Supplemental Information

Nucleoporin 160 Regulates Flowering through Anchoring HOS1 for Destabilizing CO in *Arabidopsis*

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Supplemental Figure 1. Effects of *Nup160* on the Flowering Response to GA, Vernalization, and Ambient Temperature.

(A) Flowering time of wild-type (WT) and *nup160-2* in response to GA treatment under short days. Exogenous GA (100 μ M) or mock treatment was applied weekly after germination (n \geq 16, \pm SD).

(B) Flowering time of wild-type (WT) and *nup160-2* in response to vernalization. Seeds were germinated on MS medium and vernalized at 4°C under low light conditions for 6 weeks and then transferred to soil to grow under short days ($n \ge 20, \pm SD$).

(C) Flowering time of wild-type (WT) and *nup160-2* in response to changes in ambient temperature (16°C, 23°C, and 28°C) under long days. The ratios of flowering time between 16°C and 23°C (16°C/23°C) and between 23°C and 28°C (23°C/28°C) for wild type and *nup160-2* are shown in the table ($n \ge 16, \pm$ SD).



Supplemental Figure 2. Effects of Various Flowering Pathways on Nup160 Expression.

(A) *Nup160* expression in 9-day-old seedlings of photoperiod pathway mutants grown under long days.

(B) Expression of *Nup160* in response to vernalization. Seeds were germinated on MS medium and vernalized at 4°C under low light conditions for 6 weeks before being transferred to soil to grow under long days ($n \ge 20, \pm$ SD). 9-day-old seedlings were harvested for expression analysis.

(C) Expression of *Nup160* in 9-day-old wild-type seedlings in response to GA treatment. Samples were harvested 6 h after GA or mock treatment.

(D) *Nup160* expression in in 9-day-old seedlings of autonomous pathway mutants grown under long days.

(E) Effects of ambient temperature on Nup160 expression. 9-day-old wild-type seedlings grown at 16°C, 23°C, and 28°C under long days were harvested for expressions analysis. Gene expression levels in all panels were normalized against the expression of TUB2 and shown as relative values to the highest level in each panel as 1. Error bars indicate SD.



Supplemental Figure 3. Rescue of *nup160-2* and *hos1-3* Mutants by Various Gene-Tagging Lines.

(A) Schematic diagrams showing the *Nup160* and *HOS1* genomic fragments used for complementation experiments. Exons are represented by black boxes.

(B) Flowering time of wild-type, *nup160-2* and *hos1-3* as well as their rescued lines grown under long days ($n \ge 16, \pm SD$). Asterisks indicate statistically significant differences in flowering time between specified genotypes (two-tailed paired Student's *t* test, *P* < 0.005).



Supplemental Figure 4. Diurnal Expression of *FT*, *SOC1*, and *CO* in Wild-Type and *nup160-2* Plants under Short Days or Long Days.

(A-C) Diurnal expression of FT (A), CO (B), and SOC1 (C) in 2-week-old wild-type and nup160-2 seedlings grown under short days.

(D) Diurnal expression of SOC1 in 9-day-old wild-type and nup160-2 seedlings grown under long days. Gene expression levels were normalized against the expression of TUB2 and shown as relative values to the highest level in each panel as 1.0. Error bars indicate SD.



Supplemental Figure 5. Characterization of *SUC2:3FLAG-CO* in Wild-Type and *nup160-2* Backgrounds.

(A) Flowering phenotype of wild-type, *nup160-2*, *SUC2:3FLAG-CO*, and *SUC2:3FLAG-CO nup160-2* plants grown under long days.

(B) Immunoblot analysis of nuclear protein extracts using anti-FLAG antibody shows detection of 3FLAG-CO in *SUC2:3FLAG-CO* and *SUC2:3FLAG-CO nup160-2*, but not in wild-type and *nup160-2* seedlings (upper panel). 9-day-old seedlings grown under long days were collected at ZT4. Histone H3 is shown as a loading control (lower panel).



Supplemental Figure 6. Characterization of Genetic or Molecular Interaction between *Nup160* and Regulators of *CO*.

(A) Diurnal expression of *Nup160* in 9-day-old wild-type seedlings grown under long days. Samples were harvested every 4 hours from the onset of illumination, which are shown in hours as Zeitgeber time (ZT). Gene expression levels were normalized against the expression of *TUB2* and shown as relative values to the highest level as 1.0. Error bars indicates SD.

(B) Subcellular localization of Nup160-GFP detected in meristematic root cells (left) and mature root cells (right) of 5-day-old seedlings grown under continuous light or transferred to dark for 4 hours. Scale bar corresponds to $10 \mu m$.

(C) Flowering time of various mutants grown under long days. Asterisks indicate statistically significant differences in flowering time between specified genotypes (two-tailed paired Student's *t* test, P < 0.005) (n $\ge 16, \pm$ SD).

(**D**) Diurnal expression of *HOS1* in 9-day-old wild-type and *nup160-2* seedlings grown under long days. Gene expression levels were normalized against the expression of *TUB2* and shown as relative values to the highest level as 1.0. Error bars indicate SD.



