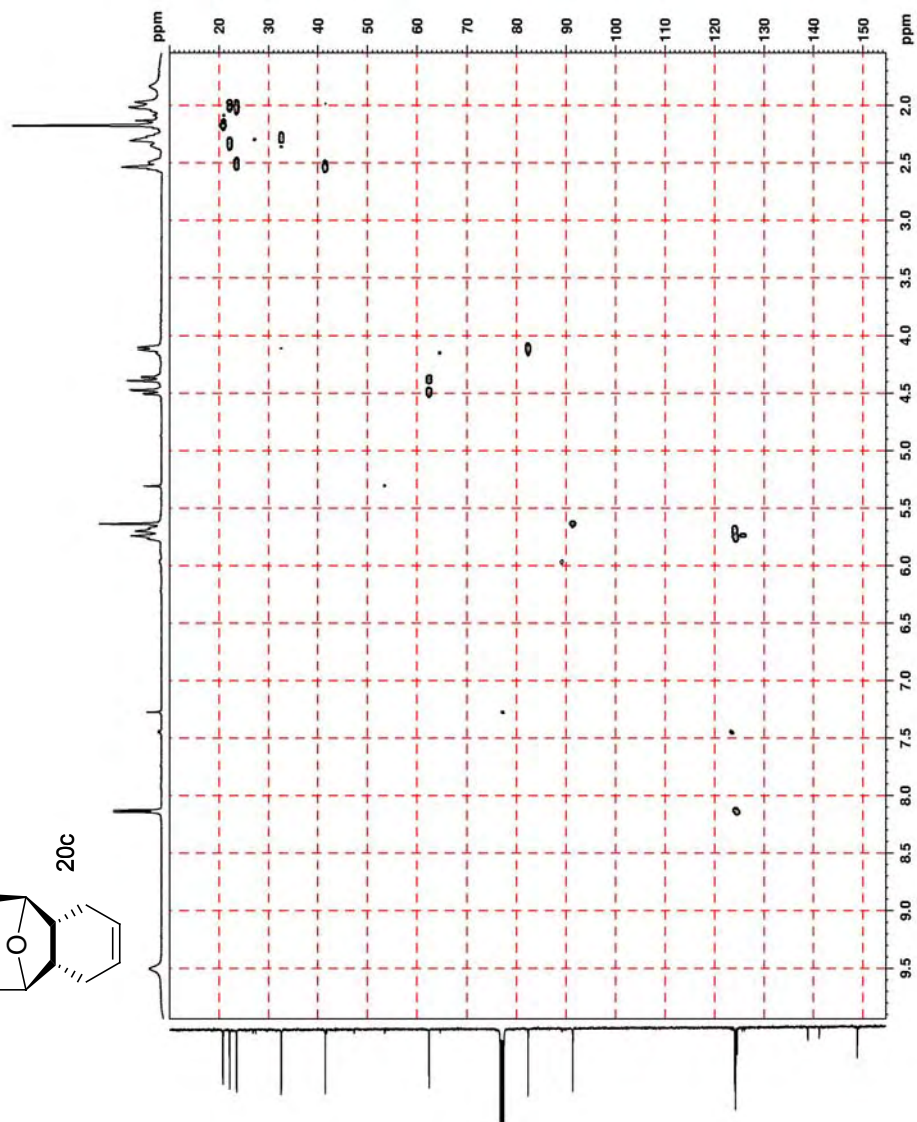
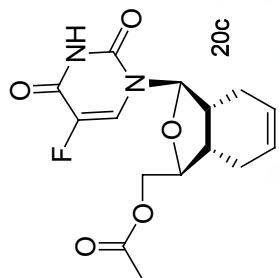
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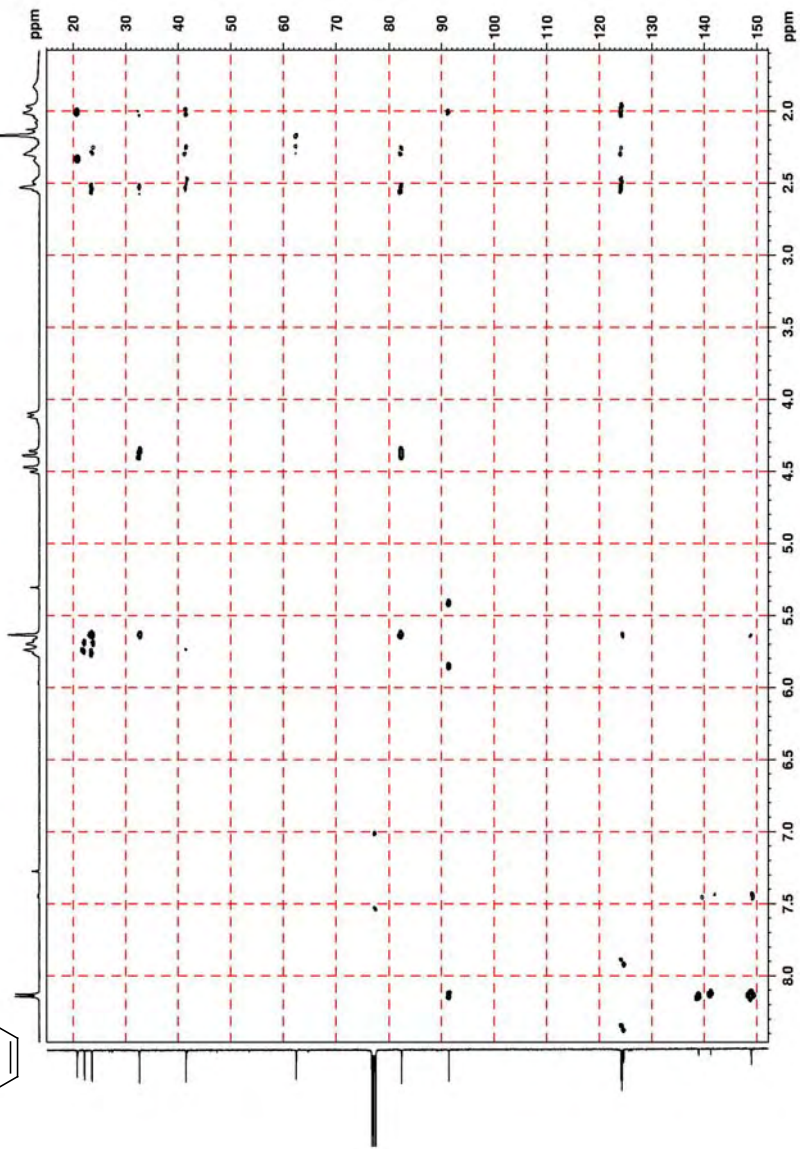
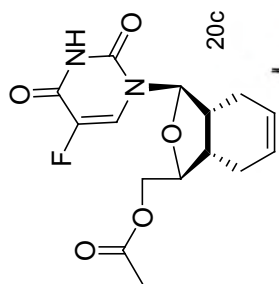
COSY



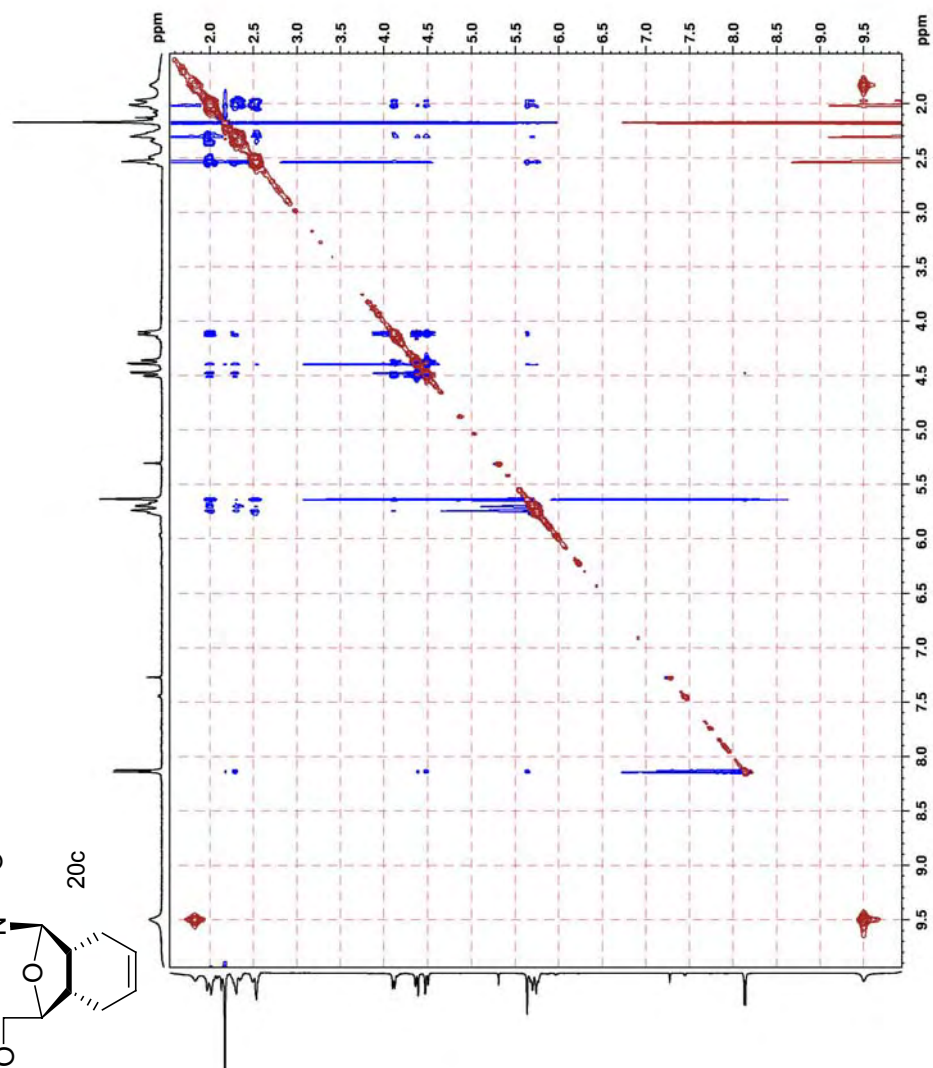
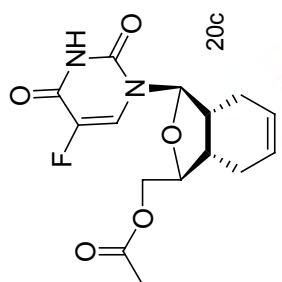
HSQC

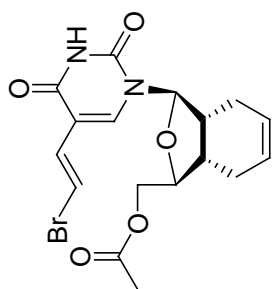


HMBC

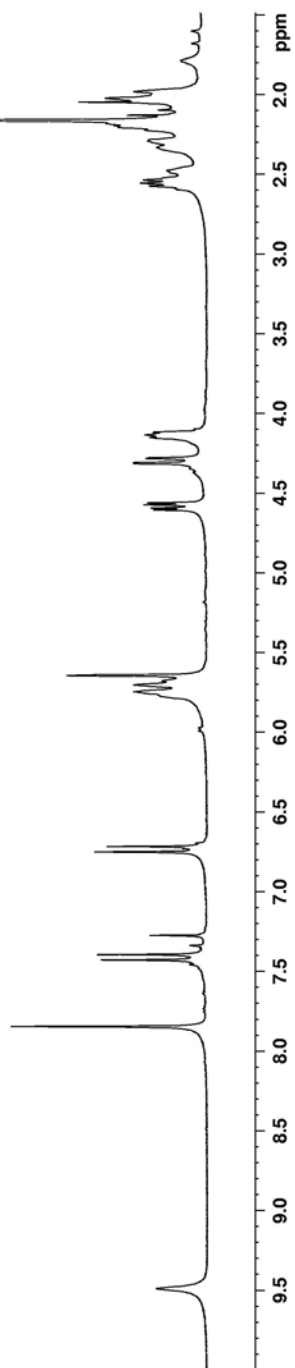


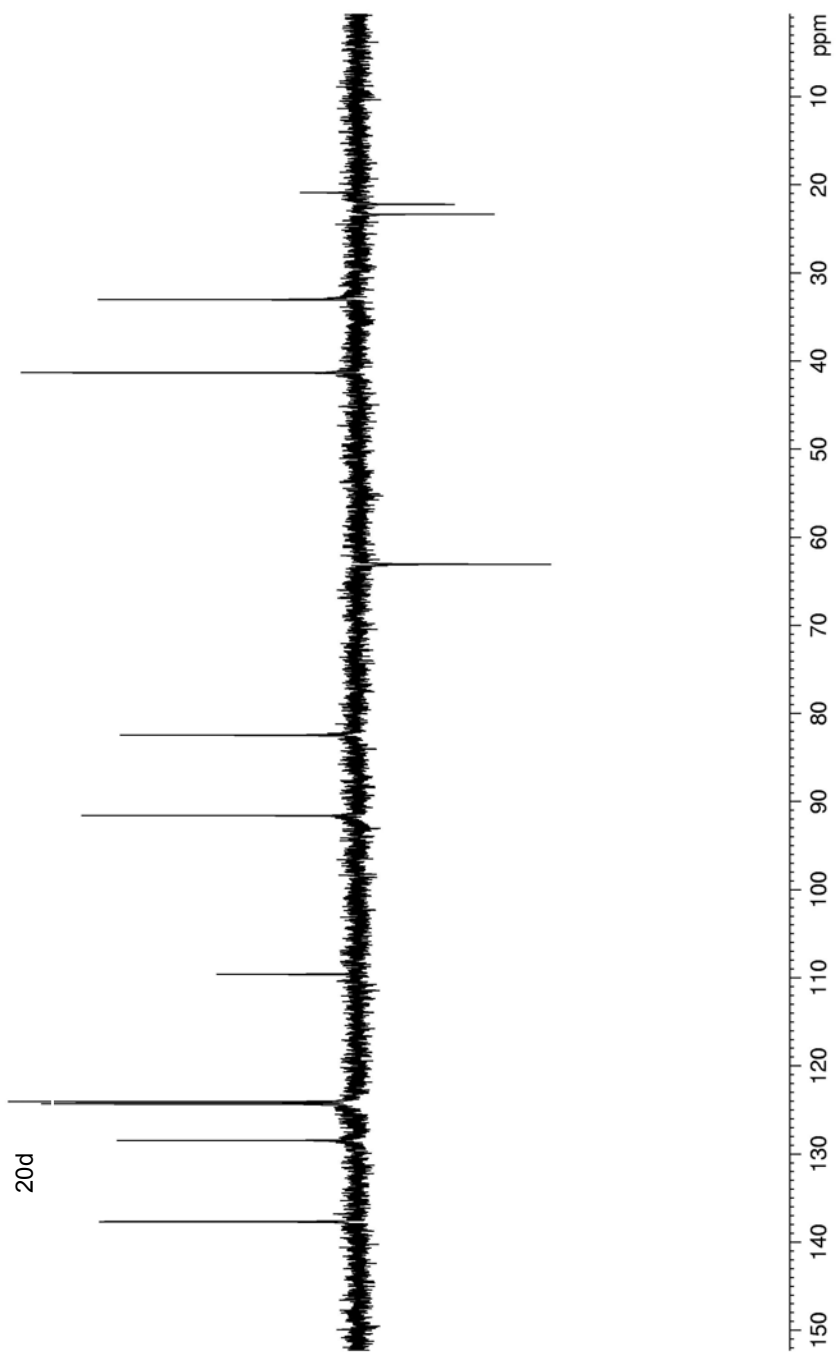
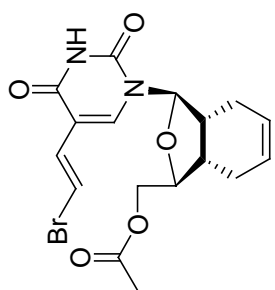
NOESY

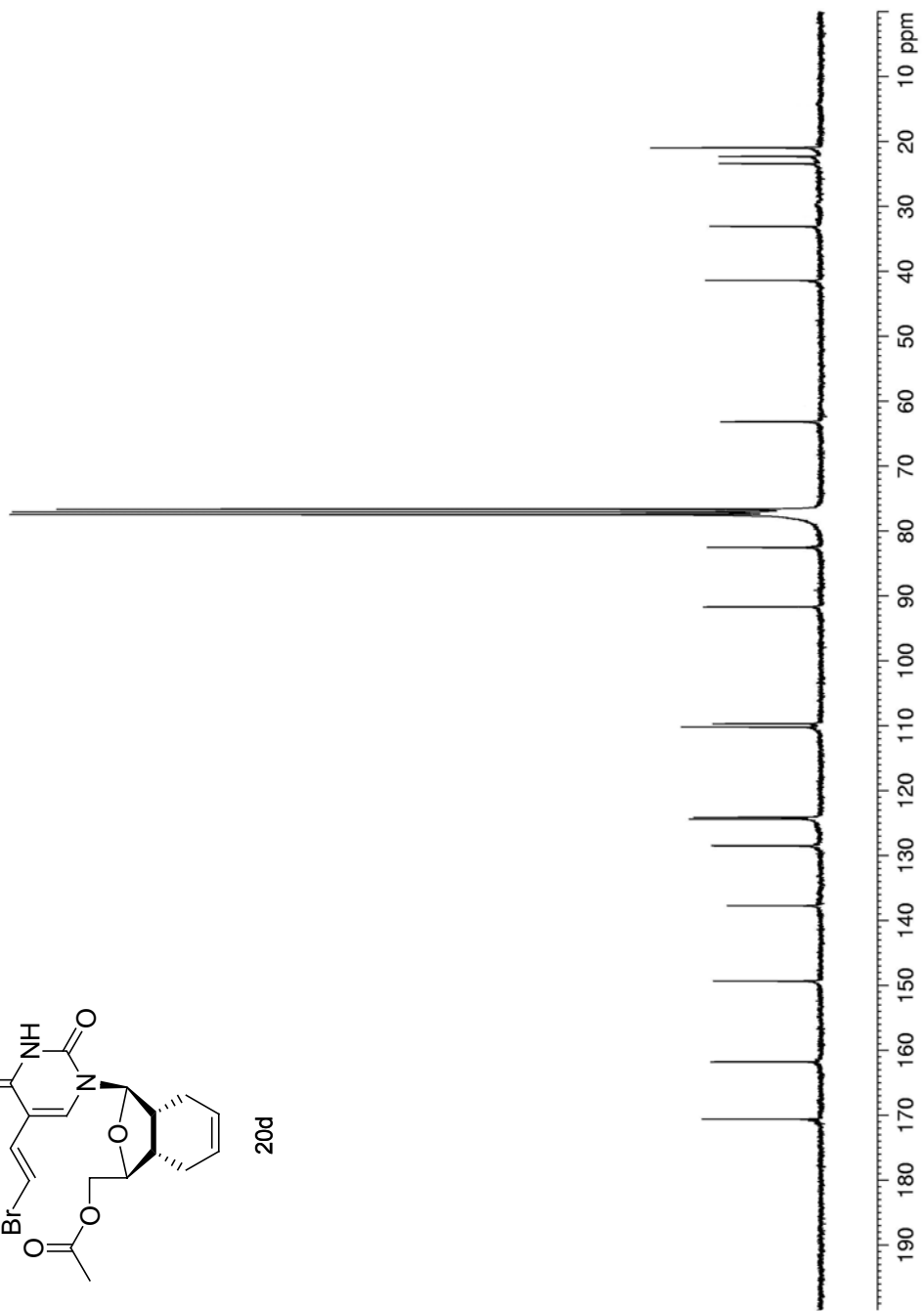
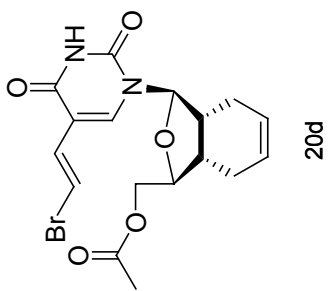


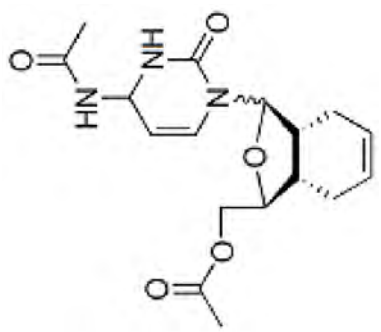


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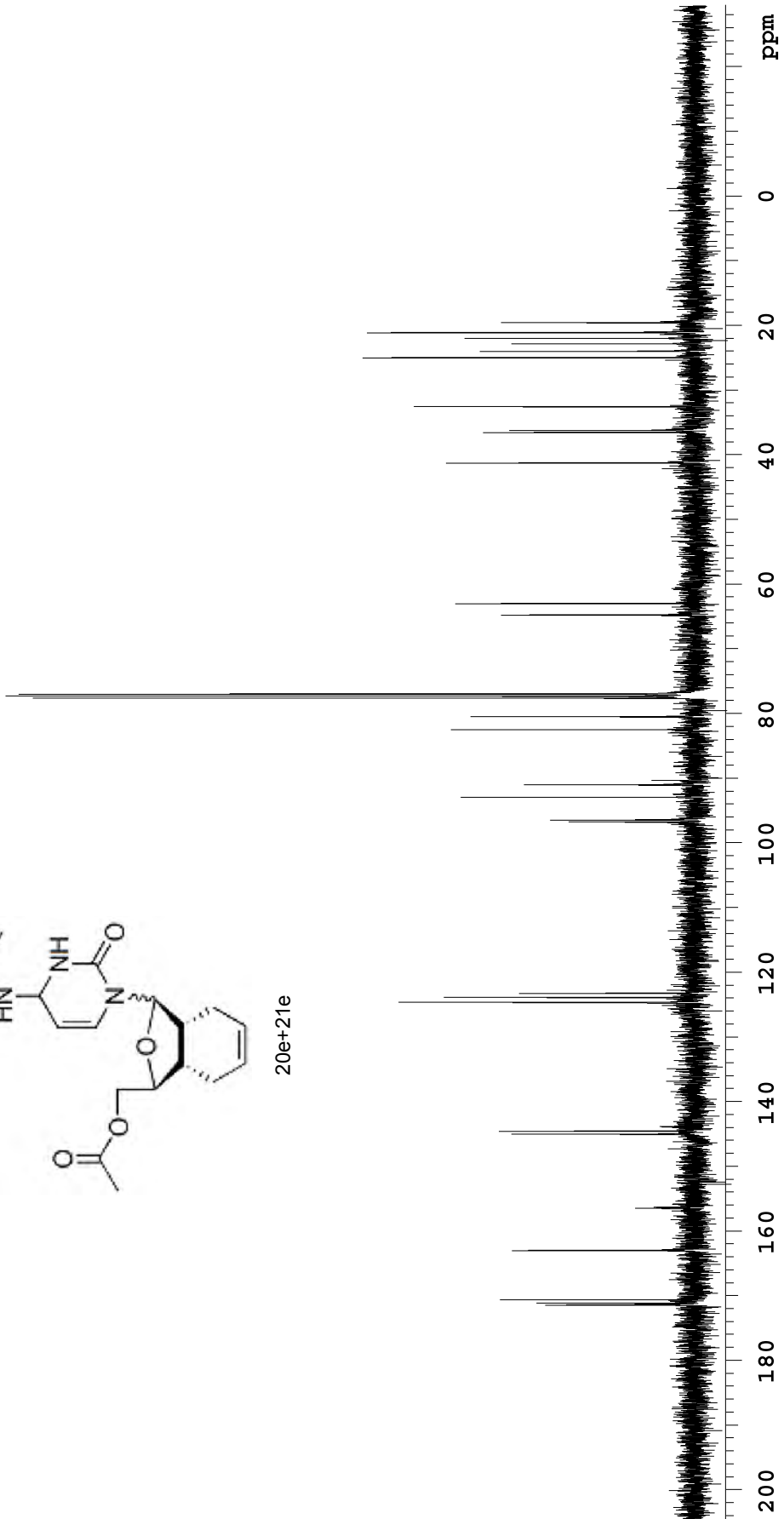


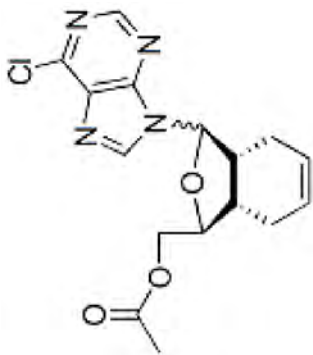




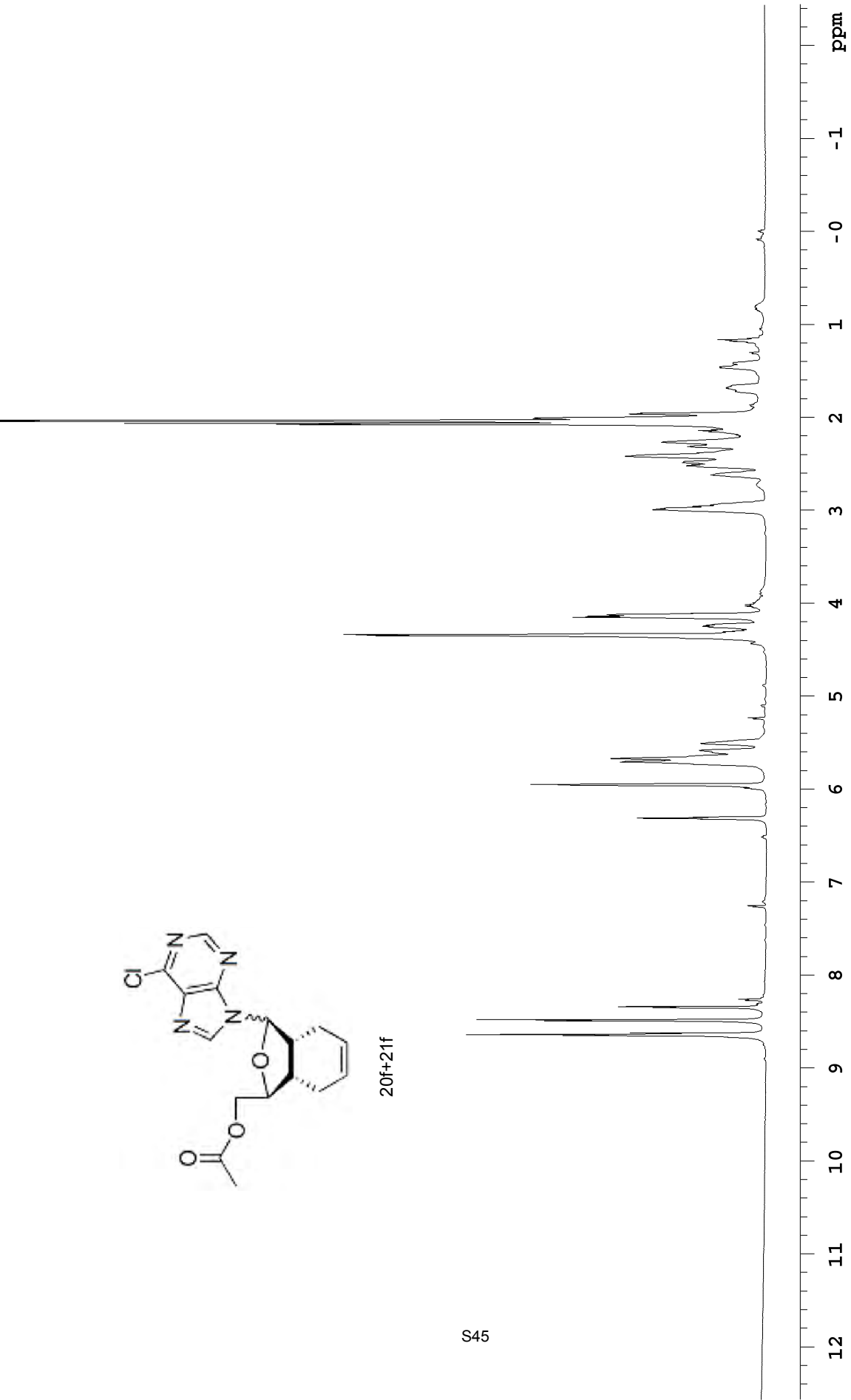


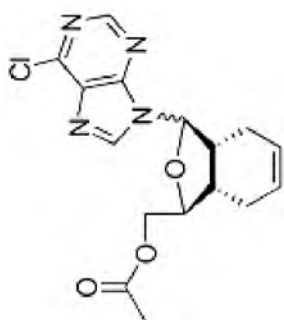
20e+21e



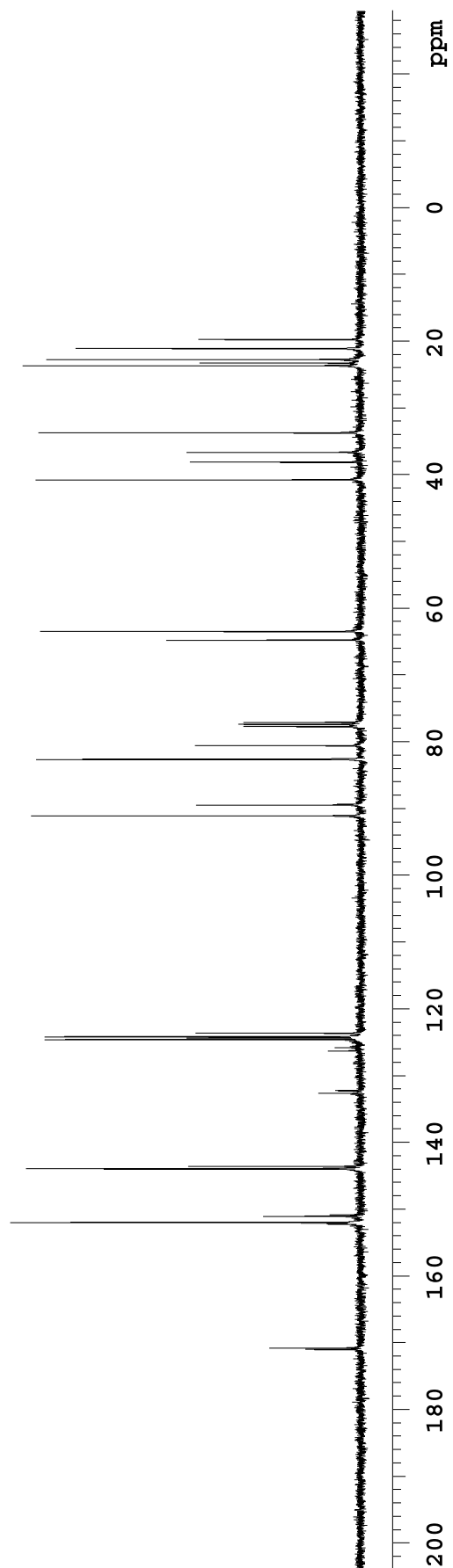


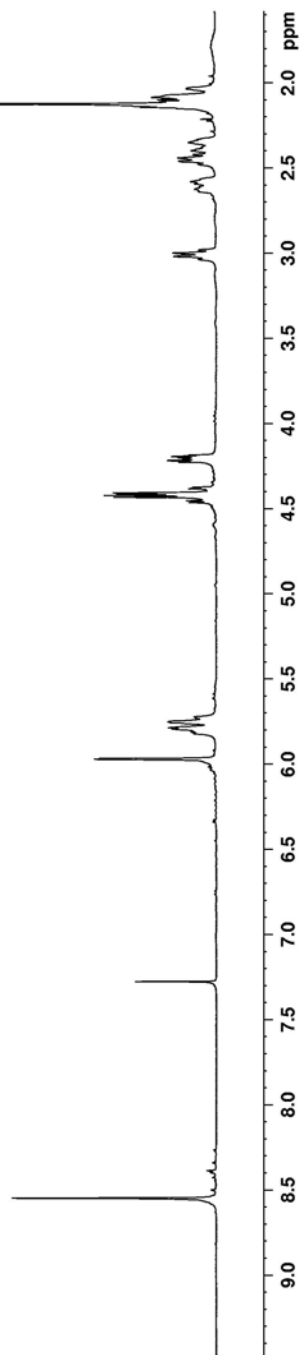
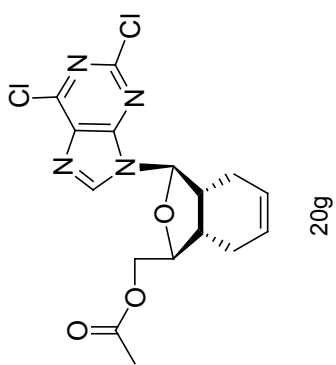
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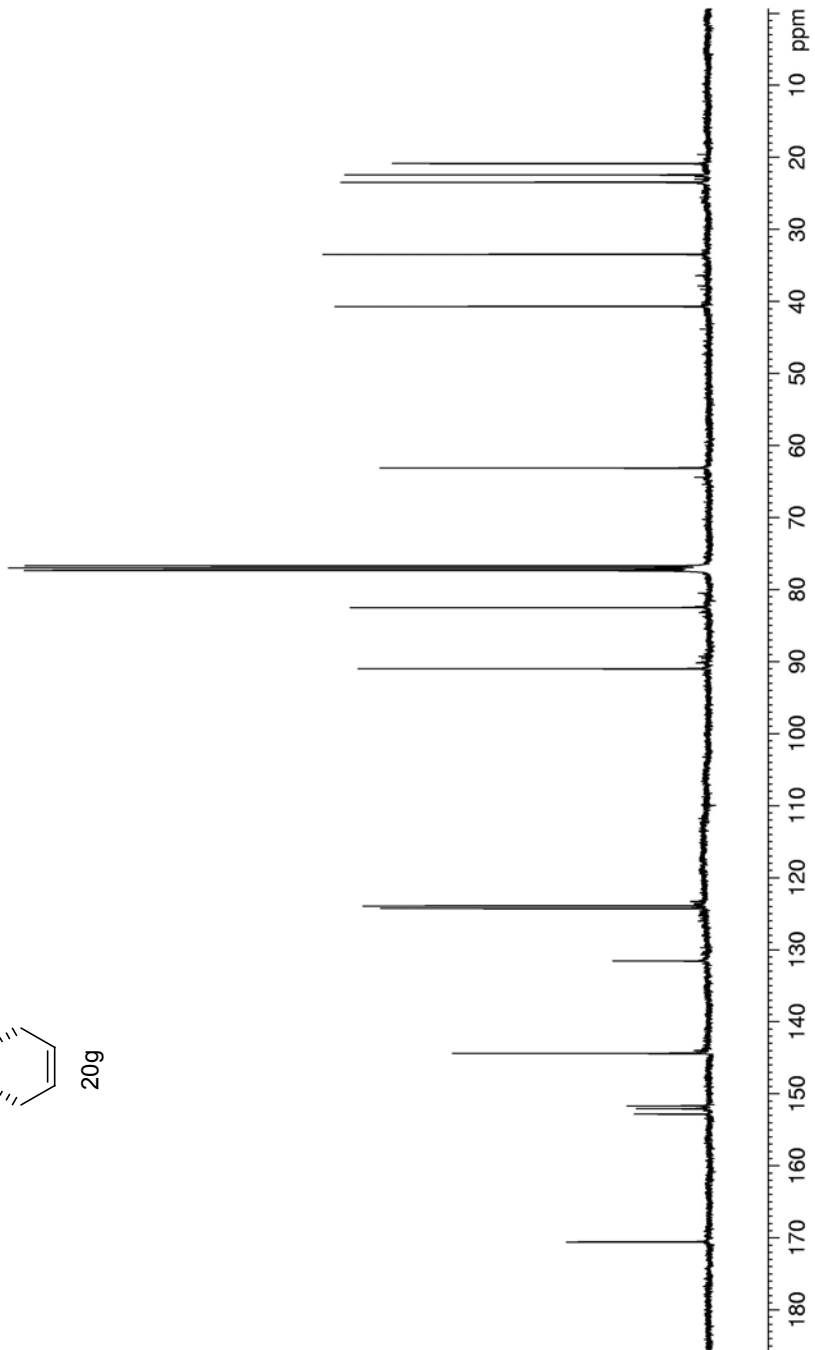
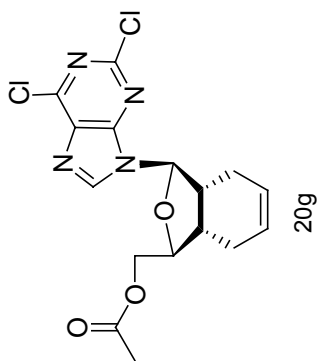


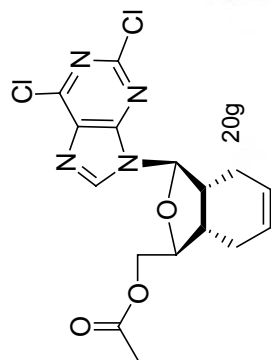


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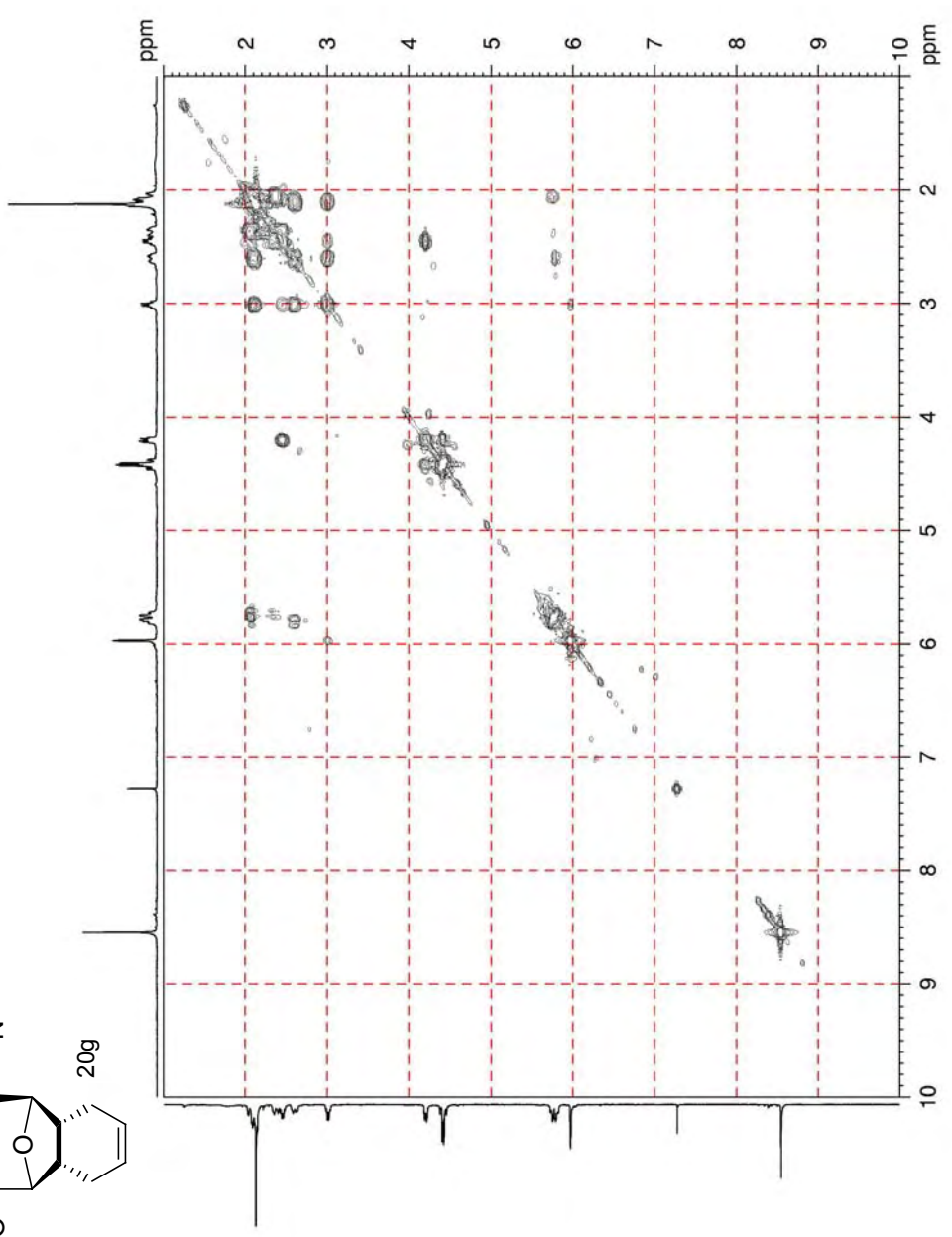




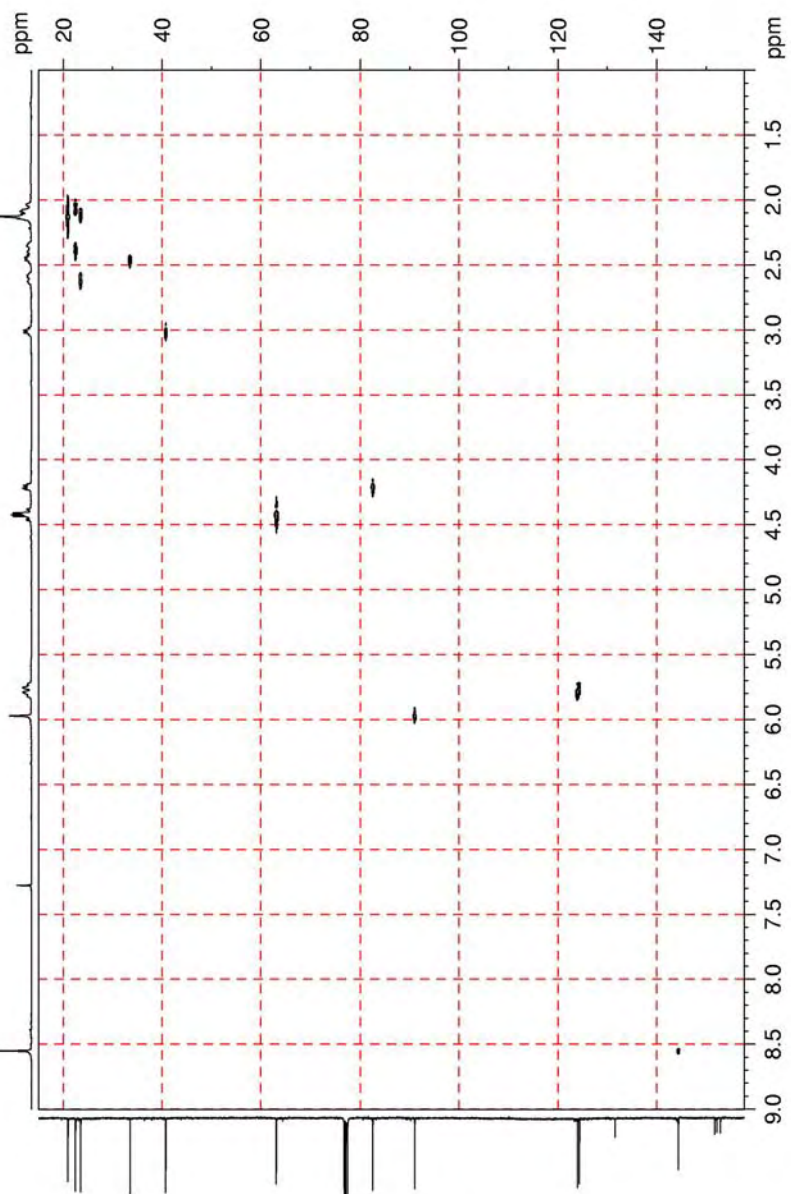
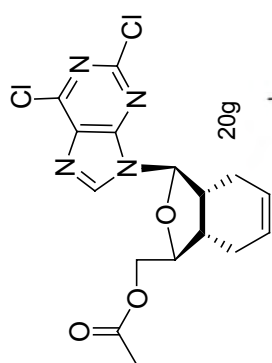




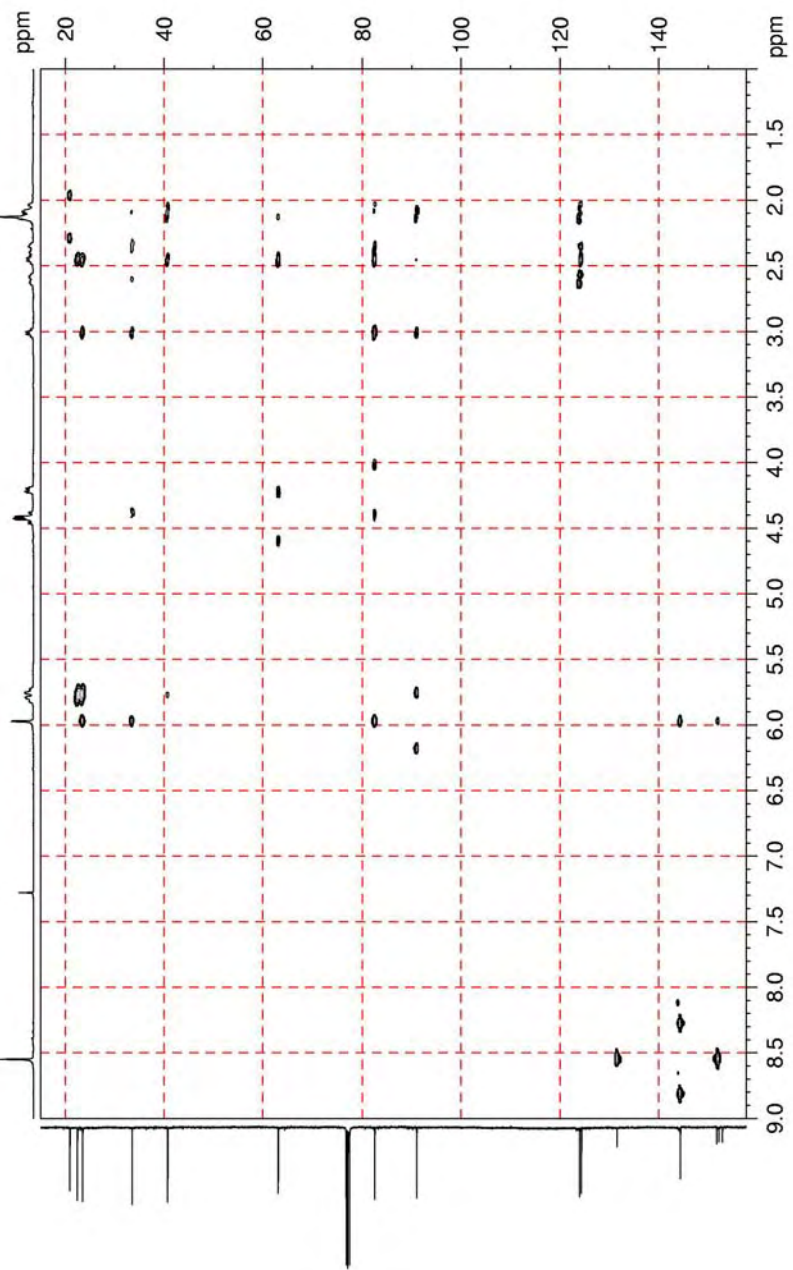
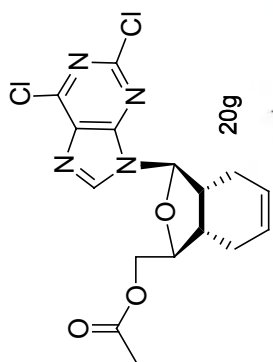
COSY



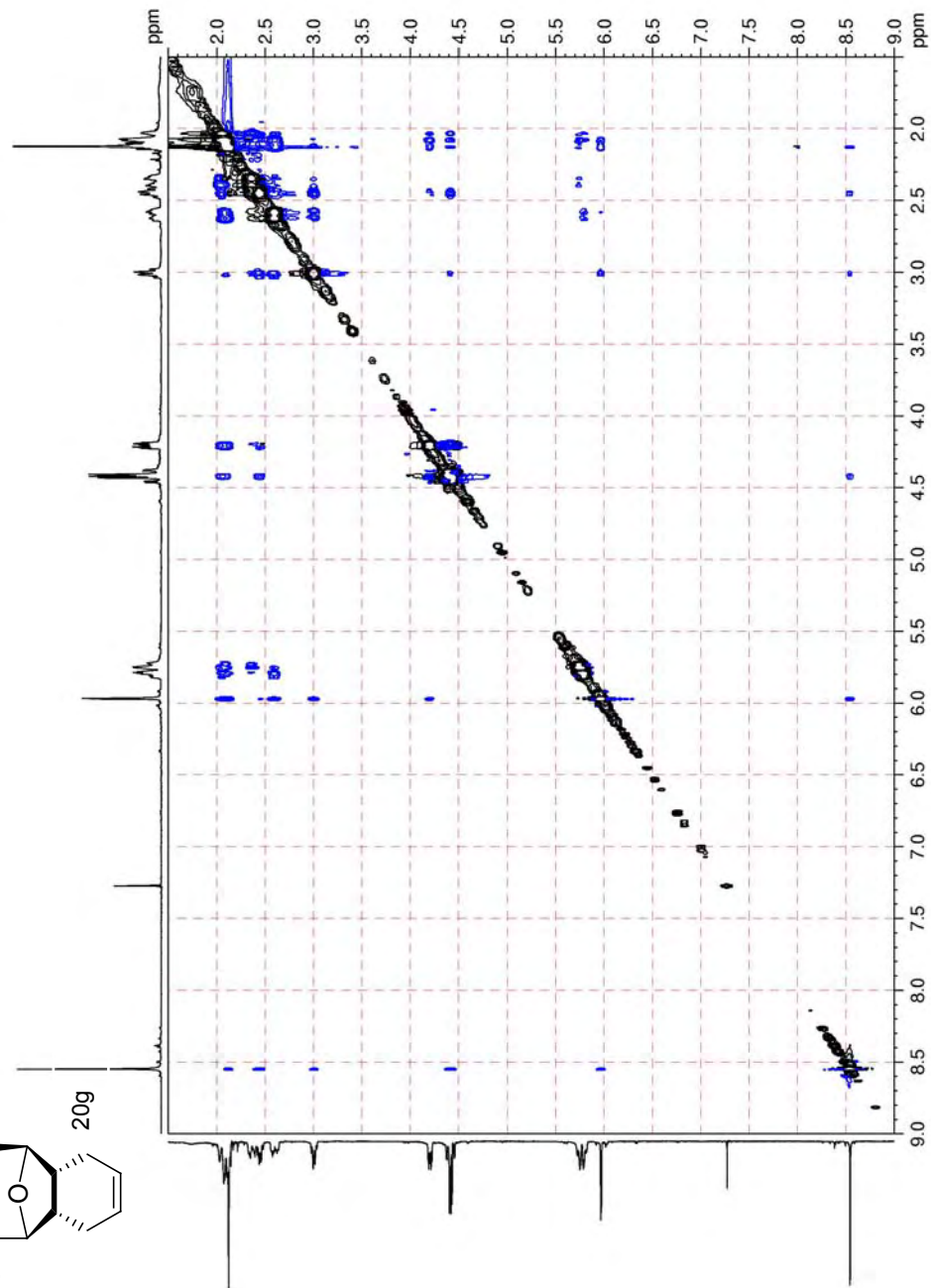
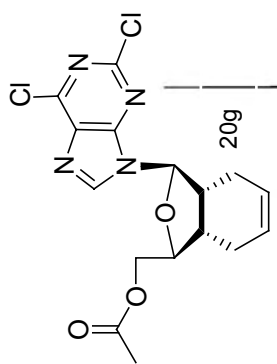
HSQC

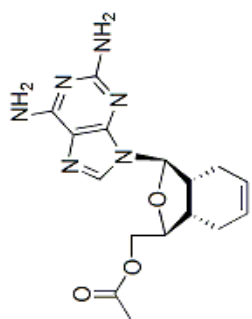


HMBC



NOESY





20m

ppm

2

3

4

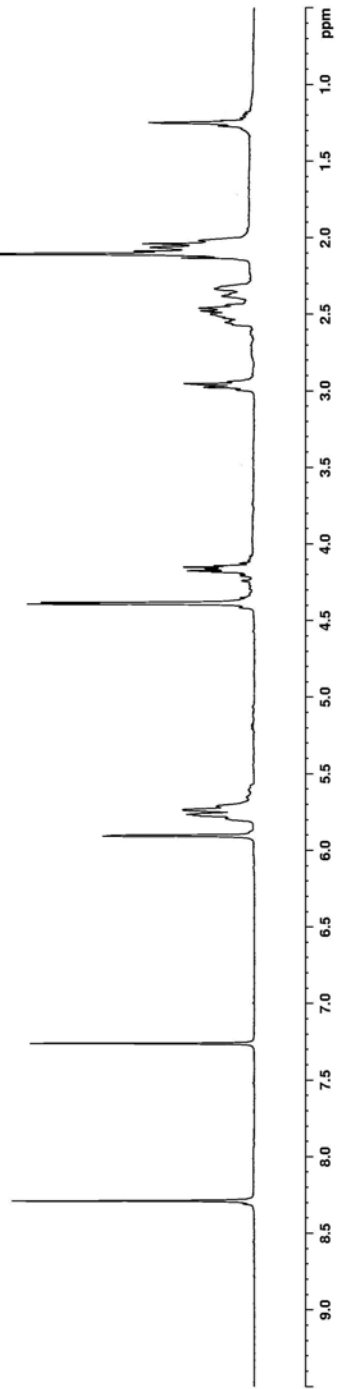
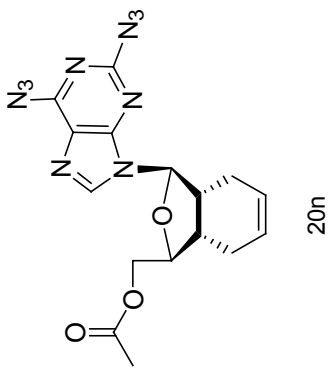
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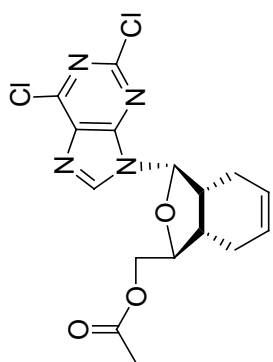
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7

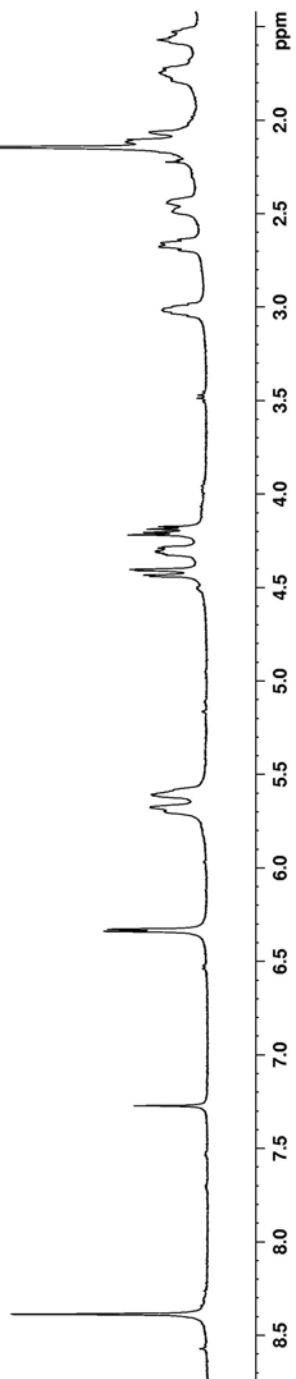
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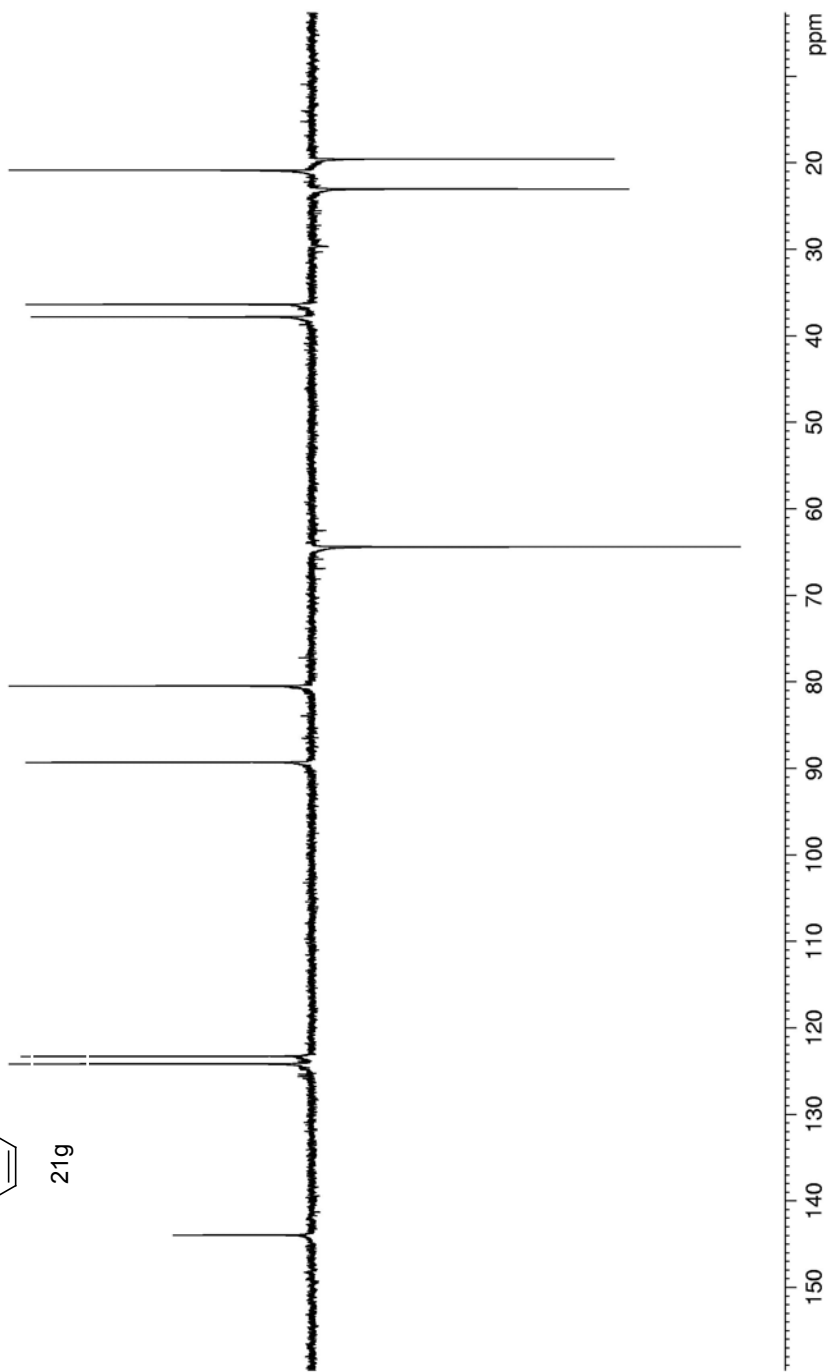
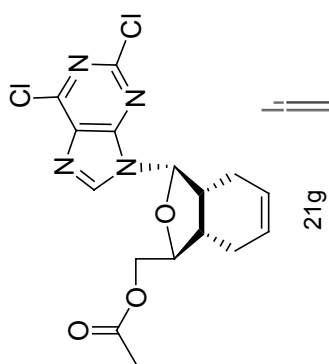
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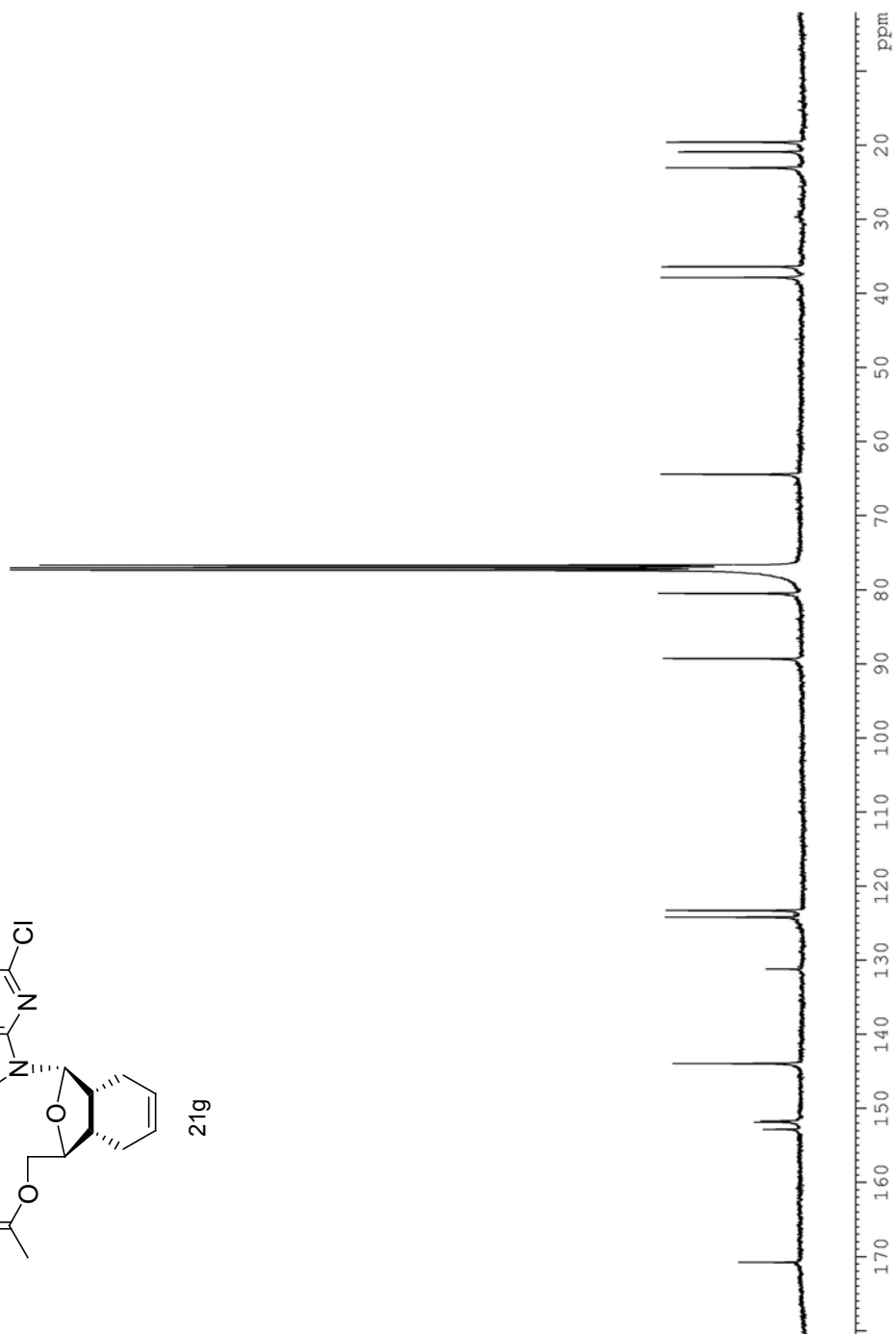
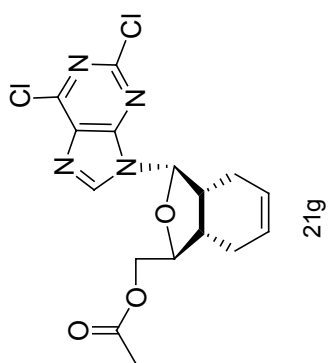




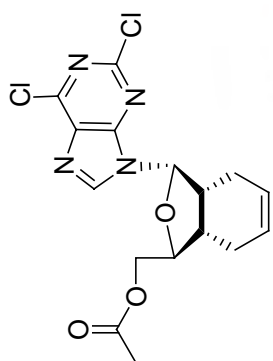
21g



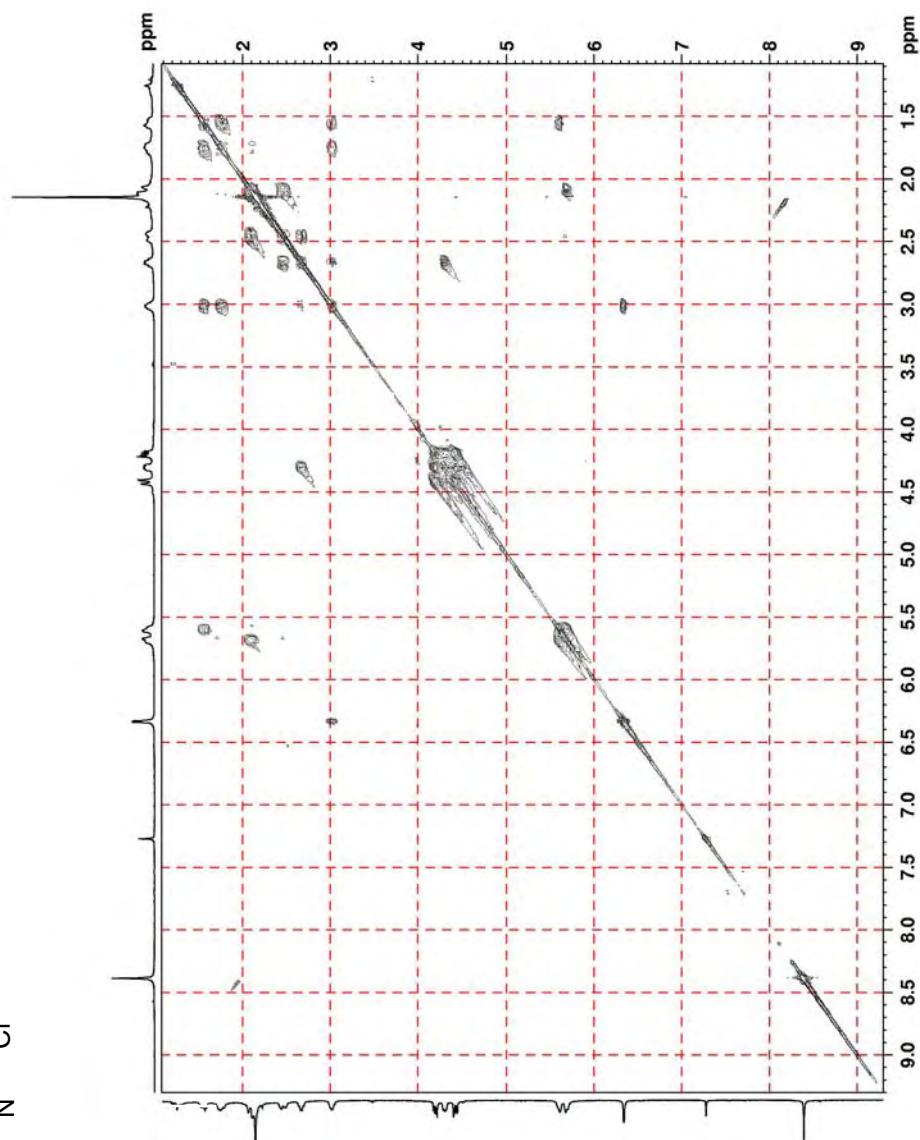


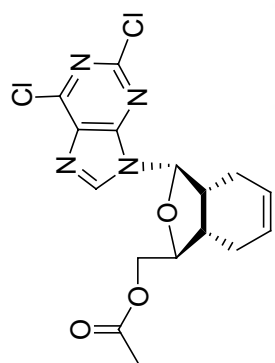


COSY



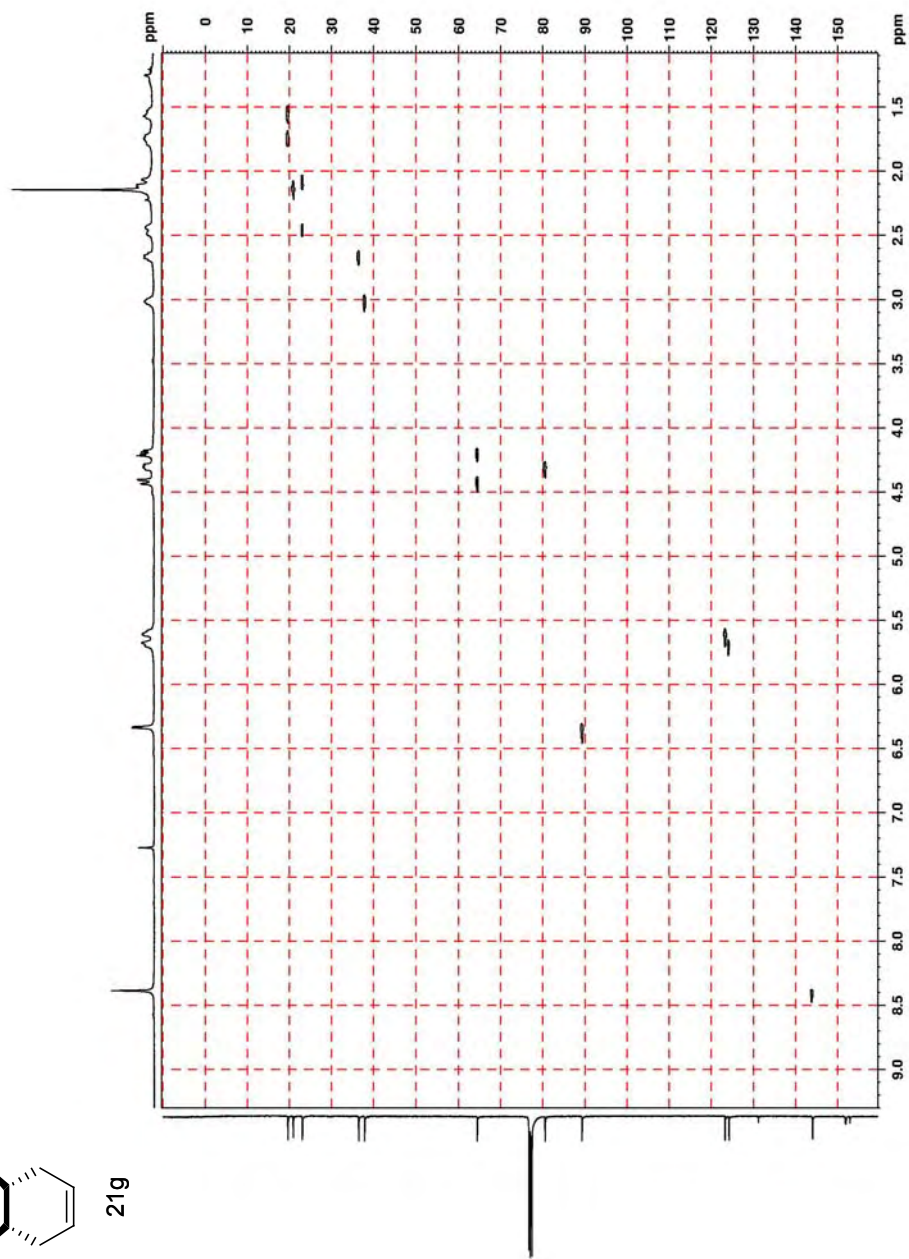
21g



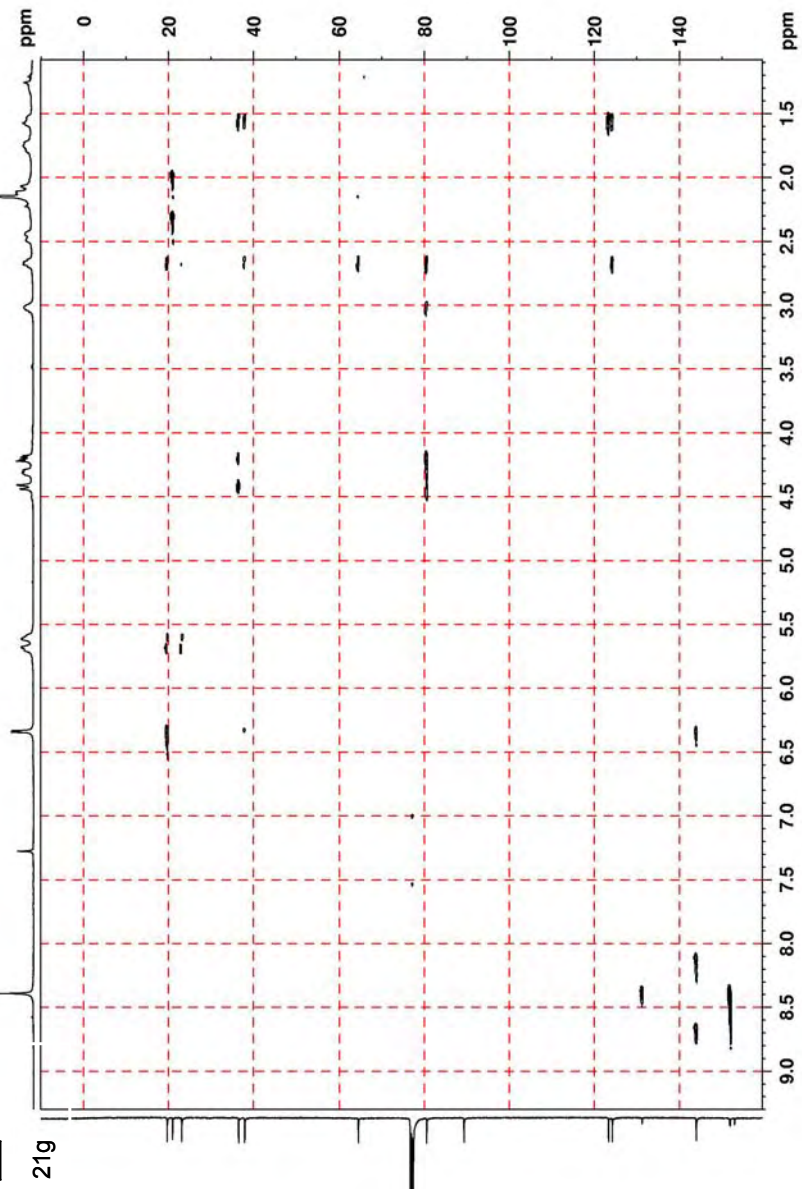
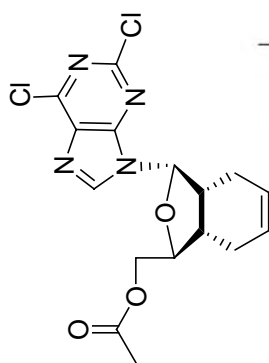


