## **Supplementary Materials**

## Mitochondrial ROS promote mitochondrial dysfunction and inflammation in ischemic acute kidney injury by disrupting TFAM-mediated mtDNA maintenance

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**Figure S1**. The vehicle (DMSO/PBS) alone had no influence on renal lesions in IRI-AKI mice. (A) Serum CREA and UREAL concentrations of mice on day 5 after IRI-AKI (n = 6; \*p < 0.05 vs. CON group). (B) Representative micrographs of renal H&E staining in mice with different treatments. (C) Real-time PCR analysis of Bax, ICAM-1, MCP-1, TFAM, ATP5a-1, and PGC-1 $\alpha$  mRNA levels in kidneys of mice on day 5 after IRI-AKI (n = 3; \*p < 0.05 vs. CON group). (D) mtDNA copy number in kidneys of mice on day 5 after IRI-AKI (n = 3; \*p < 0.05 vs. CON group).



**Figure S2**. Measurement of ROS production in HK2 cells under different stress conditions. (A) Determination of intracellular ROS in the HK2 cells under H/R conditions using the DCFH-DA staining (scale bar = 50  $\mu$ m). (B) Determination of intracellular ROS and mtROS in HK2 cells under t-BHP conditions with the DCFH-DA and MitoSOX staining (Scale bar = 50  $\mu$ m). (C) Determination of mtROS level in HK2 cells under t-BHP conditions using flow cytometry with MitoSOX staining.



**Figure S3**. TFAM degradation was enhanced in HK2 cells under oxidative stress. (A) Cell viability was determined by CCK8 assay (n = 3; \*p < 0.05 *vs*. Control group; <sup>#</sup>p < 0.05 *vs*. t-BHP group). (B) Western blotting of TFAM, Lon, and p53 proteins in HK2 cells under t-BHP conditions with or without bortezomib treatment. (C) Quantitative analysis of protein expression detected by western blotting (n = 3; \*p < 0.05 *vs*. Control group; <sup>#</sup>p < 0.05 *vs*. t-BHP group).



**Figure S4**. (A) Observation of mitochondria and quantification of mitochondrial length in HK2 cells under H/R conditions. HK2 cells were treated with or without MT and then stained with anti-TOM20. (B) Real-time PCR analysis of PGC-1 $\alpha$ , ATP5a-1, and NDUFS8 mRNA levels in HK2 cells under H/R conditions with or without MT treatment (n = 3; \*p < 0.05 *vs*. Control group; <sup>#</sup>p < 0.05 *vs*. H/R group).



Figure S5. (A) Double-IF staining of TOM20 (red) and TFAM (green) in the HK2 cells under H/R conditions with or without TFAM siRNA treatment (scale = 10  $\mu$ m). (B) Quantitative analysis of TFAM protein expression in the HK2 cells (n = 6; \*p < 0.05 *vs*. Control group; <sup>#</sup>p < 0.05 *vs*. H/R group; <sup>&</sup>p < 0.05 vs. MT group).



Figure S6. TFAM deficiency abolished the protective role of mtROS scavenger in the HK2 cells. (A) Real-time PCR analysis of TFAM mRNA levels in HK2 cells. (B) Western blotting analysis of TFAM protein levels in the HK2 cells at 48 h after transfection. (C) Real-time PCR analysis of TFAM and PGC-1 $\alpha$  mRNA levels in HK2 cells under H/R conditions (n = 3; \*p < 0.05 *vs*. Control group; <sup>#</sup>p < 0.05 *vs*. H/R group; <sup>&</sup>P < 0.05 *vs*. MT group).



Figure S7. Loss of TFAM under oxidative stress induced cytokine production in HK2 cells. (A) Western blotting analysis of ICAM-1, Bax, and TFAM protein levels in HK2 cells under H/R conditions with various treatments. (B) Quantitative analysis of ICAM-1 and TFAM protein expression detected by western blotting (n = 3; \*p < 0.05 *vs*. Control group;  $^{\#}p < 0.05$  *vs*. H/R group;  $^{\&}P < 0.05$  *vs*. MT group).



**Figure S8**. (A) Double-IF staining of TFAM (red) and dsDNA (green) in HK2 cells under H/R conditions with various treatments (scale bar = 10  $\mu$ m). (B) Average size of mtDNA nucleoid in HK2 cells of different groups (n = 20 cells; \*p < 0.05 *vs*. Control group; <sup>#</sup>p < 0.05 *vs*. H/R group). (C) Quantification of cytoplasmic dsDNA (yellow arrows) intensity in HK2 cells (n = 16 cells; \*p < 0.05 *vs*. Control group; <sup>#</sup>p < 0.05 *vs*. H/R group; <sup>&</sup>p < 0.05 vs. MT group).



Figure S9. Representative images of the TFAM (red), dsDNA (green), and DAPI costaining in kidney of normal control mice (scale bar =  $10 \mu m$ ).

Gene	Sequence 5'-3'	Species
TFAM	AGCTCAGAACCCAGATGCAA	
	CCGCCCTATAAGCATCTTGA	
PGC1-α	TGCTGAAGAGGCAAGAGACA	
	CACACGCACACTCCATC	
NDUFS8	CATCTACTGCGGCTTCTGC	
	GGGCGTCACCGATACAAGT	
ATP5a-1	AGAGGACAGGAGCCATTGTG	
	TCAGACCAACTCGCCTACG	
UQCRC1	CAGTCCTCTCAGCCCACTTG	Human
	CCGATTCTTTGTTCCCTTGA	
IL-1β	TGGCAGAAAGGGAACAGAAA	
	CTGGCTGATGGACAGGAGAT	
TNF-α	TGCTGCACTTTGGAGTGATCG	
	TGTCACTCGGGGTTCGAGAAG	
GAPDH	ACCACAGTCCATGCCATCAC	
	TCCACCACCCTGTTGCTGTA	
TFAM	CACCCAGATGCAAAACTTTCAG	
	CTGCTCTTTATACTTGCTCACAG	
PGC1-α	CACCAAACCCACAGAAAACAG	
	GGGTCAGAGGAAGAGATAAAGTTG	
ATP5a-1	CATTGGTGATGGTATTGCGC	
	TCCCAAACACGACAACTCC	
Bax	TGGAGATGAACTGGACAGCA	Mouse
	TGAAGTTGCCATCAGCAAAC	
MCP-1	AGTTGACCCGTAAATCTGAAGC	
	GTGGTTGTGGAAAAGGTAGTGG	
ICAM-1	ACCCAACTGGAAGCTGTTTG	
	CACACTCTCCGGAAACGAAT	
TNF-α	CCAGGAGAAAGTCAGCCTCCT	
	TCATACCAGGGCTTGAGCTCA	
GAPDH	CAGATCCACAACGGATATATTGGG	
	CATGACAACTTTGGCATTGTGG	

Table S1 Primers used in Real-time PCR assay

Gene	Sequence 5'-3'	Species
hB2M	TGTTCCTGCTGGGTAGCTCT	Human
	CCTCCATGATGCTGCTTACA	
mtND1	CACTTTCCACACAGACATCA	
	TGGTTAGGCTGGTGTTAGGG	
COX2	ATAACCGAGTCGTTCTGCCAAT	Mouse
	TTTCAGAGCATTGGCCATAGAA	
Rsp18	TGTGTTAGGGGACTGGTGGACA	
	CATCACCCACTTACCCCCAAAA	

 Table S2 Primers used in mtDNA copy number assay