Supplemental Figure 7. Nup160 Affects NPC-Localization of HOS1.

(A and B) Subcellular localization of GFP-HOS1 in a mature root cell (A) and a leaf petiole cell (B) of *GFP-gHOS1 hos1-3* in the wild-type (WT) background. Scale bars correspond to 10 μ m. (C and D) Subcellular localization of GFP-HOS1 in a mature root cell (C) and a leaf petiole cell (D) of *GFP-gHOS1 hos1-3* in the *nup160-2* background. Arrowheads outline the nucleus localization observed in the *nup160-2* background. GFP, GFP fluorescence; BF, bright field; Chlorophyll, autofluorescence of chlorophyll; Merge, merge of GFP and BF with/without Chlorophyll images. Scale bars correspond to 10 μ m.



Supplemental Figure 8. Nup160 Affects NPC-Localization of HOS1 When *HOS1* Is Overexpressed.

(A) Flowering time (upper panel; $n \ge 16, \pm SD$) and *HOS1* expression (lower panel) in wild-type, *hos1-3*, 35S: *GFP-HOS1 hos1-3*, *nup160-2*, and 35S: *GFP-HOS1 nup160-2 hos1-3* seedlings grown under long days. 9-day-old seedlings were collected for semi-quantitative PCR analysis. *TUB2* was amplified as an internal control.

(**B** and C) Subcellular localization of GFP-HOS1 in meristematic root cells (left panels) and mature root cells (right panels) of 5-day-old 35S: GFP-HOS1 hos1-3 (**B**) and 35S: GFP-HOS1 nup160-2 hos1-3 (**C**) plants. Scale bars correspond to 10 µm.



Supplemental Figure 9. Detection of nlsRFP in Leaf Vasculature Cells of SUC2:nlsRFP.

Confocal analysis of nlsRFP expressed in the first true leave of 9-day-old *SUC2:nlsRFP GFP-gHOS1 hos1-3* (indicated as WT; upper panel) and *SUC2:nlsRFP GFP-gHOS1 nup160-2 hos1-3* (indicated as *nup160-2*; lower panel). RFP, RFP fluorescence; BF, bright field; Merge, merge of RFP and BF images. Scale bars correspond to 200 µm.



Supplemental Figure 10. NUP160 Does Not Affect *GFP-HOS1* mRNA and Protein Expression.

(A) Semi-quantitative PCR shows comparable *GFP-HOS1* mRNA expression in 9-day-old *GFP-gHOS1 hos1-3* and *GFP-gHOS1 nup160-2 hos1-3* seedlings. The fragment covering a 500-bp *HOS1* coding region was amplified, while *TUB2* was amplified as an internal control.
(B) Immunoblot analysis shows comparable GFP-HOS1 protein expression in *GFP-gHOS1 hos1-3* and *GFP-gHOS1 nup160-2 hos1-3*. Total protein was extracted from 9 and 11-day-old seedlings and analyzed using anti-GFP antibody. The membrane stained with Ponceau Red showing the ribulose bisphosphate carboxylase large subunit (RbcL) was included as a loading control.

(C-F) *HOS1-GUS* expression in 9-day-old *gHOS1-GUS* in the wild-type (WT) (C and D) and *nup160-2* (E and F) backgrounds. (D and F) High-magnification images of the black squares shown in (C and E), respectively. Scale bars correspond to 1 mm (C-F).



Supplemental Figure 11. Nup160 and HOS1 Similarly Affect *FLC* Expression. (A) *FLC* expression in 9-day-old of wild-type (WT), *nup160-2*, *hos1-3*, and *nup160-2 hos1-3* seedlings grown under long days. Gene expression levels were normalized against the expression of *TUB2* and shown as relative values to the highest level as 1. Error bars indicate SD. (B) Flowering time of various mutants grown under long days ($n \ge 16, \pm$ SD). Asterisks indicate statistically significant differences in flowering time between specified genotypes (two-tailed paired Student's *t* test, *P* < 0.05).

Supplemental Table 1. List of Primers Used in This Study.