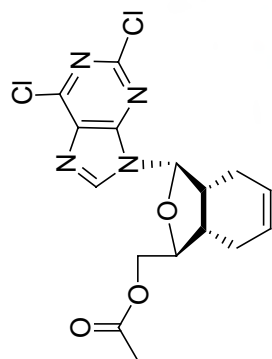
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HSQC



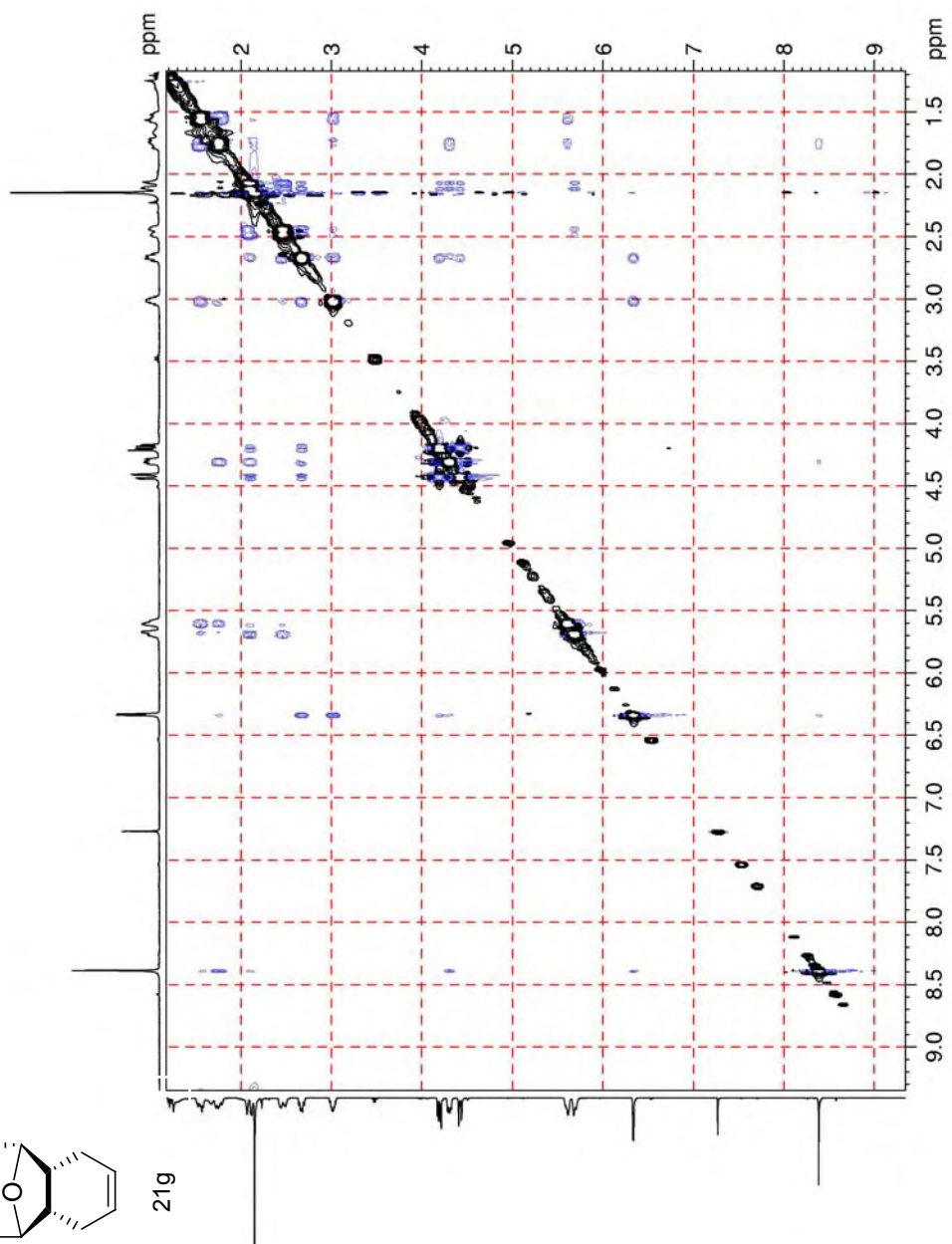
HMBC

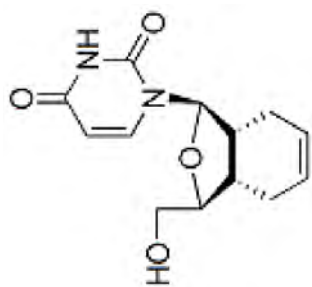




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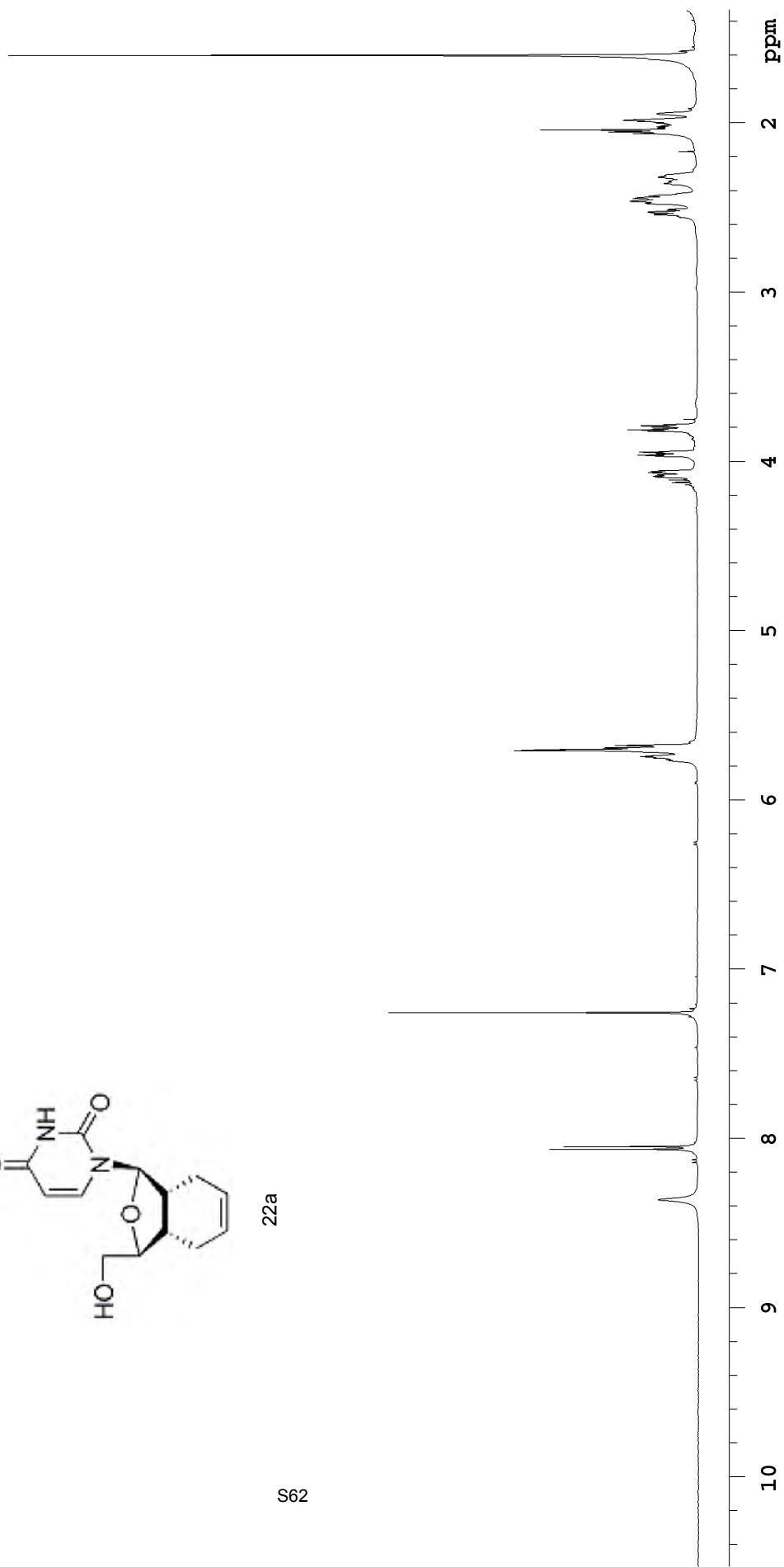
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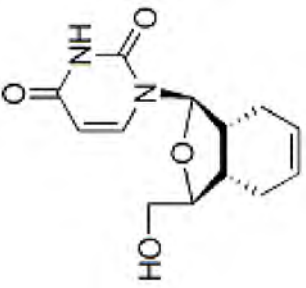




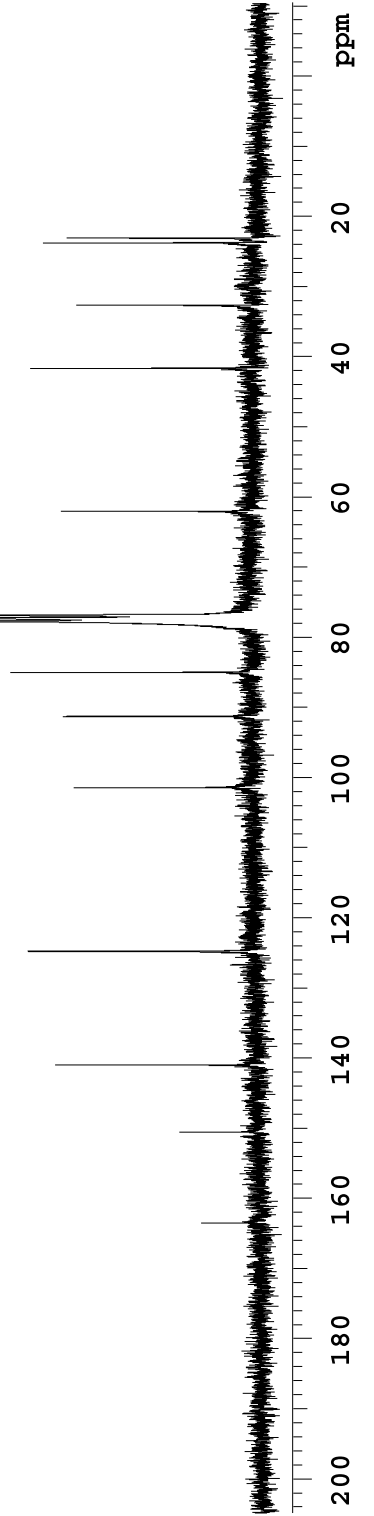
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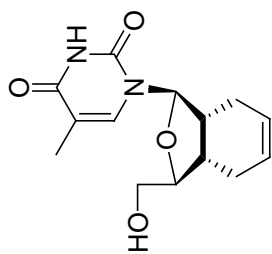
S62



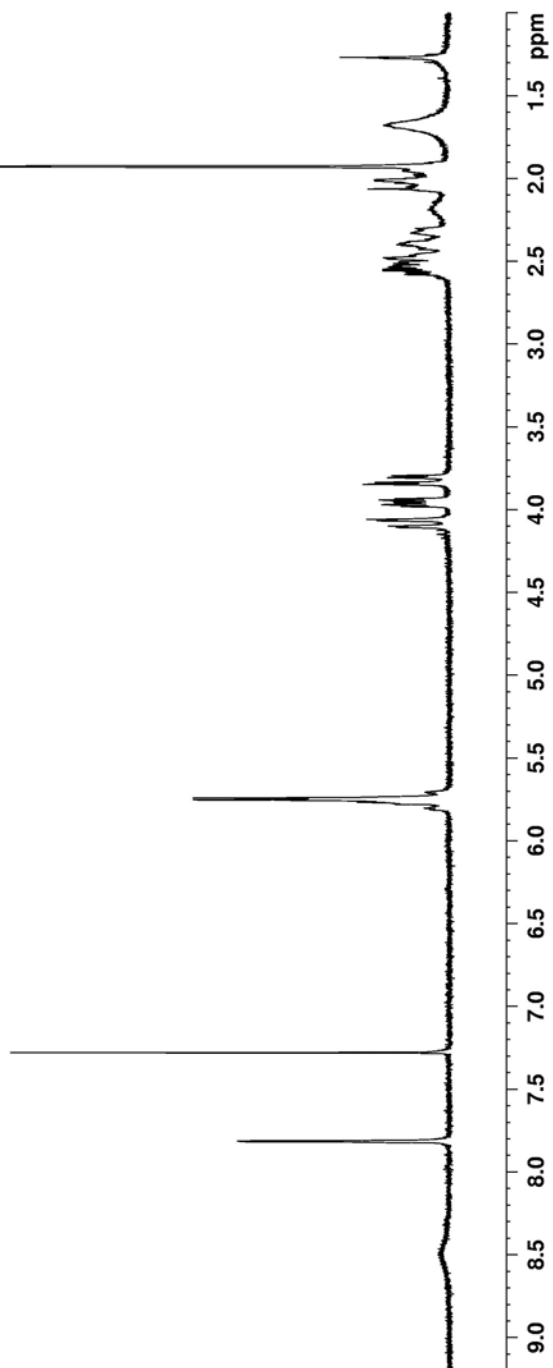


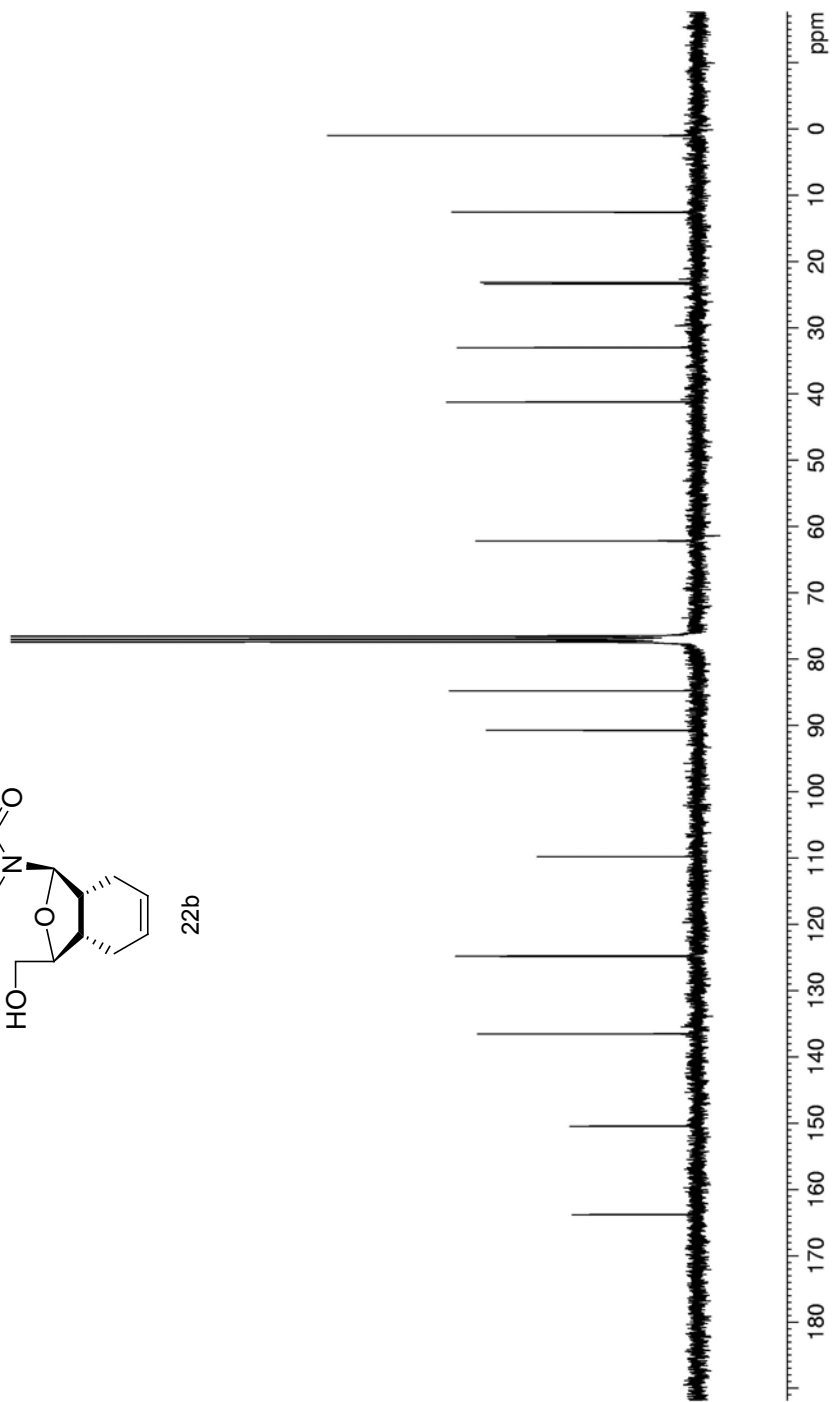
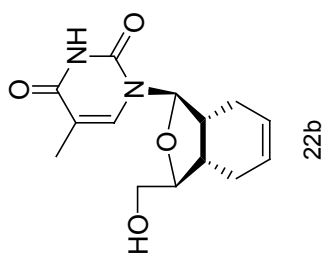
22a



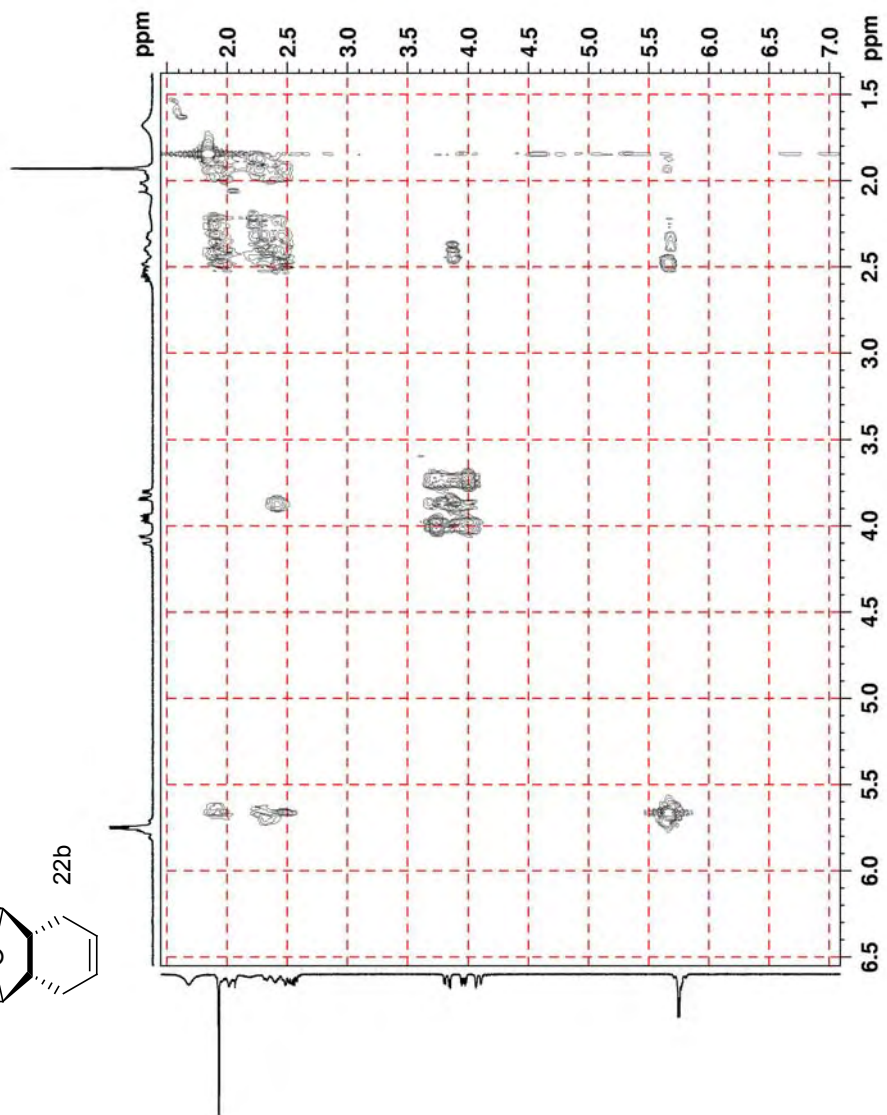
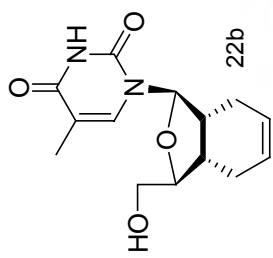


22b

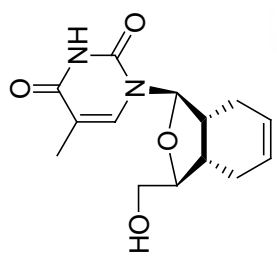




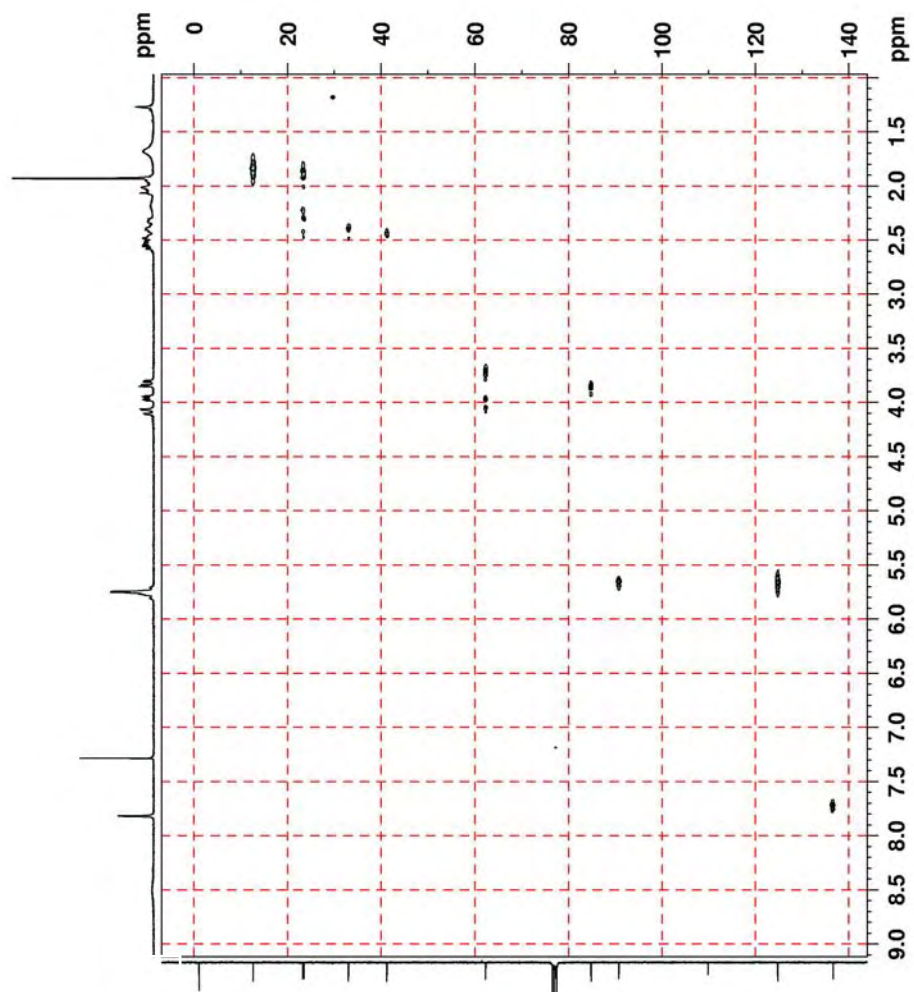
COSY



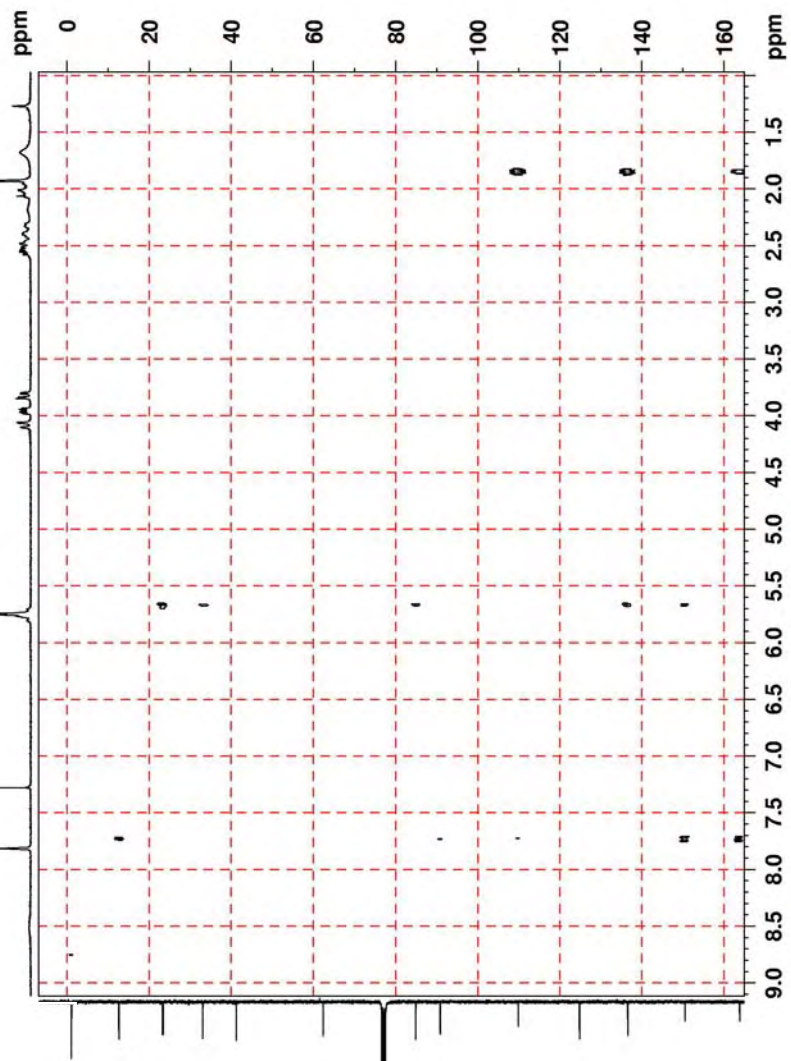
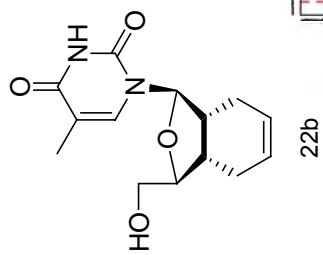
HSQC

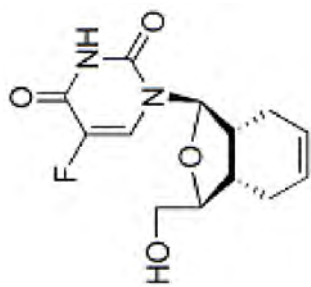


22b

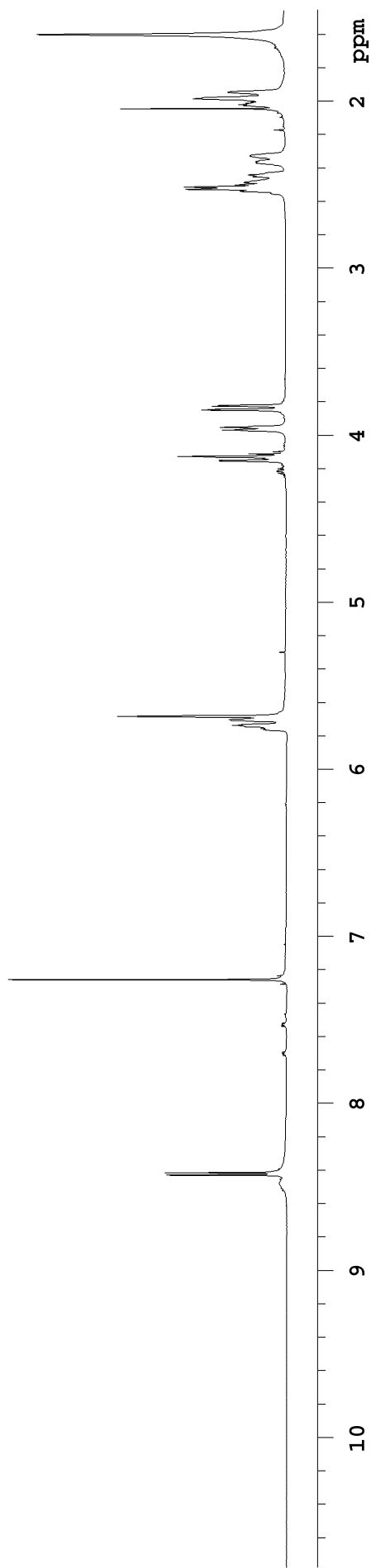


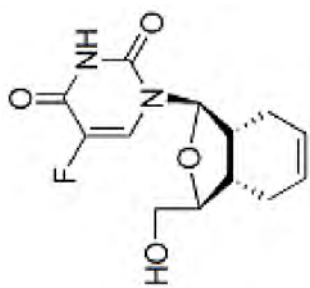
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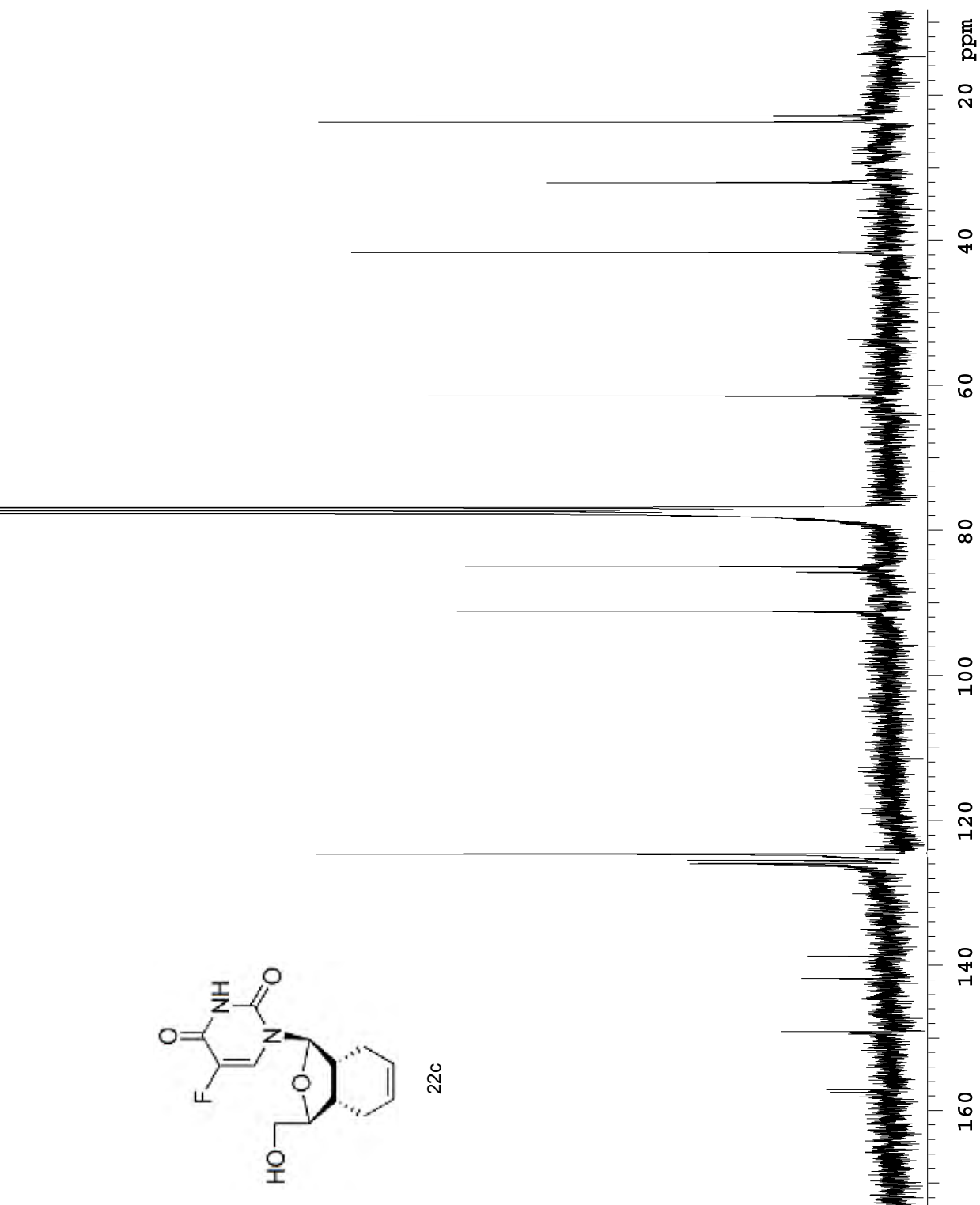


