

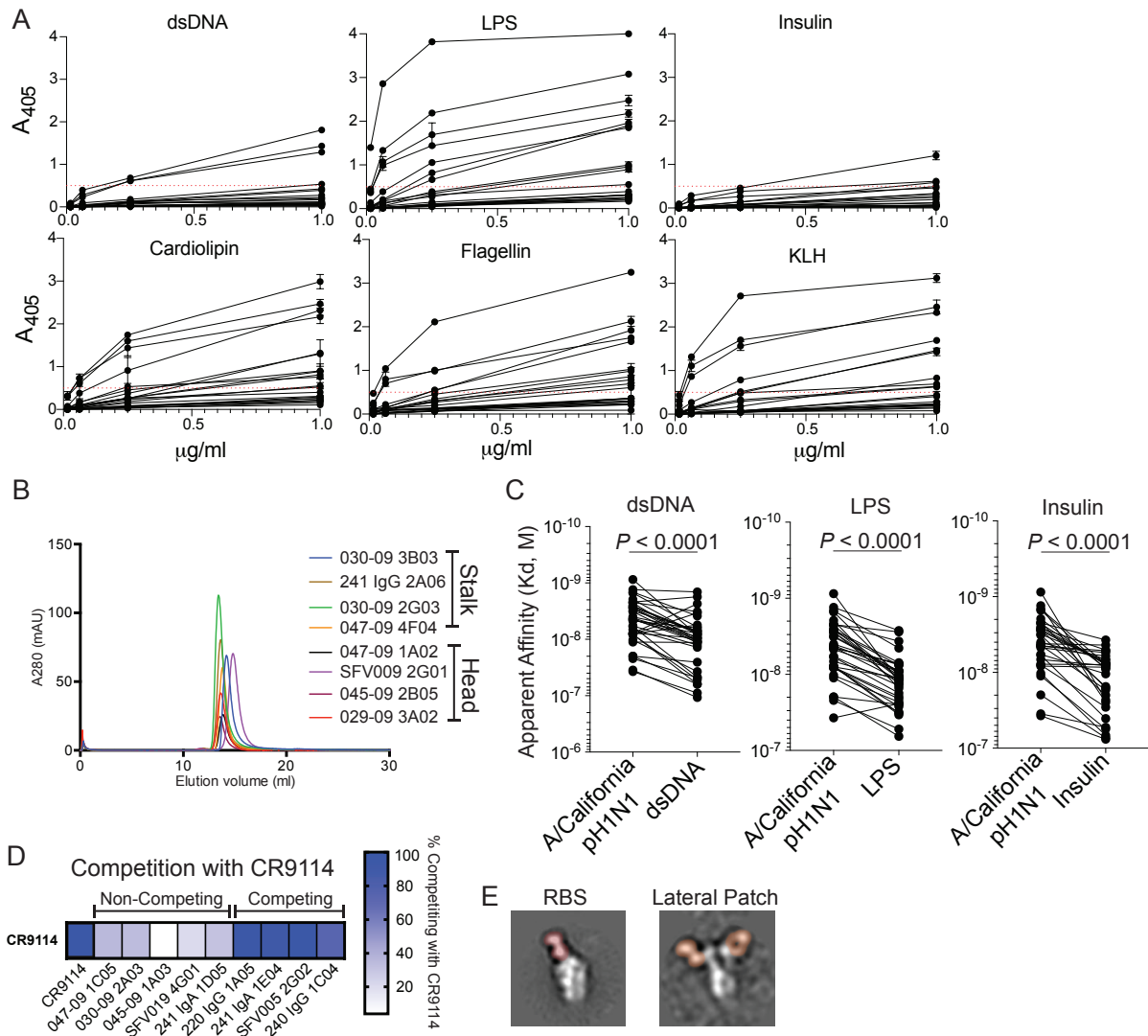
**Supplemental Information**

**Polyreactive Broadly Neutralizing B cells**

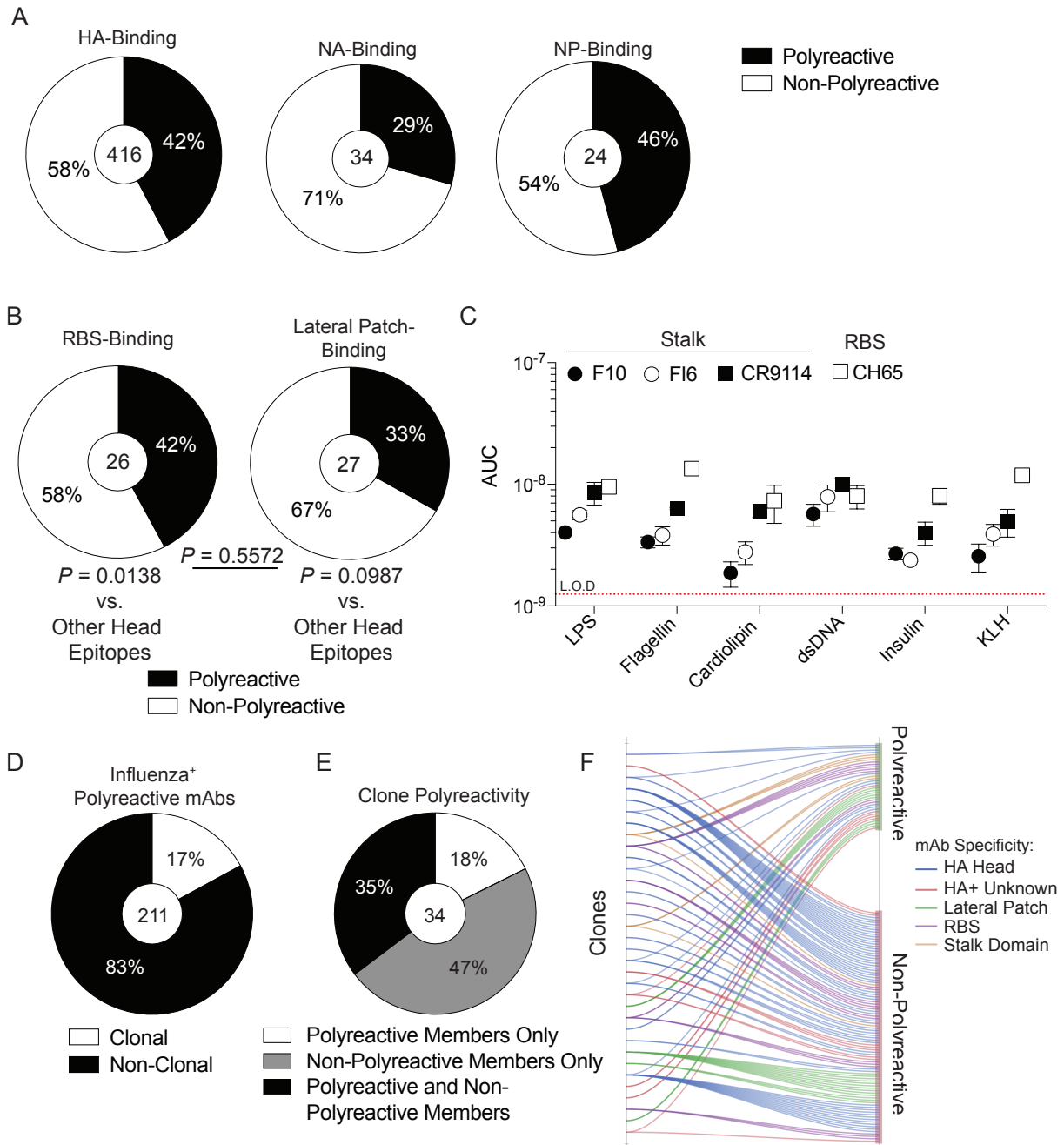
**Are Selected to Provide Defense**

**against Pandemic Threat Influenza Viruses**

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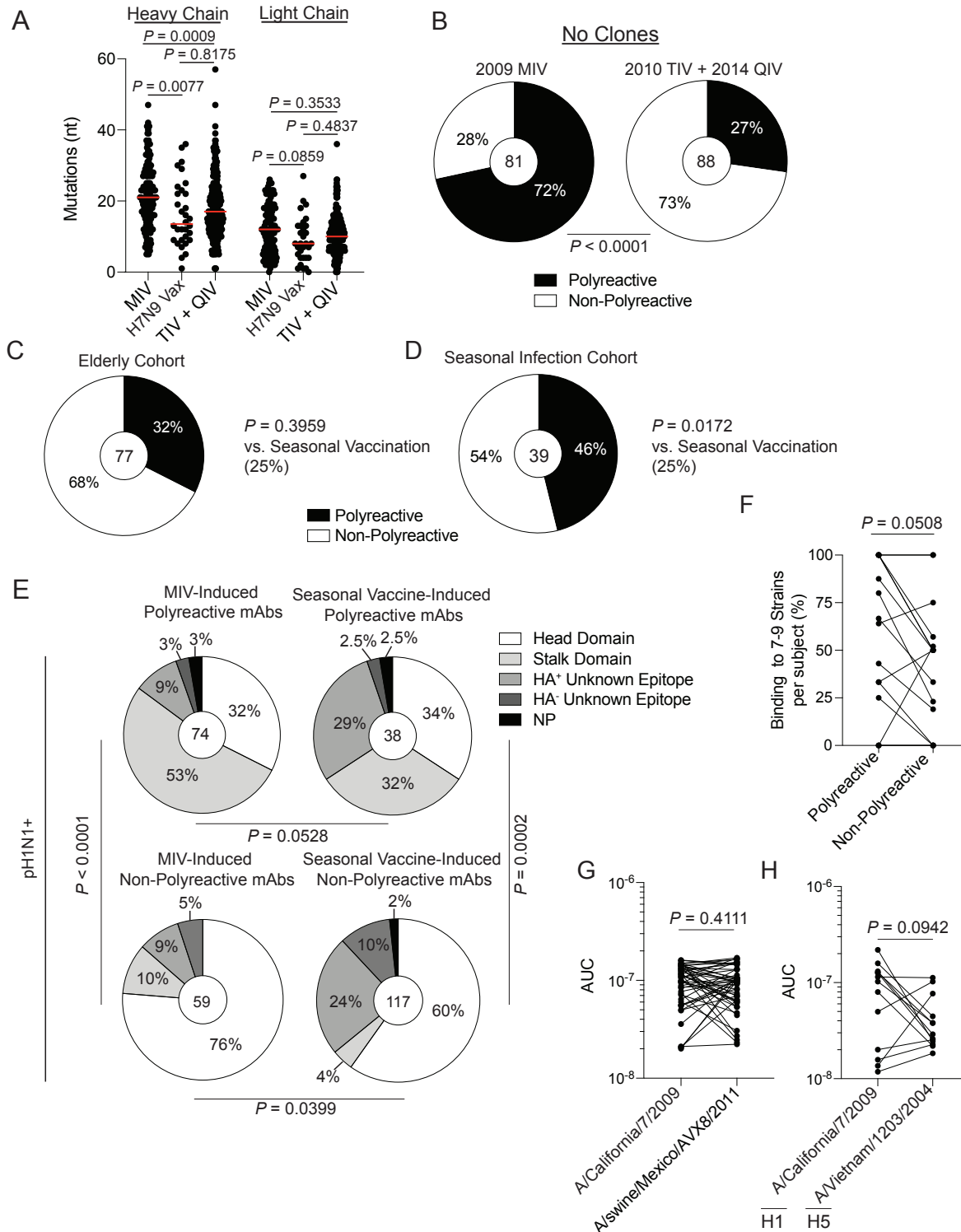


**Figure S1: Polyreactive binding of influenza virus-binding antibodies. Related to Figure 1.** (A) Polyreactivity ELISA example graphs. Representative of 22 antibodies tested for polyreactivity against the 6 antigens used in the polyreactivity ELISA panel. (B) Size exclusion chromatography of polyreactive mAbs indicating that polyreactive mAbs are monomers and do not form aggregates. (C) Paired apparent affinity ( $K_d$ ) of polyreactive mAbs binding to A/California/7/2009 (pH1N1) virus and dsDNA ( $n=37$ ), insulin ( $n=30$ ), or LPS ( $n=36$ ). Each line connects the same mAb. (D) Representative competition ELISA results of mAbs competing or not competing with CR9114, an antibody that specifically targets the BN stalk epitope. (E) Representative negative stain electron microscopy of an RBS binding antibody (SFV018 2D01 fab in red) and lateral patch binding antibody (045-09 2B05 fab in orange). Data in C were analyzed by paired non-parametric Wilcoxon matched-pairs signed rank Tests. Limit of Detection (L.O.D.) represented as dashed red line.



