

Supporting Information

CRISPR-generated *Nrf2a* loss- and gain-of-function mutants facilitate mechanistic analysis of chemical oxidative stress-mediated toxicity in zebrafish

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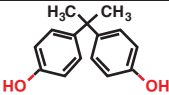
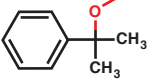
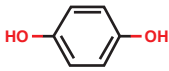
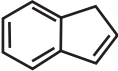
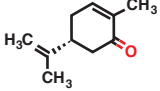
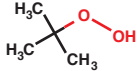
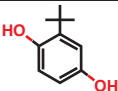
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Figure S1.

| Chemical | CAS | Structure | LC ₅₀ | DMSO conc. | 40% LC ₅₀ | 20% LC ₅₀ | Gene Expression | | | | | | | |
|----------------------------------|-----------|--|------------------|------------|----------------------|----------------------|-----------------|-------------|-------------|--------------|--------------|-------------|-------------|-------------|
| | | | | | | | <i>hmx1a</i> | <i>gclc</i> | <i>gstp</i> | <i>prdx1</i> | <i>gpx1a</i> | <i>ngq1</i> | <i>sod1</i> | <i>sod2</i> |
| Bisphenol A | 80-05-7 |  | 12.8 | 0.003% | 5.11 | -- | | | | | | | | |
| <i>R</i> -(-)-Carvone | 6485-40-1 |  | 58.2 | -- | 23.3 | 11.6 | | * | * | * | | | * | * |
| Cumene hydroperoxide | 80-15-9 |  | 23.4 | 0.00009% | 9.35 | 4.68 | | * | * | * | * | * | | |
| Hydroquinone | 123-31-9 |  | 0.24* | -- | 0.095 | -- | | | | | | | | |
| Indene | 95-13-6 |  | 50.9 | 0.05% | 20.4 | -- | * | * | | | | * | * | |
| <i>tert</i> -Butyl hydroperoxide | 75-91-2 |  | 176 | -- | 70.3 | 35.2 | | * | * | * | * | * | * | * |
| <i>tert</i> -Butyl hydroquinone | 1948-33-0 |  | 0.35* | 0.000002% | 0.142 | -- | | | * | * | | * | | |

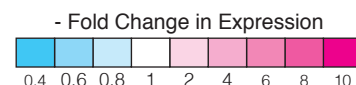


Figure S1. All chemicals used in the present study. All concentrations are mg/L. LC₅₀ concentrations marked with * were determined for this work whereas all others were identified in ref 1. DMSO concentrations (in % v/v where indicated) are final concentrations in all exposures including the 0 mg/L control. Heat map represents expression changes from exposure to 40% LC₅₀ for each chemical from 5 hpf to 96 hpf with daily renewal. Asterisks in boxes indicate significantly different expression ($p < 0.05$) relative to untreated controls. Control (untreated) expression in all cases is set to 1.0. All data represent the mean \pm SEM ($n = 5$ biological replicates, 10 larvae per replicate).

Note. While researchers have used *t*BHQ as a strong activator of Nrf2a in zebrafish,²⁻⁵ we found that effects of *t*BHQ on the Nrf2a-responsive genes *gstp* and *prdx1* were only moderate at 96 hpf relative to other compounds tested. It is likely that the length of exposure used in this study accounted for this difference relative to previous work. For example, *t*BHQ strongly activates Nrf2a under brief 3-6 hr exposures especially at 2 dpf.^{5,6} In addition, an *in vivo* study using rat embryos indicated that NRF2 activation by *t*BHQ was greater after acute exposures than subacute exposures.⁷ Because our group was examining the effects of several compounds on Nrf2a-dependent gene expression, it was beyond the scope of the study to test the effects of the test agents at multiple time points.

Figure S2.

A.

| | | |
|--------------|---|----|
| <i>WT</i> | <u>GTCTGTTTTCTCTGCAGGACATGGATCTGATCGATATCCTGTGGCGGCAGGATGTGGATC</u> | 60 |
| <i>w210</i> | GTCTGTTTTCTCTGCAGGACATGGATCTGATCGATATCCTGTGGCGGCAGGATGTGGATC | 60 |
| <i>w211</i> | GTCTGTTTTCTCTGCAGGACATGGATCTGATCGATATCCTGTGGCGGCAGGATGTGGATC | 60 |
| <i>w212</i> | GTCTGTTTTCTCTGCAGGACATGGATCTGATCGATATCCTGTGGCGGCAGGATGTGGATC | 60 |
| <i>dw213</i> | GTCTGTTTTCTCT----- | 13 |
| <i>dw214</i> | GTCTGTTTTCTCTGCAGGACATGGATCTGATCGATATCCTGTGGCGGCAGGATGTGGATC | 60 |

| | | |
|--------------|---|-----|
| <i>WT</i> | <u>TGGGCGCGGGCCGTGAGGTGTTTCGACTTCAGCTACCGGCAGAAGGAGGTGGAGCTGCGCA</u> | 120 |
| <i>w210</i> | TGGGC----- TCGTGAGGTGTTTCGACTTCAGCTACCGGCAGAAGGAGGTGGAGCTGCGCA | 115 |
| <i>w211</i> | TGGGCGCGGGCCGTGAGGTGTTTCGACTTCAGCTACCGGCAGAAGGAGGTGGAGCTGCGCA | 120 |
| <i>w212</i> | TGGGCGC----- | 67 |
| <i>dw213</i> | ----- | 13 |
| <i>dw214</i> | TGGGCGCGGGC----- | 71 |