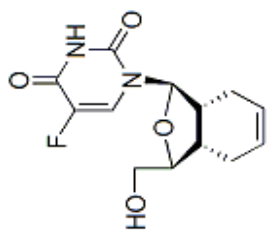
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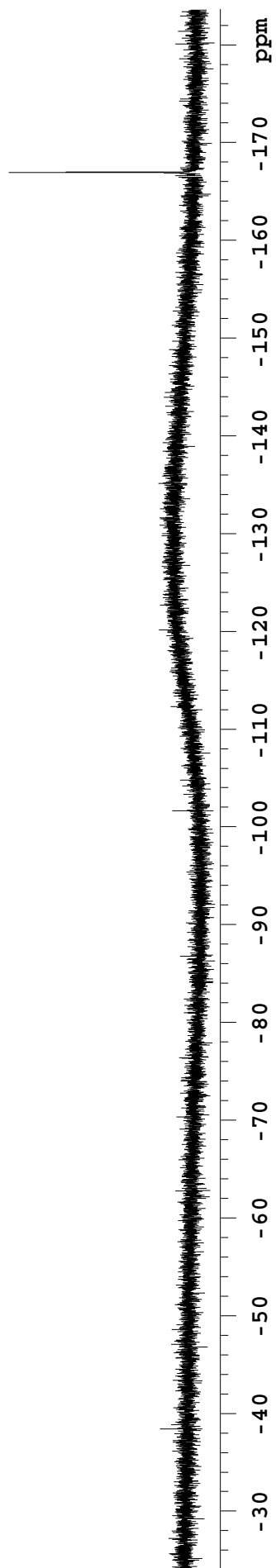


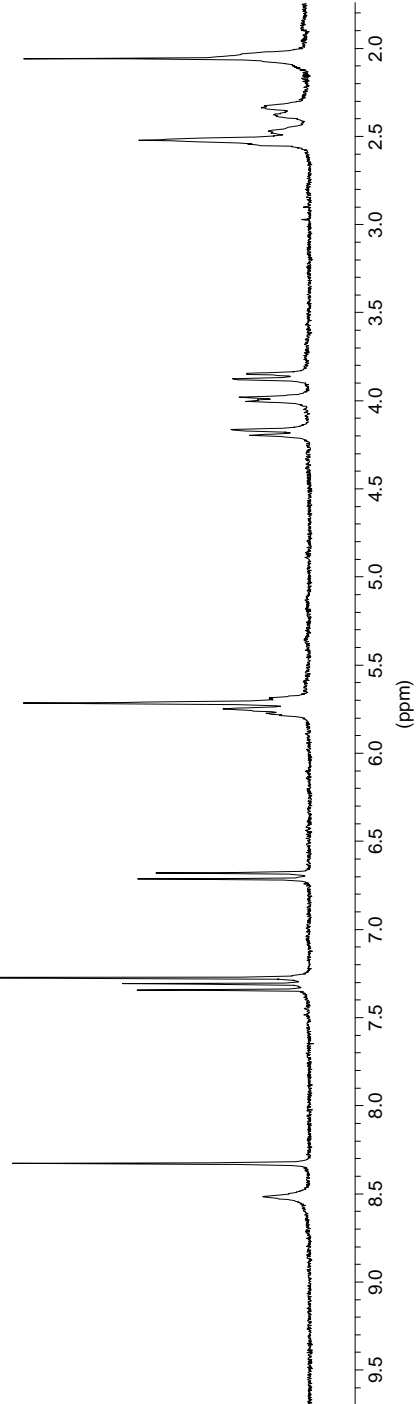
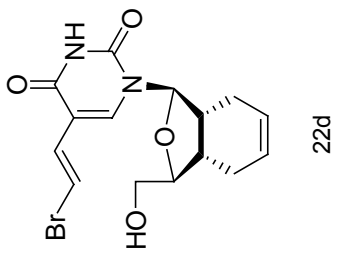
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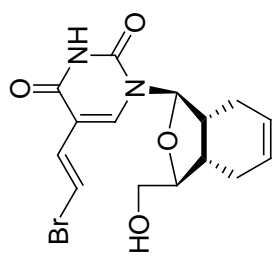




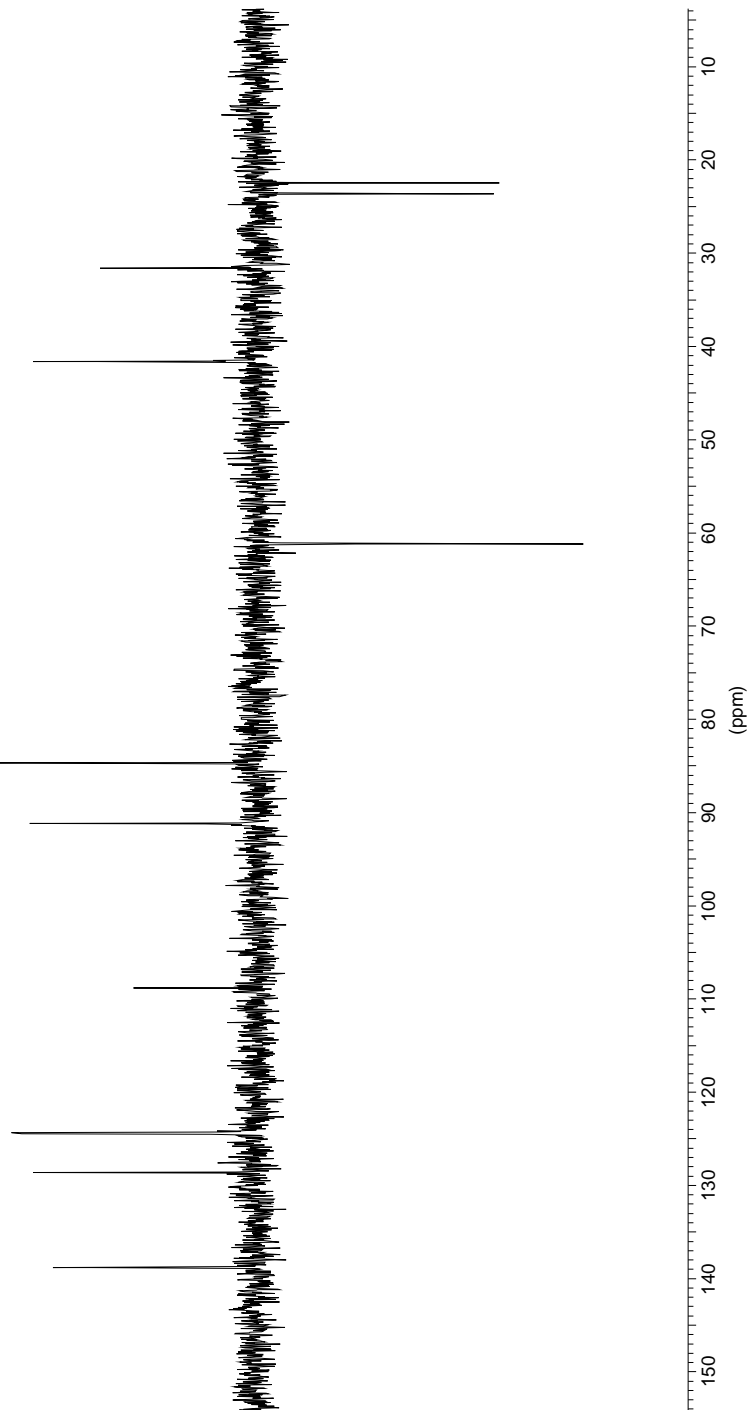
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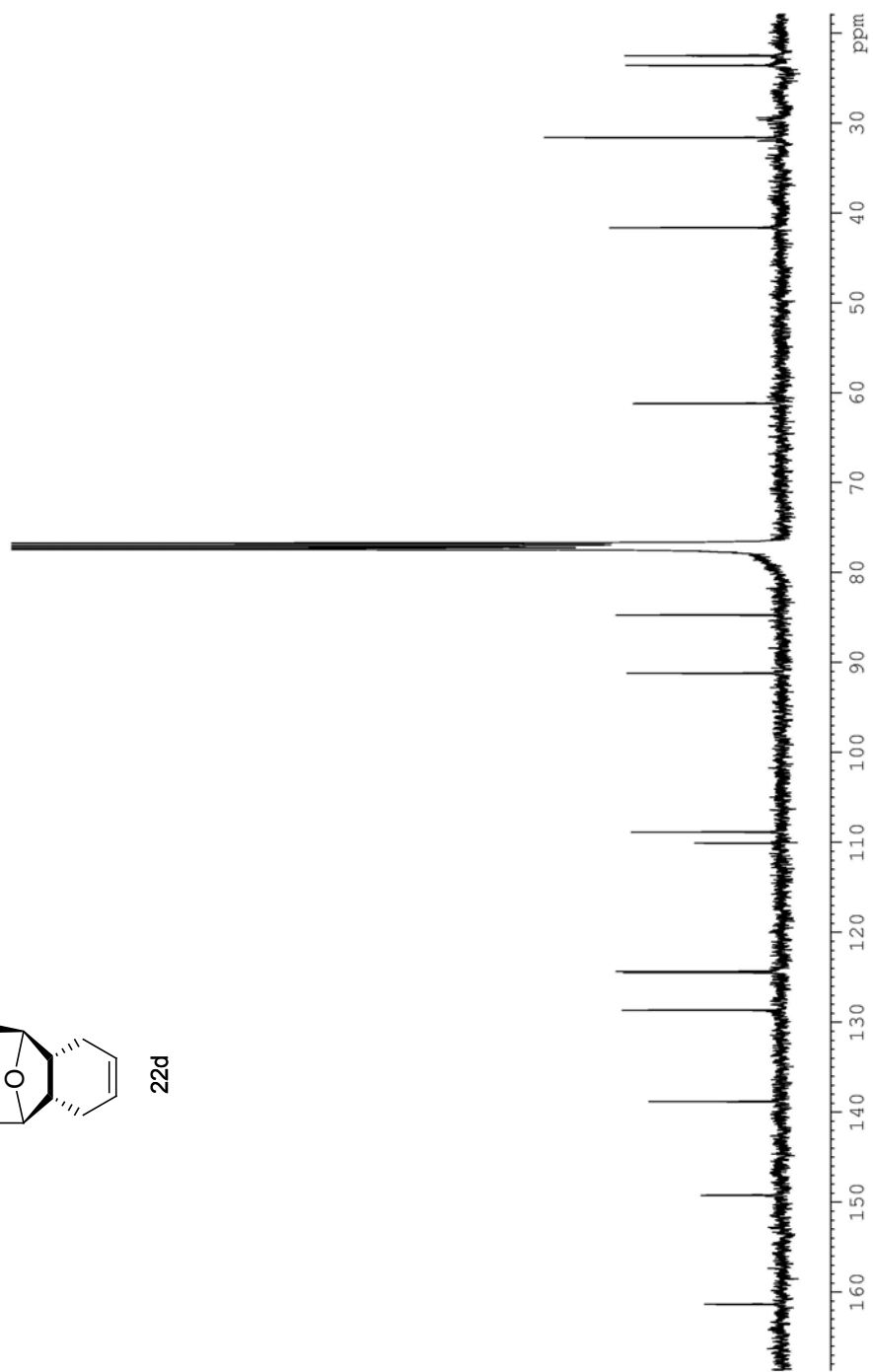
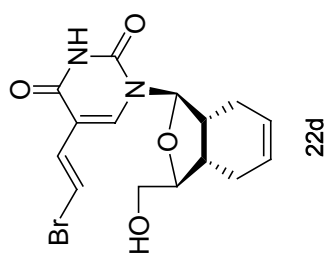




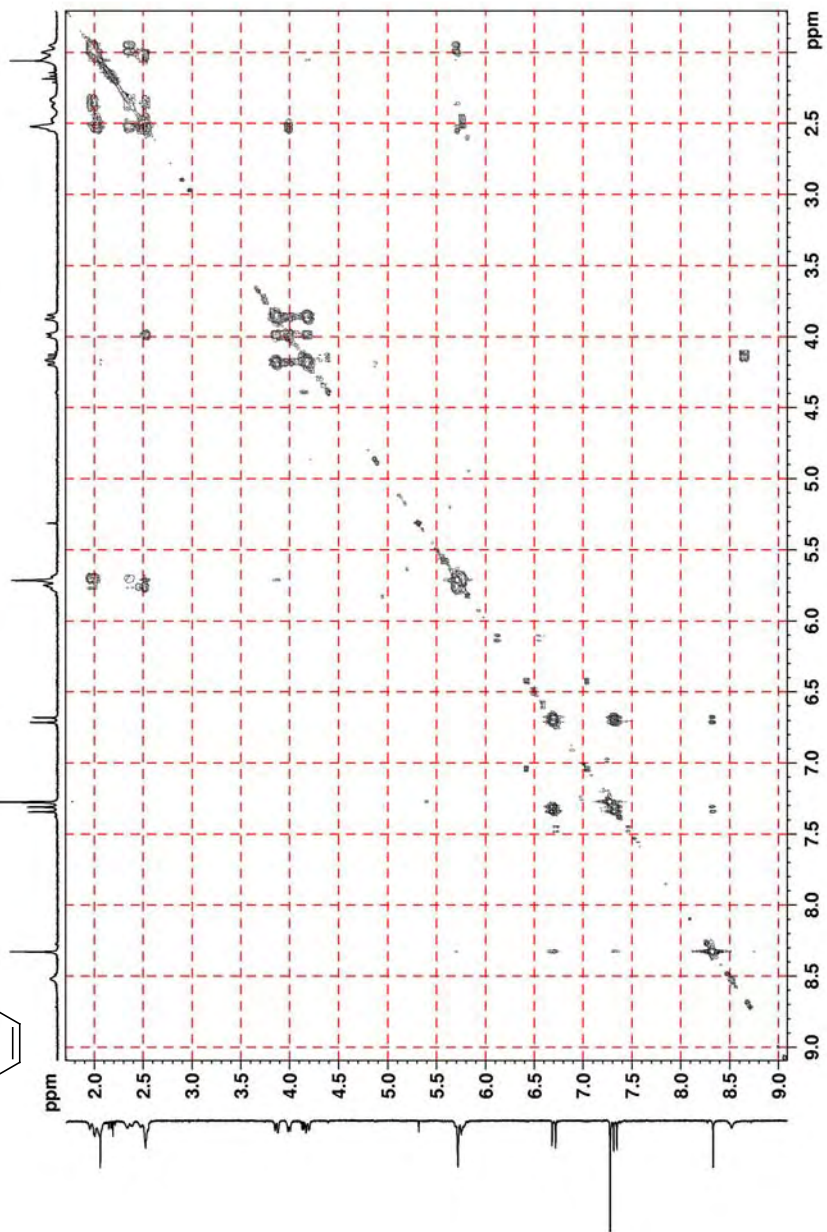
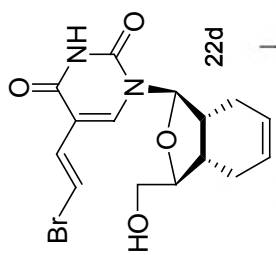


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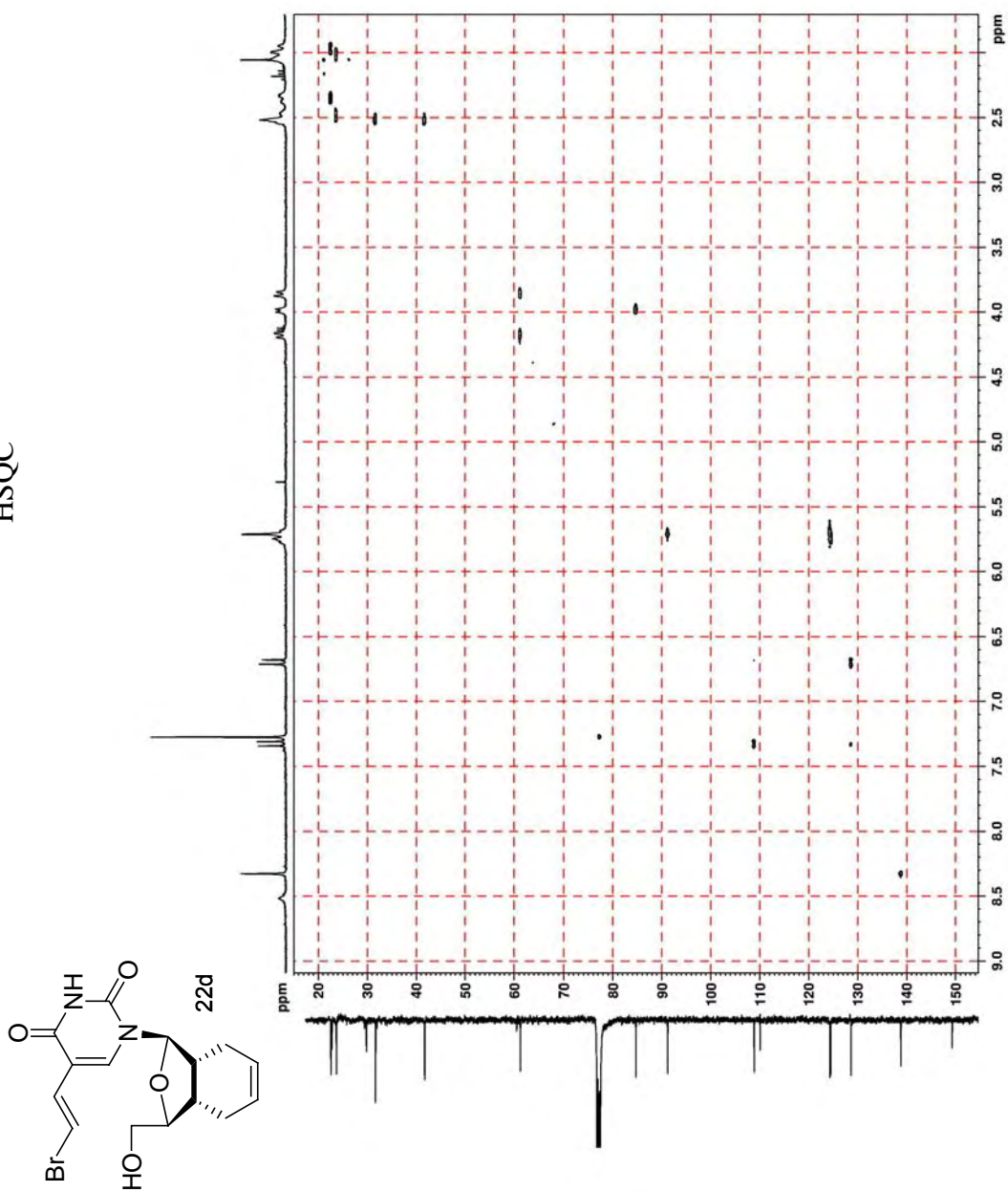


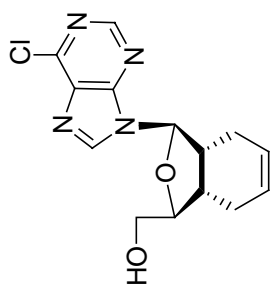


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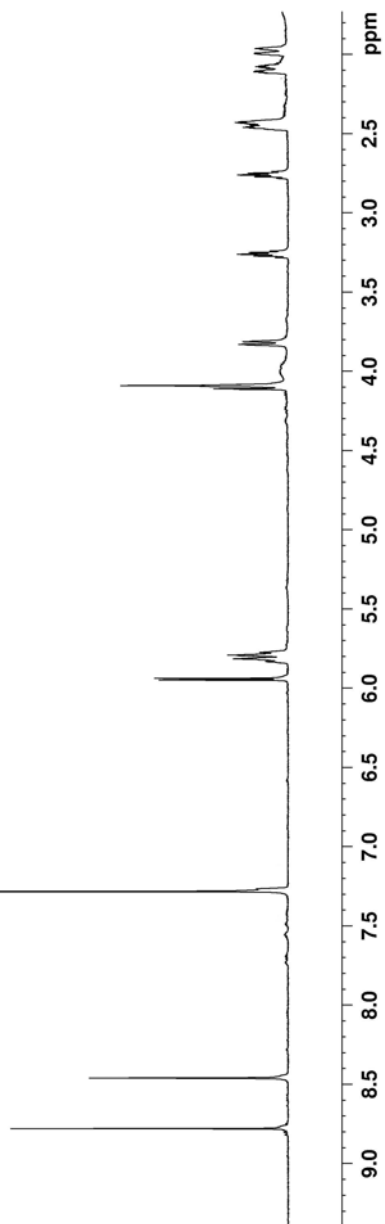


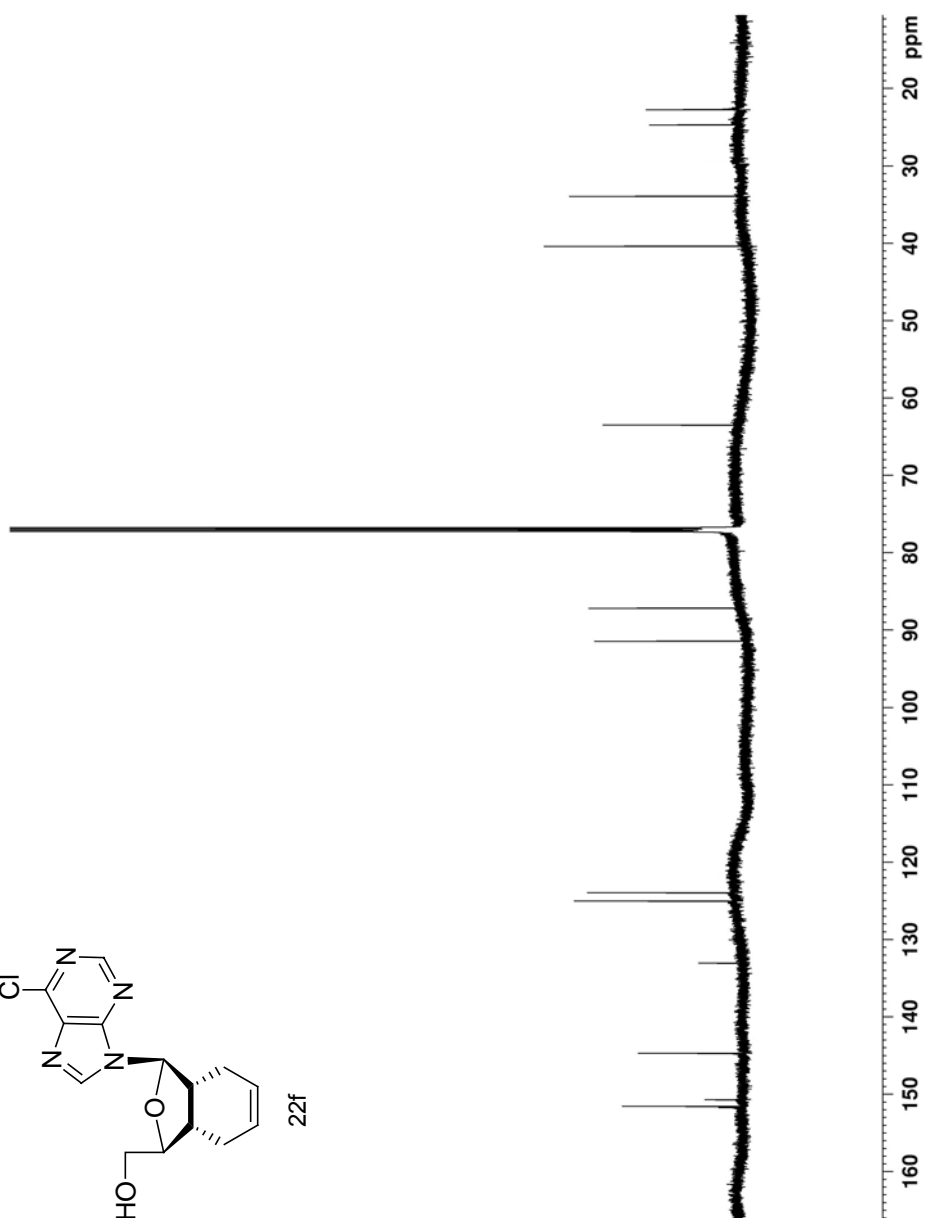
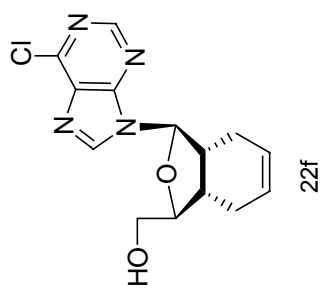
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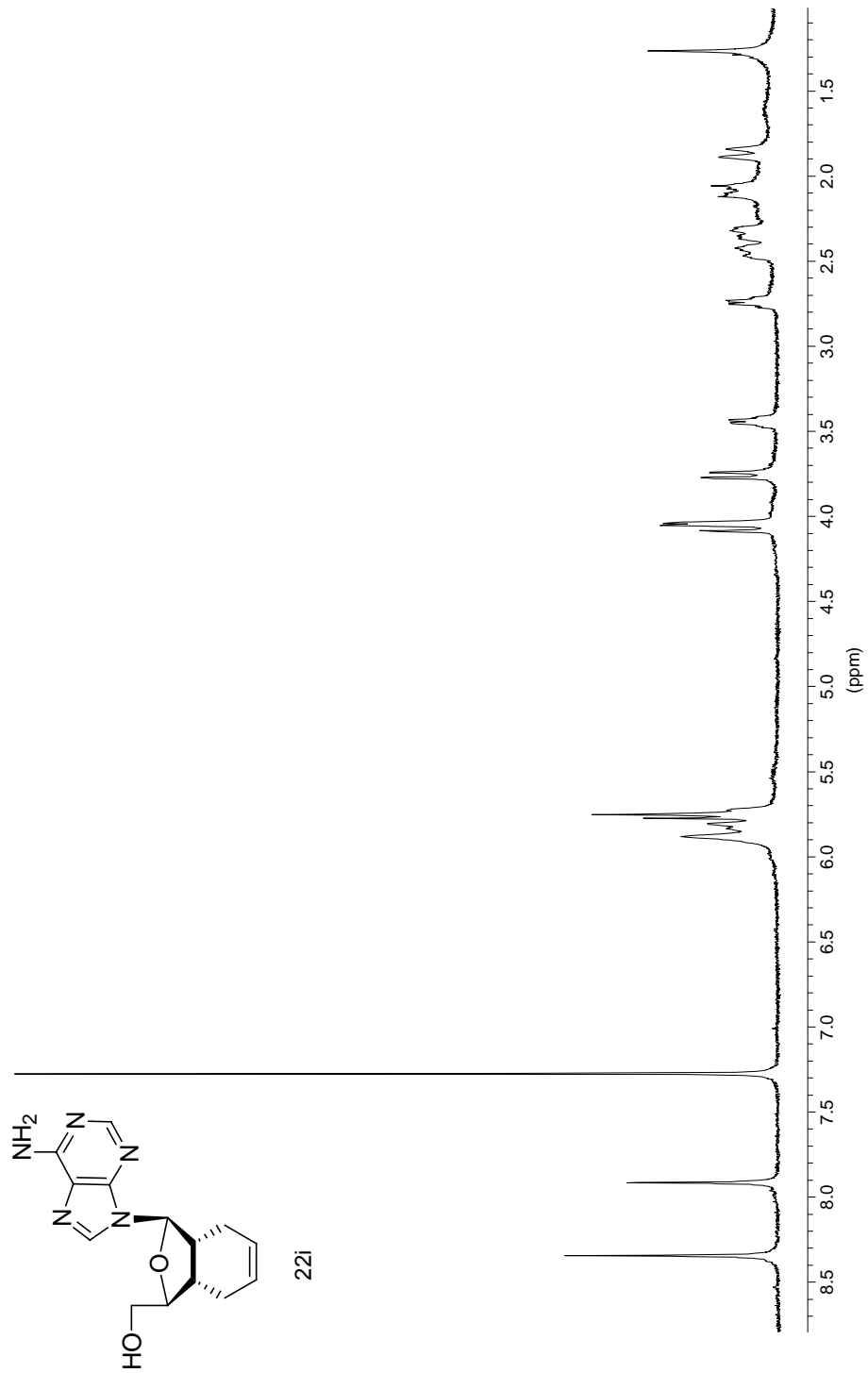


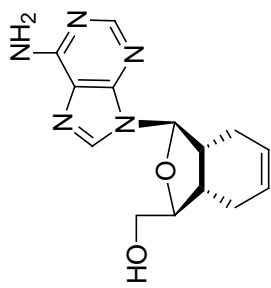


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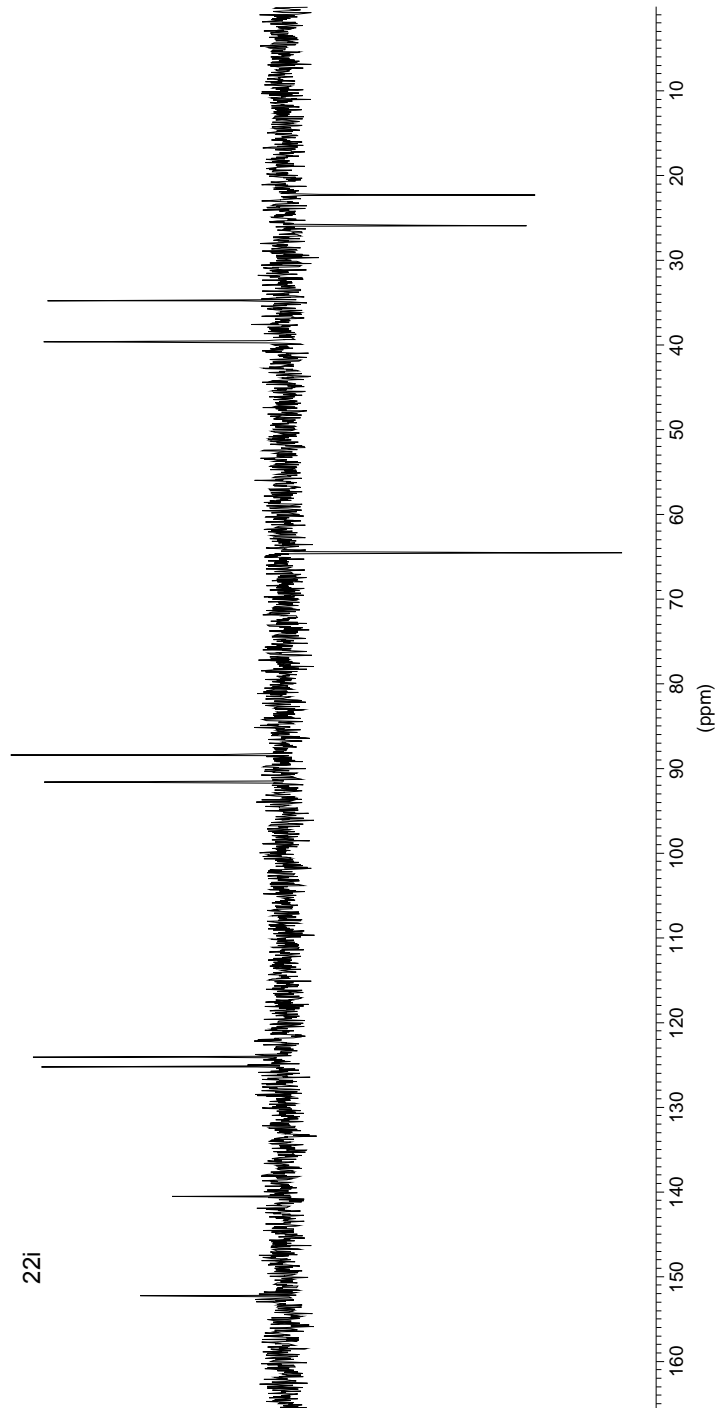


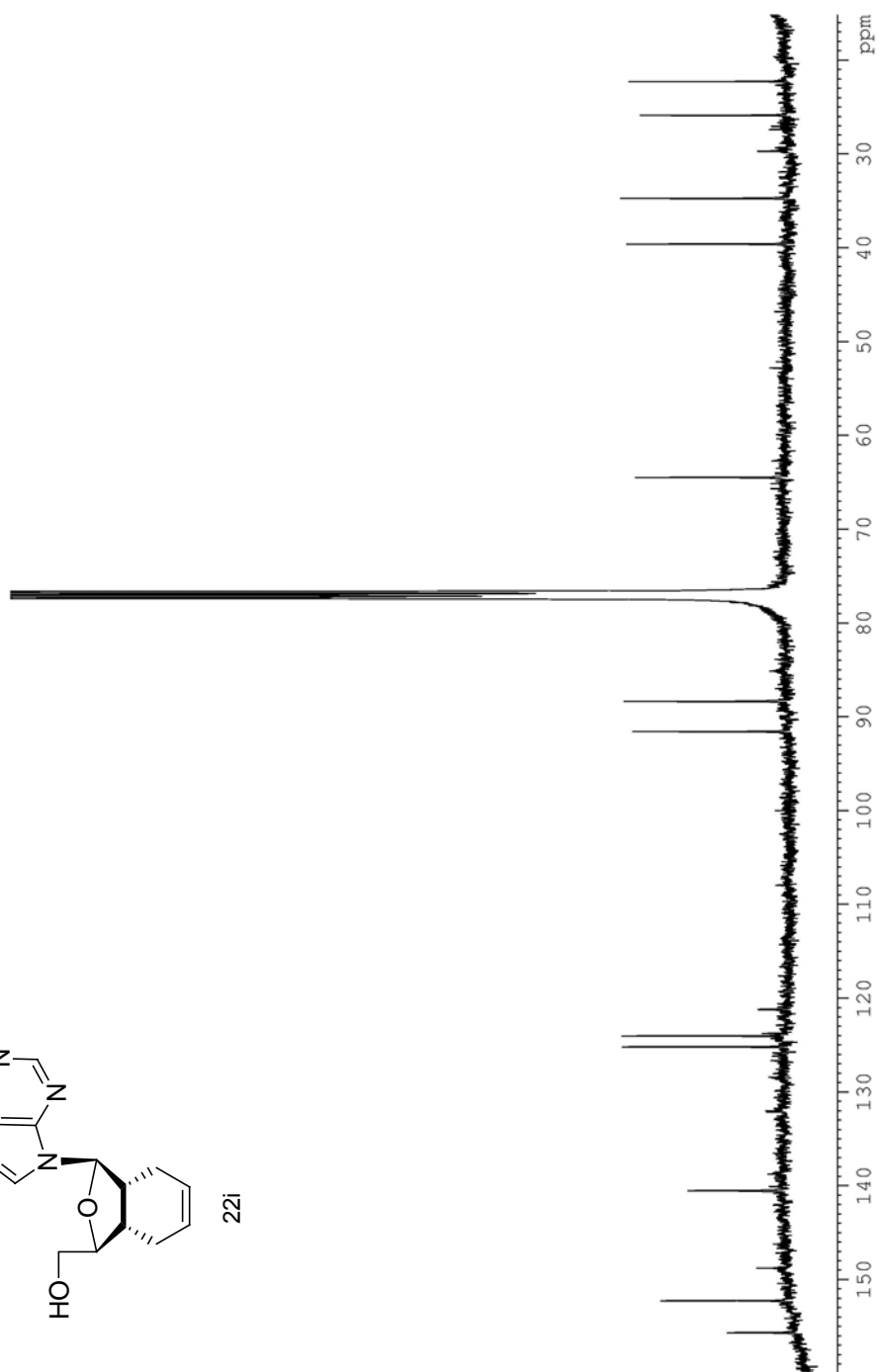
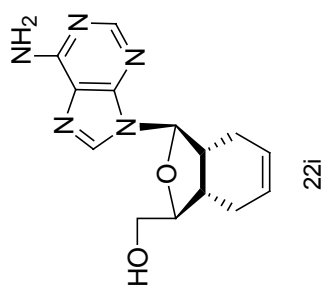