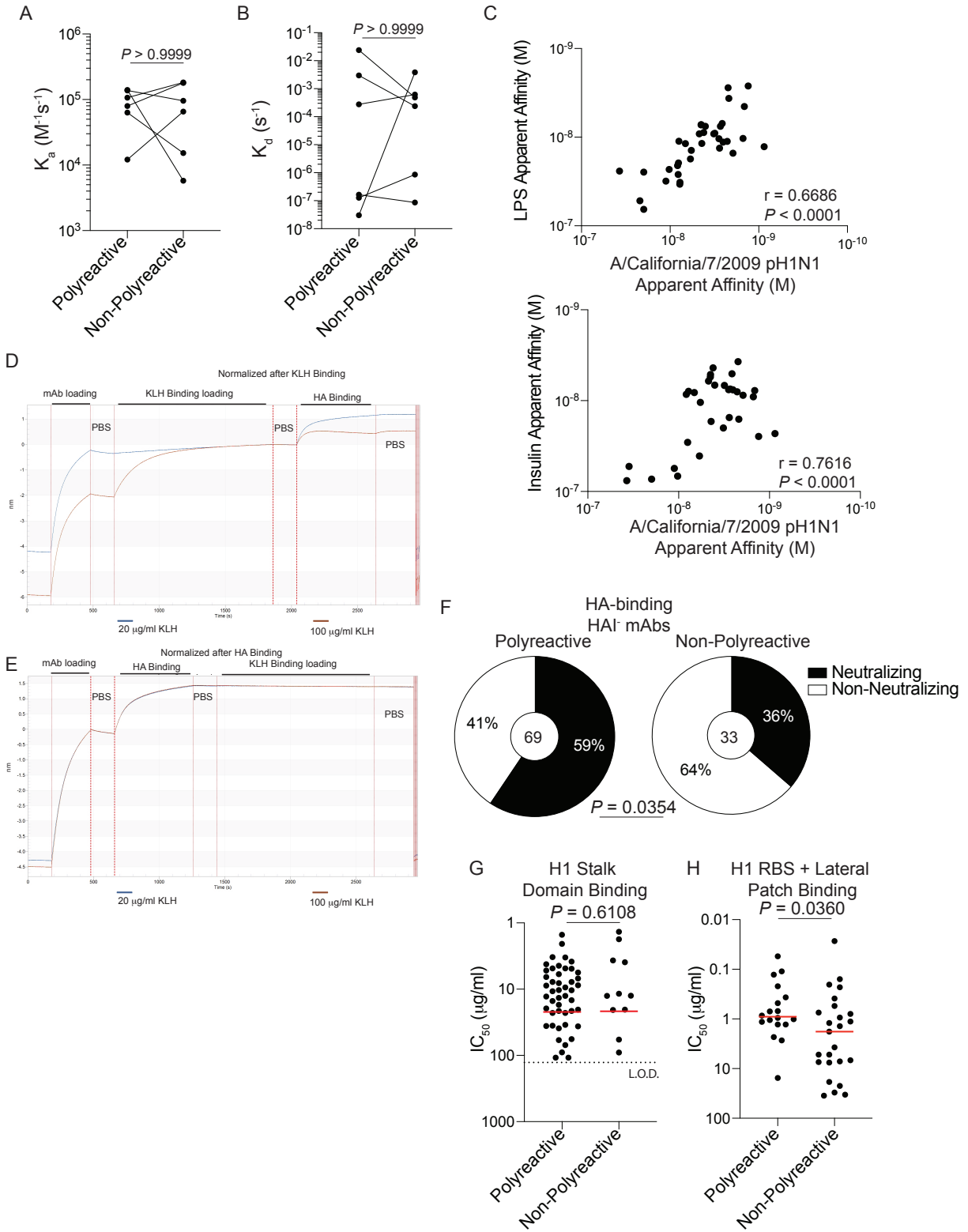
**Figure S2: Polyreactivity of antigen-specific mAbs. Related to Figure 1. (A)** Proportion of mAbs binding HA, NA, and NP that are polyreactive. **(B)** Proportion of RBS or lateral patch-binding mAbs that are polyreactive. **(C)** Polyreactivity of 4 published broadly neutralizing mAbs. The number in the center of pie graphs indicates the number of mAbs tested. Each antibody was tested in duplicate twice and the data are mean  $\pm$  S.E.M. **(D)** Proportion of influenza virus positive polyreactive mAbs that are part of a clonal expansion. **(E)** Proportion of clones that only have polyreactive members, non-polyreactive members, or a mix of polyreactive and non-polyreactive

members. **(F)** Polyreactivity of clones based on antigen specificity. Each clone line on the left-hand side is one clonal expansion against HA (n=34 clones). For data in **A**, **B**, and **D**, the number in the center of each pie-graph is the number of mAbs tested. For data in **E**, the number in the center of the pie graph is the number of influenza virus specific clones analyzed. Data in **B** were analyzed by Fisher's Exact Test relative to other head epitope data in **Figure 1D**. Limit of Detection (L.O.D.) represented as dashed red line.



**Figure S3: Polyreactive mAb induction by different influenza exposures and cross-reactivity of polyreactive mAbs. Related to Figure 2 and Figure 3. (A)** Number of nucleotide mutations of heavy and light chains of mAbs generated from the 2009 MIV (heavy n=131; light n=123), H7N9 vaccine (heavy n=32; light n=31), and seasonal vaccination (TIV+QIV; heavy

n=259; light n=249). **(B)** Proportion of pH1N1<sup>+</sup> mAbs that are polyreactive from individuals vaccinated with the 2009 MIV or 2010-2011 TIV + 2014-2015 QIV, excluding any clonal expansions. **(C-D)** MAbs isolated from elderly subjects ( $\geq 65$  years old) immunized with seasonal influenza vaccines **(C)** or from adults infected with seasonal influenza A viruses **(D)** were tested for polyreactivity. **(E)** Epitope targeting of polyreactive and non-polyreactive mAbs induced by the 2009 MIV (left) or seasonal vaccination (right). **(F)** Proportion of polyreactive and non-polyreactive mAbs per subject (n=12) binding to 7-9 H1N1 strains, based on data in **Figure 3A**. Each line connects the proportion of polyreactive and non-polyreactive mAbs binding 7-9 H1N1 strains from each subject. **(G)** Binding affinity (as shown as AUC) of polyreactive mAbs (n=50) induced by the 2009 MIV against A/California/7/2009 and A/swine/Mexico/AVX8/2011 (H1N2). **(H)** Binding affinity (as shown as AUC) polyreactive mAbs (n=13) induced by the 2009 MIV and the 2014 QIV against A/California/7/2009 and A/Vietnam/1203/2004 recombinant H5. For data in **A**, each symbol represents one mAb and the red bar is the median. Lines in **F** and **G** connect the same mAb binding A/California/7/2009 and A/swine/Mexico/AVX8/2011 (**F**) or A/Vietnam/1203/2004 rH5 (**G**). For data in **B-E**, the number in the center of each pie graph is the number of mAbs tested. Data in **A** were analyzed by a non-parametric Kruskal-Wallis Test, data in **B-D** were analyzed by Fisher's Exact Test, data in **E** were analyzed by using Chi-square Tests, and data in **F-H** were analyzed by a paired non-parametric Wilcoxon matched-pairs signed rank Test.

