

***Schistosoma mansoni* eggs induce Wnt/ β -catenin signaling and activate the protooncogene c-Jun in human and hamster colon**

Short Title: *S. mansoni* activates CRC-signaling

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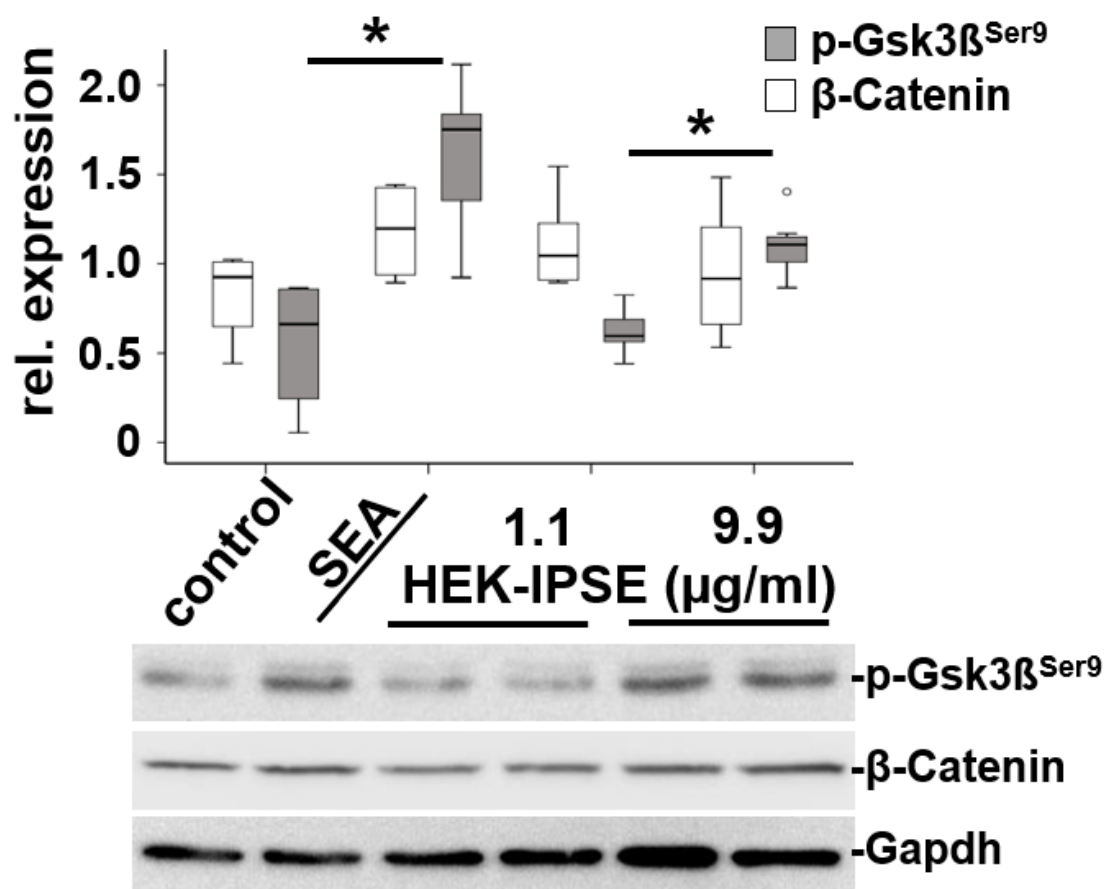
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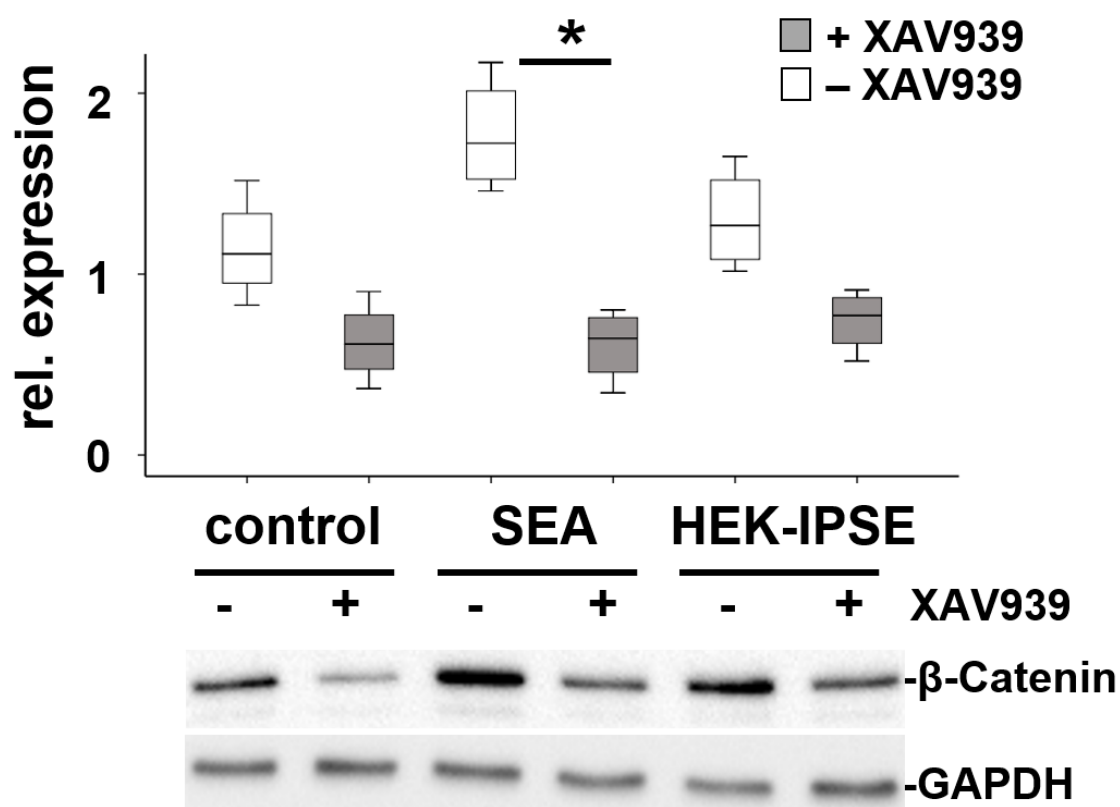
Suppl. Fig. 1



Suppl. Fig. 1: Quantification of western blots shown in Fig. 1c.

Western blot analysis showed the phosphorylation of GSK-3 β and indicated a slightly enhanced β -catenin expression in SW620 cells stimulated with SEA and HEK-IPSE for 4h. The level of significance for the difference between the signals for β -catenin in control and HEK-IPSE was lost after Bonferroni correction ($p=0.014$ before- and $p=0.083$ after Bonferroni correction). $n=3-8$ /group. GAPDH was used as loading control. The experiment and assay was reproduced at least two times and analyzed by optical densitometry, nonparametric ANOVA (Kruskall-Wallis) and subsequent comparison of the groups as well as Bonferroni correction for statistical analysis. Representative blots are shown.

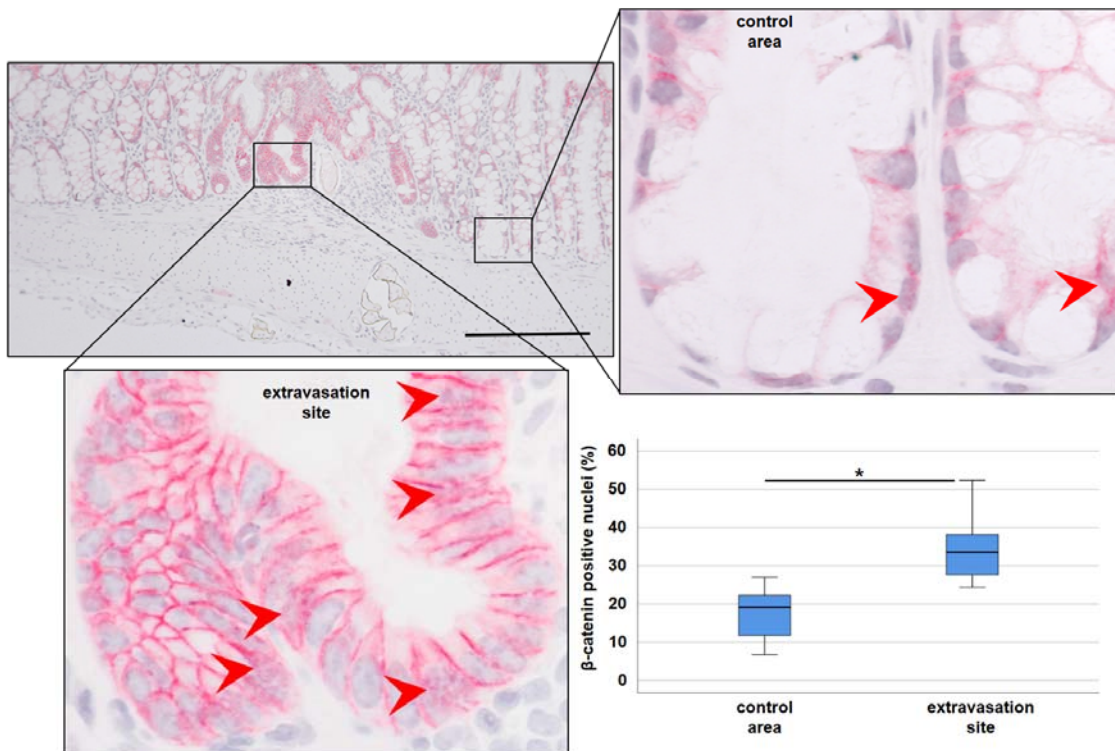
Suppl. Fig. 2



Suppl. Fig. 2: Quantification of western blots shown in Fig. 1d.

