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Supporting Information

Amino Acid Modified RNA Bases as Building Blocks of an Early Earth RNA-Peptide World

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1. General Experimental Methods

Chemicals were purchased from Sigma-Aldrich, TCI, Fluka, ABCR, Carbosynth or Acros Organics and used without further purification. Some of the strands were purchased from Metabion or Ella Biotech. Reagent-grade dry solvents (Sigma-Aldrich, Acros Organics) were stored over molecular sieves and handled under inert gas atmosphere. Reactions and chromatography fractions were monitored by qualitative thin-layer chromatography (TLC) on silica gel F254 TLC plates from Merck KGaA. Flash column chromatography was performed on Geduran® Si60 (40-63 µm) silica gel from Merck KGaA. NMR spectra were recorded on Bruker AVIIIHD 400 spectrometers (400 MHz). ¹H NMR shifts were calibrated to the residual solvent resonances: DMSO-d₆ (2.50 ppm), CDCl₃ (7.26 ppm), Acetone-d₆ (2.05 ppm), CD₂Cl₂ (5.32 ppm). ¹³C NMR shifts were calibrated to the residual solvent: DMSO-d₆ (39.52 ppm), CDCl₃ (77.16 ppm), Acetone-d₆ (29.84 ppm), CD₂Cl₂ (53.84 ppm). All NMR spectra were analysed using the program MestreNova 10.0.1 from Mestrelab Research S. L. High resolution mass spectra were measured by the analytical section of the Department of Chemistry of the Ludwigs-Maximilians-Universität München on the spectrometer MAT 90 (ESI) from Thermo Finnigan GmbH. IR spectra were recorded on a PerkinElmer Spectrum BX II FT-IR system. All substances were directly applied as solids or on the ATR unit. Analytical RP-HPLC was performed on an analytical HPLC Waters Alliance (2695 Separation Module, 2996 Photodiode Array Detector) equipped with the column Nucleosil 120-2 C18 from Macherey Nagel using a flow of 0.5 ml/min, a gradient of 0-30% of buffer B in 45 min was applied. Preparative RP-HPLC was performed on a HPLC Waters Breeze (2487 Dual λ Array Detector, 1525 Binary HPLC Pump) equipped with the column VP 250/32 C18 from Macherey Nagel using a flow of 5 ml/min, a gradient of 0-25% of buffer B in 45 min was applied for the purifications. Oligonucleotides were purified using the following buffer system: buffer A: 100 mM NEt₃/HOAc (pH 7.0) in H₂O and buffer B: 100 mM NEt₃/HOAc in 80% (v/v) acetonitrile. The pH values of buffers were adjusted using a MP 220 pH-meter (Metter Toledo). Oligonucleotides were detected at wavelength: 260 nm. Melting profiles were measured on a JASCO V-650 spectrometer. Calculation of concentrations was assisted using the software OligoAnalyzer 3.0 (Integrated DNA Technologies: https://eu.idtdna.com/calc/analyzer). For strands containing artificial bases, the extinction coefficient of their corresponding canonical-only strand was employed without corrections. Matrix-assisted laser desorption/ionization-time-of-flight (MALDI-TOF) mass spectra were recorded on a Bruker Autoflex II. For MALDI-TOF

measurements, the samples were desalted on a 0.025 µm VSWP filter (Millipore) against ddH₂O and co-crystallized in a 3-hydroxypicolinic acid matrix (HPA).

2. Synthesis of the Phosphoramidite Building-Blocks

2.1. Synthesis of RNA building blocks

Npe-protection of carboxy group of amino acid

The reaction was performed according to the procedure published before.¹

L-amino acid (1 eq.), 2-(4-nitrophenyl)ethanol (npe-OH, 3 eq.) and TsOH (3 eq.) were refluxed in toluene overnight in a *Dean-Stark* apparatus. The solution was cooled to room temperature and Et₂O was added. The oily residue was decanted, and the upper layer was removed to collect the oil. Precipitation of was induced by adding to the oil MeOH and Et₂O.

Compound 4



Yield: 70%; **IR**: $\tilde{v} = 3401$ (w), 2930 (s), 2892 (s), 2858 (s), 1730 (s), 1510 (vs), 1465 (s), 1300 (vs), 1258 (s), 1167 (s), 1010 (s), 895 (w), 832 (s) cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ : 8.2 – 8.1 (m, 5H), 7.58 (d, *J* = 8.8 Hz, 2H), 7.48 (d, *J* = 8.2 Hz, 2H), 7.11 (d, *J* = 8.2 Hz, 2H), 5.6 (br. s., 1H), 4.45 (t, *J* = 6.2 Hz, 2H), 4.1 – 4.0 (m, 1H), 3.89 (d, *J* = 4.0 Hz, 1H), 3.10 (t, *J* = 6.2 Hz, 2H), 2.29 (s, 3H), 1.14 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ : 168.2, 146.5, 146.3, 137.7, 130.4, 128.1, 125.6, 123.5, 65.4, 64.9, 57.9, 33.8, 20.7, 20.0; HRMS (ESI): calculated for C₁₂H₁₇N₂O₅⁺: m/z = 269.1137 [M+H]⁺; found: m/z = 269.1140 [M+H]⁺.

The analytical data is in agreement with the literature.¹

Compound 5



Yield: 75%; **IR**: $\tilde{v} = 3400$ (w), 2931 (s), 2894 (s), 2858 (s), 1730 (s), 1510 (vs), 1465 (s), 1300 (vs), 1258 (s), 1167 (s), 1010 (s), 895 (w), 832 (s) cm⁻¹; ¹H **NMR (400 MHz, DMSO-***d***₆) \delta**: 8.34 (br. s, 3H), 8.16 (d, J = 8.9 Hz, 2H), 7.58 (d, J = 8.9 Hz, 2H), 7.50 (d, J = 8.0 Hz, 2H), 7.12 (d, J = 8.0 Hz, 2H), 4.49 – 4.36 (m, 2H), 4.11 (br. s, 1H), 3.74 – 3.73 (m, 2H), 3.08 (t, J = 6.4 Hz, 2H), 2.28 (s, 3H); ¹³C **NMR (101 MHz, DMSO-***d***₆) \delta**: 168.0, 146.3, 138.0, 130.4, 128.2, 125.6, 123.5, 65.4, 59.5, 54.2, 33.9, 20.9; **HRMS (ESI)**: calculated for C₁₁H₁₅N₂O₅⁺: m/z = 255.0981 [M+H]⁺; found: m/z = 255.0977 [M+H]⁺.

Compound 6



Yield: 82 %; **IR**: $\tilde{v} = 3402$ (w), 2933 (s), 2894 (s), 2858 (s), 1730 (s), 1689 (s), 1514 (vs), 1469 (s), 1310 (vs), 1258 (s), 1167 (s), 1010 (s), 895 (w), 834 (s) cm⁻¹; ¹H NMR (400 MHz, DMSOd₆) δ : 8.42 (s, 3H), 8.16 – 8.11 (m, 4H), 7.55 – 7.48 (m, 6H), 7.12 (d, J = 7.9 Hz, 2H), 4.37 (t, J = 6.3 Hz, 2H), 4.34 – 4.30 (m, 1H), 4.26 (q, J = 6.3 Hz, 2H), 3.01 (q, J = 6.5 Hz, 4H), 2.85 (qd, J = 17.5, 5.1 Hz, 2H), 2.28 (s, 3H); ¹³C NMR (101 MHz, DMSO-d₆) δ : 169.0, 168.1, 146.4, 146.3, 145.3, 138.0, 130.3, 130.2, 128.2, 125.6, 123.5, 123.5, 65.7, 64.7, 48.4, 34.1, 33.8, 33.7, 20.8; HRMS (ESI): calculated for C₂₀H₂₂N₃O₈⁺: m/z = 432.1401 [M+H]⁺; found: m/z = 432.1405 [M+H]⁺.

General procedure of hydroxy group protection with TBSCl

The reaction was performed according to the procedure published before.¹

Npe-protected ester (1 eq.) was dissolved in pyridine and treated with one half of TBSCl (3 eq.) and 1*H*-imidazole (3 eq.). After 10 min, the second half was added, and the reaction mixture was left to stir at room temperature overnight. The mixture was diluted with CH_2Cl_2 and washed successively with sat. NaHCO₃ solution and H₂O. The organic layer was dried, evaporated and purified by flash chromatography eluting with $CH_2Cl_2/MeOH$ (10/1, v/v) to afford the target compound as an oil.



Yield: 94%; IR: $\tilde{v} = 3854$ (w), 3745 (w), 2930 (w), 2856 (w), 1735 (vs), 1601 (s), 1518 (vs), 1472 (w), 1463 (w), 1374 (w), 1344 (vs), 1251 (s), 1155 (s), 1076 (s), 967 (s), 835 (s), 775 (s), 747 (s) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 8.16 (d, J = 8.6 Hz, 2H), 7.38 (d, J = 8.6 Hz, 2H), 4.41 (dt, J = 11.0, 6.8 Hz, 1H), 4.29 – 4.16 (m, 2H), 3.24 (d, J = 2.8 Hz, 1H), 3.06 (t, J = 6.8 Hz, 2H), 1.20 (d, J = 6.3 Hz, 3H), 0.80 (s, 9H), -0.01 (s, 3H), -0.10 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ : 174.3, 147.0, 145.6, 129.9, 123.9, 69.6, 64.5, 60.9, 35.0, 25.7, 21.0, 17.9, -4.2, -5.2; HRMS (ESI): calculated for C₁₈H₃₁N₂O₅Si⁺: m/z = 383.2002 [M+H]⁺; found: m/z = 383.1997 [M+H]⁺.

The analytical data is in agreement with the literature.¹

Compound 8



Yield: 96%; **IR**: $\tilde{v} = 3854$ (w), 3745 (w), 2955 (w), 2930 (w), 2856 (w), 1735 (vs), 1601 (s), 1518 (vs), 1472 (w), 1374 (w), 1344 (vs), 1251 (s), 1155 (s), 1075 (s), 967 (s), 855 (w), 835 (s), 775 (s), 747 (s) cm⁻¹; ¹**H NMR (400 MHz, CDCl₃)** δ : 8.13 (d, J = 8.6 Hz, 2H), 7.36 (d, J = 8.6 Hz, 2H), 4.34 (q, J = 6.7 Hz, 2H), 3.82 (dd, J = 9.8, 3.7 Hz, 1H), 3.73 (dd, J = 9.8, 3.0 Hz, 1H), 3.47 (s, 1H), 3.04 (t, J = 6.7 Hz, 2H), 0.81 (s, 9H); ¹³**C NMR (101 MHz, CDCl₃)** δ : 173.9, 146.9, 145.6, 129.8, 123.8, 65.4, 64.4, 56.5, 34.9, 25.7, 18.2, -5.5, -5.6; **HRMS (ESI)**: calculated for C₁₇H₂₉N₂O₅Si⁺: m/z = 369.1846 [M+H]⁺; found: m/z = 369.1842 [M+H]⁺.

General procedure for npe-ester formation of Boc-protected amino acid

Boc-amino acid (1 eq.) was dissolved in CH_2Cl_2 under inert atmosphere and cooled to 0 °C. Then npeOH (1.3 eq.) and PPh₃ (1.3 eq.) were added followed by slow addition of DIAD (1.3 eq.). The reaction mixture was left to stir for 2 h at room temperature. Then the solution was washed with water, organic phase was dried over Na₂SO₄ and evaporated. The crude product was purified by flash chromatography to afford the target product.



Eluent: Hex/EtOAc (4/1, v/v).

Yield: 96%; **IR**: $\tilde{v} = 3397$ (w), 2975 (w), 1751 (s), 1709 (vs), 1519 (vs), 1391 (w), 1366 (w), 1345 (vs), 1159 (vs), 1056 (w), 905 (vs), 723 (vs) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 8.15 (d, J = 8.6 Hz, 2H), 7.38 (d, J = 8.6 Hz, 2H), 4.95 (d, J = 8.7 Hz, 1H), 4.38 (t, J = 6.6 Hz, 2H), 4.15 (dd, J = 9.0, 4.8 Hz, 1H), 3.06 (t, J = 6.6 Hz, 2H), 2.05 – 1.97 (m, 1H), 1.41 (s, 9H), 0.88 (d, J = 6.8 Hz, 3H), 0.78 (d, J = 6.8 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ : 172.4, 155.7, 146.9, 145.5, 129.8, 123.8, 79.9, 64.5, 58.6, 34.9, 31.2, 28.4, 19.0, 17.6; HRMS (ESI): calculated for C₁₈H₂₇N₂O₆⁺: m/z = 367.1869 [M+H]⁺; found: m/z = 367.1874 [M+H]⁺.

