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Supplemental Information

D614G Mutation Alters SARS-CoV-2

Spike Conformation and Enhances

Protease Cleavage at the S1/S2 Junction

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Figure S1. Structural comparison of residues 986-987 in the SARS-CoV-2 S ectodomain with the MERS and SARS-CoV-1 S ectodomains, Related to Figure 2. A. Cryo-EM reconstruction map of the 1-RBD-up states of the SARS-CoV-2 S-GSAS ectodomain (EMDB-22822; PDB ID 7KDH) colored by chains and zoomed-in view of residues K986-V987 in the 1-RBD-up and 3-RBD-down structures. The cryo-EM map is shown as a transparent surface and the fitted model is in cartoon representation, with residues shown as balls and sticks. B. Sequence alignment of residues 981-992 of SARS-CoV-2 and corresponding residues of SARS-CoV-1 and MERS spike proteins. C. Magnified view of one protomer showing residues V1060 and L1061 from MERS (PDB 5X5C; in yellow) and SARS-CoV-1 (PDB 5X58; in yellow) overlayed with residues K986 and V987 from SARS-CoV-2 S-GSAS ectodomain (colored as in A).



Figure S2. Overlay of the S-GSAS/D614G 1-up and 3-down substates, Related to Figure 3. Side and top view of the superposition of the **A.** 1-up substate structures of S-GSAS/D614G from Figure **3D** (PDB ID 7KE9, 7KEA, 7KEB and 7KEC) and **C.** of the 3-down substate structures (PDB ID 7KE4, 7KE6, 7KE7 and 7KE8) from Figure **3E.** The structures were superimposed using S2 subunit residues 908-1035 (spanning the HR1 and CH regions) with the S1 subunit colored by domain and the S2 subunit colored grey. RBD is colored red, NTD green, SD1 dark blue, SD2 orange and the linker between the NTD and RBD colored cyan. **B.** Ribbon representation of the overlay of the 1-up substate structures of S-GSAS/D614G colored by chain (PDB ID 7KE9 and 7KEC). **D.** Ribbon representation of the overlay of the 3-down substate structures of S-GSAS/D614G colored by state (PDB ID 7KE4, 7KE7 and 7KE8).



Figure S3. Difference distance matrices (DDM) analysis of S-GSAS and S-RRAR/D614G fully cleaved by furin showing structural changes between different protomers, Related to Figure 4. The S-GSAS DDM are shown in A

and S-RRAR/D614G in **B**. The blue to white to red coloring scheme is illustrated at the bottom.

RBD "up" (1-RBD-up S) vs

RBD "down" protomer #2 (1-RBD-up S)

1027

827

1027

821

621

427

227

27

227

427 Residu

627

RBD "down" protomer (3-RBD-down S)

Residue numbe





RBD "up" (1-RBD-up S) vs RBD "down" protomer (3-RBD-down S)

Δ RBD "up" (1-RBD-up S) vs RBD "down" protomer #1 (1-RBD-up S) 1027 827 numbe 627 Residue 227 27 427 Residue 627 827 1027 umbe

RBD "down" protomer #1 (1-RBD-up S) vs

RBD "down" protomer #2 (1-RBD-up S)

Table S1. Cryo-EM data collection and refinements statistics for the S-GSAS, S-GSAS/D614G and furin cleaved S-RRAR/D614G SARS-CoV-2 spike ectodomains, Related to Figures 2, 3 and 7.

	S-G	SAS	S-GSAS Conse	S/D614G ensus	S-RRAR/D614G Furin cleaved						
	3-down	1-up	3-down	1-up	3-down	1-up					
PDB ID	7KDG	7KDH	7KDK	7KDL	7KDI	7KDJ					
EMDB ID	22821	22822	22825	22826	22823	22824					
Data collection and processing											
Microscope	FEI Titan Krios										
Detector	Gatan K3										
Magnification	81,0	000	81,	000	81,000						
Voltage (kV)	30	00	30	00	300						
Electron exposure (e–/Å2)	65.	.24	51	.8	65.94						
Defocus range (µm)	0.8-	2.5	0.40	-2.94	0.38-2.88						
Pixel size (Å)	1.0	69	1.0)58	1.058						
Reconstruction software			cryoS	parc							
Symmetry imposed	C3	C1	C3	C1	C3	C1					
Initial particle images (no.)	2,566,724		2,343,150		479,208						
Final particle images (no.)	581,495	175,529	782,485	613,271	224,310	99,165					
Map resolution (Å)	3.01	3.33	2.8	2.96	3.26	3.49					
FSC threshold	0.143	0.143	0.143	0.143	0.143	0.143					
Refinement											
Initial model used	6VXX	6VYB	6VXX	6VYB	6VXX	6VYB					
Model resolution (Å)	3.01	3.33	2.8	2.96	3.26	3.49					
FSC threshold	0.143	0.143	0.143	0.143	0.143	0.143					
Map sharpening B factor (Å ²)	-129.3	-101.3	-121.5	-106.9	-104.2	-94.8					
Model composition											
Nonhydrogen atoms	23,700	22,698	23,688	22,365	23,688	22,365					
Protein residues	2,916	2,891	2,916	2,875	2,916	2,875					
R.m.s. deviations											
Bond lengths (Å)	0.013	0.012	0.013	0.012	0.012	0.012					
Bond angles (°)	1.845	1.838	1.87	1.835	1.789	1.757					
Validation											
MolProbity score	1.03	1.13	0.98	0.96	0.97	1.03					
Clashscore	0.28	0.43	0.3	0.09	0.15	0.23					
Poor rotamers (%)	0.31	0.56	0.12	0.27	0.27	0.44					
Ramachandran plot											
Favored (%)	93.46	92.3	94.66	93.6	93.81	93.21					
Allowed (%)	6.22	7.31	5.24	6.11	6.12	6.4					
Disallowed (%)	0.32	0.39	0.11	0.29	0.07	0.39					

Table S1 continued. Cryo-EM data collection and refinements statistics for the S-GSAS, S-GSAS/D614G and furin cleaved S-RRAR/D614G SARS-CoV-2 spike protein ectodomains, Related to Figures 2, 3 and 7.

	S-GSAS/D614G									
		Substates								
		3-de				<u>1-I</u>				
	7KE4 22831	7NE0 22832	/NE/ 22833	7KE0 22834	7KE9 22835	7NEA 22836	7NED 22837	7 NEU 22838		
Data collection and processin	22001	22052	22000	22004	22000	22030	22007	22030		
Magnification	9			81	000					
Voltage (kV)	300									
Electron exposure $(e_{-}/Å^{2})$	51 8									
Defocus range (µm)	0.40-2.94									
Pixel size (Å)	1.058									
Symmetry imposed	C1									
Initial particle images (no.)	2,343,150									
Final particle images (no.)	182,326	196,173	157,254	133,373	287,765	145,566	127,429	58,580		
Map resolution (Å)	3.21	3.1	3.32	3.26	3.08	3.33	3.48	3.84		
FSC threshold	0.143	0.143	0.143	0.143	0.143	0.143	0.143	0.143		
Refinement										
Initial model used	6VXX	6VXX	6VXX	6VXX	6VYB	6VYB	6VYB	6VYB		
Model resolution (Å)	3.21	3.1	3.32	3.26	3.08	3.33	3.48	3.84		
FSC threshold	0.143	0.143	0.143	0.143	0.143	0.143	0.143	0.143		
Map sharpening B factor (Å ²)	-95.3	-98.8	-92.8	-91.2	-104.5	-88.9	-86.2	-81		
Model composition										
Nonhydrogen atoms	23,684	23,688	23,688	23,684	22,365	22,365	22,365	22,33 7		
Protein residues	2,916	2,916	2,916	2,916	2,875	2,875	2,875	2,875		
R.m.s. deviations										
Bond lengths (Å)	0.013	0.013	0.013	0.013	0.012	0.012	0.012	0.012		
Bond angles (°)	1.845	1.838	1.81	1.836	1.813	1.893	1.856	1.845		
Validation										
MolProbity score	1.02	1	1.1	1.13	0.95	1.23	1.3	1.58		
Clashscore	0.13	0.15	0.34	0.41	0.11	0.64	0.83	0.74		
Poor rotamers (%)	1.06	1.02	0.08	1.1	0.04	1.24	0.13	2.44		
Ramachandran plot										
Favored (%)	92.93	93.25	92.23	92.9	93.99	92.42	90.31	89.92		
Allowed (%)	6.72	6.47	7.49	6.82	5.79	7.15	9.08	9.05		
Disallowed (%)	0.35	0.28	0.28	0.28	0.21	0.43	0.61	1.04		



