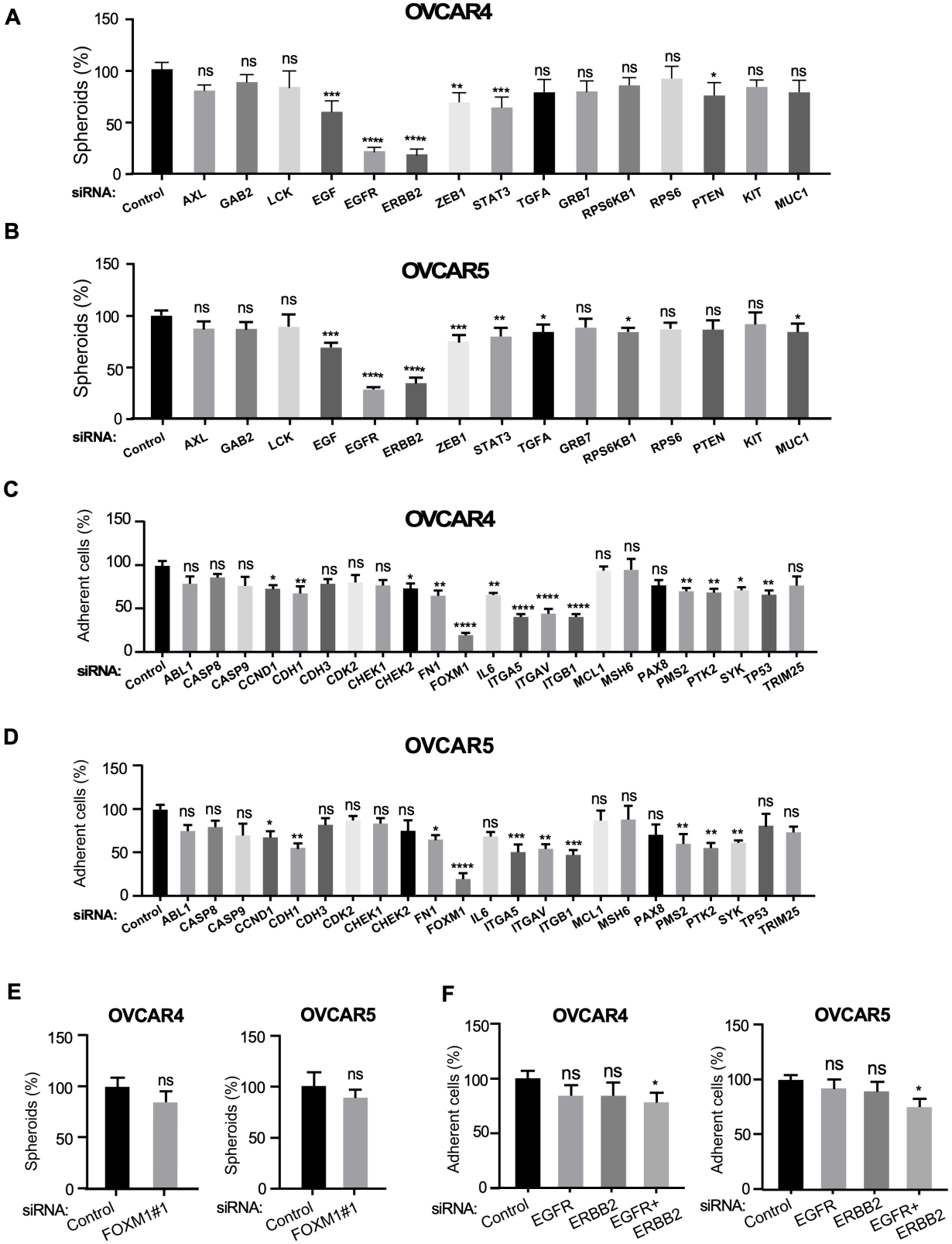
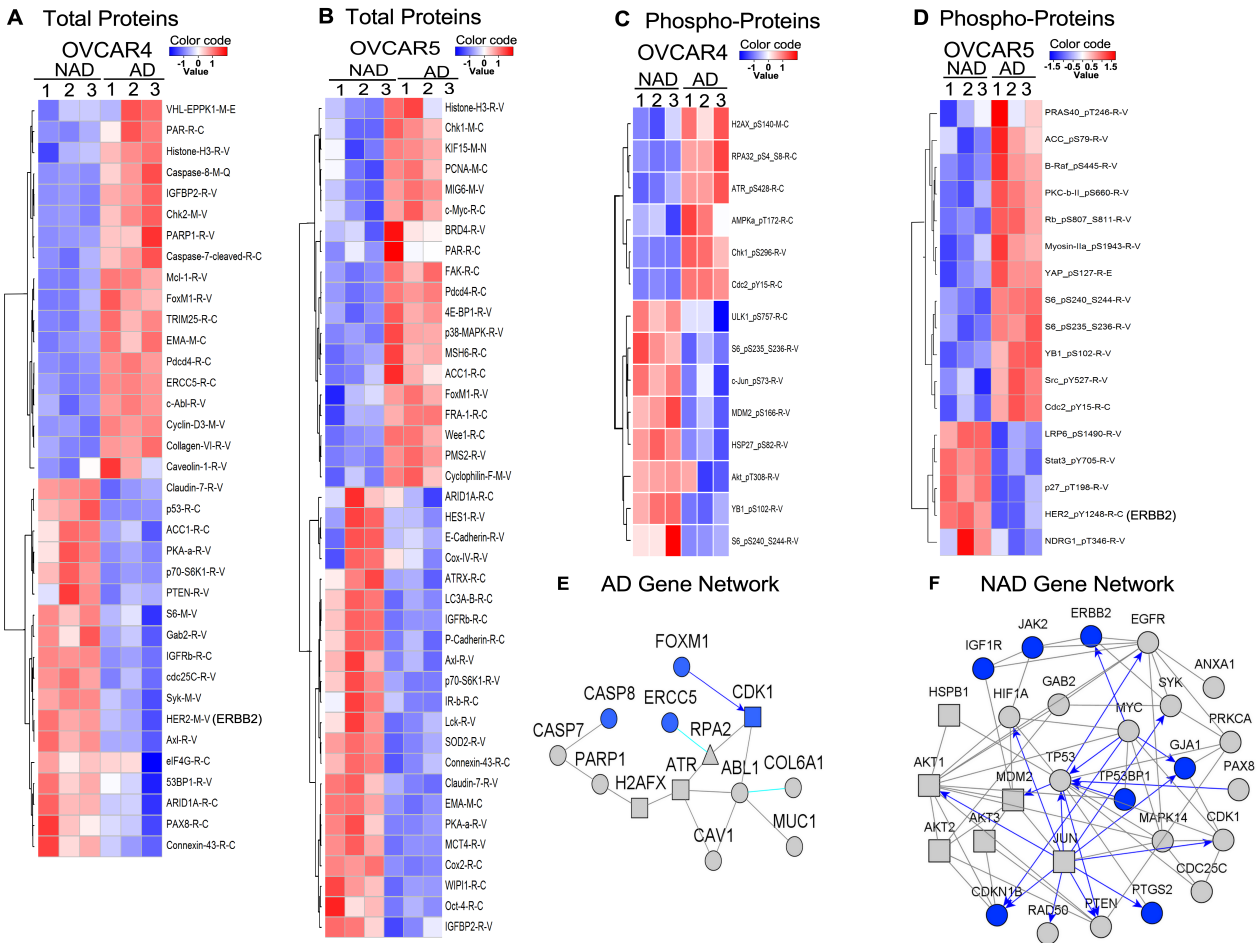
Supplementary Figures, Legends to Supplementary Figures and Supplementary Tables

Supplementary Figure-1



Supplementary Figure-1: A-B: OVCAR4 and OVCAR5 cells were transfected with control siRNA or siRNAs of all genes indicated NAD network in Fig-2C, then trypsinized 12h after transfection and cells seeded on non-adherent culture plate for 6 days and number of spheroids formed were quantitated. **C-D:** OVCAR4 and OVCAR5 cells were transfected with control siRNA or siRNAs of all genes indicated in the AD gene network in Fig. 2D. Cells were trypsinized 18h after transfection and 1000 cells from each group were plated on ~90% confluent mesothelial cells in three biological replicates in 96-well culture plate for 3h at 37ºC. Non-adherent cells were washed and removed and the fluorescent signaling intensity was determined by fluorimetry. **E**. FOXM1 was knocked down using target-specific siRNA in OVCAR4 and OVCAR5 cells and then plated for spheroid formation as described in A. **F.** EGFR, and ERBB2 were knocked individually or in combination using siRNAs and the cells were plated as in C and cell adhesion was determined.