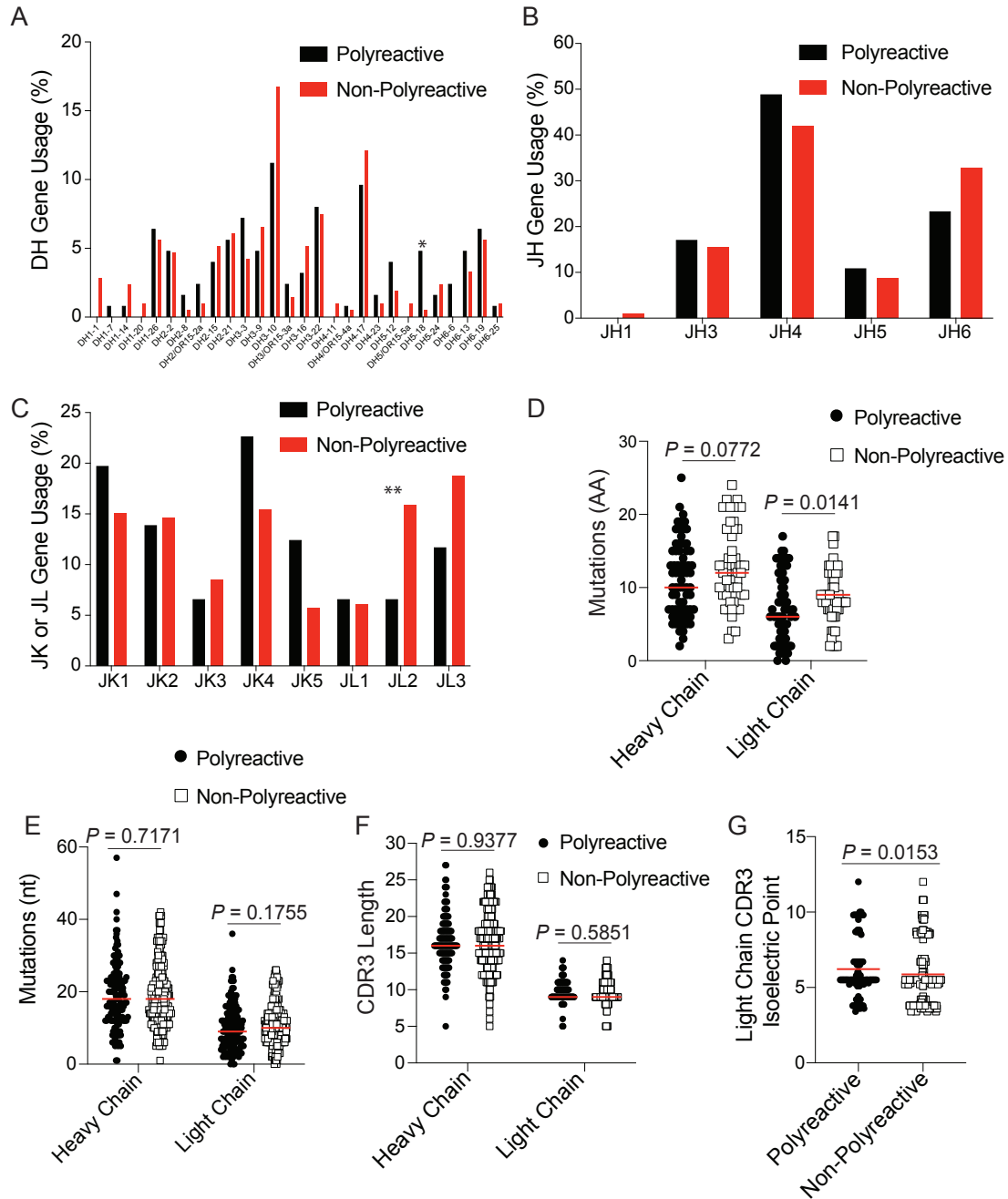


**Figure S4: Polyreactivity augments viral binding and neutralization. Related to Figure 4.**

**(A-B)**  $K_a$  **(A)** and  $K_d$  **(B)** of polyreactive and non-polyreactive mAbs from the same clone binding to A/California/7/2009 HA. Each line connects polyreactive and non-polyreactive clonal members