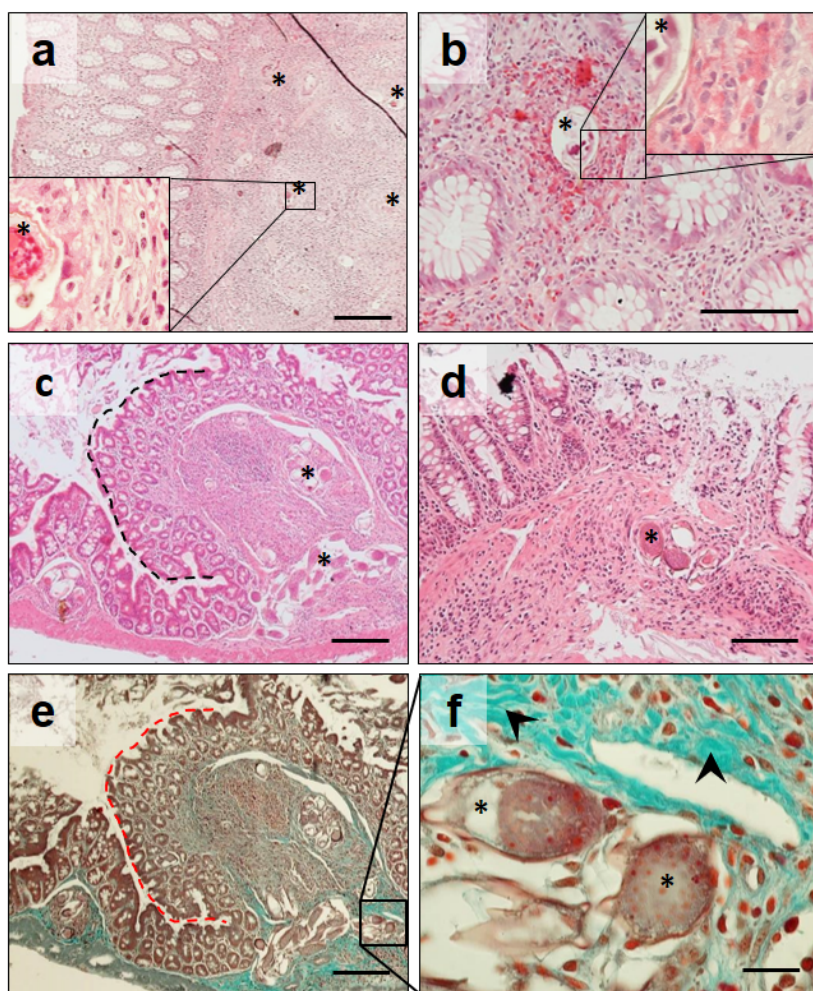
The inhibition of Wnt-signaling by the tankyrase inhibitor XAV939 led to a clear reduction of β -catenin after SEA stimulation. $n=4$ /group. GAPDH was used as loading control. The experiment and assay was reproduced at least two times and analyzed by optical densitometry, nonparametric ANOVA (Kruskall-Wallis) and subsequent comparison of the groups as well as Bonferroni correction for statistical analysis. Representative blots are shown.

Suppl. Fig. 3



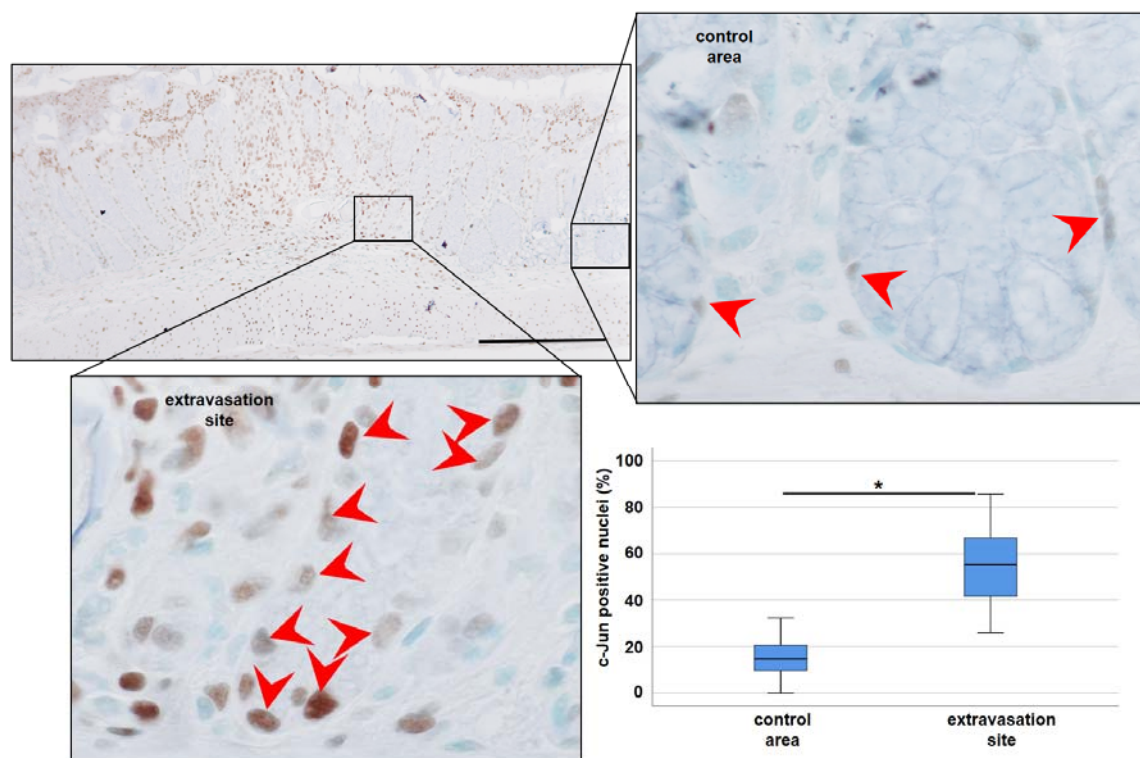
Suppl. Fig. 3: β -catenin positive nuclei of enterocytes in the lower part of crypts in direct vicinity to extravasation sites of *S. mansoni* eggs in bisex infected hamster colon were counted. For control, β -catenin positive nuclei of enterocytes in the lower third of crypts with normal appearance were counted. At least 10 positive and negative area in at least three specimen were used for quantification at 1000x.

Suppl. Fig. 4

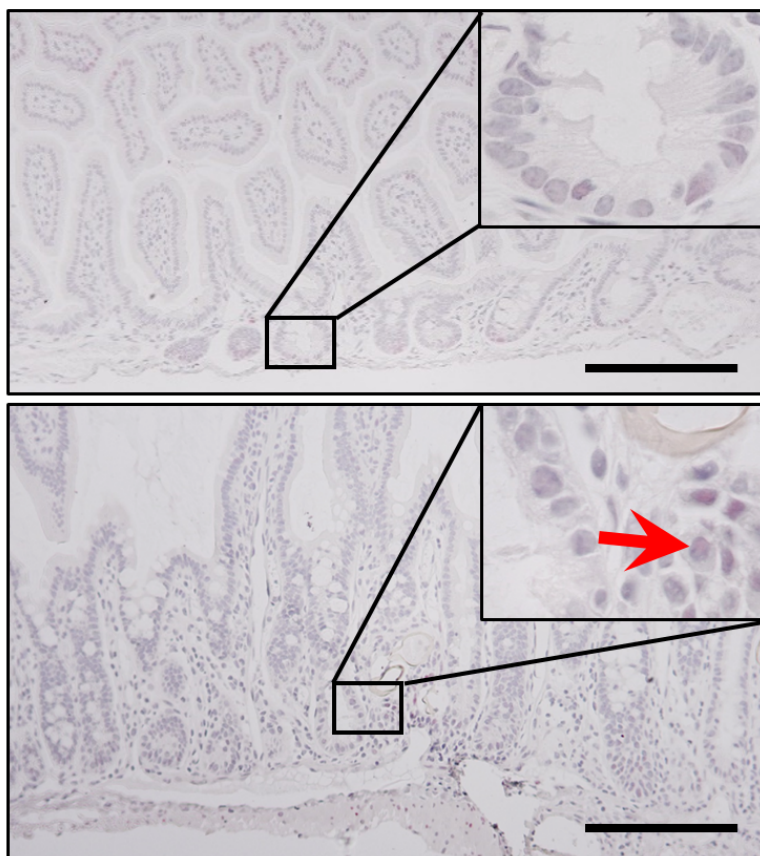


Suppl. Fig. 4: *S. mansoni* eggs provoke fibrogranulomatous colitis in patients and infected hamsters. (A and B) Histologic sections of a rectal biopsy from a 29 year-old male Egypt patient with *S. mansoni*-induced colitis. *S. mansoni* eggs (*) with lateral spines cross the submucosa and mucosa causing a fibroinflammatory reaction with massive infiltration of leucocytes. H.E. staining, magnification 50 x (A) and 200 x (B), bars 200 μ m (A) and 100 μ m (B). **(C and D)** H.E.-stained histologic sections of *S. mansoni*-infected hamster colon. Pronounced multifocal granulomatous inflammation in the lamina propria with multiple parasitic eggs (*) and distinct edema. Magnification 50 x (C) and 100 x (D), bars 200 μ m (A) and 50 μ m (B). **(E and F)** Masson's trichrome-stained histologic sections of *S. mansoni*-infected hamster colon. *S. mansoni* eggs (*) passing the bowel wall were surrounded by **fibrovascular fibrogranulomatous** tissue (arrowheads) leading to **granulomatous** thickening of the submucosa. (F) shows magnification of the area in the box in (E). Magnification 50 x (E) and 1000 x (F), bars 200 μ m (E) and 10 μ m (F).

