



Supplemental Figure 1. Analysis of genes differentially regulated by TS in ccRCCs and NKTs. (A) Numbers of genes differentially regulated in ccRCC and NKTs from LTS compared to NS. (B) Lack of overlap between genes regulated differentially by TS in ccRCCs and NKTs. (C) Heatmap of all differentially regulated genes in NKTs from LTS compared to NS showing stratification with TS (marked by black in the top bar). Four samples from LTS stratified with NS. (D) Heatmap of pathways regulated by TS in ccRCCs. Magenta boxes on the right of the heatmap mark the GSEA mitochondrial/oxidative phosphorylation categories. See Supplemental Table 2C for definitions of the categories. (E) Box-whisker plots for expression of mitochondrial genes in NKTs from NS and LTS. The boxes represent lower and upper guartiles separated by the median (thick horizontal line) and the whiskers extend to the minimum and maximum values, excluding points that are outside the 1.5 interguartile range from the box (marked as circles). P values from Mann-Whitney-Wilcoxon test are provided at the top. (F) Heatmap of pathways regulated by TS in NKTs. Boxes on the right of the heatmap mark the GSEA categories including genes included in the response to arsenic (vellow), genes regulated by PRC2 (magenta), and extracellular matrix regulatory genes (blue). See Supplemental Table 3C for definition of GSEA categories. (G) Box-whisker plots and P values from Mann-Whitney-Wilcoxon test for expression of arsenic response genes in NKTs from NS and LTS. (H) Box-whisker plots and P values from Mann-Whitney-Wilcoxon test for expression of arsenic response genes in ccRCCs from NS and LTS. NKTs from 15 NS and 16 LTS were analyzed.





Supplemental Figure 2. Analysis of metabolites' steady-state levels in ccRCCs and NKTs from LTS

and NS. (A) Heatmap of all differentially abundant metabolites showing stratification with tissue type but not with smoking status. (B) Metabolic pathways enriched for metabolites with differential abundance in ccRCCs and NKTs.





Supplemental Figure 3. Alterations in the intercorrelations of metabolites. (A) Circos showing unique edges for Spearman correlations among 133 metabolites with Spearman correlation coefficient ≥ 0.5 comparing NKTs and ccRCCs in NS and LTS. (B) Circos showing unique edges for Spearman correlations among 133 metabolites comparing NKTs from NS and LTS.



Supplemental Figure 4. Reprogramming of glycolytic metabolism in ccRCCs from LTS. (A) Bar graph showing the number of correlations for each individual glycolytic intermediate for each tissue shown in Fig. 4C, left. Numbers above the bars indicate number of correlations. (B) Enrichment analysis of pathways associated with metabolites correlated with glycolytic intermediates in ccRCCs from NS using MetaboAnalyst. (C) Enrichment analysis of pathways associated with metabolites correlated with glycolytic intermediates correlated with glycolytic intermediates in ccRCCs from NS using MetaboAnalyst. (C) Enrichment analysis of pathways associated with metabolites correlated with glycolytic intermediates in ccRCCs from LTS using MetaboAnalyst.



Supplemental Figure 5. Reprogramming of purine connections in ccRCCs from LTS. (A) Box-whisker plots of ATP concentration in NKTs and ccRCCs from NS and LTS. (B) Circos showing correlations of IMP in NKTs and tumors from NS and LTS. Numbers indicate number of correlations in each tissue.