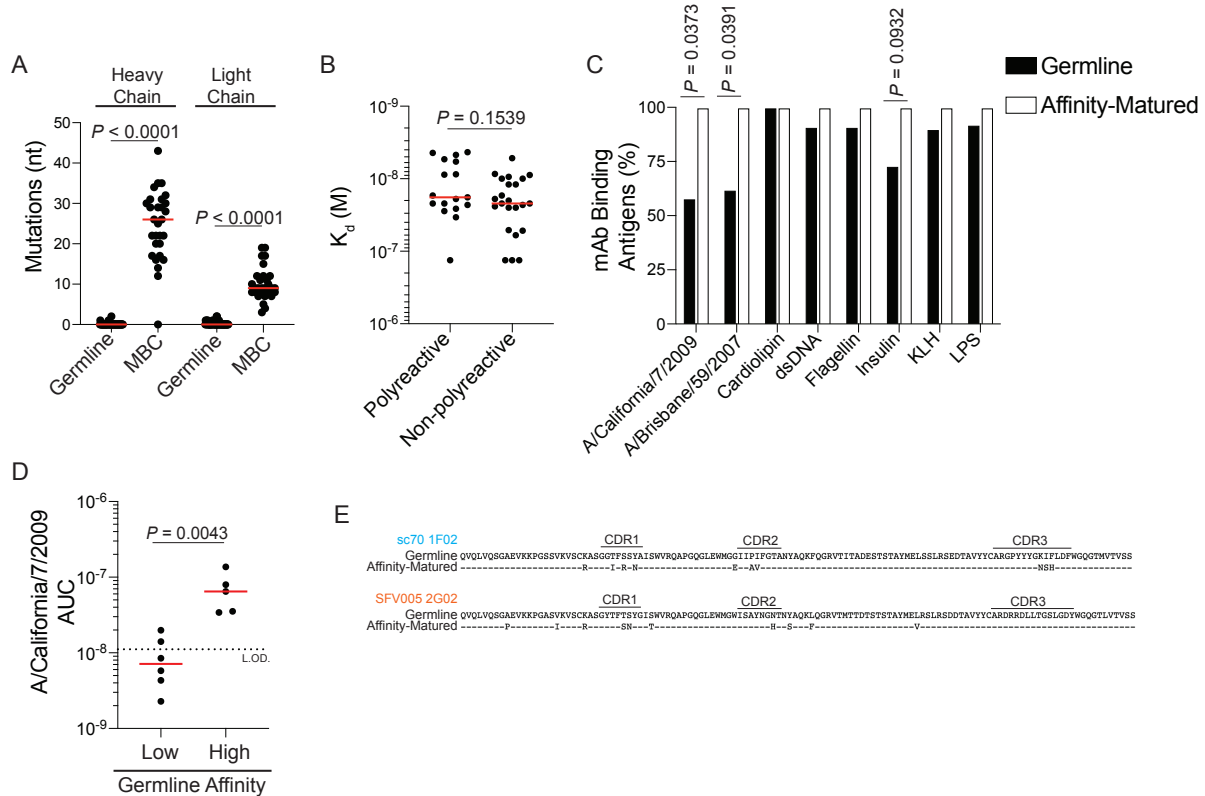
(n=6). **(C)** Spearman Correlation of the apparent affinity ( $K_d$ ) of polyreactive mAb binding to A/California/7/2009 virus and LPS (top; n=36) or Insulin (bottom; n=30). **(D-E)** Using biolayer interferometry, a Protein A sensor was loaded with SFV005 2G02 (polyreactive mAb). **(D)** The sensor was then dipped in 20  $\mu\text{g/ml}$  or 100  $\mu\text{g/ml}$  of KLH, followed by 10  $\mu\text{g/ml}$  A/California/7/2009 HA. **(E)** After SFV005 2G02, the sensor was dipped into 10  $\mu\text{g/ml}$  A/California/7/2009 HA, and then dipped in 20  $\mu\text{g/ml}$  or 100  $\mu\text{g/ml}$  of KLH. Data are representative of 10 mAbs. The assays were performed twice for each antibody. **(F)** Polyreactive and non-polyreactive antibodies targeting HA<sup>+</sup> HAI<sup>-</sup> epitopes were tested for neutralization against A/California/7/2009. Proportion of polyreactive and non-polyreactive antibodies that are neutralizing. **(G-H)** Neutralization potency ( $\text{IC}_{50}$ ) against A/California/7/2009 virus of polyreactive (n=47) and non-polyreactive (n=11) mAbs targeting the stalk domain **(G)** and of polyreactive (n=18) and non-polyreactive (n=25) mAbs targeting the RBS and lateral patch **(H)**. For data in **C**, **G**, and **H**, each symbol represents one mAb and the red bar indicates the median. For data in **F**, the number in the center of each pie graph is the number of mAbs tested. Data in **A** and **B** were analyzed by a paired non-parametric Wilcoxon matched-pairs signed rank Test. Data in **F** were analyzed using a Fisher's Exact Test and data in **G** and **H** were analyzed using a non-parametric Mann-Whitney Test. Limit of Detection (L.O.D.) represented as dashed black line.