S-GSAS (lot SG7): 96% spike

Data S1. NSEM and Cryo-EM data processing for the S-GSAS SARS-CoV-2 S ectodomain, Related to Figure 2. A. Representative NSEM micrograph. **B**. NSEM Particle picks in green. **C**. 2D class averages; the particles in the classes marked with a red box are recognizable as the pre-fusion spike. The rest of the classes are marked as "junk". **D**. NSEM Particles present in the boxed class averages in panel **C**. are shown with a green circle around them.



9,815 micrographs; 4,291,642 particles

Ε



G_{Representative 2-D class averages}



2,566,724 particles selected



Data S1 continued. NSEM and Cryo-EM data processing for the S-GSAS SARS-CoV-2 S ectodomain, Related to Figure 2. E. Representative cryo-EM micrograph. **F**. Cryo-EM CTF fit. **G**. Representative 2D class averages from Cryo-EM dataset. **H-I.** *Ab initio* reconstructions for the cryo-EM **H**. 1-up state (the RBD in the up position is identified by an asterisk) and **I**. 3-down state. **J-K**. Refined maps for the cryo-EM **J**. 1-up state (the RBD in the up position is identified by an asterisk) and **K**. 3-down state. **L-M**. Fourier shell correlation curves for the cryo-EM **L**. 1-up state and **M**. 3-down state.



Data S1 continued. NSEM and Cryo-EM data processing for the S-GSAS SARS-CoV-2 S ectodomain, Related to Figure 2. N-O. Refined cryo-EM maps colored by local resolution for the **N.** 3-RBD-down (EMDB-22821) and **O.** 1-RDB-up (EMD-22822) states. **P.** Zoom-in images showing the S2, SD2, D614, S1', S2' and K986-V987 regions in the 3-RBD-down (top) and 1-RDB-up structures (bottom). The cryo-EM map is shown as a transparent surface and the fitted model is in cartoon representation, with residues shown as balls and sticks (PDB ID 7KDG and 7KDH).

Data S1. NSEM and Cryo-EM data processing for the S-GSAS SARS-CoV-2 S ectodomain, Related to Figure 2. A. Representative NSEM micrograph. **B.** NSEM Particle picks in green. **C.** 2D class averages; the particles in the classes marked with a red box are recognizable as the pre-fusion spike. The rest of the classes are marked as "junk". **D.** NSEM Particles present in the boxed class averages in panel C. are shown with a green circle around them. **E.** Representative cryo-EM micrograph. **F.** Cryo-EM CTF fit. **G.** Representative 2D class averages from Cryo-EM dataset. **H-I.** *Ab initio* reconstructions for the cryo-EM **H.** 1-up state (the RBD in the up position is identified by an asterisk) and **I.** 3-down state. **J-K.** Refined maps for the cryo-EM **J.** 1-up state (the RBD in the up position is identified by an asterisk) and **K** 3-down state. **L-M.** Fourier shell correlation curves for the cryo-EM **L.** 1-up state and **M.** 3-down state. **N-O.** Refined cryo-EM maps colored by local resolution for the **N.** 3-RBD-down (EMDB-22821) and **O.** 1-RDB-up (EMD-22822) states. **P.** Zoom-in images showing the S2, SD2, D614, S1', S2' and K986-V987 regions in the 3-RBD-down (top) and 1-RDB-up structures (bottom). The cryo-EM map is shown as a transparent surface and the fitted model is in cartoon representation, with residues shown as balls and sticks (PDB ID 7KDG and 7KDH).