Supplementary Figure-2

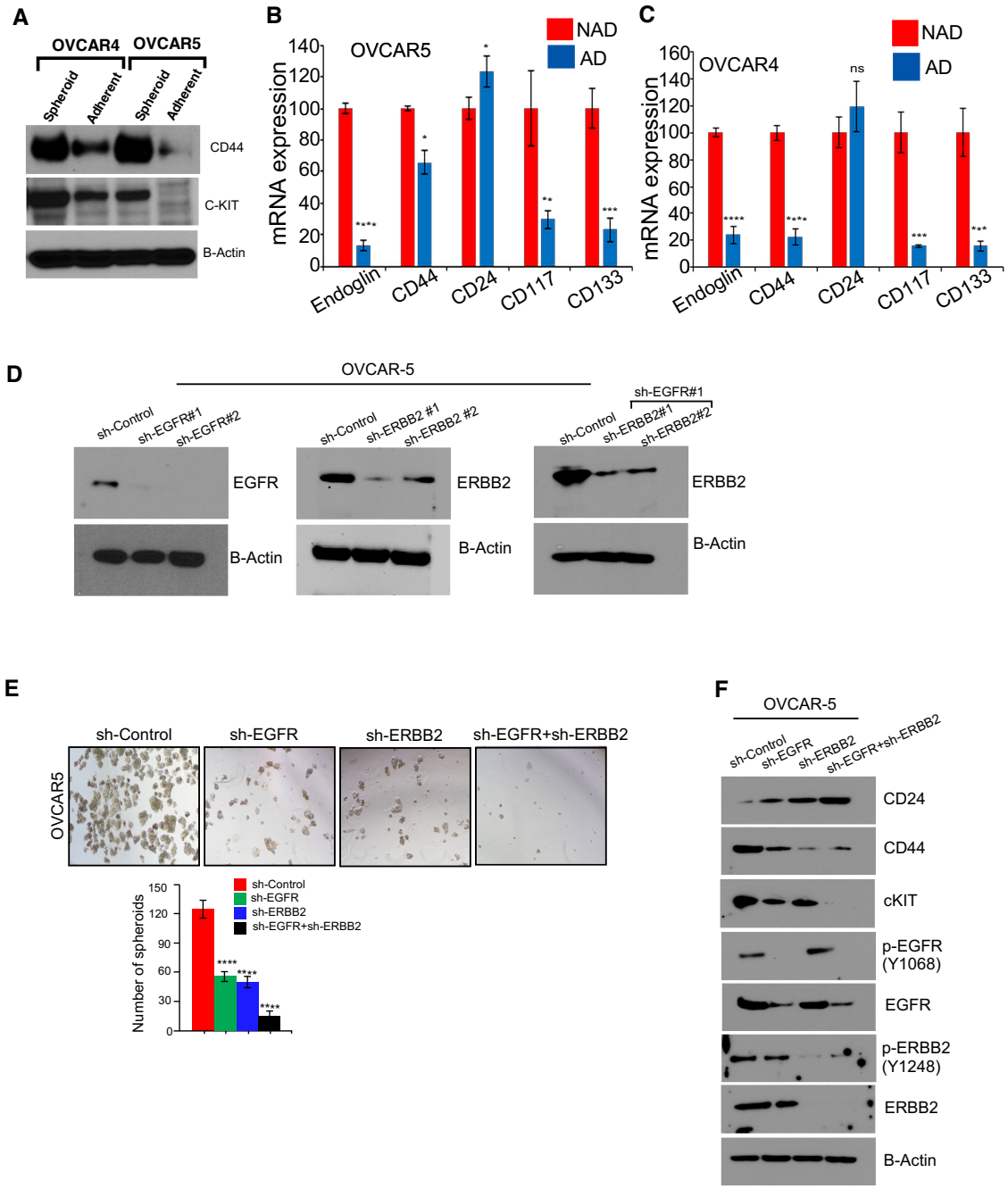


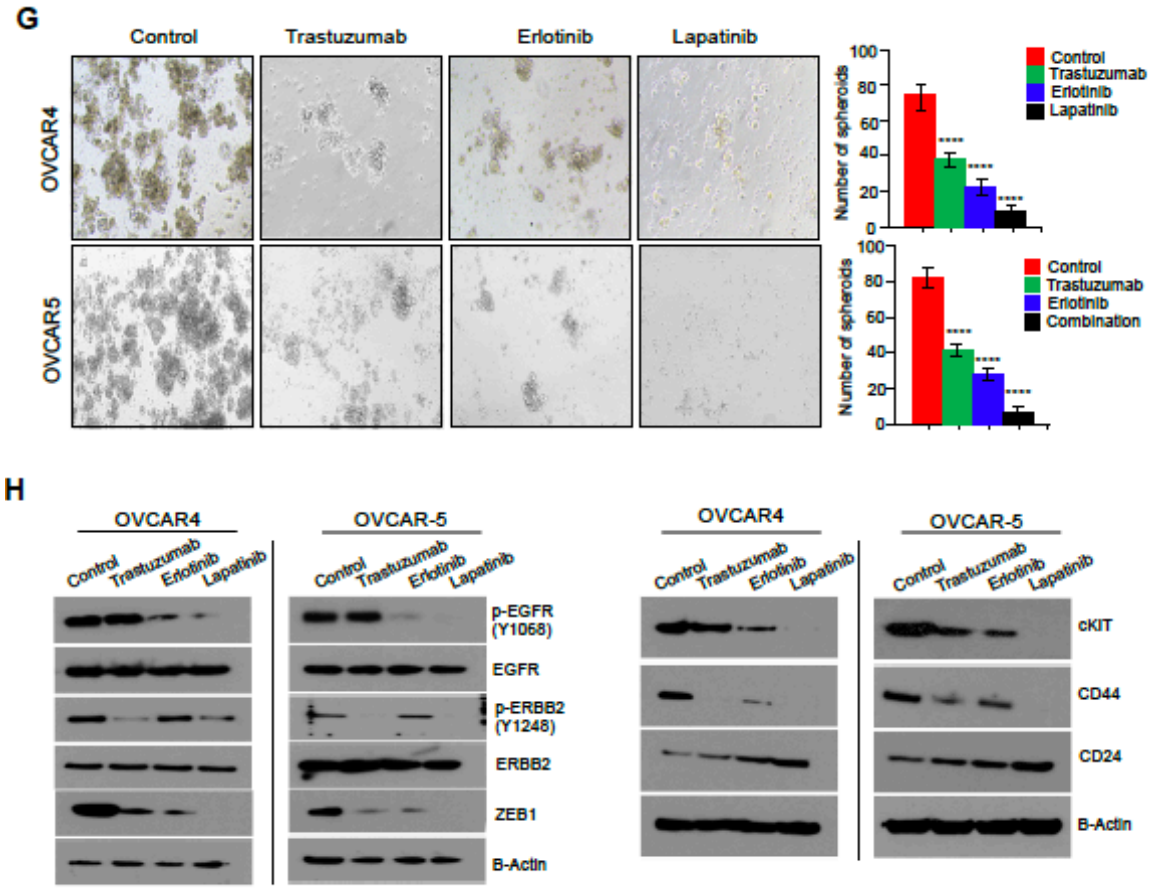
Supplementary Figure-2:

A-B, Adherent (AD) or non-adherent (NAD)cells of both OVCAR4 and OVCAR5 cell were grown on AD or NAD conditions for 24h. Tumor cell lysates were prepared and RPPA analysis was performed. Heat maps in A represents the total proteins altered with a log fold change ≥ 0.2 between AD versus NAD conditions. **C-D,** Heat maps represents the phospho-proteins altered with a log fold change ≥ 0.15 between AD versus NAD cells in triplicates. **E-F**, Protein network shows the connectivity between genes altered in AD or

NAD form of cells plotted using Netwalker. Circle indicate total protein; whereas the square indicates phospho proteins respectively. Triangle indicates the candidates are altered in both total and phospho form. Blue color nodes are altered in both cell lines. Grey shows the proteins altered in one cell line or the proteins identified for connectivity by Netwalker.

