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GALLO: An R package for Genomic Annotation and integration of multiple data source in livestock for positional candidate LOci --Manuscript Draft--

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Abstract:	<p>The development of high-throughput sequencing and genotyping methodologies and precision livestock farming allowed the identification of thousands of genomic regions associated with several complex traits. The integration of multiple sources of biological information is a crucial step to better understand patterns regulating the development of complex traits. Genomic Annotation in Livestock for positional candidate LOci (GALLO) is an R package, for the accurate annotation of genes and quantitative trait loci (QTLs) located in regions identified in the most common genomic analyses performed in livestock, such as Genome-Wide Association Studies and transcriptomics using RNA-Sequencing. Moreover, GALLO allows the graphical visualization of gene and QTL annotation results, data comparison among different grouping factors (e.g., methods, breeds, tissues, statistical models, studies, etc.), and QTL enrichment in different livestock species including cattle, pigs, sheep, chicken, etc. Consequently, GALLO is a useful package for annotation, identification of hidden patterns across datasets, datamining of previous reported associations, as well as the efficient scrutinization of the genetic architecture of complex traits in livestock.</p>	
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1 **GALLO: An R package for Genomic Annotation and integration of multiple**
2 **data source in livestock for positional candidate LOci**

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14 **Abstract**

15 The development of high-throughput sequencing and genotyping methodologies and precision
16 livestock farming allowed the identification of thousands of genomic regions associated with
17 several complex traits. The integration of multiple sources of biological information is a crucial
18 step to better understand patterns regulating the development of complex traits. Genomic
19 Annotation in Livestock for positional candidate LOci (GALLO) is an R package, for the accurate
20 annotation of genes and quantitative trait loci (QTLs) located in regions identified in the most
21 common genomic analyses performed in livestock, such as Genome-Wide Association Studies and
22 transcriptomics using RNA-Sequencing. Moreover, GALLO allows the graphical visualization of
23 gene and QTL annotation results, data comparison among different grouping factors (e.g.,
24 methods, breeds, tissues, statistical models, studies, etc.), and QTL enrichment in different
25 livestock species including cattle, pigs, sheep, chicken, etc. Consequently, GALLO is a useful
26 package for annotation, identification of hidden patterns across datasets, datamining of previous
27 reported associations, as well as the efficient scrutinization of the genetic architecture of complex
28 traits in livestock.

29 **Keywords:** Multi-omics integration; QTL annotation; Gene annotation; Datamining; QTL
30 enrichment analysis; Livestock

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35 **Background**

36 The identification of quantitative trait loci (QTLs), genomic regions linked to complex traits
37 through association tests using genetic markers and phenotypic traits, is a crucial step in the
38 improvement of genomic selection and economic profitability in livestock [1–4]. Additionally, in
39 the last decades, the development of precision livestock farming strategies resulted in the
40 possibility to obtain a huge volume and diversity of phenotypic data [5–7]. The development of
41 high-throughput methodologies (e.g., Genome-Wide Association Studies, Transcriptomics,
42 Metabolomics, Proteomics, etc.) for the study of the genetic architecture of complex traits allows
43 the identification of potential candidate genes associated with economically relevant traits in
44 livestock. Taken together, these new technologies can substantially improve the accuracy of
45 detection of candidate regions associated with economical important traits across the genome in
46 livestock species [8]. Consequently, the number of QTLs identified across the genome in livestock
47 species increased substantially in the last years. Currently, in the Animal QTLdb it is possible to
48 retrieve information about QTLs previously identified in cattle (127,191), chicken (11,340), horse
49 (2,260), pig (29,865), rainbow trout (584) and sheep (3,001) [9]. The proper integration of the
50 results obtained from different methodologies and technologies available is a crucial step for the
51 accurate identification of the biological processes regulating the development of complex traits as
52 well as the identification of potential functional candidate genes for each trait or shared among
53 traits [8,10–12]. The integration of both structural and functional data can help to scrutinize the
54 genetic architecture of economically relevant traits, consequently, helping to better understand
55 complex biological patterns regulating the expression of these traits, such as pleiotropic effect,
56 epistasis and genetic hitchhiking, among others.

57 In spite of the great potential to improve the identification of functional candidate genes and/or
58 QTLs through the integration of multiple data sources, currently this process is not very
59 straightforward due to limitations in the pipelines and algorithms implemented in the tools
60 available for livestock. Currently, there are several tools that implement functions for gene (i.e.,
61 Biomart and BEDTools) and QTL annotation (Animal QTLdb) [9,13,14]. However, these tools
62 have limitations regarding the automatization process to analyze results from multiple candidate
63 regions (Biomart web application and the R package and Animal QTLdb) or for the visualization
64 of the results. Moreover, although the automatization is possible, the direct link between the
65 candidate regions and/or markers with the annotated genes and QTLs is missed. Consequently,
66 this gap is forcing the user to back solve the overlap between the input and output files in order to
67 perform the proper association between the candidate region and/or markers and the annotated
68 genes and/or positional co-localized QTLs. In addition, nowadays there is still a gap for
69 customized QTL enrichment analyses in the available software and databases. The Genomic
70 functional Annotation in Livestock for positional candidate LOci (GALLO) is an R package
71 designed to provide an automatized and a straightforward environment for gene and QTL
72 annotation in multiple candidate regions, as well as data integration from multiple data sources.
73 The QTL enrichment analyses can additionally be performed directly by GALLO using the output
74 obtained from the QTL annotation step. In addition, GALLO also provides a set of functions for
75 graphical visualization of the annotation, comparison, integration and QTL enrichment results. In
76 this context, the GALLO package was developed as an alternative tool: 1) to allow the integration
77 and the simultaneous annotation of multiple datasets for genes and QTLs; 2) to provide graphical
78 visualization tools to integrate visually the annotation and the similarity against the datasets; 3) to

79 perform QTL enrichment analysis for the positional candidate genomic regions and/or markers
80 associated with economically relevant traits in livestock.

81 **Implementation**

82 The GALLO package was wrote in the R language [15]. The stable release is available as an R
83 package on CRAN (<https://cran.r-project.org/web/packages/GALLO/index.html>). The code was
84 extensively tested with several datasets from different sources and methodologies and reviewed to
85 ensure the package quality standards. Additionally, the vignettes were created to be comprehensive
86 and to present practical examples in order to provide a user-friendly and an easier understanding
87 and usability for the user.

88 The GALLO package provides a useful set of functions that allows the user a straightforward data
89 integration, comparison, gene and QTL annotation, and visualization of several data sources and
90 methodologies, such as data from genome-wide association study (GWAS), RNA-Sequencing,
91 whole-genome sequencing, etc. (Figure 1 and Table 1). The main advantage to perform an
92 automated analysis from multiple datasets results in the flexibility to handle the output using
93 different subsets (traits, populations, models, etc.) in the same environment, without generating
94 multiple intermediate output files.

95 **Methods**

96 *Case study – Candidate regions for scrotal circumference and fertility in cattle*

97 The dataset used to present the basic usage and advantages of GALLO package is composed by
98 the markers significantly associated with scrotal circumference in Canchim breed [16] and
99 uncompensable fertility in Holstein cattle [17]. These two studies were previously analyzed

100 together in a systematic review regarding male fertility in cattle [11]. Therefore, the data used
101 herein comprises a multi-study and multi-breed analysis. These candidate markers (527 single
102 nucleotide polymorphisms (SNPs)) are available on Supplementary Table 1. In addition to the
103 candidate markers, we present as Supplementary Files 1 and 2, the annotation gff file containing
104 the QTL database information for cattle (obtained from the Animal QTLdb;
105 https://www.animalgenome.org/cgi-bin/QTLdb/BT/download?file=gffUMD_3.1) and the gtf file
106 containing the genes annotated in the cattle genome obtained from Ensembl
107 (ftp://ftp.ensembl.org/pub/release-94/gtf/bos_taurus/). The genomic coordinates of both files were
108 based on the bovine reference genome version UMD 3.1 due to the original coordinates used to
109 report the location of the candidate markers in the original studies. Here, the analysis performed
110 follows the same logical order to the one presented in the GALLO vignette
111 (https://rpubs.com/pablo_bio/GALLO_vignette). However, the dataset used in the user practical
112 tutorial is a subset of the data presented here, aiming to reduce computational demand for the users.
113 The script with all the commands used to perform the analysis present here are available in
114 Supplementary File 3. All the tests were performed using a desktop with a processor Intel Core i5
115 2.4 GHz with 8 Gb of RAM memory.

116 *Importing datasets and annotating genes and QTLs around candidate markers*

117 The first step in the pipeline consists in importing the databases which will be used for the analyses
118 with the *import_gff_gtf()* function. In our specific example, we imported both, cattle gene
119 annotation (gtf) and QTL (gff) databases. The *import_gff_gtf()* function receives as arguments the
120 database file (db_file) and the file type (file_type= gff or gtf) and creates a dataframe with the
121 respective information from each file. The system time demanded to import the gtf and gff files
122 were 0.045 and 0.311 seconds, respectively, indicating an efficient importing process. The file

123 containing the candidate markers can be imported using any available function in the R
124 environment such as *read.table()* and *read.csv()*.

125 The main function of GALLO, *find_genes_qtls_around_markers()*, is responsible to perform the
126 annotation of genes and/or co-localized QTLs within or nearby candidate markers or genomic
127 regions (using a user's defined interval/window). This function uses the information provided in
128 the .gtf file (for gene annotation) or .gff (for QTL annotation) to retrieve the requested information.
129 The output combines the information available in the input file provided by the user with the
130 information available for the genes and QTLs mapped in the candidate genomic regions.
131 Consequently, for example, for an input file composed of three genomic coordinates where 4 genes
132 are annotated in each of the intervals determined by the user, the output file of
133 *find_genes_qtls_around_markers()* will contain 12 rows. The minimum information necessary
134 for the gene and QTL annotation procedures is a data frame with two columns with the
135 chromosome (CHR) and position in base pairs (BP) in the case of candidate SNPs input. In the
136 case of candidate haplotypes, windows, copy number variations (CNVs) or candidate regions; the
137 input file is composed by three columns corresponding to the chromosome (CHR), the start
138 position in base pairs (BP1) and the end position in base pairs (BP2). Data examples for the
139 candidate markers and windows input files can be obtained using the *data(QTLmarkers)* and
140 *data(QTLwindows)* commands in R. Additionally, examples of QTL and gene annotation results
141 are accessible through the commands *data(gtfGenes)* and *data(gffQTLs)* commands, respectively.
142 These outputs can be easily handled by summary functions in R, such as *table()*, to obtain
143 information such as the total number of genes and QTLs, the number of genes and QTLs annotated
144 per variants, etc. The performance of GALLO package in terms of efficiency is similar to other
145 currently packages and software which allow a similar annotation of candidate genomic loci. In

146 this sense, the gene annotation process was compared with the *getBM()* function from the biomaRt
147 package. The gene annotation process on GALLO needed 0.424 seconds to completely annotate
148 the genes in a 200 Kb interval (upstream and downstream) from candidate markers, while the
149 biomaRt function required 0.019 seconds. The QTL annotation on GALLO was compared with
150 the Bedtools `-wao -C` command, resulting in 0.851 and 0.12 seconds required for each approach,
151 respectively. It is important to highlight that for both gene and QTL annotation using biomaRt and
152 bedtools, respectively, a posterior processing of the output file is required in order to match the
153 candidate markers and the genes and QTLs mapped within the candidate intervals. On the order
154 hand, the output file from *find_genes_qtls_around_markers()* function was designed to allow this
155 match in an intuitive way, combining the rows of both candidate markers file and database files
156 (gff and gtf). Additionally, GALLO allows the user to perform both annotations for genes and
157 QTLs with a single software and programming language. Consequently, GALLO obtains a more
158 elaborate and informative output with compromise the computational demand for the analysis. The
159 output files obtained in the gene and QTL annotation are available on Supplementary Tables 2 and
160 3, respectively.

161 *Comparing and visualizing the overlapping of genes and QTLs annotated within the candidate*
162 *regions*

163 The output file generated by the *find_genes_qtls_around_markers()* function can be used as an
164 input file for the other set of GALLO functions. An advantage from the output of
165 *find_genes_qtls_around_markers()* function is any additional information present in the input file
166 will be retained in the output file. Consequently, this information can be used compare the retrieved
167 information between groups of population, methodologies, statistical models, etc. For example,
168 the functions *overlapping_among_groups()* and *plot_overlapping()* can be used to create matrices

169 with the overlapping values among groups and to visualize this overlapping. The Figure 2 shows
170 the results of gene and QTL overlapping between the positional markers obtained in the two studies
171 selected for the dataset of markers analyzed, Feugang et al. (2009) [17] and Buzanskas et al. (2017)
172 [16]. It is important to highlight that these overlapping matrices are not symmetrical. The
173 percentage of genes from study A shared with the study B, and vice-versa, are calculated in
174 function of the total number of genes in A or B, respectively. In the current example, it is possible
175 to note that only a small percentage of the positional candidate genes were shared between the
176 studies. However, the analysis of QTL (using the trait name as reference ID) overlapping indicated
177 a higher similarity between the studies, 46% of the all the QTLs annotated in the candidate regions
178 from Feugang et al. (2010) [17] were also present in Buzanskas et al. (2017) [16] and 93% of the
179 QTLs annotated in the candidate regions from Buzanskas et al. (2017) were also present in
180 Feugang et al. (2010) [16,17]. These results may suggest that even with a small proportion of
181 shared genes, the candidate regions of both studies are frequently associated with similar
182 processes. Similar roles played by the positional candidate genes in those regions in related
183 biological process would be one of the reasons of the observed result.

184 *Understanding the QTL context of the candidate regions*

185 A more precise investigation of the QTL representativeness and diversity can help to better
186 understand the genomic context of the candidate regions. The recurrent association of particular
187 genomic regions with multiple traits might suggest the presence of complex genetic mechanisms
188 regulating that region, such as pleiotropy, epistasis, hitchhiking effect, among others [18,19]. The
189 *plot_qtl_info()* function from GALLO allows the graphical visualization of the summary of QTL
190 types and traits annotated. The percentage of each QTL type annotated within the candidate regions
191 is presented in a pie plot through the use of the argument `qtl_plot="qtl_type"`, while the percentage

192 of each trait associated with a specific QTL type can be plotted setting the argument
193 `qtl_plot="qtl_name"` and informing the additional argument `qtl_class` (that must receive the name
194 of the QTL class to be plotted). Figure 3 shows that for Feugang et al. (2009) [17] the two most
195 frequent QTL types were Milk (50.42%) and Reproduction (16.97%), while for Buzanskas et al.
196 (2017) [16] the most frequent QTL types were Reproduction (87.06%) and Meat and Carcass
197 (5.03%). An in depth analyses can be performed for each QTL type in order to observe the
198 frequency of each trait associated with a specific QTL type. The most frequent traits related with
199 Reproduction QTLs were calving ease (>3%) and scrotal circumference (>60%) for Feugang et al.
200 (2009) and Buzanskas et al. (2017) [16,17], respectively (Figure 3). The comparison between the
201 frequency of traits related with Reproduction QTLs annotated in Feugang et al. (2009) and
202 Buzanskas et al. (2017) [16,17] indicated that among the top 10 more frequent QTLs, calving ease,
203 inhibin levels, stillbirth, interval to first estrus after calving, and birth index were shared between
204 the studies. The combined analysis (not filtering by study) indicated that the Reproduction and
205 Milk QTL types were the two most frequent classes with 76.99% and 10.62% of all QTL types,
206 respectively. In addition, scrotal circumference, inhibin level and calving easy were the most
207 frequent Reproduction QTL related traits in the combined analysis.

208 *QTL enrichment analysis*

209 In some cases, the biases produced by a greater research into certain areas/traits of higher relevance
210 to animal production (such as milk production related traits in the QTL database for cattle) may
211 result in a larger proportion of records for these traits in the QTL database. Consequently, the
212 simple investigation of the proportion of each QTL type might not be totally useful. The GALLO
213 package allows the user to perform a QTL enrichment analysis to test the significance of the QTL
214 representativeness. The QTL enrichment analysis function on GALLO package is based in a

215 hypergeometric test approach, where the number of QTLs annotated within the candidate regions,
216 for each QTL type or trait, is compared with the observed number of QTLs in the reference
217 database. The *qtl_enrich()* function allow the user to perform the QTL enrichment analysis for
218 both QTL types and traits (*qtl_type*= “QTL_type” or “Name”), for the whole genome or
219 chromosome-wise (*enrich_type*= “genome” or “chromosome”) and for all the annotated
220 chromosomes or a subset (*chr.subset*= NULL or the object with the subset of chromosomes). The
221 use of chromosome-wise enrichment analysis might help to detect specific regions across the
222 genome with high number of QTLs for some specific trait, i.e. BTA14 in cattle for milk production
223 [20]. A total of 161 unique pairs of traits and chromosomes were tested for the enrichment using
224 the annotated QTLs for both studies. The system time required to perform the enrichment analysis
225 was 5.32 seconds, suggesting efficient processing. The top 10 enriched QTLs (False Discovery
226 Rate (FDR) < 0.05) for the combined analysis are shown in Table 2 and the enrichment results for
227 all the annotated QTLs are shown in Supplementary Table 4. Additionally, GALLO also allows
228 the user to obtain a graphical visualization, in a bubble plot, of the enrichment results using the
229 *QTLenrich_plot()* function. This function received as arguments the enriched table obtained from
230 *qtl_enrich()*, the name of a column with the traits names to be plotted and the name of a column
231 with the p-values to be plotted. A total of 28 pairs of traits and chromosomes were found enriched
232 in the combined analysis, with scrotal circumference (BTA 5, 18, 9, and 21), milk glycosylated
233 kappa-casein percentage (BTA 6 and 16), inhibin level (BTA 5), triglyceride level (BTA 5), milk
234 kappa-casein percentage (BTA 6) and milk iron content (BTA 23) in the list of top 10 most
235 enriched traits (Figure 4).

236 *Relationship between studies and enriched QTLs*

237 An interesting functionality of GALLO is the graphical visualization of the relationship between
238 groups using a chord plot. The *relationship_plot()* function receives as argument a dataframe (it
239 can use the gene or QTL annotation results, the QTL enrichment, or any other table with two
240 groups of information to be compared), the two groups to be compared (arguments x and y) and
241 the set graphical arguments or set the size, color and gap between the sector in the chord plot. The
242 Figure 5 shows the chord plot obtained using a subset of the QTL annotation dataframe composed
243 only by the top 10 enriched traits and the studies which these traits were annotated. This plot
244 indicated that only inhibin levels and scrotal circumference on BTA5 are shared between Feugang
245 et al. (2009) and Buzanskas et al. (2017) [16,17]. Additionally, milk glycosylated kappa-casein
246 percentage (BTA 6 and 16), milk kappa-casein percentage (BTA 6) and milk iron content (BTA
247 23) were annotated only in Feugang et al. (2009) [17] and scrotal circumference (BTA 9, 18, 21)
248 and triglyceride level (BTA 5) were annotated only in Buzanskas et al. (2017) [16]. Inhibin is
249 produced by the Sertoli cells and can be used as a biomarker for sexual development [21]. In
250 addition, the inhibin levels were already associated with both scrotal circumference and sperm
251 quality traits in several studies, suggesting an important role in male fertility [22–26]. The results
252 obtained here through the integration of the GWAS results from two independent studies followed
253 by QTL annotation reinforce this association. Additionally, QTLs not associated with
254 Reproduction phenotypes were identified in the enrichment analysis, suggesting the presence of
255 complex such as pleiotropic effect, epistasis and genetic hitchhiking effect. Previous studies
256 already highlight the possible role of genomic regions with this kind of processes in the cattle
257 genome [27,28]. An additional integration of the QTL annotation and enrichment analysis
258 performed here with the gene annotation and prospection for functional candidate genes can be a

259 powerful tool to better understand the genetic architecture and the relationship among complex
260 traits.

261 **Discussion**

262 The GALLO package is composed of a group of functions designed to perform an efficient and
263 direct downstream analysis for the gene and QTL annotation for candidate markers/SNPs,
264 haplotypes, genomic windows, runs of homozygosity, CNVs, etc. The functions implemented in
265 GALLO were designed in order to allow the integration of multiple datasets simultaneously. A
266 brief summary of these functions is shown in Table 1. For example, GWAS results from multiple
267 traits and/or populations or breeds can be analyzed together and compared or individually analyzed
268 in the downstream analysis. This can be easily performed by adding an extra column in the input
269 file with the grouping factors to classify each dataset. These input files can be easily adapted from
270 the output of the most common used software to analyze high-throughput genomic data, such as
271 PLINK, BLUPF90, DESeq2, etc. [29–31]. In addition, GALLO provides a set of functions
272 designed for the visualization of the annotation results, overlapping among groups, relationship
273 between groups (i.e., markers and candidate genes, datasets and QTLs, models and positional
274 candidate genes, etc.), and QTL enrichment results. This set of functions provides the user the
275 capability to integrate several results from multiple sources including different methodologies
276 (GWAS, RNA-sequencing, proteomics, etc.), populations (breeds, time-points, etc.), traits or the
277 different combination of these groups or others.

278 A summary of usage examples and output descriptions, for all the functions available on GALLO
279 can be find in the reference manual (Supplementary File 4). It is important to highlight that the
280 two studies used as an example here are also part of the bovine QTL database present in the Animal

281 QTLdb. Consequently, the results obtained here for annotation and enrichment would be expected,
282 once the candidate regions are necessarily present in the annotation database. This approach was
283 used as proof of concept of the methodology and indicates a precise annotation of the candidate
284 regions.

285 **Conclusion**

286 The integration of multiple datasets for gene and QTL annotation is one of the major bottlenecks
287 for the automatization of functional analysis of the results obtained using high-throughput
288 methodologies. The GALLO package provides a user-friendly and straightforward environment to
289 perform gene and QTL annotation, visualization, data comparison and QTL enrichment for
290 functional studies in livestock species. Consequently, the use of GALLO in the analysis of data
291 generated from high-throughput methodologies may improve the identification of hidden patterns
292 across datasets, datamining of previous reported associations, as well as the efficiency in the
293 scrutinization of the genetic architecture of complex traits in livestock.

294 **Availability and requirements**

295 Project name: Genomic Annotation in Livestock for positional candidate LOci (GALLO)

296 Project home page: <https://github.com/pablobio/GALLO>

297 Operating system(s): Platform independent

298 Programming language: R

299 Other requirements: Depends: R ($\geq 3.5.0$)

300 License: GPL-3

301

302

303 **Availability of supporting data**

304 All the data analyzed in the present study can be accessed in the public repository hosting the R
305 package (<https://github.com/pablobio/GALLO>). The input files and results used as examples in the
306 manuscript preparation are available in the supplementary Tables 1-4. A manual including usage
307 examples and output descriptions, for all the functions available on GALLO can be find in the
308 reference manual (Supplementary File 4).

309 **Declarations**

310 *List of abbreviations*

311 BP: position in base pairs; BP1: start position in base pairs; BP2: end position in base pairs; CHR:
312 Chromosome; CNV: Copy Number Variation; GALLO: Genomic Annotation in Livestock for
313 positional candidate Loci; GWAS: Genome-Wide Association Study; QTL: Quantitative trait loci;
314 SNP: Single Nucleotide Polymorphism.

315 *Ethics approval and consent to participate*

316 Not applicable.

317 *Consent for publication*

318 Not applicable.

319 *Competing interests*

320 The authors declare that they have no competing interests.

321

322

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329 preparation of the manuscript.

330 *Authors' contributions*

331 PASF, ASV and AC were responsible for the conceptualization, data processing and review of the
332 codes. PASF and ASV were responsible for data curation. PASF and GM were responsible to
333 implement the bioinformatic pipeline, integrate datasets, and the coding. AC was responsible for
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419

420 **Tables**

421 Table 1: Description of the functions implemented in the GALLO package.

Function	Description	Output
Gene and QTL annotation		
import_gff_gtf	Import the gff and gtf files used for QTL and gene annotation, respectively	A dataframe composed by the information present in the gtf and gff files
find_genes_qtls_around_markers	Annotation of genes and QTLs around candidate regions	A data frame composed of the columns present in the input file and the genes or QTLs mapped within or around (if interval provided) the candidate regions
Data visualization		
overlapping_among_groups	Overlapping between grouping factors (such as different traits, statistical models, populations, studies, stc.)	A list with three matrices: 1) A matrix with the number of overlapping data; 2) A matrix with the percentage of overlapping; 3) A matrix with the combination of the two previous ones
plot_overlapping	Plot overlapping between data and grouping factors	A heatmap with the overlapping between groups
plot_qtl_info	Plot QTLs information from the gene or QTL annotation output	A pie plot (if QTL class is chosen) or a bar plot (if trait name is chosen) for the annotated QTLs
relationship_plot	Plot the relationship among	A chord plot linking a grouping factor (genomic regions, traits, populations,

the candidate regions or grouping factors with the annotated genes and QTLs

etc.) with the annotated genes or QTLs

QTL enrichment

qtl_enrich

Performs a QTL enrichment analysis based on a Bootstrap simulation for each QTL class or trait

A data frame composed of the enrichment results for QTL classes or traits present in the input file. 1) QTL: The QTL class or trait used for the enrichment; 2) CHR: The chromosome for that specific QTL or trait (if the option "chromosome" is informed to the argument enrich_type); 3) N_QTLs: Number of observed QTLs or traits in the dataset; 4) N_QTLs_db: Number of each annotated QTL in the qTL database; 5) Total_annotated_QTLs: Total number of annotated QTLs; 6) Total_QTLs_db: Total number of QTLs in the QTL database; 7) pvalue: P-value for the enrichment analysis; 8) adj.pval: The adjusted p-value based on the multiple test correction selected by the user; 9) QTL_type= The QTL type for each annotated trait.

QTLenrich_plot

Creates a bubble plot with the QTL enrichment results

A plot with the QTL enrichment results

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428 Table 2: Top 10 enriched QTLs for the combined analysis performed with the candidate regions from the two studies, Feugang et al.
 429 (2009) and Buzanskas et al. (2017), used in the example dataset.

QTL	CHR	# QTLs	# QTLs db	Total # QTLs	Total # QTLs db	p-value	FDR	QTL type
Scrotal circumference	5	132	134	347	5942	1.56E-171	4.98E-169	Reproduction
Scrotal circumference	18	11	13	41	2147	2.20E-18	3.52E-16	Reproduction
Scrotal circumference	9	11	14	30	1395	2.04E-17	2.18E-15	Reproduction
Milk glycosylated kappa-casein percentage	6	71	1607	204	12158	1.86E-15	1.49E-13	Milk
Inhibin level	5	47	285	347	5942	3.38E-11	2.16E-09	Reproduction
Scrotal circumference	21	4	5	12	3606	3.51E-10	1.87E-08	Reproduction
Milk kappa-casein percentage	6	76	2637	204	12158	2.39E-07	1.01E-05	Milk
Triglyceride level	5	6	7	347	5942	2.53E-07	1.01E-05	Health
Milk glycosylated kappa-casein percentage	16	7	44	21	1440	1.29E-06	4.58E-05	Milk
Milk iron content	23	4	8	19	1159	3.48E-06	0.00011329	Milk

430

431 **Figure legends:**

432 **Figure 1:** Workflow explaining the main functions implemented on GALLO. The grey rectangles represent
433 the functions, while the rounded and sharp rectangles represent the main goal of that respective function
434 and its input, respectively.

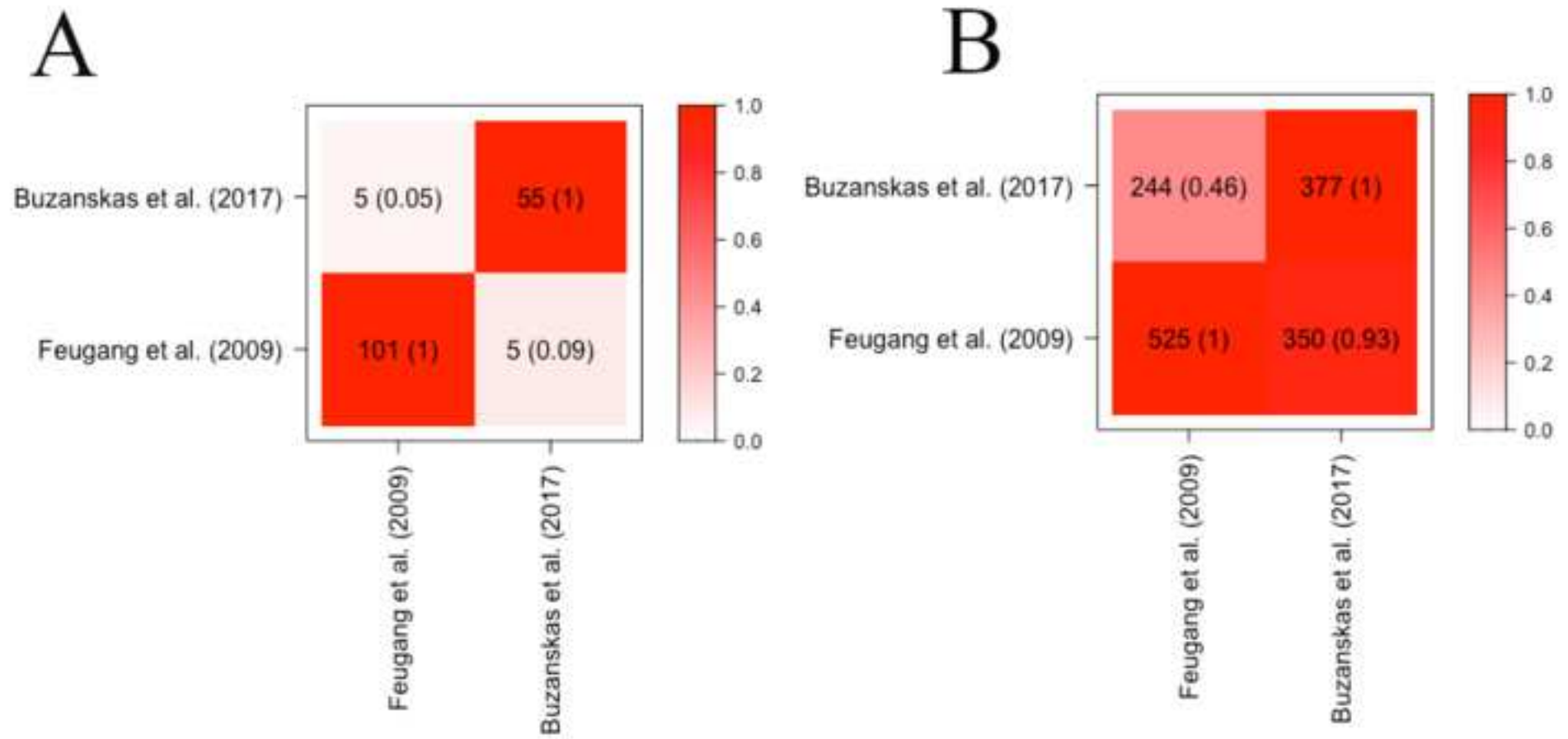
435 **Figure 2:** Overlapping between genes (A) and QTLs (B) annotated within the candidate regions
436 (100 Kb downstream and upstream from the significant markers) from Feugang et al. (2009) and
437 Buzanskas et al. (2017). The darker the color within the squares, the higher is the percentage of
438 shared genes or QTLs.

439 **Figure 3:** Percentage of QTL type (pie plot) and trait related to Reproduction QTLs (barplots) for
440 the QTL annotation results obtained for Feugang et al. (2009) (A), Buzanskas et al. (2017) (B) and
441 the combined analysis (using both studies) (C).

442 **Figure 4:** Bubble plot displaying the enrichment results for the top 10 enrich QTLs identified
443 using the QTLs annotated within the candidate regions from Feugang et al. (2009) and Buzanskas
444 et al. (2017). The darker the red shade in the circles, stronger is the enrichment. The area of the
445 circles is proportional to the number of QTLs. The x-axis shows a richness factor obtained by the
446 ratio of number of QTLs annotated in the candidate regions and the total number of each QTL (and
447 chromosome in the case of this plot) in the reference database.

448 **Figure 5:** Chord plot showing the relationship between the top 10 enriched QTLs (Scrotal
449 circumference – SCRCIR, Inhibin level – INHIB, Triglyceride level – TRIGLY, Milk glycosylated
450 kappa-casein percentage – MGKCASP, Milk iron content – MFE, Milk kappa-casein percentage
451 - MKCASP) and the studies (Feugang et al. (2009) in purple and Buzanskas et al. (2017) in pink).





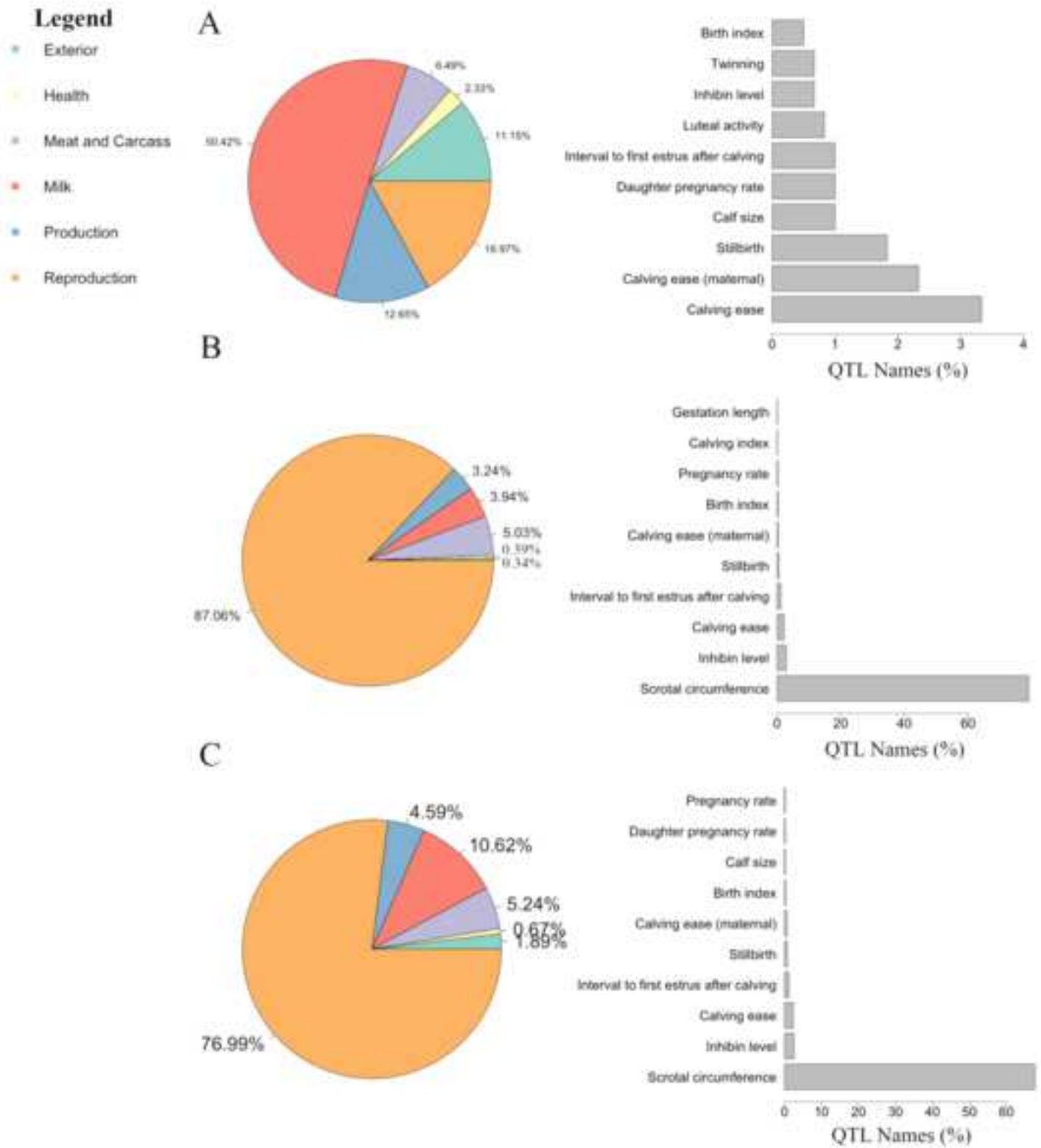
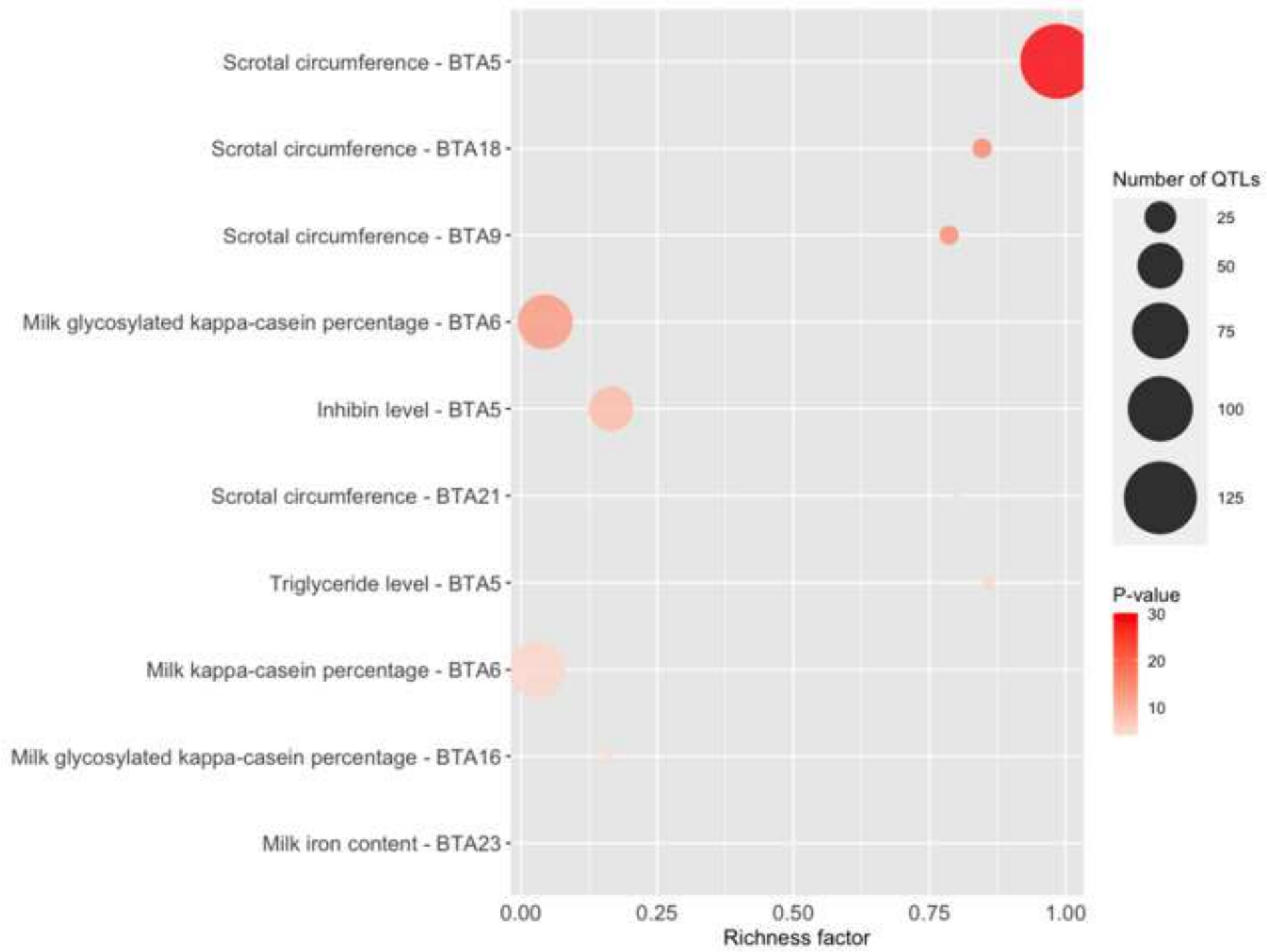
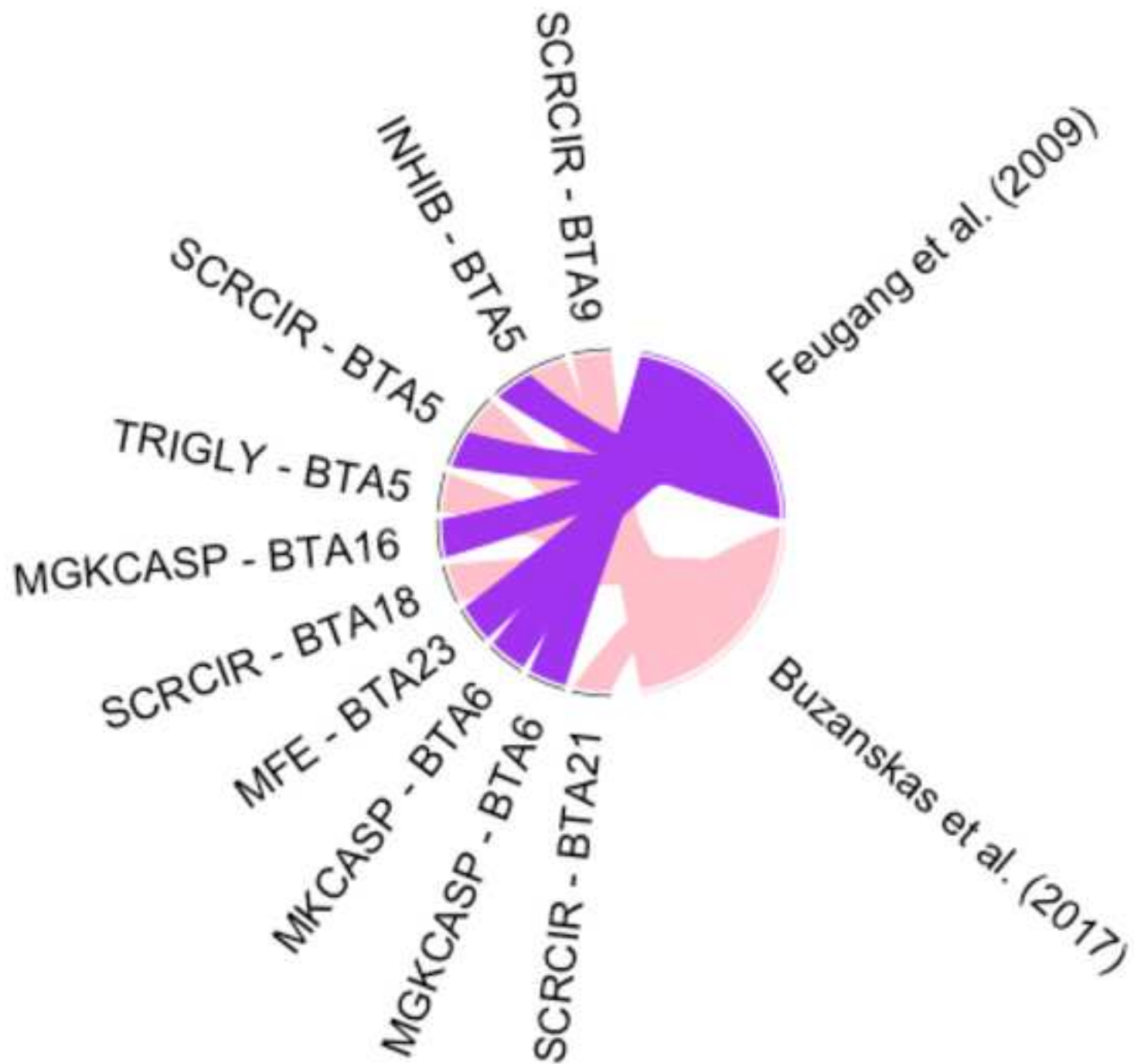


Figure 4







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Guelph, September 1st, 2020

Dear Editorial Office,

We are pleased to re-submit the manuscript entitled “GALLO: An R package for Genomic Annotation and integration of multiple data source in livestock for positional candidate LOci” for consideration to publish it in the GigaScience. This is a resubmission of this manuscript after the inclusion of all the suggestion and considerations raised by the editor and the prior publication of the package in an official repository, in this case, the CRAN.

The present study introduces the applicability and the functionalities of GALLO package, developed in the R environment.

The identification of quantitative trait loci (QTLs) is a crucial step in the improvement of genomic selection and economic profitability in livestock. The development of high-throughput sequencing and genotyping methodologies and precision livestock farming allowed the identification of thousands of genomic regions associated with several complex traits. Consequently, the number of QTLs identified across the genome in livestock species increased substantially in the last years. Currently, in the Animal QTLdb it is possible to retrieve information about QTLs previously identified in cattle (127,191), chicken (11,340), horse (2,260), pig (29,865), rainbow trout (584) and sheep (3,001). The proper integration of the results obtained from different methodologies and technologies available is a crucial step for the accurate identification of the biological processes regulating the development of complex traits as well as the identification of potential functional candidate genes. However, currently, the integration of multiple data sources is not very straightforward due to limitations in the pipelines and algorithms implemented in the tools available for livestock. Moreover, although the automatization is possible, the direct link between the candidate regions and/or markers with the annotated genes and QTLs is missed. Consequently, this gap is forcing the user to back solve the overlap between the input and output files in order to perform the proper association between the candidate region and/or markers and the annotated genes and/or positional co-localized QTLs. In addition, nowadays there is still a lack of for customized QTL enrichment analyses in the available software and databases. Genomic Annotation in Livestock for positional candidate LOci (GALLO) is an R package, for the accurate annotation of genes and QTLs located in regions identified using the most common genomic analyses performed in livestock, such as Genome-Wide Association Studies and transcriptomics using RNA-Sequencing. Moreover, GALLO allows the graphical visualization of gene and QTL annotation results, data comparison among different grouping factors (e.g., methods, breeds, tissues, statistical models, studies, etc.), and QTL enrichment in different livestock species including cattle, pigs, sheep, chicken, etc. Consequently, GALLO is a useful package for annotation, identification of hidden patterns across datasets, datamining of previous reported associations, as well as the efficient scrutinization of the genetic architecture of complex traits in livestock.

We affirm that this manuscript has not been published elsewhere and is not under consideration by any other journal. All authors have approved the manuscript and agree with its submission to GigaScience.

The authors declare that they have no competing interests. With my best regards,

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