S-GSAS/D614G (lot SG15): 83% spike

Data S2. NSEM and Cryo-EM data processing for the S-GSAS/D614G ectodomain consensus structures, Related to Figure 3. A. Representative NSEM micrograph. **B.** NSEM Particle picks in green. **C.** 2D class averages; the particles in the classes marked with a red box are recognizable as the pre-fusion spike. The rest of the classes are marked as "junk". **D.** NSEM Particles present in the boxed class averages in panel **C**. are shown with a green circle around them.



Ε

9021 micrographs; 3,962,746 particles

CTF fit

G_{Representative 2-D class averages}



2,343,150 particles selected



Data S2 continued. NSEM and Cryo-EM data processing for the S-GSAS/D614G ectodomain consensus structures, Related to Figure 3. E. Representative cryo-EM micrograph. F. Cryo-EM CTF fit. G. Representative 2D class averages from Cryo-EM dataset. H-I. *Ab initio* reconstructions for the cryo-EM H. 1-up state (the RBD in the up position is identified by an asterisk) and I. 3-down state. J-K. Refined maps for the cryo-EM J. 1-up state (the RBD in the up position is identified by an asterisk) and K 3-down state. L-M. Fourier shell correlation curves for the cryo-EM L. 1-up state and M. 3-down state.





Data S2 continued. NSEM and Cryo-EM data processing for the S-GSAS/D614G ectodomain consensus structures, Related to Figure 3. N-O. Refined cryo-EM maps colored by local resolution for the consensus **N.** 3-RBD-down (EMDB-22825) and **O.** 1-RDB-up (EMD-22826) states. **P.** Zoom-in images showing the S2, SD2, D614, S1', S2' and K986-V987 regions in the consensus 3-RBD-down (top) and 1-RDB-up structures (bottom). The cryo-EM map is shown as a transparent surface and the fitted model is in cartoon representation, with residues shown as balls and sticks (PDB ID 7KDK and 7KDL).

Data S2. NSEM and Cryo-EM data processing for the S-GSAS/D614G ectodomain consensus structures, Related to Figure 3. A. Representative NSEM micrograph. **B.** NSEM Particle picks in green. **C.** 2D class averages; the particles in the classes marked with a red box are recognizable as the pre-fusion spike. The rest of the classes are marked as "junk". **D.** NSEM Particles present in the boxed class averages in panel **C**. are shown with a green circle around them. **E.** Representative cryo-EM micrograph. **F.** Cryo-EM CTF fit. **G.** Representative 2D class averages from Cryo-EM dataset. **H-I.** *Ab initio* reconstructions for the cryo-EM **H.** 1-up state (the RBD in the up position is identified by an asterisk) and **I.** 3-down state. **J-K.** Refined maps for the cryo-EM **J.** 1-up state (the RBD in the up position is identified by an asterisk) and **K** 3-down state. **L-M.** Fourier shell correlation curves for the cryo-EM **L.** 1up state and **M.** 3-down state. **N-O.** Refined cryo-EM maps colored by local resolution for the consensus **N.** 3-RBDdown (EMDB-22825) and **O.** 1-RDB-up (EMD-22826) states. **P.** Zoom-in images showing the S2, SD2, D614, S1', S2' and K986-V987 regions in the consensus 3-RBD-down (top) and 1-RDB-up structures (bottom). The cryo-EM map is shown as a transparent surface and the fitted model is in cartoon representation, with residues shown as balls and sticks (PDB ID 7KDK and 7KDL).



Data S3. Cryo-EM data processing details for S-GSAS/D614G 1-up and 3-down subpopulations, Related to Figure 3D and E. A-E. *Ab initio* reconstructions, refined maps and Fourier shell correlation curves for the 1-up subpopulations (the RBD in the up position is identified by an asterisk).





3-down state 182,326 particles C1 symmetry



3-down state 196,173 particles C1 symmetry



3-down state 157,254 particles C1 symmetry



3-down state 133,373 particles C1 symmetry



[#]3-down state 182,326 particles 3.2 Å, C1 symmetry; EMD-22831



3-down state 196,173 particles 3.1 Å, C1 symmetry; EMD-22832



Fourier shell

Correlation GSFSC Resolution: 3.21Å

^{~~}3-down state 157,254 particles 3.3 Å, C1 symmetry; EMD-22833





3-down state 133,373 particles 3.3 Å, C1 symmetry; EMD-22834

Data S3 continued. Cryo-EM data processing details for S-GSAS/D614G 1-up and 3-down subpopulations, Related to Figure 3D and E. F-K. *Ab initio* reconstructions, refined maps and Fourier shell correlation curves for the 3-down subpopulations.



Data S3 continued. Cryo-EM data processing details for S-GSAS/D614G 1-up and 3-down subpopulations, Related to Figure 3D and E. F-K. *Ab initio* reconstructions, refined maps and Fourier shell correlation curves for the 3-down subpopulations.

Data S3. Cryo-EM data processing details for S-GSAS/D614G 1-up and 3-down subpopulations, Related to Figure 3D and E. A-E and overlay of the S-GSAS/D614G 1-up and 3-down substates. *Ab initio* reconstructions, refined maps and Fourier shell correlation curves for the 1-up subpopulations (the RBD in the up position is identified by an asterisk). F-K. *Ab initio* reconstructions, refined maps and Fourier shell correlation curves for the 3-down subpopulations.







S-HRV3C (lot 115KJ): 54% spike

Data S4. NSEM quality control workflow for the S-HRV3C and S-HRV3C/D614G SARS-CoV-2 S ectodomains, Related to Figure 5. A and E. Representative NSEM micrograph of **A.** S-HRV3C and **E.** S-HRV3C/D614G SARS-CoV-2 S ectodomains. **B and F.** Particle picks in green. **C and G.** 2D class averages; the particles in the classes marked with a red box are recognizable as the pre-fusion spike. The rest of the classes are marked as "junk". **D and H.** Particles present in the boxed class averages in panel C. are shown with a green circle around them.







S-HRV3C/D614G (lot 116KJ): 91% spike

Data S4 continued. NSEM quality control workflow for the S-HRV3C and S-HRV3C/D614G SARS-CoV-2 S ectodomains, Related to Figure 5. A and E. Representative NSEM micrograph of **A.** S-HRV3C and **E.** S-HRV3C/D614G SARS-CoV-2 S ectodomains. **B and F.** Particle picks in green. **C and G.** 2D class averages; the particles in the classes marked with a red box are recognizable as the pre-fusion spike. The rest of the classes are marked as "junk". **D and H.** Particles present in the boxed class averages in panel C. are shown with a green circle around them.







S-RRAR (lot 117KJ): 56% spike

Data S5. NSEM and Cryo-EM data processing for the S-RRAR and S-RRAR/D614G SARS-CoV-2 S ectodomain, Related to Figure 6 and 7. A-D. S-RRAR NSEM. A. Representative NSEM micrograph. B. NSEM Particle picks in green. C. 2D class averages (the particles in the classes marked with a red box are recognizable as the pre-fusion spike). The rest of the classes are marked as "junk". D. NSEM Particles present in the boxed class averages in panel C. are shown with a green circle around them.



S-RRAR/D614G (lot 120KJ): 89% spike

Data S5 continued. NSEM and Cryo-EM data processing for the S-RRAR and S-RRAR/D614G SARS-CoV-2 S ectodomain, Related to Figure 6 and 7. E-H. S-RRAR/D614G NSEM. **E.** Representative NSEM micrograph. **F.** NSEM Particle picks in green. **G.** 2D class averages (the particles in the classes marked with a red box are recognizable as the pre-fusion spike). The rest of the classes are marked as "junk". **H.** NSEM Particles present in the boxed class averages in panel **G**. are shown with a green circle around them.



Data S5 continued. NSEM and Cryo-EM data processing for the S-RRAR and S-RRAR/D614G SARS-CoV-2 S ectodomain, Related to Figure 6 and 7. I. Representative cryo-EM micrograph for furin cleaved S-RRAR-D614G. **J.** Cryo-EM CTF fit furin cleaved S-RRAR-D614G. **K.** Representative 2D class averages from Cryo-EM dataset for furin cleaved S-RRAR-D614G. **L-N.** *Ab initio* reconstructions for the cryo-EM for furin cleaved S-RRAR-D614G **L.** 1-up state (the RBD in the up position is identified by an asterisk), **M**. missing-RBD state (the RBD in the up position or missing is identified by an asterisk) and **N.** 3-down state. **O-Q.** Refined maps for the cryo-EM for furin cleaved S-RRAR-D614G **O.** 1-up state (the RBD in the up position is identified by an asterisk) and **Q.** 3-down state. **R-T.** Fourier shell correlation curves for the cryo-EM furin cleaved S-RRAR-D614G **R.** 1-up state, **S.** missing-RBD state and **T.** 3-down state.





Data S5 continued. NSEM and Cryo-EM data processing for the S-RRAR and S-RRAR/D614G SARS-CoV-2 S ectodomain, Related to Figure 6 and 7. U-V. Refined cryo-EM maps colored by local resolution for the consensus U. 3-RBD-down (EMDB-22823) and V. 1-RDB-up (EMD-22824) states. W. Zoom-in images showing the S2, SD2, D614G, S1', S2' and K986-V987 regions in the consensus 3-RBD-down (top) and 1-RDB-up structures (bottom). The cryo-EM map is shown as a transparent surface and the fitted model is in cartoon representation, with residues shown as balls and sticks (PDB ID 7KDJ and 7KDI).

Data S5. NSEM and Cryo-EM data processing for the S-RRAR and S-RRAR/D614G SARS-CoV-2 S ectodomain, Related to Figure 6 and 7. A-D. S-RRAR NSEM. A. Representative NSEM micrograph. B. NSEM Particle picks in green. C. 2D class averages (the particles in the classes marked with a red box are recognizable as the pre-fusion spike). The rest of the classes are marked as "junk". D. NSEM Particles present in the boxed class averages in panel C. are shown with a green circle around them. E-H. S-RRAR/D614G NSEM. E. Representative NSEM micrograph. F. NSEM Particle picks in green. G. 2D class averages (the particles in the classes marked with a red box are recognizable as the pre-fusion spike). The rest of the classes are marked as "junk". H. NSEM Particles present in the boxed class averages in panel G. are shown with a green circle around them. I. Representative cryo-EM micrograph for furin cleaved S-RRAR-D614G. J. Cryo-EM CTF fit furin cleaved S-RRAR-D614G. K. Representative 2D class averages from Cryo-EM dataset for furin cleaved S-RRAR-D614G. L-N. Ab initio reconstructions for the cryo-EM for furin cleaved S-RRAR-D614G L. 1-up state (the RBD in the up position is identified by an asterisk), M. missing-RBD state (the RBD in the up position or missing is identified by an asterisk) and N. 3-down state. O-Q. Refined maps for the cryo-EM for furin cleaved S-RRAR-D614G O. 1-up state (the RBD in the up position is identified by an asterisk) P. missing-RBD state (the RBD in the up position or missing is identified by an asterisk) and Q. 3-down state. R-T. Fourier shell correlation curves for the cryo-EM furin cleaved S-RRAR-D614G R. 1-up state, S. missing-RBD state and T. 3-down state. U-V. Refined cryo-EM maps colored by local resolution for the consensus U. 3-RBD-down (EMDB-22823) and V. 1-RDB-up (EMD-22824) states. W. Zoom-in images showing the S2, SD2, D614G, S1', S2' and K986-V987 regions in the consensus 3-RBD-down (top) and 1-RDB-up structures (bottom). The cryo-EM map is shown as a transparent surface and the fitted model is in cartoon representation, with residues shown as balls and sticks (PDB ID 7KDJ and 7KDI).