Supplementary Figure-3

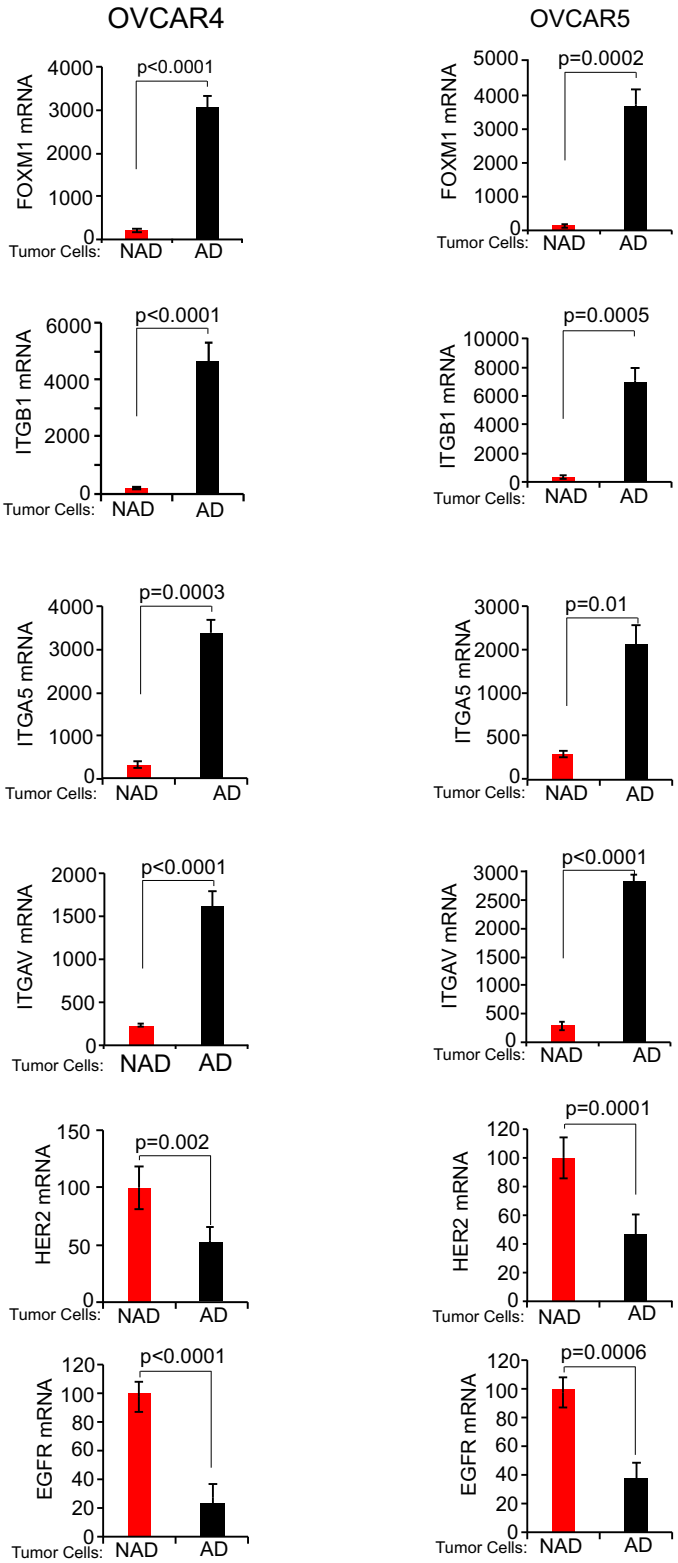




Supplementary Figure-3: A, Ovarian cancer cell lines OVCAR4 and OVCAR5 were cultured on adherent or non-adherent culture plate for 24h; then cell lysates were prepared and immunoblot was performed using antibodies indicated. **B-C,** OVCAR4 and OVCAR5 lines were grown on adherent and non-adherent conditions for 24h. Cell lysate was prepared, mRNA isolated and qPCR was performed to determine the expression of the genes indicated. Values were presented as mean ± SE of triplicates and p-values were determined by the Student's t-test. *p<0.05, **p<0.01, ***p<0.001 and ****p<0.0001. **D,** OVCAR5 cells were transfected with lentiviral particles encodes either control shRNAs, or two different shRNAs of EGFR or ERBB2. Stable knockdown of EGFR clone#1 was selected and infected with shERBB2 encoding viral particle to make stable knocked down

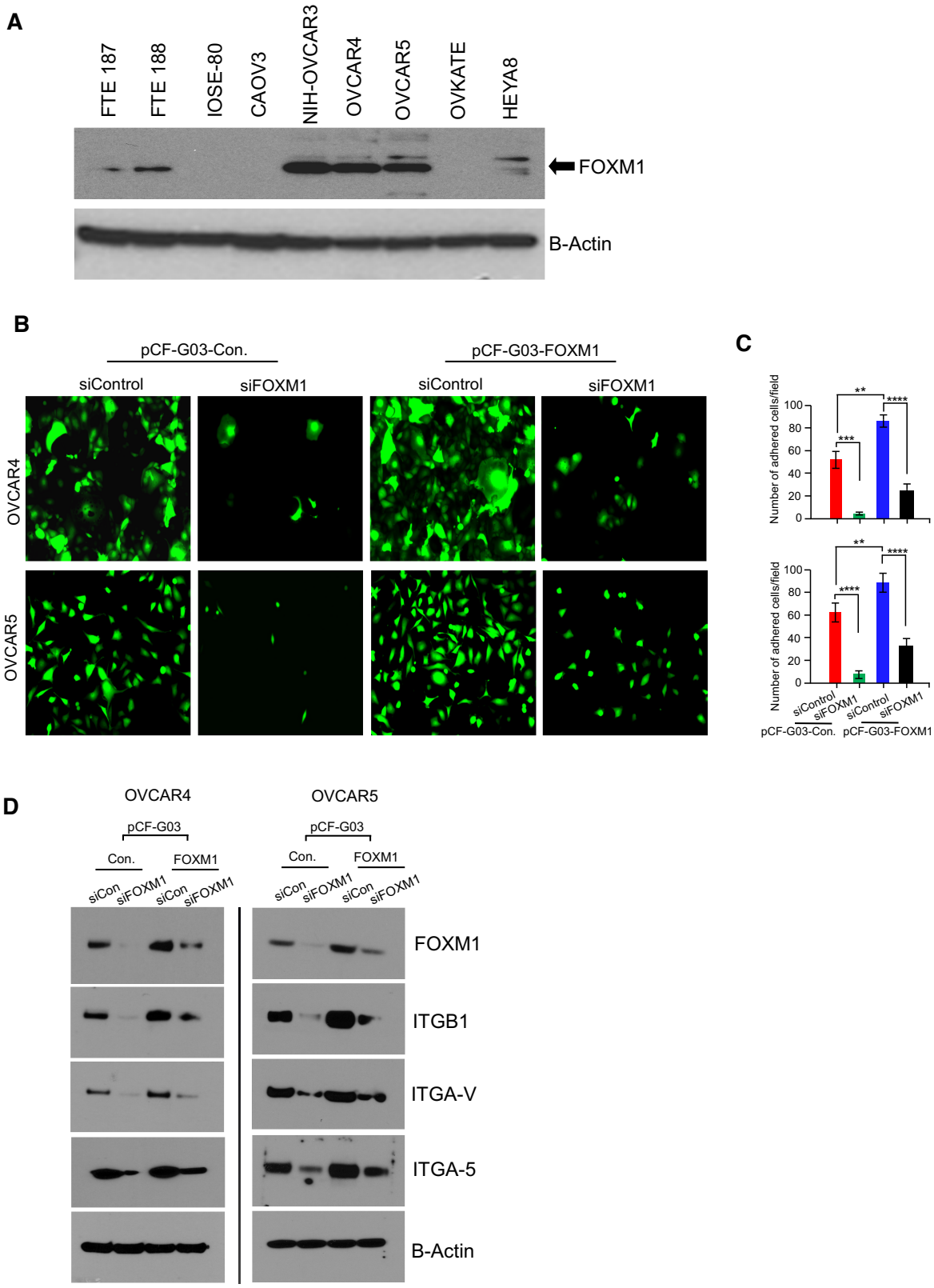
of EGFR or ERBB2. Cell lysates were prepared and immunoblotted using the antibodies indicated. **E,** OVCAR5 cells were selected for EGFR, ERBB2 or both EGFR and ERBB2 from (D) were plated on low-attachment plates and the number of spheroids were quantitated after two-weeks **F,** Tumor spheroids from (E) were lysed and immunoblotted using the antibodies indicated. **G.** OVCAR 4 and OVCAR 5 spheroids were grown in the presence of Trastuzumab (5 μg/mL), Erlotinib (4 μM), or Lapatinib (4 μM) for 7 days. 3D-spheroids were photographed, and the number of spheroids were quantitated. **H,** Tumor spheroids from (G) were lysed and immunoblotted using the antibodies indicated. β-actin was used as a loading control.