**Figure S5: Repertoire and biochemical characteristics of polyreactive and non-polyreactive antibodies. Related to Figure 5. (A-B) DH (A) and JH (B) gene usage by polyreactive and non-polyreactive antibodies. (C) JK or JL gene usage by polyreactive and non-polyreactive antibodies. (D) Somatic hyper mutations (amino acid changes) of polyreactive (heavy n=71; light 68) and non-polyreactive (heavy n=55; light n=53) mAbs induced by the 2009 MIV. (E) Somatic hypermutations (nucleotide mutations) of heavy and light chains of all polyreactive (n=137) and non-polyreactive (n=246) mAbs. (F) Heavy chain and light chain CDR3 length of**

polyreactive (n=137) and non-polyreactive mAbs (heavy n=245; light n=246). (**G**) Light chain CDR3 isoelectric point of polyreactive (n=137) and non-polyreactive (n=246) mAbs. For data in **D-G**, each symbol represents one mAb and the red bar indicates the median. Data in **A-C** were analyzed by Fisher's Exact Tests, and data in **D-G** were analyzed by unpaired non-parametric Mann-Whitney Tests. Each symbol represents a single antibody. \*  $P \leq 0.05$ ; \*\*  $P \leq 0.01$



**Figure S6: Germline precursors of broadly-reactive antibodies are polyreactive. Related to Figure 6.** (A) Somatic hypermutations (nucleotide mutations) of heavy and light chains of stalk domain-binding germline (n=50) and MBC (n=29) mAbs tested in Figure 6A. (B) Affinity of polyreactive (n=17) and non-polyreactive (n=23) germline mAbs binding the stalk domain. (C) Proportion of reverted germline mAbs generated from affinity-matured polyreactive and corresponding affinity-matured mAbs binding influenza viruses and polyreactive panel antigens. (D) Area under the curve (AUC) of reverted germline mAbs categorized as high (n=5) or low (n=6) affinity binding to A/California/7/2009 related to Figure 6E. (E) Heavy chain sequences of germline and affinity-matured sc70 1F02 and SFV005 2G02. For data in A, B, and D, each symbol represents one mAb and the red bar indicates the median. Data in A, B, and D were analyzed by an unpaired non-parametric Mann-Whitney Test and data in C were analyzed by Fisher's Exact Test.

**Table S1: Influenza vaccination and infection and influenza-negative naïve B cell and MBC cohorts.** Related to STAR Methods.

Cohort	# of Subjects	# of mAbs	Average # mAbs per subject (range)	Reference
pH1N1 MIV	11	133	12 (1 – 29)	(Andrews et al., 2015a)
2010-2011 TIV	12	48	4 (1 – 6)	(Andrews et al., 2015a)
2014-2015 QIV	8	166	21 (8 – 53)	(Neu et al., 2019)
H7N9 LAIV/IIV	5	31	6 (2 – 16)	(Henry Dunand et al., 2016)
Elderly pre-H1N1 TIV	13	77	6 (1 – 21)	(Henry et al., 2019)
Chimeric HA Vaccine Germline	12	50	4 (1 – 16)	(Bernstein et al., 2019)
Chimeric HA Vaccine MBCs	12	29	2 (1 – 11)	(Bernstein et al., 2019)
2014-2015 H3N2 Infected	3	18	6 (2 – 10)	(Chen et al., 2018)
2015-2016 H1N1 Infected	4	21	5 (1 – 10)	(Chen et al., 2018)
Influenza-Negative Naïve B cells	3	52	17 (11 – 28)	(Duty et al., 2009)
Influenza-Negative MBCs	4	56	14 (7 – 22)	(Koelsch et al., 2007)

**Table S2: Subject demographics for cohorts.** Related to STAR Methods. \*No demographic information was obtained from the H7N9 LAIV/IIV cohort and the influenza-negative naïve B cell and MBC cohorts.

Cohort	# of Subjects	Male (%)	Mean Age [Range]
pH1N1 MIV	11	36.4	41.1 (24 – 64)
2010-2011 TIV	12	58.3	29.3 (23 – 43)
2014-2015 QIV	8	37.5	29.9 (24 – 34)
Elderly pre-H1N1 TIV	13	53.8	75.7 (71 – 89)
Chimeric HA Vaccine Germline	12	25	27.7 (20 – 37)
Chimeric HA Vaccine MBCs	12	25	30.9 (24 – 36)
2014-2015 H3N2 Infected	3	66.7	43 (34 – 49)
2015-2016 H1N1 Infected	4	31.25	31.3 (23 – 46)