Compound 13



Eluent: Hex/EtOAc (3/1, v/v).

Yield: 85%; IR: $\tilde{v} = 3396$ (w), 2978 (w), 2254 (w), 1750 (s), 1707 (vs), 1601 (w), 1518 (vs), 1391 (w), 1366 (w), 1345 (vs), 1250 (w), 1159 (vs), 1056 (w), 905 (vs), 727 (vs) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 8.10 (d, J = 8.6 Hz, 2H), 7.38 (d, J = 8.6 Hz, 2H), 4.97 (d, J = 9.2 Hz, 1H), 4.39 (t, J = 6.6 Hz, 2H), 3.87 (d, J = 5.8 Hz, 2H), 3.06 (t, J = 6.6 Hz, 2H), 1.43 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ : 170.3, 155.8, 147.0, 145.4, 129.9, 123.9, 80.3, 64.7, 42.4, 34.9, 28.4; HRMS (ESI): calculated for C₁₅H₂₁N₂O₆⁺: m/z = 325.1400 [M+H]⁺; found: m/z = 325.1398 [M+H]⁺.

The analytical data is in agreement with the literature.²

Compound 14a



Eluent: Hex/EtOAc (4/1, v/v).

Yield: 81%; IR: $\tilde{v} = 3426$ (w), 3356 (w), 1740 (w), 1709 (vs), 1602 (w), 1518 (vs), 1495 (s), 1344 (vs), 1249 (w), 1159 (vs), 1056 (w), 855 (s), 733 (s), 698 (vs) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 8.15 (d, J = 8.7 Hz, 2H), 7.31 (d, J = 8.7 Hz, 2H), 7.25 – 7.20 (m, 3H), 7.04 (d, J = 5.6 Hz, 2H), 4.90 (d, J = 8.4 Hz, 1H), 4.54 (q, J = 6.4 Hz, 1H), 4.39 – 4.25 (m, 2H), 3.07 – 2.93 (m, 4H), 1.41 (s, 9H); ¹³C NMR (101 MHz, CD₂Cl₂) δ : 171.8, 155.0, 146.9, 145.3, 135.8, 129.8, 129.2, 128.6, 127.1, 123.8, 80.1, 64.7, 54.5, 38.4, 34.7, 28.3; HRMS (ESI): calculated for C₂₂H₂₆N₂O₆Na⁺: m/z = 437.1683 [M+Na]⁺; found: m/z = 437.1684 [M+Na]⁺.

Compound 14b



Eluent: Hex/EtOAc (4/1, v/v).

Yield: 85%; IR: $\tilde{v} = 3443$ (w), 3375 (w), 2977 (w), 2929 (w), 1740 (w), 1709 (vs), 1602 (w), 1517 (vs), 1495 (s), 1343 (vs), 1248 (w), 1157 (vs), 1055 (w), 855 (s), 747 (s), 698 (vs) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 8.16 (d, J = 8.7 Hz, 2H), 7.33 (d, J = 8.7 Hz, 2H), 7.30 – 7.20 (m, 3H), 7.07 (d, J = 6.2 Hz, 2H), 4.97 (d, J = 8.4 Hz, 1H), 4.56 (q, J = 6.2 Hz, 1H), 4.40 – 4.27 (m, 2H), 3.08 – 2.93 (m, 4H), 1.43 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ : 171.8, 155.1, 146.9, 145.4, 135.9, 129.8, 129.2, 128.6, 127.1, 123.8, 80.0, 64.7, 54.5, 38.4, 34.7, 28.3; HRMS (ESI): calculated for C₂₂H₃₀N₃O₆⁺: m/z = 432.2129 [M+NH₄]⁺; found: m/z = 432.2131 [M+NH₄]⁺.



Boc-histidine (0.5 g, 1.96 mmol, 1 eq.), 2-(4-nitrophenyl)ethanol (0.655 g, 3.92 mmol, 2 eq.), DMAP (0.048 g, 0.39 mmol, 0.20 eq.) and HBTU (0.967 g, 2.55 mmol, 1.3 eq.) were dissolved in DMF (4 ml) under inert atmosphere. Diisopropylamine (686 μ l, 4.90 mmol, 2.5 eq.) was added dropwise and the reaction mixture was stirred overnight at room temperature. The resulting solution was diluted with EtOAc (30 ml) and quenched with saturated NH₄Cl solution (15 ml). The organic layer was washed with water, dried and the solvents were removed *in vacuo*. The crude product was purified by flash chromatography on silica gel (2% to 5% MeOH in DCM) to obtain the target product as a pale-yellow foam.

Yield: 93%; **IR**: $\tilde{v} = 2977$ (w), 1699 (vs), 1600 (s), 1516 (vs), 1391 (w), 1365 (w), 1344 (vs), 1250 (w), 1160 (vs), 1108 (w), 1054 (w), 1016 (w), 855 (vs), 748 (w), 697 (w) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 8.18 – 8.07 (m, 2H), 7.58 (s, 1H), 7.41 – 7.30 (m, 2H), 6.72 (s, 1H), 5.77 (d, J = 8.2 Hz, 1H), 4.51 (q, J = 6.2 Hz, 1H), 4.34 (t, J = 6.6 Hz, 2H), 3.06 – 3.00 (m, 4H), 1.41 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ : 172.1, 155.7, 147.1, 145.7, 135.2, 134.2, 130.0, 123.9, 115.9, 80.2, 64.9, 53.6, 34.9, 29.7, 28.5; HRMS (ESI): calculated for C₁₉H₂₅N₄O₆⁺: m/z = 405.1769 [M+H]⁺; found: m/z = 405.1765 [M+H]⁺.

General procedure for deprotection of Boc-protecting group

Npe-protected amino acid was dissolved in 4M HCl/Dioxane mixture at 0 °C. The reaction mixture was left to stir for 2 h and afterwards was evaporated to dryness. The resulting product was used for further steps without additional purification.

Compound 15



Yield: 99%; **IR**: $\tilde{v} = 3335$ (w), 2964 (w), 2850 (w), 1741 (s), 1604 (w), 1516 (vs), 1464 (w), 1379 (vs), 1232 (vs), 1215 (s), 1170 (w), 1043 (w), 969 (w), 857 (s), 751 (s), 700 (s) cm⁻¹; ¹H

NMR (400 MHz, DMSO-*d*₆**) \delta**: 8.66 (br. s, 3H), 8.17 (d, *J* = 8.6 Hz, 2H), 7.62 (d, *J* = 8.6 Hz, 2H), 4.46 (dtd, *J* = 23.3, 11.1, 6.3 Hz, 2H), 3.76 (d, *J* = 4.5 Hz, 1H), 3.11 (t, *J* = 6.3 Hz, 2H), 2.10 (tt, *J* = 11.6, 5.8 Hz, 1H), 0.84 (d, *J* = 3.0 Hz, 3H), 0.82 (d, *J* = 3.0 Hz, 3H). ¹³C **NMR (101 MHz, DMSO-***d*₆**)** δ : 168.8, 146.3, 130.4, 123.4, 65.3, 57.2, 33.8, 29.2, 18.3, 17.4 **HRMS (ESI)**: calculated for C₁₃H₁₉N₂O₄⁺: m/z = 267.1339 [M+H]⁺; found: m/z = 267.1139 [M+H]⁺.

Compound 16



Yield: 99%; IR: $\tilde{v} = 2949$ (w), 1746 (s), 1515 (vs), 1310 (vs), 1238 (vs), 1053 (w), 955 (s), 905 (s) 856 (s), 698 (s) cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ : 8.30 (br. s, 3H), 8.18 (d, *J* = 8.7 Hz, 2H), 7.59 (d, *J* = 8.7 Hz, 2H), 4.44 (t, *J* = 6.4 Hz, 2H), 3.77 (s, 2H), 3.09 (t, *J* = 6.4 Hz, 2H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ : 167.6, 156.2, 146.3, 130.4, 123.5, 67.9, 33.9, 28.2, 22.0; HRMS (ESI): calculated for C₁₀H₁₃N₂O₄⁺: m/z = 225.0870 [M+H]⁺; found: m/z = 225.0868 [M+H]⁺. The analytical data is in agreement with the literature.²

Compound 17a



Yield: 99%; IR: $\tilde{v} = 3142$ (w), 2988 (w), 2802 (w), 1740 (vs), 1601 (s), 1518 (vs), 1490 (vs), 1351 (vs), 1232 (vs), 1191 (s), 1102 (s), 981 (s), 856 (vs), 755 (vs), 736 (vs), 706 (vs) cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ : 8.74 (br. s, 3H), 8.16 (d, *J* = 8.7 Hz, 2H), 7.49 (d, *J* = 8.7 Hz, 2H), 7.33 – 7.21 (m, 3H), 7.12 (dd, *J* = 7.8, 1.8 Hz, 2H), 4.33 (t, *J* = 6.3 Hz, 2H), 4.20 (dd, *J* = 7.8, 5.5 Hz, 1H), 3.16 (dd, *J* = 14.0, 5.5 Hz, 1H), 3.06 – 2.89 (m, 3H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ : 168.9, 146.3, 134.9, 130.4, 129.4, 128.5, 127.2, 123.4, 65.4, 53.3, 35.8, 33.7; HRMS (ESI): calculated for C₁₇H₁₉N₂O₄⁺: m/z = 315.1339 [M+H]⁺; found: m/z = 315.1332 [M+H]⁺.

Compound 17b



Yield: 99%; **IR**: $\tilde{v} = 3146$ (w), 2988 (w), 2802 (w), 1740 (vs), 1602 (s), 1518 (vs), 1490 (vs), 1351 (vs), 1232 (vs), 1192 (s), 1102 (s), 856 (vs), 755 (vs), 736 (vs), 705 (vs) cm⁻¹; ¹H NMR **(400 MHz, DMSO-***d*₆**)** δ : 8.91 (br. s., 3H), 8.12 (d, J = 8.7 Hz, 2H), 7.46 (d, J = 8.7 Hz, 2H), 7.27 – 7.20 (m, 3H), 7.12 (d, J = 6.4 Hz, 2H), 4.29 (t, J = 6.4 Hz, 2H), 4.19 – 4.08 (m, 1H), 3.21 (dd, J = 14.1, 5.0, 1H), 3.10 – 2.88 (m, 3H); ¹³C NMR (101 MHz, DMSO-*d*₆**)** δ : 168.9, 146.3, 134.8, 130.4, 129.4, 128.5, 127.2, 123.4, 65.4, 53.3, 35.8, 33.7; HRMS (ESI): calculated for C₁₇H₁₉N₂O₄⁺: m/z = 315.1339 [M+H]⁺; found: m/z = 315.1332 [M+H]⁺.

Compound 21



Yield: 98%; **IR**: $\tilde{v} = 2960$ (w), 1737 (vs), 1624 (w), 1598 (w), 1516 (vs), 1454 (w), 1415 (w), 1351 (vs), 1282 (w), 1188 (w), 1124 (vs), 1045 (w), 1008 (w), 854 (s), 774 (w), 749 (w), 701 (w) cm⁻¹; ¹**H NMR (400 MHz, DMSO-***d*₆**)** δ : 8.69 (s, 3H), 8.21 – 8.15 (m, 2H), 7.59 – 7.55 (m, 2H), 7.51 (s, 1H), 6.04 (s, 2H), 4.39 (m, 3H), 3.56 (s, 1H), 3.17 (d, *J* = 6.9 Hz, 2H), 3.04 (td, *J* = 6.4, 3.9 Hz, 2H), 1.13 (s, 9H); ¹³**C NMR (101 MHz, DMSO-***d*₆**)** δ : 176.7, 168.2, 146.4, 146.1, 137.4, 130.4, 123.5, 120.0, 69.2, 66.4, 65.6, 51.1, 38.2, 33.7, 26.5; **HRMS (ESI)**: calculated for C₂₀H₂₇N₄O₆⁺: m/z = 419.1925 [M+H]⁺; found: 419.1919 [M+H]⁺.

Compound 20



The histidine derivative **19** (0.745 g, 1.84 mmol, 1 eq.) was dissolved in DMF (12 mL) under N₂ atmosphere at 0 °C. Subsequently, K_2CO_3 (0.509 g, 3.68 mmol, 2 eq.) was added and the mixture was stirred for 40 min. Chloromethylpivalate (319 µl, 2.21 mmol, 1.2 eq.) was added dropwise at 0°C and the reaction mixture was left to warm to room temperature while stirring for 5 h. Catalytic amounts of KI were added and the mixture was stirred for another 1 h. The resulting suspension was diluted with EtOAc (75 ml) and quenched with saturated NH₄Cl solution (35 ml). The organic phase was washed with water, dried over Na₂SO₄ and concentrated *in vacuo*. The crude mixture was purified by flash chromatography on silica gel (40% isohexane in EtOAc to pure EtOAc). The pivalate protected histidine derivative was obtained as a yellow oil.

Yield: 55%; IR: 2850 (w), 2600 (w), 1738 (vs), 1624 (w), 1598 (w), 1573 (w), 1516 (vs), 1454 (w), 1414 (w), 1350 (vs), 1282 (w), 1191 (w), 1124 (vs), 1044 (w), 1008 (w), 854 (vs), 773 (w), 749 (w), 701 (w) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ: 8.19 – 8.14 (m, 2H), 7.54 (d, J = 1.4 Hz, 1H), 7.42 – 7.36 (m, 2H), 6.71 (s, 1H), 5.81 (d, J = 8.2 Hz, 1H), 5.73 (s, 2H), 4.51 (dt, J = 8.2, 5.3 Hz, 1H), 4.34 (t, J = 6.7 Hz, 2H), 3.04 (t, J = 6.7 Hz, 2H), 2.97 (t, J = 5.2 Hz, 2H), 1.42 (s, 9H), 1.14 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ: 177.9, 172.0, 155.7, 147.0, 145.8, 138.3, 130.0, 123.9, 117.3, 79.9, 77.4, 67.7, 64.7, 53.5, 38.9, 35.0, 30.1, 28.5, 27.0; HRMS (ESI): calculated for C₂₅H₃₅N₄O₈⁺: m/z = 519.2449 [M+H]⁺; found: m/z = 519.2441 [M+H]⁺.

Compound 22

The reaction was conducted according to a published procedure.³

Phenyl chloroformate (4 ml, 31.9 mmol, 1 eq.) was dissolved in dry CH_2Cl_2 under nitrogen and cooled to 0 °C. Then *N*-methylimidazole (2.54 ml, 31.9 mmol, 1 eq.) was added dropwise. The mixture was allowed to stir at room temperature for 2 hours. Afterwards the reaction mixture was filtered, the precipitate was washed with CH_2Cl_2 and dried.

Yield: 95%; **IR**: $\tilde{v} = 2926$ (w), 1783 (vs), 1588 (w), 1536 (w), 1372 (s), 1330 (s), 1232 (vs), 749 (vs), 689 (s) cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ : 10.29 (s, 1H), 8.37 (s, 1H), 8.02 (s, 1H), 7.43 - 7.58 (m, 5H), 4.01 (s, 3H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ : 157.5, 135.6, 129.3, 123.1, 121.3, 119.5, 118.6, 115.3, 35.4.

The analytical data is in agreement with the literature.³



Compound was synthesized following the procedure published earlier.⁴

Adenosine **23** (1 g, 3.74 mmol, 1 eq.) was suspended in DMF and di-*tert*-butylsilyl ditriflate (1.46 ml, 4.49 mmol, 1.2 eq.) was added dropwise under stirring at 0 °C. The resulting solution was stirred at 0°C for 45 min. Then imidazole (1.27 g, 18.7 mmol, 5 eq.) was added and the reaction was warmed to room temperature over a period of 30 min. Then TBSC1 (0.68 g, 4.49 mmol, 1.2 eq.) was added and the reaction was heated to 60 °C overnight. Subsequently, the reaction mixture was diluted with EtOAc and washed with water and brine. The organic layer was dried and evaporated. The residue was purified by flash chromatography (Hex/EtOAc, 1/1, v/v).

Yield: 76%; **IR**: $\tilde{v} = 3148$ (w), 2933 (w), 2859 (w), 2361 (w), 1677 (s), 1604 (s), 1598 (w), 1576 (w), 1473 (w), 1426 (w), 1363 (w), 1329 (w), 1302 (w), 1258 (w), 1200 (w), 1166 (w), 1136 (w), 1105 (w), 1064 (vs), 1009 (s), 890 (w), 828 (vs), 786 (w), 754 (w), 729 (w) cm⁻¹; ¹H NMR (400 **MHz, CDCl₃**) δ : 8.31 (s, 1H), 7.83 (s, 1H), 6.12 (br. s, 2H), 5.91 (s, 1H), 4.61 (d, J = 4.7 Hz, 1H), 4.50 (ddd, J = 16.5, 9.3, 4.7 Hz, 2H), 4.25 – 4.17 (m, 1H), 4.03 (dd, J = 10.5, 9.3 Hz, 1H), 1.07 (s, 9H), 1.04 (s, 9H), 0.92 (s, 9H), 0.16 (s, 3H), 0.14 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ : 155.5, 152.8, 149.3, 138.9, 120.4, 92.6, 75.9, 75.6, 74.8, 67.9, 27.6, 27.1, 26.0, 22.9, 20.5, 18.4, -4.2, -4.8; HRMS (ESI): calculated for C₂₄H₄₄N₅O₄Si₂⁺: m/z = 522.2932 [M+H]⁺; found: m/z = 522.2926 [M+H]⁺.

The analytical data is in agreement with the literature.⁴

General procedure for amino acid attachment to the adenosine derivative



The silyl-protected adenosine derivative 24 (1 eq.) was dissolved in dry CH_2Cl_2 under nitrogen atmosphere. 1-*N*-methyl-3-phenoxycarbonyl-imidazolium chloride (22, 2 eq.) was added to the reaction mixture and the resulting suspension was stirred at room temperature for 2 hours (the solution in time becomes clear). Afterwards the protected amino acid (2 eq.) was added together with TEA (2 eq.) as a solution in CH_2Cl_2 and the resulting solution was stirred overnight at room temperature. The reaction was quenched by addition of saturated aqueous NaHCO₃ solution. The solution was extracted three times with CH_2Cl_2 , and the organic phase was dried, filtered and concentrated *in vacuo*. The residue was purified by flash chromatography eluting with Hex/EtOAc to give product as white foam.

Compound 25



Eluent: Hex/EtOAc (4/3, v/v).

Yield: 91%; **IR**: $\tilde{v} = 3237$ (w), 2931 (s), 2857 (s), 1737 (s), 1701 (vs), 1610 (s), 1520 (vs), 1465 (s), 1345 (s), 1250 (s), 1136 (w), 1057 (s), 998 (w), 894 (w), 840 (s), 777 (s) cm⁻¹; ¹H NMR (400 **MHz, CDCl₃)** δ : 10.05 (d, J = 9.0 Hz, 1H), 8.56 (s, 1H), 8.41 (s, 1H), 8.16 (s, 1H), 7.94 (d, J = 8.6 Hz, 2H), 7.32 (d, J = 8.6 Hz, 2H), 5.99 (s, 1H), 4.64 (d, J = 4.6 Hz, 1H), 4.60-4.46 (m, 3H), 4.33-4.24 (m, 2H), 4.20-4.29 (m, 1H), 4.05 (dd, J = 10.5, 9.1 Hz, 1H), 3.03 (t, J = 6.5 Hz, 2H), 1.25 (d, J = 6.5 Hz, 3H), 1.08 (s, 9H), 1.05 (s, 9H), 0.95 (s, 9H), 0.90 (s, 9H), 0.19 (s, 3H), 0.16 (s, 3H), 0.07 (s, 3H), -0.04 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ : 171.1, 154.6, 151.3, 150.3, 149.8, 146.8, 145.7, 141.6, 129.9, 123.7, 121.1, 92.6, 76.0, 75.7, 75.0, 68.8, 68.0, 64.8, 59.8, 35.0, 27.7, 27.2, 26.1, 25.7, 22.9, 21.3, 20.6, 18.5, 18.0, -4.1, -4.8, -5.2; HRMS (ESI): calculated for C₄₃H₇₂N₇O₁₀Si₃⁺: m/z = 930.4643 [M+H]⁺; found: m/z = 930.4640 [M+H]⁺.



Eluent: 10% CH₂Cl₂ in EtOAc to pure EtOAc.

Yield: 86%; **IR**: $\tilde{v} = 3854$ (w), 3745 (w), 3650 (w), 2932 (w), 2858 (w), 2361 (w), 2341 (w), 1735 (s), 1670 (s), 1654 (w), 1610 (w), 1587 (w), 1521 (vs), 1472 (s), 1395 (w), 1345 (vs), 1252 (w), 1166 (w), 1118 (vs), 1055 (vs), 999 (w), 894 (w), 826 (vs), 781 (s), 750 (w) cm⁻¹; ¹H NMR (400 MHz, CDCI₃) δ : 10.05 (dd, J = 7.6, 3.7 Hz, 1H), 8.45 (s, 1H), 8.08 (s, 1H), 8.06 – 8.02 (m, 3H), 7.59 (d, J = 1.3 Hz, 1H), 7.41 – 7.35 (m, 2H), 6.86 (dd, J = 5.3, 1.3 Hz, 1H), 5.96 (d, J = 7.0 Hz, 1H), 5.74 (d, J = 2.6 Hz, 2H), 4.94 – 4.85 (m, 1H), 4.60 (dd, J = 6.0, 4.6 Hz, 1H), 4.53 – 4.46 (m, 2H), 4.42 (tq, J = 6.5, 1.7 Hz, 2H), 4.24 (tdd, J = 9.8, 5.0, 2.8 Hz, 1H), 4.08 – 4.00 (m, 1H), 3.23 – 3.12 (m, 2H), 3.07 (t, J = 6.5 Hz, 2H), 1.09 (s, 9H), 1.08 (s, 9H), 1.05 (s, 9H), 0.94 (s, 9H), 0.17 (s, 3H), 0.15 (s, 3H); ¹³C NMR (101 MHz, CDCI₃) δ : 177.8, 171.7, 153.6, 151.3, 150.2, 149.8, 146.9, 145.8, 141.2, 138.4, 138.1, 123.0, 123.8, 121.1, 117.3, 92.6, 76.0, 75.7, 74.9, 67.9, 67.7, 64.7, 55.5, 38.8, 35.0, 30.7, 27.7, 27.2, 26.9, 26.1, 22.9, 20.5, 18.5, 1.3, -4.2; HRMS (ESI): calculated for C₄₅H₆₈N₉O₁₁Si₂⁺: m/z = 966.4571 [M+H]⁺; found: m/z = 966.4576 [M+H]⁺.

Compound 27



Eluent: Hex/EtOAc (4/3, v/v).

Yield: 78%; IR: $\tilde{v} = 3230$ (w), 2960 (w), 2960 (w), 2858 (w), 1741 (s), 1702 (vs), 1611 (s), 1520 (vs), 1466 (s), 1345 (vs), 1250 (s), 1139 (vs), 1057 (vs), 999 (s), 894 (s), 810 (vs), 781 (s) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 10.01 (d, J = 8.4 Hz, 1H), 8.50 (s, 1H), 8.17 (s, 1H), 8.07 (d, J = 8.3 Hz, 2H), 7.28 (d, J = 8.3 Hz, 2H), 5.98 (s, 1H), 4.61 (d, J = 4.6 Hz, 1H), 4.56 – 4.38 (m, 5H), 4.20-4.29 (m, 1H), 4.07 (dd, J = 10.5, 9.1 Hz, 1H), 3.09 (t, J = 6.5 Hz, 2H), 2.28 – 2.21 (m, 1H), 1.08 (s, 9H), 1.05 (s, 9H), 1.00 (d, J = 6.5 Hz, 3H), 0.95 (s, 12H), 0.18 (s, 3H), 0.16 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ : 171.1, 154.1, 152.2, 151.1, 149.8, 146.9, 145.6, 141.5, 129.9, 126.4, 123.7, 121.1, 92.6, 75.9, 75.7, 74.9, 67.9, 64.8, 58.8, 35.0, 30.9, 27.7, 27.2, 26.1, 22.9, 20.4, 19.5, 18.4, 18.0, -4.2, -4.9; HRMS (ESI): calculated for C₃₈H₆₀N₇O₉Si₂⁺: m/z = 814.3991 [M+H]⁺; found: m/z = 814.3976.

Compound 28



Eluent: Hex/EtOAc (4/3, v/v).