Supplementary Figure-4



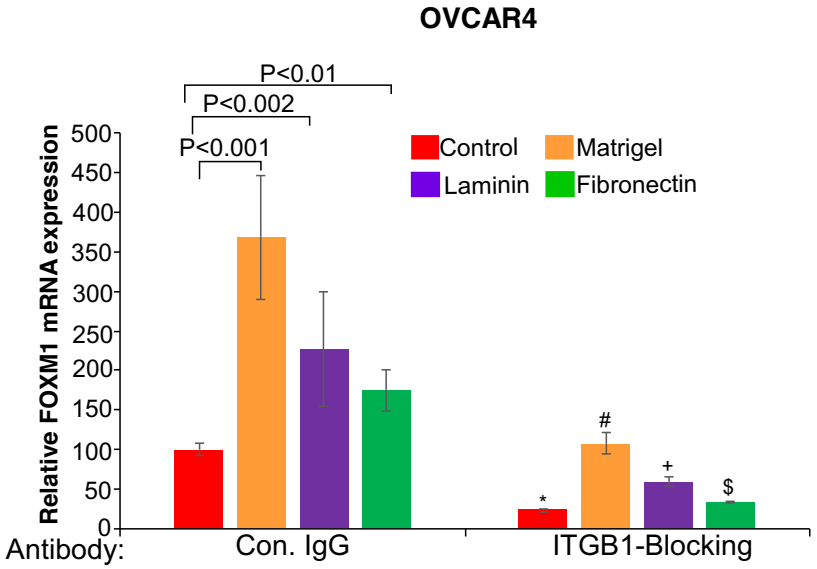
Supplementary Figure-4: OVCAR4 or OVCAR5 cells were injected ($1X10^6$ cells/mice) injected intraperitoneally into the nude mice. Adherent tumor cells were collected from omentum and non-adherent cells in the peritoneum were collected by peritoneal wash using sterile PBS. Tumor cells were lysed, total RNA was isolated, and qPCR was performed. β-actin was used as the internal standard and the values were presented as mean ± SE of triplicates and p-values were determined by the Student's t-test.

Supplementary Figure-5



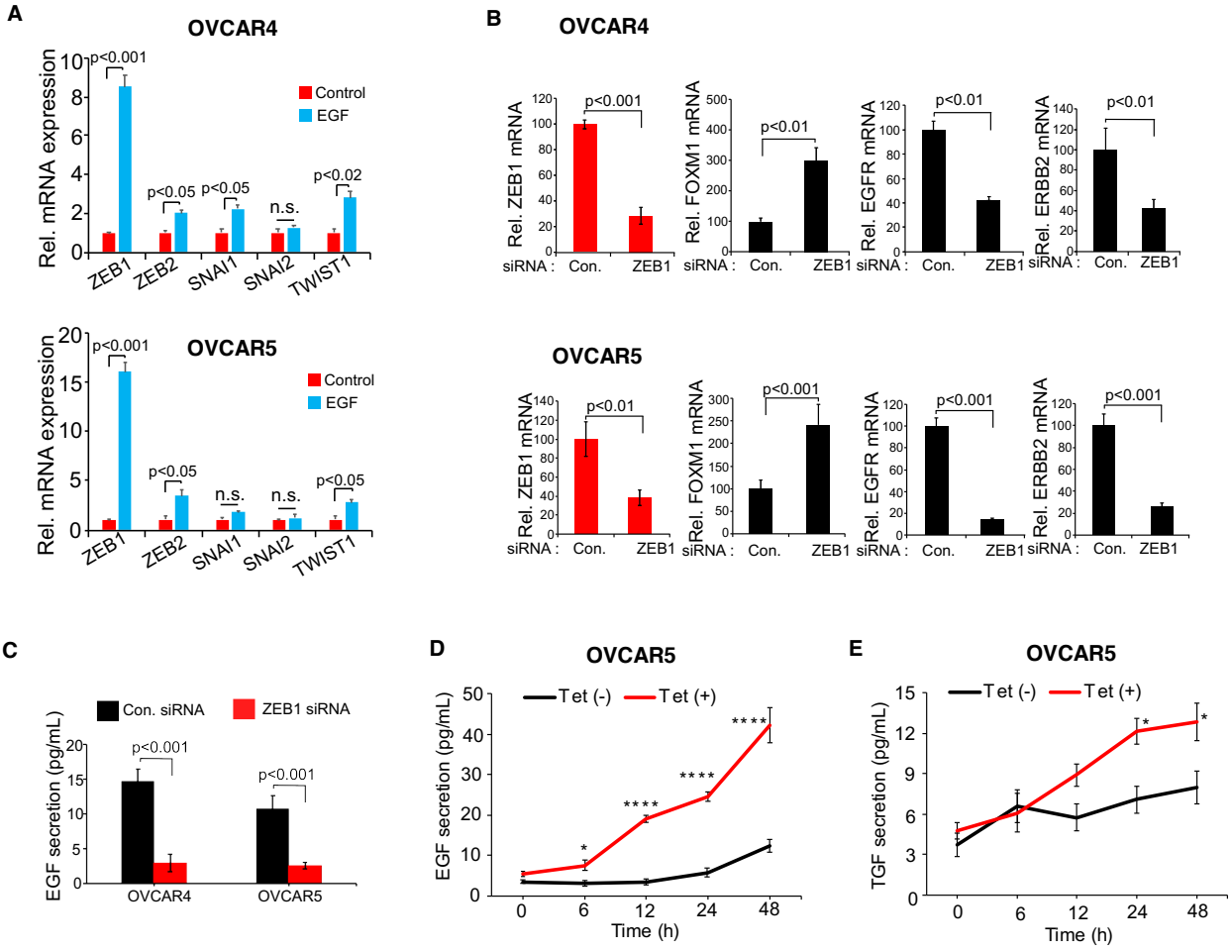
Supplementary Figure-5: A, Ovarian cancer cell lines and fallopian tube epithelial cells were lysed and immunoblotted using indicated antibodies. **B-C**. pCF-G03-control vector or pCF-G03-FOXM1 plasmid were transfected in OVCAR4 and OVCAR5 cells stably expressing for 24h, which were pre-transfected with either con siRNAs or FOXM1 siRNAs for 12h. Cells were trypsinized from above and plated on culture plates were coated with extracellular matrix cocktail for 4h for cell adhesion. Non-adherent cells were removed by gentle washing using PBS and number of adherent cells were photographed and quantitated. **D**, Immunoblot was performed using the lysates prepared from (B) with indicated antibodies.

Supplementary Figure-6



Supplementary Figure-6: Ovarian cancer cells were trypsinized and incubated with control IgG or with integrin beta-1 blocking antibody for 1h, then plated on matrigel, Laminin or Fibronectin coated plate. Adherent cells were isolated 3h after adhesion and FOXM1 mRNA expression was determined by qPCR.

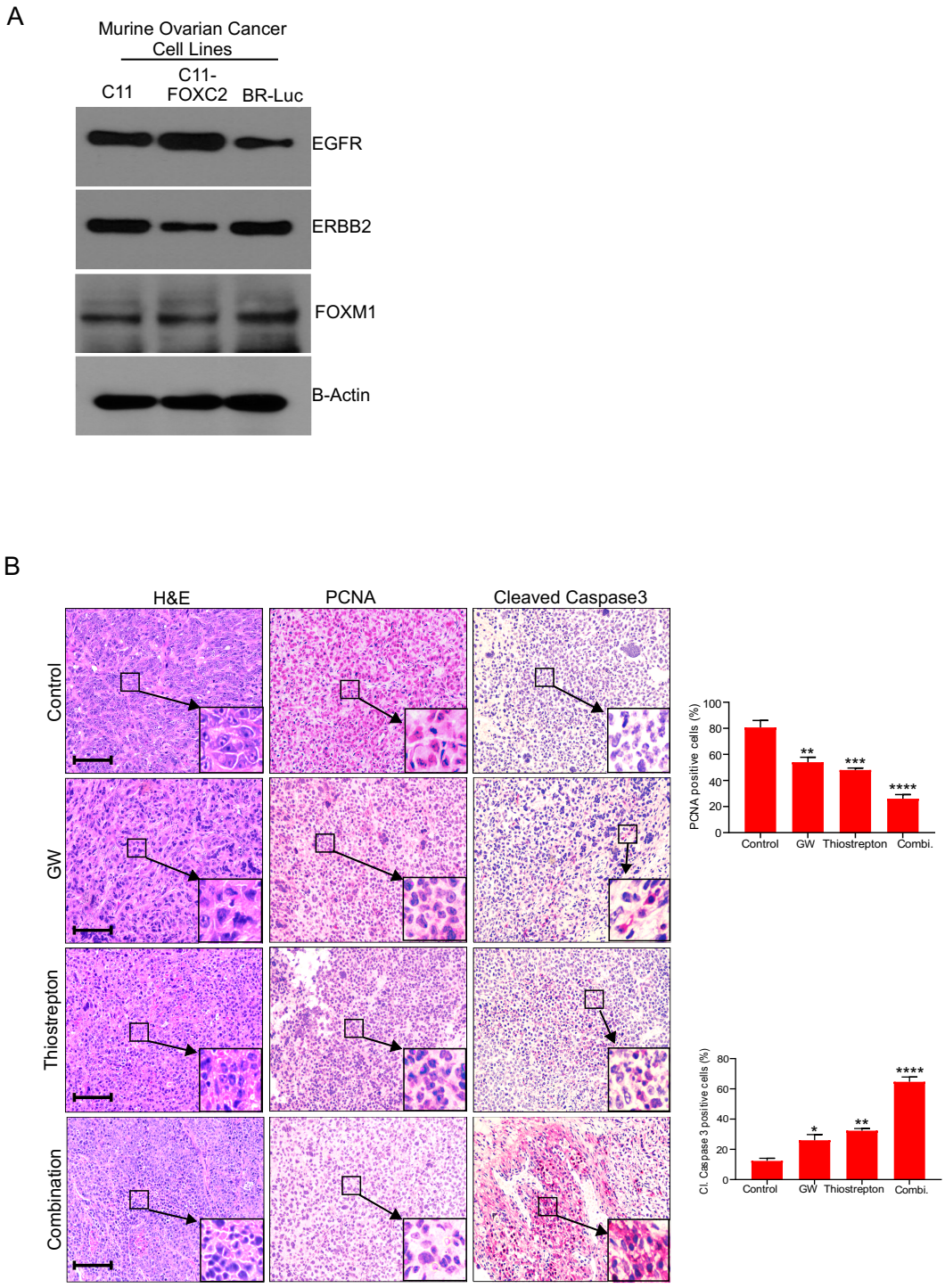
Supplementary Figure-7



Supplementary Figure-7: A, Ovarian cancer cells OVCAR4 were stimulated with EGF and determined the expression of EMT inducing transcription factors by qPCR. **B**, OVCAR4 and OVCAR5 cells were transfected with control or ZEB1 siRNAs, then mRNA was collected 24h after transfection and qPCR was performed to determine the expression of indicated genes. **C,** OVCAR4 and OVCAR5 cells were transfected with control or ZEB1 siRNAs, then culture supernatant was collected after 24h after transfection and ELISA was performed to detect the levels of EGF **D-E.** Culture supernatant were collected from the OVCAR5 cells stably express inducible ZEB1 after the treatment of tetracycline (1µg/ml) at the indicated time points and performed ELISA

to detect the levels of EGF and TGF-α. Results are from triplicate experiments and error bars represent SE. Significance was calculated by Student's t test by comparing the groups in the same time point. *$p<0.01$, ****$p<0.0001$

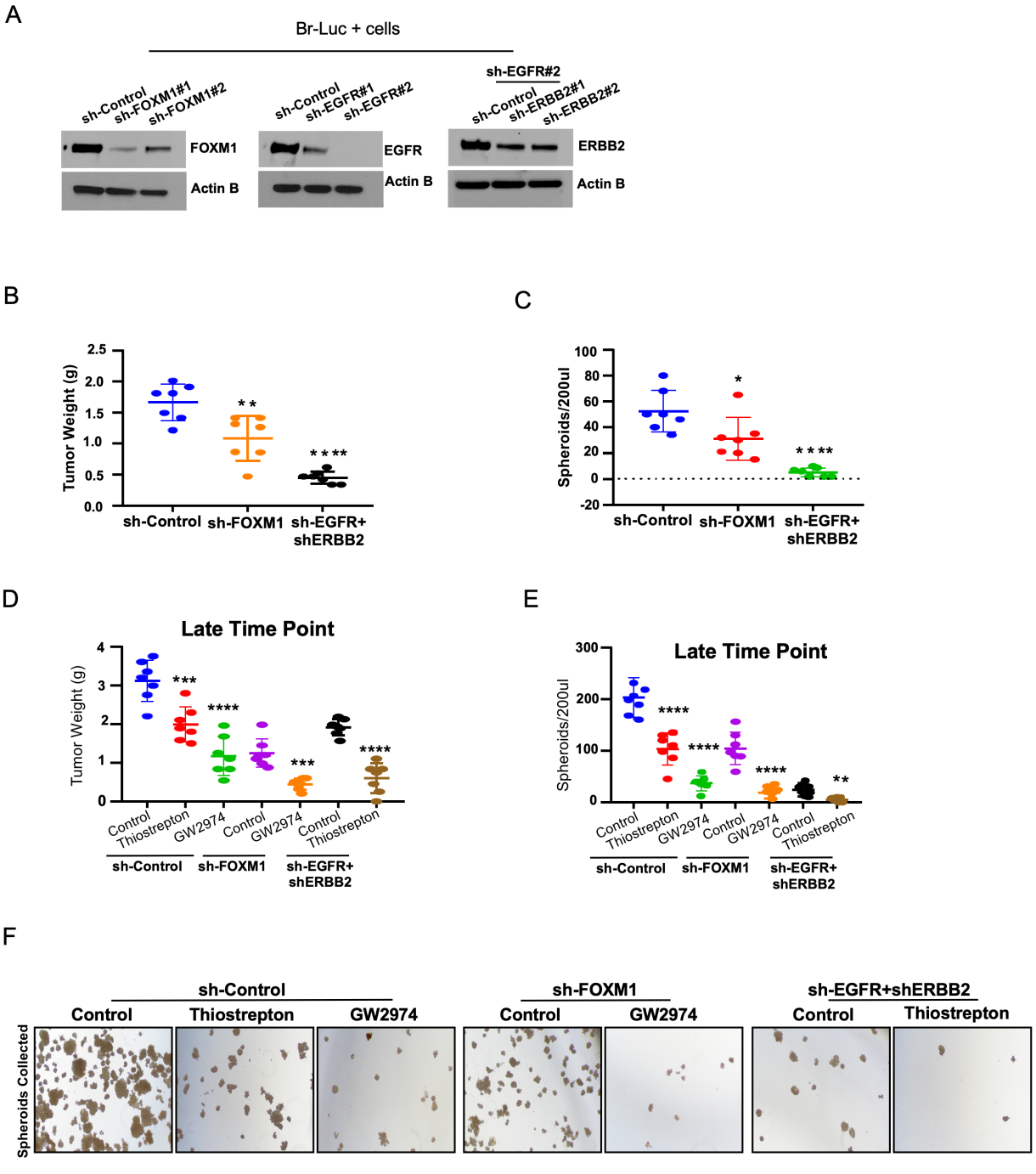
Supplementary Figure-8



Supplementary Figure-8. A, Mouse ovarian cancer cells were lysed and immunoblot was performed using the antibodies indicated. **B,** Representative IHC images of tumor

tissue collected from **Figure-7A,** and stained using Hematoxylin and Eosin, or using the antibodies indicated (left). Images were captured under 20X magnification staining intensity of PCNA and cleaved caspase-3 were quantitated from 5 randomly selected fields (right). Error bars indicate SE of 5 independent determinations. **p<0.01, ***p<0.001, ****p<0.0001.

Supplementary Figure-9



Supplementary Figure-9. A. BR-Luc cells, which stably express control shRNA, two different shRNAs target FOXM1 or EGFR. To develop stable knockouts of both EGFR and ERBB2, we used ERBB2- shRNA viral particles to infect BR-Luc cells stably express sh-EGFR (clone #2)). Stable cells were selected and immunoblotted using the indicated antibodies. **B-C.** BR-Luc cells stably knockdown of FOXM1, or knockdown of both EGFR and ERBB2 or BR-Luc cells express control-shRNAs were injected intraperitoneally ($1X10^5$ cells) in female FVB mice (n=7). Mice were sacrificed on 25^{th} day after tumor cell inoculation and tumors from all organ sites and ascites fluid were collected and tumor weight was quantitated. Tumor spheroids in the peritoneum were collected by filtering peritoneal wash through 50 μM filter and the number tumor spheroids in the peritoneal fluid were quantitated. **D-E**, Mice bearing BR-Luc cells express control-shRNA cells were treated with either Thiostrepton (FOXM1 inhibitor) or Lapatinib (GW2974). The mice bearing BR-Luc cells expressing sh-FOXM1 were treated with lapatinib; whereas mice bearing BR-Luc cells expressing sh-EGFR and sh-ERBB2, were treated with Thiostrepton. All the drugs were continued twice a week for 7 weeks and monitored tumor growth. Total tumor weight and number tumor spheroids in ascites fluid were determined as described in B and C above. **F**, Tumor spheroids collected from (E) were plated in a 6 well culture plate and the number of spheroids were imaged. Error bars indicates average ± SE and p-values were determined by one-way ANOVA.

Supplementary Table-1: Sequences of siRNAs used for transfections

Gene		Sequence
FoxM1	Sense	5'CACUAUCAACAAUAGCCUA 3'
	Antisense	5'UAGGCUAUUGUUGAUAGU3'
FoxM1	Sense	5'GCUCAUACCUGGUACCUAU3'
	Antisense	5'AUAGGUACCAGGUAUGAGC3'
Zeb1	Sense	5'GGAAGAACGUGACAGCACA3'
	Antisense	5'UGUGCUGUCACGUUCUUCC3'
Zeb1	Sense	5'GGACAGCACAGUAAAUCUA 3'
	Antisense	5'UAGAUUUACUGUGCUGUCCT3'
ERBB2	Sense	5'CAUCAAAGUGUUGAGGGAA3'
	Antisense	5'UUCCCUCAACACUUUGAUG3'
EGFR	Sense	5'GCUUUGGUGCCACCUGCGU3'
	Antisense	5'ACGCAGGUGGCACCAAAGC3'
TP53	Sense	5'GUCUUUGAACCCUUGCUUG3'
	Antisense	5'CAAGCAAGGGUUCAAAGAC3'
EGF	Sense	5'GGAUUAAUCCACGAAUUGA3'
	Antisense	5'UCAAUUCGUGGAUUAAUCC3'
TGFA	Sense	5'CAGUUGGUGUCUGAGUCCA3'
	Antisense	5'UGGACUCAGACACCAACUG3'
GRB7	Sense	5'GCCUGUACUUCAGCAUGGA3'
	Antisense	5'UCCAUGCUGAAGUACAGGC3'
AXL	Sense	5'GGAAGAUUUGGAGAACACA3'
	Antisense	5'UGUGUUCUCCAAAUCUUCC3'
RPS6KB1	Sense	5'CCACAAUCGUGCUGUGGAU3'
	Antisense	5'AUCCACAGCACGAUUGUGG3'
TRIM25	Sense	5'CCAUUUAUCAGAUUCUCCU3'
	Antisense	5'AGGAGAAUCUGAUAAAUGG3'
CDK2	Sense	5'GCACGUACGGAGUUGUGUA3'
	Antisense	5'UACACAACUCCGUACGUGC3'
MSH6	Sense	5'GAAAUGAUGGAGGCACGAU3'
	Antisense	5'AUCGUGCCUCCAUCAUUUC3'
PMS2	Antisense	5'GAAGAUACCGGAUGUAAAU3'
	Sense	5'AUUUACAUCCGGUAUCUUC3'
CCND1	Antisense	5'GCAUGUUCGUGGCCUCUAA3'
	Sense	5'UUAGAGGCCACGAACAUGC3'
CDH1	Antisense	5'GCUGUAUACACCAUAUUGA3'
	Sense	5'UCAAUAUGGUGUAUACAGC3'
MCL1	Antisense	5'GUAAUAGAACUAUGACUGU3'
	Sense	5'ACAGUCAUAGUUCUAUUAC3'
CASP9	Antisense	5'CAGACAGUGGUGUUGAUGA3'
	Antisense	5'UCAUCAACACCACUGUCUG3'
CDH3	Sense	5'GCAACUUUAUAAUUGAGAA3'
	Antisense	5'UUCUCAAUUAUAAAGUUGC3'
FN1	Sense	5'CACUUAUGAGCGUCCUAAA3'
	Antisense	5'UUUAGGACGCUCAUAAGUG3'
TP53	Sense	5'GAGGUUGGCUCUGACUGUA3'
	Antisense	5'UACAGUCAGAGCCAACCUC3'