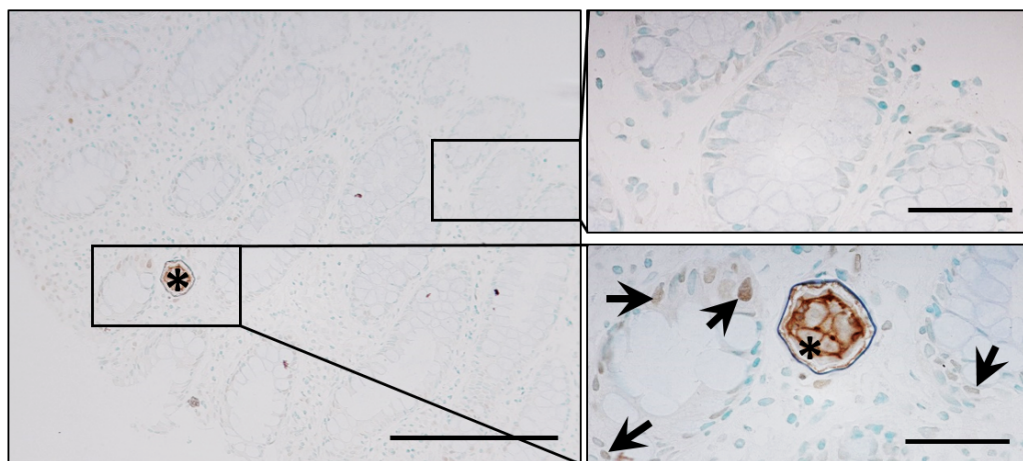
Suppl. Fig. 5



Suppl. Fig. 5: c-Jun positive nuclei of enterocytes in the lower third of crypts in direct vicinity to extravasation sites of *S. mansoni* eggs in bisex infected hamster colon were counted. For control, c-Jun positive nuclei in the lower third of crypts with normal appearance were counted. Red arrowheads depict some of the c-Jun positive nuclei. At least 10 positive and negative area in at least 3 representative specimen were used for quantification at 1000x.

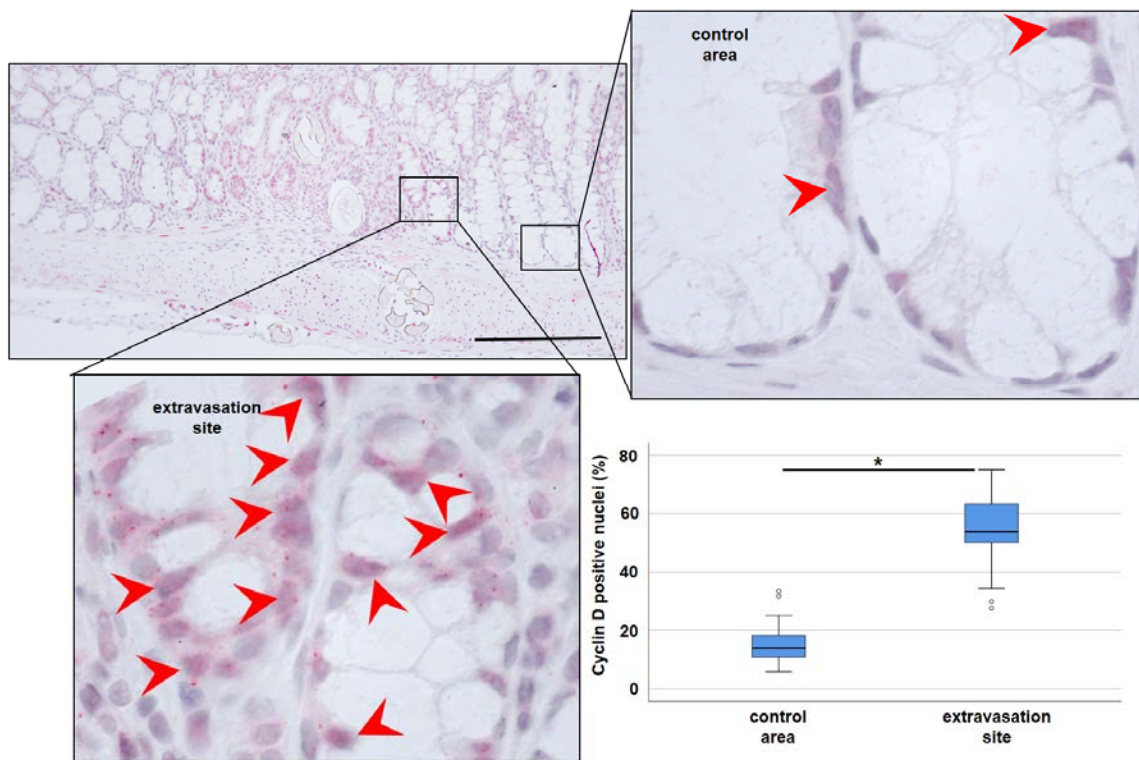
Suppl. Fig. 6

Suppl. Fig. 6: Immunostaining revealed marginal epithelial activation of c-Jun (arrows) around parasite eggs (*) in the ileum of *S. mansoni* infected hamsters (lower panels), which did not differ substantially from c-Jun activation in the ileum of control hamsters (upper panels). Magnification 200 x, bar 200 µm.

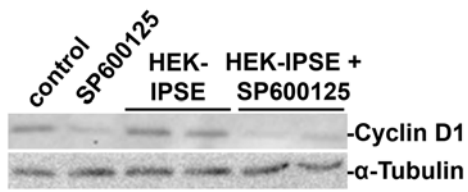
Suppl. Fig. 7

Suppl. Fig. 7: Immunostaining of Cyclin D1 (brown) depicted nuclear localization of Cyclin D1 (arrows) in enterocytes of crypts in direct vicinity to *S. mansoni* eggs (*). The upper panel on the right depicts the magnification of the boxed control area showing normal appearing crypts with marginal Cyclin D1 staining. A representative histologic section of a colon biopsy from an 25-year-old Eritrean male with schistosomal colitis is shown. Nuclear co-staining in blue, magnification 200 x and 1000 x, bars 200 µm and 20 µm.

Suppl. Fig. 8

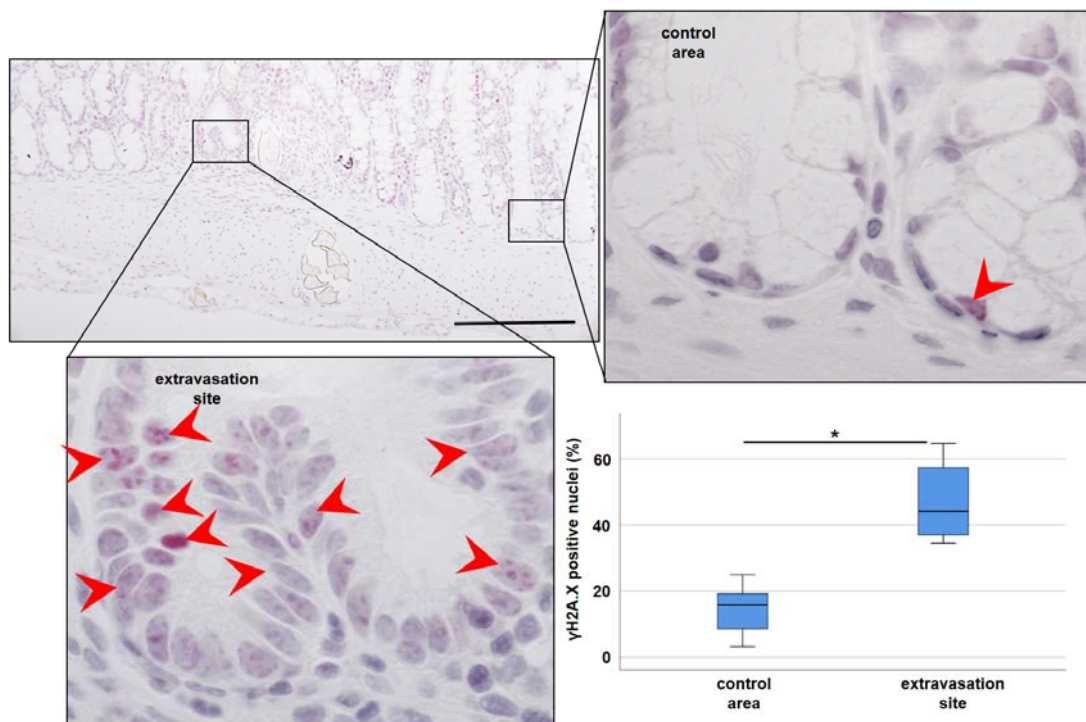


Suppl. Fig. 8: Cyclin D positive nuclei of enterocytes in the lower third of crypts in direct vicinity to extravasation sites of *S. mansoni* eggs in bisex infected hamster colon were counted. For control, Cyclin D positive nuclei of enterocytes in the lower third of crypts with normal appearance were counted. Red arrowheads depict some of the Cyclin D positive nuclei. At least 10 positive and negative area in at least 3 representative specimens were used for quantification at 1000 x.

Suppl. Fig. 9

Suppl. Fig. 9: Inhibition of the JNK/c-Jun pathway in SW620 cells by SP600125 reduced SEA activated Cyclin D1 expression. One representative western blot of three.