Yield: 72%; IR: $\tilde{v} = 3239$ (w), 2932 (s), 2858 (s), 1749 (s), 1703 (vs), 1611 (s), 1520 (vs), 1468 (s), 1345 (s), 1252 (s), 1141 (w), 1055 (s), 990 (w), 894 (w), 826 (s), 750 (s) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 9.95 (br. s, 1H), 8.50 (s, 1H), 8.21 – 8.03 (m, 3H), 7.38 (d, J = 8.6 Hz, 2H), 5.98 (s, 1H), 4.60 (d, J = 4.7 Hz, 1H), 4.57 – 4.40 (m, 4H), 4.30 – 4.17 (m, 3H), 4.10 – 4.03 (m, 1H), 3.09 (t, J = 6.6 Hz, 2H), 1.08 (s, 9H), 1.05 (s, 9H) 0.94 (s, 9H), 0.17 (s, 3H), 0.16 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ : 170.0, 154.2, 151.2, 150.2, 149.9, 147.0, 145.5, 141.4, 129.9, 123.8, 121.1, 92.5, 76.0, 75.7, 74.9, 67.9, 42.2, 35.0, 27.6, 27.2, 26.0, 22.9, 20.5, 18.5, -4.1, -4.9; HRMS (ESI): calculated for C₃₅H₅₄N₇O₉Si₂⁺: m/z = 772.3522 [M+H]⁺; found: m/z = 772.3504 [M+H]⁺.



Eluent: Hex/EtOAc (2/1, v/v).

Yield: 87%; **IR**: $\tilde{v} = 3229$ (w), 2953 (w), 2929 (w), 2857 (w), 1735 (s), 1693 (s), 1607 (s), 1589 (s), 1517 (vs), 1469 (s), 1391 (w), 1310 (vs), 1292 (w), 1251 (w), 1210 (w), 835 (s) cm⁻¹; ¹**H NMR (400 MHz, CDCl₃)** δ : 10.28 (s, 1H), 8.42 (s, 1H), 8.22 – 8.12 (m, 1H), 8.07 (d, J = 8.6Hz, 2H), 7.96 (d, J = 8.6 Hz, 2H), 7.37 – 7.30 (m, 4H), 6.00 (s, 1H), 4.93 – 4.89 (m, 1H), 4.64 (d, J = 4.6 Hz, 1H), 4.51 (ddd, J = 9.2, 4.9, 2.7 Hz, 2H), 4.46 – 4.40 (m, 2H), 4.39 – 4.22 (m, 3H), 4.09 – 4.05 (m, 1H), 3.08 – 2.94 (m, 6H), 1.07 (s, 9H), 1.05 (s, 9H), 0.95 (s, 9H), 0.19 (s, 3H), 0.17 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ : 179.8, 170.7, 153.7, 149.8, 146.9, 146.8, 145.5, 145.4, 129.8, 123.8, 123.6, 120.9, 92.6, 75.9, 75.7, 74.9, 67.9, 65.0, 64.5, 49.7, 36.5, 34.9, 34.8, 27.6, 27.1, 26.0, 22.8, 20.5, 18.4, -4.2, -4.9; **HRMS (ESI)**: calculated for C₄₅H₆₃N₈O₁₃Si₂⁺: m/z = 979.4053 [M+H]⁺; found: m/z = 979.4056.

Compound 30a



Yield: 68%; **IR**: $\tilde{v} = 3190$ (w), 2933 (w), 2858 (w), 1742 (w), 1702 (vs), 1612 (s), 1587 (w), 1521 (vs), 1469 (vs), 1345 (vs), 1253 (s), 1057 (s), 1000 (w), 828 (vs) cm⁻¹; ¹H NMR (400 MHz, CD₂Cl₂) δ : 10.00 (d, J = 7.5 Hz, 1H), 8.94 (s, 1H), 8.37 (d, J = 1.2 Hz, 2H), 8.00 (d, J = 7.5 Hz, 1H), 8.94 (s, 1H), 8.37 (d, J = 1.2 Hz, 2H), 8.00 (d, J = 7.5 Hz, 1H), 8.94 (s, 1H), 8.97 (d, J = 1.2 Hz, 2H), 8.00 (d, J = 7.5 Hz, 1H), 8.94 (s, 1H), 8.97 (d, J = 1.2 Hz, 2H), 8.00 (d, J = 1.2 Hz, 8.00 (d, J =

8.7 Hz, 2H), 7.39 – 7.26 (m, 5H), 7.21 – 7.17 (m, 2H), 6.02 (s, 1H), 4.89 – 4.76 (m, 1H), 4.60 (d, J = 4.6 Hz, 1H), 4.54 – 4.46 (m, 2H), 4.40 (td, J = 6.5 Hz, 1.7 Hz, 2H), 4.31 – 4.22 (m, 1H), 4.16 – 4.06 (m, 1H), 3.18 (d, J = 6.8 Hz, 2H), 3.03 (t, J = 6.5 Hz, 2H), 1.10 (s, 9H), 1.06 (s, 9H), 0.96 (s, 9H), 0.20 (s, 3H), 0.18 (s, 3H); ¹³C NMR (101 MHz, CD₂Cl₂) δ : 171.5, 153.8, 150.7, 150.2, 149.8, 146.7, 145.9, 142.2, 136.4, 129.8, 129.5, 128.5, 127.1, 123.4, 120.8, 92.2, 75.8, 75.7, 74.8, 67.7, 64.6, 54.9, 37.8, 34.7, 27.3, 26.9, 25.7, 22.6, 20.2, 18.2, -4.6, -5.2; HRMS (ESI): calculated for C₄₂H₆₀N₇O₉Si₂⁺: m/z = 862.3986 [M+H]⁺; found: m/z = 862.3995 [M+H]⁺.

Compound 30b



Eluent: Hex/EtOAc (4/1, v/v).

Yield: 71%; **IR**: $\tilde{v} = 3192$ (w), 2933 (w), 2858 (w), 1740 (w), 1700 (vs), 1611 (s), 1520 (vs), 1466 (vs), 1345 (vs), 1252 (s), 1166 (w), 1139 (w), 1054 (vs), 998 (s), 893 (s), 826 (vs), 736 (vs) cm⁻¹; ¹**H NMR (400 MHz, CD₂Cl₂)** δ : 10.01 (d, J = 7.6 Hz, 1H), 8.96 (s, 1H), 8.37 (s, 2H), 8.00 (d, J = 8.7 Hz, 1H), 7.37 – 7.22 (m, 5H), 7.22 – 7.14 (m, 2H), 6.03 (s, 1H), 4.90 – 4.81 (m, 1H), 4.61 (d, J = 4.5 Hz, 1H), 4.54 – 4.45 (m, 2H), 4.39 (t, J = 6.4 Hz, 2H), 4.32 – 4.21 (m, 1H), 4.16 – 4.05 (m, 1H), 3.19 (d, J = 6.2 Hz, 2H), 3.02 (t, J = 6.4 Hz, 2H), 1.09 (s, 9H), 1.06 (s, 9H), 0.96 (s, 9H), 0.20 (s, 3H), 0.18 (s, 3H); ¹³C NMR (101 MHz, CD₂Cl₂) δ : 171.4, 153.6, 150.8, 150.2, 149.8, 146.7, 145.9, 141.8, 136.4, 129.8, 129.5, 128.5, 127.1, 123.4, 120.8, 92.2, 75.9, 75.7, 74.8, 67.7, 64.6, 54.9, 37.9, 34.7, 27.3, 26.8, 25.7, 22.6, 20.2, 18.2, -4.6, -5.3.; HRMS (ESI): calculated for C₄₂H₆₀N₇O₉Si₂⁺: m/z = 862.3986 [M+H]⁺; found: m/z = 862.3996 [M+H]⁺.



Eluent: Hex/EtOAc (1/1, v/v).

Yield: 85%; **IR**: $\tilde{v} = 3238$ (w), 2931 (s), 2859 (s), 1737 (s), 1701 (vs), 1610 (s), 1520 (vs), 1470 (s), 1345 (s), 1251 (s), 1136 (w), 1057 (s), 998 (w), 899 (w), 840 (s), 778 (s) cm⁻¹; ¹**H NMR (400 MHz, CDCl₃)** δ : 10.18 (d, J = 8.3 Hz, 1H), 8.61 (s, 1H), 8.44 (s, 1H), 8.19 (s, 1H), 8.02 (d, J = 8.6 Hz, 2H), 7.35 (d, J = 8.7 Hz, 2H), 5.99 (s, 1H), 4.72 (dt, J = 8.4, 2.8 Hz, 1H), 4.61 (d, J = 4.6 Hz, 1H), 4.50 (td, J = 9.8, 4.9 Hz, 2H), 4.43 (t, J = 6.5 Hz, 2H), 4.24 (td, J = 10.0, 5.0 Hz, 1H), 4.14 (dd, J = 10.1, 2.7 Hz, 1H), 4.07 (dd, J = 10.5, 9.1 Hz, 1H), 3.91 (dd, J = 10.1, 3.1 Hz, 1H), 3.06 (t, J = 6.5 Hz, 2H), 1.08 (s, 9H), 1.05 (s, 9H), 0.94 (s, 9H), 0.88 (s, 10H), 0.18 (s, 3H), 0.16 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ : 170.6, 153.9, 151.2, 150.3, 149.8, 146.9, 145.6, 141.6, 129.8, 123.7, 121.1, 92.5, 75.9, 75.7, 74.9, 67.9, 64.8, 64.6, 55.7, 34.9, 27.6, 27.1, 26.0, 25.7, 22.8, 20.5, 18.4, 18.2, -4.2, -4.8, -5.3, -5.6; HRMS (ESI): calculated for C₄₂H₇₀N₇O₁₀Si₃⁺: m/z = 916.4492 [M+H]⁺; found: m/z = 916.4501 [M+H]⁺.

General procedure for deprotection 3'-5'-silyl protecting group:



The modified adenosine (0.86 mmol) was dissolved in CH_2Cl_2 under N_2 atmosphere and transferred into a plastic flask. Pyridine (1 ml) was added and the solution was cooled in an ice-bath. Then $Py^*(HF)_n$ (140 µl) was added and the mixture was stirred at 0 °C for 2 h. The reaction was quenched with sat. NaHCO₃ and extracted with CH_2Cl_2 . Organic phase was washed with

water and dried over Na₂SO₄. The solvents were removed in vacuo. The crude product was purified by flash chromatography eluting with $CH_2Cl_2/MeOH$ (9/1, v/v) to afford the product as a colourless foam.

Compound 32



Yield: 95%; **IR**: $\tilde{v} = 3244$ (w), 2952 (w), 2929 (w), 2856 (w), 1736 (w), 1695 (s), 1610 (s), 1588 (s), 1520 (vs), 1469 (s), 1345 (vs), 1313 (w), 1250 (vs), 1129 (w), 1093 (s), 835 (vs), 760 (vs) cm⁻¹; ¹**H NMR (400 MHz, CDCl₃)** δ : 9.90 (d, J = 9.1 Hz, 1H), 8.50 (s, 1H), 8.46 (s, 1H), 8.10 – 8.08 (m, 3H), 7.37 (d, J = 8.6 Hz, 2H), 5.85 (d, J = 7.0 Hz, 1H), 5.10 (dd, J = 7.0, 4.7 Hz, 1H), 4.59 (dd, J = 9.1, 1.5 Hz, 1H), 4.53 – 4.41 (m, 2H), 4.41 – 4.34 (m, 2H), 4.34 – 4.26 (m, 1H), 3.98 (dd, J = 13.0, 1.5 Hz, 1H), 3.78 (dd, J = 13.0, 1.5 Hz, 1H), 3.07 (t, J = 6.7 Hz, 2H), 1.24 (d, J = 6.7 Hz, 3H), 0.89 (s, 9H), 0.81 (s, 9H), 0.05 (s, 3H), -0.06 (s, 3H), -0.14 (s, 3H), -0.34 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ : 170.9, 154.0, 151.3, 150.9, 149.3, 147.1, 145.6, 143.0, 130.0, 123.9, 91.5, 87.7, 74.9, 72.7, 68.8, 65.13, 63.3, 59.8, 35.0, 25.7, 21.3, 18.1, 18.0, -4.0, -5.0, -5.1, -5.2; HRMS (ESI): calculated for C₃₅H₅₆N₇O₁₀Si₂⁺: m/z = 790.3622 [M+H]⁺; found: m/z = 790.3612 [M+H]⁺.

Compound 33



Yield: 75%; **IR**: $\tilde{v} = 3854$ (w), 3745 (w), 3650 (w), 3230 (w), 2930 (w), 2857 (w), 2361 (w), 2341 (w), 1740 (s), 1699 (vs), 1611 (w), 1587 (w), 1520 (vs), 1472 (s), 1395 (w), 1345 (vs), 1253 (w), 1119 (vs), 1091 (w), 1058 (w), 1030 (w), 983 (w), 914 (w), 856 (vs), 779 (vs), 747(w), 670 (w) cm⁻¹; ¹**H NMR (400 MHz, CDCl₃)** δ : 10.05 – 9.98 (m, 1H), 8.50 (d, J = 3.1 Hz, 1H), 8.18 – 8.10 (m, 3H), 8.01 (d, J = 9.1 Hz, 1H), 7.60 (dd, J = 8.2, 1.3 Hz, 1H), 7.44 – 7.37 (m, 2H), 6.84 (d, J = 1.4 Hz, 1H), 5.99 – 5.93 (m, 1H), 5.81 (dd, J = 7.3, 5.4 Hz, 1H), 5.77 – 5.71 (m, 2H), 5.08 (dt, J = 7.3, 5.1 Hz, 1H), 4.90 (dt, J = 7.2, 5.4 Hz, 1H), 4.42 (q, J = 6.3 Hz, 2H), 4.39 – 4.34 (m, 2H), 3.98 – 3.92 (m, 1H), 3.82 – 3.72 (m, 1H), 3.23 – 3.13 (m, 2H), 3.08 (dt, J = 13.8, 6.6 Hz, 2H), 2.83 (s, 1H), 1.09 (s, 9H), 0.80 (s, 9H), -0.18 (s, 3H), -0.40 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ : 177.8, 171.8, 153.2, 151.1, 150.9, 149.3, 147.0, 145.8, 143.0, 138.2, 138.1, 130.0, 123.9, 123.1, 117.4, 91.4, 87.8, 74.7, 73.0, 67.7, 65.0, 63.5, 53.5, 38.8, 35.0, 30.6, 26.9, 25.7, 18.0, -5.1, -5.2; HRMS (ESI): calculated for C₃₇H₅₂N₉O₁₁Si⁺: m/z = 826.3550 [M+H]⁺; found: m/z = 826.3559 [M+H]⁺.

Compound 34



Yield: 95%; **IR**: $\tilde{v} = 3245$ (w), 2930 (w), 2857 (w), 1743 (s), 1695 (vs), 1590 (s), 1518 (vs), 1471 (s), 1344 (vs), 1252 (s), 1212 (w), 1188 (s), 1130 (w), 1085 (s), 836 (vs) cm⁻¹; ¹**H NMR (400 MHz, Acetone-***d***6)** δ : 10.01 (d, J = 8.4 Hz, 1H), 8.50 (s, 1H), 8.17 (s, 1H), 8.07 (d, J = 8.3 Hz, 2H), 7.28 (d, J = 8.3 Hz, 2H), 5.98 (s, 1H), 4.61 (d, J = 4.6 Hz, 1H), 4.56 – 4.38 (m, 6H), 4.20-4.29 (m, 1H), 4.07 (dd, J = 10.5, 9.1 Hz, 1H), 3.09 (t, J = 6.5 Hz, 2H), 1.00 (d, J = 6.5 Hz, 3H), 0.96 (d, J = 6.5 Hz, 3H), 0.95 (s, 9H), 0.18 (s, 3H), 0.16 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ : 172.2, 154.4, 151.6, 149.5, 147.0, 145.5, 143.3, 136.5, 129.9, 123.7, 122.0, 91.2, 87.6, 74.7, 72.8, 64.7, 63.4, 58.7, 35.0, 30.9, 25.6, 19.4, 17.9, -5.2, -5.3; HRMS (ESI): calculated for $C_{30}H_{44}N_7O_9Si^+$: m/z = 674.2970 [M+H]⁺; found: m/z = 674.2974 [M+H]⁺.



Yield: 97%; **IR**: $\tilde{v} = 3250$ (w), 2932 (w), 2859 (w), 1740 (s), 1680 (vs), 1595 (s), 1520 (vs), 1470 (s), 1340 (vs), 1254 (s), 1213 (w), 1168 (s), 1130 (w), 1085 (s), 836 (vs) cm⁻¹; ¹**H NMR (400 MHz, CDCl₃)** δ : 9.92 (t, J = 5.7 Hz, 1H), 8.92 (br. s, 1H), 8.54 (s, 1H), 8.25 (s, 1H), 8.13 (d, J = 8.7 Hz, 2H), 7.39 (d, J = 8.7 Hz, 2H), 5.87 (d, J = 7.2 Hz, 1H), 5.07 (dd, J = 7.2, J = 4.7 Hz, 1H), 4.44 (t, J = 6.7 Hz, 2H), 4.37 (m, 2H), 4.20 (t, J = 5.2 Hz, 2H), 3.97 (d, J = 13.0 Hz, 1H), 3.77 (d, J = 13.0 Hz, 1H), 3.09 (t, J = 6.7 Hz, 2H), 2.94 (br. s, 1H), 0.78 (s, 9H), -0.19 (s, 3H), -0.38 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ : 169.9, 154.0, 150.8, 149.5, 147.0, 145.4, 143.6, 129.9, 123.9, 122.0, 91.2, 87.6, 74.8, 72.8, 64.9, 63.3, 42.2, 34.9, 25.6, 17.9, -5.2; HRMS (ESI): calculated for C₂₇H₃₇N₇O₉Si⁺: m/z = 632.2495 [M + H]⁺; found m/z = 632.2492.

Compound 36



Yield: 98%; **IR**: $\tilde{v} = 3229$ (w), 2953 (w), 2929 (w), 2857 (w), 1735 (s), 1693 (s), 1607 (s), 1589 (s), 1517 (vs), 1469 (s), 1391 (w), 1310 (vs), 1292 (w), 1251 (w), 1210 (w), 835 (s) cm⁻¹; ¹**H NMR (400 MHz, Acetone-***d*₆**)** δ : 10.30 (d, J = 7.9 Hz, 1H), 9.41 (s, 1H), 8.77 (s, 1H), 8.50 (s, 1H), 8.10 (d, J = 8.8 Hz, 2H), 8.06 (d, J = 8.8 Hz, 2H), 7.54 (dd, J = 8.7, 7.4 Hz, 4H), 6.12 (d, J = 5.8 Hz, 1H), 5.07 (s, 1H), 5.00 – 4.95 (m, 1H), 4.90 (dt, J = 7.9, 5.3 Hz, 1H), 4.49 – 4.31 (m, 5H), 4.23 (q, J = 2.4 Hz, 1H), 4.06 (s, 1H), 3.92 (dd, J = 12.4, 2.1 Hz, 1H), 3.79 (d, J = 12.2 Hz, 1H), 3.10 (dt, J = 12.6, 6.4 Hz, 4H), 3.02 (d, J = 5.5 Hz, 2H), 0.79 (s, 9H), -0.08 (s, 3H), -0.21 (s, 160) (s), 1200 (s), 1200

3H); ¹³C NMR (101 MHz, Acetone- d_6) δ : 171.3, 171.2, 154.2, 151.4, 150.9, 150.6, 147.6, 147.4, 147.2, 144.3, 136.6, 131.0, 124.1, 121.9, 90.6, 87.6, 76.8, 72.4, 65.8, 65.1, 62.9, 50.6, 37.1, 35.2, 27.2, 26.0, 18.6, -4.9, -5.1; HRMS (ESI): calculated for C₃₇H₄₇N₈O₁₃Si⁺: m/z = 839.3032 [M+H]⁺; found: m/z = 839.3041 [M+H]⁺.

Compound 37a



Yield: 80%; **IR**: $\tilde{v} = 3240$ (w), 2929 (w), 2857 (w), 1742 (w), 1699 (vs), 1613 (s), 1520 (vs), 1471 (s), 1345 (vs), 1255 (w), 839 (s) cm⁻¹; ¹**H NMR (400 MHz, CD₂Cl₂)** δ : 9.83 (d, J = 7.4 Hz, 1H), 8.63 (s, 1H), 8.40 (s, 1H), 8.18 (s, 1H), 8.10 (d, J = 8.7 Hz, 2H), 7.38 (d, J = 8.7 Hz, 2H), 7.33 – 7.21 (m, 2H), 7.19 – 7.14 (m, 2H), 5.87 (d, J = 7.2 Hz, 1H), 5.64 (dd, J = 11.6 Hz, J = 2.3 Hz, 1H), 5.07 (dd, J = 7.2 Hz, J = 4.7 Hz, 1H), 4.87 – 4.80 (m, 1H), 4.45 – 4.32 (m, 4H), 3.93 (d, J = 12.9 Hz, 1H), 3.79 – 3.69 (m, 1H), 3.17 (d, J = 6.2 Hz, 2H), 3.05 (t, J = 6.6 Hz, 2H), 2.93 (s, 1H), 0.79 (s, 9H), -0.19 (s, 3H), -0.38 (s, 3H); ¹³C NMR (101 MHz, CD₂Cl₂) δ : 171.4, 153.1, 150.7, 150.5, 149.8, 149.3, 146.8, 145.8, 143.3, 136.3, 129.8, 129.4, 128.3, 127.2, 123.5, 91.0, 87.6, 74.8, 72.7, 64.7, 63.1, 54.9, 37.8, 34.7, 25.3, 17.7, -5.6, -5.7; HRMS (ESI): calculated for C₃₄H₄₄N₇O₉Si⁺: m/z = 722.2965 [M+H]⁺; found: m/z = 722.2971 [M+H]⁺.

Compound 37b



Yield: 78%; IR: $\tilde{v} = 3229$ (w), 2930 (w), 2858 (w), 1740 w), 1697 (vs), 1612 (s), 1519 (vs), 1470 (s), 1345 (vs), 1254 (s), 1216 (w), 838 (vs), 781 (s), 736 (vs), 700 (s) cm⁻¹; ¹H NMR (400 MHz, CD₂Cl₂) δ : 9.79 (br. s, 1H), 8.58 (br. s, 1H), 8.39 (s, 1H), 8.17 (t, J = 4.0 Hz, 1H), 8.05 (d, J = 8.7 Hz, 2H), 7.35 (d, J = 8.7 Hz, 2H), 7.32 – 7.23 (m, 2H), 7.21-7.15 (m, 2H), 5.87 (d, J = 7.1 Hz, 1H), 5.61 (dd, J = 11.6 Hz, 2.4 Hz, 1H), 5.05 (dd, J = 7.2 Hz, 4.7 Hz, 1H), 4.82 (q, J = 6.2 Hz, 1H), 4.46 – 4.30 (m, 4H), 3.93 (d, J = 12.9 Hz, 1H), 3.79 – 3.68 (m, 1H), 3.17 (d, J = 6.2 Hz, 2H), 3.03 (t, J = 6.5 Hz, 3H), 2.88 (s, 1H), 0.80 (s, 9H), -0.17 (s, 3H), -0.36 (s, 3H); ¹³C NMR (101 MHz, CD₂Cl₂) δ : 171.4, 153.1, 150.7, 150.5, 149.8, 149.3, 146.8, 145.8, 143.3, 136.3, 129.8, 129.4, 128.5, 128.3, 127.2, 123.5, 91.0, 87.6, 74.8, 72.7, 64.7, 63.1, 54.9, 37.8, 34.7, 25.3, 17.7, -5.6, -5.7; HRMS (ESI): calculated for C₃₄H₄₃N₇O₉SiNa⁺: m/z = 744.2784 [M+Na]⁺; found: m/z = 744.2776 [M+Na]⁺.