| | | |
|--------------|---|-----|
| <i>WT</i> | <u>GGCGGAGGGAGCAGGAGGAGCAGGAGCTGCAGGAGCGTCTGCAGGAGCAGGAGAAGACAC</u> | 180 |
| <i>w210</i> | GGCGGAGGGAGCAGGAGGAGCAGGAGCTGCAGGAGCGTCTGCAGGAGCAGGAGAAGACAC | 175 |
| <i>w211</i> | GGCGGAGGGAGCAGGAGGAGCAGGAGCTGCAGGAGCGTCTGCAGGAGCAGGAGAAGACAC | 180 |
| <i>w212</i> | ----- | 67 |
| <i>dw213</i> | -----GCGTCTGCAGGAGCAGGAGAAGACAC | 39 |
| <i>dw214</i> | ----- | 71 |

| | | |
|--------------|--|-----|
| <i>WT</i> | <u>TGCTGGCTCAGCTGCAGCTCGACGAGGAGACCGGAGAGTTCCTG<u>CCGCGCAGCACACCGC</u></u> | 240 |
| <i>w210</i> | TGCTGGCTCAGCTGCAGCTCGACGAGGAGACCGGAGAGTTCCTG <u>CCGCGCAGCACACCGC</u> | 235 |
| <i>w211</i> | TGCTGGCTCAGCTGCAGCTCGACGAGGAGACCGG----- TCGGCACACCGC | 226 |
| <i>w212</i> | ----- | 67 |
| <i>dw213</i> | TGCTGGCTCAGCTGCAGCTCGACGAGGAGACCGGAGAGTTCCTG <u>CCGCGCAGCACACCGC</u> | 99 |
| <i>dw214</i> | -----ACACCGC | 78 |

| | | |
|--------------|---|-----|
| <i>WT</i> | <u>TCACACACACACCTGAAGCAGACGGAGGAGGAGCGGGAGAAATCACACAGGTACACATGC</u> | 300 |
| <i>w210</i> | TCACACACACACCTGAAGCAGACGGAGGAGGAGCGGGAGAAATCACACAGGTACACATGC | 295 |
| <i>w211</i> | TCACACACACACCTGAAGCAGACGGAGGAGGAGCGGGAGAAATCACACAGGTACACATGC | 286 |
| <i>w212</i> | TCACACACACACCTGAAGCAGACGGAGGAGGAGCGGGAGAAATCACACAGGTACACATGC | 127 |
| <i>dw213</i> | TCACACACACACCTGAAGCAGACGGAGGAGGAGCGGGAGAAATCACACAGGTACACATGC | 159 |
| <i>dw214</i> | TCACACACACACCTGAAGCAGACGGAGGAGGAGCGGGAGAAATCACACAGGTACACATGC | 138 |

B.

| | | | |
|--------------|----|--|----|
| <i>WT</i> | 15 | DMDLIDILWRQD <u>VDL</u> GAGREVFDFSYR <u>Q</u> KEVELRRRREQEEQELQERLQEQEK <u>TLLAQLQ</u> | 75 |
| <i>w210</i> | 15 | DMDLIDILWRQD <u>VDLGS</u> * | 32 |
| <i>w211</i> | 15 | DMDLIDILWRQD <u>VDL</u> GAGREVFDFSYR <u>Q</u> KEVELRRRREQEEQELQERLQEQEK <u>TLLAQLQ</u> | |
| <i>w212</i> | 15 | DMDLIDILWRQD <u>VDLGAHTHT</u> * | 36 |
| <i>dw213</i> | 15 | -----EQEK <u>TLLAQLQ</u> | 26 |
| <i>dw214</i> | 15 | DMDLIDILWRQD <u>VDL</u> GAG----- | 33 |

| | | |
|-------|---|-----|
| WT | <u>LDEETGE</u> FLPRSTPLTHTPEADGGGAGEITQ | 106 |
| w210 | --- | 32 |
| w211 | <u>LDEETGR</u> HTAHTHT * | 89 |
| w212 | --- | 36 |
| dw213 | <u>LDEETGE</u> FLPRSTPLTHTPEADGGGAGEITQ | 57 |
| dw214 | -----TPLTHTPEADGGGAGEITQ | 52 |

Figure S2. Independently-isolated *nfe2l2a* mutant lines encode null (LOF) or hyperactive (GOF) alleles. (a) Genomic sequences of Intron 1 (grey letters) and Exon 2 (black letters) for wild type and five mutant alleles show deletions (dashes) and insertions (bold text) near CRISPR guide sites (underlined in wild type, with PAM sites double underlined). (b) Protein sequences predicted from Exon 2 cDNA show that three out-of-frame deletions result in putative LOF alleles with altered amino acids (bold) and novel stop codons (*), while the two in-frame deletions result in putative GOF alleles by removing of some or all of the DLG and ETGE motifs bound by Keap1 (underlined) and lysine residues that are targets of ubiquitination (double underline).

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