Supplemental Figure 6. Reprogramming of amino acid metabolism. (A) Circos showing correlation for Asp (blue edges) in ccRCCs from NS and LTS. (B) Schematic representation of pathways utilizing Asp. Metabolites correlated with Asp marked in pink. Legend represents values of Spearman correlation coefficient (SCC) for Asp. (C) Circos show correlation for Glu (blue edges) in ccRCCs from NS and LTS. (D) Schematic representation of pathways utilizing Glu. Metabolites correlated with Glu marked in blue. Legend represents values of SCC for Glu. (E) Circos show correlations of His (blue edges) with other metabolites in ccRCCs from NS and LTS. (F) Circos show correlations of Lys (blue edges) with other metabolites in ccRCCs from NS and LTS. (G) Circos show correlations of Gln (blues edges) with other metabolites in ccRCCs from LTS and NS (H) Schematic representation of Gln metabolic connections in ccRCCs from NS and LTS. Green scale represents values of SCC for Gln. (I) Inserts derived from the Circos show correlations of ATP (green edges) with amino acid in ccRCC from NS and LTS. Numbers above all Circos indicate number of edges in the indicated tissues.



Supplemental Figure 7. Metallomic analysis of TS effects in NKTs and ccRCCs from NS and LTS. (A) Principal component analysis showing different distribution of metals between NKTs (blue) and ccRCCs (red). (B) Levels of MT in NKTs and ccRCCs from NS and LTS. (C) SEC-ICP-MS chromatogram shown in Fig. 6E. Arrows indicate retention times for the molecular weight standards used to calibrate the SEC column size. (D) SEC-ICP-MS chromatograms from indicated tissues for Cd and As. Y-axis on the left represents the intensity of signal for the metals. HMW- high molecular weight fraction, LMW- low molecular weight fraction, MT- metallothioneins.



Supplemental Figure 8. Identification of ccRCC subtypes based on stratification using metabolic set of genes. (A) Box Whisker plots showing distribution of average expression of MG-154 (right) compared to the distribution of average expression of all genes (18,161) (left) measured in transcriptomics analysis in the entire TS cohort. P=3.72e⁻⁷¹ (Wilcoxon test). (B) Box Whisker plots showing distribution of average expression of MG-154 (right) compared to the distribution of average expression of all genes (left) in the TCGA cohort of 309 Caucasian males. P=3.51e⁻⁴² (Wilcoxon test). Note that TS and TCGA cohorts were processed and normalized differently resulting in different scales for expression values without affecting analysis of relative expression levels. (C) Box Whisker plots showing distribution of average expression of MG-154 in the three subtypes identified in the CTSC cohort. ****P=1.98e⁻⁰⁵, **P=0.0052 (Wilcoxon test). (D) Percent similarity between indicated CTSC and TCGA subtypes determined by the nearest centroid approach. (E) Box Whisker plots showing distribution of average expression of MG-154 in the six subtypes identified in the CTSC cohort. ****P=1.98e⁻⁰⁵, **P=0.0052 (Wilcoxon test). (D) Percent similarity between indicated CTSC and TCGA subtypes determined by the nearest centroid approach. (E) Box Whisker plots showing distribution of average expression of MG-154 in the six subtypes identified in the CTSC cohort. ****P=1.98e⁻⁰⁵, **P=0.0052 (Wilcoxon test). (D) Percent similarity between indicated CTSC and TCGA subtypes determined by the nearest centroid approach. (E) Box Whisker plots showing distribution of average expression of MG-154 in the six subtypes identified in TCGA cohort. (F) P values determined by Wilcoxon test for all the comparison among 6 TCGA subtypes.



Supplemental Figure 9. Characterization of metabolic subtypes in TCGA. (A) Stacked bar graphs showing distribution of tumor grades among 6 TCGA subtypes. The only significant difference in G2+G1 vs. G3+G4 is between subtypes 4 and 2. *, P=0.02 (chi-square). (B) Stacked bar graphs showing distribution of tumor stage among 6 subtypes. There is a significant difference in stage I+II vs III+IV in subtype 4 when compared to subtype 2 and 1. *, P=0.028, ***, P=0.008 (chi-square test). (C) Stacked bar graph showing status of metastases in the indicated subtype. M0- no distant metastases; M1- metastatic spread to different tissues. There is no difference between subtype 4 and other subtypes. (D) Average expression of MG-154 in TCGA tumors with either wild type or mutated indicted tumor suppressor genes.