Compound 38



Yield: 96%; **IR**: $\tilde{v} = 3240$ (w), 2950 (w), 2927 (w), 2856 (w), 1737 (w), 1695 (s), 1611 (s), 1589 (s), 1520 (vs), 1469 (s), 1346 (vs), 1313 (w), 1250 (vs), 1130 (w), 1093 (s), 835 (vs), 780 (vs) cm⁻¹; ¹**H NMR (400 MHz, Acetone-***d*₆**)** δ : 10.11 (d, J = 8.2 Hz, 1H), 9.13 (s, 1H), 8.72 (s, 1H), 8.52 (s, 1H), 8.11 (d, J = 8.7 Hz, 2H), 7.59 (d, J = 8.7 Hz, 2H), 6.13 (d, J = 6.1 Hz, 1H), 5.07 (dd, J = 8.7, 3.8 Hz, 1H), 4.99 (dd, J = 6.1, 4.7 Hz, 1H), 4.66 (dt, J = 8.3, 3.0 Hz, 1H), 4.45 (td, J

= 6.5, 1.2 Hz, 2H), 4.39 (td, J = 4.3, 2.5 Hz, 1H), 4.22 (p, J = 2.8 Hz, 1H), 4.15 (dd, J = 10.3, 2.9 Hz, 1H), 4.04 (d, J = 3.8 Hz, 1H), 3.98 (dd, J = 10.2, 3.2 Hz, 1H), 3.94 – 3.87 (m, 1H), 3.78 (ddd, J = 12.4, 8.5, 2.5 Hz, 1H), 3.16 (t, J = 6.3 Hz, 2H), 0.88 (s, 9H), 0.79 (s, 9H), 0.07 (s, 3H), 0.02 (s, 3H), -0.08 (s, 3H), -0.23 (s, 3H); ¹³C NMR (101 MHz, Acetone-*d*₆) δ: 171.0, 154.1, 151.4, 151.0, 147.4, 144.1, 131.1, 124.2, 122.1, 90.5, 87.8, 76.8, 72.6, 65.7, 64.3, 63.0, 56.4, 35.3, 27.2, 26.1, 26.0, 18.7, 18.6, -4.9, -5.2, -5.3, -5.6; HRMS (ESI): calculated for $C_{34}H_{54}N_7O_{10}Si_2Na^+$: m/z = 798.3290 [M+Na]⁺; found: m/z = 798.3288 [M+Na]⁺.

General procedure for DMT protection:



The 3'-5'-deprotected adenosine derivative (1 eq.) was dissolved in pyridine under N_2 atmosphere. DMT chloride (1.5 eq.) was added in two portions and the mixture was stirred at room temperature overnight. Then the volatiles were evaporated, and crude product was purified by flash chromatography eluting with CH₂Cl₂/MeOH (10/1/, v/v) with an addition of 0.1% of pyridine, unless otherwise specified, to afford the DMT protected derivative as white foam.

Compound 39



Yield: 90%; **IR**: $\tilde{v} = 3350$ (w), 2930 (w), 2856 (w), 1729 (w), 1684 (s), 1608 (s), 1521 (vs), 1464 (s), 1345 (vs), 1248 (vs), 1174 (w), 1094 (w), 10033 (s), 827 (vs), 777 (vs) cm⁻¹; ¹H NMR (400 MHz, Acetone-*d*₆) δ : 10.00 (d, *J* = 9.0 Hz, 1H), 8.95 (s, 1H), 8.55 (s, 1H), 8.40 (s, 1H), 7.98 (d, *J* = 8.7 Hz, 2H), 7.59 - 7.46 (m, 4H), 7.35 (dd, *J* = 9.0, 3.0 Hz, 4H), 7.28 - 7.13 (m, 4H), 6.83

(dd, J = 9.0, 3.0 Hz, 4H), 6.16 (d, J = 4.6 Hz, 1H), 5.16 (t, J = 4.6 Hz, 1H), 4.61 – 4.48 (m, 3H), 4.45 – 4.35 (m, 2H), 4.33 – 4.26 (m, 1H), 3.99 (d, J = 5.8 Hz, 1H), 3.75 (s, 6H), 3.51 – 3.44 (m, 2H), 3.12 (t, J = 6.2 Hz, 2H), 1.27 (d, J = 6.2 Hz, 3H), 0.91 (s, 9H), 0.85 (s, 9H), 0.09 (s, 3H), 0.06 (s, 3H), -0.05 (s, 6H); ¹³C NMR (101 MHz, Acetone-*d*₆) δ : 171.6, 159.6, 154.8, 151.5, 151.3, 147.5, 146.1, 136.7, 131.1, 130.9, 129.7, 129.0, 128.5, 127.5, 126.1, 124.1, 121.9, 113.8, 90.2, 87.1, 84.8, 76.3, 71.9, 69.6, 65.6, 64.4, 60.3, 55.5, 35.3, 26.1, 25.9, 21.5, 18.7, 18.5, -4.1, -4.6, -4.8, -5.2; HRMS (ESI): calculated for C₅₆H₇₄N₇O₁₂Si₂⁺: m/z = 1092.4929 [M+H]⁺; found: m/z = 1092.4937 [M+H]⁺.

Compound 40



Eluent: 10% CH₂Cl₂ in EtOAc to pure EtOAc containing 0.1% of pyridine.

Yield: 85%; **IR**: $\tilde{v} = 3854$ (w), 3746 (w), 3650 (w), 3630 (w), 2931 (w), 2361 (w), 2341 (w), 1740 (s), 1700 (vs), 1654 (w), 1609 (s), 1587 (w), 1559 (w), 1508 (vs), 1472 (s), 1396 (w), 1345 (s), 1300 (w), 1250 (vs), 1176 (s), 1118 (vs), 1032 (vs), 986 (w), 912 (w), 835 (vs), 781 (s), 751 (w), 700 (w) cm⁻¹; **¹H NMR (400 MHz, Acetone-***d*₆**)** δ : 10.11 – 10.06 (m, 1H), 8.61 – 8.55 (m, 1H), 8.47 (s, 1H), 8.45 (s, 1H), 8.13 – 8.05 (m, 2H), 7.71 (d, J = 1.4 Hz, 1H), 7.63 – 7.55 (m, 2H), 7.53 – 7.46 (m, 2H), 7.40 – 7.32 (m, 4H), 7.28 (td, J = 8.2, 7.7, 2.0 Hz, 2H), 7.25 – 7.18 (m, 1H), 7.06 (s, 1H), 6.88 – 6.81 (m, 4H), 6.13 (d, J = 4.4 Hz, 1H), 5.88 (s, 2H), 5.10 (t, J = 4.6 Hz, 1H), 4.75 (dq, J = 8.0, 4.2, 3.3 Hz, 1H), 4.56 – 4.51 (m, 1H), 4.47 – 4.36 (m, 2H), 4.30 – 4.25 (m, 1H), 4.03 – 3.93 (m, 1H), 3.77 (s, 3H), 3.76 (s, 3H), 3.50 – 3.40 (m, 2H), 3.16 – 3.11 (m, 2H), 3.12 – 3.04 (m, 2H), 1.05 (s, 9H), 0.86 (s, 9H), 0.06 (s, 3H), -0.04 (s, 3H); ¹³C NMR (101 MHz, Acetone-*d*₆) δ : 177.9, 172.1, 159.6, 153.9, 151.7, 151.3, 147.7, 147.6, 146.1, 143.3, 139.2, 138.8, 136.7, 131.1, 131.0, 130.0, 129.0, 128.6, 127.6, 124.6, 124.1, 118.2, 113.9, 90.1, 87.1, 84.7, 76.4, 71.9, 71.8, 68.8, 66.2, 64.4, 55.5, 39.2, 35.3, 31.0, 27.0, 26.1, 18.7, -4.6, -4.8; HRMS (ESI): calculated for C₅₈H₇₀N₉O₁₃Si⁺: m/z = 1128.4857 [M+H]⁺; found: m/z = 1128.4885 [M+H]⁺.



Yield: 70%; **IR**: $\tilde{v} = 2929$ (w), 1735 (w), 1684 (w), 1569 (s), 1508 (vs), 1464 (s), 1344 (s), 1249 (vs), 1176 (w), 1015 (s), 800 (vs) 749 (s) cm⁻¹; ¹H NMR (400 MHz, Acetone-*d*₆) δ : 10.02 (d, *J* = 9.0 Hz, 1H), 8.55 (s, 1H), 8.51 (s, 1H), 8.11 (d, *J* = 8.7 Hz, 2H), 7.59 (d, *J* = 8.7 Hz, 2H), 7.52 – 7.47 (m, 2H), 7.37 (dd, *J* = 9.0, 2.2 Hz, 4H), 7.30 – 7.13 (m, 4H), 6.86 – 6.82 (m, 4H), 6.16 (d, *J* = 4.6 Hz, 1H), 5.16 (t, *J* = 4.6 Hz, 1H), 4.57 – 4.39 (m, 5H), 4.30 (q, *J* = 4.6 Hz, 1H), 4.00 (d, *J* = 5.8 Hz, 1H), 3.76 (s, 6H), 3.47 (d, *J* = 4.2 Hz, 2H), 3.16 (t, *J* = 6.5 Hz, 2H), 2.31 – 2.30 (m, 1H), 0.99 (d, *J* = 6.5 Hz, 3H), 0.95 (d, *J* = 6.5 Hz, 3H), 0.85 (s, 9H), 0.06 (s, 3H), -0.04 (s, 3H); ¹³C NMR (101 MHz, Acetone-*d*₆) δ : 172.2, 159.5, 154.3, 151.6, 151.3, 147.4, 146.1, 143.5, 136.6, 131.0, 130.0, 128.9, 128.9, 127.5, 126.1, 124.2, 121.7, 113.8, 90.3, 87.0, 84.7, 76.4, 71.9, 65.2, 64.3, 59.5, 55.4, 35.3, 31.5, 26.1, 19.6, 18.7, 18.2, -4.6, -4.8; HRMS (ESI): calculated for C₅₁H₆₂N₇O₁₁Si⁺: m/z = 976.4277 [M+H]⁺; found: m/z = 976.4287 [M+H]⁺.

Compound 42



Yield: 85%; **IR**: $\tilde{v} = 2931$ (w), 1748 (w), 1703 (s), 1609 (s), 1588 (w), 1509 (s), 1469 (s), 1345 (vs), 1250 (vs), 1177 (vs), 1035 (w), 835 (s) cm⁻¹; ¹**H NMR (400 MHz, Acetone-***d*₆**)** δ : 9.93 (t, *J* = 5.5 Hz, 1H), 9.05 (br. s, 1H), 8.58 (s, 1H), 8.46 (s, 1H), 8.11 (d, *J* = 8.7 Hz, 2H), 7.58 (d, *J* = 8.7 Hz, 2H), 7.54 – 7.47 (m, 2H), 7.41 – 7.33 (m, 4H), 7.31 – 7.23 (m, 2H), 7.23 – 7.16 (m, 1H), 6.88 – 6.78 (m, 4H), 6.16 (d, *J* = 4.4 Hz, 1H), 5.10 (dd, *J* = 4.7 Hz, 1H), 4.53 (t, *J* = 5.1 Hz, 1H), 4.44 (t, *J* = 6.4 Hz, 2H), 4.30 (dt, *J* = 4.4 Hz, 1H), 4.13 (d, *J* = 5.5 Hz, 2H), 4.00 (d, *J* = 5.7 Hz, 2H), 4.30 (dt, *J* = 4.4 Hz, 1H), 4.13 (d, *J* = 5.5 Hz, 2H), 4.00 (d, *J* = 5.7 Hz, 2H), 4.50 (dt, *J* = 5.7 Hz), 4.50 (dt, *J* = 5.7 Hz), 4.50 (dt, *J* = 5.7 Hz), 5.5 Hz (dt), 5.5 Hz

1H), 3.75 (s, 6H), 3.53 - 3.42 (m, 2H), 3.13 (t, J = 6.4 Hz, 2H), 0.85 (s, 9H), 0.05 (s, 3H), -0.05 (s, 3H); ¹³C NMR (101 MHz, Acetone- d_6) δ : 170.6, 159.6, 154.6, 151.7, 151.3, 151.2, 147.6, 147.4, 146.1, 143.6, 136.7, 131.1, 130.9, 129.1, 129.0, 128.6, 127.5, 124.2, 121.7, 113.8, 90.0, 87.1, 84.8, 76.4, 72.0, 65.2, 64.4, 55.5, 42.6, 35.3, 26.1, 18.7, -4.7, -4.8; HRMS (ESI): calculated for C₄₈H₅₅N₇O₁₁Si⁺: m/z = 934.3802 [M + H]⁺; found: m/z = 934.3812 [M + H]⁺.