Suppl. Fig. 10



Suppl. Fig. 10: γ H2A.X positive nuclei of enterocytes in the lower third of crypts in direct vicinity to extravasation sites of *S. mansoni* eggs in bisex infected hamster colon were counted. For control, γ H2A.X positive nuclei of enterocytes in the lower third of crypts with normal appearance were counted. Red arrowheads depict some of the γ H2A.X positive nuclei. At least 10 positive and negative area in at least 3 representative specimen were used for quantification.

Original Blots:

Fig. 1a: Array consists of 2 membranes with different analytes for each group.

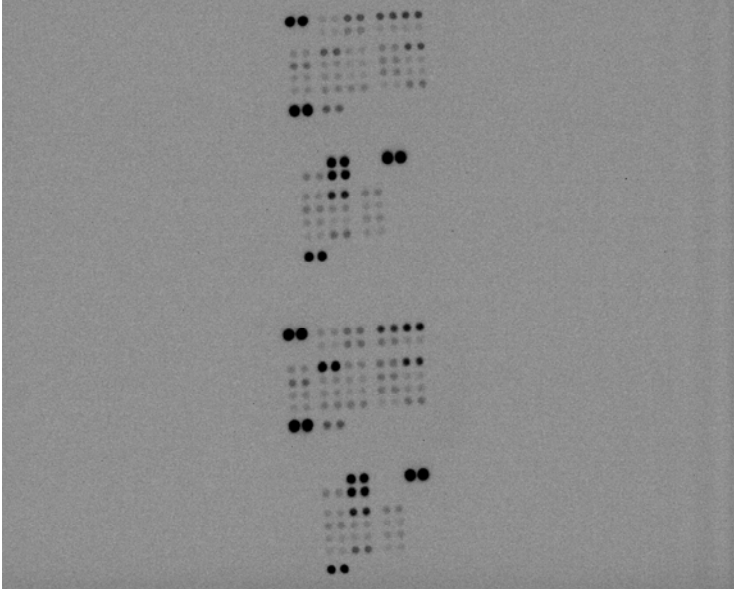


Fig. 1c p-Gsk3 β ^{Ser9}: please note, that the standard lane is an incident light photograph introduced by INTAS imager

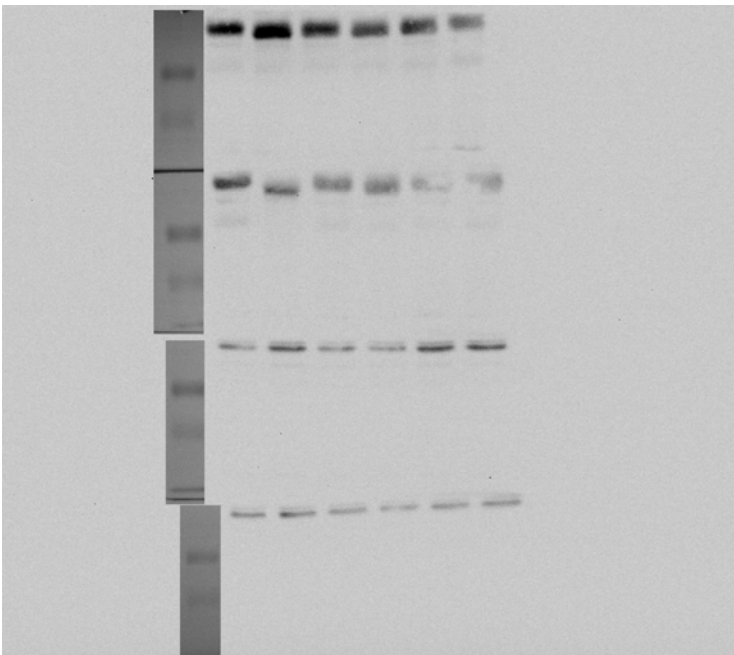


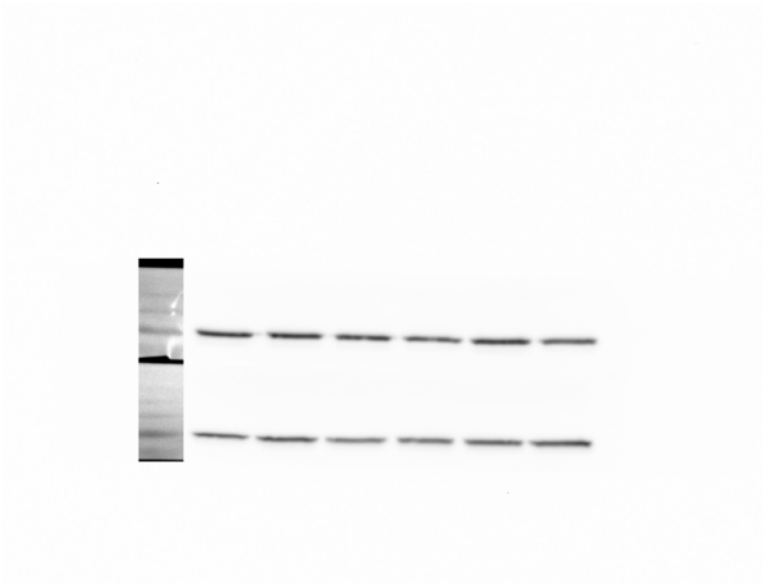
Fig. 1c β -Catenin:

Fig. 1c Gapdh:

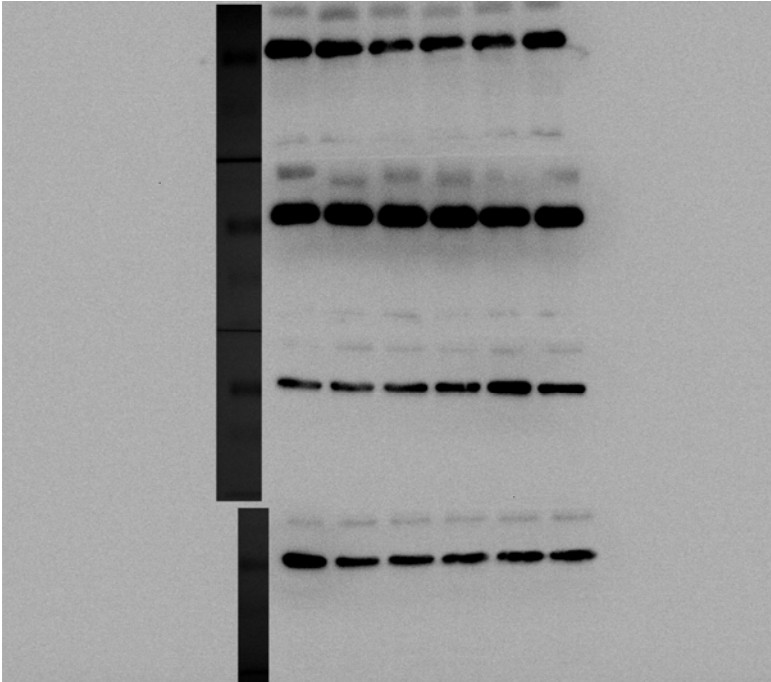


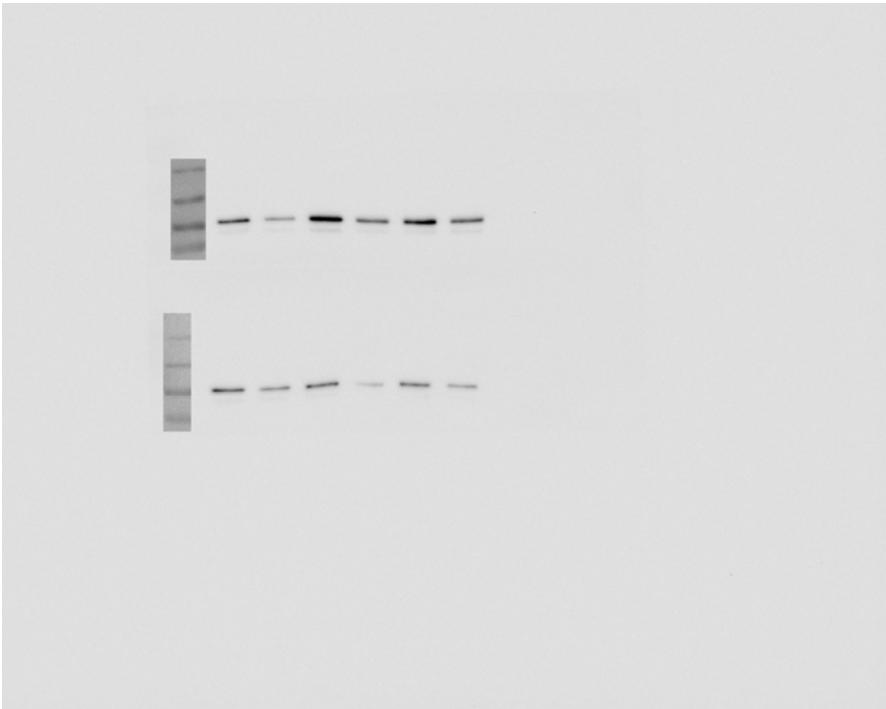
Fig. 1d β -catenin

Fig. 1d Gapdh

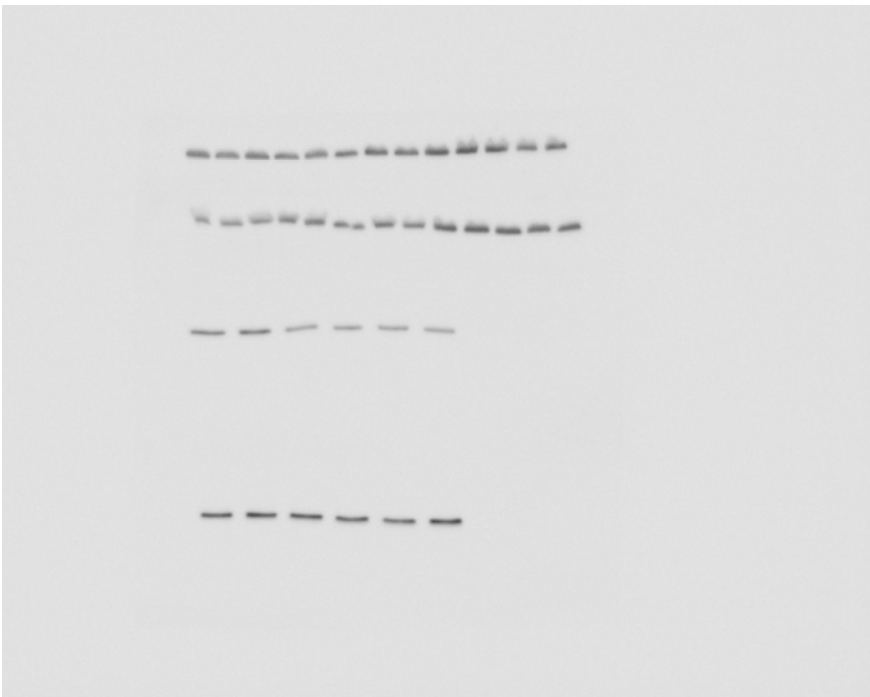


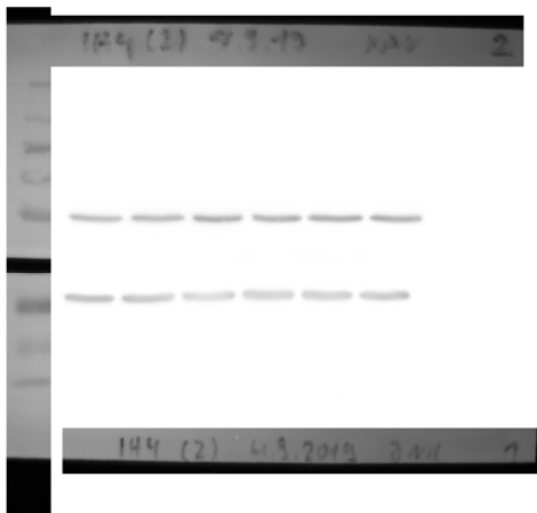
Fig. 1d p- β -Catenin: (old before revision)Fig 1d α -Tubulin: (old before revision)

Fig. 2b p-c-Jun:

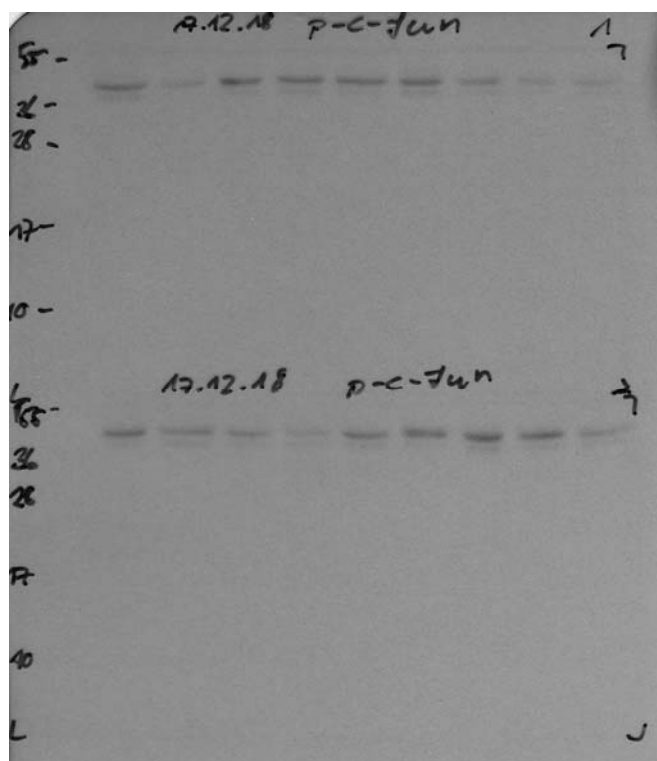


Fig 2b c-Jun:

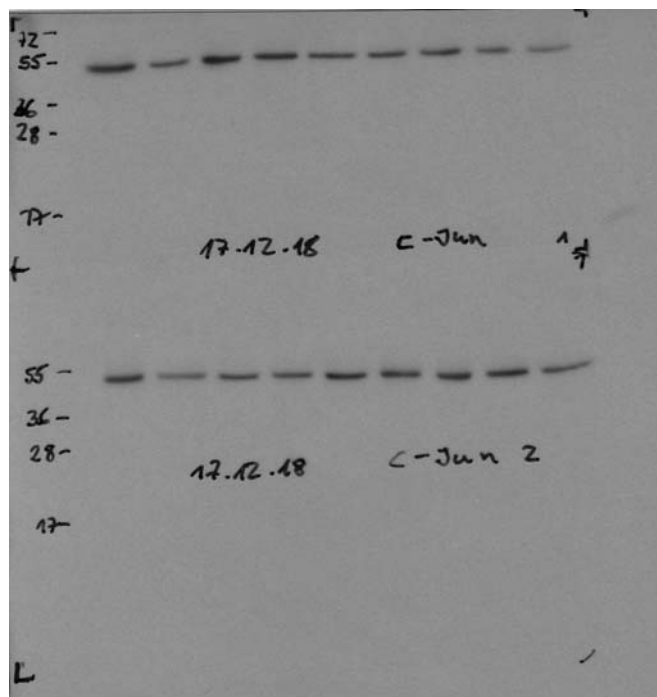


Fig. 2b Gapdh:

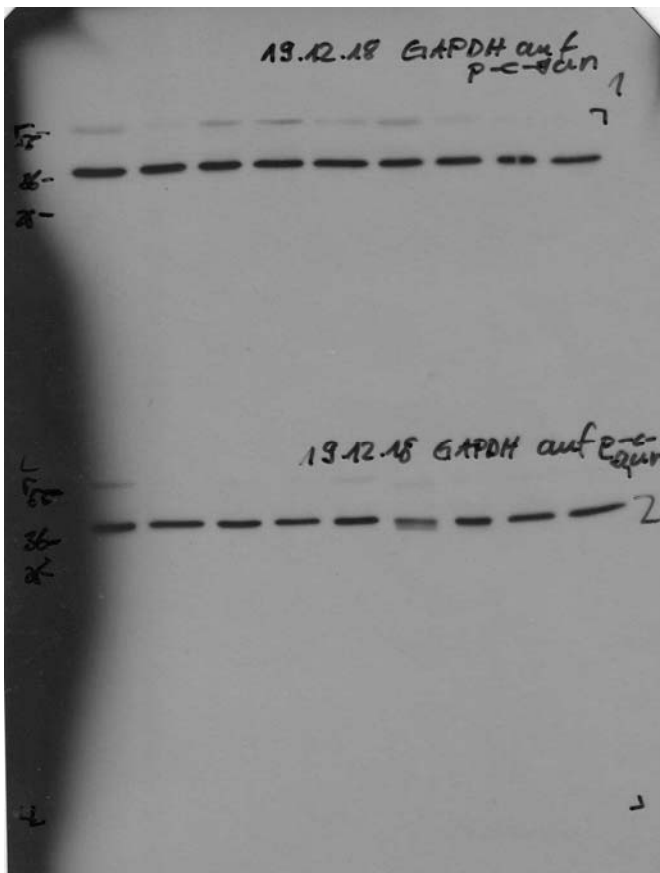


Fig. 2e AP-1 EMSA:



Fig. 3a p-c-Jun:

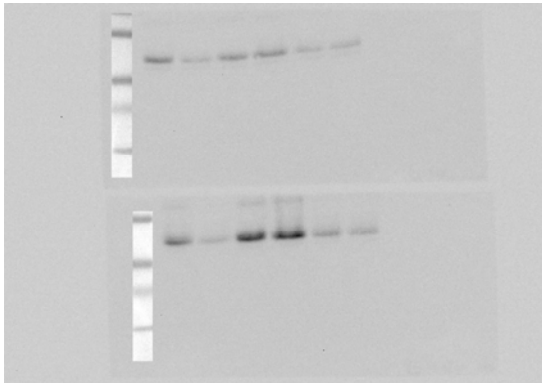


Fig 3a c-Jun:

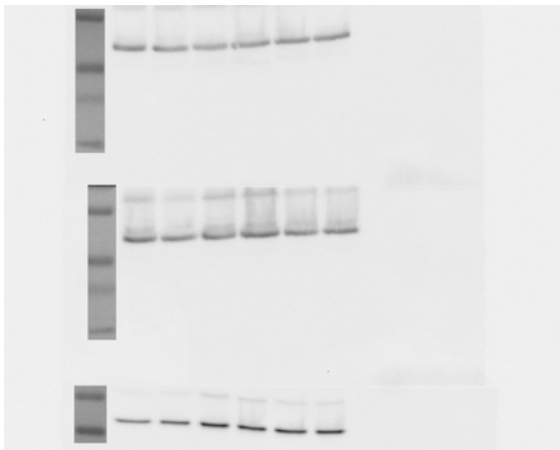


Fig. 3a Gapdh:

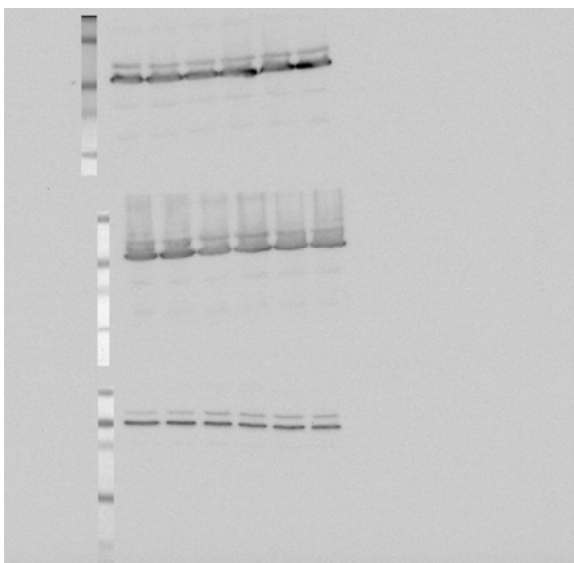


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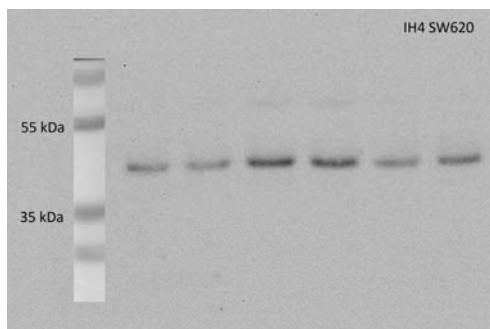


Fig. 3b c-Jun:

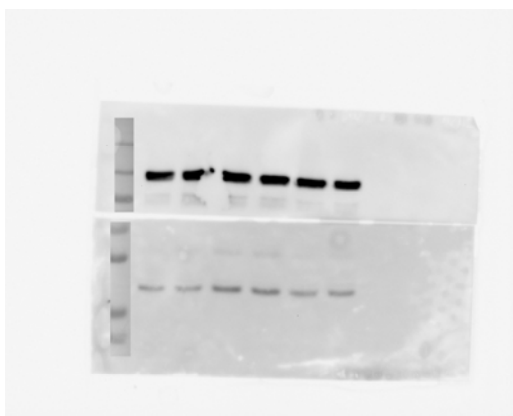


Fig. 3b Gapdh:

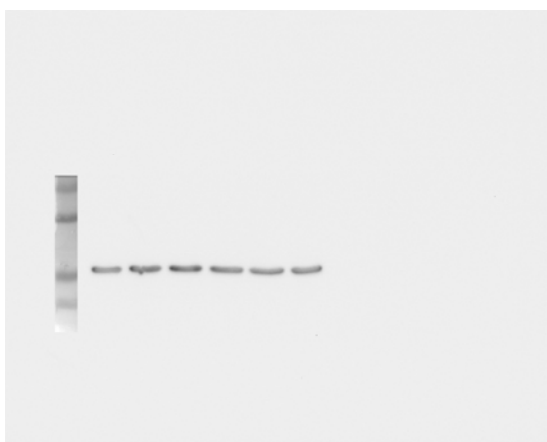


Fig. 3c + d p-c-Jun:

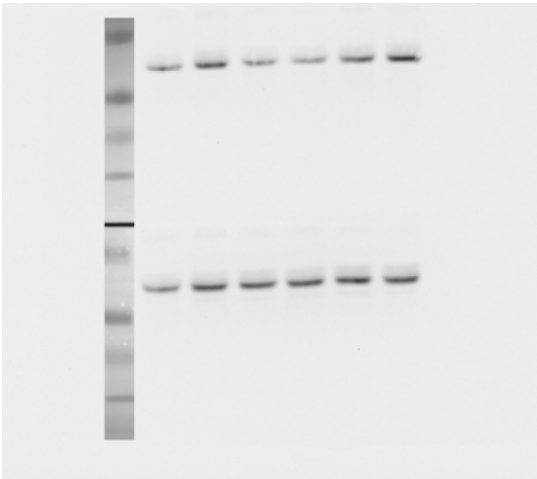


Fig. 3c+d c-Jun:

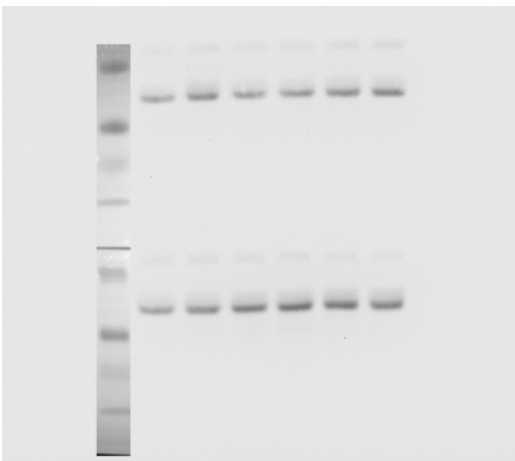


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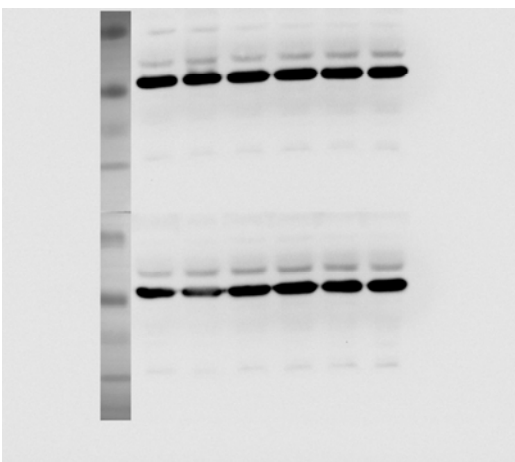


Fig. 3e p-c-Jun:

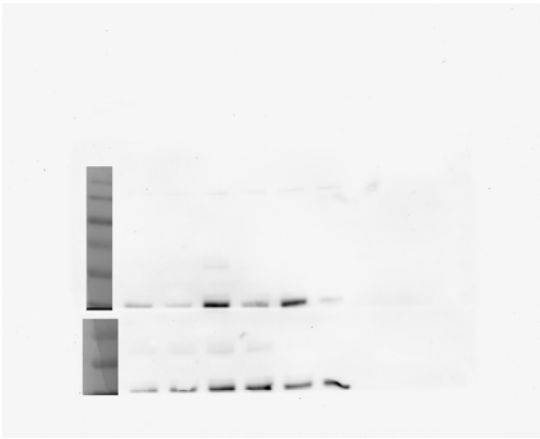


Fig. 3 e c-Jun:

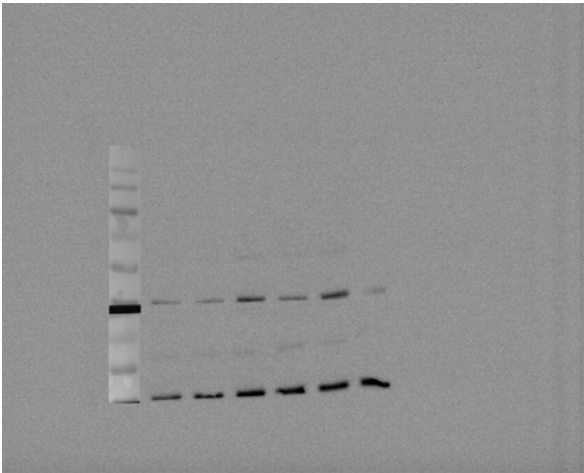


Fig. 3e Gapdh:

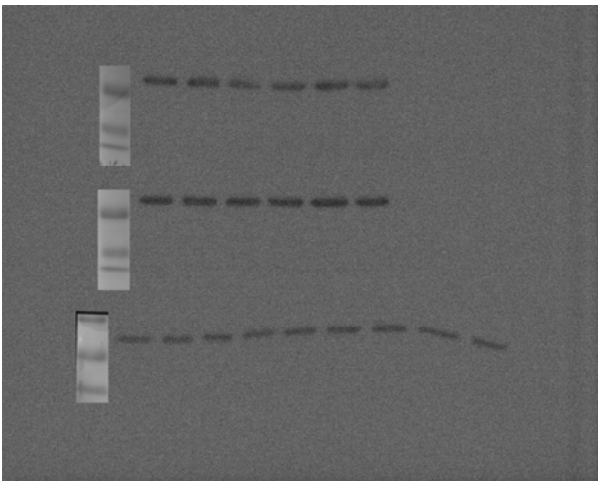


Fig. 4c Cyclin D1:

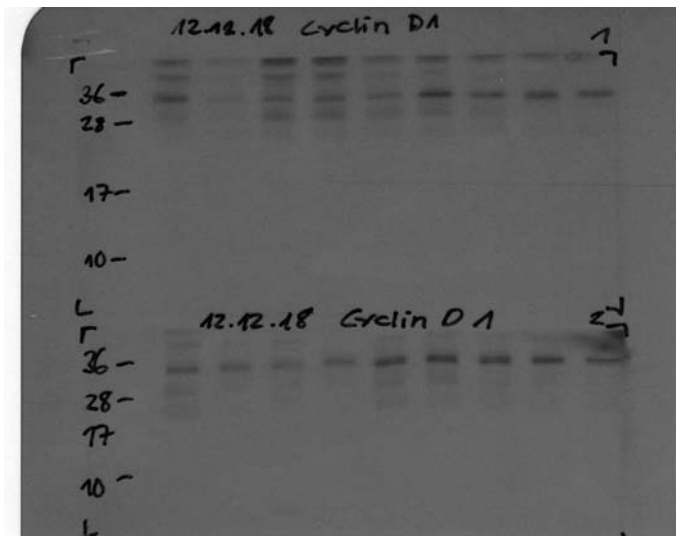


Fig. 4c Gapdh_a:

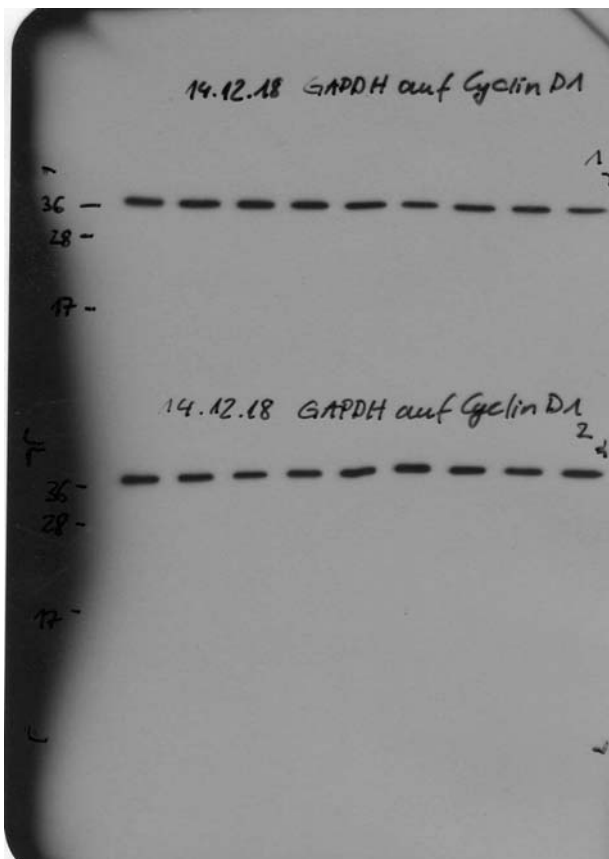


Fig. 4c Mcm2:

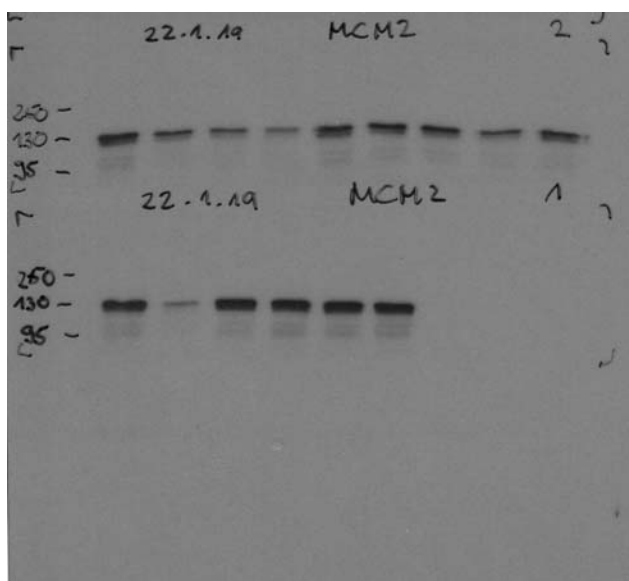


Fig. 4c Gapdh_b:

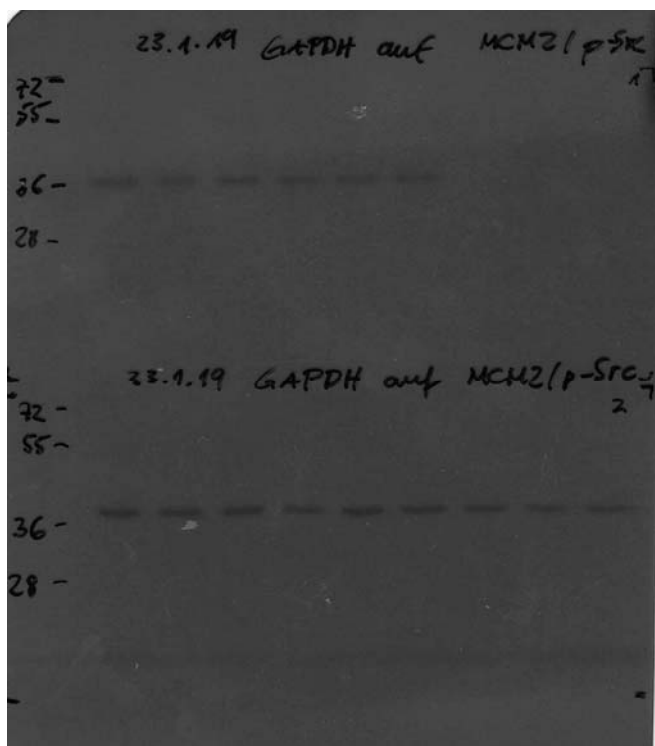


Fig. 4d Cyclin D1_a:

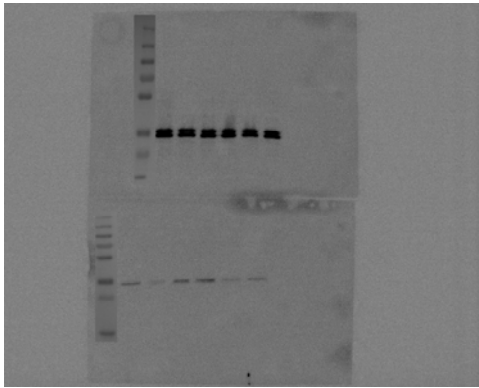
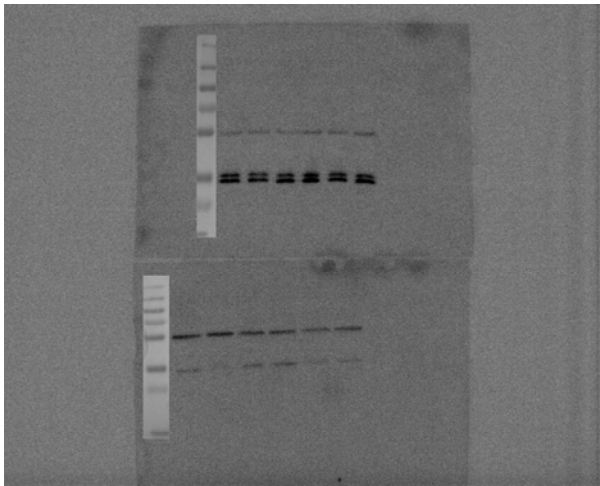
Fig. 4d α -Tubulin_a:

Fig. 4d Cyclin D1_b:

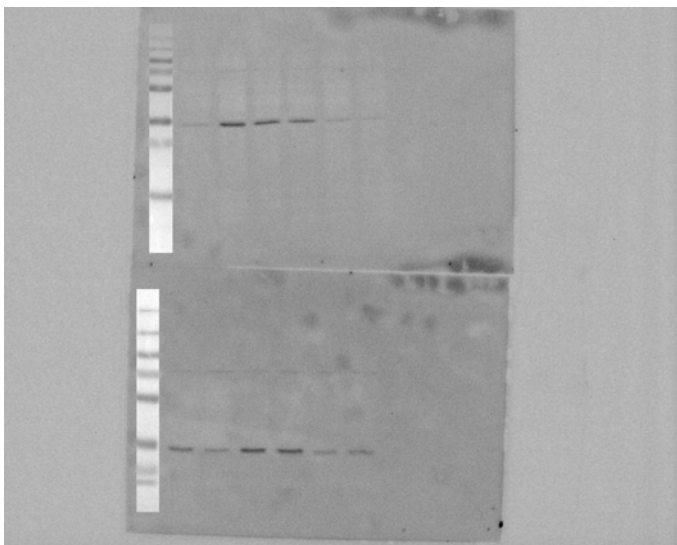


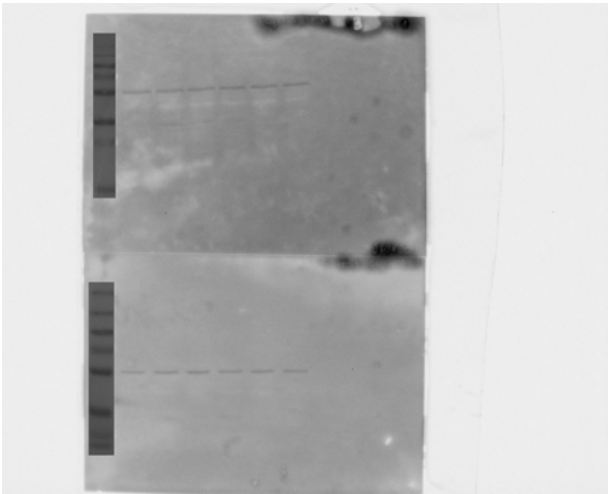
Fig. 4d α -Tubulin_b:

Fig. 4e Cyclin D1:

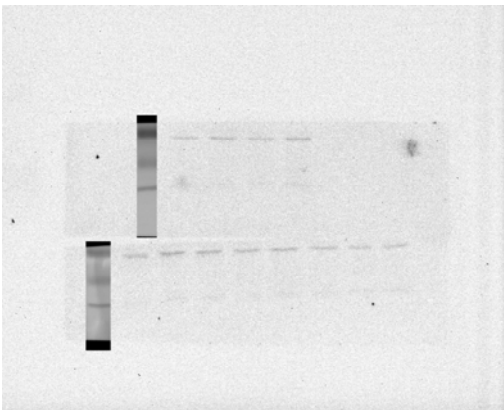
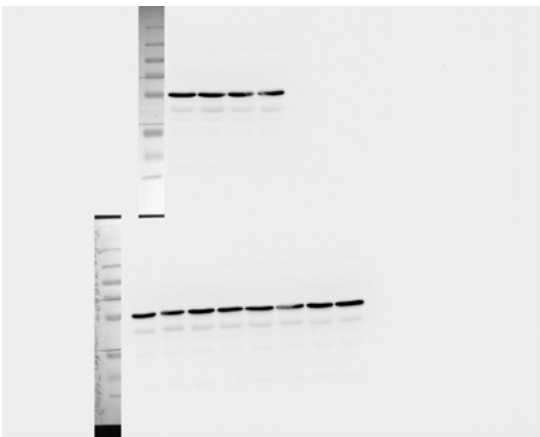
Fig. 4e α -Tubulin:

Fig. 4f Cyclin D1:

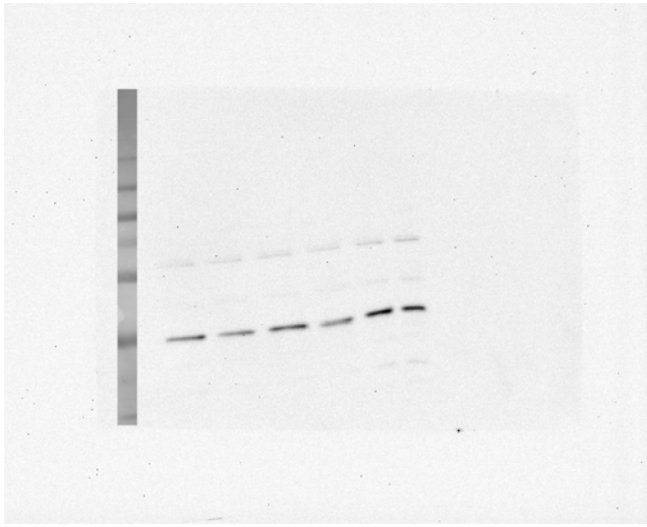


Fig. 4f Gapdh:

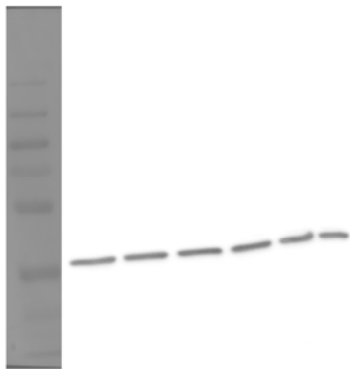


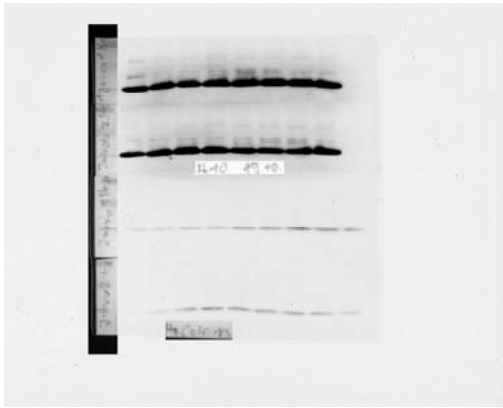
Fig. 5a γ H2a.x:

Fig. 5a Gapdh_a:

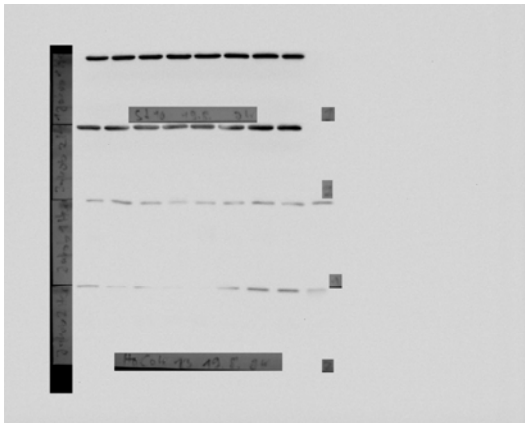


Fig. 5a Parp1:

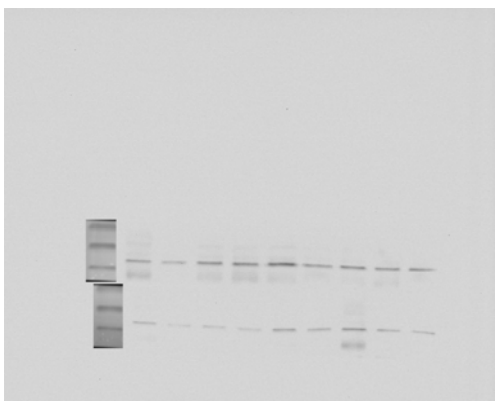


Fig. 5a Gapdh_b:

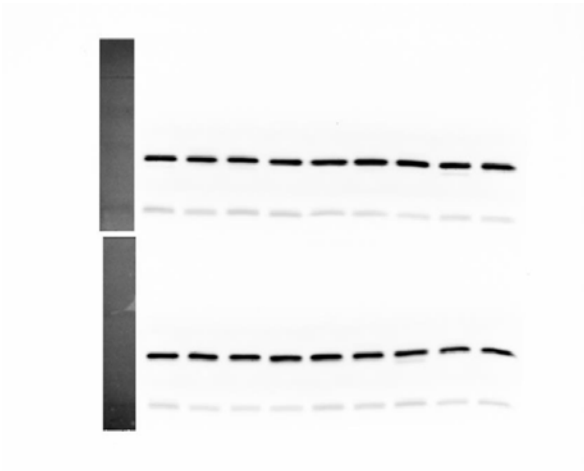
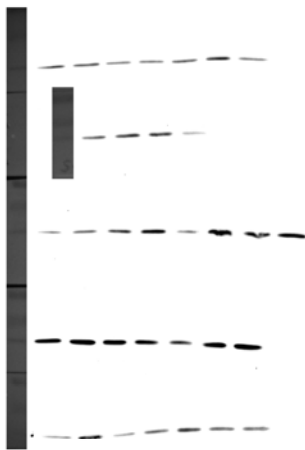
Fig. 5c γ H2a.x:

Fig. 5c Gapdh_a:

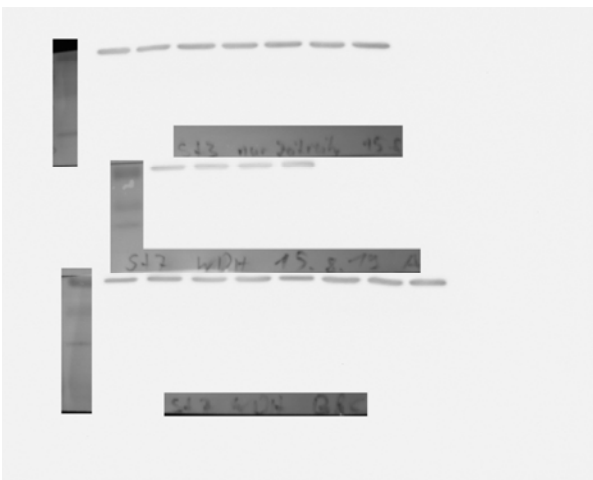


Fig. 5 c Parp1:

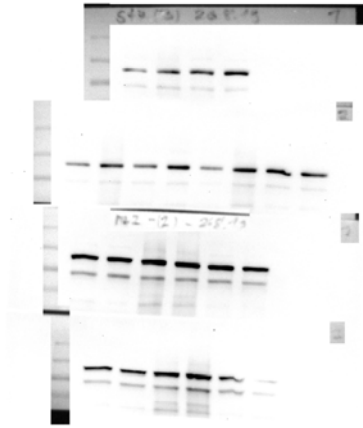
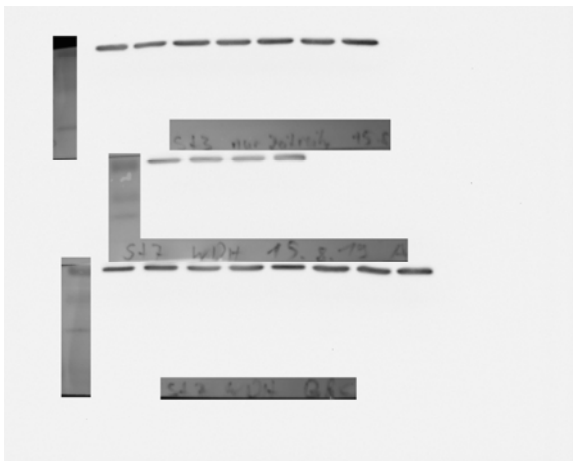
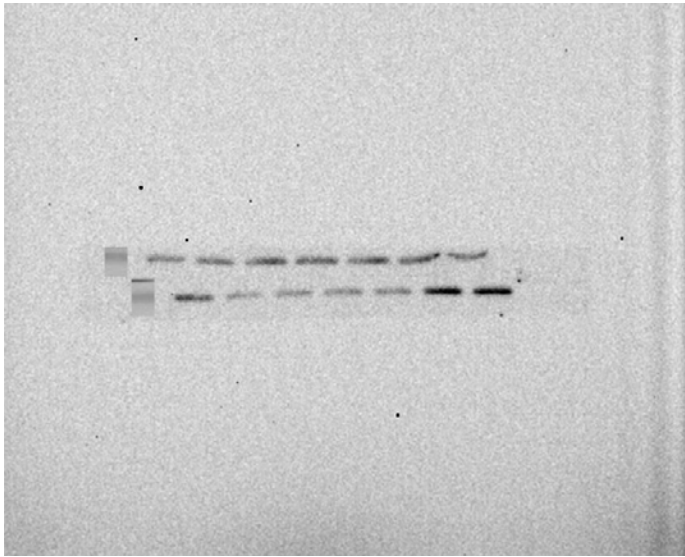


Fig. 5c Gapdh_b:



Suppl. Fig. 9 α -Tubulin:

Suppl. Fig. 9 Cyclin D1:

