Supporting Information Dataset 3

Development and validation of a potent and specific inhibitor for the CLC-2 chloride channel

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Dataset 3: Chemical Synthesis, General

All reagents were obtained commercially unless otherwise noted. Meclofenamate sodium, *N*-phenylanthranilic acid, and lubiprostone were purchased commercially from Sigma-Aldrich. Diclofenac sodium, indomethacin, and niflumic acid were purchased from Santa Cruz Biotechnology. Salsalate was purchased from ACROS Organics. Aceclofenac was purchased from AK Scientific. Organic solutions were concentrated under reduced pressure (~20 Torr) by rotary evaporation. Air- and moisture-sensitive liquids and solutions were transferred via syringe or stainless steel cannula. Chromatographic purification of the desired carboxylate, phosphonate, and sulfate inhibitors was accomplished using high performance liquid chromatography on a C18 column (Alltima C18, 10 μ M, 22 × 250 mm or SiliaChrom AQ C18, 5 μ M, 10 × 250 mm). Thin layer chromatography was performed on EM Science silica gel 60 F254 plates (250 mm). Visualization of the developed chromatogram was accomplished by fluorescence quenching and by staining with aqueous potassium permanganate or aqueous ceric ammonium molybdate (CAM) solution.

Nuclear magnetic resonance (NMR) spectra were acquired on a Varian Inova spectrometer operating at 300, 400, 500, or 600 MHz for ¹H spectra are referenced internally according to residual solvent signals. Data for ¹H NMR are recorded as follows: chemical shift (δ , ppm), multiplicity (s, singlet; d, doublet; t, triplet; q, quartet; quintet; m, multiplet; br, broad), coupling constant (Hz), integration. Compound concentrations were determined by quantitative NMR in DMSO-*d*₆ using *N*,*N*-dimethylformamide as an internal standard. Infrared spectra were recorded as thin films using NaCl plates on a Thermo-Nicolet 300 FT-IR spectrometer and are reported in frequency of absorption. Low-resolution mass spectra were obtained from the Vincent Coates Foundation Mass Spectrometry Laboratory and the Stanford ChEM-H facility, using a Shimadzu 20-20 ESI mass spectrometer and a Phenomenex Synergi 4 μ m Hydro-RP 80 Å reversed phase column (30 × 2 mm column, gradient flow 0:1 \rightarrow 1:0 MeCN/H₂O with 0.1% formic acid over 4 min). Microwave reactions were performed in a Biotage Initiator microwave reactor.

Dataset 3, Inhibitor Stock Solution Preparation and Quantification

Meclofenamic acid (MCFA) derivatives were quantified by ¹H NMR spectroscopy using distilled *N*,*N*dimethylformamide (DMF) as an internal standard. Use of DMF as the internal standard allows for recovery of pure material following lyophilization. Each MCFA derivative was weighed into an Eppendorf tube, using a calibrated analytical balance (Mettler Toledo, Model XS105), and dissolved into DMSO- d_6 to a final concentration of 30–100 mM. To ensure complete dissolution, the sealed Eppendorf tube was inverted at least 5 times and then sonicated for ~60 seconds. Using a calibrated analytical balance, DMF (22 mg) was weighed into a scintillation vial, 3.0 mL DMSO- d_6 (stored in a desiccator jar and preferably newly opened to minimize water contamination in the solvent) was added to the vial via a p1000 micropipette, and the solution thoroughly mixed by inverting the capped vial at least 10 times (note: to prevent crosscontamination between samples and for convenience, disposable micropipette tips may be used without affecting the accuracy of the measurements). To ensure robustness of the quantification protocol, the 100 mM DMF stock solution was independently quantified in triplicate against stock solutions with known concentrations of fumaronitrile dissolved in DMSO- d_6 . Fumaronitrile produces a sharp singlet that integrates to 2H at 7.03 ppm in DMSO- d_6 (DMSO solvent residual peak referenced to 2.50 ppm) and may be integrated against either of the DMF methyl signals that appear at 2.73 ppm (3H) and 2.89 (3H) ppm. The average of these three NMR measurements was used for calculation of the final DMF internal standard concentration. All measurements were performed at room temperature. Stock solutions were stored frozen, and left at ambient temperature for several hours to thaw and vortexed prior to quantification. A relaxation delay time (d1) of 20 s and an acquisition time (at) of 10 s were used during spectral acquisition. The number of scans (nt) was typically set to 32 unless a particular compound concentration was low (< 10 mM), such that more scans were required to improve signal/noise. The concentration of each MCFA derivative was determined by comparison with signal integrations for the MCFA derivative and the DMF internal standard.

Dataset 3: Experimental Protocols and Characterization Data

Meclofenamate derivatives. A general two-step protocol (Buchwald-Hartwig cross-coupling,¹ followed by ester hydrolysis) was used to transform commercially available anilines into meclofenamate derivatives. Additional transformations were required to obtain several derivatives for which the requisite starting materials are not commercially available. Experimental details are provided for these compounds.

Synthesis of C3-substituted 2,6-dichloronitrobenzenes

Methylation of 2,6-dichloro-3-nitrophenol



2,4-Dichloro-3-nitrophenol (0.5 g, 2.4 mmol) was dissolved in 5 mL of acetone, and the solution was cooled to 0 °C. Potassium hydroxide (146 mg, 2.6 mmol, 1.1 equiv) was added, followed by dropwise addition of dimethyl sulfate (656 mg, 5.2 mmol, 2.2 equiv). The mixture was warmed to room temperature and stirred for 3 h. The reaction was stirred for an additional 30 min at 60 °C until disappearance of the yellow color was observed. The reaction mixture was cooled to room temperature and diluted with 10 mL of EtOAc. The contents were transferred to a separatory funnel with an additional ~10 mL of EtOAc, and the combined filtrates were washed with 2 x 20 mL of H₂O and 1 x 20 mL of saturated aqueous NaCl. The organic fraction was dried over MgSO₄, filtered and concentrated under reduced pressure. The oily residue was redissolved in CH₂Cl₂ to which ~1 g of silica gel was then added. The suspension was concentrated under reduced pressure and the solid material dry loaded onto a silica gel column pre-packed in pentane. Purification of this material by chromatography on silica gel (gradient elution: 0:1 \rightarrow 1:19 Et₂O/pentane)

^{1.} Sadighi, J. P.; Harris, M. C.; Buchwald, S. L. A Highly Active Palladium Catalyst System for the Arylation of Anilines. *Tetrahedron Lett.* **1998**, 39 (30), 5327–5330.

furnished the desired product as a white solid (112 mg, 21%). TLC R_f = 0.3 (9:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 7.38 (d, *J* = 9.1 Hz, 1H), 6.98 (d, *J* = 9.1 Hz, 1H), 3.95 (s, 3H) ppm; IR (thin film) v 1539, 1480, 1454, 1438, 1362, 1300, 1291 cm⁻¹.

Triflation of 2,6-dichloro-3-nitrophenol

$$\begin{array}{c} CI \\ \hline \\ \hline \\ CI \\ CI \\ OH \end{array} \begin{array}{c} Tf_2O \\ \hline \\ Pyridine \\ CH_2CI_2 \end{array} \begin{array}{c} CI \\ \hline \\ CI \\ OTf \end{array}$$

To a 250-mL flame-dried flask under N₂ was added 2,4-dichloro-3-nitrophenol (1.0 g, 4.8 mmol), 48 mL of CH₂Cl₂, and freshly distilled pyridine (0.46 g, 5.8 mmol, 1.2 equiv). The flask was placed in an ice bath, and triflic anhydride (1.36 g, 4.8 mmol) was add dropwise via syringe. The solution was stirred at 0 °C for 15 min, then warmed to room temperature and stirred for an additional 40 min. Following this time, the reaction mixture was poured into a separatory funnel and the organic layer was washed successively with 1 x 50 mL of saturated aqueous NaHCO₃, 1 x 50 mL of H₂O, and 1 x 50 mL of saturated aqueous NaCl. The organic fraction was dried over Na₂SO₄, filtered and concentrated under reduced pressure. The oily residue was re-dissolved in CH₂Cl₂ to which ~1 g of silica gel was then added. The suspension was concentrated under reduced pressure and the solid material dry loaded onto a silica gel column pre-packed in hexanes. Purification of this material by chromatography on silica gel (gradient elution: 0:1→1:3 acetone/hexanes) furnished the desired product as a white solid (1.43 g, 88%). TLC R_f = 0.5 (9:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 7.57 (d, *J* = 9.0 Hz, 1H), 7.49 (d, *J* = 9.1 Hz, 1H) ppm; IR (thin film) v 1556, 1456, 1436, 1359, 1221, 1135 cm⁻¹.

Cross-coupling of aryl triflates and trifluoroborate salts



To a two-necked 25-mL flask was added 2,4-dichloro-3-nitrophenyl trifluoromethanesulfonate (300 mg, 0.88 mmol), potassium vinyltrifluoroborate (118 mg, 0.88 mmol), and Na₂CO₃ (281 mg, 2.6 mmol, 3.0 equiv). The flask was stoppered with a rubber septum and a reflux condenser, and flushed for ~5 min with argon. Dimethoxyethane (6.0 mL) was sparged with argon for 10 min, and then added via cannula to the flask containing the solid materials. To this mixture was added a solution of Pd(PPh₃)₄ (51 mg, 0.06 mmol, 0.05 equiv) in 2.5 mL of benzene under argon. The contents were stirred at 85 °C under argon for 6 h. Following this time, the reaction mixture was cooled to room temperature, diluted with 100 mL of Et₂O, and transferred to a separatory funnel. The organic phase was washed with 2 x 100 mL of H₂O and 1 x 100 mL of saturated aqueous NaCl, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The oily residue was re-dissolved in CH₂Cl₂ to which ~1 g of silica gel was then added. The suspension was concentrated under reduced pressure and the solid material dry loaded onto a silica gel column pre-packed

in hexanes. Purification of this material by chromatography on silica gel (gradient elution: $0:1 \rightarrow 1:19$ acetone/hexanes) furnished the desired product as a white solid (115 mg, 60%). TLC R_f = 0.33 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 7.62 (dd, *J* = 8.6, 0.6 Hz, 1H), 7.40 (dd, *J* = 8.6, 0.6 Hz, 1H), 7.01 (ddt, *J* = 17.5, 11.0, 0.6 Hz, 1H), 5.83 (dd, *J* = 17.4, 0.6 Hz, 1H), 5.57 (dd, *J* = 11.0, 0.6 Hz, 1H) ppm; IR (thin film) v 1536, 1456, 1361 cm⁻¹.



Prepared according to the above procedure using the appropriate tetrafluoroborate salt; colorless oil (30 mg, 44%). TLC R_f = 0.40 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 7.39 (d, *J* = 8.4 Hz, 1H), 7.28 (d, *J* = 8.4 Hz, 1H), 5.34 (q, *J* = 1.5 Hz, 1H), 5.03 (t, *J* = 1.1 Hz, 1H), 2.08 (s, 3H) ppm; IR (thin film) v 1548, 1460, 1362 cm⁻¹.



Prepared according to the above procedure using the appropriate tetrafluoroborate salt; white solid (78 mg, 33%). TLC R_f = 0.36 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.51–7.45 (m, 4H), 7.43 (d, *J* = 8.4 Hz, 1H), 7.41–7.38 (m, 2H) ppm; IR (thin film) v 3089, 3063, 3031, 2892, 1539, 1447, 1456, 1359, 1253 cm⁻¹.



Prepared according to the above procedure using the appropriate tetrafluoroborate salt; colorless film (99 mg, 20%). TLC R_f = 0.24 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.45 (d, *J* = 8.4 Hz, 1H), 7.44 (td, *J* = 7.5, 1.8 Hz, 1H), 7.37 (d, *J* = 8.4 Hz, 1H), 7.16 (dd, *J* = 7.5, 1.8 Hz, 1H), 7.05 (td, *J* = 7.5, 1.0 Hz, 1H), 7.00 (d, *J* = 8.4 Hz, 1H), 3.79 (s, 3H) ppm; IR (thin film) v 2934, 2838, 1583, 1548, 1498, 1464, 1436, 1360, 1275, 1241 cm⁻¹.

Cross-coupling of aryl triflates and vinyl boronic acids



The reaction was performed following a general protocol described by Littke, et al.² To a 5-mL oven-dried round-bottom flask was added 2,4-dichloro-3-nitrophenyl trifluoromethanesulfonate (300 mg, 0.88 mmol), cyclohex-1-en-1-yl boronic acid (122 mg, 0.97 mmol, 1.1 equiv), anhydrous potassium fluoride (169 mg, 2.9 mmol, 3.3 equiv), and Pd(OAc)₂ (9.9 mg, 0.044 mmol, 0.05 equiv). The flask was stoppered with a rubber septum and flushed with argon for 5 min. In a glovebox, tricyclohexylphosphine (15 mg, 0.053 mmol, 0.06 equiv) was transferred to a round-bottom flask and sealed with a new rubber septum. The flask was removed from the glovebox, and 2.0 mL of dry, argon-purged THF was added. The phosphine solution was sparged with argon for 5 min and then added via cannula to the flask containing the starting materials and Pd catalyst. The reaction was stirred under argon for 8 h. Following this time, the reaction mixture was diluted with 5 mL of Et₂O and filtered through a plug of silica gel. The flask and filter cake were rinsed with 3 x 5 mL of Et₂O. The combined filtrates were concentrated under reduced pressure to an oily residue. Purification of this material by chromatography on silica gel (gradient elution: $0:1 \rightarrow 1:19$ EtOAc/hexanes) furnished the desired product as a white solid (136 mg, 57%). TLC $R_f = 0.53$ (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 500 MHz) δ 7.35 (d, J = 8.3 Hz, 1H), 7.23 (d, J = 8.3 Hz, 1H), 5.73 (tt, J = 3.7, 1.8 Hz, 1H), 2.25–2.22 (m, 2H), 2.21–2.16 (m, 2H), 1.79–1.72 (m, 2H), 1.71–1.66 (m, 2H) ppm; IR (thin film) v 2933, 2859, 2836, 1549, 1456, 1362, 1256, 1188, 1137 cm⁻¹.

Alkylation of 2,6-dichloro-3-nitrophenol



To an ice-cold suspension of sodium hydride (35 mg, 1.44 mmol, 1.2 equiv) in 1.0 mL of anhydrous DMF was added via syringe a solution of 2,4-dichloro-3-nitrophenol (250 mg, 1.20 mmol) in 1.0 mL DMF. After evolution of H₂ ceased (~30 min), neat benzyl bromide (206 mg, 1.20 mmol) was added via syringe. The reaction mixture was warmed to room temperature and stirred for 1.5 h. During this time, the color of the reaction changed from bright red-orange to pale yellow (Note: reaction times vary according to the reactivity of the electrophile and require up to 24 h at ambient temperature until the yellow color is largely

^{2.} Littke, A. F.; Dai, C.; Fu, G. C. Versatile Catalysts for the Suzuki Cross-Coupling of Arylboronic Acids with Aryl and Vinyl Halides and Triflates under Mild Conditions. *J. Am. Chem. Soc.* **2000**, *1*22, 4020–4028.

extinguished. Certain reactions, particularly those using mesylate-base electrophiles, require elevated temperatures, as indicated below). Following completion, the contents were transferred to a separatory funnel with 50 mL of Et₂O. The organic layer was washed with 3 x 50 mL of H₂O and 2 x 50 mL of 1.0 M aqueous NaOH, dried over Na₂SO₄, filtered and concentrated under reduced pressure to a white solid. Purification of this material by recrystallization from ~25 mL of hot/cold hexanes afforded the desired product as white crystals (146 mg, 41%). TLC R_f = 0.40 (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.46–7.34 (m, 5H), 7.32 (d, *J* = 9.0 Hz, 1H), 7.00 (d, *J* = 9.0 Hz, 1H), 5.21 (s, 2H) ppm; IR (thin film) v 1541, 1479, 1449, 1364, 1298 cm⁻¹.



Prepared according to the above procedure with 2,4-dichloro-3-nitrophenol (250 mg, 1.20 mmol) and freshly prepared cyclohexylmethyl methanesulfonate (231 mg, 1.20 mmol). After stirring for 1.5 h at room temperature, the reaction flask was heated to 120 °C in an oil bath and stirred for an additional 4 h. Product obtained as an off-white solid (141 mg, 39%). TLC R_f = 0.56 (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.33 (d, *J* = 9.0 Hz, 1H), 6.94 (d, *J* = 9.1 Hz, 1H), 3.84 (d, *J* = 5.9 Hz, 2H), 1.91–1.81 (m, 3H), 1.81–1.75 (m, 2H), 1.75–1.68 (m, 1H), 1.36–1.26 (m, 2H), 1.21 (tt, *J* = 12.7, 3.2 Hz, 1H), 1.13–1.03 (m, 2H) ppm; IR (thin film) v 2925, 2852, 1544, 1478, 1459, 1360, 1293, 1267 cm⁻¹.

Prepared according to the above procedure with 2,4-dichloro-3-nitrophenol (1.5 g, 7.21 mmol) and 1,2dibromoethane (208 mg, 8.65 mmol, 1.2 equiv). Following the addition of 1,2-dibromoethane, the mixture was stirred at 110 °C for 6 h. Product was obtained as a white solid (662 mg, 32%). TLC R_f = 0.20 (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 7.38 (d, *J* = 9.0 Hz, 1H), 6.99 (d, *J* = 9.0 Hz, 1H), 4.39 (t, *J* = 6.2 Hz, 2H), 3.69 (t, *J* = 6.2 Hz, 2H) ppm; IR (thin film) v 1537, 1364, 1296 cm⁻¹.



Prepared according to the above procedure with 2,4-dichloro-3-nitrophenol (250 mg, 1.2 mmol) and 2-fluorobenzyl bromide (227 mg, 1.2 mmol). The product was isolated as a white solid (294 mg, 77%). TLC $R_f = 0.22$ (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.51 (td, J = 7.5, 1.7 Hz, 1H), 7.39–7.34 (m,

2H), 7.20 (td, *J* = 7.6, 1.1 Hz, 1H), 7.11 (ddd, *J* = 9.7, 8.2, 1.1 Hz, 1H), 7.06 (d, *J* = 9.1 Hz, 1H), 5.26 (s, 2H) ppm; IR (thin film) v 3093, 2911, 1585, 1542, 1492, 1478, 1455, 1364, 1303, 1274, 1234 cm⁻¹.



Prepared according to the above procedure with 2,4-dichloro-3-nitrophenol (250 mg, 1.2 mmol) and 2methylbenzyl bromide (266 mg, 1.44 mmol, 1.2 equiv). The product was isolated as an off-white solid (293 mg, 78%). TLC R_f = 0.18 (9:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 7.38 (d, *J* = 7.2 Hz, 1H), 7.35 (dd, *J* = 9.0, 1.1 Hz, 1H), 7.32–7.27 (m, 1H), 7.25–7.21 (m, 2H), 7.05 (d, *J* = 9.0 Hz, 1H), 5.16 (s, 2H), 2.39 (s, 3H) ppm; IR (thin film) v 2919, 1592, 1542, 1495, 1479, 1451, 1364, 1300, 1285, 1272, 1194, 1100 cm⁻¹.



Prepared according to the above procedure with 2,4-dichloro-3-nitrophenol (250 mg, 1.2 mmol) and 3methylbenzyl bromide (222 mg, 1.2 mmol). The product was isolated as a white solid (239 mg, 64%). TLC $R_f = 0.26$ (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.31 (d, J = 9.1, 1H), 7.29 (t, J = 7.4, 1H), 7.23–7.16 (m, 3H), 7.00 (d, J = 9.1 Hz, 1H), 5.17 (s, 2H), 2.38 (s, 3H) ppm; IR (thin film) v 3101, 3043, 2916, 1592, 1539, 1479, 1450, 1363, 1302, 1273, 1100 cm⁻¹.



Prepared according to the above procedure with 2,4-dichloro-3-nitrophenol (312 mg, 1.5 mmol) and 4methylbenzyl bromide (333 mg, 1.8 mmol, 1.2 equiv). The product was isolated as a white solid (342 mg, 73%). TLC R_f = 0.24 (9:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 7.34–7.28 (m, 3H), 7.21 (d, *J* = 7.8 Hz, 2H), 7.00 (d, *J* = 9.2 Hz, 1H), 5.16 (s, 2H), 2.37 (s, 3H) ppm; IR (thin film) v 3094, 2908, 2867, 1866, 1594, 1537, 1482, 1455, 1363, 1312, 1299, 1274 cm⁻¹.



Prepared according to the above procedure with 2,4-dichloro-3-nitrophenol (312 mg, 1.5 mmol) and freshly prepared (perfluorophenyl)methyl methanesulfonate (425 mg, 1.54 mmol, 1.03 equiv). The product was isolated as a pale yellow solid (425 mg, 73%). TLC R_f = 0.30 (4:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 7.43 (d, *J* = 9.0 Hz, 1H), 7.14 (d, *J* = 9.0 Hz, 1H), 5.22 (s, 2H) ppm; ¹⁹F NMR (CDCl₃, 376 MHz) δ –142.18 (dd, *J* = 22.1, 8.2 Hz, 2F), –151.06 (t, *J* = 20.8 Hz, 1F), –160.92 (td, *J* = 20.6, 7.0 Hz, 2F) ppm; IR (thin film) v 1658, 1590, 1549, 1525, 1508, 1465, 1435, 1389, 1362, 1313, 1288, 1264, 1195, 1135, 1104 cm⁻¹.



Prepared according to the above procedure with 2,4-dichloro-3-nitrophenol (250 mg, 1.2 mmol) and (2-bromoethyl)benzene (266 mg, 1.44 mmol, 1.2 equiv). The product was isolated as a white solid (169 mg, 45%). TLC R_f = 0.16 (9:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.37–7.25 (m, 6H), 6.92 (d, *J* = 9.1 Hz, 1H), 4.24 (t, *J* = 6.8 Hz, 2H), 3.17 (t, *J* = 6.7 Hz, 2H) ppm; IR (thin film) v 3084, 3031, 2955, 2884, 1889, 1603, 1584, 1541, 1496, 1478, 1457, 1431, 1389, 1367, 1293, 1269, 1193, 1157 cm⁻¹.



Prepared according to the above procedure with 2,4-dichloro-3-nitrophenol (1.24 g, 6.0 mmol) and methyl 2-(bromomethyl)benzoate (1.50 g, 6.6 mmol, 1.1 equiv). The product was isolated as a white solid (1.71 g, 81%). TLC R_f = 0.5 (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 8.08 (dd, *J* = 7.8, 1.3 Hz, 1H), 7.80 (d, *J* = 7.9 Hz, 1H), 7.62 (td, *J* = 7.8, 1.1 Hz 1H), 7.44 (t, *J* = 7.8 Hz, 1H), 7.35 (dd, *J* = 9.0, 0.7 Hz, 1H), 7.09 (d, *J* = 9.0, 1H), 5.63 (s, 2H), 3.92 (s, 3H) ppm; IR (thin film) v 1722, 1543, 1475, 1432, 1365, 1302, 1267, 1143 cm⁻¹.



Prepared according to the above procedure with 2,4-dichloro-3-nitrophenol (1.24 g, 6.0 mmol) and methyl 3-(bromomethyl)benzoate (1.50 g, 6.6 mmol, 1.1 equiv). The product was isolated as an off-white solid (1.68 g, 79%). TLC R_f = 0.39 (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 8.09 (s, 1H), 8.04 (d, *J* = 7.8 Hz, 1H), 7.65 (d, *J* = 7.7 Hz, 1H), 7.51 (t, *J* = 7.7 Hz, 1H), 7.34 (d, *J* = 9.0 Hz, 1H), 7.01 (d, *J* = 9.0 Hz, 1H), 5.23 (s, 2H), 3.94 (s, 3H) ppm; IR (thin film) v 1717, 1544, 1476, 1362, 1306, 1102 cm⁻¹.



Prepared according to the above procedure with 2,4-dichloro-3-nitrophenol (1.24 g, 6.0 mmol) and methyl 4-(bromomethyl)benzoate (1.50 g, 6.6 mmol, 1.1 equiv). The product was isolated as a white solid (1.75 g, 83%). TLC R_f = 0.32 (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 8.08 (d, *J* = 8.0 Hz, 2H), 7.50 (d, *J* = 8.0 Hz, 2H), 7.33 (d, *J* = 9.0 Hz, 1H), 6.98 (d, *J* = 9.0 Hz, 1H), 5.25 (s, 2H), 3.93 (s, 3H) ppm; IR (thin film) v 1705, 1551, 1480, 1449, 1309, 1287, 1120 cm⁻¹.



Prepared according to the above procedure with 2,4-dichloro-3-nitrophenol (194 mg, 0.93 mmol) and freshly prepared 3-acetamidobenzyl methanesulfonate (250 mg, 1.20 mmol, 1.3 equiv). The product was isolated as a tan solid (270 mg, 82%). TLC R_f = 0.43 (19:1 CH₂Cl₂/MeOH); ¹H NMR (acetone-d₆, 500 MHz) δ 9.27 (s, 1H), 7.84 (d, *J* = 2.8 Hz, 1H), 7.66–7.56 (m, 2H), 7.47 (d, *J* = 9.0 Hz, 1H), 7.35–7.27 (m, 1H), 7.20 (d, *J* = 7.6 Hz, 1H), 5.33 (s, 2H), 2.08 (s, 3H) ppm; IR (thin film) v 3303, 3088, 1669, 1615, 1596, 1545, 1476, 1364, 1295, 1267, 1196 cm⁻¹.

Phthalimide synthesis



To a solution of 1-(2-bromoethoxy)-2,4-dichloro-3-nitrobenzene (250 mg, 0.79 mmol) in 1.6 mL of DMF was added potassium phthalimide (176 mg, 0.95 mmol, 1.2 equiv). The reaction was stirred at 80 °C for 7.5 h. Following this time, the reaction mixture was transferred to a separatory funnel with 100 mL of EtOAc. The organic layer was washed with 1 x 100 mL of H₂O, 2 x 100 mL of saturated aqueous Na₂CO₃, and 2 x 100 mL of saturated aqueous NaCl (2 x 100 mL), dried over Na₂SO₄, filtered and concentrated under reduced pressure to a white solid. Purification of this material by recrystallization from ~10 mL of hot/cold acetone afforded the desired product as a white solid (173 mg, 57%). TLC R_f = 0.44 (3:2 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.88 (dd, *J* = 5.4, 3.0 Hz, 2H), 7.75 (dd, *J* = 5.5, 3.0 Hz, 2H), 7.34 (d, *J* = 9.1 Hz, 1H), 6.98 (d, *J* = 9.1 Hz, 1H), 4.33 (t, *J* = 5.7 Hz, 2H), 4.18 (t, *J* = 5.7 Hz, 2H) ppm; IR (thin film) v 1714, 1544, 1467, 1394, 1365, 1295, 1268 cm⁻¹.

Ester hydrolysis



A screw-capped vial was charged with methyl 2-((2,4-dichloro-3-nitrophenoxy)methyl)benzoate (500 mg, 1.40 mmol) and 20 mL of a 1.0 M methanolic solution of KOH. The mixture was stirred for 24 h then warmed to 50 °C and stirred for an additional 2 h. During this time the opaque mixture transitioned to a clear yellow solution. The solution was cooled to ambient temperature and acidified to pH 1 by dropwise addition of 5 mL of concentrated aqueous HCI. Acidification of the reaction mixture resulted in precipitation of a white solid. Following the addition of 50 mL of H₂O, the desired product was isolated by vacuum filtration through a Büchner funnel as a white solid (405 mg, 84%). TLC R_f = 0.28 (19:1 CH₂Cl₂/MeOH); ¹H NMR (acetone-d₆, 400 MHz) δ 8.12 (dd, *J* = 7.8, 1.5 Hz, 1H), 7.81 (d, *J* = 7.8, 1H), 7.73–7.63 (m, 2H), 7.54 (t, *J* = 7.5, 1H), 7.47 (d, *J* = 9.0, 1H), 5.74 (s, 2H) ppm; IR (thin film) v 2851, 1686, 1541, 1475, 1415, 1363, 1298, 1271, 1198 cm⁻¹.



Prepared according to the above procedure; white solid (436 mg, 91%). TLC $R_f = 0.28$ (19:1 CH₂Cl₂/MeOH); ¹H NMR (acetone-d₆, 400 MHz) δ 8.20 (s, 1H), 8.04 (d, *J* = 7.8 Hz, 1H), 7.82 (d, *J* = 7.8 Hz, 1H), 7.69 (d, *J* = 9.2 Hz, 1H), 7.62–7.49 (m, 2H), 5.48 (s, 2H) ppm; IR (thin film) v 2824, 1567, 1680, 1544, 1480, 1365, 1297, 1210 cm⁻¹.



Prepared according to the above procedure; white solid (410 mg, 86%). TLC $R_f = 0.28$ (19:1 CH₂Cl₂/MeOH); ¹H NMR (acetone-d₆, 400 MHz) δ 8.10 (d, J = 8.4 Hz, 2H), 7.68 (d, J = 9.2 Hz, 3H), 7.53 (d, J = 9.2 Hz, 1H), 5.50 (s, 2H) ppm; IR (thin film) v 2900, 2551, 1687, 1542, 1478, 1447, 1426, 1363, 1297, 1263 cm⁻¹.

MOM protection of 2,6-dichloro-3-nitrophenol



To an ice-cold suspension of NaH (70 mg, 2.92 mmol, 1.2 equiv) in 3.2 mL of anhydrous Et₂O was added 2,4-dichloro-3-nitrophenol (505 mg, 2.43 mmol) in 1.0 mL of DMF under N₂. After evolution of H₂ ceased (approximately 30 min), chloromethyl methyl ether (196 mg, 1.0 mmol) was added dropwise via syringe. The reaction was warmed to ambient temperature and stirred for 1 h. Following this time, the reaction mixture was quenched by the addition of 1 mL of H₂O and transferred to a separatory funnel with 50 mL of Et₂O. The organic layer was washed with 2 x 50 mL of H₂O and 1 x 50 mL of saturated aqueous NaCl, dried over MgSO₄, filtered and concentrated under reduce pressure to a white solid (588 mg, 95%). This material was judged to be sufficiently pure by ¹H NMR for use in the subsequent step without further purification. TLC R_f = 0.25 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 300 MHz) 7.35 (d, *J* = 9.1 Hz, 1H) 7.26 (d, *J* = 9.1 Hz, 1H) 5.28 (s, 2H) 3.51 (s, 3H) ppm; IR (thin film) v 3092, 2971, 2948, 2909, 1589, 1545, 1465, 1367, 1265, 1152 cm⁻¹.



AK-36. The reaction was performed following a general protocol described by Suzuki, et al.³ To a suspension of meclofenamic sodium acetate (75 mg, 0.24 mmol), Et₃N (0.33 mL, 2.4 mmol, 10 equiv), and NH₄Cl (126 mg, 2.4 mmol, 10 equiv) in 2.0 mL of THF were added successively 1-ethyl-3(3-dimethylamino)carbodiimide (131 mg, 0.68 mmol, 2.9 equiv) and 1-hydroxybenzotriazole monohydrate (112 mg, 0.73 mmol, 3.1 equiv). The reaction mixture was stirred for 23 h, then poured into a separatory funnel containing 50 mL of H₂O. The aqueous layer was extracted with 1 x 50 mL of EtOAc. The organic fraction was washed with saturated aqueous 1 x 50 mL of NaHCO₃ and 1 x 50 mL saturated aqueous NaCl, dried over Na₂SO₄, filtered and concentrated under reduced pressure to a white solid. Purification of this material by chromatography on silica gel (1:1 hexanes/EtOAc) furnished the desired product as a white solid (53 mg, 76%). TLC R_f = 0.50 (1:1 hexanes/EtOAc); ¹H NMR (DMSO-d₆, 600 MHz) δ 10.09 (s, 1H), 8.09 (s, 1H), 7.72 (dd, *J* = 7.9, 1.5 Hz, 1H), 7.48 (d, *J* = 8.3 Hz, 2H), 7.30 (d, *J* = 8.5 Hz, 1H), 7.22 (td, *J* = 8.5, 1.5 Hz, 1H), 6.75 (td, *J* = 7.9, 1.1 Hz, 1H), 6.18 (dd, *J* = 8.4, 1.0 Hz, 1H), 2.37 (s, 3H) ppm; IR (thin film) v 3482, 3191, 1660, 1616, 1581, 1506, 1449, 1392, 1287 cm⁻¹; LRMS (ES⁺) calcd 295.03 for C₁₄H₁₂Cl₂N₂O [M+H]⁺ found 294.7 (M⁺).



Prepared according to the above procedure from the corresponding carboxylic acid (See *Synthesis of C3-substituted 2,6-dichloronitrobenzenes* for carboxylic acid synthesis); off-white solid (187 mg, 63%). TLC R_f = 0.37 (19:1 CH₂Cl₂/MeOH); ¹H NMR (acetone-d₆, 400 MHz) δ 7.73–7.68 (m, 2H), 7.66 (d, *J* = 9.2 Hz, 1H), 7.57–7.49 (m, 2H), 7.46–7.42 (m, 2H), 6.84 (s, 1H), 5.62 (s, 2H) ppm; IR (thin film) v 3467, 1679, 1540, 1470, 1361, 1278 cm⁻¹.

Suzuki, T.; Imai, K.; Nakagawa, H.; Miyata, N. 2-Anilinobenzamides as SIRT Inhibitors. *ChemMedChem* 2006, 1, 1059–1062.



Prepared according to the above procedure from the corresponding carboxylic acid (See *Synthesis of C3-substituted 2,6-dichloronitrobenzenes* for carboxylic acid synthesis); off-white solid (223 mg, 74%). TLC R_f = 0.33 (19:1 CH₂Cl₂/MeOH); ¹H NMR (acetone-d₆, 400 MHz) δ 8.10 (s, 1H), 7.94 (d, *J* = 7.9 Hz, 1H), 7.75–7.64 (m, 2H), 7.55–7.51 (m, 3H), 6.70 (s, 1H), 5.44 (s, 2H) ppm; IR (thin film) v 3357, 3182, 1654, 1539, 1500, 1476, 1407, 1364, 1301, 1199 cm⁻¹.



Prepared according to the above procedure from the corresponding carboxylic acid (See *Synthesis of C3-substituted 2,6-dichloronitrobenzenes* for carboxylic acid synthesis); off-white solid (222 mg, 74%). TLC R_f = 0.33 (19:1 CH₂Cl₂/MeOH); ¹H NMR (acetone-d₆, 400 MHz) δ 7.99 (d, *J* = 8.2 Hz, 2H), 7.67 (d, *J* = 8.9 Hz, 1H), 7.63 (d, *J* = 8.2 Hz, 2H), 7.50 (d, *J* = 9.2 Hz, 2H), 6.68 (s, 1H), 5.46 (s, 2H) ppm; IR (thin film) v 3405, 1672, 1615, 1544, 1476, 1448, 1363, 1291, 1263, 1197 cm⁻¹.

Synthesis of substituted anilines



To a solution of 1,3-dichloro-4-methoxy-2-nitrobenzene (60 mg, 0.27 mmol) in 2.7 mL of EtOH was added Fe powder (106 mg, 1.89 mmol, 7.0 equiv), anhydrous FeCl₃ (6.7 mg, 0.04 mmol, 0.15 equiv), and glacial acetic acid (0.1 mL, 1.70 mmol, 6.3 equiv). The reaction flask was sealed and the contents stirred at 70 °C for 2 h. Following this time, the solution was cooled to room temperature, diluted with 10 mL of EtOAc, and filtered through a pad of Celite. The flask and filter cake were washed with EtOAc and the combined filtrates were concentrated under reduced pressure. The oily residue was re-dissolved in CH₂Cl₂ to which ~0.5 g of silica gel was then added. The suspension was concentrated under reduced pressure and the solid material dry loaded onto a silica gel column pre-packed in hexanes. Purification of this material by chromatography on silica gel (gradient elution: 0:1 \rightarrow 1:3 EtOAc/hexanes) furnished the desired product as a white solid (36 mg, 70%). TLC R_f = 0.27 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 7.12 (d, *J* = 8.9 Hz, 1H), 6.30 (d, *J* = 8.9 Hz, 1H), 3.86 (s, 3H) ppm; IR (thin film) v 3485, 3387, 1610, 1475, 1306, 1247, 1121 cm⁻¹.



Prepared according to the above procedure; white solid (475 mg, 92%). TLC $R_f = 0.54$ (9:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 300 MHz) δ 7.15 (d, J = 8.5 Hz, 1H), 7.02 (dd, J = 17.4, 10.7 Hz, 1H), 6.91 (d, J = 8.4 Hz, 1H), 5.71 (d, J = 17.4 Hz, 1H), 5.37 (d, J = 10.7 Hz, 1H), 4.49 (br s, 2H) ppm; IR (thin film) v 3488, 3391, 1603, 1471, 1401 cm⁻¹.



Prepared according to the above procedure; colorless oil (20 mg, 65%). TLC R_f = 0.59 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 500 MHz) δ 7.13 (d, *J* = 8.2 Hz, 1H), 6.54 (d, *J* = 8.2 Hz, 1H), 5.20 (quintet, *J* = 1.6 Hz, 1H), 4.94 (dq, *J* = 1.9, 1.0 Hz, 1H), 4.52 (s, 2H), 2.06 (s, 3H) ppm; IR (thin film) v 3490, 3393, 2920, 1604, 1468, 1422 cm⁻¹.



Prepared according to the above procedure; colorless oil (65 mg, 94%). TLC $R_f = 0.50$ (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.46–7.36 (m, 5H), 7.23 (d, J = 8.2 Hz, 1H), 6.68 (d, J = 8.3 Hz, 1H), 4.60 (s, 2H) ppm; IR (thin film) v 3488, 3390, 3060, 3031, 1603, 1579, 1548, 1464, 1419 cm⁻¹.



Prepared according to the above procedure; colorless oil (74 mg, 83%). TLC R_f = 0.37 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.38 (td, *J* = 7.5, 1.8 Hz, 1H), 7.22 (d, *J* = 8.2 Hz, 1H), 7.16 (dd, *J* = 7.5, 1.8 Hz, 1H), 7.02 (td, *J* = 7.4, 1.1 Hz, 1H), 6.98 (dd, *J* = 8.3, 1.0 Hz, 1H), 6.64 (d, *J* = 8.2 Hz, 1H), 4.54 (s, 2H), 3.79 (s, 3H) ppm; IR (thin film) v 3486, 3387, 2937, 2835, 1603, 1582, 1548, 1497, 1467, 1435, 1420, 1280, 1254, 1239 cm⁻¹.



Prepared according to the above procedure; pale yellow film (38 mg, 85%). TLC R_f = 0.69 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.10 (d, *J* = 8.2 Hz, 1H), 6.50 (d, *J* = 8.2 Hz, 1H), 5.64–5.62 (m, 1H), 4.48 (s, 2H), 2.25–2.21 (m, 2H), 2.19–2.14 (m, 2H), 1.79–1.71 (m, 2H), 1.69–1.65 (m, 2H) ppm; IR (thin film) v 3490, 3392, 2931, 2857, 2835, 1603, 1547, 1467, 1421 cm⁻¹.

Prepared according to the above procedure; white solid (111 mg, 85%). TLC R_f = 0.39 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 7.44 (d, *J* = 7.5 Hz, 2H), 7.39 (t, *J* = 6.8 Hz, 2H), 7.32 (t, *J* = 7.1 Hz, 1H), 7.08 (d, *J* = 8.9 Hz, 1H), 6.34 (d, *J* = 8.9 Hz, 1H), 5.12 (s, 2H), 4.49 (s, 2H) ppm; IR (thin film) v 3302, 1615, 1476, 1452, 1303, 1248 cm⁻¹.



Prepared according to the above procedure; off-white solid (104 mg, 83%). TLC $R_f = 0.45$ (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 7.08 (d, J = 8.8 Hz, 1H), 6.27 (d, J = 8.9 Hz, 1H), 4.45 (s, 2H), 3.77 (d, J = 6.1 Hz, 2H), 1.92–1.82 (m, 3H), 1.81–1.66 (m, 4H), 1.36–1.17 (m, 2H), 1.08 (m, 2H) ppm; IR (thin film) v 3332, 2921, 2857, 1614, 1478, 1449, 1309, 1118 cm⁻¹.



Prepared according to the above procedure; white solid (107 mg, 79%). TLC $R_f = 0.54$ (9:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.11 (d, J = 8.9 Hz, 1H), 6.29 (d, J = 8.9 Hz, 1H), 4.40 (s, 2H), 4.30 (t, J = 6.5 Hz, 2H), 3.66 (t, J = 6.5 Hz, 2H) ppm; IR (thin film) v 3438, 3330, 2942, 2882, 1615, 1476, 1462, 1306, 1246 cm⁻¹.



Prepared according to the above procedure; white solid (301 mg, 85%). TLC $R_f = 0.39$ (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 7.10 (d, J = 8.9 Hz, 1H), 6.54 (d, J = 8.9 Hz, 1H), 5.21 (s, 2H), 4.48 (s, 2H), 3.50 (s, 3H) ppm; IR (thin film) v 3438, 3391, 3317, 2961, 2833, 1615, 1476, 1446, 1160 cm⁻¹.



Prepared according to the above procedure; white solid (225 mg, 98%). TLC $R_f = 0.30$ (19:1 hexanes/Et₂O); ¹H NMR (CDCl₃, 400 MHz) δ 7.57 (td, J = 7.5, 0.9 Hz, 1H), 7.35–7.28 (m, 1H), 7.17 (t, J = 7.5 Hz, 1H), 7.13–7.04 (m, 2H), 6.38 (d, J = 8.9 Hz, 1H), 5.18 (s, 2H), 4.51 (s, 2H) ppm; ¹⁹F NMR (CDCl₃, 376 MHz) δ –119.19 to –119.31 (m, 1F) ppm; IR (thin film) v 3293, 1616, 1587, 1493, 1477, 1451, 1388, 1313, 1302, 1230, 1114 cm⁻¹.



Prepared according to the above procedure; white solid (192 mg, 97%). TLC R_f = 0.30 (19:1 hexanes/ Et₂O); ¹H NMR (CDCl₃, 400 MHz) δ 7.45 (d, *J* = 7.3 Hz, 1H), 7.29–7.17 (m, 3H), 7.11 (d, *J* = 8.9 Hz, 1H), 6.38 (d, *J* = 8.9 Hz, 1H), 5.07 (s, 2H), 2.38 (s, 3H) ppm; IR (thin film) v 3482, 3384, 2926, 1608, 1450, 1376, 1304, 1247, 1109 cm⁻¹.



Prepared according to the above procedure; white solid (186 mg, 94%). TLC R_f = 0.32 (19:1 hexanes/ Et₂O); ¹H NMR (CDCl₃, 300 MHz) δ 7.29–7.20 (m, 3H), 7.13 (d, *J* = 7.2 Hz, 1H), 7.07 (d, *J* = 8.9 Hz, 1H), 6.34 (d, *J* = 8.9 Hz, 1H), 5.08 (s, 2H), 2.37 (s, 3H) ppm; IR (thin film) v 3488, 3389, 3027, 2920, 1610, 1567, 1475, 1377, 1307, 1244, 1168, 1117 cm⁻¹.



Prepared according to the above procedure; white solid (192 mg, 97%). TLC R_f = 0.30 (19:1 hexanes/ Et₂O); ¹H NMR (CDCl₃, 400 MHz) δ 7.33 (d, *J* = 8.0 Hz, 2H), 7.19 (d, *J* = 8.3 Hz, 2H), 7.07 (d, *J* = 8.9 Hz, 1H), 6.34 (d, *J* = 8.9 Hz, 1H), 5.08 (s, 2H), 2.36 (s, 3H) ppm; IR (thin film) v 3441, 3319, 2920, 1614, 1586, 1519, 1477, 1451, 1380, 1302, 1247, 1166, 1115 cm⁻¹.



Prepared according to the above procedure; white solid (180 mg, 72%). TLC R_f = 0.27 (19:1 hexanes/ Et₂O); ¹H NMR (CDCl₃, 400 MHz) δ 7.14 (d, *J* = 8.8 Hz, 1H), 6.43 (d, *J* = 8.9 Hz, 1H), 5.14 (s, 2H) ppm; ¹⁹F NMR (CDCl₃, 376 MHz) δ –142.32 (dd, *J* = 22.2, 8.6 Hz, 2F), –152.59 (t, *J* = 20.8 Hz, 1F), –161.74 (td, *J* = 21.1, 7.5 Hz, 2F) ppm. IR (thin film) v 3442, 3338, 1659, 1613, 1525, 1510, 1479, 1468, 1452, 1389, 1300, 1248, 1135 cm⁻¹.



Prepared according to the above procedure; colorless residue that slowly solidified (146 mg, 99%). TLC R_f = 0.30 (19:1 hexanes/Et₂O); ¹H NMR (CDCl₃, 400 MHz) δ 7.36–7.30 (m, 4H), 7.29–7.23 (m, 1H), 7.08 (d, J = 8.9 Hz, 1H), 6.27 (d, J = 8.9 Hz, 1H), 4.49 (s, 2H), 4.18 (t, J = 7.0 Hz, 2H), 3.15 (t, J = 7.0 Hz, 2H) ppm; IR (thin film) v 3487, 3388, 2928, 1610, 1497, 1476, 1465, 1386, 1307, 1247, 1168, 1117 cm⁻¹.



Prepared according to the above procedure; off-white solid (212 mg, 77%). TLC R_f = 0.37 (9:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 8.04 (dd, *J* = 7.8, 1.4 Hz, 1H), 7.87 (dd, *J* = 8.0, 1.1 Hz, 1H), 7.59 (td, *J* = 7.7, 1.4 Hz, 1H), 7.39 (t, *J* = 7.4 Hz, 1H), 7.09 (d, *J* = 8.9 Hz, 1H), 6.39 (d, *J* = 8.9 Hz, 1H),

5.54 (s, 2H), 4.52 (s, 2H), 3.91 (s, 3H) ppm; IR (thin film) v 3432, 3320, 2942, 1719, 1616, 1474, 1313, 1262, 1140 cm⁻¹.

Prepared according to the above procedure; off-white solid (243 mg, 89%). TLC $R_f = 0.20$ (9:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 8.10 (s, 1H), 8.00 (d, *J* = 8.0 Hz, 1H), 7.68 (d, *J* = 7.6 Hz, 1H), 7.47 (t, *J* = 7.6 Hz, 1H), 7.08 (d, *J* = 8.8 Hz, 1H), 6.32 (d, *J* = 8.8 Hz, 1H), 5.15 (s, 2H), 4.52 (s, 2H), 3.93 (s, 3H) ppm; IR (thin film) v 3484, 3381, 2951, 1720, 1611, 1475, 1308, 1289, 1248, 1205, 1110 cm⁻¹.



Prepared according to the above procedure; off-white solid (228 mg, 83%). TLC $R_f = 0.27$ (9:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 8.06 (d, J = 7.8 Hz, 2H), 7.52 (d, J = 7.8 Hz, 2H), 7.08 (d, J = 8.9 Hz, 1H), 6.30 (d, J = 8.9 Hz, 1H), 5.17 (s, 2H), 4.53 (s, 2H), 3.92 (s, 3H) ppm; IR (thin film) v 3471, 3381, 2952, 1716, 1614, 1475, 1435, 1310, 1283, 1111 cm⁻¹.



Prepared according to the above procedure; tan solid (155 mg, 91%). TLC $R_f = 0.43$ (19:1 CH₂Cl₂/MeOH); ¹H NMR (DMSO-d₆, 500 MHz) δ 9.98 (s, 1H), 7.65 (s, 1H), 7.53 (d, J = 7.9 Hz, 1H), 7.30 (t, J = 7.8 Hz, 1H), 7.15 (d, J = 8.9 Hz, 1H), 7.09 (d, J = 7.6 Hz, 1H), 6.44 (d, J = 9.0 Hz, 1H), 5.48 (s, 2H), 5.11 (s, 2H), 2.03 (s, 3H) ppm; IR (thin film) v 3311, 1666, 1612, 1552, 1473, 1370, 1306, 1246, 1199 cm⁻¹.



Prepared according to the above procedure; off-white solid (133 mg, 97%). TLC R_f = 0.37 (19:1 CH₂Cl₂/MeOH); ¹H NMR (DMSO-d₆, 500 MHz) δ 7.91 (s, 1H), 7.60 (d, *J* = 7.6 Hz, 1H), 7.56 (d, *J* = 7.6 Hz, 1H), 7.50 (t, *J* = 7.6 Hz, 1H), 7.47 (s, 1H), 7.40 (t, *J* = 7.4 Hz, 1H), 7.16 (d, *J* = 8.8 Hz, 1H), 6.38 (d, *J* = 8.9 Hz, 1H), 5.50 (s, 2H), 5.34 (s, 2H) ppm; IR (thin film) v 3384, 3188, 1645, 1607, 1594, 1473, 1310, 1246, 1200, 1117 cm⁻¹.



Prepared according to the above procedure; off-white solid (190 mg, 99%). TLC R_f = 0.33 (19:1 CH₂Cl₂/MeOH); ¹H NMR (DMSO-d₆, 500 MHz) δ 8.01 (s, 1H), 7.95 (s, 1H), 7.83 (d, *J* = 7.6 Hz, 1H), 7.60 (d, *J* = 7.6 Hz, 1H), 7.49 (t, *J* = 7.6 Hz, 1H), 7.41 (s, 1H), 7.17 (d, *J* = 8.9 Hz, 1H), 6.49 (d, *J* = 8.9 Hz, 1H), 5.50 (s, 2H), 5.19 (s, 2H) ppm; IR (thin film) v 3345, 3182, 1651, 1606, 1474, 1452, 1406, 1375, 1305, 1244, 1200, 1116 cm⁻¹.



Prepared according to the above procedure; off-white solid (174 mg, 95%). TLC $R_f = 0.33$ (19:1 CH₂Cl₂/MeOH); ¹H NMR (DMSO-d₆, 500 MHz) δ 7.98 (s, 1H), 7.89 (d, *J* = 8.1 Hz, 2H), 7.51 (d, *J* = 8.1 Hz, 2H), 7.39 (s, 1H), 7.17 (d, *J* = 8.9 Hz, 1H), 6.47 (d, *J* = 9.0 Hz, 1H), 5.51 (s, 2H), 5.22 (s, 2H) ppm; IR (thin film) v 3338, 3178, 1641, 1614, 1572, 1477, 1452, 1304, 1120 cm⁻¹.



Prepared according to the above procedure; pale yellow solid (79 mg, 54%). TLC R_f = 0.44 (3:2 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.86 (dd, *J* = 5.4, 3.0 Hz, 2H), 7.72 (dd, *J* = 5.4, 3.0 Hz, 2H), 7.06 (d, *J* = 8.9 Hz, 1H), 6.28 (d, *J* = 8.9 Hz, 1H), 4.44 (s, 2H), 4.23 (t, *J* = 5.7 Hz, 2H), 4.15 (t, *J* = 5.7 Hz, 2H) ppm; IR (thin film) v 3480, 3374, 2918, 2849, 1712, 1611, 1464, 1393, 1306 cm⁻¹.

Hydrogenation of styrenyl olefins



Pd/C (5 wt %, 1.0 mg) was added to a solution of 2,6-dichloro-3-vinylaniline (20 mg, 0.11 mmol) in 2.5 mL of MeOH. The reaction flask was sealed with a septum and fitted with a N₂ gas inlet and an 18-gauge needle. The flask was flushed with N₂ for 5 min. The N₂ line was then replaced with a balloon of H₂ and the headspace of the flask was swept with H₂. The outlet needle was removed, the flask equipped with a fresh H₂ balloon, and the black suspension was stirred for exactly 1 h (Note: reaction times beyond 1 h resulted in inseparable mixtures of proto-dehalogenation products). Following this time, the mixture was sparged for 5 min with a gentle-stream of N₂ gas, diluted with 10 mL of EtOAc, and filtered through a pad of Celite. The flask and filter cake were washed with EtOAc and the combined filtrates were concentrated under reduced pressure. The oily residue was re-dissolved in CH₂Cl₂ to which ~0.5 g of silica gel was then added. The suspension was concentrated under reduced pressure and the solid material dry loaded onto a silica gel column pre-packed in hexanes. Purification of this material by chromatography on silica gel (gradient elution, 0:1→1:3 EtOAc/hexanes) furnished the desired product as a clear oil (10.3 mg, 51%). TLC R_f = 0.38 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 300 MHz) δ 7.13 (d, *J*= 8.3 Hz, 1H), 6.61 (d, *J*= 8.4 Hz, 1H), 4.49 (s, 2H), 2.72 (q, *J* = 7.5 Hz, 2H), 1.23 (t, *J* = 7.5 Hz, 3H) ppm; IR (thin film) v 3489, 3392, 2970, 2935, 2873, 1607, 1471, 1432 cm⁻¹.

Synthesis of substituted 2-bromo esters

Fischer esterification



2-Bromo-4-methylbenzoic acid (500 mg, 2.3 mmol) was dissolved in 6.0 mL of EtOH to which activated 3 Å molecular sieves (~0.5 g) were then added. Concentrated H₂SO₄ (1.0 mL) was added dropwise, the flask was fitted with a refluxed condenser, and the suspension was stirred at 70 °C for 3 h. Following this time, the mixture was cooled to room temperature and filtered through a pad of Celite. The flask and filter cake were rinsed with ~20 mL of EtOAc. The combined filtrates were transferred to a separatory funnel containing 70 mL of EtOAc. The organic fraction was washed with 1 x 70 mL of H₂O, 1 x 70 mL of saturated aqueous NaHCO₃, and 1 x 70 mL of saturated aqueous NaCl. The organic layer was collected, dried over MgSO₄, filtered and concentrated under reduced pressure. Purification of this material by chromatography on silica gel (gradient elution, 0:1→1:19 acetone/hexanes) furnished the desired product as a clear, colorless oil (395 mg, 71%). TLC R_f = 0.57 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 7.71 (d, *J* = 7.9 Hz, 1H), 7.48 (s, 1H), 7.15 (d, *J* = 7.9 Hz, 1H), 4.38 (q, *J* = 7.1 Hz, 2H), 2.36 (s, 3H), 1.39 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 2982, 1732, 1447, 1408, 1366, 1291, 1245, 1182, 1146 cm⁻¹.



Prepared according to the above procedure; colorless oil (461 mg, 82%). TLC R_f = 0.57 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 7.57 (dd, *J* = 2.3, 0.8 Hz, 1H), 7.51 (d, *J* = 8.2 Hz, 1H), 7.12 (ddq, *J* = 8.2, 2.3, 0.7 Hz, 1H), 4.39 (q, *J* = 7.1 Hz, 2H), 2.33 (s, 3H), 1.40 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 2982, 1732, 1471, 1297, 1251, 1203, 1110 cm⁻¹.



Prepared according to the above procedure; colorless oil (374 mg, 66%). TLC R_f = 0.41 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 7.86 (dd, *J* = 8.7, 6.0 Hz, 1H), 7.40 (dd, *J* = 8.3, 2.5 Hz, 1H), 7.07 (ddd, *J* = 8.8, 7.7, 2.5 Hz, 1H), 4.39 (q, *J* = 7.1 Hz, 2H), 1.40 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3081, 2983, 1732, 1596, 1487, 1385, 1366, 1286, 1253, 1208, 1109 cm⁻¹.

Prepared according to the above procedure; colorless oil (364 mg, 64%). TLC $R_f = 0.13$ (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 8.61 (dd, J = 4.6, 1.4 Hz, 1H), 7.99 (dd, J = 8.2, 1.4 Hz, 1H), 7.29 (dd, J = 8.2, 4.6 Hz, 1H), 4.49 (q, J = 7.1 Hz, 2H), 1.44 (t, J = 7.1 Hz, 3H) ppm; IR (thin film) v 3056, 2983, 1732, 1570, 1424, 1367, 1301, 1205, 1174, 1141 cm⁻¹.



Prepared according to the above procedure (the corresponding methyl ester may be prepared in >90%

yield via the protocol described in *Esterification through EDC coupling* and alternatively used for the synthesis of AK-44); colorless oil (140 mg, 31%). TLC R_f = 0.37 (3:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz, appears as a mixture of protonated and free base) δ 8.48 (dd, *J* = 4.7, 2.1 Hz, 1H), 8.07 (dd, *J* = 7.7, 2.1 Hz, 1H), 7.34 (dd, *J* = 7.7, 4.8 Hz, 1H), 4.43 (q, *J* = 7.1 Hz, 2H), 1.42 (t, *J* = 7.1, 3H) ppm; IR (thin film) v 2983, 1732, 1578, 1557, 1402, 1367, 1301, 1275, 1245, 1142 cm⁻¹.

Prepared according to the above procedure; colorless oil (130 mg, 29%). TLC R_f = 0.40 (3:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 8.85 (s, 1H), 8.62 (d, *J* = 4.9 Hz, 1H), 7.62 (dd, *J* = 4.9, 0.7 Hz, 1H), 4.44 (q, *J* = 7.1 Hz, 2H), 1.42 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 2983, 1737, 1470, 1398, 1367, 1303, 1270, 1214, 1181, 1129 cm⁻¹.

Esterification with Mel on sterically hindered 2,6-disubstituted substrates



To an ice-cold suspension of NaH (59 mg, 2.45 mmol, 1.2 equiv) in 3.0 mL of anhydrous DMF was added dropwise via cannula a solution of 2-bromo-6-methylbenzoic acid (439 mg, 2.04 mmol) in 9 mL of DMF. Gas evolution ensued immediately and stirring at 0 °C continued until bubbling ceased (~30 min). Iodomethane (0.25 mL, 4.08 mmol, 2.0 equiv) was added dropwise via syringe, the solution was warmed to ambient temperature and stirred for 2 h. The flask was then fitted with a reflux condenser and the contents stirred at 80 °C for 1 h. Upon heating, the opaque reaction mixture slowly transitioned to clear yellow. The reaction was cooled to room temperature and transferred to a separatory funnel with 200 mL of Et₂O. The ethereal layer was washed with 4 x 200 mL H₂O, dried over MgSO₄, filtered and concentrated under reduced pressure to an oily residue. Purification of this material by chromatography on silica gel (gradient elution, 0:1 \rightarrow 1:19 EtOAc/hexanes) furnished the desired product as a colorless oil (363 mg, 78%). TLC R_f = 0.30 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 7.44–7.36 (m, 1H), 7.21–7.12 (m, 2H), 3.95 (s, 3H), 2.33 (s, 3H) ppm; IR (thin film) v 2952, 1738, 1594, 1565, 1451, 1278, 1245, 1178, 1150, 1103 cm⁻¹.

Esterification through EDC coupling



The following procedure is adapted from Stangeland, et al.⁴ To an ice-cold suspension of 4-bromonicotinic acid (102 mg, 0.50 mmol) in 2.0 mL of CH₂Cl₂ was added successively 4-dimethylaminopyridine (6.2 mg, 0.05 mmol, 0.1 equiv), freshly distilled MeOH (80 μ L, 2.0 mmol, 4.0 equiv), and 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (107 mg, 0.55 mmol, 1.1 equiv). Upon addition of EDC, the solution changed from opaque to clear. The reaction was stirred for 3 h at 0 °C and then quenched by the addition of 3 mL of H₂O. The reaction was transferred to a separatory funnel with 30 mL of H₂O and extracted with 1 x 30 mL of CH₂Cl₂. The organic layer was washed with 2 x 30 mL of saturated aqueous NaCl, dried over Na₂SO₄, filtered and concentrated under reduced pressure to a yellow oil (97 mg, 89%). This material was judged to be sufficiently pure by ¹H NMR for use in the subsequent step without further purification. TLC R_f = 0.32 (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 8.97 (s, 1H), 8.45 (d, *J* = 5.3 Hz, 1H), 7.61 (d, *J* = 5.3 Hz, 1H), 3.96 (s, 3H) ppm; IR (thin film) v 2953, 1738, 1569, 1548, 1466, 1435, 1396, 1291, 1272, 1217, 1127 cm⁻¹.

Buchwald-Hartwig amination



The following procedure is adapted from Sadighi, et al.¹ An oven-dried 5-mL microwave vial was charged with 2,6-diethylaniline (200 mg, 1.34 mmol), ethyl 2-bromobenzoate (307 mg, 1.34 mmol), and DPEphos (54 mg, 0.10 mmol, 0.075 equiv). The vial was sealed with a septum and purged with argon. Anhydrous toluene (3.0 mL) was added and the solution was sparged with a gentle stream of argon for ~5 min. The septum was quickly removed and Pd(OAc)₂ (15 mg, 0.067 mmol, 0.05 equiv) was added in a single portion. The vial was resealed and the yellow solution was stirred under argon for 10 min. Following this time, the septum was quickly removed and Cs₂CO₃ (611 mg, 1.88 mmol, 1.4 equiv) was added. The headspace of the vial was flushed with argon and the vessel was quickly resealed with a crimped microwave vial cap. The reaction was stirred at 160 °C in a microwave reactor for 2 h (Note: the reaction may also be performed with conventional heating at 115 °C for 20 h; reaction yields are comparable). The reaction mixture was cooled to room temperature, diluted with 10 mL of EtOAc, and filtered through a plug of Celite. The flask and filter cake were rinsed with ~30 mL of EtOAc, and the combined filtrates were concentrated under reduced pressure. The brown residue was re-dissolved in CH_2Cl_2 to which ~1 g of silica gel was then added. The suspension was concentrated under reduced pressure and the solid material dry loaded onto a silica gel column pre-packed in pentane. Purification by chromatography on silica gel (gradient elution, $0:1 \rightarrow 1:19$ Et₂O/pentane) furnished the desired product as a white solid (180 mg, 45%). TLC $R_f = 0.40$ (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 9.05 (s, 1H), 7.97 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.26–7.13 (m,

^{4.} Stangeland, E. L.; Patterson, L. J.; Zipfel, S. 1-(2-phenoxymethylheteroaryl)piperidine and piperazine compounds. US20110230495 A1, September 22, 2011.

4H), 6.61 (ddd, J = 8.1, 7.1, 1.2 Hz, 1H), 6.19 (dd, J = 8.5, 1.2 Hz, 1H), 4.39 (q, J = 7.1 Hz, 2H), 2.55 (dq, J = 14.6, 7.3 Hz, 4H), 1.43 (t, J = 7.1 Hz, 3H), 1.13 (t, J = 7.6 Hz, 6H) ppm; IR (thin film) v 3320, 2967, 2934, 1680, 1578, 1506, 1453, 1253, 1231 cm⁻¹.



Prepared according to the above procedure; pale yellow residue (207 mg, 67%). TLC R_f = 0.6 (9:1 hexanes/acetone); ¹H NMR (CDCl₃, 400 MHz) δ 9.70 (s, 1H), 8.03 (d, *J* = 8.0, 1H), 7.45–7.34 (m, 2H), 7.30 (d, *J* = 8.4 Hz, 1H), 7.15–7.12 (m, 2H), 6.87 (t, *J* = 7.6 Hz, 1H), 4.39 (q, *J* = 7.1 Hz, 2H), 1.42 (t, *J* = 7.1, 3H) ppm; IR (thin film) v 3299, 3261, 2982, 1687, 1581, 1522, 1448, 1402, 1368, 1320, 1249, 1185, 1164, 1144, 1084 cm⁻¹.



Prepared according to the above procedure; white crystalline solid (174 mg, 56%). TLC R_f = 0.72 (9:1 hexanes/acetone); ¹H NMR (CDCl₃, 400 MHz) δ 9.68 (s, 1H), 8.03 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.49 (d, *J* = 2.4 Hz, 1H), 7.45–7.40 (m, 1H), 7.38–7.32 (m, 2H), 6.94–6.86 (m, 2H), 4.39 (q, *J* = 7.1 Hz, 2H), 1.42 (t, *J* = 7.2 Hz, 3H) ppm; IR (thin film) v 3293, 2986, 1689, 1583, 1520, 1456, 1410, 1256, 1224, 1085 cm⁻¹.



Prepared according to the above procedure (Note: the reaction was stirred for 1 h at 160 °C; these conditions were employed prior to reaction optimization); off-white solid (62 mg, 17%). TLC $R_f = 0.58$ (9:1 hexanes/acetone); ¹H NMR (CDCl₃, 400 MHz) δ 9.41 (s, 1H), 8.02 (d, *J* = 8.0 Hz, 1H), 7.42 (d, *J* = 8.1 Hz, 2H), 7.30–7.24 (m, 1H), 7.15 (t, *J* = 8.1 Hz, 1H), 6.78 (t, *J* = 7.5 Hz, 1H), 6.35 (d, *J* = 8.5 Hz, 1H), 4.40 (q, *J* = 7.1 Hz, 2H), 1.43 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3297, 3081, 2981, 1683, 1586, 1506, 1452, 1314, 1256, 1162, 1144, 1084 cm⁻¹.



Prepared according to the above procedure with conventional heating at 100 °C for 16 h (unoptimized reaction conditions); off-white solid (23 mg, 7%). TLC 0.60 R_f = (9:1 hexanes/acetone); ¹H NMR (CDCl₃, 500 MHz) δ 9.61 (s, 1H), 8.02 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.46–7.40 (m, 2H), 7.37 (ddd, *J* = 8.7, 7.1, 1.7 Hz, 1H), 7.22 (d, *J* = 8.5, 1H), 7.18 (dd, *J* = 8.7, 2.4 Hz, 1H), 6.85 (t, *J* = 8.1 Hz, 1H), 4.39 (q, *J* = 7.1 Hz, 2H), 1.42 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3294, 2925, 1690, 1592, 1521, 1479, 1455, 1321, 1255, 1225 cm⁻¹.

Prepared according to the above procedure with conventional heating at 100 °C for 16 h. rac-BINAP (0.15 equiv) was used in place of DPEphos (unoptimized reaction conditions); white solid (42 mg, 14%). TLC R_f = 0.60 (9:1 hexanes/acetone); ¹H NMR (CDCl₃, 500 MHz) δ 9.56 (s, 1H), 8.02 (dd, *J* = 8.1, 1.7 Hz, 1H), 7.41–7.35 (m, 3H), 7.27 (d, *J* = 7.9 Hz, 1H), 7.08 (dd, *J* = 8.7, 2.6 Hz, 1H), 6.84 (t, *J* = 8.1 Hz, 1H), 4.38 (q, *J* = 7.1 Hz, 2H), 1.43 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3307, 2985, 2927, 1683, 1587, 1562, 1517, 1476, 1454, 1368, 1323, 1257, 1230, 1149 cm⁻¹.



Prepared according to the above procedure; off-white solid (188 mg, 55%). TLC R_f = 0.68 (24:1 pentane/Et₂O); ¹H NMR (CDCl₃, 500 MHz) δ 9.37 (s, 1H), 8.01 (dd, *J* = 8.1, 1.7 Hz, 1H), 7.44 (s, 2H), 7.29 (ddd, *J* = 8.7, 7.2, 1.7 Hz, 1H), 6.80 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1H), 6.32 (dd, *J* = 8.4, 1.1 Hz, 1H), 4.40 (q, *J* = 7.1 Hz, 2H), 1.43 (t, *J* = 7.3 Hz, 3H) ppm; IR (thin film) v 3307, 3074, 2984, 1682, 1583, 1505, 1450, 1369, 1312, 1253, 1145, 1091 cm⁻¹.



Prepared according to the above procedure; 1 h reaction time (unoptimized reaction conditions); white solid (20 mg, 7%). TLC R_f = 0.60 (9:1 hexanes/acetone); ¹H NMR (CDCl₃, 500 MHz) δ 9.65 (s, 1H), 8.04 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.39–7.35 (m, 2H), 7.31–7.29 (m, 1H), 7.13 (t, *J* = 7.8 Hz, 1H), 6.96 (d, *J* = 7.6 Hz, 1H), 6.83 (t, *J* = 7.7 Hz, 1H), 4.42 (q, *J* = 7.1 Hz, 2H), 2.45 (s, 3H), 1.45 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3307, 2981, 1687, 1594, 1577, 1520, 1472, 1454, 1323, 1253, 1224 cm⁻¹.



Prepared according to the above procedure; yellow oil (80 mg, 28%). TLC $R_f = 0.68$ (24:1 pentane/Et₂O); ¹H NMR (CDCl₃, 400 MHz) δ 9.54 (s, 1H), 8.00 (dd, J = 8.0, 1.7 Hz, 1H), 7.38–7.28 (m, 3H), 7.27–7.21 (m, 1H), 6.83–6.77 (m, 2H), 4.38 (q, J = 7.5 Hz, 2H), 2.30 (s, 3H), 1.41 (t, J = 7.1 Hz, 3H) ppm; IR (thin film) v 3306, 2980, 2927, 1687, 1585, 1522, 1455, 1368, 1319, 1255, 1228, 1163, 1145, 1083 cm⁻¹.



Prepared according to the above procedure; colorless oil (140 mg, 50%). TLC R_f = 0.61 (24:1 pentane/Et₂O); ¹H NMR (CDCl₃, 400 MHz) δ 9.21 (s, 1H), 8.01 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.33 (t, *J* = 7.9 Hz, 1H), 7.20–7.11 (m, 1H), 6.99 (t, *J* = 8.0 Hz, 2H), 6.79 (t, *J* = 7.6 Hz, 1H), 6.60 (d, *J* = 8.0 Hz, 1H), 4.39 (q, *J* = 7.2 Hz, 2H), 1.42 (t, *J* = 7.2 Hz, 3H) ppm; IR (thin film) v 3308, 2983, 1683, 1584, 1519, 1474, 1455, 1320, 1280, 1245, 1164, 1145, 1085 cm⁻¹.



Prepared according to the above procedure; 1.5 h reaction time (unoptimized reaction conditions); yellow oil (190 mg, 40%). TLC R_f = 0.58 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 500 MHz) δ 9.04 (s, 1H), 7.99 (dd, *J* = 8.0, 1.4 Hz, 1H), 7.20 (m, 1H), 7.14 (m, 3H), 6.63 (ddd, *J* = 8.1, 7.0, 1.1 Hz, 1H), 6.20 (dd, *J* = 8.6, 1.2 Hz, 1H), 4.39 (q, *J* = 7.1 Hz, 2H), 2.21 (s, 6H), 1.43 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3320, 2980, 1682, 1579, 1507, 1454, 1251, 1160, 1142 cm⁻¹.



Prepared according to the above procedure; white solid (115 mg, 40%). TLC R_f = 0.52 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 9.37 (s, 1H), 7.90 (d, *J* = 8.1 Hz, 1H), 7.32 (d, *J* = 8.1 Hz, 1H), 7.11 (d, *J* = 8.3 Hz, 1H), 6.58 (d, *J* = 8.2 Hz, 1H), 6.10 (s, 1H), 4.38 (q, *J* = 7.1 Hz, 2H), 2.42 (s, 3H), 2.22 (s, 3H), 1.42 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3287, 2978, 2926, 1677, 1615, 1572, 1510, 1460, 1241, 1156 cm⁻¹.



Prepared according to the above procedure; white solid (32 mg, 10%). TLC R_f = 0.52 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 9.22 (s, 1H), 7.83–7.78 (m, 1H), 7.29 (d, *J* = 8.2 Hz, 1H), 7.11–7.07 (m, 2H), 6.25 (d, *J* = 8.5 Hz, 1H), 4.40 (q, *J* = 7.1 Hz, 2H), 2.40 (s, 3H), 2.27 (s, 3H), 1.43 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3306, 2981, 2924, 1685, 1584, 1511, 1461, 1256, 1234, 1207 cm⁻¹.



Prepared according to the above procedure; white solid (120 mg, 50%). TLC R_f = 0.42 (19:1 hexanes/acetone); ¹H NMR (CDCI₃, 300 MHz) δ 8.24 (s, 1H), 7.27 (d, *J* = 8.0 Hz, 1H), 7.13–7.02 (m, 2H),

6.69 (d, *J* = 7.6 Hz, 1H), 6.23 (d, *J* = 8.4 Hz, 1H), 3.96 (s, 3H), 2.48 (s, 3H), 2.39 (s, 3H) ppm; IR (thin film) v 3289, 2947, 1677, 1583, 1492, 1460, 1303, 1253 cm⁻¹.

Prepared according to the above procedure; white solid (33 mg, 52%). TLC R_f = 0.32 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 9.42 (s, 1H), 8.01 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.36 (d, *J* = 9.0 Hz, 1H), 7.27 (ddd, *J* = 8.6, 7.1, 1.7 Hz, 1H), 6.82 (d, *J* = 9.0 Hz, 1H), 6.77 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1H), 6.37 (dd, *J* = 8.4, 1.1 Hz, 1H), 4.40 (q, *J* = 7.1 Hz, 2H), 3.93 (s, 3H), 1.43 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3300, 2980, 1683, 1578, 1508, 1466, 1455, 1252 cm⁻¹.



Prepared according to the above procedure; isolated as an inseparable mixture of products and resolved by reversed-phase HPLC following ester hydrolysis (see below for details); white solid (137 mg); TLC $R_f = 0.33$ (19:1 hexanes/EtOAc, single spot for both products); ¹H NMR (CDCl₃, 400 MHz), see spectrum below; IR (thin film) v 3435, 1688, 1586, 1501, 1322, 1257, 1134 cm⁻¹.



Prepared according to the above procedure; isolated as an inseparable mixture of products and resolved by HPLC following ester hydrolysis (see below for details); white solid (93 mg); TLC $R_f = 0.9$ (19:1 hexanes/acetone); IR (thin film) v 3492, 3392, 3299, 2976, 1683, 1604, 1588, 1506, 1454, 1369, 1318, 1251 cm⁻¹.



Prepared according to the above procedure; white solid (6 mg, 6%). TLC $R_f = 0.43$ (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 9.49 (s, 1H), 8.02 (dd, J = 8.0, 1.6 Hz, 1H), 7.48–7.38 (m, 6H), 7.31 (ddd, J = 8.6, 7.1, 1.7 Hz, 1H), 7.21 (d, J = 8.3 Hz, 1H), 6.78 (ddd, J = 8.1, 7.1, 1.1 Hz, 1H), 6.44 (dd, J = 8.4, 1.1 Hz, 1H), 4.40 (q, J = 7.1 Hz, 2H), 1.43 (t, J = 7.1 Hz, 3H) ppm; IR (thin film) v 3298, 1684, 1535, 1508, 1452, 1368, 1316, 1251, 1162, 1144 cm⁻¹.



Prepared according to the above procedure; colorless film (26 mg, 22%). TLC R_f = 0.23 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 9.49 (s, 1H), 8.02 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.45 (d, *J* = 8.2 Hz, 1H), 7.40 (ddd, *J* = 8.3, 7.4, 1.7 Hz, 1H), 7.31 (t, *J* = 7.3 Hz, 1H), 7.21 (dd, *J* = 7.5, 1.7 Hz, 1H), 7.17 (d, *J* = 8.3 Hz, 1H), 7.04 (td, *J* = 7.4, 1.1 Hz, 1H), 6.99 (d, *J* = 8.3 Hz, 1H), 6.78 (t, *J* = 7.6 Hz, 1H), 6.48 (d, *J* = 52.5 Hz, 1H), 4.40 (q, *J* = 7.1 Hz, 2H), 3.80 (s, 3H), 1.43 (t, *J* = 7.1, 1.0 Hz, 3H) ppm; IR (thin film) v 3299, 2980, 1683, 1584, 1508, 1463, 1451, 1251 cm⁻¹.



Prepared according to the above procedure; colorless film (21 mg, 12%). TLC R_f = 0.40 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 9.40 (s, 1H), 8.01 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.34 (d, *J* = 8.3 Hz, 1H), 7.38 (td, *J* = 8.4, 1.6 Hz, 1H), 7.03 (d, *J* = 8.3 Hz, 1H), 6.76 (td, *J* = 7.1, 1.1 Hz, 1H), 6.36 (dd, *J* = 8.4, 1.0 Hz, 1H), 5.69 (tt, *J* = 3.7, 1.8 Hz, 1H), 4.40 (q, *J* = 7.1 Hz, 2H), 2.32–2.24 (m, 2H), 2.22–2.13 (m, 2H), 1.80–1.72 (m, 2H), 1.72–1.64 (m, 2H), 1.42 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) 3299, 2932, 1685, 1585, 1508, 1453, 1368, 1251, 1162, 1144 cm⁻¹.



Prepared according to the above procedure; white solid (42 mg, 24%). TLC $R_f = 0.31$ (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 9.42 (s, 1H), 8.01 (ddd, J = 8.0, 1.7, 0.4 Hz, 1H), 7.49–7.27 (m, 7H), 6.85 (d, J = 9.0 Hz, 1H), 6.78 (ddd, J = 8.2, 7.1, 1.1 Hz, 1H), 6.38 (dd, J = 8.4, 1.1 Hz, 1H), 5.19 (s, 2H), 4.40 (q, J = 7.1 Hz, 2H), 1.43 (t, J = 7.1 Hz, 3H) ppm; IR (thin film) v 3299, 1683, 1578, 1507, 1451, 1369, 1296, 1251, 1162 cm⁻¹.

Prepared according to the above procedure; colorless film (16 mg, 10%). TLC R_f = 0.43 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 9.38 (s, 1H), 8.00 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.32 (d, *J* = 8.9 Hz, 1H), 7.27 (td, *J* = 8.5, 7.8, 1.6 Hz, 1H), 6.81–6.74 (m, 2H), 6.37 (d, *J* = 8.4 Hz, 1H), 4.40 (q, *J* = 7.1 Hz, 2H), 3.83 (d, *J* = 6.1 Hz, 2H), 1.92–1.84 (m, 3H), 1.81–1.68 (m, 4H), 1.42 (t, *J* = 7.1 Hz, 3H), 1.36–1.18 (m, 2H), 1.15–1.05 (m, 2H) ppm; IR (thin film) v 3300, 2925, 2853, 1684, 1577, 1509, 1452, 1298, 1250, 1162 cm⁻¹.

Prepared according to the above procedure; white solid (135 mg, 37%). TLC $R_f = 0.31$ (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 9.42 (s, 1H), 8.01 (dd, J = 8.0, 1.7 Hz, 1H), 7.33 (d, J = 9.0 Hz, 1H), 7.29 (ddd, J = 8.6, 7.1, 1.7 Hz, 1H), 7.09 (d, J = 9.0 Hz, 1H), 6.78 (ddd, J = 8.2, 7.1, 1.1 Hz, 1H), 6.37 (dd, J = 8.5, 1.0 Hz, 1H), 5.27 (s, 2H), 4.40 (q, J = 7.1 Hz, 2H), 3.54 (s, 3H), 1.43 (t, J = 7.1 Hz, 3H) ppm; IR (thin film) v 3299, 2980, 1682, 1580, 1505, 1454, 1251, 1162 cm⁻¹.

Prepared according to the above procedure. Reaction mixture was diluted with 1:9 MeOH/CH₂Cl₂ instead of EtOAc before filtering through Celite due to limited compound solubility in less polar solvents; white solid (45 mg, 14%). TLC R_f = 0.10 (9:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 9.40 (s, 1H), 8.17 (d, *J* = 4.3 Hz, 1H), 7.32 (d, *J* = 8.2 Hz, 1H), 7.21 (dd, *J* = 8.6, 4.3 Hz, 1H), 7.14 (d, *J* = 8.3 Hz, 1H), 6.71 (d, *J* = 8.6 Hz, 1H), 4.53 (q, *J* = 7.1 Hz, 2H), 2.41 (s, 3H), 1.49 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3301, 2982, 1683, 1591, 1489, 1453, 1402, 1373, 1321, 1249, 1199 cm⁻¹.



Prepared according to the above procedure. Reaction mixture was diluted with 1:9 MeOH/CH₂Cl₂ instead of EtOAc before filtering through Celite due to limited compound solubility in less polar solvents. In the filtration step, the ethyl ester transesterified to the methyl ester; pale yellow solid (48 mg, 26%). TLC R_f = 0.42 (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 9.00 (s, 1H), 8.07 (d, *J* = 5.2 Hz, 1H), 7.82 (s, 1H),

7.72 (d, *J* = 5.1 Hz, 1H), 7.33 (d, *J* = 8.2 Hz, 1H), 7.15 (d, *J* = 8.2 Hz, 1H), 3.98 (s, 3H), 2.42 (s, 3H) ppm; IR (thin film) v 3320, 2951, 1694, 1562, 1494, 1459, 1441, 1424, 1301, 1226, 1202, 1179 cm⁻¹.

Prepared according to the above procedure. Reaction mixture was diluted with 1:9 MeOH/CH₂Cl₂ instead of EtOAc before filtering through Celite due to limited compound solubility in less polar solvents; pale yellow solid (72 mg, 36%). TLC R_f = 0.65 (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 9.56 (s, 1H), 8.30–8.20 (m, 2H), 7.30 (d, *J* = 8.3 Hz, 1H), 7.11 (d, *J* = 8.2 Hz, 1H), 6.71 (dd, *J* = 7.7, 4.9 Hz, 1H), 4.42 (q, *J* = 7.1 Hz, 2H), 2.40 (s, 3H), 1.43 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3313, 2984, 1679, 1584, 1487, 1458, 1382, 1295, 1244, 1141 cm⁻¹.

$$\overbrace{CI}^{CI} \xrightarrow{H} \overbrace{CI}^{CO_2Me}_{N}$$

Prepared according to the above procedure. Reaction mixture was diluted with 1:9 MeOH/CH₂Cl₂ instead of EtOAc before filtering through Celite due to limited compound solubility in less polar solvents; off-white solid (7 mg, 5%). TLC R_f = 0.20 (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 9.60 (s, 1H), 9.03 (s, 1H), 8.23 (d, *J* = 6.1 Hz, 1H), 7.34 (d, *J* = 8.3 Hz, 1H), 7.20 (d, *J* = 8.3 Hz, 1H), 6.16 (d, *J* = 6.0 Hz, 1H), 3.98 (s, 3H), 2.42 (s, 3H) ppm; IR (thin film) v 3288, 1692, 1596, 1572, 1502, 1459, 1316, 1226, 1113 cm⁻¹.

$$\overbrace{Me}^{CI} \xrightarrow{H}_{CO_2Et} CO_2Et$$

Prepared according to the above procedure; colorless oil (110 mg, 34%). TLC R_f = 0.26 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.58 (ddd, *J* = 7.7, 1.6, 1.0 Hz, 1H), 7.41 (dd, *J* = 2.6, 1.6 Hz, 1H), 7.27 (td, *J* = 8.0, 2.0 Hz, 2H), 7.05 (d, *J* = 8.3 Hz, 1H), 6.81 (ddd, *J* = 8.1, 2.5, 1.0 Hz, 1H), 5.88 (s, 1H), 4.35 (q, *J* = 7.1 Hz, 2H), 2.40 (s, 3H), 1.37 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3353, 2981, 1715, 1608, 1590, 1510, 1484, 1462, 1392, 1368, 1290, 1231, 1106 cm⁻¹.



Prepared according to the above procedure; white solid (145 mg, 45%). TLC $R_f = 0.19$ (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.92 (d, J = 8.6 Hz, 2H), 7.30 (d, J = 8.3 Hz, 1H), 7.10 (d, J = 8.3 Hz, 1H), 6.63 (d, J = 8.6 Hz, 2H), 5.95 (s, 1H), 4.33 (q, J = 7.1 Hz, 2H), 2.41 (s, 3H), 1.36 (t, J = 7.1 Hz, 3H) ppm; IR (thin film) v 3390, 3329, 2981, 1694, 1607, 1519, 1462, 1367, 1311, 1278, 1174, 1108 cm⁻¹.



Prepared according to the above procedure; isolated as an inseparable mixture of products (~2.5:1 desired product/carbazole) and resolved by reversed-phase HPLC following ester hydrolysis; pale pink solid (184 mg); TLC $R_f = 0.64$ (9:1 hexanes/EtOAc, single spot for both products); ¹H NMR (CDCl₃, 400 MHz), see spectrum below; IR (thin film) v 3427, 3293, 2982, 2930, 1687, 1617, 1591, 1513, 1461, 1258, 1128 cm⁻¹.



Prepared according to the above procedure; colorless residue (205 mg, 55%). TLC $R_f = 0.43$ (4:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 9.56 (s, 1H), 8.29–8.20 (m, 2H), 7.46 (d, *J* = 7.6 Hz, 2H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.35–7.30 (m, 2H), 6.87 (d, *J* = 8.9 Hz, 1H), 6.75–6.69 (m, 1H), 5.17 (s, 2H), 3.96 (s, 3H) ppm; IR (thin film) v 1737, 1693, 1577, 1495, 1452, 1396, 1292, 1245, 1137 cm⁻¹.



Prepared according to the above procedure; colorless residue (97 mg, 80%). TLC R_f = 0.23 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 9.43 (s, 1H), 8.02 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.59 (t, *J* = 7.6 Hz, 1H), 7.36–7.26 (m, 3H), 7.19 (t, *J* = 7.6 Hz, 1H), 7.12–7.07 (m, 1H), 6.90 (d, *J* = 9.0 Hz, 1H), 6.78 (t, *J* = 7.8 Hz, 1H), 6.38 (d, *J* = 8.3 Hz, 1H), 5.25 (s, 2H), 4.41 (q, *J* = 7.1 Hz, 2H), 1.43 (t, *J* = 7.1 Hz, 3H) ppm; ¹⁹F NMR (CDCl₃, 376 MHz) δ –119.10 (dt, *J* = 10.2, 6.6 Hz, 1F) ppm; IR (thin film) v 3298, 2981, 1683, 1608, 1578, 1507, 1453, 1382, 1297, 1252, 1163, 1146 cm⁻¹.



Prepared according to the above procedure; viscous, colorless oil that slowly solidified (82 mg, 48%). TLC $R_f = 0.19$ (19:1 hexanes/Et₂O); ¹H NMR (CDCl₃, 400 MHz) δ 9.43 (s, 1H), 8.00 (dd, J = 8.0, 1.7 Hz, 1H), 7.44 (d, J = 6.9 Hz, 1H), 7.31 (dd, J = 8.9, 0.5 Hz, 1H), 7.29–7.17 (m, 4H), 6.87 (d, J = 8.9 Hz, 1H), 6.80–6.72 (m, 1H), 6.37 (d, J = 8.4 Hz, 1H), 5.11 (s, 2H), 4.38 (q, J = 7.1 Hz, 2H), 2.38 (s, 3H), 1.41 (t, J = 7.1

Hz, 3H) ppm; IR (thin film) v 3298, 2980, 1683, 1606, 1577, 1506, 1453, 1369, 1295, 1251, 1163, 1145 cm⁻¹.



Prepared according to the above procedure; viscous, colorless oil (87 mg, 50%). TLC R_f = 0.20 (19:1 hexanes/ Et₂O); ¹H NMR (CDCl₃, 400 MHz) δ 9.43 (s, 1H), 8.01 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.33–7.22 (m, 5H), 7.14 (d, *J* = 7.0 Hz, 1H), 6.84 (d, *J* = 9.0 Hz, 1H), 6.77 (ddd, *J* = 8.0, 7.2, 0.9 Hz, 1H), 6.38 (d, *J* = 8.4 Hz, 1H), 5.13 (s, 2H), 4.40 (q, *J* = 7.1 Hz, 2H), 2.37 (s, 3H), 1.42 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3298, 2980, 1684, 1606, 1578, 1508, 1452, 1369, 1298, 1250, 1162, 1145 cm⁻¹.



Prepared according to the above procedure; viscous, colorless oil that slowly solidified (85 mg, 49%). TLC $R_f = 0.19$ (19:1 hexanes/ Et₂O); ¹H NMR (CDCl₃, 400 MHz) δ 9.42 (s, 1H), 8.01 (dd, J = 8.0, 1.7 Hz, 1H), 7.35 (d, J = 8.1 Hz, 2H), 7.32–7.25 (m, 2H), 7.20 (d, J = 8.2 Hz, 2H), 6.85 (d, J = 9.0 Hz, 1H), 6.77 (ddd, J = 8.2, 7.2, 1.1 Hz, 1H), 6.37 (d, J = 8.4 Hz, 1H), 5.15 (s, 2H), 4.40 (q, J = 7.1 Hz, 2H), 2.37 (s, 3H), 1.43 (t, J = 7.1 Hz, 3H) ppm; IR (thin film) v 3298, 2981, 2926, 1683, 1605, 1578, 1507, 1452, 1369, 1314, 1296, 1250, 1163, 1145 cm⁻¹.



Prepared according to the above procedure; white solid (57 mg, 37%). TLC R_f = 0.25 (9:1 hexanes/ Et₂O); ¹H NMR (CDCl₃, 400 MHz) δ 9.43 (s, 1H), 8.00 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.39 (d, *J* = 8.9 Hz, 1H), 7.31– 7.25 (m, 1H), 6.95 (d, *J* = 9.0 Hz, 1H), 6.78 (ddd, *J* = 8.0, 7.2, 1.1 Hz, 1H), 6.34 (d, *J* = 8.5 Hz, 1H), 5.20 (s, 2H), 4.40 (q, *J* = 7.1 Hz, 2H), 1.42 (t, *J* = 7.1 Hz, 3H) ppm; ¹⁹F NMR (CDCl₃, 376 MHz) δ –142.23 (dd, *J* = 22.3, 8.6 Hz, 2F), –152.13 (t, *J* = 20.8 Hz, 1F), –161.38 to –161.60 (m, 2F) ppm; IR (thin film) v 3299, 2984, 1685, 1658, 1581, 1508, 1453, 1383, 1312, 1292, 1251, 1163, 1134 cm⁻¹.



Prepared according to the above procedure; viscous, colorless oil (67 mg, 39%). TLC R_f = 0.29 (9:1 hexanes/Et₂O); ¹H NMR (CDCl₃, 400 MHz) δ 9.39 (s, 1H), 8.00 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.35–7.20 (m, 7H), 6.79–6.72 (m, 2H), 6.34 (d, *J* = 8.4 Hz, 1H), 4.38 (q, *J* = 7.1 Hz, 2H), 4.21 (t, *J* = 6.9 Hz, 2H), 3.15 (t, *J* = 6.9 Hz, 2H), 1.41 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3299, 3028, 2980, 1683, 1605, 1578, 1508, 1453, 1388, 1368, 1296, 1251, 1163, 1145 cm⁻¹.



Prepared according to the above procedure; off-white solid (22 mg, 30%). TLC $R_f = 0.17$ (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 9.44 (s, 1H), 8.04 (dd, *J* = 17.5, 7.9 Hz, 2H), 7.90 (d, *J* = 7.9 Hz, 1H), 7.61 (t, *J* = 7.7 Hz, 1H), 7.40 (t, *J* = 7.7 Hz, 1H), 7.35–7.28 (m, 2H), 6.92 (d, *J* = 8.9 Hz, 1H), 6.78 (t, *J* = 7.6 Hz, 1H), 6.40 (d, *J* = 8.4 Hz, 1H), 5.61 (s, 2H), 4.41 (q, *J* = 7.2 Hz, 2H), 3.93 (s, 3H), 1.43 (t, *J* = 7.0 Hz, 3H) ppm; IR (thin film) v 3299, 2981, 2951, 1718, 1684, 1579, 1509, 1452, 1300, 1251, 1139 cm⁻¹.



Prepared according to the above procedure; white solid (16 mg, 27%). TLC R_f = 0.59 (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 9.43 (s, 1H), 8.13 (s, 1H), 8.07–7.98 (m, 2H), 7.70 (d, *J* = 7.7 Hz, 1H), 7.49 (t, *J* = 7.7 Hz, 1H), 7.34–7.26 (m, 2H), 6.85 (d, *J* = 8.9 Hz, 1H), 6.78 (t, *J* = 7.5 Hz, 1H), 6.37 (d, *J* = 8.4 Hz, 1H), 5.22 (s, 2H), 4.41 (q, *J* = 7.2 Hz, 2H), 3.93 (s, 3H), 1.43 (t, *J* = 7.2 Hz, 3H) ppm; IR (thin film) v 3446, 1727, 1693, 1606, 1505, 1431, 1273, 1208, 1181, 1146 cm⁻¹.



Prepared according to the above procedure; white solid (111 mg, 75%). TLC $R_f = 0.48$ (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 9.44 (s, 1H), 8.07 (d, J = 8.2 Hz, 2H), 8.01 (dd, J = 8.2, 1.7 Hz, 1H), 7.55 (d, J = 8.2 Hz, 2H), 7.32 (d, J = 9.0 Hz, 1H), 7.31–7.28 (m, 1H), 6.82 (d, J = 9.0 Hz, 1H), 6.78 (ddd, J = 8.2, 7.2, 1.1 Hz, 1H), 6.37 (d, J = 8.3 Hz, 1H), 5.24 (s, 2H), 4.41 (q, J = 7.1 Hz, 2H), 3.93 (s, 3H), 1.43 (t, J = 7.1 Hz, 3H) ppm; IR (thin film) v 3299, 2951, 1722, 1683, 1578, 1508, 1451, 1280, 1251, 1109 cm⁻¹.



Prepared according to the above procedure. Reaction mixture was diluted with 1:4 MeOH/CH₂Cl₂ instead of EtOAc before filtering through Celite due to limited compound solubility in less polar solvents; white solid (33 mg, 24%). TLC R_f = 0.17 (49:1 CH₂Cl₂/MeOH); ¹H NMR (CDCl₃, 500 MHz) δ 9.42 (s, 1H), 8.01 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.61 (s, 1H), 7.49 (d, *J* = 8.0 Hz, 1H), 7.38–7.27 (m, 3H), 7.21 (d, *J* = 6.9 Hz, 2H), 6.83 (d, *J* = 9.0 Hz, 1H), 6.78 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1H), 6.38 (dd, *J* = 8.5, 1.0 Hz, 1H), 5.17 (s, 2H), 4.40 (q, *J* = 7.1 Hz, 2H), 2.19 (s, 3H), 1.43 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3302, 1678, 1561, 1487, 1451, 1298, 1249 cm⁻¹.



Prepared according to the above procedure with 15 mol % catalyst. Reaction mixture was diluted with 1:4 MeOH/CH₂Cl₂ instead of EtOAc before filtering through Celite due to limited compound solubility in less polar solvents; white solid (59 mg, 50%). TLC R_f = 0.22 (49:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 11.74 (s, 1H), 8.83 (d, *J* = 8.4 Hz, 1H), 8.10 (d, *J* = 8.0 Hz, 1H), 7.85 (d, *J* = 7.8 Hz, 1H), 7.78 (d, *J* = 7.6 Hz, 1H), 7.60 (t, *J* = 7.9 Hz, 1H), 7.55 (t, *J* = 7.7 Hz, 1H), 7.46 (t, *J* = 7.5 Hz, 1H), 7.15 (t, *J* = 7.8 Hz, 1H), 7.05 (d, *J* = 8.9 Hz, 1H), 6.41 (d, *J* = 8.9 Hz, 1H), 5.52 (s, 2H), 4.46 (s, 2H), 4.38 (q, *J* = 7.1 Hz, 2H), 1.41 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3299, 1676, 1607, 1531, 1509, 1474, 1448, 1313, 1251 cm⁻¹.



Prepared according to the above procedure with 15 mol % catalyst. Reaction mixture was diluted with 1:4 MeOH/CH₂Cl₂ instead of EtOAc before filtering through Celite due to limited compound solubility in less polar solvents; white solid (58 mg, 49%). TLC R_f = 0.16 (9:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 12.13 (s, 1H), 8.93 (d, *J* = 8.5 Hz, 1H), 8.14–8.07 (m, 2H), 7.99 (d, *J* = 7.9 Hz, 1H), 7.67 (d, *J* = 7.8 Hz, 1H), 7.61 (t, *J* = 7.8 Hz, 1H), 7.55 (t, *J* = 7.7 Hz, 1H), 7.13 (t, *J* = 7.6 Hz, 1H), 7.08 (d, *J* = 8.9 Hz, 1H), 6.36 (d, *J* = 8.9 Hz, 1H), 5.21 (s, 2H), 4.52 (s, 2H), 4.42 (q, *J* = 7.1 Hz, 2H), 1.43 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3377, 1675, 1608, 1589, 1531, 1475, 1449, 1308, 1254 cm⁻¹.



Prepared according to the above procedure. Reaction mixture was diluted with 1:4 MeOH/CH₂Cl₂ instead of EtOAc before filtering through Celite due to limited compound solubility in less polar solvents. In the filtration step, the ethyl ester transesterified to the methyl ester; white solid (8 mg, 7%). TLC R_f = 0.52 (49:1 CH₂Cl₂/MeOH); ¹H NMR (DMSO-d₆, 500 MHz) δ 11.60 (s, 1H), 8.55 (d, *J* = 8.2 Hz, 1H), 8.02 (dd, *J* = 7.9, 1.7 Hz, 1H), 7.99 (d, *J* = 8.1 Hz, 2H), 7.72–7.64 (m, 3H), 7.26 (t, *J* = 7.7 Hz, 1H), 7.18 (d, *J* = 8.9 Hz, 1H), 6.49 (d, *J* = 8.9 Hz, 1H), 5.53 (s, 2H), 5.27 (s, 2H), 3.90 (s, 3H) ppm; IR (thin film) v 3309, 2920, 1679, 1609, 1590, 1535, 1477, 1450, 1386, 1312, 1251 cm⁻¹.



Prepared according to the above procedure; colorless oil (109 mg, 97%). TLC R_f = 0.58 (3:2 hexanes/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 9.34 (s, 1H), 7.98 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.85 (dd, *J* = 5.5, 3.0 Hz, 2H), 7.71 (dd, *J* = 5.5, 3.0 Hz, 2H), 7.31 (d, *J* = 8.9 Hz, 1H), 7.26–7.20 (m, 1H), 6.82 (d, *J* = 9.0 Hz, 1H), 6.75 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1H), 6.32 (d, *J* = 8.4 Hz, 1H), 4.37 (q, *J* = 7.1 Hz, 2H), 4.31 (t, *J* = 5.6 Hz, 2H), 4.18–4.17 (m, 2H), 1.41 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3299, 2982, 1775, 1715, 1683, 1579, 1507, 1453, 1394, 1297, 1251 cm⁻¹.
MOM group deprotection



A flame-dried 5-mL round-bottom flask was charged with ethyl 2-((2,6-dichloro-3-(methoxy-methoxy)phenyl)amino)benzoate (25.5 mg, 0.069 mmol). The flask was sealed with a septum, and the headspace flushed with N₂, and 2.0 mL of anhydrous CH₂Cl₂ was added. Following dissolution of the starting material, the flask was placed in an ice bath, and bromotrimethylsilane (0.05 mL, 0.379 mmol, 5.5 equiv) was added dropwise via syringe. The mixture was stirred at 0 °C for 1 h, following which time the reaction was warmed to ambient temperature and stirred for an additional 3 h. The reaction was quenched by the addition of 1 mL of H₂O and transferred to a separatory funnel with 20 mL of EtOAc. The organic fraction was washed with 2 x 20 mL of H₂O and 1 x 20 mL of saturated aqueous NaCl, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a white solid (19 mg, 85%). This material was judged to be sufficiently pure by ¹H NMR for use in the subsequent step without further purification. TLC R_f = 0.16 (9:1 hexanes/acetone); ¹H NMR (CDCl₃, 400 MHz) δ 9.40 (s, 1H), 8.01 (d, *J* = 8.1 Hz, 1H), 7.36–7.22 (m, 2H), 6.94 (d, *J* = 8.9 Hz, 1H), 6.79 (t, *J* = 7.5 Hz, 1H), 6.37 (d, *J* = 8.5 Hz, 1H), 5.61 (s, 1H), 4.40 (q, *J* = 7.1 Hz, 2H), 1.43 (t, *J* = 7.1 Hz, 3H) ppm; IR (thin film) v 3308, 2926, 1683, 1580, 1506, 1452, 1253 cm⁻¹.

Ester hydrolysis



AK-21. A screw-capped vial was charged with ethyl 2-((2,6-diethylphenyl)amino)benzoate (180 mg, 0.6 mmol) and 3.0 mL of a 1.0 M solution of NaOH in 1:1:1 mixture of H₂O/EtOH/THF. The reaction was stirred until thin-layer chromatography indicated the complete consumption of starting material (~24 h). Following this time, the solution was acidified to pH 2 with 2–3 drops of concentrated aqueous HCl, which resulted in the immediate precipitation of a white solid. The solid product was isolated by vacuum filtration through a small Büchner funnel and purified by reversed-phase HPLC (Alltima C18, 10 µM, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30–100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 23.3–25.2 min = 86–88% MeCN); white solid (32 mg, 20%); ¹H NMR (acetone-d₆, 500 MHz) δ 9.32 (s, 1H), 7.99 (dd, *J* = 8.1, 1.7 Hz, 1H), 7.29–7.17 (m, 4H), 6.65 (t, *J* = 7.5 Hz, 1H), 6.16 (d, *J* = 8.5 Hz, 1H), 2.54 (dq, *J* = 14.6, 7.2 Hz, 4H), 1.10 (t, *J* = 7.6 Hz, 6H) ppm. LRMS (ES⁻) calcd 268.13 for C₁₇H₁₈NO₂⁻ found 267.7 (M⁻).

AK-1. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1→1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 30.6–31.6 min); white solid (5 mg, 26%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.87 (s, 1H), 7.96 (d, *J* = 8.0 Hz, 1H), 7.70 (d, *J* = 2.5 Hz, 1H), 7.57 (d, *J* = 8.7 Hz, 1H), 7.49–7.44 (m, 1H), 7.39 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.23 (d, *J* = 8.4 Hz, 1H), 6.92 (t, *J* = 7.4 Hz, 1H) ppm; LRMS (ES⁻) calcd 279.99 for C₁₃H₈Cl₂NO₂⁻ found 280.0 (M⁻).



AK-2. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1→1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 30.8–31.9 min); white solid (24 mg, 63%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.64 (s, 1H), 7.93 (d, *J* = 7.8 Hz, 1H), 7.53 (d, *J* = 8.7 Hz, 1H), 7.48–7.44 (m, 2H), 7.30 (d, *J* = 8.4 Hz, 1H), 7.23 (dd, *J* = 9.0, 2.4 Hz, 1H), 6.90 (t, *J* = 7.6 Hz, 1H) ppm; LRMS (ES⁻) calcd 279.99 for C₁₃H₈Cl₂NO₂⁻ found 280.0 (M⁻).



AK-3. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1 \rightarrow 1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 29.5–30.7 min); white solid (4 mg, yield not determined); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.83 (s, 1H), 7.93 (d, *J* = 7.9 Hz, 1H), 7.43 (t, *J* = 7.8 Hz, 1H), 7.38 (d, *J* = 8.0 Hz, 1H), 7.22 (t, *J* = 7.8 Hz, 1H), 7.19 (d, *J* = 8.5 Hz, 1H), 7.06 (d, *J* = 7.4 Hz, 1H), 6.86 (t, *J* = 7.5 Hz, 1H), 2.37 (s, 3H) ppm; LRMS (ES⁻) calcd 260.05 for C₁₄H₁₁CINO₂⁻ found 260.0 (M⁻).



AK-4. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1 \rightarrow 1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 27.8–28.9 min); white solid (12 mg, 49%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.51 (s, 1H), 7.90 (d, *J* = 7.6 Hz, 1H), 7.63 (d, *J* = 8.1 Hz, 2H), 7.37 (t, *J* = 8.1 Hz, 1H), 7.33 (t, *J* = 7.8 Hz, 1H), 6.79 (t, *J* = 7.4 Hz, 1H), 6.22 (d, *J* = 8.1 Hz, 1H) ppm; LRMS (ES⁻) calcd 279.99 for C₁₃H₈Cl₂NO₂⁻ found 280.0 (M⁻).

CI H CO₂H

AK-5. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1 \rightarrow 1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 30.3–32.0 min); white solid (21 mg, 38%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.90 (s, 1H), 7.96 (d, *J* = 8.0 Hz, 1H), 7.58–7.48 (m, 3H), 7.32 (d, *J* = 8.4 Hz, 1H), 7.10 (dd, *J* = 8.7, 2.5 Hz, 1H), 6.96 (t, *J* = 7.2 Hz, 1H) ppm; LRMS (ES⁻) calcd 279.99 for C₁₃H₈Cl₂NO₂⁻ found 280.0 (M⁻).



AK-6. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1→1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 29.4–30.6 min); white solid (23 mg, 48%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.77 (s, 1H), 7.93 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.45 (ddd, *J* = 8.7, 7.1, 1.7 Hz, 1H), 7.41 (d, *J* = 8.1 Hz, 1H), 7.37 (d, *J* = 2.1 Hz, 1H), 7.21 (dd, *J* = 8.5, 1.0 Hz, 1H), 6.91 (dd, *J* = 8.2, 2.1 Hz, 1H), 6.87 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1H), 2.29 (s, 3H) ppm; LRMS (ES⁻) calcd 260.05 for C₁₄H₁₁CINO₂⁻ found 260.0 (M⁻).

AK-7. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1 \rightarrow 1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 30.7–31.6 min); white solid (9 mg, 17%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.48 (s, 1H), 7.89 (dd, *J* = 7.9, 1.7 Hz, 1H), 7.82 (s, 2H), 7.32 (ddd, *J* = 8.6, 7.1, 1.7 Hz, 1H), 6.79 (t, *J* = 7.7 Hz, 1H), 6.24 (d, *J* = 8.4 Hz, 1H) ppm; LRMS (ES⁻) calcd 313.95 for C₁₃H₇Cl₃NO₂⁻ found 315.9 (M⁻).



AK-8. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1 \rightarrow 1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 26.0–27.6 min); white solid (32 mg, 31%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.31 (s, 1H), 7.90 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.41–7.31 (m, 2H), 7.25 (t, *J* = 8.3 Hz,

2H), 6.81 (t, *J* = 7.6 Hz, 1H), 6.49 (d, *J* = 8.2 Hz, 1H) ppm; LRMS (ES⁻) calcd 248.05 for C₁₃H₈F₂NO₂⁻ found 248.0 (M⁻).

AK-10. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1 \rightarrow 1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 30.4–30.6 min); white solid (10 mg, 37%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.51 (s, 1H), 7.78 (d, *J* = 8.1 Hz, 1H), 7.51 (d, *J* = 8.3 Hz, 1H), 7.35 (d, *J* = 8.3 Hz, 1H), 6.60 (dd, *J* = 8.0, 1.1 Hz, 1H), 5.99 (s, 1H), 2.38 (s, 3H), 2.15 (s, 3H) ppm; LRMS (ES⁻) calcd 308.03 for C₁₅H₁₂Cl₂NO₂⁻ found 308.1 (M⁻).



AK-11. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1 \rightarrow 1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 30.8–32.0 min); white solid (7 mg, 24%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.37 (s, 1H), 7.70 (d, *J* = 2.1 Hz, 1H), 7.50 (d, *J* = 8.3 Hz, 1H), 7.33 (d, *J* = 8.4 Hz, 1H), 7.13 (dd, *J* = 8.2, 2.1 Hz, 1H), 6.54 (s, 1H), 6.12 (d, *J* = 8.5 Hz, 1H), 2.37 (s, 3H), 2.21 (s, 3H) ppm; LRMS (ES⁻) calcd 308.03 for C₁₅H₁₂Cl₂NO₂⁻ found 308.0 (M⁻).



AK-12. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1 \rightarrow 1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 29.9–30.6 min); white solid (2 mg, 13%); ¹H NMR (DMSO-d₆, 600 MHz) δ 8.33 (s, 1H), 7.47 (d, *J* = 8.3 Hz, 1H), 7.29 (d, *J* = 8.3 Hz, 1H), 7.09 (t, *J* = 7.9 Hz, 1H), 6.67 (d, *J* = 7.5 Hz, 1H), 6.53 (s, 1H), 6.05 (d, *J* = 8.3 Hz, 1H), 2.42 (s, 3H), 2.37 (s, 3H) ppm; LRMS (ES⁻) calcd 308.03 for C₁₅H₁₂Cl₂NO₂⁻ found 308.1 (M⁻).



AK-13. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1 \rightarrow 1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 27.6–28.2 min); white solid (2 mg, 24%); ¹H NMR

(DMSO-d₆, 600 MHz) δ 9.50 (s, 1H), 7.89 (dd, *J* = 7.8, 1.6 Hz, 1H), 7.57 (d, *J* = 9.1 Hz, 1H), 7.35–7.30 (m, 1H), 7.17 (d, *J* = 9.1 Hz, 1H), 6.78 (t, *J* = 7.7 Hz, 1H), 6.23 (d, *J* = 8.5 Hz, 1H), 3.91 (s, 3H) ppm; LRMS (ES⁻) calcd 310.00 for C₁₄H₁₀Cl₂NO₃⁻ found 310.1 (M⁻).

AK-14. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1 \rightarrow 1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 31.0–31.5 min); white solid (1 mg, 7%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.56 (s, 1H), 7.90 (dd, *J* = 7.9, 1.6 Hz, 1H), 7.59 (dd, *J* = 8.4, 0.9 Hz, 1H), 7.34 (t, *J* = 7.6 Hz, 1H), 7.26 (d, *J* = 8.3 Hz, 1H), 6.78 (t, *J* = 7.5 Hz, 1H), 6.24 (d, *J* = 8.4 Hz, 1H), 5.31 (t, *J* = 1.7 Hz, 1H), 5.01 (s, 1H), 2.07 (s, 3H) ppm. LRMS (ES⁻) calcd 320.03 for C₁₆H₁₂Cl₂NO₂⁻ found 320.0 (M⁻).



AK-15. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1 \rightarrow 1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 29.5–30.2 min); white solid (1 mg, 2%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.16 (s, 1H), 7.86 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.25 (t, *J* = 7.5 Hz, 1H), 7.20–7.12 (m, 3H), 6.64 (t, *J* = 7.5 Hz, 1H), 6.53 (s, 1H), 6.07 (d, *J* = 8.5 Hz, 1H), 2.12 (s, 6H) ppm; LRMS (ES⁻) calcd 240.11 for C₁₅H₁₄NO₂⁻ found 240.2 (M⁻).



AK-16. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1→1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 30.8–32.2 min); white solid (32 mg, 39%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.53 (s, 1H), 7.90 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.55 (d, *J* = 8.4 Hz, 1H), 7.35 (d, *J* = 8.4 Hz, 1H), 7.34–7.30 (m, 1H), 6.77 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1H), 6.19 (dd, *J* = 8.4, 1.1 Hz, 1H), 2.75 (q, *J* = 7.4 Hz, 2H), 1.20 (t, *J* = 7.5 Hz, 3H) ppm; LRMS (ES⁻) calcd 308.03 for C₁₅H₁₂Cl₂NO₂⁻ found 308.0 (M⁻).



AK-17. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow over 30 min of 0:1 \rightarrow 1:0 MeCN/0.1% TFA in H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 33.2–34.2 min); white solid (2 mg, 9%); ¹H NMR (acetonitrile-d₃, 500 MHz) δ 9.50 (s, 1H), 7.99 (dd, *J* = 7.9, 1.6 Hz, 1H), 7.77–7.63 (m, 2H), 7.34 (ddd, *J* = 8.6, 7.2, 1.7 Hz, 1H), 6.84 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1H), 6.30 (dd, *J* = 8.5, 1.1 Hz, 1H) ppm; LRMS (ES⁻) calcd 347.98 for C₁₄H₇Cl₂F₃NO₂⁻ found 348.0 (M⁻).



AK-19. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 17.1–17.6 min = 79% MeCN); white solid (32 mg, 29%); ¹H NMR (DMSO-d₆, 500 MHz) δ 8.11 (s, 1H), 7.47 (d, *J* = 8.3 Hz, 1H), 7.31–7.27 (m, 2H), 7.24 (t, *J* = 7.8 Hz, 1H), 7.08 (s, 1H), 6.74 (dt, *J* = 8.0, 1.2 Hz, 1H), 2.37 (s, 3H) ppm. LRMS (ES⁻) calcd 294.01 for C₁₄H₁₀Cl₂NO₂⁻ found 293.6 (M⁻).



AK-20. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 16.7–17.8 min = 78–79% MeCN); white solid (12 mg, 9%); ¹H NMR (acetone-d₆, 500 MHz) δ 7.86 (d, *J* = 8.7 Hz, 2H), 7.69 (s, 1H), 7.45 (d, *J* = 8.3 Hz, 1H), 7.32 (d, *J* = 8.3 Hz, 1H), 6.66 (d, *J* = 8.7 Hz, 2H), 2.41 (s, 3H) ppm. LRMS (ES⁻) calcd 294.01 for C₁₄H₁₀Cl₂NO₂⁻ found 293.5 (M⁻).



AK-22. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100→70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30→100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 21.6–23.2 min = 84–86% MeCN); white solid (55 mg, 45%); ¹H NMR (DMSO-d₆, 500 MHz) δ 9.71 (s, 1H), 7.97 (t, *J* = 7.8 Hz, 1H), 7.53 (d, *J* = 8.3 Hz, 1H), 7.38 (d, *J* = 8.3 Hz, 1H), 6.59 (td, *J* = 8.5, 2.3 Hz, 1H), 5.87 (dd, *J* = 11.7, 2.1 Hz, 1H), 2.38 (s, 3H) ppm. LRMS (ES⁻) calcd 312.00 for C₁₄H₉Cl₂FNO₂⁻ found 311.6 (M⁻).



AK-23. Prepared according to the above procedure. The solution containing the sodium carboxylate salt was not acidified prior to isolation of the impure material to avoid MOM group deprotection. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:1 MeCN/H₂O over 5 min, then 0:1 \rightarrow 1:0 MeCN/H₂O over 30 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 19.8–20.1 min = 49–50% MeCN); white solid (10 mg, 9%); ¹H NMR (DMSO-d₆, 500 MHz) δ 7.86 (dd, *J* = 7.6, 1.8 Hz, 1H), 7.46 (d, *J* = 9.0 Hz, 1H), 7.10 (d, *J* = 9.0 Hz, 1H), 7.02 (td, *J* = 7.5, 1.9 Hz, 1H), 6.60 (td, *J* = 7.4, 1.1 Hz, 1H), 6.09 (dd, *J* = 8.2, 1.1 Hz, 1H), 5.31 (s, 2H), 3.42 (s, 3H) ppm. LRMS (ES⁻) calcd 340.01 for C₁₅H₁₂Cl₂NO₄⁻ found 339.6 (M⁻).

AK-24. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 22.2–24.0 min = 85–87% MeCN); white solid (13 mg, 32%); ¹H NMR (DMSO-d₆, 500 MHz) δ 9.52 (s, 1H), 7.90 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.56 (d, *J* = 9.0 Hz, 1H), 7.50–7.47 (m, 2H), 7.42 (t, *J* = 7.4 Hz, 2H), 7.37–7.30 (m, 2H), 7.26 (d, *J* = 9.1 Hz, 1H), 6.78 (ddd, *J* = 8.0, 7.1, 1.1 Hz, 1H), 6.23 (dd, *J* = 8.4, 1.0 Hz, 1H), 5.26 (s, 2H) ppm. LRMS (ES⁻) calcd 386.04 for C₂₀H₁₄Cl₂NO₃⁻ found 385.7 (M⁻).



AK-25. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 23.3–24.5 min = 86–87% MeCN); white solid (1 mg, 18%); ¹H NMR (acetone-d₆, 500 MHz) δ 9.69 (s, 1H), 8.05 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.64 (d, *J* = 8.3 Hz, 1H), 7.51–7.36 (m, 7H), 6.83 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1H), 6.45 (dd, *J* = 8.4, 1.0 Hz, 1H) ppm. LRMS (ES⁻) calcd 356.03 for C₁₉H₁₂Cl₂NO₂⁻ found 355.6 (M⁻).

AK-26. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100→70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30→100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 30.2–31.6 min = 94–96% MeCN); white solid (1 mg, 9%); ¹H NMR (acetone-d₆, 500 MHz) δ 9.57 (s, 1H), 8.03 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.48 (d, *J* = 9.0 Hz, 1H), 7.35 (td, *J* = 8.6, 7.1, 1.5 Hz, 1H), 7.12 (d, *J* = 9.0 Hz, 1H), 6.81 (td, *J* = 8.1, 7.1, 1.1 Hz, 1H), 6.35 (dd, *J* = 8.6, 1.1 Hz, 1H), 3.95 (d, *J* = 6.2 Hz, 2H), 1.95–1.82 (m, 3H), 1.80–1.66 (m, 3H), 1.32 (tt, *J* = 12.4, 3.2 Hz, 2H), 1.23 (tt, *J* = 12.4, 3.1 Hz, 1H), 1.15 (qd, *J* = 12.3, 3.4 Hz, 2H) ppm. LRMS (ES⁻) calcd 392.08 for C₂₀H₂₀Cl₂NO₃⁻ found 391.8 (M⁻).



AK-27. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100→70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30→100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 22.2–23.2 min = 85–86% MeCN); white solid (3 mg, 12%); ¹H NMR (acetone-d₆, 500 MHz) δ 9.70 (s, 1H), 8.04 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.60 (d, *J* = 8.4 Hz, 1H), 7.42 (ddd, *J* = 8.3, 7.4, 1.7 Hz, 1H), 7.39 (s, 1H), 7.28 (d, *J* = 8.3 Hz, 1H), 7.23 (dd, *J* = 7.5, 1.8 Hz, 1H), 7.12 (dd, *J* = 8.3, 1.0 Hz, 1H), 7.04 (td, *J* = 7.5, 1.0 Hz, 1H), 6.82 (t, *J* = 7.6 Hz, 1H), 6.47–6.41 (m, 1H), 3.80 (s, 3H) ppm. LRMS (ES⁻) calcd 386.04 for C₂₀H₁₄Cl₂NO₃⁻ found 385.7 (M⁻).



AK-28. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100→70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30→100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 15.4–16.0 min = 76–77% MeCN); white solid (4 mg, 23%); ¹H NMR (acetone-d₆, 600 MHz) δ 9.55 (s, 1H), 8.03 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.41–7.33 (m, 2H), 7.05 (d, *J* = 8.9 Hz, 1H), 6.80 (ddd, *J* = 8.2, 7.1, 1.1 Hz, 1H), 6.37 (ddd, *J* = 8.5, 3.3, 1.0 Hz, 1H) ppm. LRMS (ES⁻) calcd 295.99 for C₁₃H₈Cl₂NO₃⁻ found 295.6 (M⁻).

AK-29. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100→70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30→100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 29.0–30.0 min = 93–94% MeCN); white solid (3 mg, 28%); ¹H NMR (acetone-d₆, 600 MHz) δ 9.60 (s, 1H), 8.04 (dd, *J* = 8.1, 1.7 Hz, 1H), 7.50 (d, *J* = 8.3 Hz, 1H), 7.35 (ddd, *J* = 8.6, 7.1, 1.7 Hz, 1H), 7.19 (d, *J* = 8.3 Hz, 1H), 6.81 (ddd, *J* = 8.2, 7.1, 1.2 Hz, 1H), 6.37–6.30 (m, 1H), 5.71 (tt, *J* = 3.7, 1.8 Hz, 1H), 2.32–2.29 (m, 2H), 2.20–2.16 (m, 2H), 1.80–1.73 (m, 2H), 1.72–1.64 (m, 2H) ppm. LRMS (ES⁻) calcd 360.06 for C₁₉H₁₆Cl₂NO₂⁻ found 359.7 (M⁻).



AK-30. Prepared according to the above procedure. The solution containing the sodium carboxylate salt was not acidified prior to isolation of the impure material. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:1 MeCN/H₂O over 5 min, then 0:1 \rightarrow 1:0 MeCN/H₂O over 30 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 19.8–22.0 min = 49–56% MeCN); white solid (35 mg, 79%); ¹H NMR (methanol-d₄, 500 MHz) δ 7.96 (dd, *J* = 4.6, 1.3 Hz, 1H), 7.39 (d, *J* = 8.3 Hz, 1H), 7.27 (dd, *J* = 8.5, 4.6 Hz, 1H), 7.24 (dd, *J* = 8.3, 0.7 Hz, 1H), 6.77 (dd, *J* = 8.5, 1.3 Hz, 1H), 2.41 (s, 3H) ppm. LRMS (ES⁻) calcd 295.00 for C₁₃H₉Cl₂N₂O₂⁻ found 294.6 (M⁻).



AK-31. Prepared according to the above procedure. The solution containing the sodium carboxylate salt was not acidified prior to isolation of the impure material. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:1 MeCN/H₂O over 5 min, then 0:1 \rightarrow 1:0 MeCN/H₂O over 30 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 19.6–20.8 min = 49–56% MeCN); white solid (7 mg, 90%); ¹H NMR (DMSO-d₆, 500 MHz) δ 8.74 (s, 1H), 8.13 (d, *J* = 6.6 Hz, 1H), 7.56 (d, *J* = 8.4 Hz, 1H), 7.44 (d, *J* = 8.4 Hz, 1H), 6.27 (d, *J* = 6.7 Hz, 1H), 2.38 (s, 3H) ppm. LRMS (ES⁻) calcd 295.00 for C₁₃H₉Cl₂N₂O₂⁻ found 294.6 (M⁻).

AK-32. Prepared according to the above procedure. The solution containing the sodium carboxylate salt was not acidified prior to isolation of the impure material. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:1 MeCN/H₂O over 5 min, then 0:1 \rightarrow 1:0 MeCN/H₂O over 30 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 16.0–19.2 min = 36–47% MeCN); white solid (40 mg, 81%); ¹H NMR (methanol-d₄, 500 MHz) δ 7.89 (d, *J* = 5.0 Hz, 1H), 7.81 (d, *J* = 5.1 Hz, 1H), 7.48 (s, 1H), 7.39 (d, *J* = 8.3 Hz, 1H), 7.22 (d, *J* = 8.3 Hz, 1H), 2.42 (s, 3H) ppm. LRMS (ES⁻) calcd 295.00 for C₁₃H₉Cl₂N₂O₂⁻ found 294.6 (M⁻).



AK-33. Prepared according to the above procedure. The solution containing the sodium carboxylate salt was not acidified prior to isolation of the impure material. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:1 MeCN/H₂O over 5 min, then 0:1 \rightarrow 1:0 MeCN/H₂O over 30 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 14.4–18.2 min = 31–44% MeCN); white solid (70 mg, 99%); ¹H NMR (methanol-d₄, 500 MHz) δ 8.27 (dd, *J* = 7.5, 1.9 Hz, 1H), 7.89 (dd, *J* = 5.0, 2.0 Hz, 1H), 7.33 (d, *J* = 8.3 Hz, 1H), 7.18 (d, *J* = 8.3 Hz, 1H), 6.70 (dd, *J* = 7.5, 5.0 Hz, 1H), 2.40 (s, 3H) ppm. LRMS (ES⁻) calcd 295.00 for C₁₃H₉Cl₂N₂O₂⁻ found 294.6 (M⁻).



AK-37. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100→70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30→100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 21.2–22.2 min = 71–73% MeCN); white solid (22 mg, yield not determined); ¹H NMR (DMSO-d₆, 600 MHz) δ 8.70 (d, *J* = 8.4 Hz, 1H), 8.06 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.99 (d, *J* = 8.2 Hz, 2H), 7.65 (d, *J* = 7.9 Hz, 3H), 7.20 (t, *J* = 7.5 Hz, 1H), 7.18 (d, *J* = 8.9 Hz, 1H), 6.49 (d, *J* = 8.9 Hz, 1H), 5.50 (s, 2H), 5.27 (s, 2H) ppm; LRMS (ES⁻) calcd 429.04 for C₂₁H₁₅Cl₂N₂O₄⁻ found 429.1 (M⁻).



AK-38. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 12.0–15.2 min = 72–76% MeCN); white solid (22 mg, 65%); ¹H NMR (DMSO-d₆, 500 MHz) δ 9.53 (s, 1H), 7.98 (d, *J* = 8.2 Hz, 2H), 7.90 (dd, *J* = 8.0, 1.8 Hz, 1H), 7.60 (d, *J* = 8.3 Hz, 2H), 7.57 (d, *J* = 9.0 Hz, 1H), 7.33 (t, *J* = 7.4 Hz, 1H), 7.24 (d, *J* = 9.2 Hz, 1H), 6.79 (t, *J* = 7.4 Hz, 1H), 6.24 (d, *J* = 8.4 Hz, 1H), 5.36 (s, 2H) ppm; LRMS (ES⁻) calcd 430.03 for C₂₁H₁₄Cl₂NO₅⁻ found 430.0 (M⁻).



AK-39. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 19.4–20.2 min = 81–82% MeCN); white solid (21 mg, 38%); ¹H NMR (DMSO-d₆, 600 MHz) δ 11.77 (s, 1H), 8.57 (d, *J* = 8.4 Hz, 1H), 8.02 (dd, *J* = 7.9, 1.7 Hz, 1H), 7.76 (d, *J* = 7.7 Hz, 1H), 7.70 (d, *J* = 7.7 Hz, 1H), 7.64–7.59 (m, 2H), 7.53 (t, *J* = 7.5 Hz, 1H), 7.20 (t, *J* = 7.7 Hz, 1H), 7.13 (d, *J* = 8.9 Hz, 1H), 6.43 (d, *J* = 8.9 Hz, 1H), 5.40 (s, 4H) ppm; LRMS (ES⁻) calcd 429.04 for C₂₁H₁₅Cl₂N₂O₄⁻⁻ found 429.1 (M⁻).



AK-40. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100→70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30→100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 20.0–20.8 min = 82–83% MeCN); white solid (41 mg, 75%); ¹H NMR (DMSO-d₆, 600 MHz) δ 12.24 (s, 1H), 8.70 (d, *J* = 8.3 Hz, 1H), 8.07–8.05 (m, 2H), 7.91 (d, *J* = 7.8 Hz, 1H), 7.71 (d, *J* = 7.7 Hz, 1H), 7.67 (t, *J* = 7.7 Hz, 1H), 7.63 (t, *J* = 7.7 Hz, 1H), 7.22 (t, *J* = 7.6 Hz, 1H), 7.18 (d, *J* = 8.9 Hz, 1H), 6.51 (d, *J* = 8.9 Hz, 1H), 5.49 (s, 2H), 5.26 (s, 2H) ppm; LRMS (ES⁻) calcd 429.04 for C₂₁H₁₅Cl₂N₂O₄⁻ found 429.1 (M⁻).



AK-41. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100→70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30→100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 14.2–15.0 min = 75–76% MeCN); white solid (10 mg, 33%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.99 (s, 1H), 9.54 (s, 1H), 7.90 (d, *J* = 8.0 Hz, 1H), 7.70 (s, 1H), 7.56–7.54 (m, 2H), 7.32 (t, *J* = 7.7 Hz, 2H), 7.23 (d, *J* = 9.1 Hz, 1H), 7.14 (d, *J* = 7.5 Hz, 1H), 6.78 (t, *J* = 7.5 Hz, 1H), 6.24 (d, *J* = 8.4 Hz, 1H), 5.23 (s, 2H), 2.04 (s, 3H) ppm; LRMS (ES⁻) calcd 443.06 for C₂₂H₁₇Cl₂N₂O₄⁻ found 443.1 (M⁻).



AK-42. Prepared according to the above procedure. The solution containing the sodium carboxylate salt was not acidified prior to isolation of the impure material. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/ H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/H₂O over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 12.2–13.7 min = 73–75% MeCN); white solid (11 mg, 78%); ¹H NMR (DMSO-d₆, 600 MHz) δ 8.07 (dd, *J* = 7.3, 2.1 Hz, 1H), 7.88 (d, *J* = 3.4 Hz, 1H), 7.49 (d, *J* = 7.5 Hz, 2H), 7.43–7.40 (m, 3H), 7.35 (t, *J* = 7.3 Hz, 1H), 7.11 (d, *J* = 9.0 Hz, 1H), 6.62 (dd, *J* = 7.4, 4.8 Hz, 1H), 5.23 (s, 2H) ppm; LRMS (ES⁻) calcd 387.03 for C₁₉H₁₃Cl₂N₂O₃⁻ found 387.0 (M⁻).



AK-43. Prepared according to the above procedure. The solution containing the sodium carboxylate salt was not acidified prior to isolation of the impure material. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/ H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/H₂O over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 11.2–14.2 min = 71–75% MeCN); white solid (6 mg, 37%); ¹H NMR (DMSO-d₆, 600 MHz) δ 8.04 (s, 1H), 7.89 (d, *J* =

7.8 Hz, 1H), 7.84 (d, J = 7.7 Hz, 1H), 7.64 (d, J = 7.3 Hz, 1H), 7.52–7.43 (m, 2H), 7.11 (d, J = 8.7 Hz, 1H), 7.02 (t, J = 6.6 Hz, 1H), 6.59 (t, J = 7.4 Hz, 1H), 6.09 (d, J = 8.1 Hz, 1H), 5.30 (s, 2H) ppm; LRMS (ES⁻) calcd 430.03 for C₂₁H₁₄Cl₂NO₅⁻ found 430.0 (M⁻).



AK-44. Prepared according to the above procedure. The solution containing the sodium carboxylate salt was not acidified prior to isolation of the impure material. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/ H₂O over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 10.8–12.2 min = 71–73% MeCN); white solid (9 mg, 40%); ¹H NMR (DMSO-d₆, 600 MHz) δ 7.86 (dd, *J* = 7.7, 1.8 Hz, 1H), 7.83 (d, *J* = 7.6 Hz, 1H), 7.58 (d, *J* = 7.7 Hz, 1H), 7.44 (d, *J* = 9.0 Hz, 1H), 7.41 (t, *J* = 7.8 Hz, 1H), 7.30 (t, *J* = 7.4 Hz, 1H), 7.03 (t, *J* = 7.6 Hz, 1H), 7.00 (d, *J* = 9.1 Hz, 1H), 6.60 (t, *J* = 7.5 Hz, 1H), 6.12 (d, *J* = 8.1 Hz, 1H), 5.65 (d, *J* = 8.1 Hz, 2H) ppm; LRMS (ES⁻) calcd 430.03 for C₂₁H₁₄Cl₂NO₅⁻ found 430.1 (M⁻).



AK-45. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100→70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30→100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 21.3–22.5 min = 84–85% MeCN); white solid (5 mg, 6%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.48 (s, 1H), 7.85 (dd, *J* = 7.9, 1.7 Hz, 1H), 7.58–7.52 (m, 2H), 7.43–7.39 (m, 1H), 7.30–7.21 (m, 4H), 6.74 (t, *J* = 7.8, 1H), 6.19 (d, *J* = 8.4 Hz, 1H), 5.26 (s, 2H) ppm; ¹⁹F NMR (acetone-d₆, 376 MHz) δ –120.18 (dt, *J* = 10.8, 7.0 Hz, 1F) ppm; LRMS (ES⁻) calcd 404.03 for C₂₀H₁₃Cl₂FNO₃⁻ found 404.1 (M⁻).



AK-46. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T

= 23.7–28.9 min = 86–93% MeCN); white solid (47 mg, 58%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.51 (s, 1H), 7.89 (d, J = 8.1 Hz, 1H), 7.56 (d, J = 9.1 Hz, 1H), 7.35–7.22 (m, 5H), 7.17 (d, J = 7.4 Hz, 1H), 6.78 (t, J = 7.6 Hz, 1H), 6.23 (d, J = 8.3 Hz, 1H), 5.22 (s, 2H), 2.33 (s, 3H) ppm; LRMS (ES⁻) calcd 400.05 for C₂₁H₁₆Cl₂NO₃⁻ found 400.0 (M⁻).



AK-47. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100→70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30→100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 23.2–29.2 min = 86–93% MeCN); white solid (53 mg, 69%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.52 (s, 1H), 7.89 (d, *J* = 8.0 Hz, 1H), 7.58 (d, *J* = 9.0 Hz, 1H), 7.47 (d, *J* = 7.6 Hz, 1H), 7.32 (d, *J* = 9.1 Hz, 2H), 7.28–7.21 (m, 3H), 6.78 (t, *J* = 7.6 Hz, 1H), 6.24 (d, *J* = 8.5 Hz, 1H), 5.24 (s, 2H), 2.36 (s, 3H) ppm; LRMS (ES⁻) calcd 400.05 for C₂₁H₁₆Cl₂NO₃⁻ found 400.0 (M⁻).



AK-48. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100→70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30→100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 23.8–29.9 min = 86–94% MeCN); white solid (49 mg, 61%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.51 (s, 1H), 7.89 (d, *J* = 8.0 Hz, 1H), 7.55 (d, *J* = 9.7 Hz, 1H), 7.36 (d, *J* = 7.5 Hz, 2H), 7.32 (t, *J* = 7.9 Hz, 1H), 7.24 (d, *J* = 9.4 Hz, 1H), 7.22 (d, *J* = 7.7 Hz, 2H), 6.78 (t, *J* = 7.6 Hz, 1H), 6.23 (d, *J* = 8.2 Hz, 1H), 5.21 (s, 2H), 2.31 (s, 3H) ppm; LRMS (ES⁻) calcd 400.05 for C₂₁H₁₆Cl₂NO₃⁻ found 400.0 (M⁻).



AK-49. Prepared according to the above procedure. Nucleophilic aromatic *para*-substitution of the Buchwald-Hartwig coupling product occurred with the ethanolic solvent upon stirring at ambient

temperature for 48 h. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 24.2–25.8 min = 87–89% MeCN); off-white solid (44 mg, 80%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.50 (s, 1H), 7.89 (dd, *J* = 7.9, 1.7 Hz, 1H), 7.63 (d, *J* = 9.0 Hz, 1H), 7.39 (d, *J* = 9.1 Hz, 1H), 7.32 (ddd, *J* = 8.6, 7.3, 1.7 Hz, 1H), 6.78 (t, *J* = 7.6 Hz, 1H), 6.20 (d, *J* = 8.3 Hz, 1H), 5.31 (s, 2H), 4.35 (q, *J* = 7.0 Hz, 2H), 1.34 (t, *J* = 7.0 Hz, 3H) ppm; ¹⁹F NMR (acetone-d₆, 376 MHz) δ –146.36 (dd, *J* = 20.7, 8.5 Hz, 2F), –159.17 (dd, *J* = 20.7, 8.5 Hz, 2F) ppm; LRMS (ES⁻) calcd 502.02 for C₂₂H₁₄Cl₂F₄NO₄⁻ found 502.05 (M⁻).

AK-50. Prepared according to the above procedure. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100→70:30 MeCN/0.1%TFA in H₂O over 10 min, then 70:30→100:0 MeCN/0.1%TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 23.2–26.0 min = 86–89% MeCN); white solid (53 mg, 88%); ¹H NMR (DMSO-d₆, 600 MHz) δ 9.49 (s, 1H), 7.89 (d, *J* = 7.6 Hz, 1H), 7.53 (dd, *J* = 9.1, 1.4 Hz, 1H), 7.37–7.29 (m, 5H), 7.24–7.17 (m, 2H), 6.78 (t, *J* = 7.5 Hz, 1H), 6.21 (d, *J* = 8.3 Hz, 1H), 4.31 (t, *J* = 6.5 Hz, 2H), 3.08 (t, *J* = 6.5 Hz, 2H) ppm; LRMS (ES⁻) calcd 400.05 for C₂₁H₁₆Cl₂NO₃⁻ found 400.0 (M⁻).



AK-51. Prepared according to the above procedure from the corresponding phthalimide-protected ethyl ester. Hydrolysis of the phthalimide group was incomplete, even after heating at 70 °C for 3 h at pH 12. The mixture was acidified to pH 2 with concentrated HCl and purified by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 13.0–16.0 min = 74–77% MeCN); To liberate the free amine, the white solid obtained following lyophilization was dissolved in 0.8 mL of EtOH and stirred with an excess of MeNH₂ (0.8 mL of a 2.0 M solution in THF, 1.6 mmol, 33.0 equiv) at 70 °C for 5 h. After cooling to ambient temperature, the reaction pH was adjusted to pH 2 with 6.0 M aqueous HCl. Purification by reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with a gradient flow of 0:100 \rightarrow 74:26 MeCN/10 mM heptafluoro-butryic acid (HFBA) in H₂O over 5 min, then 74:26 \rightarrow 78:22 MeCN/10 mM HFBA in H₂O over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 9.2–9.8 min = 74.7–74.8% MeCN); white solid (4 mg, 21% over

two steps); ¹H NMR (DMSO-d₆, 600 MHz) δ 7.90 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.58 (d, *J* = 9.1 Hz, 1H), 7.33 (ddd, *J* = 8.6, 7.1, 1.7 Hz, 1H), 7.19 (d, *J* = 9.1 Hz, 1H), 6.80 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1H), 6.22 (dd, *J* = 8.5, 1.1 Hz, 1H), 4.29 (t, *J* = 5.2 Hz, 2H), 3.27 (t, *J* = 5.1 Hz, 2H) ppm; LRMS (ES⁻) calcd 339.03 for C₁₅H₁₃Cl₂N₂O₃⁻ found 339.0 (M⁻).

Synthesis of sulfate, phosphonate, and tetrazole derivatives



The following procedure is adapted from Sadighi, et al.¹ An oven-dried 5-mL microwave vial was charged with 2,6-dichloro-3-methylaniline (176 mg, 1.0 mmol), 2-bromoanisole (187 mg, 1.0 mmol), and DPEphos (40 mg, 0.075 mmol, 0.075 equiv). The vial was sealed with a septum and purged with argon prior to the addition of 3.0 mL of toluene. The solution was sparged with argon for ~5 min. Under a blanket of argon, the septum was briefly removed and Pd(OAc)₂ (11 mg, 0.05 mmol, 0.05 equiv) was added. The vial was resealed, and the orange solution was stirred at room temperature for 10 min. Following this time, the septum was quickly removed and sodium t-butoxide (135 mg, 1.4 mmol, 1.4 equiv) was added. The headspace of the vial was flushed with argon and quickly resealed with a crimped microwave vial cap. The reaction was stirred at 160 °C in a microwave reactor for 2 h. The reaction mixture was cooled to room temperature, diluted with 10 mL of EtOAc, and filtered through a plug of Celite. The flask and filter cake were rinsed with ~30 mL of EtOAc, and the combined filtrates were concentrated under reduced pressure. The brown residue was re-dissolved in CH_2CI_2 to which ~1 g of silica gel was then added. The suspension was concentrated under reduced pressure and the solid material dry loaded onto a silica gel column prepacked in pentane. Purification of this material was accomplished by chromatography on silica gel (gradient elution: $0:1 \rightarrow 1:19$ Et₂O/pentane) furnished the desired product as a white solid (142 mg, 50%). TLC R_f = 0.6 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 400 MHz) δ 7.29 (d, J = 8.3 Hz, 1H), 7.04 (dd, J = 8.2, 0.7 Hz, 1H), 6.91 (dd, J = 7.9, 1.6 Hz, 1H), 6.86 (td, J = 7.9, 1.8 Hz, 1H), 6.80 (td, J = 7.6, 1.6 Hz, 1H), 6.36 (dd, J = 7.6, 1.7 Hz, 1H), 3.97 (s, 3H), 2.41 (s, 3H) ppm; IR (thin film) v 3372, 1598, 1506, 1452, 1428, 1247, 1225, 1117 cm⁻¹.



Prepared according to the above procedure substituting 1,2-dibromobenzene for 2-bromoanisole; white solid (453 mg, 55%). TLC $R_f = 0.71$ (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 300 MHz) δ 7.54 (dd, J = 7.9, 1.5 Hz, 1H), 7.30 (d, J = 8.3 Hz, 1H), 7.20–7.12 (m, 2H), 6.74 (ddd, J = 7.9, 7.3, 1.5 Hz, 1H), 6.35 (dd, J = 8.1, 1.5 Hz, 1H), 6.12 (s, 1H), 2.41 (s, 3H) ppm; IR (thin film) v 3376, 3062, 2954, 2922, 2851, 1595, 1500, 1464, 1448, 1400, 1305 cm⁻¹.



Prepared according to the above procedure substituting 2-bromobenzonitrile for 2-bromoanisole; white solid (128 mg, 46%). TLC R_f = 0.42 (19:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 500 MHz) δ 7.54 (d, *J* = 7.6 Hz, 1H), 7.37–7.29 (m, 2H), 7.15 (d, *J* = 8.3 Hz, 1H), 6.88 (t, *J* = 7.6 Hz, 1H), 6.37 (d, *J* = 8.4 Hz, 1H), 6.23 (s, 1H), 2.41 (s, 3H), ppm; IR (thin film) v 3306, 2222, 1605, 1578, 1512, 1452, 1399, 1317, 1296, 1163 cm⁻¹.



The following procedure is adapted from Ghalib, et al.⁵ To a 2 mL oven-dried conical reaction vial was added successively N-(2-bromophenyl)-2,6-dichloro-3-methylaniline (263 mg, 0.79 mmol), neat triethylphosphite (0.3 mL, 1.75 mmol, 2.2 equiv), and PdCl₂ (28 mg, 0.16 mmol, 0.2 equiv). The vial was purged with argon for 2 min then capped with a sturdy screw-cap and placed in a pre-heated sand bath at 180-190 °C. The reaction was stirred at this temperature for 3 h, over which time the mixture changed in color from pale yellow to dark brownish-black. The reaction mixture was cooled to ambient temperature, diluted with 10 mL of CH₂Cl₂, and filtered through a plug of Celite. The vial and filter cake were rinsed with 3 x 30 mL of CH₂Cl₂. The combined filtrates were concentrated under reduced pressure. The oily brown residue was re-dissolved in CH₂Cl₂ to which ~1 g of silica gel was then added. The suspension was concentrated under reduced pressure and the solid material dry loaded onto a silica gel column pre-packed in hexanes. Purification by chromatography on silica gel (gradient elution: $0:1 \rightarrow 1:3$ EtOAc/hexanes) provided the desired product as yellow-brown oil (265 mg, 86%). TLC Rf = 0.30 (3:1 hexanes/EtOAc); ¹H NMR (CDCl₃, 300 MHz) δ 8.27 (s, 1H), 7.62 (ddd, J = 14.5, 7.7, 1.6 Hz, 1H), 7.37–7.29 (m, 2H), 7.06 (d, J = 8.3 Hz, 1H), 6.84 (td, J = 7.4, 3.2 Hz, 1H), 6.33 (t, J = 7.5 Hz, 1H), 4.31–4.05 (m, 4H), 2.39 (s, 3H), 1.36 (t, J = 7.1, 1.8 Hz, 6H) ppm; ³¹P NMR (CDCl₃, 400 MHz) δ 21.73 ppm; IR (thin film) v 3259, 2982, 1600, 1512, 1449, 1391, 1218, 1141 cm⁻¹.



To an ice-cold solution of 2,6-dichloro-N-(2-methoxyphenyl)-3-methylaniline (51 mg, 0.18 mmol) in 2.2 mL of anhydrous CH₂Cl₂ was added boron trichloride (0.6 mL of 1.0 M in hexanes, 0.6 mmol, 3.3 equiv) was added dropwise. The reaction was warmed to room temperature over 1 h and stirred for an additional 4 h.

Ghalib, M.; Jones, P. G.; Lysenko, S.; Heinicke, J. W. Enantiomerically Pure N Chirally Substituted 1,3-Benzazaphospholes: Synthesis, Reactivity toward *t*-BuLi, and Conversion to Functionalized Benzazaphospholes and Catalytically Useful Dihydrobenzazaphospholes. *Organometallics* **2014**, 33, 804–816.

Following this time, the reaction was quenched with 2 mL of saturated aqueous NH₄Cl and transferred to a separatory funnel with 60 mL of EtOAc. The organic fraction was washed with 1 x 60 mL of saturated aqueous NH₄Cl and 2 x 60 mL of saturated aqueous NaCl, dried over MgSO₄, filtered and concentrated under reduced pressure. Purification of this material was accomplished by chromatography on silica gel (gradient elution, 0:1 \rightarrow 1:3 EtOAc/hexanes) to furnish the desired product as a clear oil (13 mg, 27%, note: the product is extremely prone to oxidation and turns turquoise upon prolonged exposure to air). TLC R_f = 0.15 (19:1 hexanes/acetone); ¹H NMR (CDCl₃, 300 MHz) δ 7.22 (d, *J* = 8.3 Hz, 1H), 7.05–6.92 (m, 3H), 6.78 (td, *J* = 7.4, 1.9 Hz, 1H), 6.62 (dd, *J* = 7.9, 1.5 Hz, 1H), 5.93 (s, 1H), 2.37 (s, 3H) ppm; IR (thin film) v 3388, 1595, 1511, 1494, 1451, 1222 cm⁻¹.



AK-9. A flask containing anhydrous sulfur trioxide *N*,*N*-dimethylformamide complex (50 mg, 0.33 mmol, 6.8 equiv) was placed in an ice bath to which a solution of 2-((2,6-dichloro-3-methyl-phenyl)amino)phenol (9.8 mg, 0.048 mmol) in 750 \Box L of a 1:4 mixture of freshly distilled pyridine/DMF was added dropwise via syringe. Following the addition, the flask was removed from the ice bath and the mixture was stirred at ambient temperature for 1 h, 30 °C for 2 h, and 45 °C for 2 h. The reaction was quenched by the addition of 0.1 mL of saturated aqueous KOH. All volatiles were removed in vacuo to a yellow residue, which was triturated successively with 1 mL of hexanes, 1 mL of acetone, and 1 mL of EtOAc. The impure material was dissolved in 2 mL of DMSO and filtered through a 13 mm syringe filter with a 0.45 µm PTFE membrane to remove any particulate matter. The desired product was obtained following reversed-phase HPLC (Alltima C18, 10 µM, 22 x 250 mm column, eluting with gradient flow over 30 min of 0:1→1:0 MeCN/H₂O, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 17.4–18.4 min = 58–61% MeCN); white powder (6 mg, 43%). ¹H NMR (DMSO-d₆, 600 MHz) δ 7.44 (s, 1H), 7.39 (d, *J* = 8.3 Hz, 1H), 7.15 (dd, *J* = 8.2, 0.8 Hz, 1H), 7.12 (dd, *J* = 7.9, 1.6 Hz, 1H), 6.88 (td, *J* = 7.7, 1.6 Hz, 1H), 6.77 (td, *J* = 7.6, 1.6 Hz, 1H), 6.23 (dd, *J* = 7.9, 1.6 Hz, 1H), 2.35 (s, 3H) ppm; LRMS (ES⁻) calcd 345.97 for C₁₃H₁₀Cl₂NO₄S⁻ found 345.9 (M⁻).



AK-18. The following procedure is adapted from Vorona, et al.⁶ A 1 mL microscale reaction vial was charged with 2-((2,6-dichloro-3-methylphenyl)amino)benzonitrile (21 mg, 0.076 mmol), 0.2 mL of *n*-propanol, zinc chloride (10 mg, 0.076 mmol), and sodium azide (6 mg, 0.091 mmol, 1.2 equiv). The vial was capped and sealed with Teflon tape, and the mixture was stirred at 95 °C for 3 h. Upon cooling, 0.1 mL of 1.0 M aqueous

^{6.} Vorona, S.; Artamonova, T.; Zevatskii, Y.; Myznikov, L. An Improved Protocol for the Preparation of 5-Substituted Tetrazoles from Organic Thiocyanates and Nitriles. *Synthesis* **2014**, *46*, 781–786.

NaOH was added, resulting in the immediate formation of a white precipitate. The reaction mixture was filtered through a cotton plug. The flask and cotton filter were rinsed with ~3 mL of *n*-propanol. The pH of the filtrate was adjusted to pH 1 with ~1–3 drops of concentrated aqueous HCI. The solution was transferred to a separatory funnel with 5 mL of H₂O and extracted with 5 mL of EtOAc. The organic fraction was washed with 2 x 5 mL of 1.0 M aqueous HCI, dried over Na₂SO₄, filtered and concentrated under reduced pressure. Purification of this material by chromatography on silica gel (2:3 hexanes/EtOAc) provided the tetrazole product as white solid (3 mg, 13%). TLC R_f = 0.13 (2:3 hexanes/EtOAc); ¹H NMR (CDCI₃, 500 MHz) δ 8.74 (s, 1H), 7.85 (d, *J* = 7.8 Hz, 1H), 7.34–7.28 (m, 2H), 7.13 (d, *J* = 8.3 Hz, 1H), 6.96 (t, *J* = 7.5 Hz, 1H), 6.51 (d, *J* = 8.3 Hz, 1H), 2.42 (s, 3H) ppm; IR (thin film) v 2924, 1702, 1616, 1590, 1552, 1480, 1454, 1312, 1286, cm⁻¹; LRMS (ES⁻) calcd 318.03 for C₁₄H₁₀Cl₂N₅⁻ found 317.6 (M⁻).



AK-34. Diethyl (2-((2,6-dichloro-3-methylphenyl)amino)-phenyl)phosphonate (71 mg, 0.18 mmol) was dissolved in 2.0 mL of MeCN, the solution cooled to 0 °C, and bromotrimethylsilane (0.2 mL, 1.5 mmol, 8.3 equiv) added dropwise via syringe. The flask was removed from the ice bath, equipped with a reflux condenser, and placed in an oil bath at 80 °C. The mixture was stirred at reflux under N₂ for 8 h. Following this time, the reaction was cooled to ambient temperature and stirred for an additional 18 h. The reaction was quenched by the addition of 1 mL of H₂O and stirred vigorously for 15 min prior to concentrating the solution under reduced pressure. The yellow-orange residue was transferred to a separatory funnel with 100 mL of EtOAc. The organic layer was washed with 1 x 100 mL of 1.0 M aqueous HCl and 2 x 100 mL of H₂O, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The desired product was obtained following reversed-phase HPLC (Alltima C18, 10 µM, 22 x 250 mm column, eluting with a gradient flow of 0:100–70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30–100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 13.8–15.8 min = 75–77% MeCN); white powder (49 mg, 81%). ¹H NMR (acetone-d₆, 500 MHz) δ 7.70 (ddd, *J* = 15.1, 7.6, 1.7 Hz, 1H), 7.37 (d, *J* = 8.3 Hz, 1H), 7.28–7.19 (m, 2H), 6.81 (td, *J* = 7.3, 2.9 Hz, 1H), 6.28 (t, *J* = 7.4 Hz, 1H), 2.37 (s, 3H) ppm; LRMS (ES⁻) calcd 328.98 for C₁₃H₁₀Cl₂NO₃P₂⁻ found 329.6 (M⁻).



AK-35. Diethyl (2-((2,6-dichloro-3-methylphenyl)amino)phenyl)phosphonate (35 mg, 0.09 mmol) was suspended in 2.0 mL of concentrated aqueous HCI. The flask was equipped with a reflux condenser and the contents were stirred at 110 °C for 5 h. The mixture was cooled to room temperature and stirred for an additional 12 h. Following this time, the aqueous yellow solution was decanted, leaving a bright violet residue. This purple film was dissolved in ~5 mL of EtOAc and concentrated under reduced pressure. The

desired product was obtained following reversed-phase HPLC (Alltima C18, 10 μ M, 22 x 250 mm column, eluting with gradient flow of 0:100 \rightarrow 70:30 MeCN/0.1% TFA in H₂O over 10 min, then 70:30 \rightarrow 100:0 MeCN/0.1% TFA over 25 min, 254 nm UV detection, flow rate = 12.0 mL/min, R_T = 16.8–19.0 min = 78–81% MeCN); white powder (23 mg, 72%). ¹H NMR (acetone-d₆, 500 MHz) δ 7.67 (dd, *J* = 14.4, 7.6 Hz, 1H), 7.41 (d, *J* = 8.2 Hz, 1H), 7.31–7.22 (m, 2H), 6.84 (t, *J* = 6.4 Hz, 1H), 6.27 (t, *J* = 7.4 Hz, 1H), 4.03 (quintet, *J* = 7.3 Hz, 2H), 2.39 (s, 3H), 1.22 (t, *J* = 7.0 Hz, 3H) ppm; LRMS (ES⁻) calcd 358.02 for C₁₅H₁₅Cl₂NO₃P⁻ found 357.6 (M⁻).

Dataset 3: NMR and IR characterization spectra















































































































































































































































