Compound 43



Yield: 95%; **IR**: $\tilde{v} = 2930$ (w), 1734 (s), 1698 (s), 1607 (s), 1509 (vs), 1466 (s), 1370 (vs), 1248 (s), 1174 (s), 1032 (s), 829 (vs), 699 (s) cm⁻¹; ¹H NMR (400 MHz, Acetone-*d*₆) δ : 10.29 (d, *J* = 7.9 Hz, 1H), 9.18 (s, 1H), 8.60 (s, 1H), 8.42 (s, 1H), 8.10 – 8.04 (m, 4H), 7.58 – 7.51 (m, 4H), 7.39 – 7.36 (m, 4H), 7.30 – 7.25 (m, 2H), 7.21 – 7.18 (m, 2H), 6.86 – 6.82 (m, 5H), 6.17 (d, *J* = 4.2 Hz, 1H), 5.10 (t, *J* = 4.5 Hz, 1H), 4.85 (dt, *J* = 7.9, 5.2 Hz, 1H), 4.56 (q, *J* = 5.2 Hz, 1H), 4.46 – 4.28 (m, 5H), 4.02 (d, *J* = 5.9 Hz, 1H), 3.75 (d, *J* = 2.1 Hz, 6H), 3.53 – 3.44 (m, 2H), 3.13 – 3.06 (m, 4H), 3.00 – 2.96 (m, 2H), 0.86 (s, 9H), 0.07 (s, 3H), 0.03 (s, 3H); ¹³C NMR (101 MHz, Acetone-*d*₆) δ : 171.3, 171.2, 159.5, 154.2, 151.5, 150.6, 147.4, 147.2, 146.1, 143.6, 136.7, 131.0, 130.0, 129.0, 128.6, 127.5, 124.2, 113.8, 90.1, 87.0, 84.6, 76.4, 71.8, 65.7, 65.1, 64.3, 55.4, 50.6, 37.1, 35.2, 26.1, 18.6, -4.6, -4.8; HRMS (ESI): calculated for C₅₈H₆₄N₈O₁₅SiNa⁺: m/z = 1163.4158 [M+Na]⁺; found: m/z = 1163.4159 [M+Na]⁺.

Compound 44a



Eluent: CH₂Cl₂/MeOH (20/1, v/v).

Yield: 82%; **IR**: $\tilde{v} = 3245$ (w), 2952 (w), 2931 (w), 1741 (w), 1699 (s), 1608 (s), 1587 (w), 1519 (s), 1509 (s), 1469 (w), 1345 (s), 1249 (s), 1176 (s), 1034 (w), 834 (s), 783 (w), 735 (vs), 701 (vs) cm⁻¹; ¹**H NMR (400 MHz, CD₂Cl₂)** δ : 9.83 (d, J = 7.5 Hz, 1H), 8.29 (s, 1H), 8.17 (s, 1H), 8.06 (d, J = 8.7 Hz, 2H), 8.00 (s, 1H), 7.46 (d, J = 6.8 Hz, 2H), 7.38 – 7.32 (m, 7H), 7.31 – 7.20 (m, 7H), 7.18 – 7.14 (m, 2H), 6.82 (d, J = 8.9 Hz, 4H), 6.02 (d, J = 5.0 Hz, 1H), 5.02 (t, J = 5.0 Hz, 1H), 4.80 (q, J = 6.3 Hz, 1H), 4.44 – 4.32 (m, 3H), 4.24 (q, J = 3.8 Hz, 1H), 3.77 (s, 6H), 3.48 (dd, J = 10.7 Hz, J = 3.2 Hz, 1H), 3.39 (dd, J = 10.7 Hz, J = 4.3 Hz, 1H), 3.16 (d, J = 6.3 Hz, 2H), 3.03 (t, J = 6.5 Hz, 2H), 2.67 (d, J = 4.7 Hz, 1H), 0.85 (s, 9H), 0.01 (s, 3H), -0.12 (s, 3H); ¹³C NMR (101 MHz, CD₂Cl₂) δ : 171.5, 153.3, 150.8, 150.6, 149.8, 149.4, 146.8, 145.8, 143.6, 136.2, 135.9, 129.9, 129.4, 128.5, 128.3, 127.2, 123.7, 123.6, 121.8, 91.0, 87.5, 74.8, 72.7, 64.8, 63.1, 54.8, 37.8, 34.7, 25.3, 17.7, -5.7; HRMS (ESI): calculated for C₅₅H₆₂N₇O₁₁Si⁺: m/z = 1024.4271 [M+H]⁺; found: m/z = 1024.429 [M+H]⁺.

Compound 44b



Eluent: $CH_2Cl_2/MeOH$ (20/1, v/v).

Yield: 85%; **IR**: $\tilde{v} = 3231$ (w), 2930 (w), 2858 (w), 1741 (w), 1697 (s), 1612 (s), 1587 (w), 1518 (vs), 1470 (s), 1344 (vs), 1253 (w), 1180 (w), 1134 (w), 1085 (w), 836 (s), 780 (s), 734 (vs), 698 (vs) cm⁻¹; ¹**H NMR (400 MHz, CD₂Cl₂)** δ : 9.83 (d, J = 7.5 Hz, 1H), 8.58 (br. s, 1H), 8.32 (s, 1H), 8.19 (s, 1H), 8.06 (d, J = 8.7 Hz, 2H), 7.99 (s, 1H), 7.46 (d, J = 6.9 Hz, 2H), 7.38 – 7.31 (m, 7H), 7.31 – 7.21 (m, 7H), 7.20 – 7.16 (m, 2H), 6.82 (d, J = 8.9 Hz, 4H), 6.05 (d, J = 4.7 Hz, 1H), 4.94 (t, J = 5.0 Hz, 1H), 4.81 (q, J = 6.3 Hz, 1H), 4.44 – 4.33 (m, 3H), 4.24 (q, J = 3.8 Hz, 1H), 3.77 (s, 6H), 3.48 (dd, J = 10.7 Hz, 3.1 Hz, 1H), 3.41 (dd, J = 10.7 Hz, 4.2 Hz, 1H), 3.16 (d, J = 6.9 Hz, 2H), 3.03 (t, J = 6.5 Hz, 2H), 2.66 (d, J = 5.0 Hz, 1H), 0.86 (s, 9H), 0.02 (s, 3H), -0.09 (s, 3H); ¹³C **NMR (101 MHz, CD₂Cl₂)** δ : 171.4, 153.4, 150.7, 150.5, 149.8, 149.4, 146.8, 145.9, 143.7, 136.3, 129.9, 129.5, 128.5, 128.3, 127.2, 123.5, 123.5, 121.8, 91.0, 87.5, 74.9, 72.7, 64.8, 63.1, 54.9, 37.8, 34.7, 25.3, 17.8, -5.6; **HRMS (ESI)**: calculated for C₅₅H₆₂N₇O₁₁Si⁺: m/z = 1024.4271 [M+H]⁺; found: m/z = 1024.4291 [M+H]⁺.

Compound 45



Yield: 90%; IR: $\tilde{v} = 2930$ (w), 1740 (s), 1694 (s), 1607 (w), 1582 (w), 1519 (vs), 1465 (w), 1345 (s), 1249 (s), 1176 (w), 1110 (w), 1034 (w), 820 (vs), 778 (s), 699 (w) cm⁻¹; ¹H NMR (400 MHz, Acetone-*d*₆) δ : 10.13 (d, J = 8.2 Hz, 1H), 8.99 (s, 1H), 8.58 (s, 1H), 8.44 (s, 1H), 8.09 (d, J = 8.7 Hz, 2H), 7.58 (d, J = 8.7 Hz, 2H), 7.51 (dd, J = 8.4, 1.3 Hz, 2H), 7.39 (d, J = 2.5 Hz, 2H), 7.37 (d, J = 2.4 Hz, 2H), 7.32 – 7.18 (m, 4H), 6.87 – 6.83 (m, 4H), 6.18 (d, J = 4.7 Hz, 1H), 5.13 (t, J = 5.0 Hz, 1H), 4.62 (dt, J = 8.2, 2.8 Hz, 1H), 4.53 (q, J = 5.0 Hz, 1H), 4.45 (t, J = 6.3 Hz, 2H), 4.30 (td, J = 4.6, 3.4 Hz, 1H), 4.14 (dd, J = 10.2, 2.8 Hz, 1H), 3.96 (dd, J = 10.2, 2.8 Hz, 1H), 3.77 (s, 6H), 3.48 (qd, J = 10.5, 4.1 Hz, 2H), 3.16 (t, J = 6.3 Hz, 2H), 0.89 (s, 9H), 0.85 (s, 9H), 0.07 (s, 3H), 0.06 (s, 3H), 0.03 (s, 3H), -0.06 (s, 3H); ¹³C NMR (101 MHz, Acetone-*d*₆) δ : 170.0, 159.5, 154.1, 151.6, 150.6, 147.4, 146.1, 143.6, 136.7, 131.1, 130.9, 129.0, 128.6, 127.6, 124.2, 121.8, 113.8, 89.9, 87.1, 84.9, 76.5, 72.0, 65.8, 64.3, 56.4, 55.4, 35.3, 27.2, 26.1, 18.7, 18.4, -4.6, -4.9, -5.3, -5.5; HRMS (ESI): calculated for C₅₅H₇₁N₇O₁₂Si₂Na⁺: m/z = 1100.4597 [M+Na]⁺; found: m/z = 1100.4620 [M+Na]⁺.

General synthesis of RNA phosphoramidites



In an oven-dried flask under argon atmosphere, 5'-DMT protected compound (1 eq.) was dissolved in CH₂Cl₂ and cooled to 0 °C. Hünig's base (4 eq.) was added dropwise followed by the addition of 2-cyanoethyl *N*,*N*-diisopropylchlorophosphoramidite (2.5 eq.). The solution was stirred at room temperature for 2 h. The reaction was quenched by addition of sat. NaHCO₃ solution, and then extracted three times with CH₂Cl₂. The organic phase was dried over Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by flash chromatography, eluting with Hex/EtOAc (1/1, v/v) containing 0.1% of pyridine, unless otherwise specified. The phosphoramidite was obtained as a mixture of diastereomers as white foam.

Compound 46



Yield: 85%; ³¹P NMR (162 MHz, Acetone- d_6) δ : 150.16, 148.45; HRMS (ESI): calculated for C₆₅H₉₁N₉O₁₃PSi₂⁺: m/z = 1292.6007 [M+H]⁺; found: m/z = 1292.6033 [M+H]⁺.



Eluent: 30% CH₂Cl₂ in EtOAc containing 0.1% of pyridine.

Yield: 66%; ³¹P NMR (162 MHz, Acetone-*d*₆) δ : 150.26, 148.61; HRMS (ESI): calculated for C₆₇H₈₇N₁₁O₁₄PSi⁺: m/z = 1328.5935 [M+H]⁺; found: m/z = 1328.5944 [M+H]⁺.

Compound 48



Yield: 87%; ³¹P NMR (162 MHz, Acetone-*d*₆) δ : 150.22, 148.65; HRMS (ESI): calculated for C₆₀H₇₉N₉O₁₂PSi⁺: m/z = 1176.5355 [M+H]⁺; found: m/z = 1176.5359 [M+H]⁺.



Yield: 86%; ³¹**P NMR (162 MHz, Acetone-***d*₆**)** δ : 150.32, 148.60; **HRMS (ESI)**: calculated for C₅₇H₇₂N₉O₁₂SiPSi⁺: m/z = 1134.4880 [M+H]⁺; found: m/z = 1134.4894 [M + H]⁺.

Compound 50



Yield: 65%; ³¹**P NMR (162 MHz, Acetone-***d*₆**)** δ : 150.15, 148.67; **HRMS (ESI)**: calculated for C₆₇H₈₂N₁₀O₁₆PSi⁺: m/z = 1341.5417 [M+H]⁺; found: m/z = 1341.5437 [M+H]⁺.

Compound 51a



Yield: 67%; ³¹P NMR (162 MHz, CD₂Cl₂) δ : 150.7, 149.1; HRMS (ESI): calculated for C₆₄H₇₉N₉O₁₂PSi⁺: m/z = 1224.5350 [M+H]⁺; found: m/z = 1224.5374 [M+H]⁺.

Compound 51b



Yield: 67%; ³¹P NMR (162 MHz, CD₂Cl₂) δ : 150.6, 148.9; HRMS (ESI): calculated for C₆₄H₇₉N₉O₁₂PSi⁺: m/z = 1224.5350 [M+H]⁺; found: m/z = 1224.5383 [M+H]⁺.



Yield: 70%; ³¹P NMR (162 MHz, Acetone- d_6) δ : 150.34, 148.45; HRMS (ESI): calculated for C₆₄H₈₉N₉O₁₃PSi₂⁺: m/z = 1278.5856 [M+H]⁺; found: m/z = 1278.5877 [M+H]⁺.

2.2. Synthesis of DNA building block

Compound 54



The compound was synthesized according to the published procedure.⁵

Acetic anhydride (1.1 ml, 11.5 mmol, 6.2 eq.) was added to a mixture of deoxyadenosine monohydrate **53** (0.5 g, 1.85 mmol, 1 eq.), pyridine (7 ml) and 4-(dimethylamino)pyridine (25 mg, 0.4 mmol, 0.1 eq.). The reaction mixture was stirred at room temperature for 4 hours. Subsequently, iced water was added, and the mixture was concentrated and co-evaporated with toluene. The compound was used for further steps without additional purification.