ITGA5	Sense	5’CAGAUAACUUCACCCGAAU3’
	Antisense	5’AUUCGGGUGAAGUUAUCUG3’
ITGB1	Antisense	5’CUGUUCUUUGGAUACUAGU3’
	Antisense	5’ACUAGUAUCCAAAGAACAG3’
ITGAV	Sense	5’CUUUACUGCUGAUAGUGCU3’
	Antisense	5’AGCACUAUCAGCAGUAAAG3’
SYK	Sense	5’GAUGUACGAUCUCAUGAAU3’
	Antisense	5’AUUCAUGAGAUCGUACAUC3’
IL6	Sense	5’CUCACCUCUUCAGAACGAA3’
	Antisense	5’UUCGUUCUGAAGAGGUGAG3’
PAX8	Sense	5’CAUUCAACCUCCCUAUGGA3’
	Antisense	5’UCCAUAGGGAGGUUGAAUG3’
CASP8	Sense	5’CAUCUCAGUUCACUGGUUU3’
	Antisense	5’AAACCAGUGAACUGAGAUG3’
GAB2	Sense	5’CAAAGUGGAUAAUGGACUU3’
	Antisense	5’AAGUCCAUUAUCCACUUUG3’
LCK	Sense	5’ CAUUGAAGAGCGGAAUUAU3’
	Antisense	5’ AUAAUUCCGCUCUUCAAUG3’
MUC1	Sense	5’ CCACUUCUGCCAACUUGUA3’
	Antisense	5’ UACAAGUUGGCAGAAGUGG3’
RPS6	Sense	5’ GAGCUAGCAGAAUCCGCAA3’
	Antisense	5’ UUGCGGAUUCUGCUAGCUC3’
ITGA5	Sense	5’CAGAUAACUUCACCCGAAU3’
	Antisense	5’AUUCGGGUGAAGUUAUCUG3’
ITGB1	Antisense	5’CUGUUCUUUGGAUACUAGU3’
	Antisense	5’ACUAGUAUCCAAAGAACAG3’
ITGAV	Sense	5’CUUUACUGCUGAUAGUGCU3’
	Antisense	5’AGCACUAUCAGCAGUAAAG3’
PTK2	Sense	5’ CUUAGUACAGCUCUUGCAU3’
	Antisense	5’ AUGCAAGAGCUGUACUAAG3’
STAT3	Sense	5’GAGAUUGACCAGCAGUAUA3’
	Antisense	5’CAACAUGUCAUUUGCUGAA3’
KIT	Sense	5’ UAAAGGAAACGCUCGACUA3’
	Antisense	5’ UAGUCGAGCGUUUCCUUUA3’
CHEK1	Sense	5’ ACUGCGACUGCUUCUUCAGUU3’
	Antisense	5’ AACUGAAGAAGCAGUCGCAGU3’
CHEK2	Sense	5’ ACGCCGTCCTTTGAATAACAA3’
	Antisense	5’ AACUGAAGAAGCAGUCGCAGU3’
PTEN	Sense	5’ GUAUAGAGCGUGCAGAUAA 3’
	Antisense	5’ UUAUCUGCACGCUCUAUAC3’
ABL1	Sense	5’ACGCACGGACATCACCATGAA3’
	Antisense	5’ AACGGCTGATGTGGACTGTCT3’

Supplementary Table-2: Catalogue Numbers of shRNA

shRNA	Catalogue Numbers	Species	Company Name
Sh-ERBB2-	TRCN0000039882 TRCN0000039881	Human	Sigma Aldrich
Sh-EGFR-	TRCN0000121068 TRCN0000295971	Human	Sigma Aldrich
Sh-FOXM1	TRCN0000015546 TRCN0000015544	Human	Sigma Aldrich
Sh-FOXM1	TRCN0000349136 TRCN0000304362	Mouse	Sigma Aldrich
Sh-Control	CSHCTR001-LVRU6MH	Mouse	GeneCopoeia
Sh-Control	CSHCTR001-LVRH1GP	Mouse	GeneCopoeia
Sh-ERBB-2	CS-MSH023592-LVRU6MH-01	Mouse	GeneCopoeia
Sh-EGFR-1	CS-MSH043628-LVRH1GP-01	Mouse	GeneCopoeia

Supplementary Table-3. Sequences of the primers used for qPCR reactions

Gene	Sequence
FOXM1	F: 5' TGCAGCTAGGGATGTGAATCTTC 3'
	R: 5' GGAGCCCAGTCCATCAGAACT 3'
ZEB1	F: 5' GATGCGAAACGCGAGGTTTT 3'
	R: 5' GCTTCTAGACAGGAAATCCCACA 3'
ITGB1	F: 5' CCGCGCGGAAAAGATGAA 3'
	R: 5' TTGAATTTGTGCACCACCCAC 3'
ITGAV	F: 5' AGGGAAGCAAAGGACCGTCT 3'
	R: 5' AAGAGGGCTGAGCTTCGGA 3'
ITGA5	F: 5' CAGCCTTGCCAGAGATCCAA 3'
	R: 5' TCCTTGTGTGGCATCTGTCC 3'
LAMB1	F: 5' TCGGCTTTCAAACAAAAGAGGC 3'
	R: 5' TAGCCTTGGGAGGAACAGAG 3'
FN1	F: 5' ACAAGCATGTCTCTCTGCCA 3'
	R: 5' TCAGGAAACTCCCAGGGTGA 3'
CD44	F: 5' CCTGGCAGCCCCGATTATTT 3'
	R: 5' AAGGACACACCCAAGCAAGG 3'
CD24	F: 5' GAGAGATAACCCTGCCCGAG 3'
	R: 5'CAAAAGAAAAGTCCGCGCCT 3'
C-KIT	F: 5' ACTGTGGCCGTTATCTGGAAG 3'
	R: 5' CTTGGGTGACAACACAACCC 3'
ENDOGLIN	F: 5' TACCACAGCCTTCATCTGCG 3'
	R: 5' AGTTGCTGTCCGAAGGATGG 3'
EGFR	F: 5' GGCAGGAGTCATGGGAGAA 3'
	R: 5' GCGATGGACGGGATCTTAG 3'
HER2	F: 5' TTGGACAGCACCTTCTACCG 3'
	R: 5' GCTGGTTCACATATTCCTGGT 3'
PROMININ	F: 5' CCATACCTAGGTCCCCGTCC 3'
	R: 5' TTTATGACCCGGCTTCTGGG 3'
E CADHERIN	F: 5' GCTGGACCGAGAGAGTTTCC 3'
	R: 5' CAAAATCCAAGCCCGTGGTG 3'
VIMENTIN	F: 5' GCAGGAGGCAGAAGAATGGT 3'
	R: 5' CCACTTCACAGGTGAGGGAC 3'
ACTB	F: 5' GTCATTCCAAATATGAGATGCGT 3'
	R: 5' GCTATCACCTCCCCTGTGTG 3'

Supplementary Table-4. Sequences of primers used for the qPCR after chromatin immunoprecipitation

Gene	Sequence
ITGB1	F: 5' GTTTCCAGCCCCAATCTCCA 3'
	R: 5' GTCACGAAGGGTGGTGAGAC 3'
ITGAV	F: 5' GCCCTCTCTCTCAGTCCAGT 3'
	R: 5' AAGAAAGGATTCGCAGGGCA 3'
ITGA5	F: 5' GGAGCTGAAGGTTGGGTCC 3'
	R: 5' CAGTTGACCAAAAGCCTGCG 3'
LAMB1	F: 5' TGGATACTAGGGGCGGAGAG 3'
	R: 5' GGACTCGCGCATTTACGTTC 3'
FN1	F: 5' GCCCTGGGACTGAAAAGTCT 3'
	R: 5' GGTGGTGGTAGTGTTTGAGG 3'
ZEB1	F: 5' TTCCAATACTCCGGTCACGTTT 3'
	R: 5' GCTTCCCACCTCCTTCGAATA 3'
AURKB	F: 5' GGGGTCCAAGGCACTGCTAC 3'
	R: 5' GGGGCGGGAGATTTGAAAAG 3'
B-ACTIN	F: 5' AGCGCGGCTACAGCTTCA 3'
	R: 5' CGTAGCACAGCTTCTCCTTAATGT 3'

Supplementary Table-5. Catalogue numbers and available Research Resource Identifier (RRID) number of reagents and software