		S (51.23)
Construct	Primer Name	Sequence (5'-3')
miR-Nup160	1-AmiR-Nup160-s	GATTGATACATGTAACTTGGCGCTCTCTCT TTTGTATTCC
	2-AmiR-Nup160-a	GAGCGCCAAGTTACATGTATCAATCAAAG
	3-AmiR-Nup160-*s	GAGCACCAAGTTACAAGTATCATTCACAG GTCGTGATATG
	4-AmiR-Nup160-*a	GAATGATACTTGTAACTTGGTGCTCTACAT ATATATTCCT
gNup160	gNup160-F	CACCTGATTGTAACTAACACATGACCAC
	gNup160-R	TGTTACTTCTTCTTCTCACACGCAC
3FLAG- gNup160	3FLAG-gNup160-F	GGTTTAGGGTTCAAAGAAGAAATGGATTA CAAGGATCATGATGGA
	3FLAG-gNup160-R	ATTCCGACGATTCTCCTCCTTATCGTCATCA TCTTTGTAATC
aNun 160 Vnul	gNup160-KpnI-F	CCTCTGCAACCGGTGGTACCAGAAAGTCT TGATTGTTCCGTG
gNup160-Kpn1	gNup160-KpnI-R	CAATCAAGACTTTCTGGTACCACCGGTTG CAGAGGATACAG
- 11001	gHOS1-F	CACCTGGTGTTGACCGTCTCATGGAGA
ghOSI	gHOS1-R	GTGACGGAACGATAGGAAGAGAC
gHOS1-KpnI	gHOS1-KpnI-F	AGCAAGAGGTACCTGAAATGAAACAAAC ACTCGAAAAGAC
	gHOS1-KpnI-R	TTCATTTCAGGTACCTCTTGCTGCGAATCT ACGTCTC
GUS-gHOS1	GUS(N)-KpnI-F	CGGGGTACCATGTTACGTCCTGTAGAAAC C
	GUS(N)-KpnI-R	CGGGGTACCTCCACCTCCTTGTTTGCCTCC CTGCTGCG
GFP-gHOS1	GFP-F-KpnI	CGGGGTACCATGAGTAAAGGAGAAGAACT TTTCAC
	GFP(N)-R-KpnI	CGGGGTACCTCCACCTCCTTTGTATAGTTC ATCCATG
gNup160-GUS	GUS(C)-KpnI-F	CGGGGTACCGGAGGTGGAATGTTACGTCC TGTAGAAACC
	GUS(C)-KpnI-R	CGGGGTACCCTATTGTTTGCCTCCCTGCTG CG
35S:GFP-HOS1	GFP-F-XhoI	CCGCTCGAGATGAGTAAAGGAGAAGAACT TTTCAC
	GFP-R-XhoI	CCGCTCGAGTTTGTATAGTTCATCCATGC
	HOS1-F-XmaI	CCCCCCGGGATGGATACGAGAGAAATCAA CGGT

Primers for Plasmid Construction

	HOS1-R-XmaI	CCCCCGGGTCATCTTGCTGCGAATCTACG
SUC2:3FLAG- CO	3FLAG-CO-F-XmaI	CCCCGGGATGGATCCCCGGGATGGATTAC
		AAGGATCATGATGGAGACTACAAGGATCA
		TGATATTGATTACAAAGATGATGACGATAA
		GTTGAAACAAGAGAGTAACGACA
	CO-R-XmaI	CCCCGGGTCAGAATGAAGGAACAATCCCA
gNup160- cEYFP	cEYFP-F-KpnI	CGGGGTACCGGCAGCGTGCAGCTCGCC
	cEYFP-R-KpnI	CGGGGTACCTCACTTGTACAGCTCGTCCA
nEYFP-gHOS1	nEYFP-F-KpnI	CGGGGTACCATGGTGAGCAAGGGCGAGG
		А
	nEYFP-R-KpnI	CGGGGTACCGTCCTCGATGTTGTGGCGGA
SUC2:nlsRFP	nlsRFP-F-XmaI	CCCCGGGATGCCAAAGAAAAAGAGGAAA
		GTGCCTAAGAAGAAGAGAAAGGTTGCGA
		AGGCAGATAAGAAAC
	RFP-R-XbaI	GCTCTAGACTAGGCGCCGGTGGAGTGGC

Primers for Quantitative Real-Time PCR

Primer Name	Sequence (5'-3')
TUB2-F	GAGAATGCTGATGAGTGCATGG
TUB2-R	AGAGTTGAGTTGACCAGGGAACC
Nup160-F	AAAGGAACTCTGTGGTCATCGC
Nup160-R	TGATCAGCAGGGCTTCTATCAGG
FT-F	CTTGGCAGGCAAACAGTGTATGCAC
FT-R	GCCACTCTCCCTCTGACAATTGTAGA
CO-F	TCAGGGACTCACTACAACGACAATGG
CO-R	TTGGGTGTGAAGCTGTTGTGACACAT
SOC1-F	AGCTGCAGAAAACGAGAAGCTCTCTG
SOC1-R	GGGCTACTCTCTTCATCACCTCTTCC
HOS1-F	GTCATTGCTTTGCTGGA
HOS1-R	TCTAACCACGCTTGCATCTC
FLC-F	CTAGCCAGATGGAGAATAATCATCATG
FLC-R	TTAAGGTGGCTAATTAAGTAGTGGGAG

Primers for Semi-Quantitative PCR

Primer Name	Sequence (5'-3')
TUB2-semi-F	ATCCGTGAAGAGTACCCAGAT
TUB2-semi-R	TCACCTTCTTCATCCGCAGTT
Nup160-semi-F	ATGGAGGAGAATCGTCGGAATC
Nup160-semi-R	GTCGAACATCCAAGTGAATCAAATGAT
HOS1-semi-F	CATAATGGTCTGGTGCATCAGA
HOS1-semi-R	AATCCTACAAGCTTCCTGGAGTG