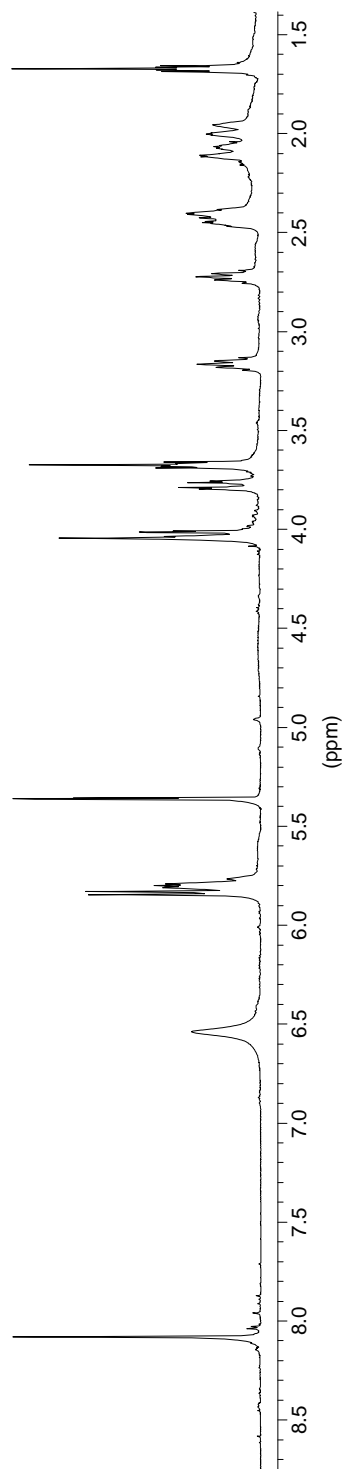
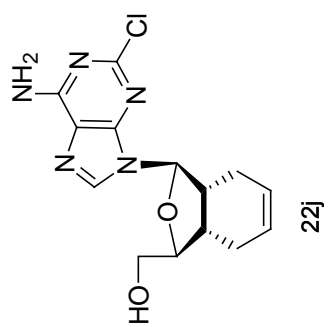


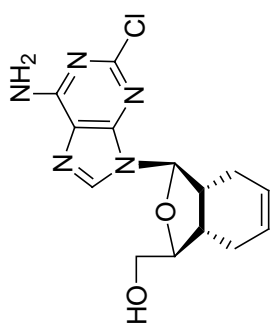


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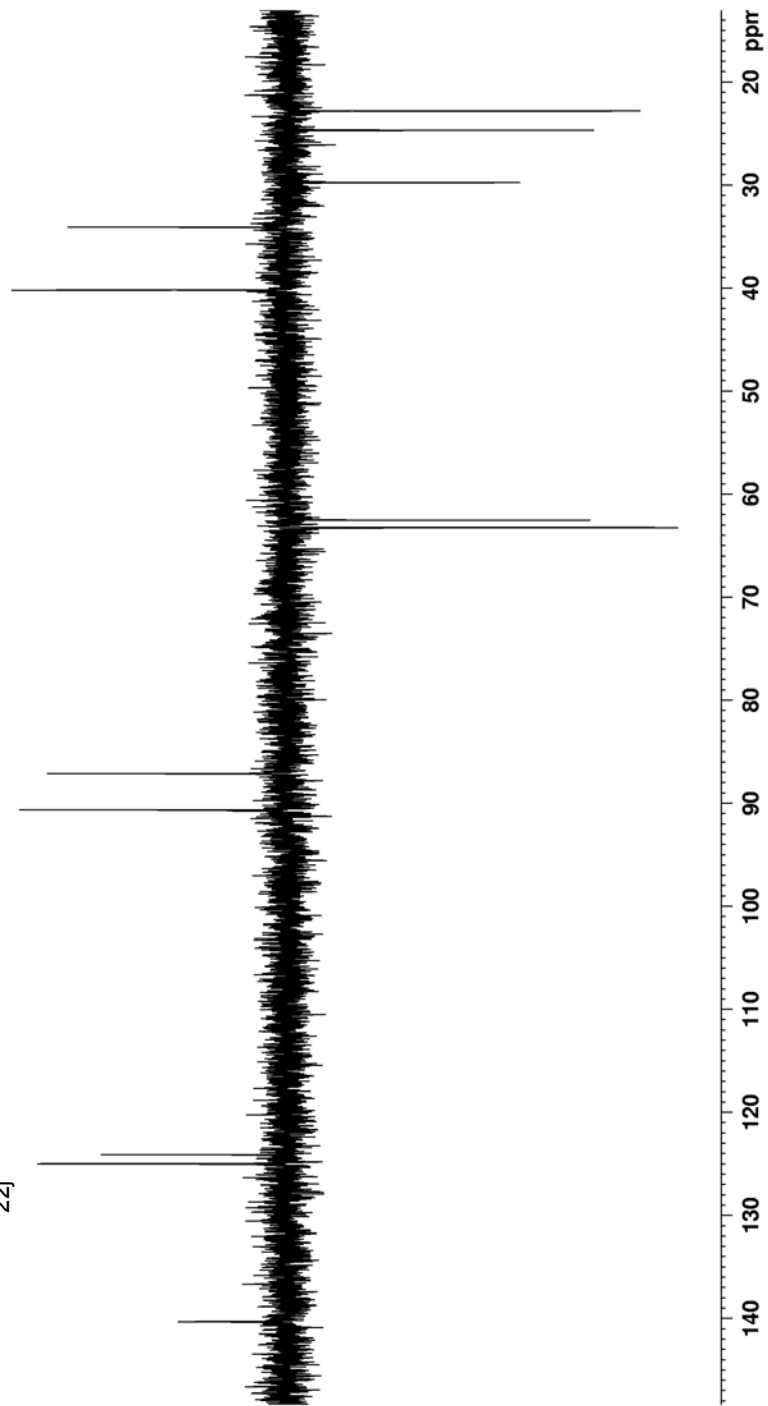


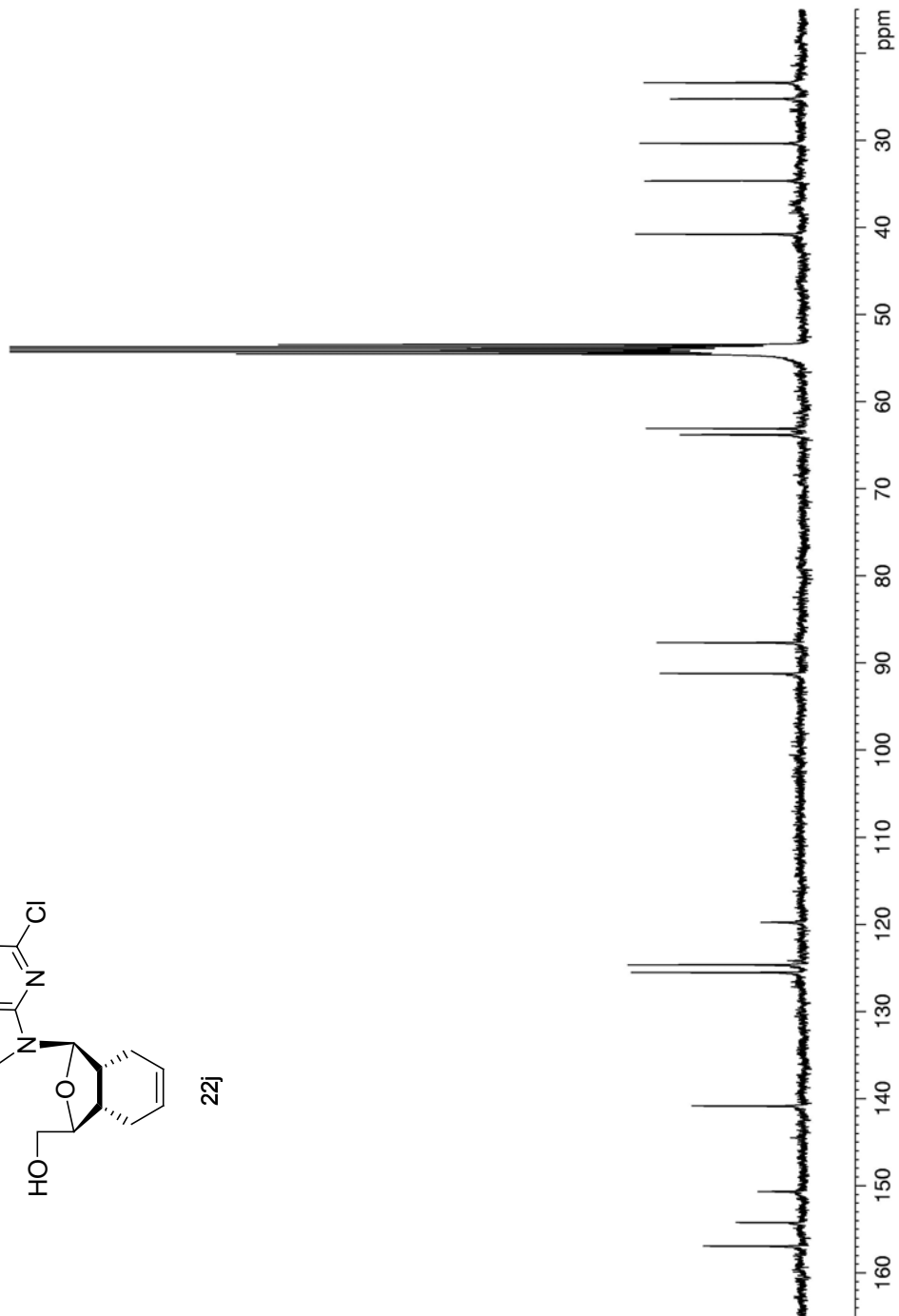
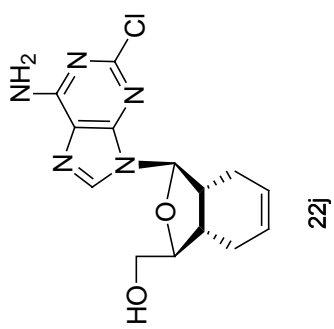




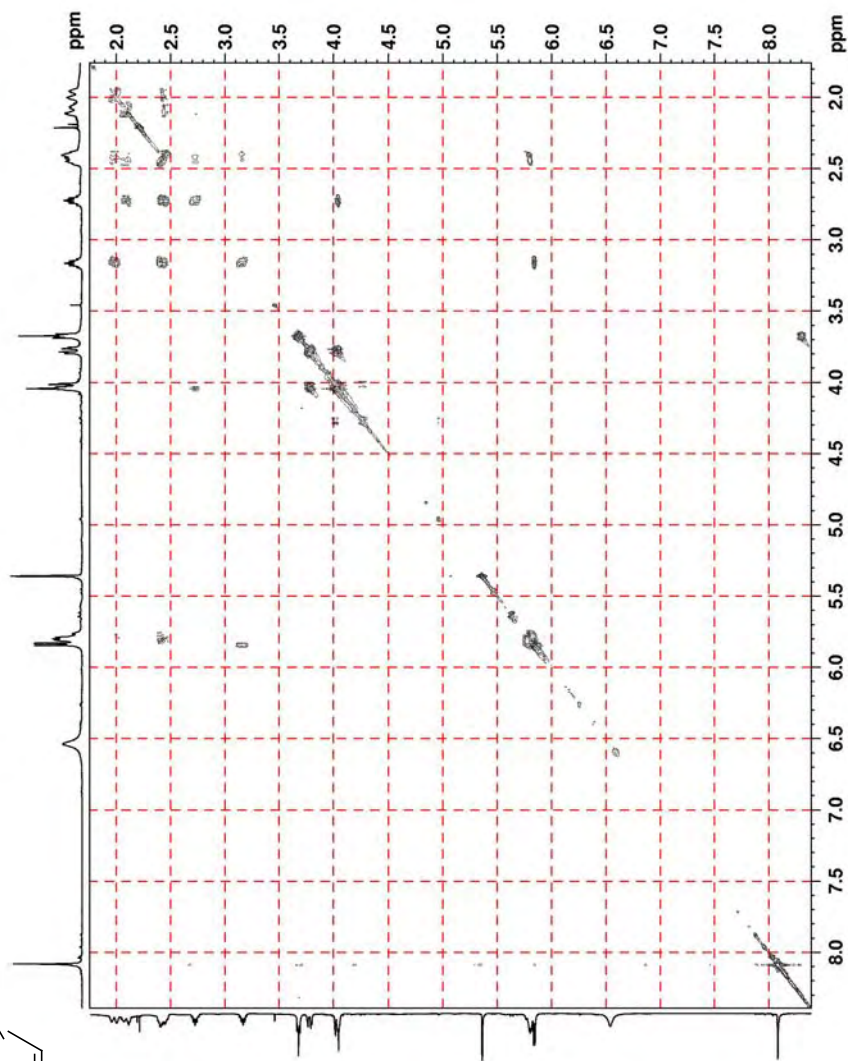
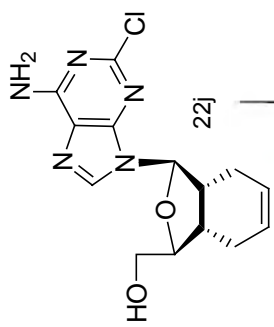


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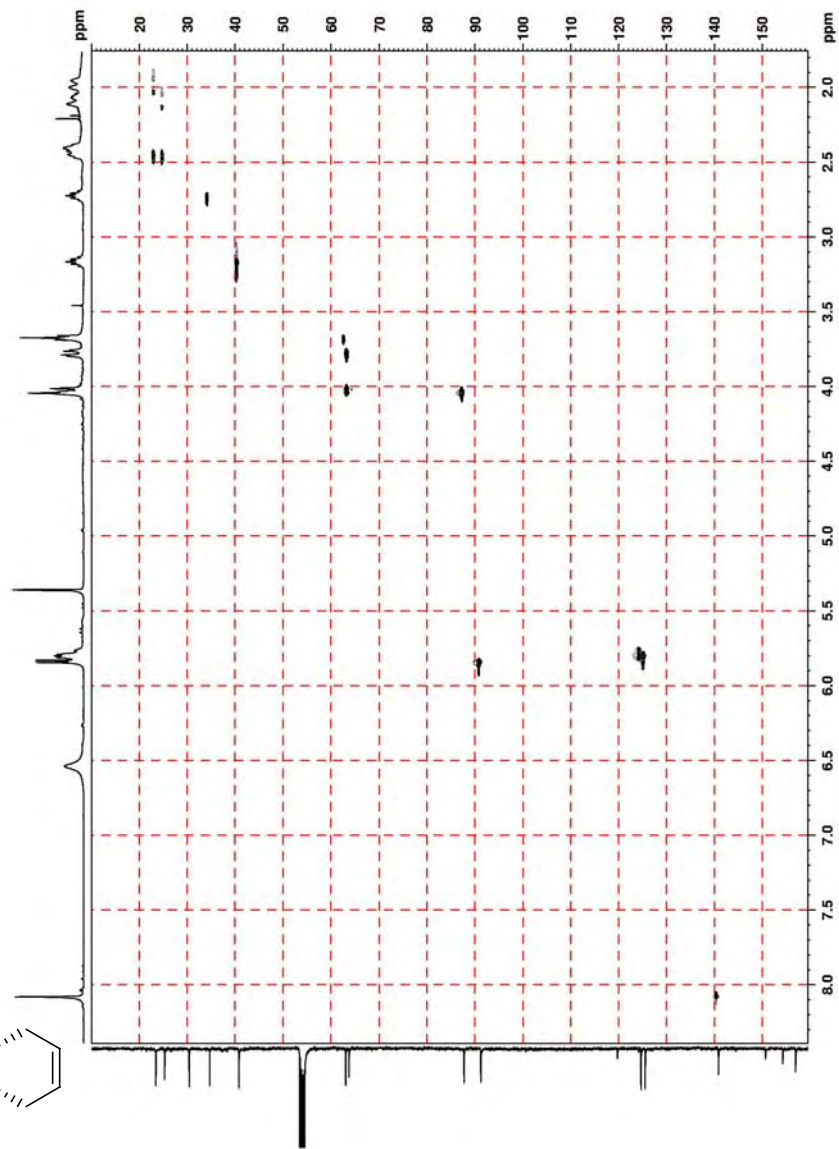
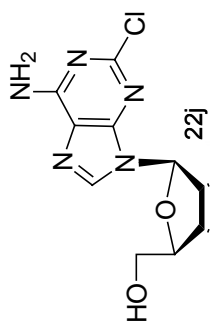


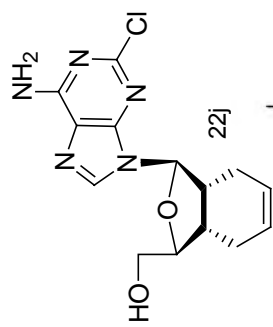


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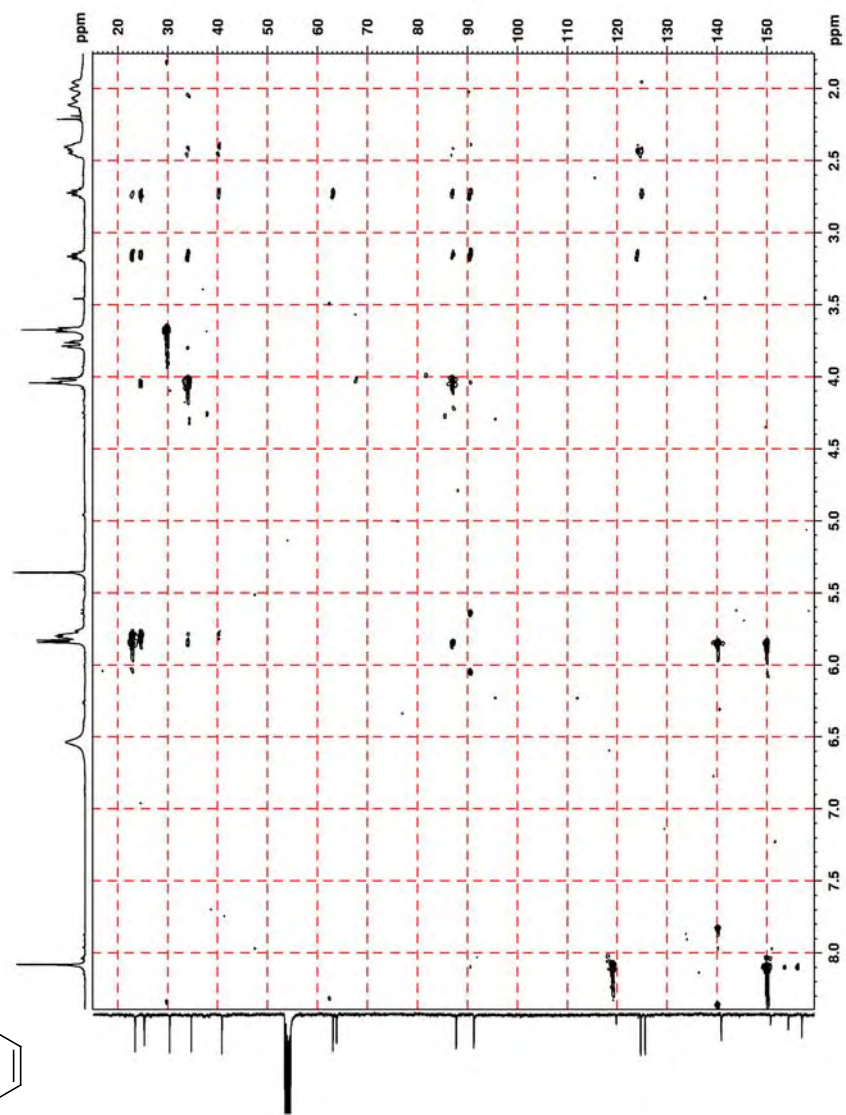


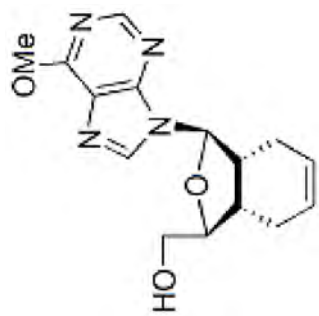
HSQC





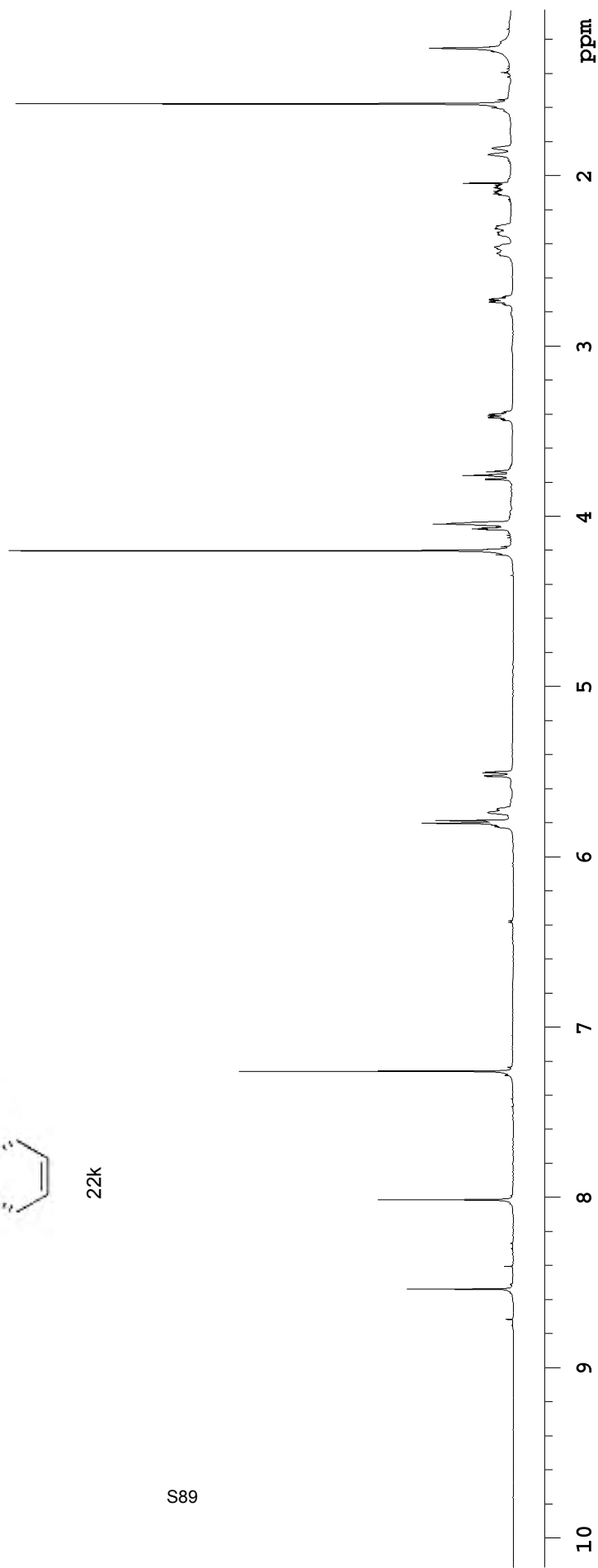
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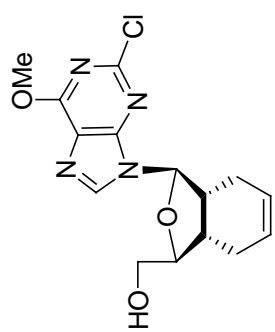




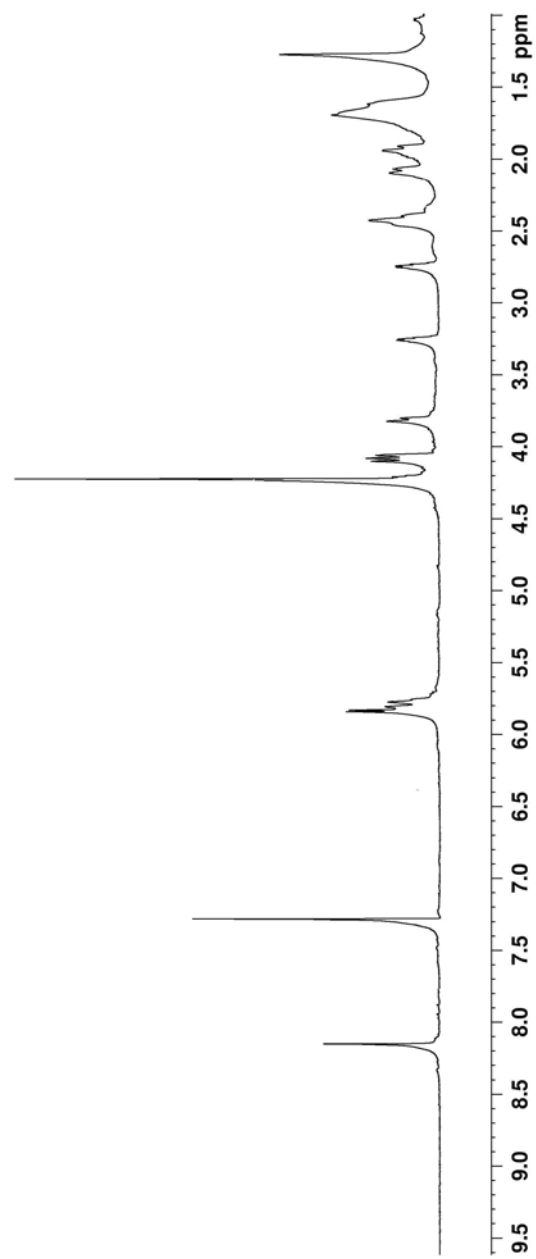
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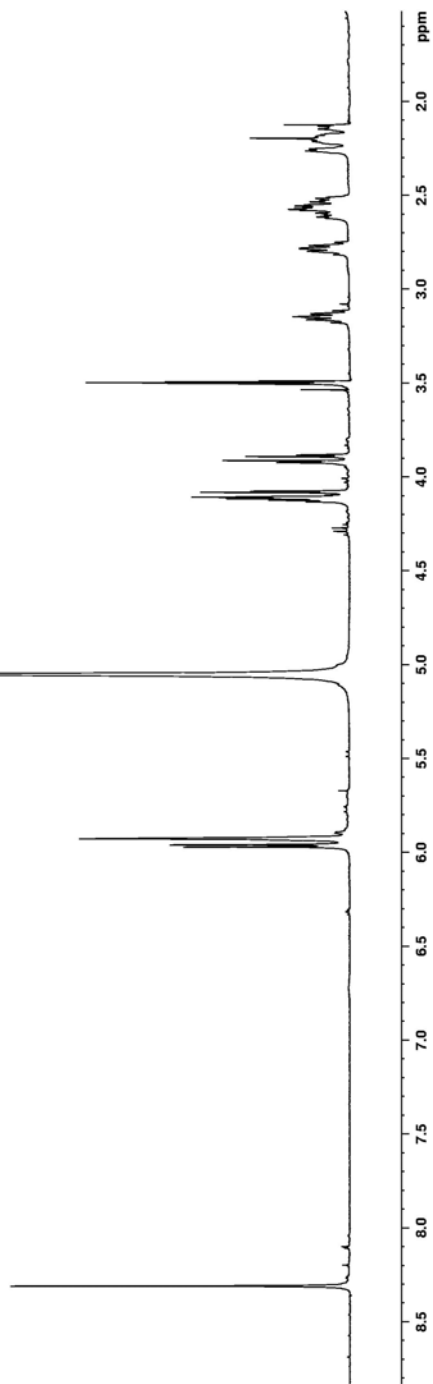
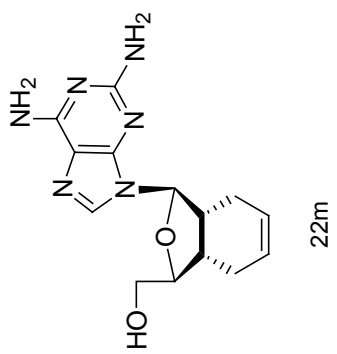
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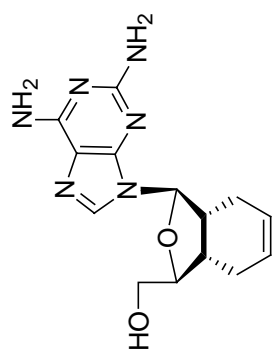




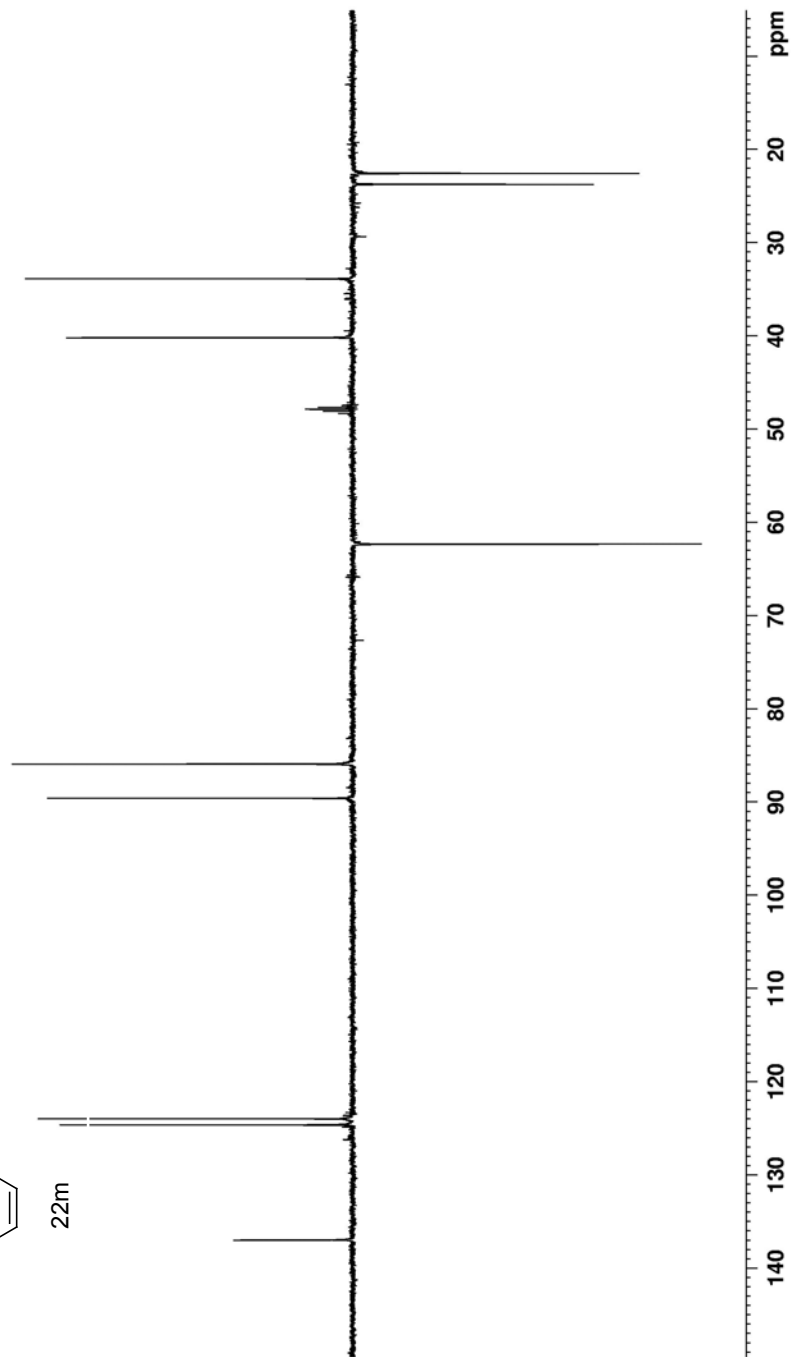
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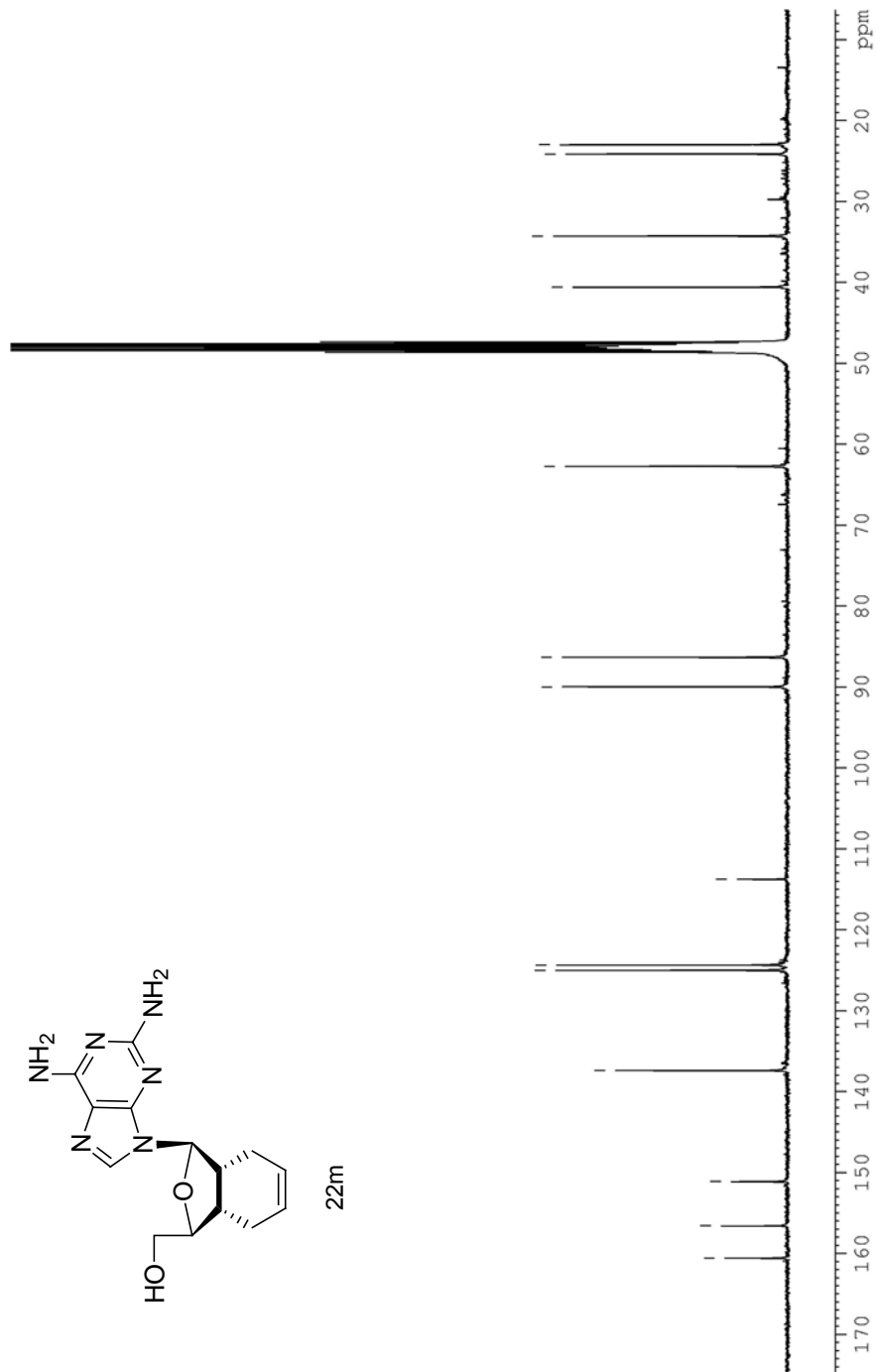
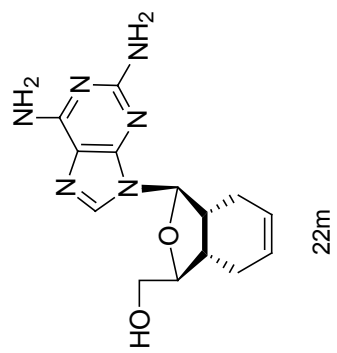