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
anti-ß-Actin	Santa Cruz Biotechnology	Cat# sc-58679; RRID: AB_772478
Purified Mouse IgG1, κ Isotype Ctrl Antibody	Biolegend	Cat# 401401, RRID: AB_2801452
Cleaved caspase3	Cell Signaling Technology	Cat# 9661, RRID: AB_2341188
Anti-rabbit IgG, HRP-linked Antibody	Cell Signaling Technology	Cat# 7074, RRID: AB_2099233
Anti-mouse IgG, HRP-linked Antibody	Cell Signaling Technology	Cat# 7076, RRID: AB_330924
Human ErbB2/Her2	R&D System	Cat# FAB9589G, RRID: AB_2800468
FOXM1	Santa Cruz Biotechnology	Cat# sc-376471, RRID: AB_11150135
EGFR	Cell Signaling Technology	Cat# 4267S RRID: RRID: AB_2246311
pEGFR Y1068	Cell Signaling Technology	Cat# 3777S RRID: RRID: AB_2096270
pEGFR Y992	Cell Signaling Technology	Cat# 2235S RRID: RRID: AB_331708
HER2	Cell Signaling Technology	Cat# 2165S RRID: RRID: AB_10692490
pHER2 Y877	Cell Signaling Technology	Cat# 2241S RRID: AB_2099407
pHER2 Y1248	Cell Signaling Technology	Cat# 2247S RRID: AB_331725
CD44	Abcam	Cat# ab157107 RRID: AB_2847859
CD24	Abcam	Cat# ab179821 RRID: Not Available
c-Kit	Abcam	Cat# ab32363 RRID: RRID: AB_731513
ITGAB1	Cell Signaling Technology	Cat# 4707P RRID: RRID: AB_2129049
ITGA-V	Cell Signaling Technology	Cat# 4711T RRID: RRID: AB_2128178
ITGA-5	Cell Signaling Technology	Cat# 4705T RRID: RRID: AB_2233962
ZEB1	Cell Signaling Technology	Cat# 3396S RRID: RRID: AB_1904164

GFP	Cell Signaling Technology	Cat# 2956S RRID: RRID: AB_1196615
EpCAM-PE	Invitrogen (Thermo Fisher Scientific)	Cat# 12-5791-83; RRID: AB_953617
CD45-FITC	Invitrogen (Thermo Fisher Scientific)	Cat#110459-42; RRID: AB_10852703
Chemicals, Peptides, and Recombinant Proteins		
DMEM	Thermo Fisher Scientific	Cat# 10569010
FBS	Atlanta Biologicals	Cat# H17112
Antibiotic (Penicillin/Streptomycin)	Thermo Fisher Scientific	Cat# 15140122
DMSO	Sigma Aldrich	Cat# D8418
Lipofectamine 2000	Invitrogen-Thermo Fisher Scientific	Cat# 11668027
Dharmafect Transfection Reagent	Dharmacon	Cat# T-2001-01
RNAiMax Transfection Reagent	Thermo Fisher Scientific	Cat# 13778030
Insulin	Sigma Aldrich	Cat# I3536
Trypan Blue	Sigma Aldrich	Cat# T8154
MTT	Sigma Aldrich	Cat# M5655
Glutaraldehyde	Sigma Aldrich	Cat# G7651
Crystal Violet	Sigma Aldrich	Cat# C6158
Acetic acid	Sigma Aldrich	Cat# A6283
Trypsin	Thermo Fisher Scientific	Cat# 15400054
PBS	Sigma Aldrich	Cat# P5493
RIPA buffer	Thermo Fisher Scientific	Cat# 89900
Protein A/G Magnetic Beads	Thermo Fisher Scientific	Cat# 88802
Paraformaldehyde	Sigma Aldrich	Cat# 158127
Triton X 100	Sigma Aldrich	Cat# T8787
Glycine	Sigma Aldrich	Cat# G8898
PIPES	Sigma Aldrich	Cat# P6757
KCl	Sigma Aldrich	Cat# P9333
NP40	Thermo Scientific	Cat# 85124
Sodium butyrate	Sigma Aldrich	Cat# B5887
Protease inhibitor cocktail	Sigma Aldrich	Cat# P8340
Dynabeads™ Protein A	Invitrogen	Cat# 10001D
Proteinase K	Invitrogen	Cat# 25530049
RNAse A	Invitrogen	Cat# AM2271
HBSS	Thermo Fisher Scientific	Cat# 14060040
10% Formalin fixative	VWR	Cat# 16004-121
TURBO DNase	Invitrogen	Cat# AM2238
Tween 20	Bio-Rad	Cat# 1706531
BSA	Sigma Aldrich	Cat# A7906
Ampicillin	Sigma Aldrich	Cat# A5354
LB Agar	Sigma Aldrich	Cat# L2897
LB Broth (Lennox)	Sigma Aldrich	Cat# L3022
Hematoxylin Solution, Harris Modified	Sigma Aldrich	Cat# HHS32-1L
IHC Antigen Retrieval solution	IHC World	Cat# IW-1100

Anti-Puromycin	Sigma Aldrich	Cat# MABE343; RRID: AB_2566826
Erlotinib	Selleckchem	Cat# S1023
GW2974	Selleckchem	Cat# S2111
Thiostrepton	Selleckchem	Cat# S4354
D-luciferin	Goldbio	Cat# LUCK-1G
EGF	Invitrogen	Cat# PHG0314
Polybrene Transfection Reagent	Millipore	Cat# TR-1003
Critical Commercial Assay Kits		
Calcein AM/EtBr based Live/Dead cell viability Kit	Molecular Probes, Thermo Fisher Scientific Inc.	Cat# L3224
Pierce BCA Protein Assay kit	Thermo Fisher Scientific	Cat# 23227
Subcellular Protein Fractionation Kit for Cultured Cells	Thermo Fisher Scientific	Cat# 78840
RNeasy kit	Qiagen	Cat# 74104
iScript cDNA synthesis kit	Bio-Rad	Cat# 1708891
iTaq Universal SYBR Green PCR Kit	Bio-Rad	Cat# 1725121
Dual-Luciferase Reporter Assay kit	Promega	Cat# E1910
ChIP assay kit	Millipore Sigma	Cat# 17-295
QIAquick PCR purification kit	Qiagen	Cat# 28104
Mycoplasma negative by PC R	Agilent Mycosensor Mycoplasma assay kit	Cat# 302109
FITC Annexin V/Dead Cell Apoptosis Kit	BD Pharmingen	Cat# 556547
Custom RT^2 PCR Array	Qiagen	Cat# 330171 CLAH36595
Human EGF Quantikine ELISA Kit	R&D	Cat# DEG00
Human TGF-alpha Quantikine ELISA Kit	R&D	Cat# DTGA00
Human Amphiregulin Quantikine ELISA Kit	R&D	Cat# DAR00
Experimental Models: Cell Lines		
IOSE80	MD Anderson Cancer Center	N/A
FTE187	MD Anderson Cancer Center	N/A
FTE188	MD Anderson Cancer Center	N/A
OVCAR4	NCI	Cat# OVCAR-4, RRID: CVCL_1627
OVCAR5	NCI	Cat# OVCAR-5, RRID: CVCL_1628
HEYA8	MD Anderson Cancer Center, Texas, USA	N/A

IGROV1	MD Anderson Cancer Center, Texas, USA	N/A
C11,	University of California, Los Angeles, CA, USA.	N/A
C11-FOXC2	University of California, Los Angeles, CA, USA.	N/A
BR-Luc	University of California, Los Angeles, CA, USA.	N/A
MeT-5A	ATCC	Cat# CRL-9444; RRID: CRL-9444
Biological Samples		
Human ovarian cancer tissues	Medical College of Wisconsin	Protocol Number: PRO00033433
Experimental Models: Organisms/Strains		
FVB/NJ - Homozygous mice	Jackson Laboratories	Cat# 001800
Oligonucleotides		
Primers for qPCR, see Table S4	IDT	N/A
Primers for CHIP-qPCR, see Table S4	IDT	N/A
Software and Algorithms		
GraphPad Prism 7	GraphPad Software	https://www.graphpad.com/scientific- software/prism/
FlowJo Software	FlowJo	https://www.flowjo.com/
qPCR software	BioRad CFX Maestro	Cat# 12004110
DNAstar	DNASTAR, Inc.	https://www.dnastar.com/software/lasergene/
Image J: J Coloc 2 image analysis plug-in	Image J	https://imagej.net/Coloc_2

Legends to Supplemental Spread Sheet 1.

List of Differentially expressed total and phosphorylated proteins present in OVCAR4 adherent cells versus non-adherent cells.

Legends to Supplemental Spread Sheet 2.

List of Differentially expressed total and phosphorylated proteins present in OVCAR5 adherent cells versus non-adherent cells.