Yield: 99%; ¹**H NMR (400 MHz, CDCl₃)** δ : 8.23 (s, 1H), 7.92 (s, 1H), 7.26 (s, 2H), 6.34 (dd, *J* = 7.9, 6.0 Hz, 1H), 5.34 – 5.32 (m, 1H), 4.32 – 4.24 (m, 3H), 2.87 – 2.80 (m, 1H), 2.57 – 2.52 (m, 1H), 2.04 (s, 3H), 2.00 (s, 3H). **HRMS (ESI)**: calculated for C₁₄H₁₈N₅O₅⁺: m/z = 336.1302 [M+H]⁺; found: m/z = 336.1305 [M+H]⁺.

The analytical data is in agreement with the literature.⁵



The acetyl-protected deoxyadenosine derivative **54** (0.5 g, 1.5 mmol, 1 eq.) was dissolved in dry CH₂Cl₂ under nitrogen atmosphere. 1-*N*-methyl-3-phenoxycarbonyl-imidazolium chloride (**22**; 0.71 g, 3 mmol, 2 eq.) was added to the reaction mixture and the resulting suspension was stirred at room temperature for 2 hours (the solution in time becomes clear). Afterwards the protected threonine derivative **7** (1.1 g, 3 mmol, 2 eq.) was added together with TEA (415 μ l, 3 mmol, 2 eq.) as a solution in CH₂Cl₂ and the resulting solution was stirred overnight at room temperature. The reaction was quenched by addition of saturated aqueous NaHCO₃ solution. The solution was extracted three times with CH₂Cl₂, and the organic phase was dried, filtered and concentrated *in vacuo*. The acetyl groups were immediately deprotected with 7N NH₃/MeOH (2 ml). After stirring 2 hours at room temperature, the reaction mixture was evaporated. The residue was purified via flash chromatography eluting with CH₂Cl₂/MeOH (10/1, v/v).

Yield: 65%; IR: $\tilde{v} = 3227$ (w), 2929 (w), 2855 (w), 1734 (w), 1686 (vs), 1610 (s), 1587 (s), 1518 (vs), 1465 (vs), 1344 (vs), 1312 (w), 1248 (s), 1213 (w), 1094 (vs), 939 (s), 827 (vs) cm⁻¹; ¹H NMR (400 MHz, Acetone-*d*₆) δ : 10.02 (d, *J* = 8.6 Hz, 1H), 9.40 (s, 1H), 8.70 (s, 1H), 8.39 (s, 1H), 7.91 (d, *J* = 8.2 Hz, 2H), 7.47 (d, *J* = 8.2 Hz, 2H), 6.54 (t, *J* = 6.7 Hz, 1H), 4.67 (s, 1H), 4.51 (d, *J* = 9.3 Hz, 2H), 4.38 (t, *J* = 5.6 Hz, 2H), 4.09 (s, 1H), 3.86 – 3.67 (m, 2H), 3.08 (t, *J* = 5.6 Hz, 2H), 2.90 (dt, *J* = 13.0, 6.7 Hz, 1H), 2.52 – 2.41 (m, 1H), 2.01 (s, 1H), 1.93 (s, 1H), 1.23 (d, *J* = 5.9 Hz, 3H), 0.88 (s, 9H), 0.05 (s, 3H), -0.06 (s, 3H); ¹³C NMR (101 MHz, Acetone-*d*₆) δ : 171.5, 155.1, 151.1, 150.9, 147.4, 144.2, 130.9, 124.0, 122.0, 89.8, 86.8, 72.6, 69.5, 65.5, 63.3, 41.4, 35.2, 30.6, 26.0, 21.5, 20.5, 18.4, -4.2, -5.2; HRMS (ESI): calculated for C₂₉H₄₂N₇O₉Si⁺: m/z = 660.2813 [M+H]⁺; found: m/z = 660.2812 [M+H]⁺.
Compound 56



The 3'-5'-deprotected adenosine derivative **55** (0.34 g, 0.52 mmol, 1 eq.) was dissolved in pyridine under N₂ atmosphere. DMT chloride (0.26 g, 0.77 mmol, 1.5 eq.) was added in two portions and the mixture was stirred at room temperature overnight. Then the volatiles were evaporated, and crude product was purified by flash chromatography eluting with $CH_2Cl_2/MeOH$ (10/1/, v/v) with an addition of 0.1% of pyridine to afford the DMT protected derivative.

Yield: 72%; **IR**: $\tilde{v} = 2960$ (w), 2930 (w), 1734 (w), 1696 (s), 1609 (s), 1586 (w), 1520 (vs), 1509 (vs), 1467 (s), 1345 (vs), 1304 (w), 1250 (vs), 1175 (s), 1095 (w), 1034 (s), 940 (w), 828 (vs), 777 (s), 699 (w) cm⁻¹; ¹**H NMR (400 MHz, Acetone-***d***₆) δ**: 10.02 (d, J = 9.1 Hz, 1H), 8.98 (s, 1H), 8.52 (s, 1H), 8.34 (s, 1H), 7.96 (d, J = 8.7 Hz, 2H), 7.51 (d, J = 8.7 Hz, 2H), 7.42 (d, J = 7.1 Hz, 2H), 7.31 – 7.28 (m, 4H), 7.24 – 7.12 (m, 3H), 6.79 – 6.74 (m, 4H), 6.55 (t, J = 6.4 Hz, 1H), 4.76 (q, J = 4.2 Hz, 1H), 4.64 – 4.58 (m, 1H), 4.55 (ddd, J = 10.6, 7.7, 1.5 Hz, 2H), 4.41 (t, J = 6.2 Hz, 2H), 4.25 – 4.18 (m, 1H), 3.74 (d, J = 3.3 Hz, 6H), 3.43 (dd, J = 10.2, 5.9 Hz, 1H), 3.35 (dd, J = 10.2, 4.0 Hz, 1H), 3.17 – 3.10 (m, 3H), 2.58 – 2.52 (m, 1H), 1.29 (d, J = 6.3 Hz, 3H), 0.93 (s, 9H), 0.11 (s, 3H), -0.01 (s, 3H); ¹³C NMR (101 MHz, Acetone-*d*₆) δ: 171.6, 159.4, 154.9, 151.2, 151.0, 147.4, 146.1, 143.9, 136.8, 131.0, 130.9, 130.8, 128.9, 128.4, 127.4, 124.0, 122.0, 113.7, 87.7, 86.8, 86.1, 72.6, 69.6, 65.4, 65.1, 60.3, 55.4, 40.0, 35.3, 26.0, 21.5, 18.4, -4.1, -5.2; HRMS (ESI): calculated for C₅₀H₆₀N₇O₁₁Si⁺: m/z = 962.4120 [M+H]⁺; found: m/z = 962.4136 [M+H]⁺.

Compound 57



In an oven-dried flask under argon atmosphere, 5'-DMT protected compound 56 (0.1 g, 0.1 mmol, 1 eq.) was dissolved in CH₂Cl₂ and cooled to 0 °C. Hünig's base (72 µl, 0.4 mmol, 4 eq.) was added dropwise followed by the addition of 2-cyanoethyl N,Ndiisopropylchlorophosphoramidite (60 µl, 0.25 mmol, 2.5 eq.). The solution was stirred at room temperature for 2 h. The reaction was quenched by addition of sat. NaHCO₃ solution, and then extracted three times with CH2Cl2. The organic phase was dried over Na2SO4, filtered and concentrated in vacuo. The residue was purified by flash chromatography, eluting with Hex/EtOAc (1/2, v/v) containing 0.1% of pyridine. The phosphoramidite was obtained as a mixture of diastereomers as white foam.

Yield: 62%; ³¹**P NMR (162 MHz, Acetone-***d*₆**)** δ : 148.00, 146.59; **HRMS (ESI)**: calculated for C₅₉H₇₆N₉NaO₁₂PSi⁺: m/z = 1184.5018 [M+Na]⁺; found: m/z = 1184.5021 [M+Na]⁺.

3. Synthesis and Purification of Oligonucleotides

All of the oligonucleotides used in this study were synthesized on a 1 µmol scale using a DNA automated synthesizer (Applied Biosystems 394 DNA/RNA Synthesizer) with standard phosphoramidite chemistry. The phosphoramidites of canonical ribonucleosides were purchased from Glen Research and Sigma-Aldrich. Oligonucleotides containing non-canonical nucleosides were synthesized in DMT-OFF mode using phosphoramidites (Bz-A, Dmf-G, Ac-C, U) with BTT in CH3CN as an activator, DCA in CH2Cl2 as a deblocking solution and Ac2O in pyridine/THF as a capping reagent. For deprotection of npe-group the solid support was treated with DBU solution (10% in THF, 1 ml) for 2 h at room temperature. Subsequently, the supernatant was removed, and the beads were washed with THF and dried under vacuum. The solid support was suspended in a mixture of aqueous ammonia and methylamine (1:1, 1 ml) at room temperature for 1 h. The supernatant was removed, and the beads were washed with water. The supernatant and washings were combined, and the solvents were evaporated under reduced pressure. The residue was subsequently heated with a solution of triethylamine trihydrofluoride (125 µl) in DMSO (50 µl) at 65 °C for 1.5 h. The RNA was precipitated by addition of aqueous NaOAc solution (3M, 25 µl) and *n*-butanol (1 ml). To ensure complete precipitation, the sample was incubated at -80 °C for 1 h. After centrifugation (4 °C, 4000 rpm, 15 min), the supernatant was removed, and the precipitated RNA was lyophilized. The oligonucleotides were further purified by reverse-phase HPLC using a Waters Breeze (2487 Dual λ Array Detector, 1525 Binary HPLC Pump) equipped with the column VP 250/32 C18 from Macherey Nagel. Oligonucleotides were purified using the following buffer system: buffer A: 100 mM NEt₃/HOAc, pH 7.0 in H₂O and buffer B: 100 mM NEt₃/HOAc in 80 % (v/v) acetonitrile. A flow rate of 5 ml/min with a gradient of 0-25 % of buffer B in 30 min was applied for the purifications. Analytical RP-HPLC was performed on an analytical HPLC Waters Alliance (2695 Separation Module, 2996 Photodiode Array Detector) equipped with the column Nucleosil 120-2 C18 from Macherey Nagel using a flow of 0.5 mL/min, a gradient of 0-30% of buffer B in 45 min was applied. Calculation of concentrations was assisted using the software OligoAnalyzer 3.0 (Integrated DNA Technologies: https://eu.idtdna.com/calc/analyzer). For strands containing non-canonical base, the extinction coefficient of their corresponding canonical-only strand was employed without corrections. The structural integrity of the synthesized oligonucleotides was analyzed by MALDI-TOF mass measurements.

Sequences of synthesized strands:

ON1: 5' GUCt⁶ACCUGA 3'

ON2: 5' GUCg⁶ACCUGA 3'

ON3: 5' GUCval⁶ACCUGA 3'

ON4: 5' GUChis⁶ACCUGA 3'

ON5: 5' GUCasp⁶ACCUGA 3'

ON6: 5' GUC-L-phe⁶ACCUGA 3'

ON7: 5' GUC-D-phe⁶ACCUGA 3'

ON8: 5' AUCGt⁶ACUACGt⁶AAUCGCt⁶AACCG 3'

ON9: 5' AGAUGUG-ser⁶A-asp⁶A-his⁶A-GAGAUGA 3'

ODN1: 5' d(GTCt⁶dACCTGA) 3'



Figure S1. HPLC and MALDI data of oligonucleotides ON1-ON5.



Figure S2. HPLC and MALDI data of oligonucleotides ON6-ODN1.

4. UV Melting Curve Measurements

The UV melting curves were measured on JASCO V-650 spectrometer using 10 mm QS cuvettes, purchased from Hellma Analytics. A solution (80 μ L) of equimolar amounts of oligonucleotides (4 μ M each) in the buffer solution containing 10 mM sodium phosphate buffer (pH 7.0) and 150 mM NaCl was heated at 90 °C for 2 min and gradually cooled to 4 °C prior to the measurement. Melting profiles were recorded at temperatures between 5 and 75 °C with a ramping and scanning rate of 1 °C/min at 260 nm. All samples were measured at least three times. T_m values from each measurement were calculated using the "fitting curve" method and presented as an average of three independent measurements.

5. NMR spectra


























































































6. References

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