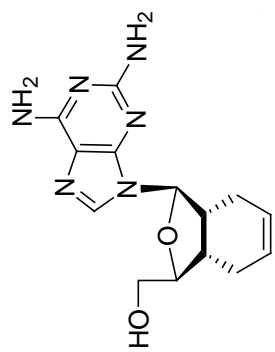




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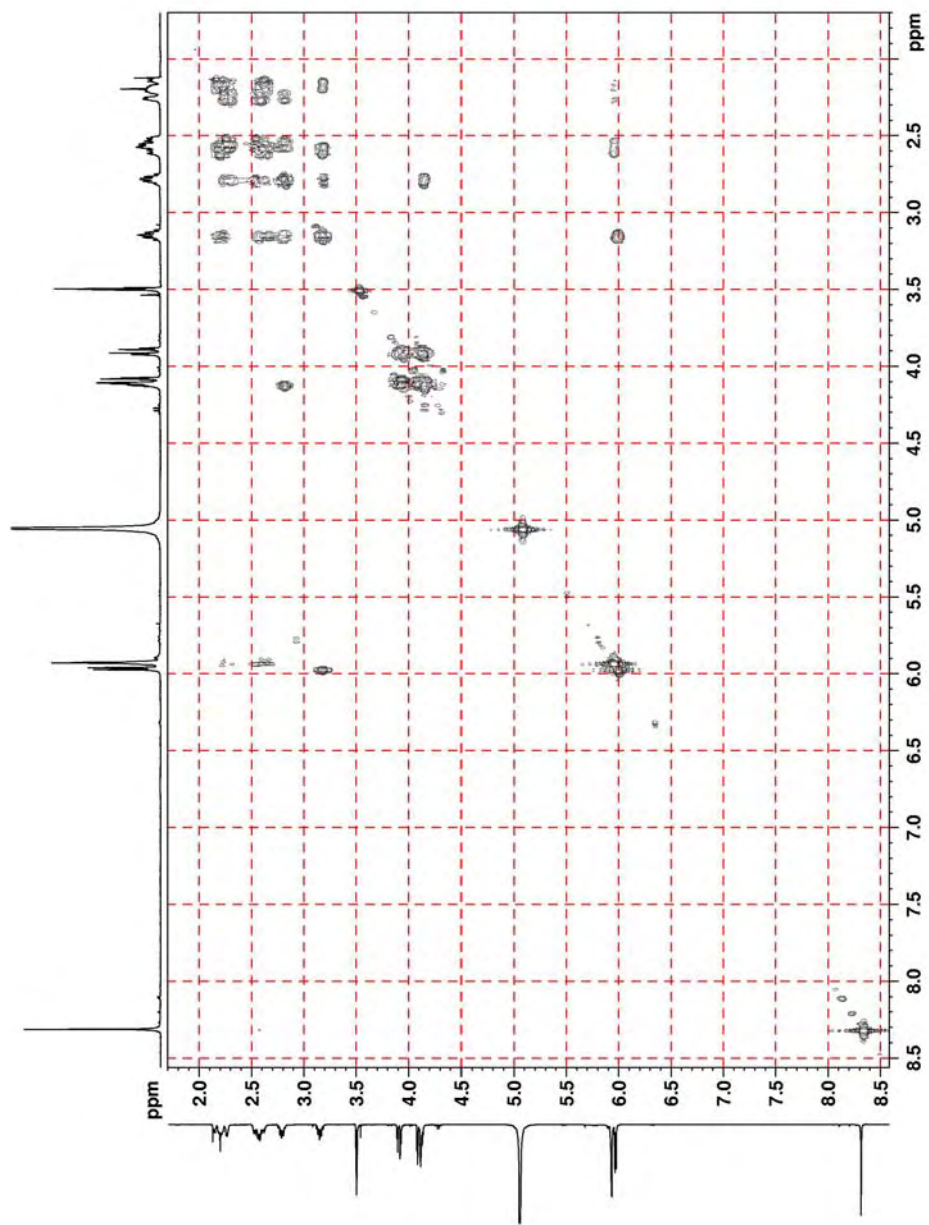




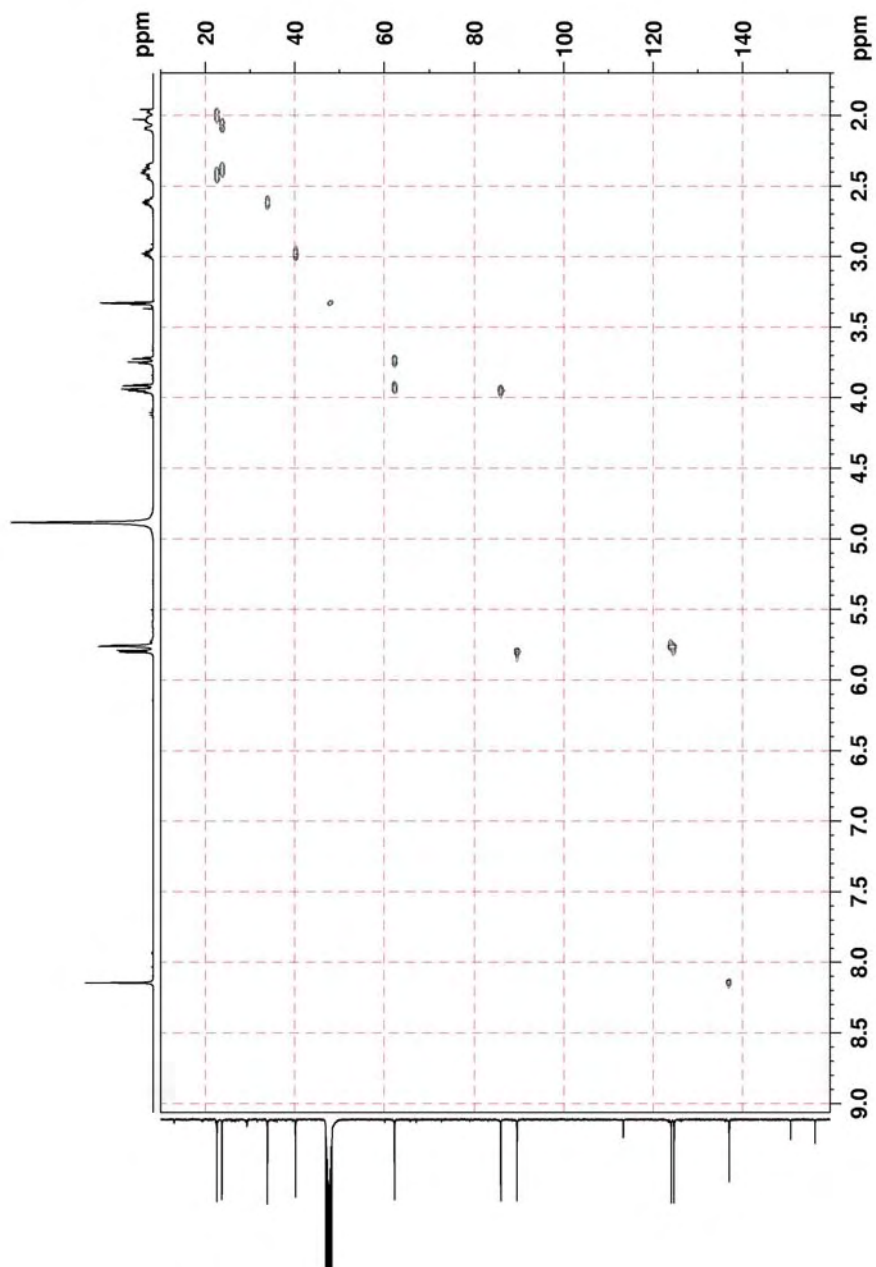
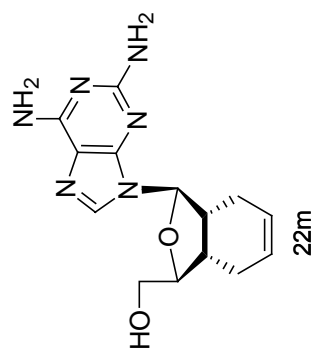


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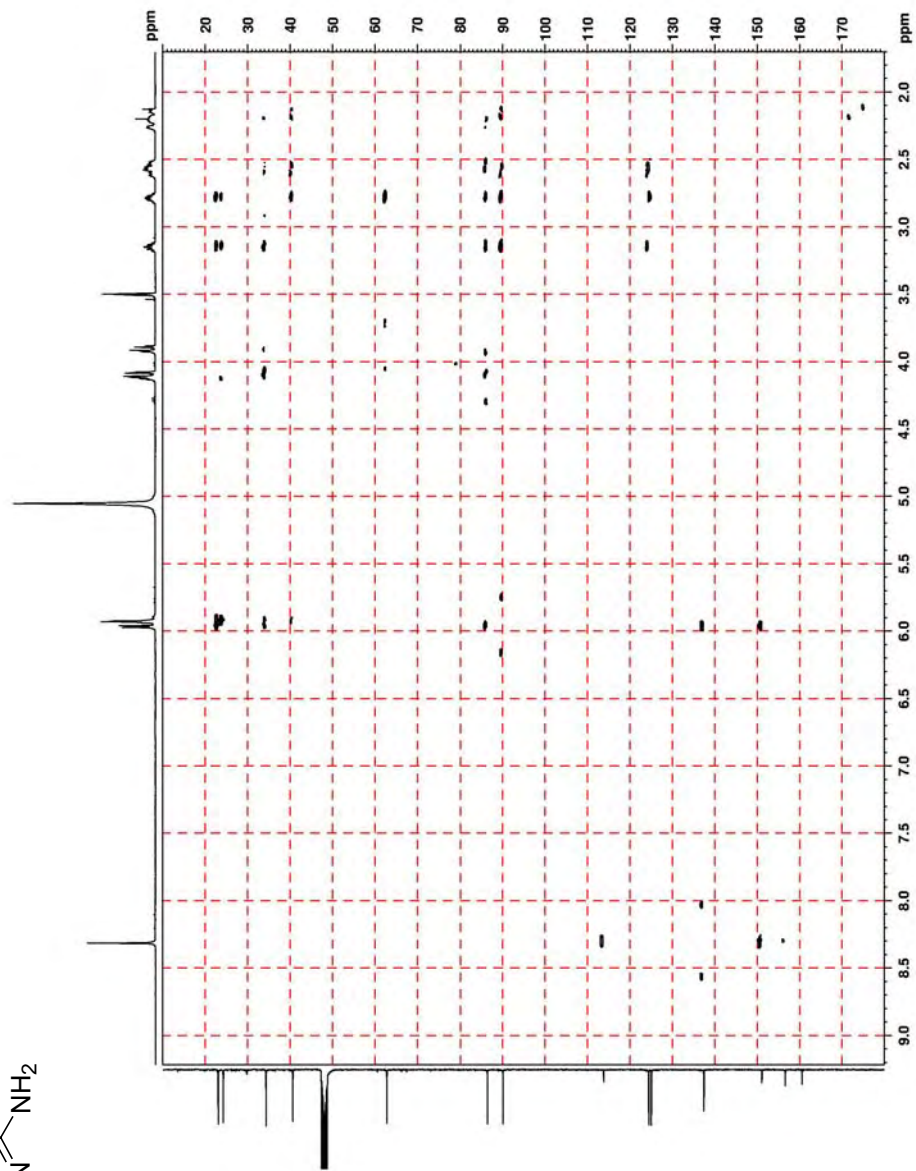
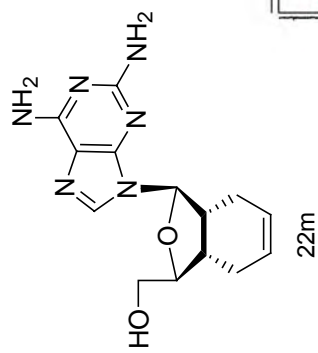
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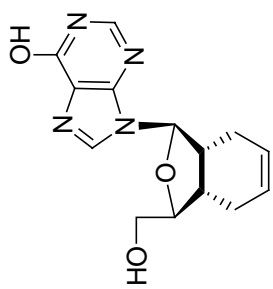


HSQC

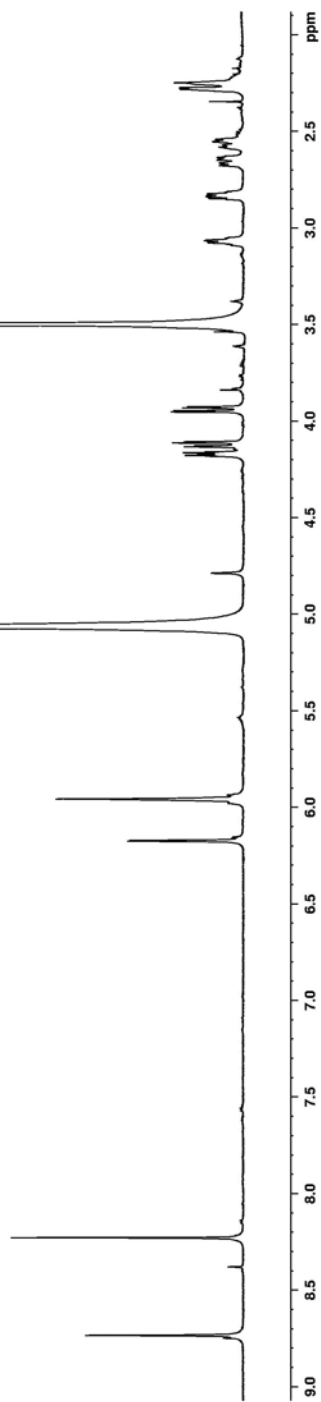


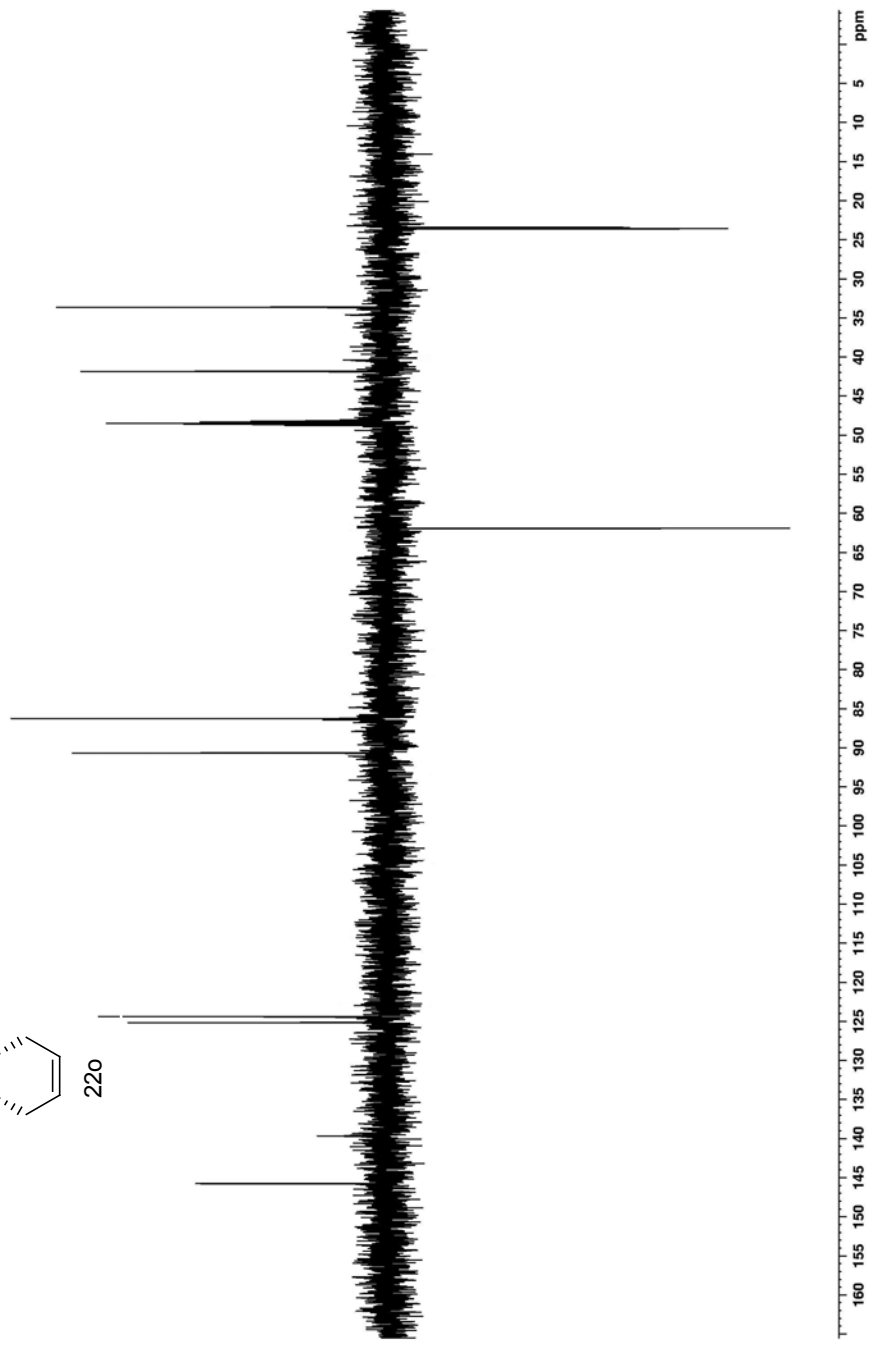
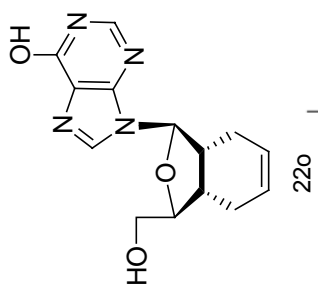
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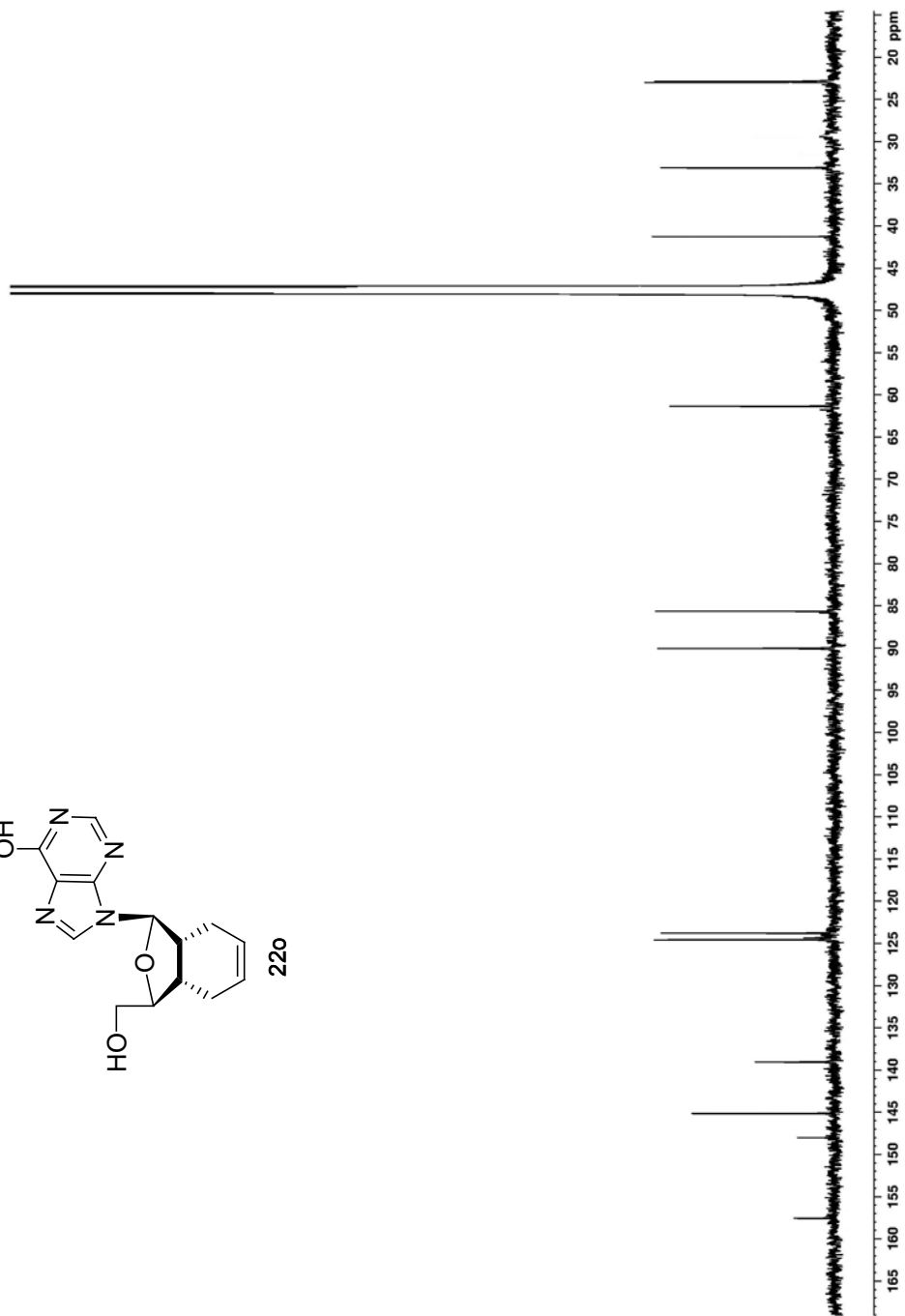
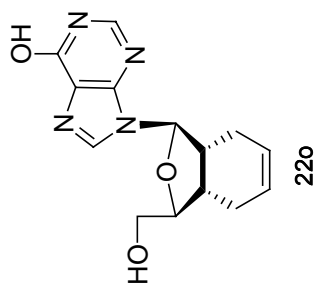




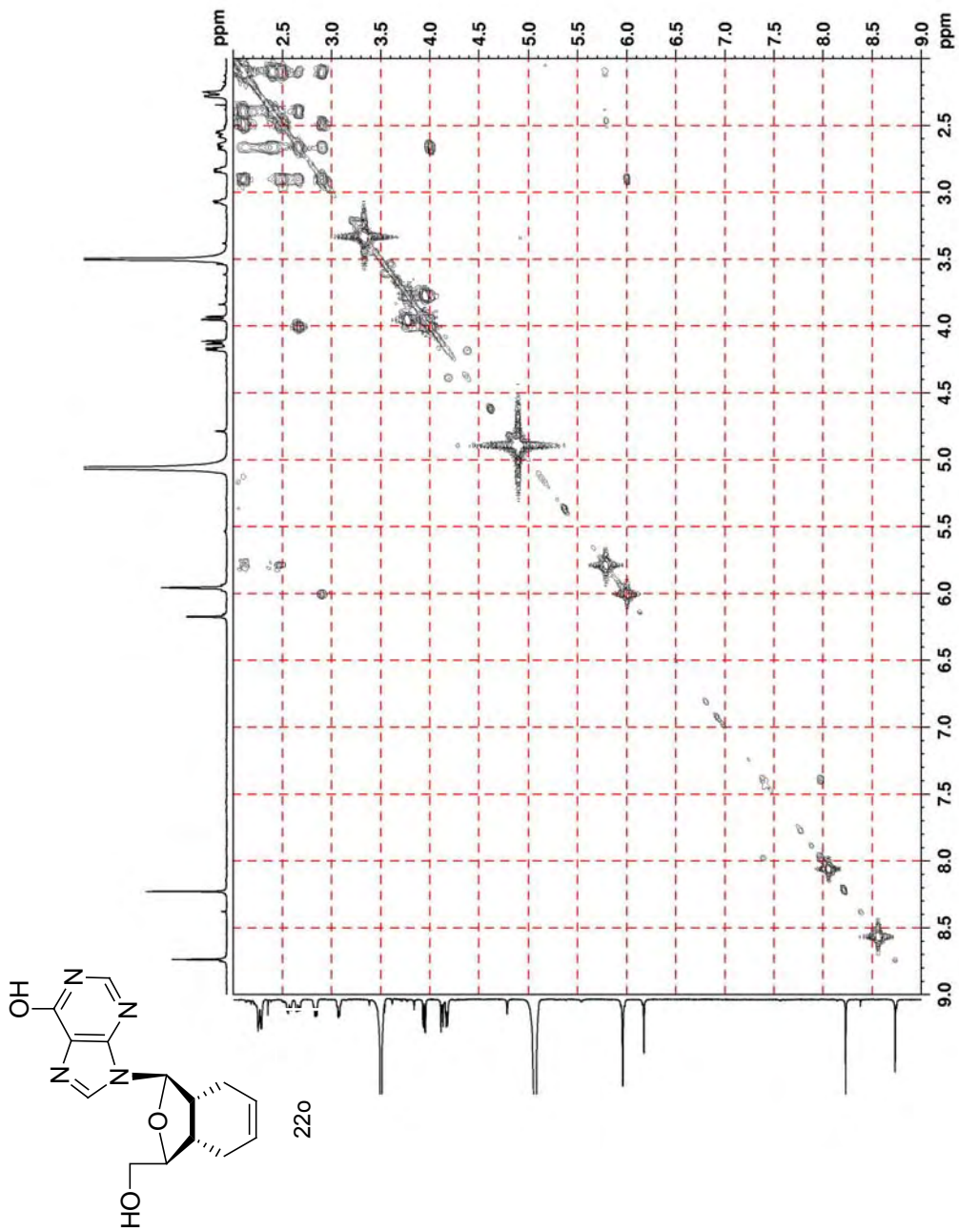
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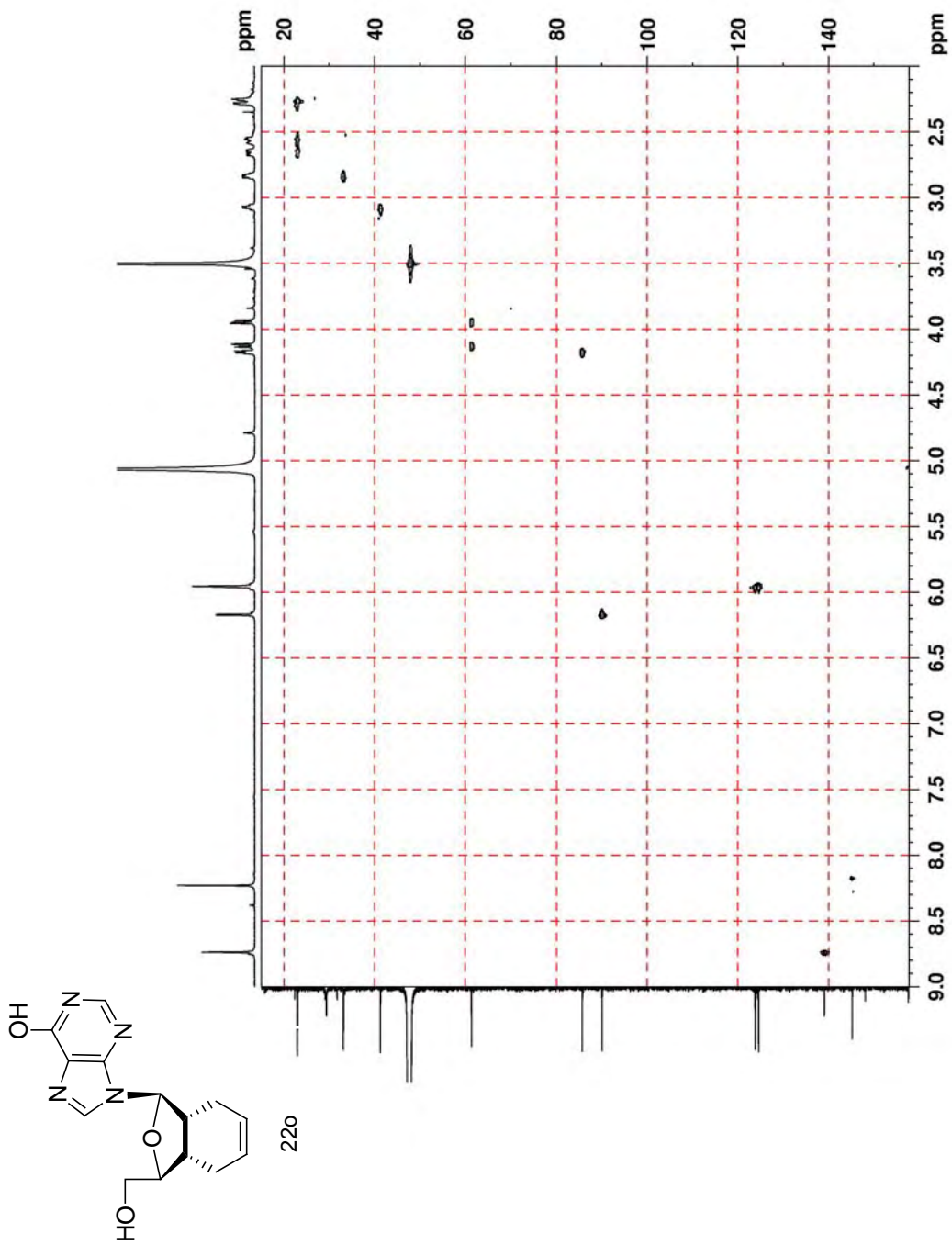




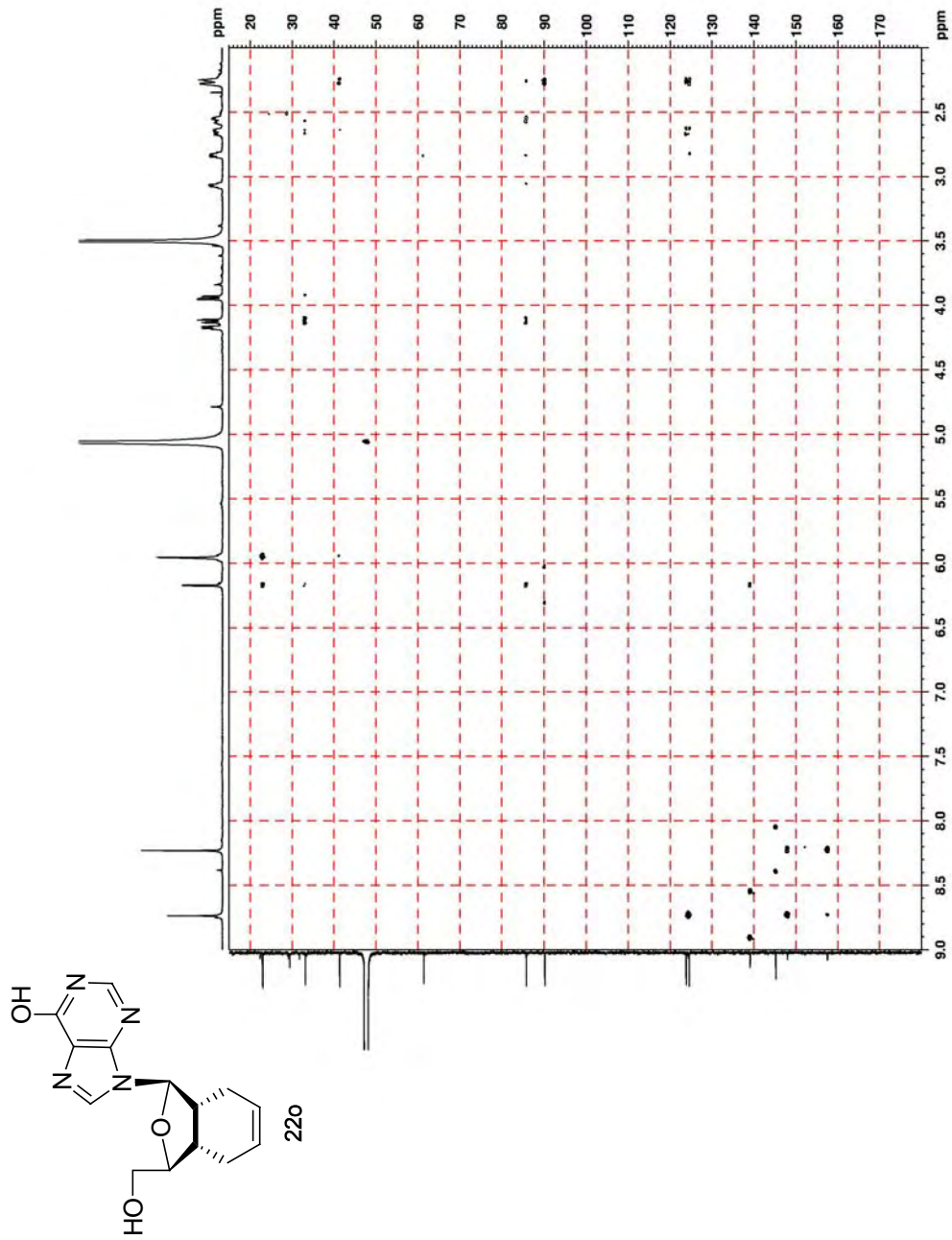
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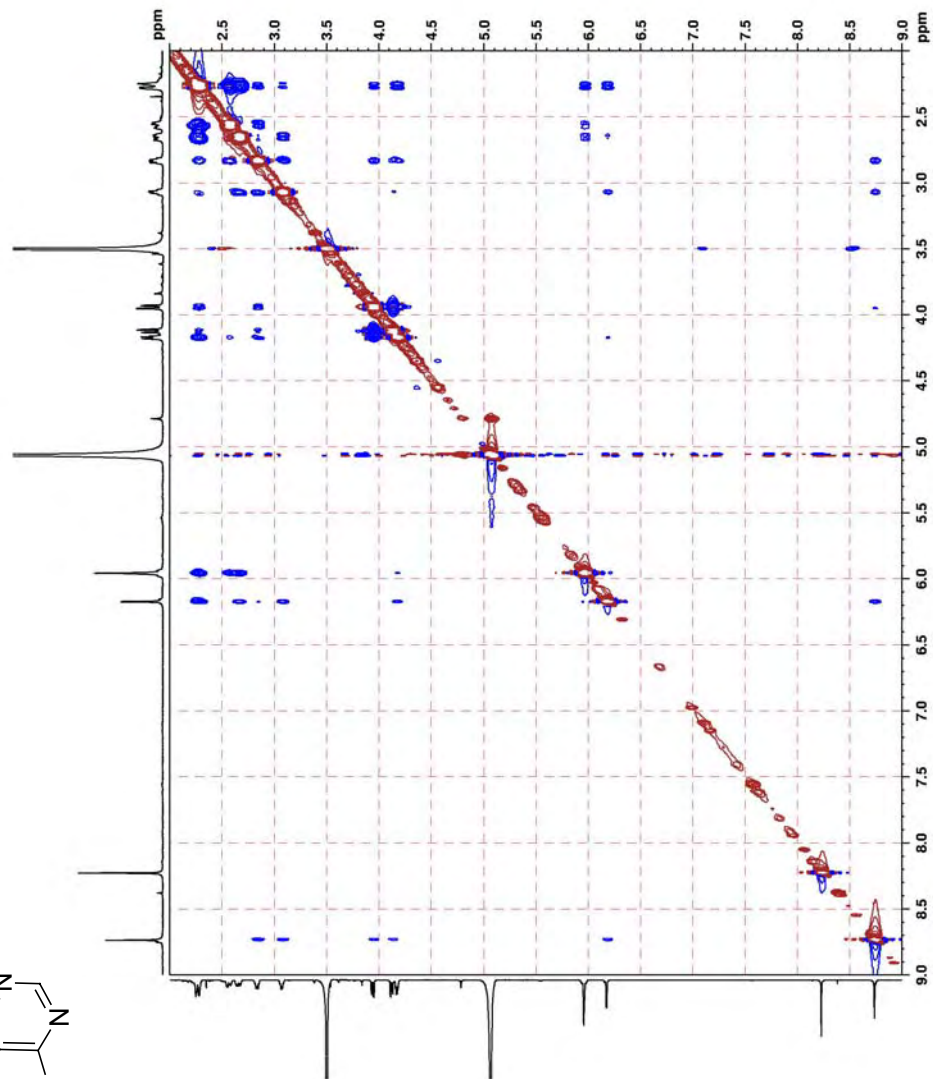
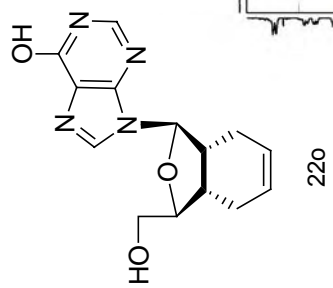
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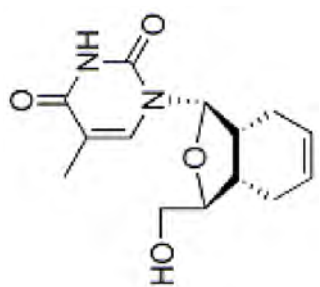


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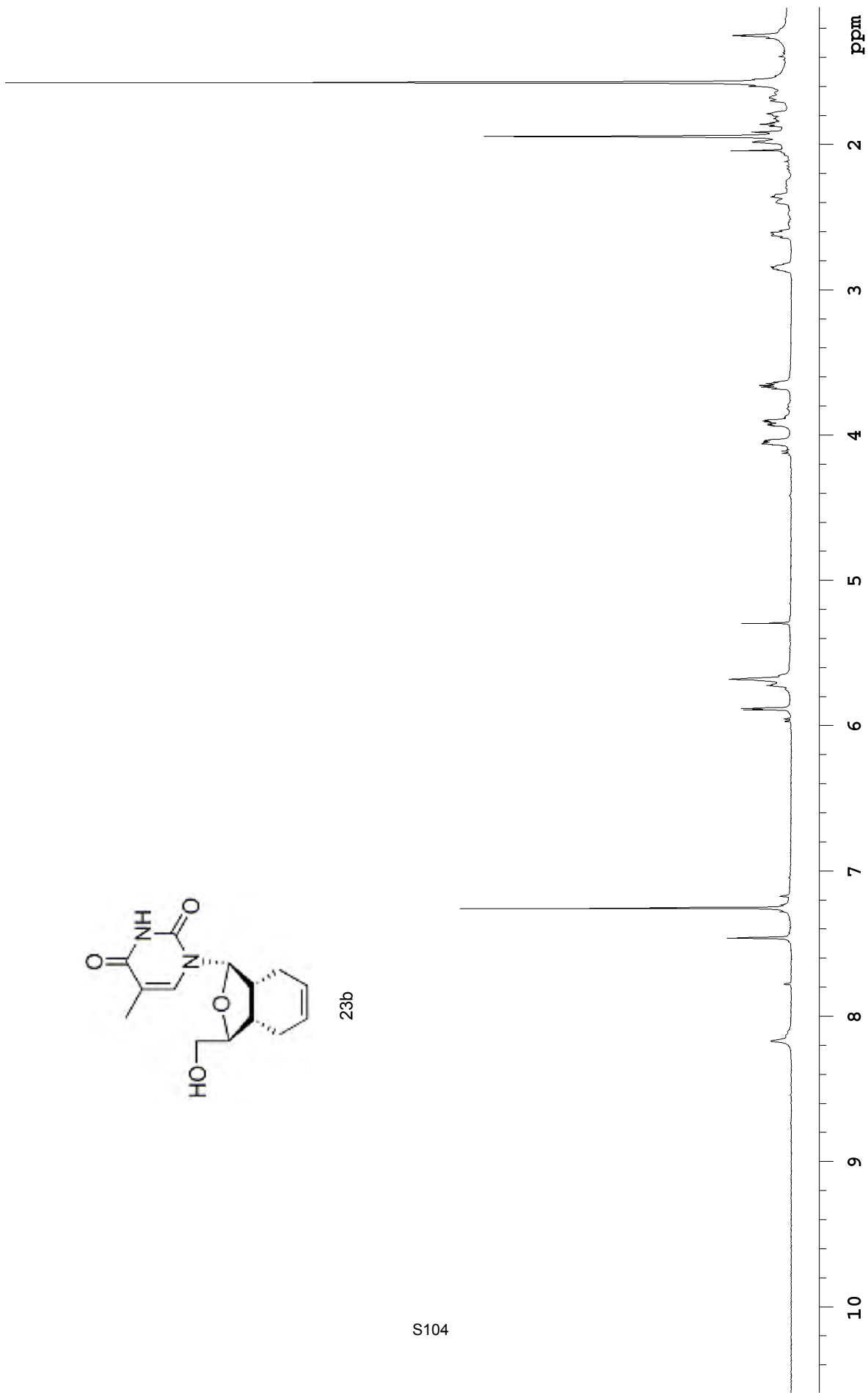
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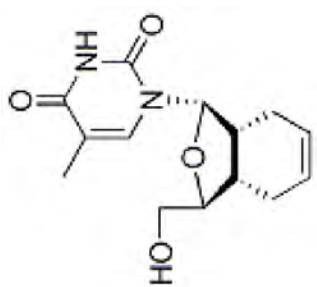




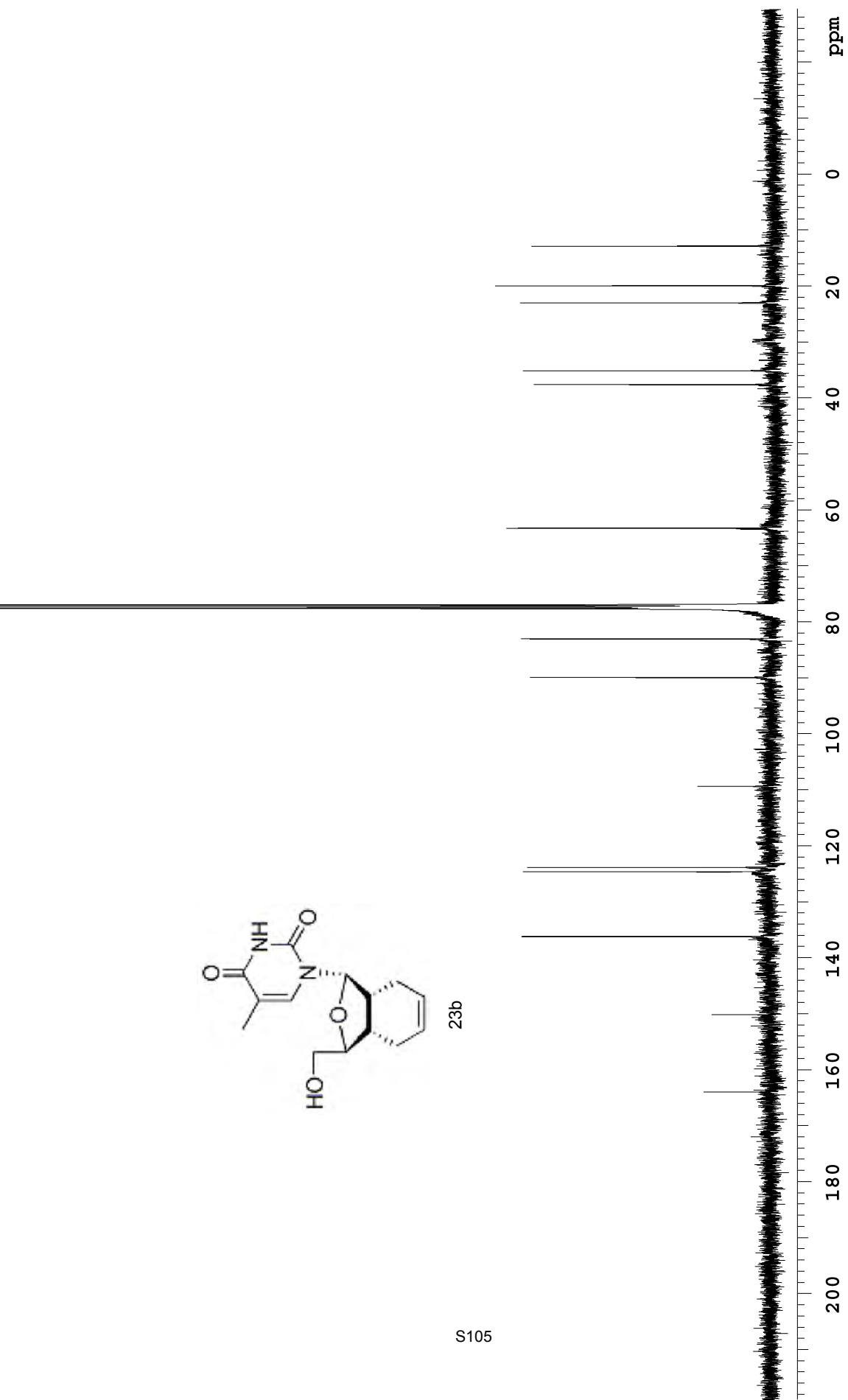
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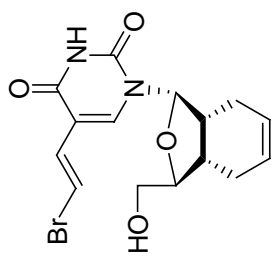
S104



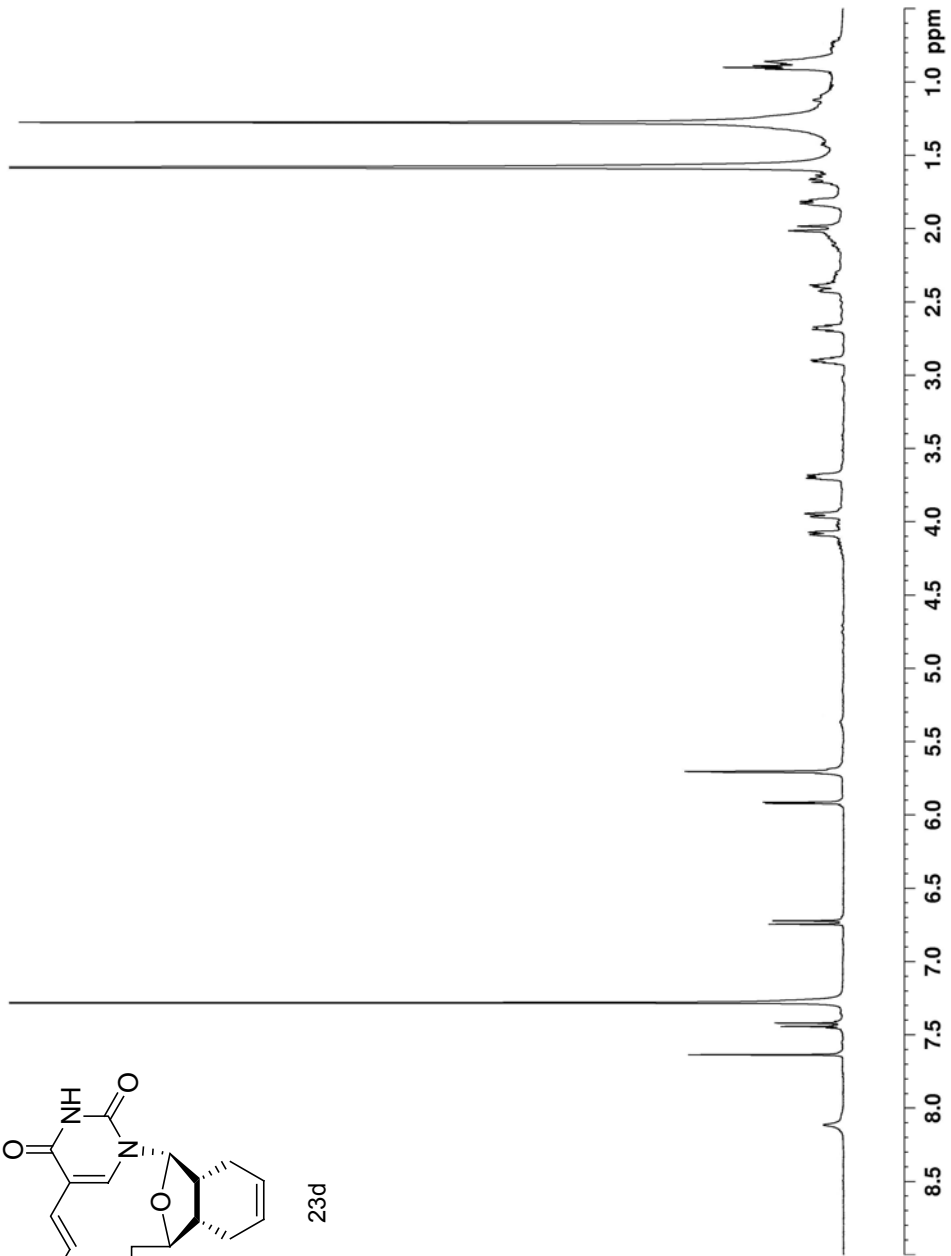


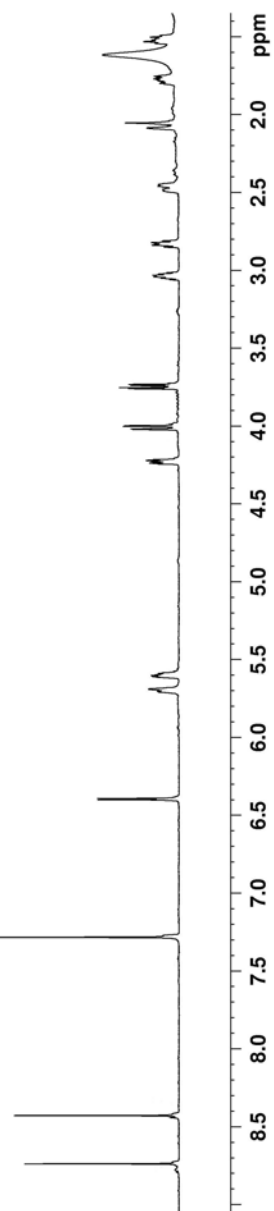
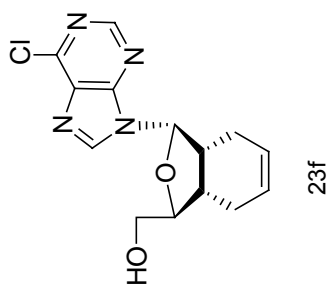
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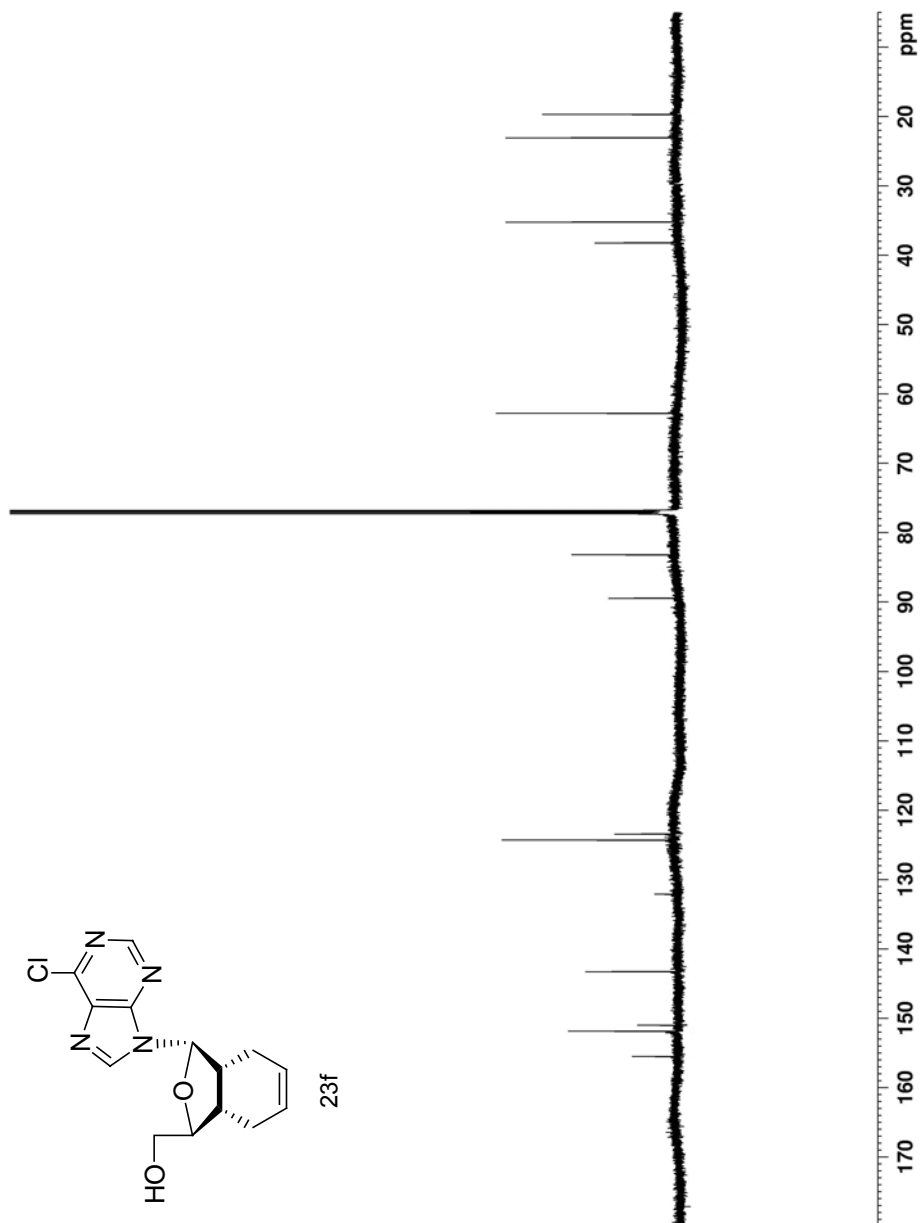
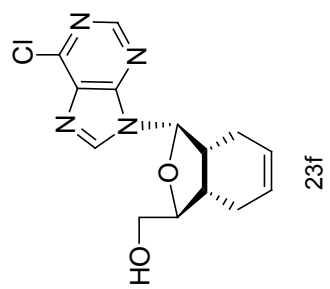


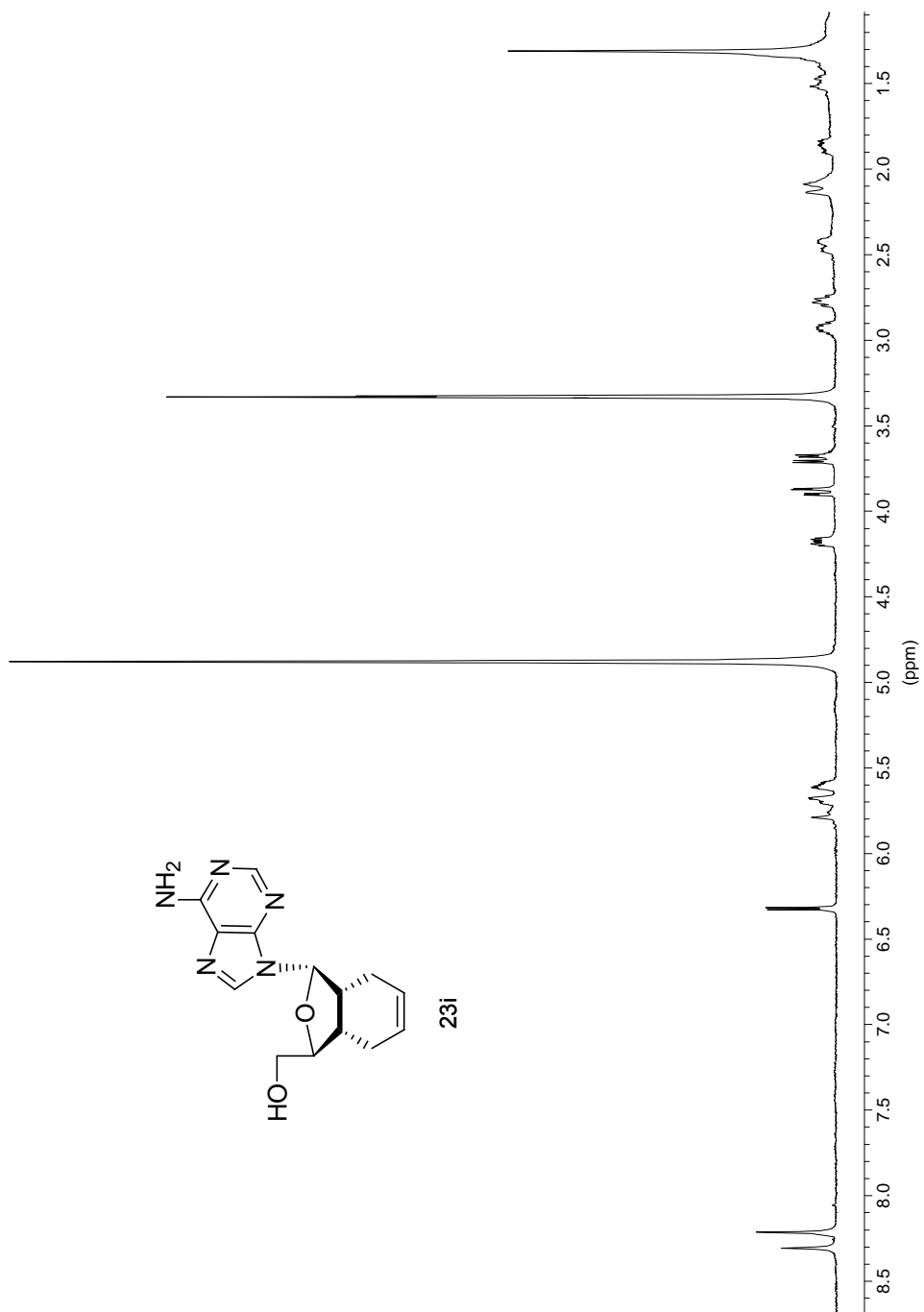
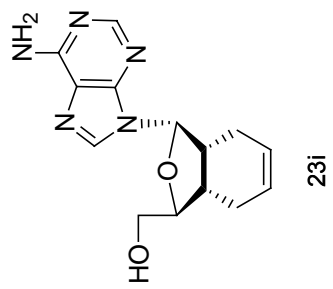


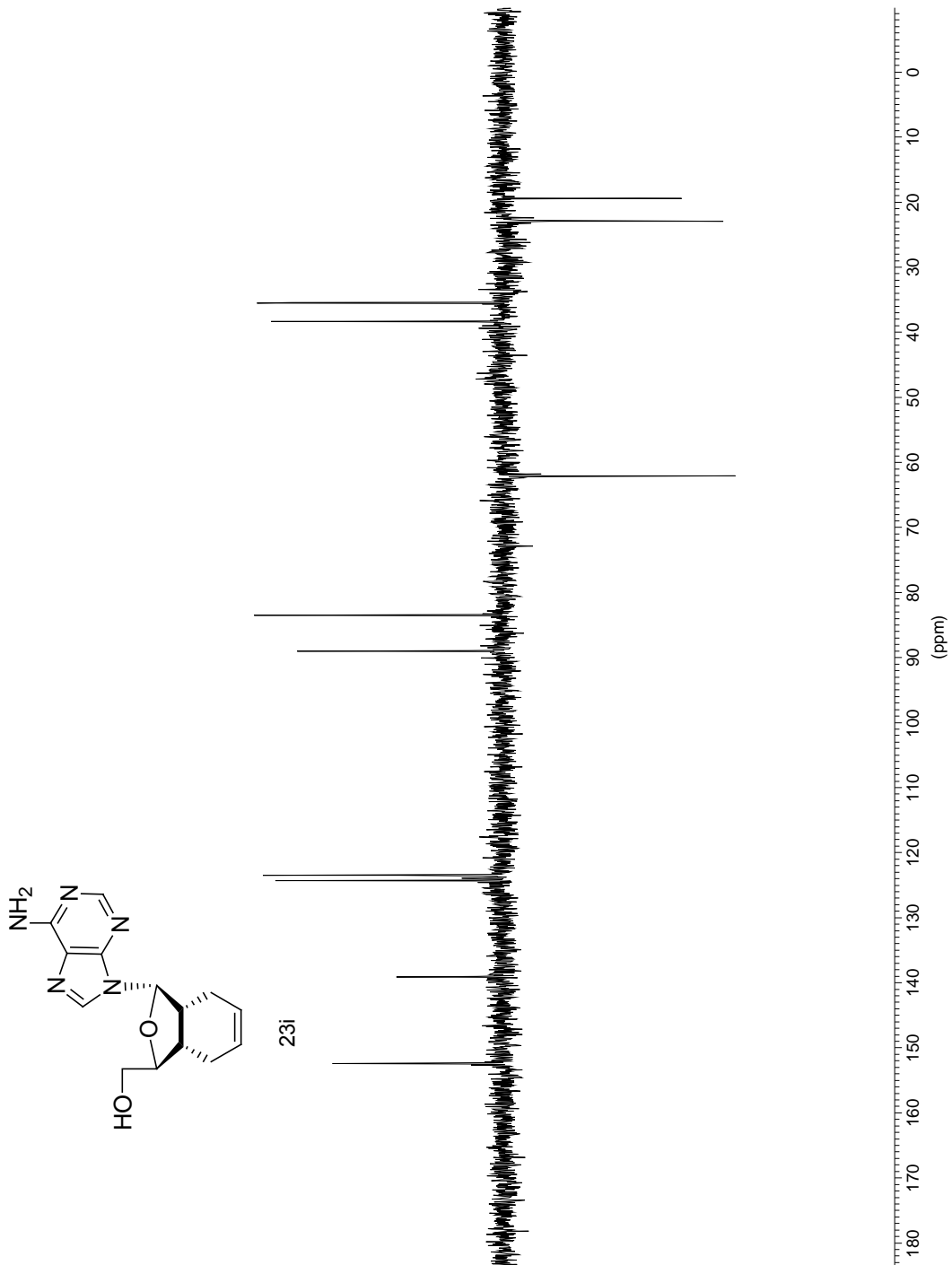
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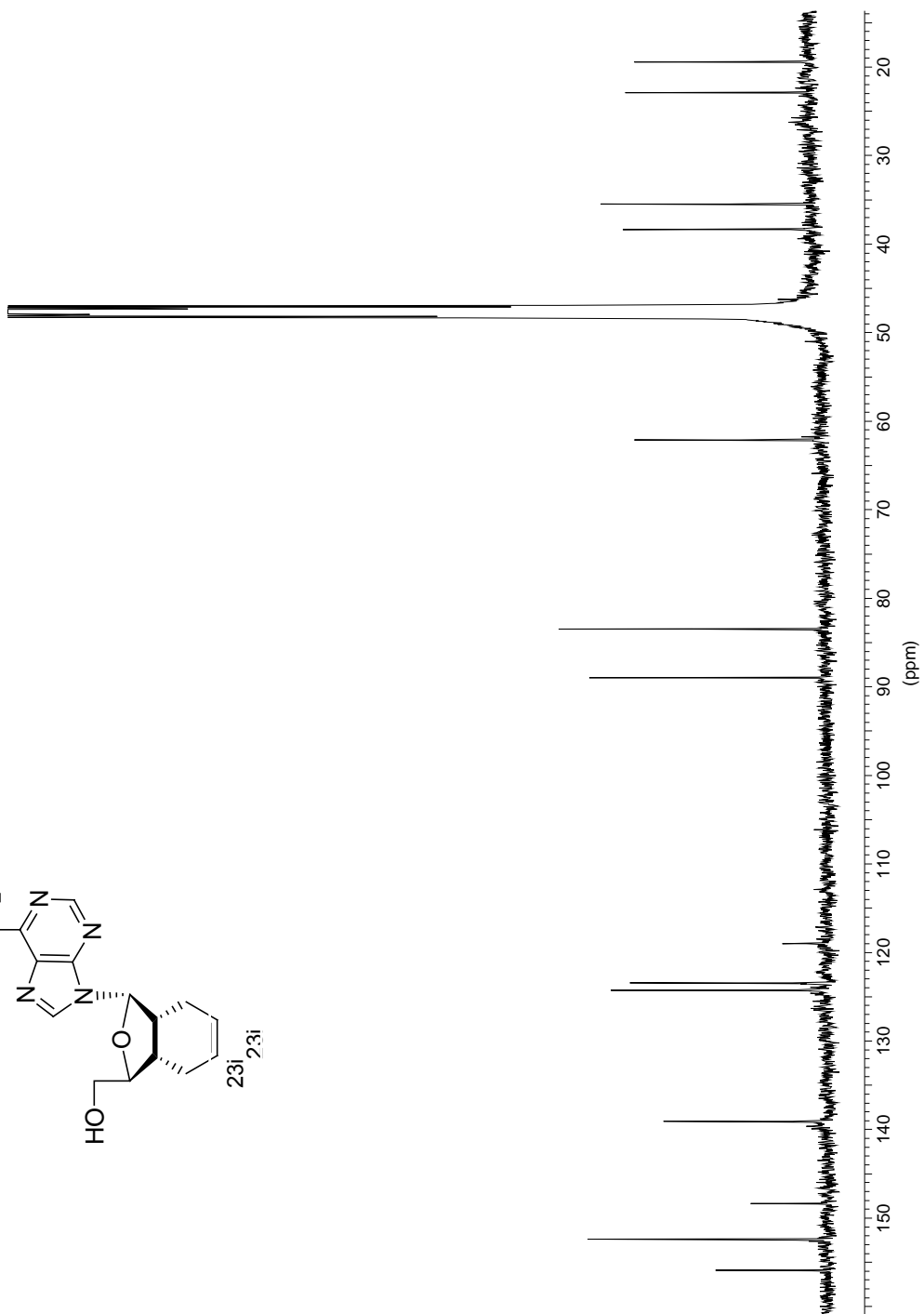
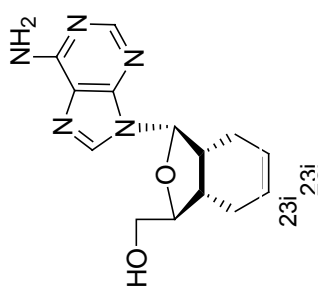


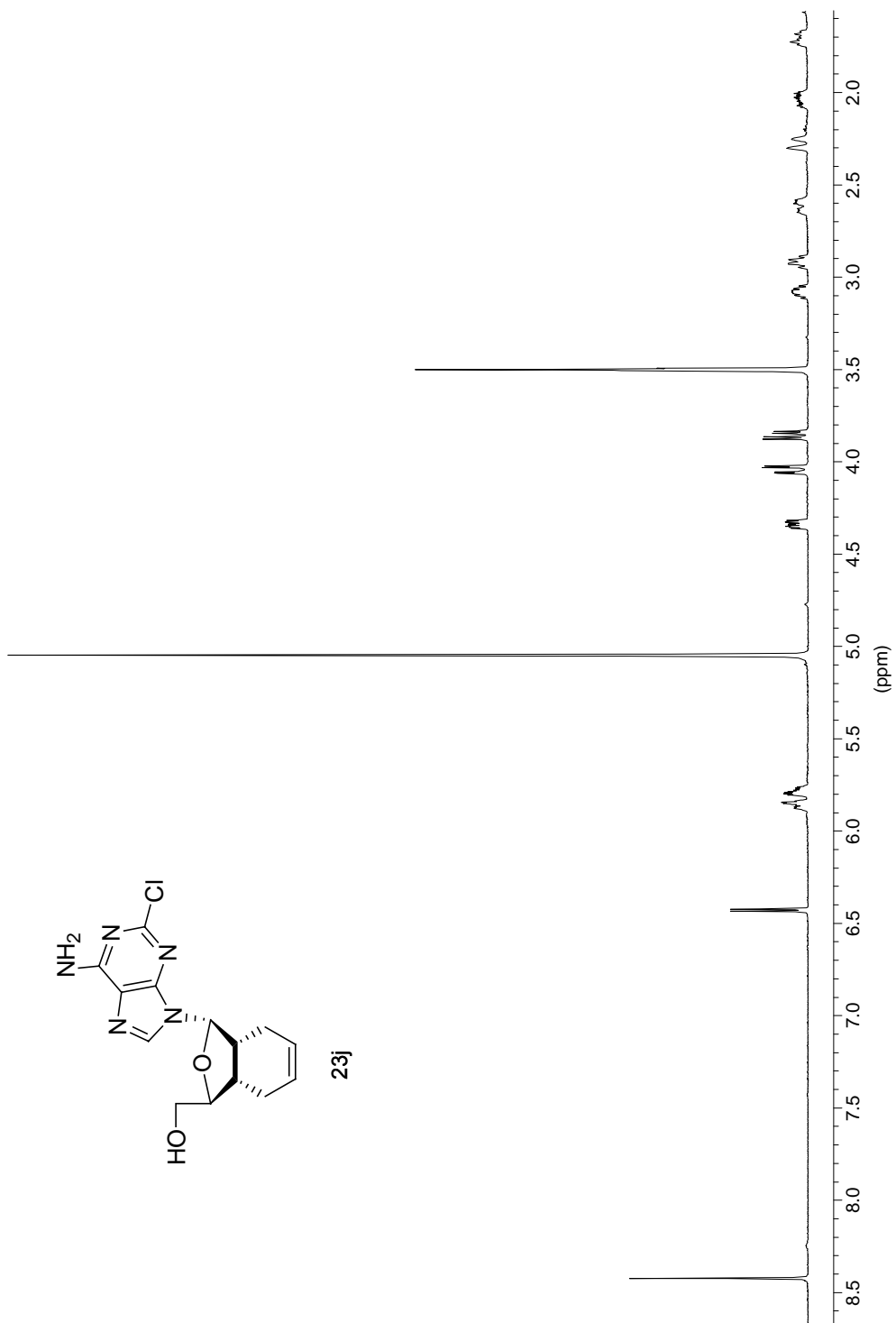
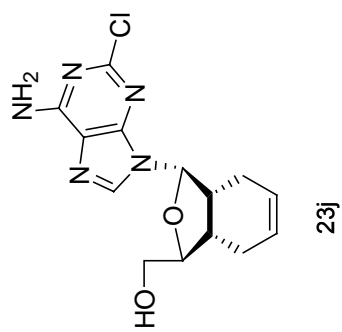


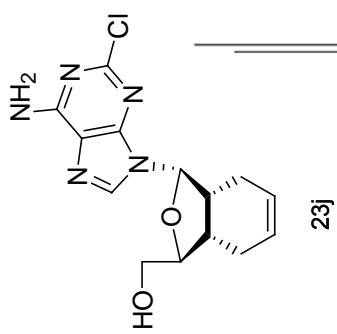




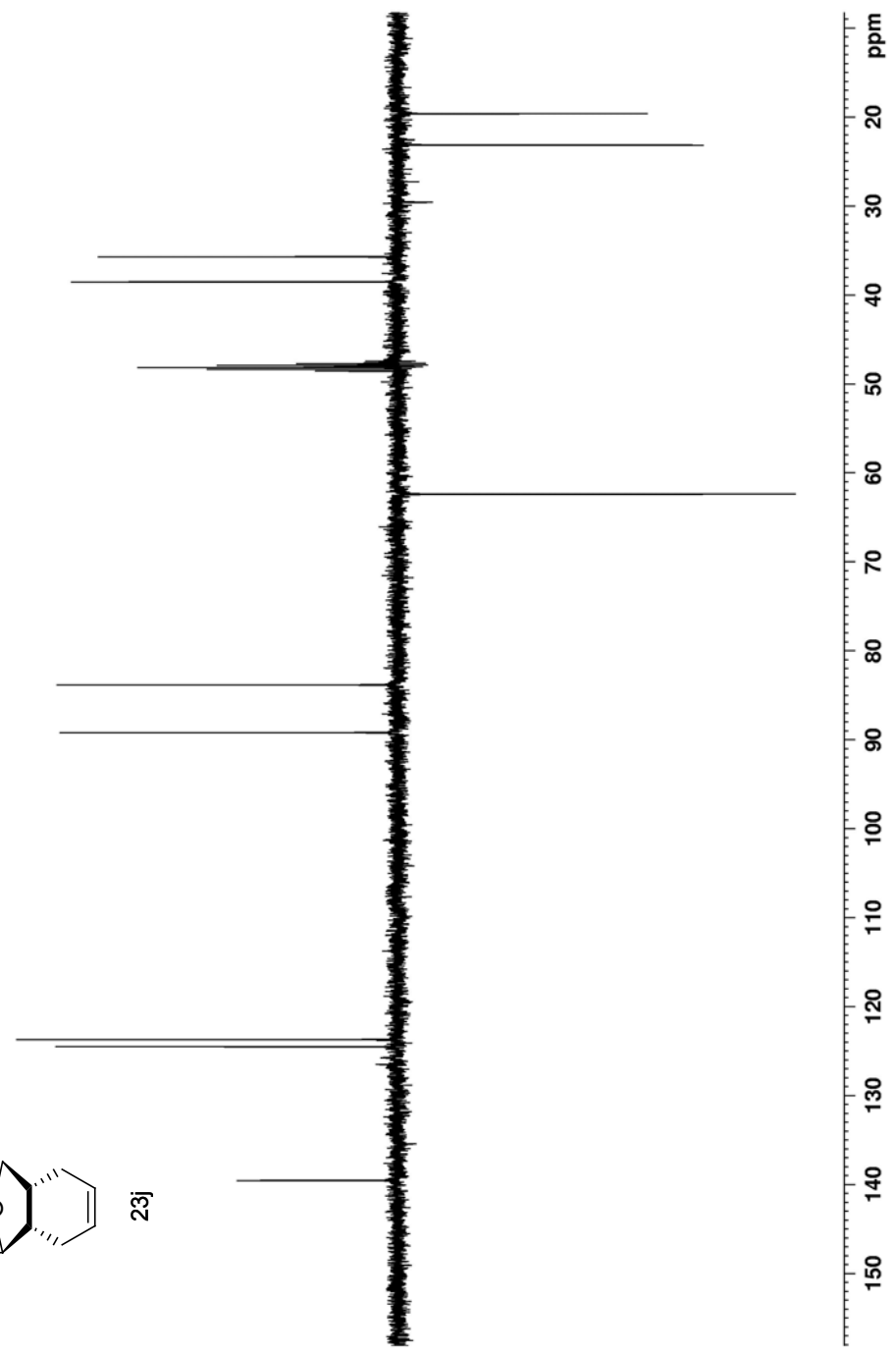


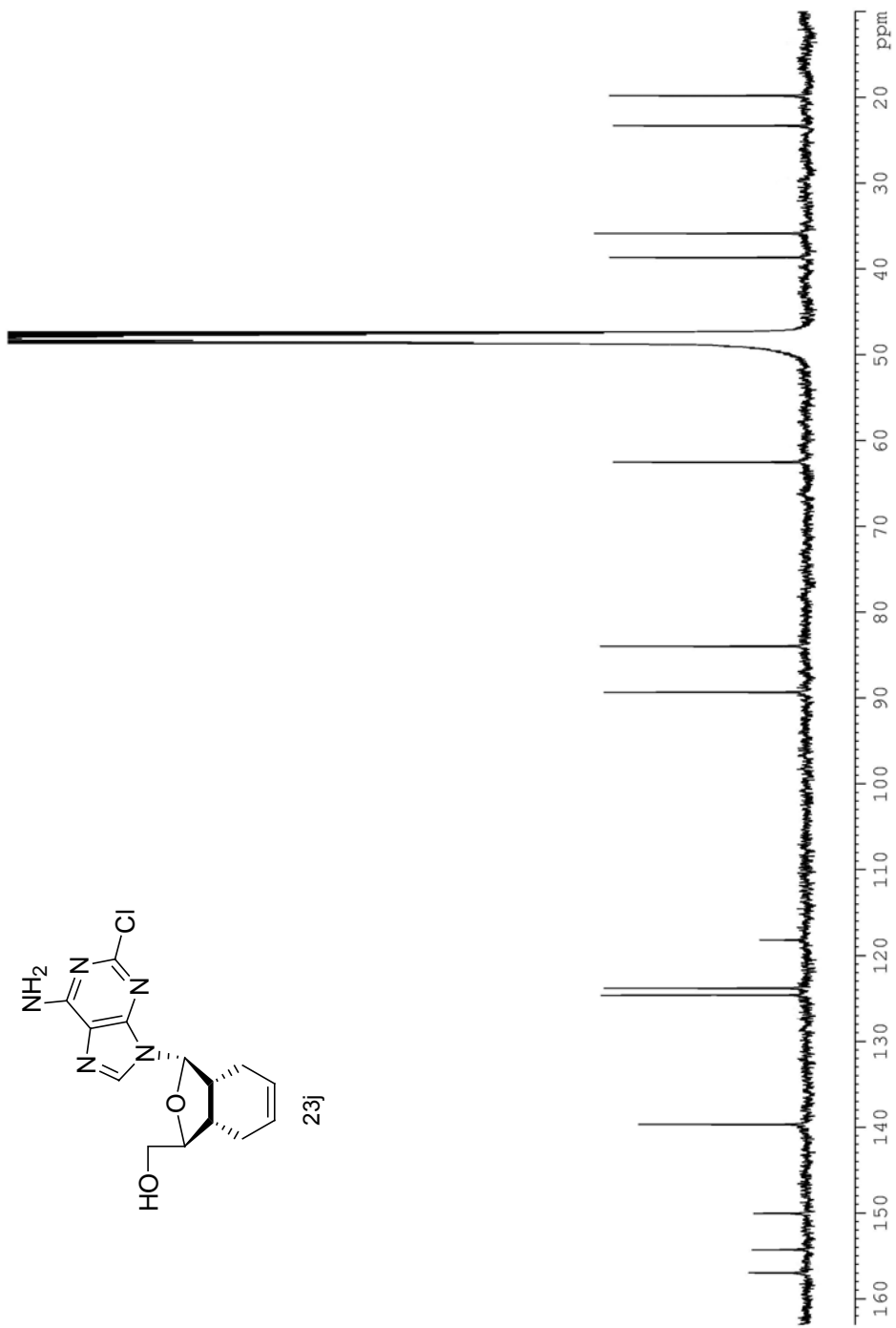
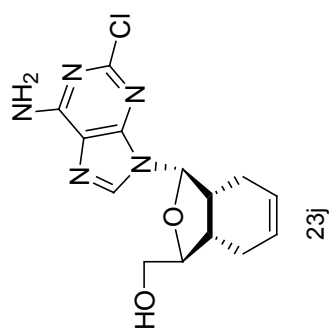


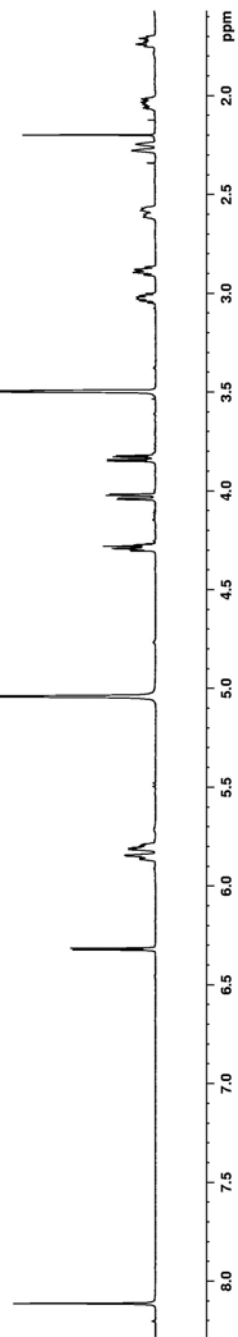
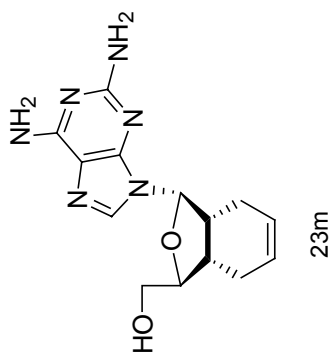


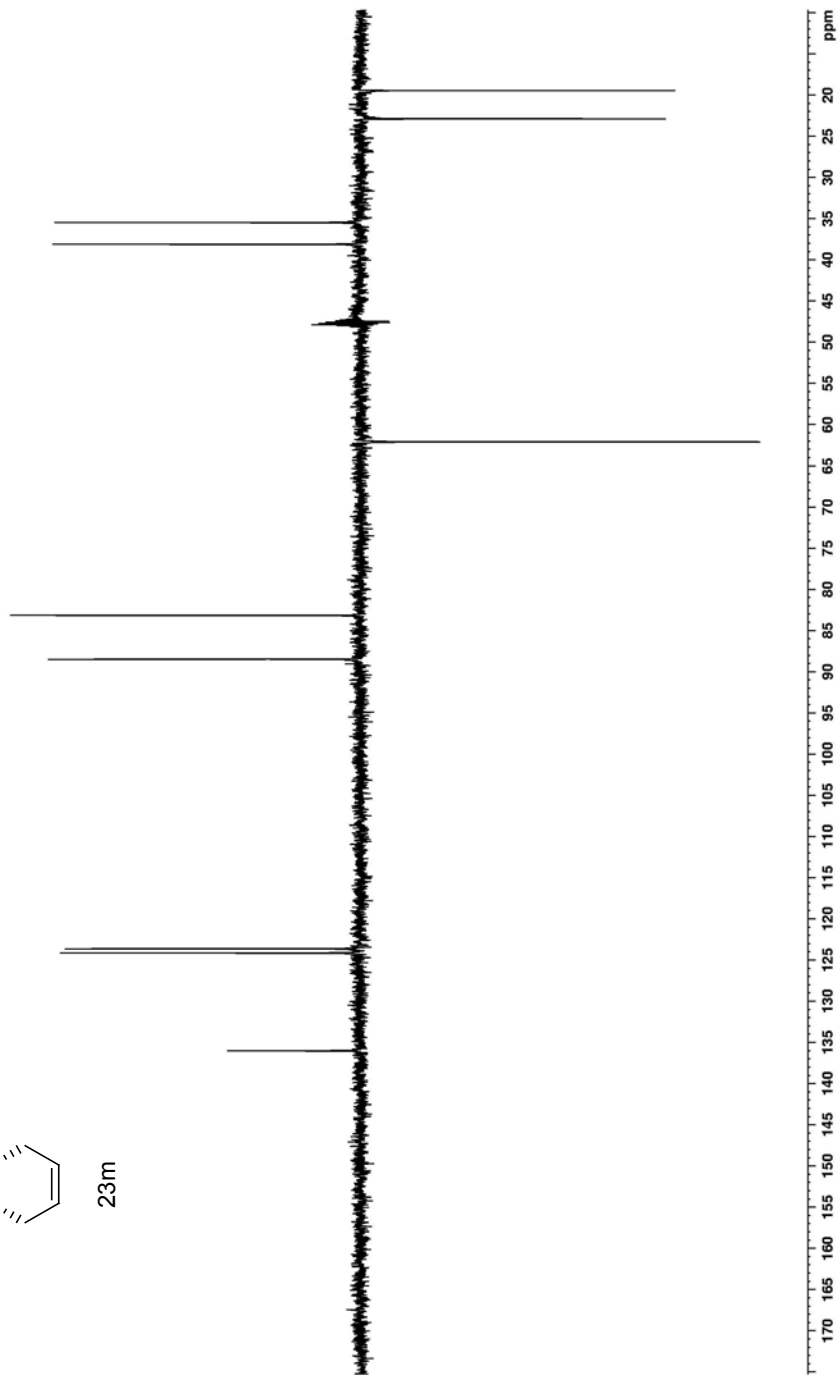
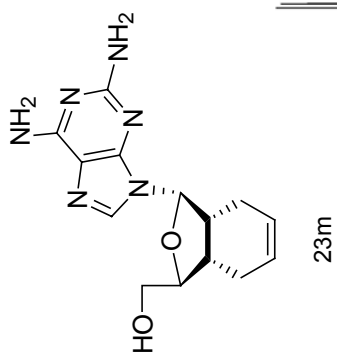


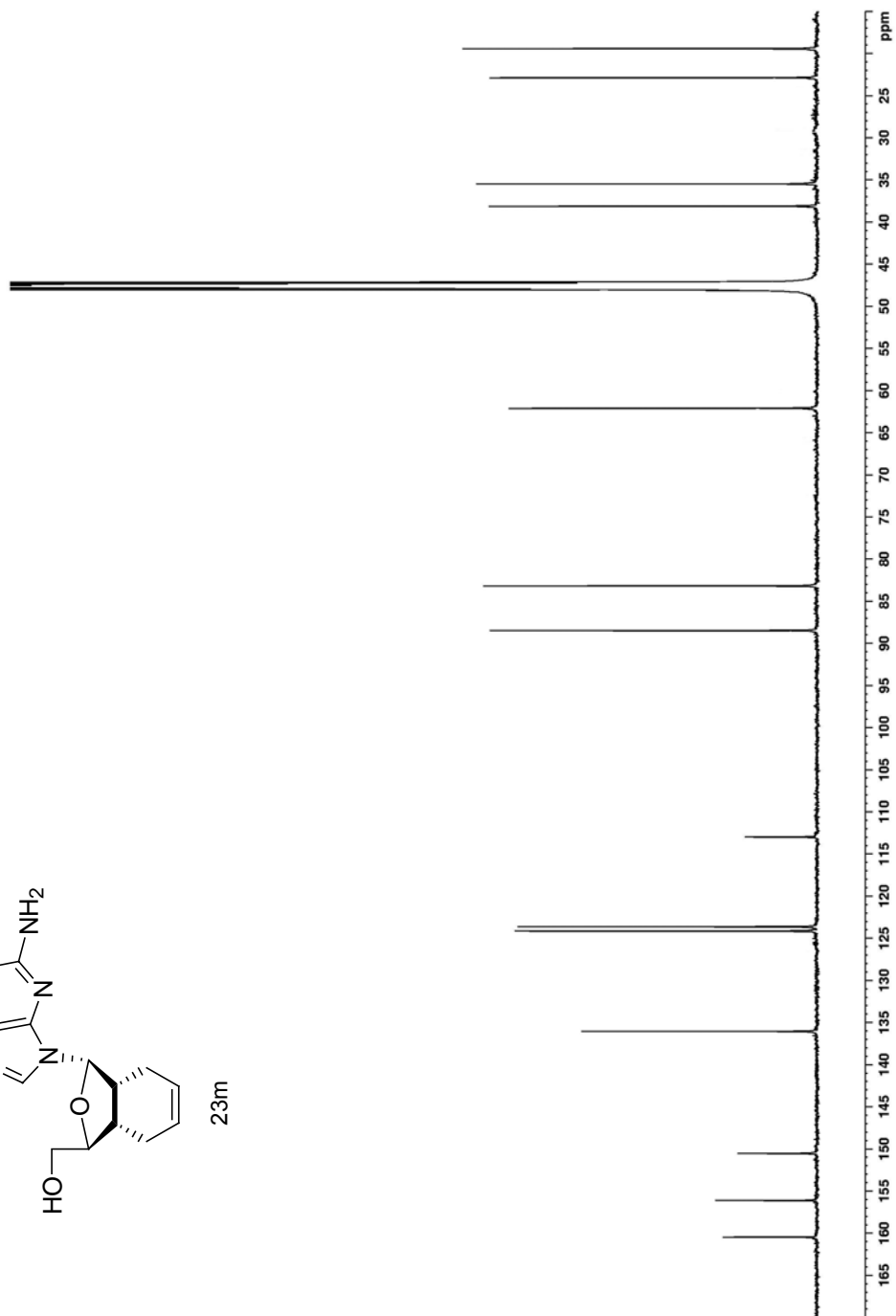
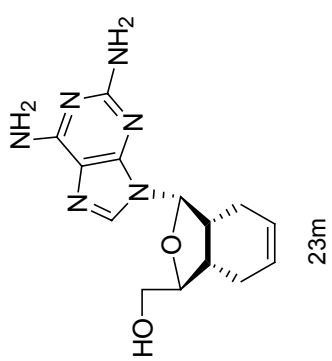
S113



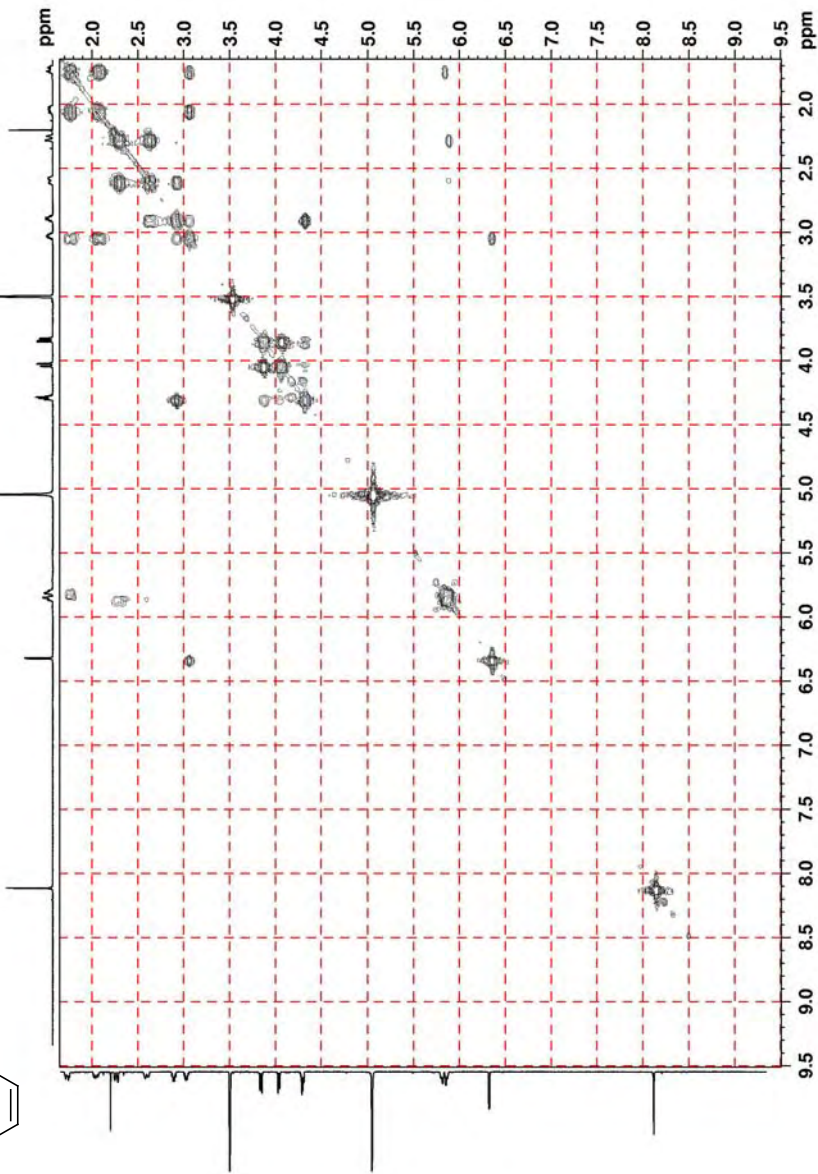
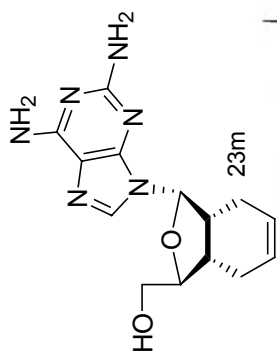


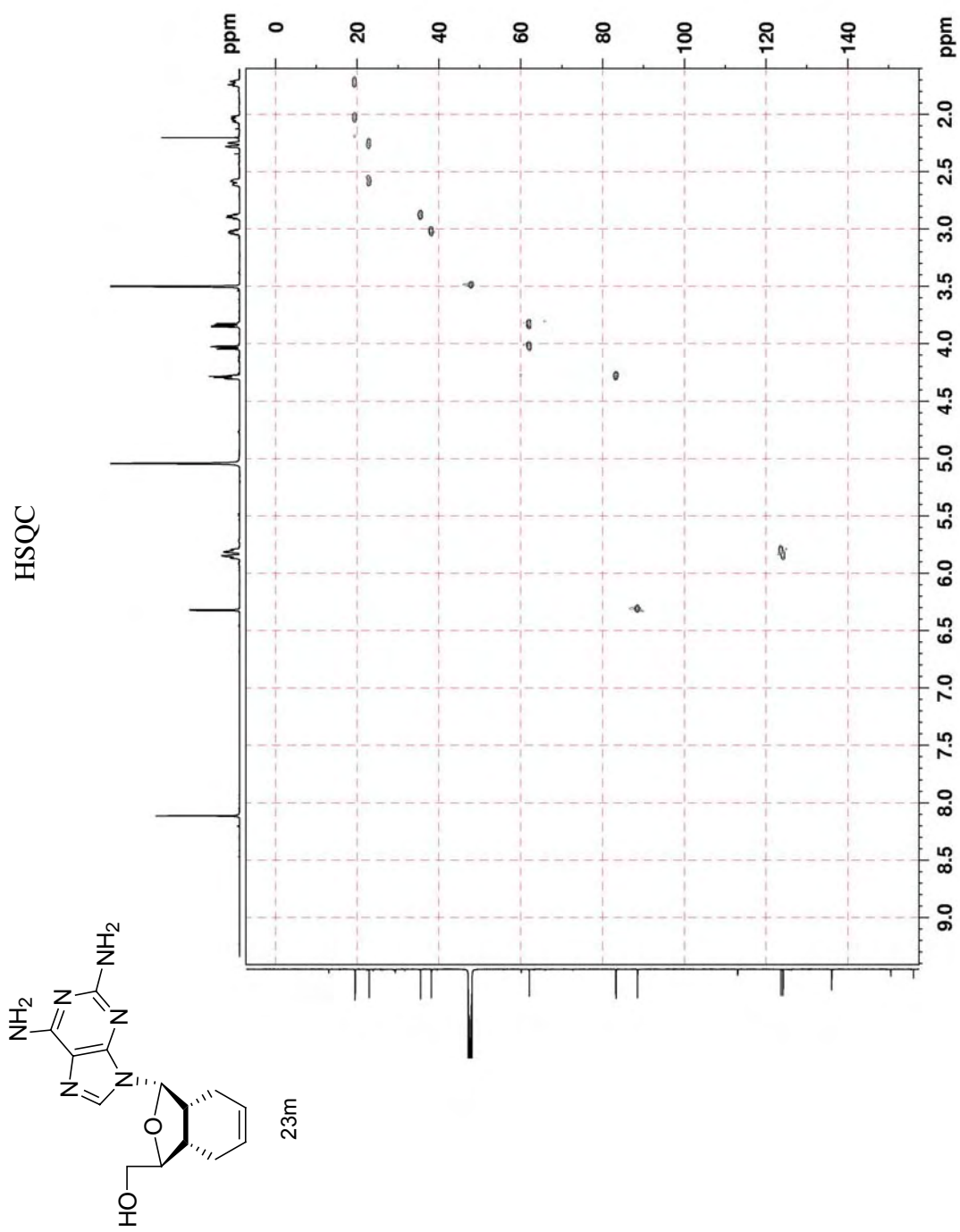


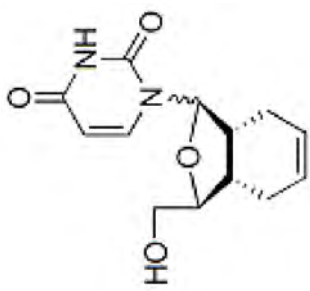




COSY

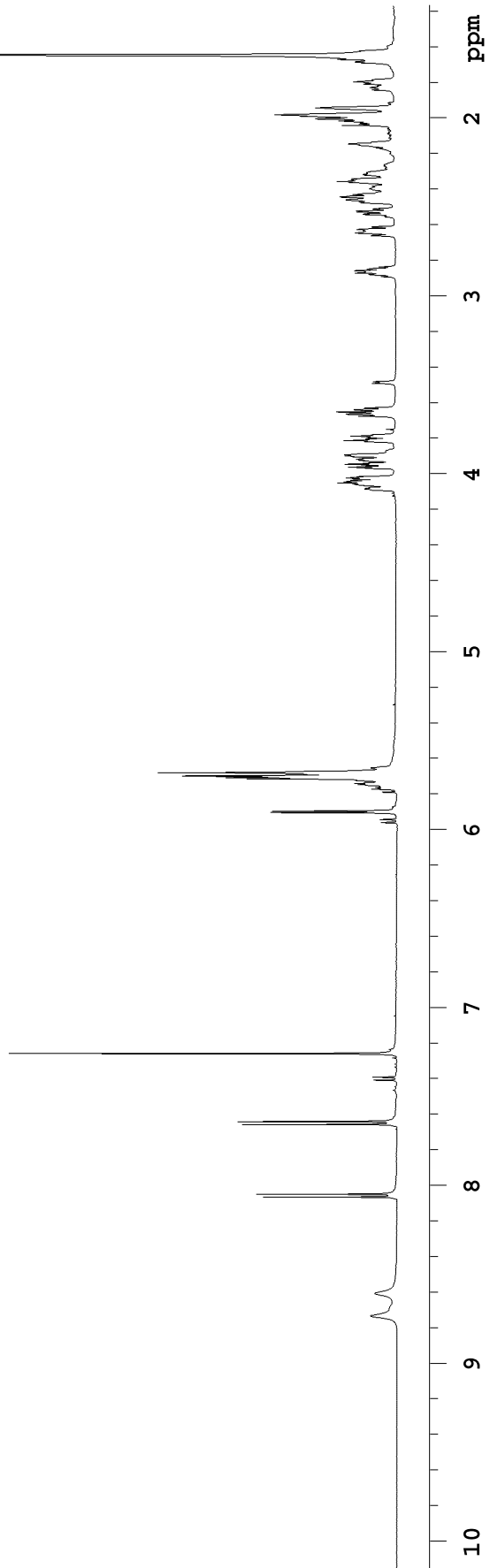


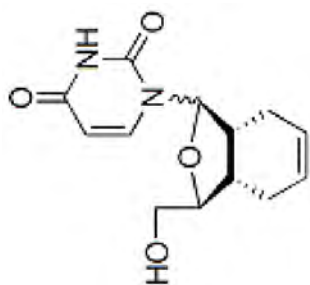




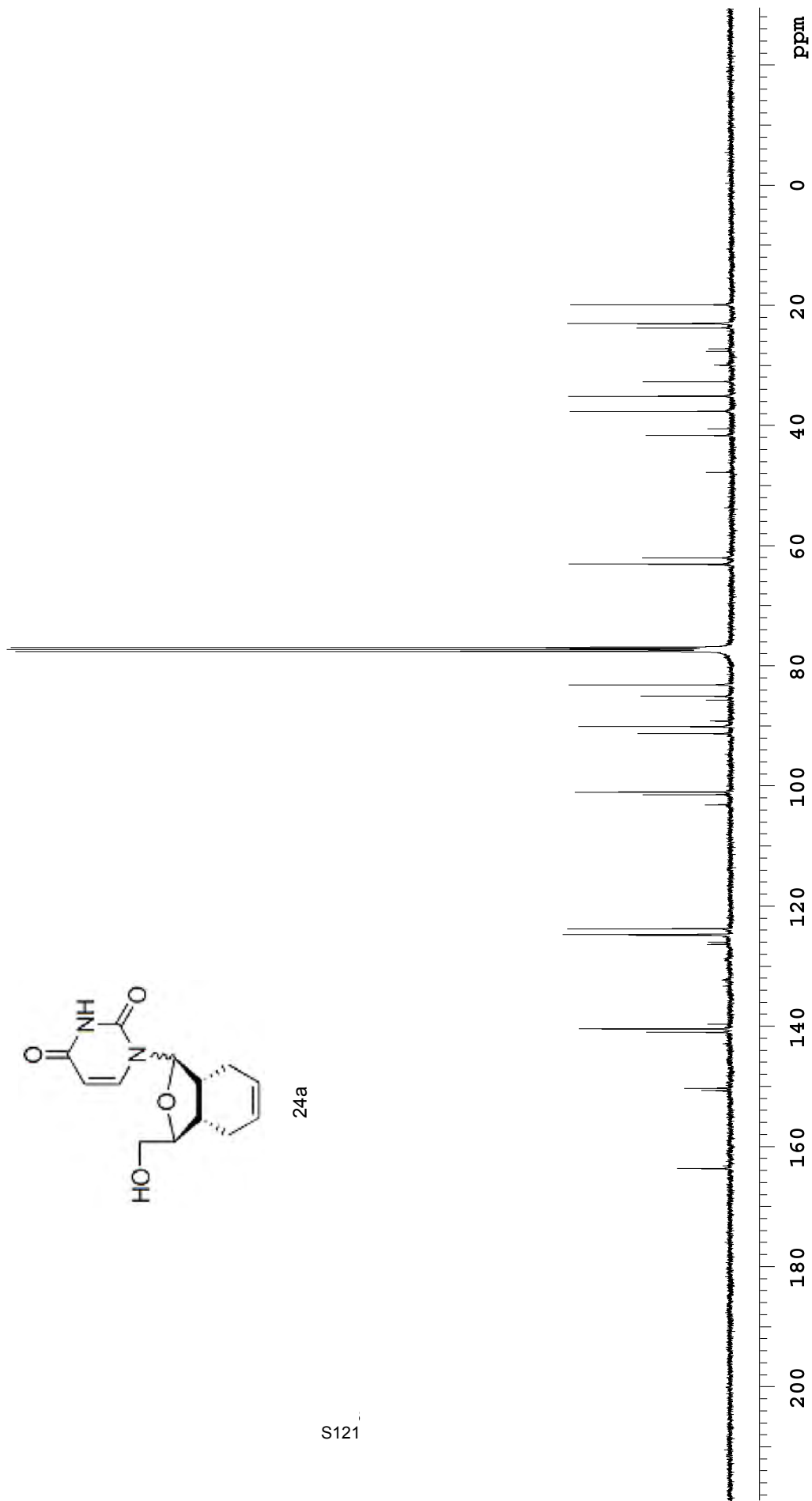
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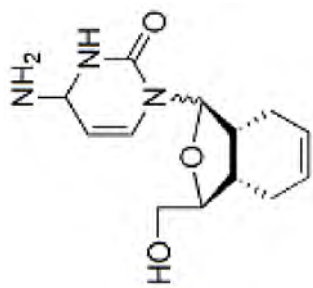
S120



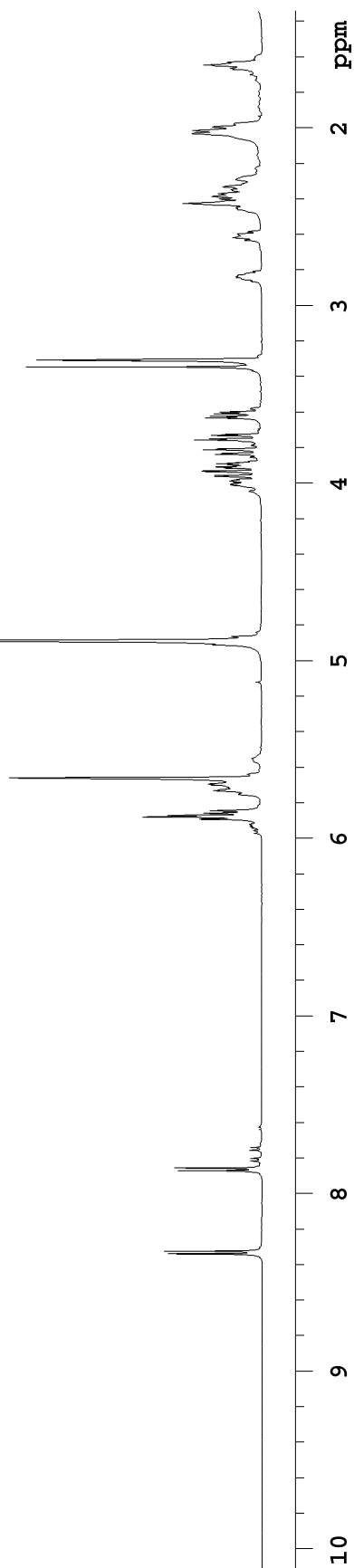


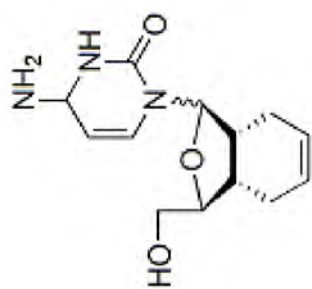
24a





24h





24h

